

Oncogenic Roles of RAD51API in Tumor Tissues Related to Overall Survival and Disease-Free Survival in Hepatocellular Carcinoma

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Abstract

Objective: This study aimed to investigate the associations between RAD51API and the outcomes of hepatocellular carcinoma (HCC).

Methods: RAD51API expression levels were compared in Gene Expression Omnibus (GEO) and The Cancer Genome Atlas (TCGA) datasets. The Liver Hepatocellular Carcinoma (TCGA, Provisional) and GSE36376 datasets were used for survival analysis. RAD51API associations with clinicopathological features were determined with the GSE36376 dataset.

Results: RAD51API mRNA expression was significantly upregulated in advanced liver fibrosis samples (S3-4 vs. S0-2 and G3-4 vs. G0-2) from hepatitis B virus (HBV)-related liver fibrosis patients and in tumor tissues and peripheral blood mononuclear cells (PBMCs) from HCC patients (all $P < 0.05$). HCC patients with high RAD51API expression had significantly worse overall survival (OS) and disease-free survival (DFS) than those with low RAD51API expression ($P = 0.0034$ and $P = 0.0012$, respectively) in the TCGA dataset, and these findings were validated with the GSE36376 dataset ($P = 0.0074$ and $P = 0.0003$, respectively). A Cox regression model indicated that RAD51API was a risk factor for OS and DFS in HCC patients in GSE36376 (HR = 1.54, 95% CI = 1.02-2.32, $P = 0.04$ and HR = 1.71, 95% CI = 1.22-2.39, $P = 0.002$, respectively). Moreover, RAD51API mRNA expression increased gradually with increasing tumor stage, including stratification by American Joint Committee on Cancer (AJCC) stages, Barcelona Clinic Liver Cancer (BCLC) stages and Edmondson grades. In addition, RAD51API was overexpressed in HCC patients with intrahepatic metastasis, major portal vein invasion, vascular invasion and/or an alpha-fetoprotein (AFP) level > 300 ng/ml.

Conclusions: Contributing to an advanced tumor stage, intrahepatic metastasis, vascular invasion and AFP level elevation, RAD51API upregulation was significantly associated with OS and DFS in HCC patients.

Keywords

RAD51API, hepatocellular carcinoma, overall survival, disease-free survival, liver fibrosis

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Introduction

An underlying hallmark of human malignancies is their genomic instability, which is associated with an increased vulnerability to accumulating DNA damage.¹ Current evidence indicates that tumors harbor defects in different DNA damage response steps, mainly those related to signaling and repair.² The pathways that determine cell fate are intertwined and play vital roles in tumorigenesis and cancer progression.^{3,4} The unravelling of the molecular mechanisms underlying the DNA damage response in human cancers offers new therapeutic approaches. Targeted therapy to inhibit the DNA damage response in cancers has created the potential for a better therapeutic strategy.^{5,6}

RAD51-associated protein 1 (RAD51AP1) plays a key role in homologous recombination (HR)-mediated chromosome damage repair,⁷ stimulates joint molecule formation and is required for cellular protection against DNA double-strand break-inducing agents.^{8,9} In human somatic cells, knockdown of RAD51AP1 expression results in increased sensitivity to DNA-damaging agents and impaired HR. The available literature indicates that RAD51AP1 is involved in cancer development and progression. Aberrant RAD51AP1 expression and its carcinogenic roles have been reported in many human cancers, including lung cancer,¹⁰ ovarian cancer,¹¹ breast cancer,^{12,13} oropharyngeal squamous cell carcinomas,¹⁴ glioblastomas,¹⁵ melanoma^{16,17} and lymphoma.¹⁸ Although RAD51AP1 expression has been found to be upregulated in hepatocellular carcinoma (HCC),^{19,20} the predictive potential of the oncogenic roles of RAD51AP1 in HCC survival prognosis is rarely studied.

In this analysis, we aimed to investigate the expression of RAD51AP1 in different stages of liver diseases and correlate it with HCC survival and clinicopathological features in TCGA and GEO datasets to offer novel insights into HCC aggressiveness and identify potential therapeutic targets.

Materials and Methods

Ethics Statement

The study protocol for this analysis and informed consent documents were reviewed and approved by the Ethics Committee of the Shanghai Public Health Clinical Center, Fudan University (approval no. 2018-Y003). All the participants provided written informed consent during their hospitalization.

Microarray Data

To compare the RAD51AP1 expression levels in serum and tissues among patients with different stages of liver diseases, including HCC, and healthy candidates, GEO (<https://www.ncbi.nlm.nih.gov/geo/>) datasets including GSE84044,²¹ GSE17548,²² GSE45436,²³ GSE55092,²⁴ GSE101685, GSE49515²⁵ and GSE36376²⁶ were included in this analysis. A TCGA dataset was used to validate RAD51AP1 expression. HCC patients in the TCGA and GSE36376²⁶ datasets were included for survival analysis.

Patients

In the TCGA dataset, 361 HCC patients were included in the analysis after matching tumor pathological type and gene expression data. In the GSE36376 dataset, 240 HCC patients who underwent surgical resection or liver transplantation at the Samsung Medical Center, Seoul, Korea, between July 2000 and May 2006 were included in this analysis. Informed consent was obtained from each patient included in the study, and this study was approved by the institutional review board of the Samsung Medical Center, Seoul, Korea, which is consistent with reports by Lim et al.²⁶

To validate RAD51AP1 mRNA expression in liver fibrosis, total RNA was extracted from 47 liver biopsy tissue specimens (21 with S0-stage fibrosis and 26 with S4-stage fibrosis), which were obtained from the Shanghai Public Health Clinical Center.

