Bifunctional dendrons for multiple carbohydrate presentation via carbonyl chemistry

Davide Bini, Francesco Nicotra and Laura Cipolla*

Letter

Address:

Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy

Email

Laura Cipolla* - laura.cipolla@unimib.it

* Corresponding author

Keywords:

bis-MPA; carbohydrates; dendrons; levulinic acid; multivalency; multivalent glycosystems

Beilstein J. Org. Chem. **2014**, *10*, 1686–1691. doi:10.3762/bjoc.10.177

Received: 28 February 2014 Accepted: 27 June 2014 Published: 25 July 2014

This article is part of the Thematic Series "Multivalent glycosystems for

Open Access

anoscience".

Guest Editor: B. Turnbull

© 2014 Bini et al; licensee Beilstein-Institut. License and terms: see end of document.

Abstract

The synthesis of new dendrons of the generations 0, 1 and 2 with a double bond at the focal point and a carbonyl group at the termini has been carried out. The carbonyl group has been exploited for the multivalent conjugation to a sample saccharide by reductive amination and alkoxyamine conjugation.

Introduction

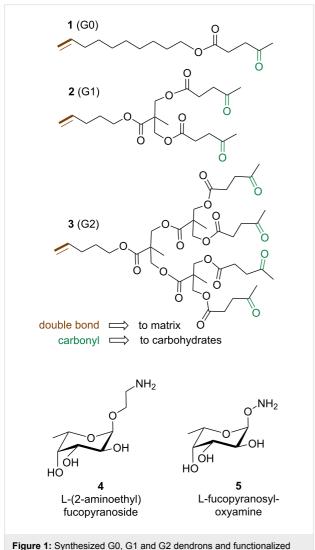
Recognition processes between glycans and their receptors are of paramount relevance in several biological phenomena, both in physiological [1,2] and in pathological [3-5] conditions. These processes can be exploited in diagnostic tools [6,7], in nanobiotechnology applications [8], and in the development of smart biomaterials for regenerative medicine [9-12]. Beside the variation of carbohydrate residues in glycoconjugates, a key issue in the recognition process is their spatial topographical presentation eliciting high affinity recognition events. In order to better understand these phenomena, dendrimers and dendrons have been developed to provide multivalent glycoconjugates [13,14]. Here, we propose the synthesis of novel dendron struc-

tures which allow for the multivalent conjugation of carbohydrates via carbonyl chemistry.

Results and Discussion

The heterobifunctional dendrons were designed in order to have bio-orthogonal functional groups at the focal point and at their termini. More specifically, a double bond was placed at the desired matrix as the focal point for further conjugation by thiol—ene chemistry, and carbonyl groups were added at the termini. The carbonyl groups can be exploited for carbohydrate functionalization [15,16] by reductive amination, oxime or hydrazone formation to yield suitably functionalized saccha-

rides (Figure 1). Given the relevance of L-fucose in mammal oligosaccharides, α-L-(2-aminoethyl) fucoside [17] and α-O-Lfucopyranosyloxyamine [18] were used as sample monosaccharides for the conjugation of the dendron (Scheme 1).



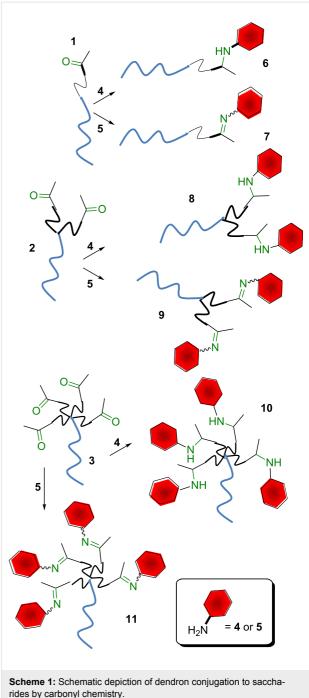
saccharides used for carbonyl conjugation.

Synthesis of dendrons

Zero, first and second generation heterobifunctional dendrons 1-3 were synthesized starting from 9-decen-1-ol (12) or selected building blocks 13 and 14 (Scheme 2) [11] by esterification reactions with levulinic acid (15). Building blocks 13 and 14 were synthesized starting from bis-(hydroxymethyl)propionic acid (bis-MPA) and bromo-1-pentene [11] in one and four steps, respectively.

L-Fucose derivatives synthesis

α-L-(2-Aminoethyl) fucopyranoside (4) and α-O-L-fucopyranosyloxyamine (5) were synthesized from commercial



rides by carbonyl chemistry.

L-fucopyranose in 4 and 5 steps, respectively, as already reported by Flitsch and co-workers [17] and Dumy and co-workers [18].