Real-Time Quantitative Polymerase Chain Reaction (RT-qPCR)

Reverse transcription and RT-qPCR analysis of RAD51AP1 were conducted according to the manufacturer's instructions (Takara Bio Inc., Shiga, Japan). The primers for RAD51AP1 used for RT-qPCR were created by Sangon Biotech (Sangon Biotech Co., Ltd., Shanghai, China). The sequences were forward: 5'-ATGACAAGCTCTACCAGAGAGAC-3', and reverse: 5'-CACATTAGTGGTGACTGTTGGAA-3'.

Survival Analysis

The Liver Hepatocellular Carcinoma (TCGA, Provisional) dataset in the cBioPortal for Cancer Genomics web service was used to evaluate the potential for RAD51AP1 to predict the overall survival (OS) and disease-free survival (DFS) of HCC patients.^{27,28} To evaluate associations between RAD51AP1 and survival in HCC patients, gene data with Z scores and clinical data for HCC patients in the Liver Hepatocellular Carcinoma (TCGA, Provisional) dataset were obtained from cBioPortal and matched using the VLOOKUP index in Microsoft Excel.

From GSE36376,²⁶ 240 HCC patients were included in this analysis. OS was defined as the time from surgery to the date of death or last follow-up. DFS was defined as the time from surgery to the date of tumor recurrence or death. The censoring time was defined as the final documented date of no evidence of tumor recurrence by imaging. The liver function parameters and clinicopathological features of HCC patients, including vascular invasion, major portal vein invasion, intrahepatic metastasis, multicentric occurrence, tumor stage and Edmondson grade, were all considered.^{26,29}

Statistical Analysis

Differences in variables between individual groups were analyzed using Student's *t* test, the Mann-Whitney *U*-test and the chi-square test based on the variable type. Factors associated with survival were assessed by univariate and multivariate Cox regression analysis. Only covariates significantly associated with

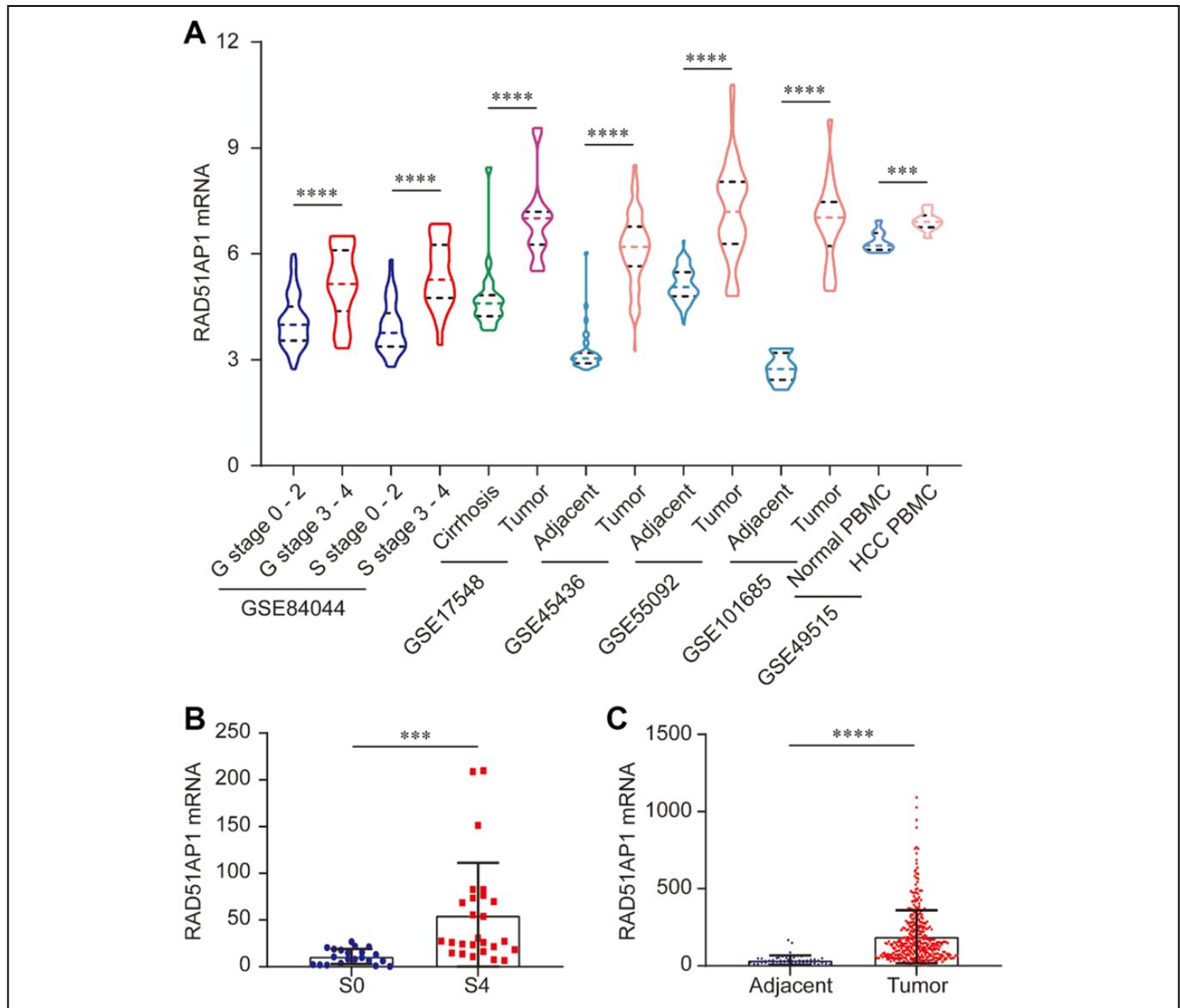


Figure 1. RAD51AP1 mRNA expression. RAD51AP1 mRNA levels in liver fibrosis, cirrhosis, tumor and adjacent tissues, and serum PBMC samples (A); RAD51AP1 mRNA expression in S0 and S4 liver tissues (B); RAD51AP1 mRNA in tumor and adjacent tissues in TCGA database (C).

outcomes in the univariate analysis (2-sided $P < 0.05$) were included in the multivariate model. Parameters significantly associated with outcomes in multivariate model were presented. Results are reported as hazard ratios (HRs) with 95% confidence intervals (CIs). The Kaplan-Meier method with the log-rank test was used to compare OS and DFS between groups. Stata software version 16.0 (Stata Corp LLC, Texas, USA) was used. A 2-tailed $P < 0.05$ were considered significant for all tests.

Results

RAD51AP1 Expression

In GSE84044, RAD51AP1 mRNA expression was significantly elevated in HBV-infected patients with histological stage (S) 3-4 fibrosis and grade (G) 3-4 inflammation compared

to those with S0-2 fibrosis and G0-2 inflammation (both $P < 0.0001$, Figure 1A). In our cohort, we validated that RAD51AP1 mRNA expression was significantly upregulated in chronic liver disease patients with S4 fibrosis compared with those with S0 fibrosis ($P < 0.001$, Figure 1B). In addition, RAD51AP1 was significantly overexpressed in tumor tissue compared to cirrhotic tissue ($P < 0.0001$, Figure 1A, GSE17548). Moreover, RAD51AP1 mRNA expression was significantly higher in tumor tissues than in adjacent tissues in HCC patients, which was confirmed in the 3 GEO datasets GSE445436, GSE55092 and GSE101685 (all $P < 0.0001$, Figure 1A). This trend was also validated in the TCGA dataset ($P < 0.0001$, Figure 1C). Additionally, serum RAD51AP1 mRNA levels were significantly higher in HCC PBMCs than in healthy PBMCs ($P < 0.001$, Figure 1A, GSE49515).

Table 1. Baseline Characteristics of HCC Patients Between RAD51API High and Low Groups in the TCGA Dataset.

Variables	RAD51API high group (n = 180)	RAD51API low group (n = 181)	P value
Age, mean \pm SD, year	58.1 \pm 1.0	61.1 \pm 1.0	0.03
Male, n (%)	117 (65.0)	127 (70.2)	0.294
BMI, mean \pm SD, kg/m ²	26.1 \pm 0.8	26.2 \pm 0.5	0.847
Race, n (%)			0.028
Asian	92 (51.1)	64 (35.4)	
White	78 (43.3)	98 (54.1)	
Others	10 (5.6)	9 (5.0)	
Tumor status, n (%)			0.175
With tumor	60 (33.3)	49 (27.1)	
Tumor free	107 (59.4)	120 (66.3)	
Radiation treatment, n (%)	1 (0.6)	3 (1.7)	0.338
Pharma treatment, n (%)	6 (3.3)	7 (3.9)	0.863
Ablation treatment, n (%)	6 (3.3)	7 (3.9)	0.836
Grade, n (%)			<0.001
1	16 (8.9)	37 (20.4)	
2	80 (44.4)	91 (50.3)	
3	76 (42.2)	45 (24.9)	
4	7 (3.9)	4 (2.2)	
AFP, n (%)			
> 200 ng/ml	119 (66.1)	67 (37.0)	<0.001
> 300 ng/ml	114 (63.3)	62 (34.3)	<0.001
> 400 ng/ml	114 (63.3)	61 (33.7)	<0.001
Platelet, mean \pm SD, 10 ³ /mm ³	219.5 \pm 8.7	217.3 \pm 7.7	0.845
Bilirubin, mean \pm SD, mg/dl	0.9 \pm 0.1	1.0 \pm 0.2	0.406
Creatinine, mean \pm SD, mg/dl	1.9 \pm 0.7	3.6 \pm 1.2	0.228
New tumor event after initial treatment, n (%)	50 (27.8)	43 (23.8)	0.298
Hepatic inflammation of adjacent tissue, n (%)			0.636
None	51 (28.3)	63 (34.8)	
Mild	48 (26.7)	49 (27.1)	
Severe	7 (3.9)	11 (6.1)	
Ishak stage, n (%)			0.178
None	25 (13.9)	47 (26.0)	
Portal fibrosis	14 (7.8)	17 (9.4)	
Fibrosis speta	15 (8.3)	13 (7.2)	
Nodular formation/cirrhosis	40 (22.2)	39 (21.5)	
Child-pugh classification, n (%)			0.726
A	118 (65.6)	115 (63.5)	
B and C	12 (6.7)	10 (5.5)	
Vascular invasion			0.858
None	116 (64.4)	105 (58.0)	
Macro and micro	54 (30.0)	51 (28.2)	
AJCC stage, n (%)			<0.001
I	21 (11.7)	0 (0)	
II	74 (41.1)	93 (51.4)	
III	42 (23.3)	40 (22.1)	
IV	53 (29.4)	35 (19.3)	
Risk factors, n (%)			0.174
None	91 (50.6)	80 (44.2)	
Alcohol assumption	48 (26.7)	57 (31.5)	
Viral hepatitis	62 (34.4)	44 (24.3)	
Estimated mean follow-up, months, mean \pm SD [†]	35.3 \pm 2.7	41.4 \pm 2.7	0.136
Deceased, n (%)	57 (31.5)	72 (40)	0.092
OS months, mean \pm SD	30.6 \pm 1.9	22.7 \pm 1.6	0.002

[†] Calculated by reverse Kaplan-Meier method.

BMI, body mass index; AFP, alpha fetoprotein; AJCC, American Joint Committee on Cancer; IQR, interquartile range; OS, overall survival.