Dendron conjugation to L-fucose by reductive amination

α-L-(2-Aminoethyl) fucopyranoside (4) was conjugated first to G0 dendron 1 by reductive amination in the presence of NaCNBH₃ (Scheme 3). The reaction afforded the desired glyco-

Scheme 3: Dendron conjugation to fucose moieties by reductive amination. Reagents and conditions: a) 4, 3 M Na_2SO_4 , AcOH, $NaCNBH_3$, EtOH, 80 °C, 6 h.

sylated dendron 6 in 27% yield. The very low yield was ascribed to the competing carbonyl reduction to the corresponding alcohol 16 as a byproduct.

The same reaction on the G1 dendron 2 gave an even more complex mixture of products, identified by mass spectrometry (System Applied Biosystems MDS SCIEX instruments: Q TRAP, LC/MS/MS, turbo ion spray and Q STAR elite nano spray) performed directly on the TLC spots following literature procedures [19]. By mass values, the mixture was composed of the desired fucosylated dendron 8 as the minor product together with the monoglycosylated derivatives 17 and 18 and the alcohols 19 and 20. In order to reduce the formation of alcohol byproducts, a "milder" reducing agent such as Na(AcO)₃BH was tried, but without any success.

Given the high extent of byproducts and the low efficiency of the glycoconjugation to the G0 dendron and G1 dendron by reductive amination, we decided to evaluate the possibility to obtain better conjugation yields by oxime ligation. Thus, G0, G1 and G2 dendrons 1–3 were reacted with α -O-L-fucopyranosyloxyamine (5) in citrate buffer at pH 3.5 [20] (Scheme 4). Due to the partial hydrophobic nature of the dendrons 1–3 they do not fully dissolve in the buffer, and the solution is not completely clear.

The dendrons 1-3 were reacted overnight at room temperature with α -O-L-fucopyranosyloxyamine (5) affording the desired glycoconjugate structures 7, 9 and 11 in quantitative yields.

Conclusion

In conclusion, novel G0, G1 and G2 dendrons suitable for glycoconjugation by carbonyl chemistry were synthesized. The conjugation of the saccharide by reductive amination was characterized by a low efficiency. On the other hand, the oxime ligation afforded the glycoconjugated dendrons in quantitative

yields. The glycosylated dendrons can be exploited for further chemoselctive thiol—ene reactions with matrices suitably functionalized with thiol groups, i.e., cysteine residues in proteins.

Experimental General methods

All chemicals were purchased from Sigma-Aldrich and used without further purification. All solvents were dried over molecular sieves, for at least 24 h prior to use, when required. When dry conditions were required, the reaction was performed under an Ar atmosphere. Thin-layer chromatography (TLC) was performed on silica gel 60 F254 coated glass plates (Merck) with UV detection when possible, or spots were visualized by charring with a conc. H₂SO₄/EtOH/H₂O solution (10:45:45 v/v/ v), or with a solution of $(NH_4)_6Mo_7O_{24}$ (21 g), $Ce(SO_4)_2$ (1 g), conc. H₂SO₄ (31 mL) in water (500 mL) and then by heating to 110 °C for 5 min. Flash column chromatography was performed on silica gel 230–400 mesh (Merck). Routine ¹H and ¹³C NMR spectra were recorded on a Varian Mercury instrument at 400 MHz (¹H) and 100.57 MHz (¹³C). Chemical shifts are reported in parts per million downfield from TMS as an internal standard, J values are given in Hz. Mass spectra were recorded on System Applied Biosystems MDS SCIEX instruments: Q TRAP, LC/MS/MS, turbo ion spray and Q STAR elite nanospray.

General procedure for levulinic acid condensation (compounds 1–3): To a 0.1 M solution of the appropriate compound dissolved in dry DCM, levulinic acid (1.2 equiv), DMAP (0.2 equiv) and DCC (1.5 equiv) were added, and the reaction mixture was stirred at room temperature (for 1 to 24 h, depending on the substrate). The precipitates were filtered off and washed with CH_2Cl_2 . The solvent was evaporated, and the residue was purified by column chromatography on a silica gel column with a suitable eluent. See Supporting Information File 1 for full experimental data.

General procedure for reductive amination reaction (compounds 6, 8): The appropriate dendron (1 equiv) and α -L-(2-aminoethyl) fucoside (4, 1 equiv) were dissolved in EtOH (0.1 M in respect to 4). AcOH (1 equiv) and 3 M Na₂SO₄ (1% of solvent volume) were added, and the mixture was heated under reflux for 2 h. NaCNBH₃ (1.5 equiv) was then added, and the reaction was heated under reflux for further 4 h. See Supporting Information File 1 for full experimental data.

General procedure for dendron/ alkoxyamine conjugation (compounds 7, 9, 11): The appropriate dendron (1 equiv) and α -O-L-fucopyranosyloxyamine (5, 1 equiv) were dissolved in citrate buffer (pH 3.5, 0.1 M in respect to 5) and stirred at room temperature overnight. The mixture was concentrated and the

product isolated. See Supporting Information File 1 for full experimental data.

Supporting Information

Supporting Information File 1

Experimental part.