Table 2. Univariate and Multivariate Logistic Regression Models for Identifying Parameters Associated With RAD51AP1.#

Variables	Univariate OR (95%CI)	p value	Multivariate OR (95%CI)	p value
Race				
Asian	Reference	-	Reference	-
White	0.55 (0.36-0.86)	0.008	1.97 (0.87-4.48)	0.106
Others	0.77 (0.3-2.01)	0.597	2.06 (0.46-9.28)	0.347
Grade				
I	Reference	-	Reference	-
II	2.12 (1.12-4.02)	0.021	4.15 (1.07-16.07)	0.04
III-IV	4.09 (2.1-7.96)	< 0.001	5.75 (1.42-23.23)	0.014
AJCC staging				
I	Reference	-	Reference	-
2	1.32 (0.78-2.24)	0.305	0.67 (0.29-1.52)	0.334
3-4	1.9 (1.13-3.22)	0.016	0.76 (0.3-1.89)	0.55
AFP > 200 ng/ml, yes vs. no	2.03 (1.34-3.1)	0.001	1.53 (0.73-3.21)	0.264
Ishak stage				
None	Reference	-	Reference	-
Portal fibrosis	1.55 (0.66-3.65)	0.318	1.45 (0.51-4.08)	0.486
Fibrosis speta	2.17 (0.89-5.27)	0.087	1.54 (0.49-4.83)	0.457
Nodular formation/cirrhosis	1.93 (1.0-3.71)	0.05	2.45 (0.95-5.33)	0.066
Family history of cancer, yes vs. no	0.49 (0.3-0.79)	0.003	0.58 (0.28-1.21)	0.148
Age, per increase 1 year	0.98 (0.97-1.0)	0.031	0.99 (0.97-1.02)	0.612

Variables including age, gender, BMI, Race, tumor status, treatment history, pathological grade, risk factor, AJCC staging, vascular invasion, child-pugh, AFP, Ishak stage, hepatic inflammation of adjacent tissue, new tumor event after initial treatment, family history of cancer, creatine, bilirubin, platelet counts were included in univariate analysis. Only variables with $p < 0.05$ in univariate model were presented and included in the multivariate analysis.

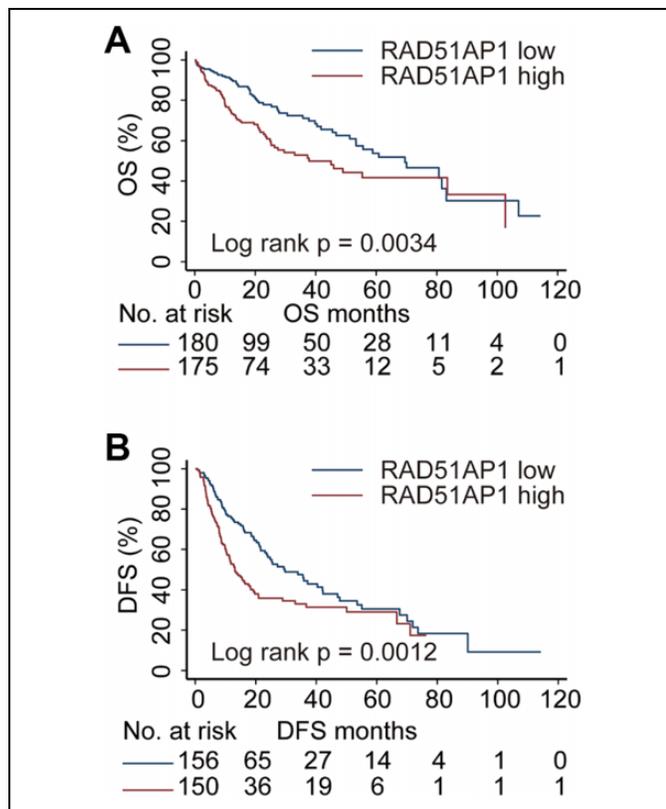


Figure 2. Associations of RAD51AP1 and OS (A) and DFS (B) in HCC patients in TCGA database.

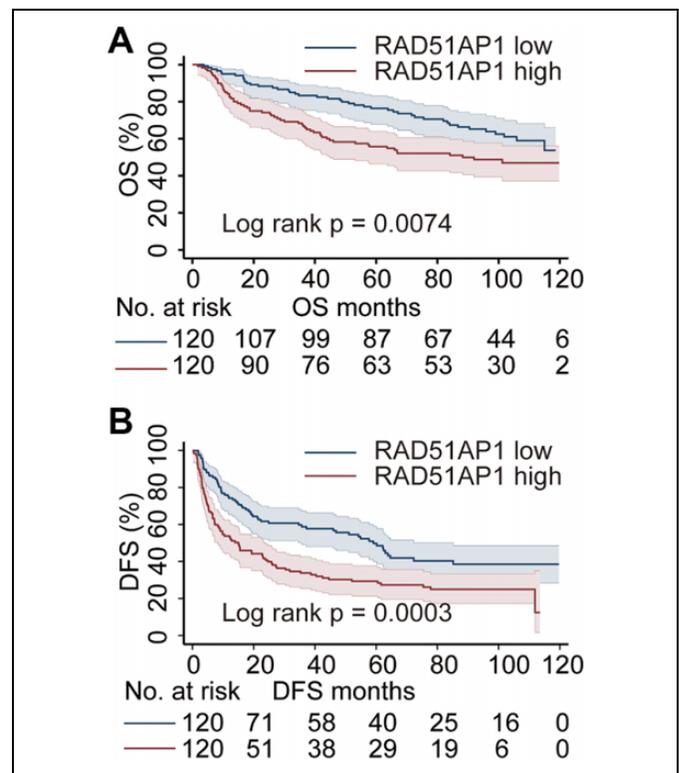


Figure 3. Associations of RAD51AP1 and OS (A) and DFS (B) in HCC patients in GSE36376 profile.

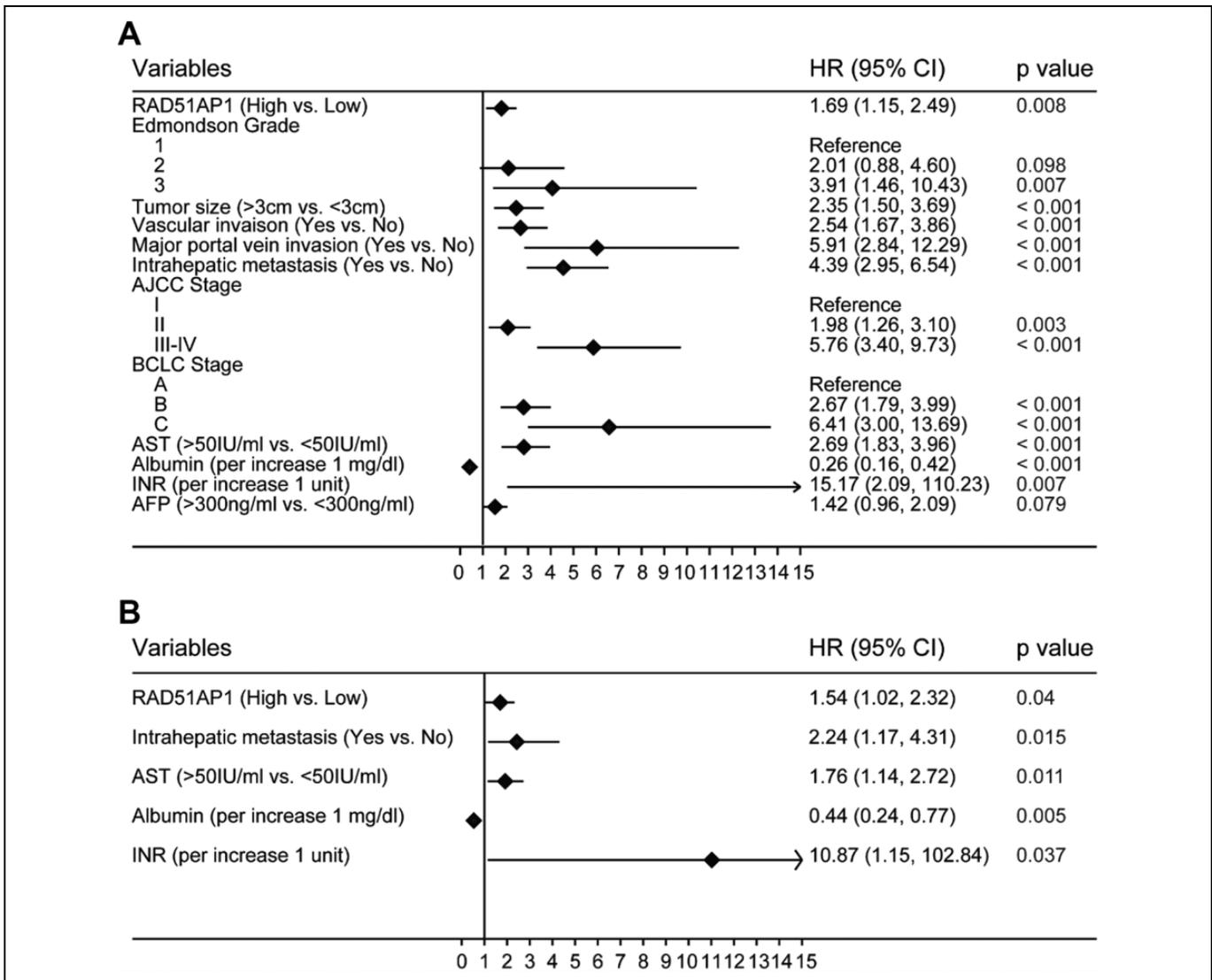


Figure 4. Univariate (A) and multivariate (B) Cox regression analysis of parameters associated with OS in HCC patients in GSE36376.

RAD51AP1 and HCC Survival

In the TCGA dataset, HCC patients were divided into high and low RAD51AP1 expression groups using the median expression level as the cutoff. As summarized in Table 1, the HCC patients in the high RAD51AP1 expression group were younger than those in the low RAD51AP1 expression group ($P = 0.03$, Table 1). The distributions of race, pathological grade and AJCC stage were significantly different between these 2 groups (all $P < 0.05$, Table 1). The patients in the high RAD51AP1 expression group had significantly higher AFP levels ($P < 0.001$, Table 1). On the other hand, univariate and multivariate logistic regression models indicated that pathological grade might contribute to RAD51AP1 expression elevation (compared to grade I, OR = 4.15 and $P = 0.04$ for grade II and OR = 5.75 and $P = 0.014$ for grade III-IV, Table 2).

As shown in Figure 2, the patients in the high RAD51AP1 expression group had significantly worse OS than those in the

low RAD51AP1 expression group (log-rank $P = 0.0034$, Figure 2A). Additionally, the patients in the high RAD51AP1 expression group had significantly shorter DFS than those in the low RAD51AP1 expression group (log rank $P = 0.0012$, Figure 2B). For validation, we evaluated 240 HCC patients in the GSE36376 dataset. The patients with RAD51AP1 upregulation in their tumor tissue had significantly poorer OS and DFS than those with RAD51AP1 downregulation (log-rank $P = 0.0074$ and 0.0003, respectively, Figure 3).