[http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-10-177-S1.pdf]

Acknowledgements

This research was financially supported by the Cariplo Fundation under the grant numbers 2010-0378, 2011-0270 and PRIN 2010L9SH3K.

References

- Johnson, J. L.; Jones, M. B.; Ryan, S. O.; Cobb, B. A. Trends Immunol. 2013, 34, 290–298. doi:10.1016/j.it.2013.01.006
- Moremen, K. W.; Tiemeyer, M.; Nairn, A. V. Nat. Rev. Mol. Cell Biol. 2012, 13, 448–462. doi:10.1038/nrm3383
- Schedin-Weiss, S.; Winblad, B.; Tjernberg, L. O. FEBS J. 2014, 281, 46–62. doi:10.1111/febs.12590
- Kleene, R.; Schachner, M. Nat. Rev. Neurosci. 2004, 5, 195–208. doi:10.1038/nrn1349
- Ohtsubo, K.; Marth, J. D. Cell 2006, 126, 855–867. doi:10.1016/j.cell.2006.08.019
- Morais, G. R.; Falconer, R. A.; Santos, I. Eur. J. Org. Chem. 2013, 1401–1410. doi:10.1002/ejoc.201201457
- Cipolla, L.; Gregori, M.; So, P.-W. Curr. Med. Chem. 2011, 18, 1002–1018. doi:10.2174/092986711794940851
- Sapsford, K. E.; Algar, W. R.; Berti, L.; Boeneman Gemmill, K.; Casey, B. J.; Oh, E.; Stewart, M. H.; Medintz, I. L. Chem. Rev. 2013, 113, 1904–2074. doi:10.1021/cr300143v
- Cipolla, L.; Russo, L.; Taraballi, F.; Lupo, C.; Bini, D.; Gabrielli, L.; Capitoli, A.; Nicotra, F. Smart biomaterials: the contribution of glycoscience. In Specialist Periodical Reports, SPR Carbohydrate Chemistry; 2012, Vol. 38, pp. 416–445.
- 10. Russo, L.; Battocchio, C.; Secchi, V.; Magnano, E.; Nappini, S.; Taraballi, F.; Gabrielli, L.; Comelli, F.; Papagni, A.; Costa, B.; Polzonetti, G.; Nicotra, F.; Natalello, A.; Doglia, S. M.; Cipolla, L. Langmuir 2014, 30, 1336–1342. doi:10.1021/la404310p
- Bini, D.; Russo, L.; Battocchio, C.; Natalello, A.; Polzonetti, G.;
 Doglia, S. M.; Nicotra, F.; Cipolla, L. *Org. Lett.* **2014**, *16*, 1298–1301.
 doi:10.1021/ol403476z
- Du, J.; Yarema, K. J. Adv. Drug Delivery Rev. 2010, 62, 671–682. doi:10.1016/j.addr.2010.01.003
- Bernardi, A.; Jimenez-Barbero, J.; Casnati, A.; De Castro, C.; Darbre, T.; Fieschi, F.; Finne, J.; Funken, H.; Jaeger, K.-E.; Lahmann, M.; Lindhorst, T. K.; et al. Chem. Soc. Rev. 2013, 42, 4709–4727. doi:10.1039/c2cs35408j
- 14. Sanchez-Navarro, M.; Rojo, J. *Drug News Perspect.* **2010**, *23*, 557–572. doi:10.1358/dnp.2010.23.9.1437246
- Paez, J. I.; Martinelli, M.; Brunetti, V.; Strumia, M. C. Polymers 2012, 4, 355–395. doi:10.3390/polym4010355
- Röglin, L.; Lempens, E. H. M.; Meijer, E. W. Angew. Chem., Int. Ed. 2011, 50, 102–112. doi:10.1002/anie.201003968

- Šardzík, R.; Noble, G. T.; Weissenborn, M. J.; Martin, A.; Webb, S. J.; Flitsch, S. L. *Beilstein J. Org. Chem.* **2010**, *6*, 699–703. doi:10.3762/bjoc.6.81
- Duléry, V.; Renaudet, O.; Philouze, C.; Dumy, P. Carbohydr. Res. 2007, 342, 894–900. doi:10.1016/j.carres.2007.02.003
- St. Hilaire, P. M.; Cipolla, L.; Tedebark, U.; Meldal, M. Rapid Commun. Mass Spectrom. 1998, 12, 1475–1484. doi:10.1002/(SICI)1097-0231(19981030)12:20<1475::AID-RCM365>3. 0.CO;2-F
- Hudak, J. E.; Yu, H. H.; Bertozzi, C. R. J. Am. Chem. Soc. 2011, 133, 16127–16135. doi:10.1021/ja206023e

License and Terms

This is an Open Access article under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The license is subject to the *Beilstein Journal of Organic Chemistry* terms and conditions: (http://www.beilstein-journals.org/bjoc)

The definitive version of this article is the electronic one which can be found at:

doi:10.3762/bjoc.10.177