Cox Regression Analysis of Variables Associated With HCC Survival

For GSE36376, patients with RAD51AP1 upregulation were younger than those with low RAD51AP1 ($p = 0.011$, supplementary Table 1), which was consistent with that in TCGA dataset. Patients with high RAD51AP1 had advanced

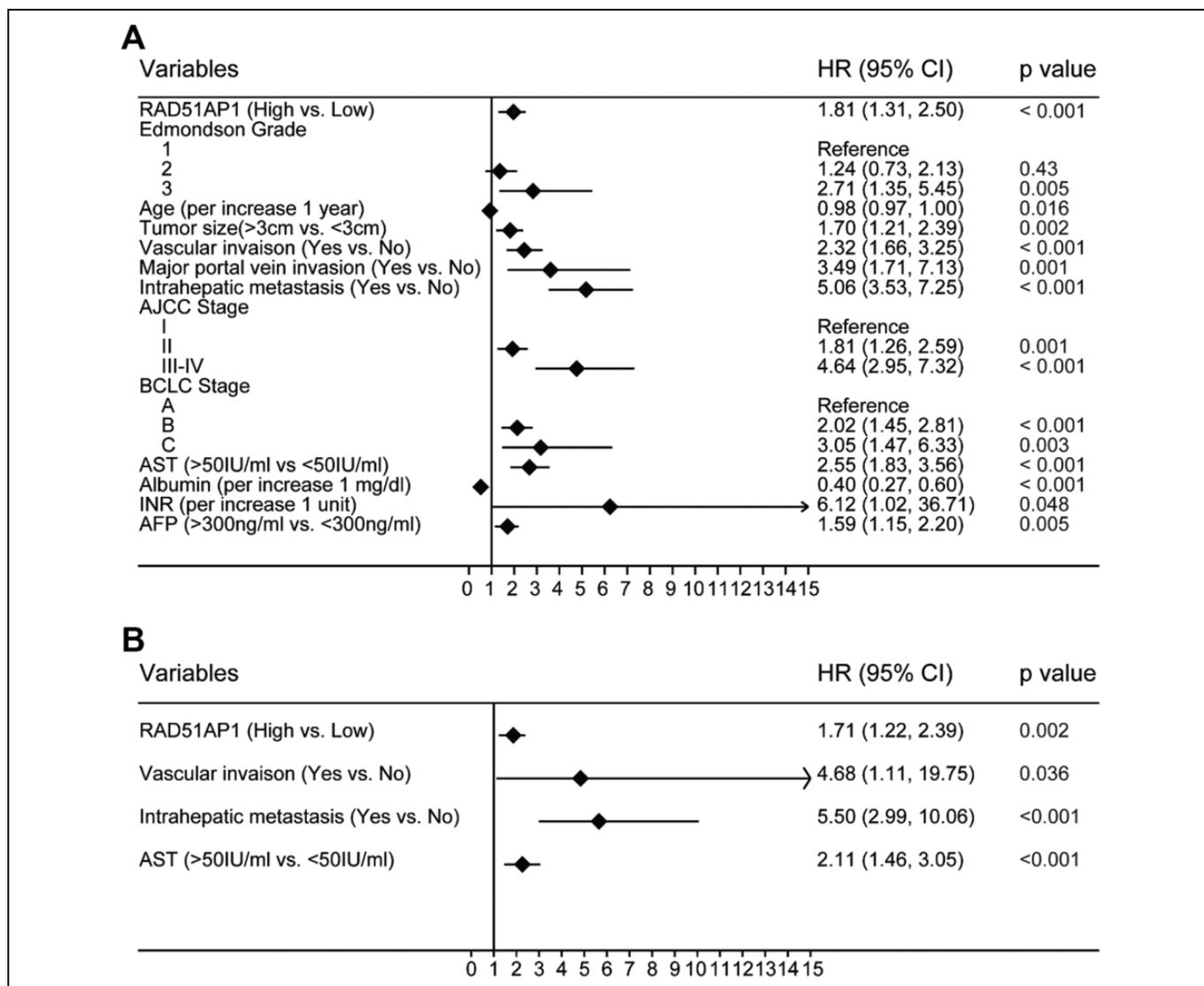


Figure 5. Univariate (A) and multivariate (B) Cox regression analysis of parameters associated with DFS in HCC patients in GSE36376.

Edmondson grade, AJCC staging, BCLC staging, more frequent of vascular invasion, major portal vein invasion and intrahepatic metastasis, and AFP elevation (all $p < 0.05$, supplementary Table 1). Univariate Cox analysis revealed that RAD51AP1, Edmondson grade, tumor size, vascular invasion status, major portal vein invasion status, intrahepatic metastasis status, AJCC stage, BCLC stage, aspartate aminotransferase (AST) elevation status, albumin, international normalized ratio (INR) and AFP were all potentially factors significantly associated with OS in HCC patients (all $P < 0.10$, Figure 4A). Multivariate analysis indicated that RAD51AP1 was a risk factor for OS in HCC patients (HR = 1.54, 95% CI = 1.02-2.32, $P = 0.04$, Figure 4B) after adjusting for the intrahepatic metastasis status, AST elevation status, INR and albumin level.

Additionally, we utilized a Cox regression model to identify the factors associated with DFS in HCC patients. As shown in Figure 5, RAD51AP1, Edmondson grade, age, tumor size, vascular invasion status, major portal vein invasion status,

intrahepatic metastasis status, AJCC stage, BCLC stage, AST elevation status, albumin, INR and AFP were all potentially factors significantly related to DFS in HCC patients (all $P < 0.10$, Figure 5A). Multivariate Cox analysis demonstrated that RAD51AP1 (HR = 1.71, 95% CI = 1.22-2.39, $P = 0.002$, Figure 5B) and the vascular invasion, intrahepatic metastasis and AST elevation statuses (all $P < 0.05$, Figure 5B) were all risk factors significantly associated with DFS in HCC patients.

Associations Between RAD51AP1 and Clinicopathological Features in HCC

In GSE36376, RAD51AP1 mRNA expression was significantly upregulated in tumor tissues compared to nontumor tissues ($P < 0.0001$, Figure 6A). Moreover, RAD51AP1 mRNA expression increased gradually with increasing tumor stage, regardless of whether AJCC staging (AJCC stage II vs. I, $P < 0.05$; AJCC stage III-IV vs. I, $P < 0.0001$; and AJCC stage

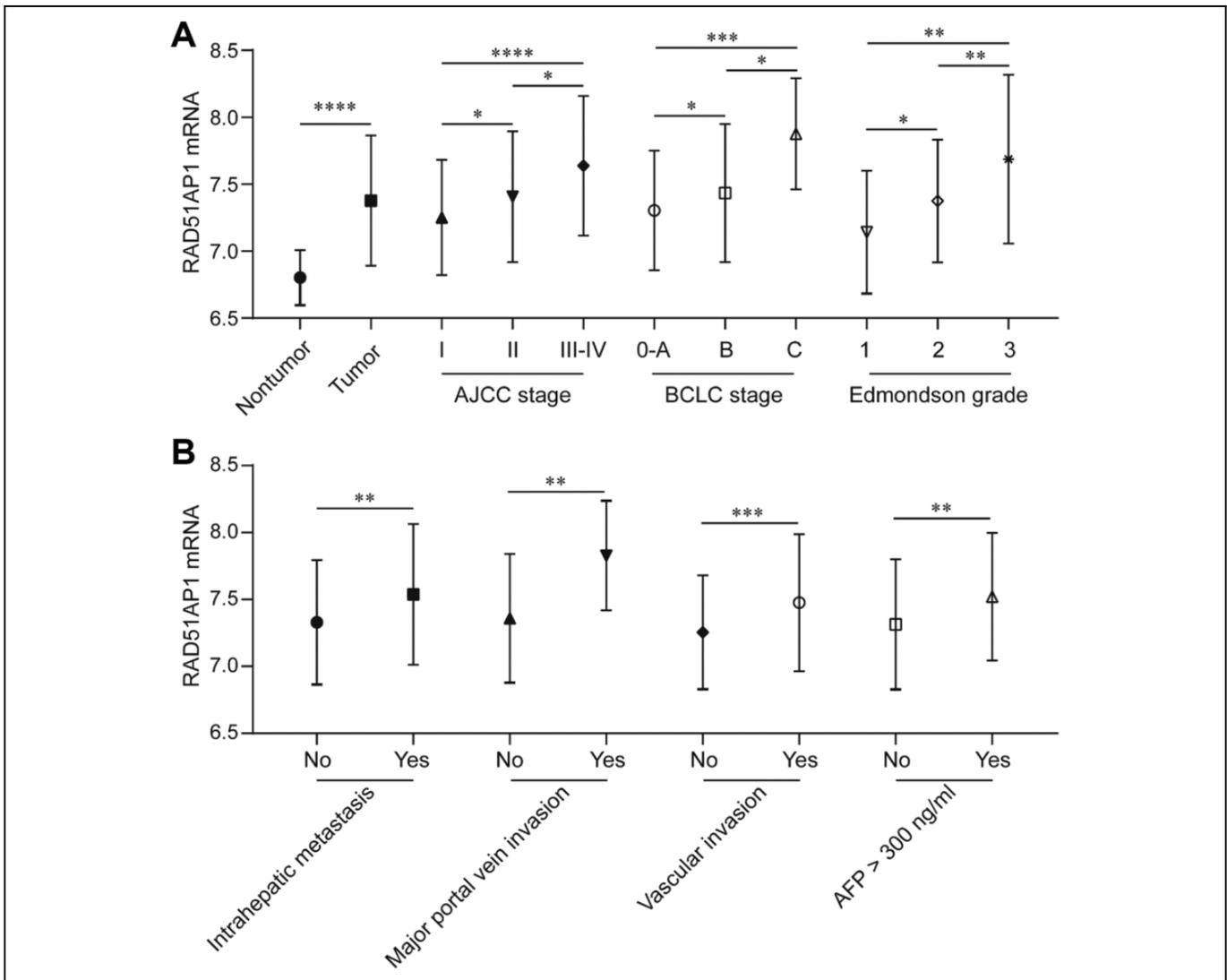


Figure 6. Validation of RAD51AP1 mRNA levels in tumor and nontumor tissues in GSE36376, RAD51AP1 expression comparison by AJCC stage, BCLC stage, Edmondson grade (A); intrahepatic metastasis status, major portal vein invasion status, vascular invasion status and AFP elevation (B).

III-IV vs. II, $P < 0.05$, Figure 6A), BCLC staging (BCLC stage B vs. 0-A, $P < 0.05$; BCLC stage C vs. B, $P < 0.05$; and BCLC stage C vs. 0-A, $P < 0.001$, Figure 6A) or Edmondson grading (grade 2 vs. grade 1, $P < 0.05$; grade 3 vs. grade 1, $P < 0.01$; and grade 3 vs. grade 2, $P < 0.01$, Figure 6A) was used. In addition, RAD51AP1 was also overexpressed in HCC patients with intrahepatic metastasis ($P < 0.01$, Figure 6B), major portal vein invasion ($P < 0.01$, Figure 6B), vascular invasion ($P < 0.001$, Figure 6B) and/or an AFP level > 300 ng/ml ($P < 0.01$, Figure 6B).

Discussion

Consistent with previous reports,^{19,20} our results revealed that RAD51AP1 was overexpressed in tumor tissues compared to adjacent tissues in HCC patients. Notably, upregulation of

RAD51AP1 expression was associated with advanced tumor stages, an advanced pathological grade, intrahepatic metastasis, vascular invasion and AFP level elevation. On the other hand, an advanced pathological grade might account for RAD51AP1 overexpression in tumor tissues. Moreover, high expression of RAD51AP1 was correlated with relatively poor OS and DFS in HCC patients. Previously, Xie et al reported that 18 HCC patients with RAD51AP1 alterations showed worse OS than 352 patients without RAD51AP1 alterations based on a cBioportal online analysis.²⁰ In addition, our results revealed that RAD51AP1 was highly expressed in liver diseases with advanced fibrosis and inflammatory stages. Intriguingly, RAD51AP1 expression is increased in hepatitis C virus (HCV)-infected liver tissues and HCV-transgenic mice. RAD51AP1 silencing results in a decrease in viral propagation. Mechanistically, RAD51AP1 is involved in the assembly steps

of the HCV life cycle by protecting viral RNA.³⁰ Considering previous studies and our results, we cautiously hypothesized that RAD51AP1 promotes liver disease progression, tumorigenesis and cancer aggressiveness in HCC patients.

Cancer cells and precancerous lesions often contain abnormally regulated DNA repair pathways,^{31,32} and upregulation of DNA repair gene expression is considered to be related to metastasis.^{16,33} Emerging evidence has shown that RAD51AP1 contributes to human cancer progression.³⁴ RAD51AP1 suppresses cell cycle arrest and apoptosis, leading to cellular resistance to DNA-damaging cancer therapies, and may increase DNA instability.³⁵ RAD51AP1 overexpression has also been found in intrahepatic cholangiocarcinoma, and silencing RAD51AP1 results in growth suppression in cholangiocarcinoma cells.³⁶ Amplification of RAD51AP1 is associated with cell immortality in ovarian cancer cells, while RAD51AP1 overexpression may account for the relatively short survival times of ovarian cancer patients³⁷ and breast cancer patients.¹³ In addition, RAD51AP1 is associated with brain metastasis and can discriminate primary and metastatic melanoma tumors, indicating that RAD51AP1 may serve as a potential driver of melanoma metastasis.¹⁷ Recent reviews have also deduced that high RAD51AP1 expression may be correlated with a selective growth advantage, enabling early neoplastic cells and metastatic cells to overcome replication stress.^{32,34,38} This might explain why DFS in HCC patients seemed to be affected by RAD51AP1 to a greater degree than OS in our analysis.

DNA replication stress induced by many cancer therapeutic agents has emerged as a significant source of genomic instability during the early stages of carcinogenesis.³⁸ Experimental studies have demonstrated that loss of RAD51AP1 leads to slow progression of the DNA replication fork and increased DNA replication stress.^{39,40} During the early steps of neoplasia, the RAD51AP1 level is elevated as a consequence of the high levels of replication stress.³⁹ Overcoming this obstacle to cell survival by elevating RAD51AP1 expression can cause further genomic changes to occur, thereby promoting the progression of precancerous lesions into cancer.³⁹ That is, RAD51AP1 plays an important role in protecting against DNA replication stress.^{34,39} Deletion of RAD51AP1 in human cells increases their sensitivity to the cytotoxic effects of DNA cross-linkers. Knockdown of RAD51AP1 expression also results in increased levels of genomic instability and reduced homology-directed repair.^{8,9,39} Hence, inhibiting RAD51AP1 during the initial stages of neoplasia may provide a useful approach for targeted therapy.³⁹

This study has some limitations. The primary limitation is that no experiments were conducted to address the effects of RAD51AP1 on hepatoma cellular functions. Second, this is a study based on public datasets, and no follow-up data for HCC patients in our medical center were available. Third, the follow-up period of patients in TCGA dataset is relatively short. Future studies with long follow-up are essentially needed. Nonetheless, our results indicated that high RAD51AP1 contributed to an advanced tumor stage, intrahepatic metastasis, vascular invasion and AFP level elevation, and RAD51AP1 upregulation was significantly associated with OS and DFS in HCC

patients. Unfortunately, current research on the relationship between RAD51AP1 and HCC development is mostly based on public databases,²⁰ and no well-designed, prospective, large-sample studies are available. There are no sufficient data indicating whether RAD51AP1 expression should guide treatment in the current stage. Future research should focus on the mechanisms involving RAD51AP1 in promoting cancer progression and on determining the therapeutic intervention value of targeting RAD51AP1 in HCC patients.

Declaration of Conflicting Interests

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Supplemental Material

Supplemental material for this article is available online.

References

1. Roos WP, Thomas AD, Kaina B. DNA damage and the balance between survival and death in cancer biology. *Nat Rev Cancer*. 2016;16(1):20-33.
2. Lord CJ, Ashworth A. The DNA damage response and cancer therapy. *Nature*. 2012;481(7381):287-294.
3. Hoeijmakers JH. DNA damage, aging, and cancer. *N Engl J Med*. 2009; 361(15):1475-1485.
4. Turgeon MO, Perry NJS, Poulgiannis G. DNA damage, repair, and cancer metabolism. *Front Oncol*. 2018;8:15.
5. O'Connor MJ. Targeting the DNA damage response in cancer. *Mol Cell*. 2015;60(4):547-560.
6. Carrassa L, Damia G. DNA damage response inhibitors: mechanisms and potential applications in cancer therapy. *Cancer Treat Rev*. 2017;60:139-151.
7. Dunlop MH, Dray E, Zhao W, et al. Mechanistic insights into RAD51-associated protein 1 (RAD51AP1) action in homologous DNA repair. *J Biol Chem*. 2012;287(15):12343-12347.
8. Modesti M, Budzowska M, Baldeyron C, Demmers JA, Ghirlando R, Kanaar R. RAD51AP1 is a structure-specific DNA binding protein that stimulates joint molecule formation during RAD51-mediated homologous recombination. *Mol Cell*. 2007; 28(3):468-481.
9. Wiese C, Dray E, Groesser T, et al. Promotion of homologous recombination and genomic stability by RAD51AP1 via RAD51 recombinase enhancement. *Mol Cell*. 2007;28(3):482-490.
10. Wu Y, Wang H, Qiao L, Jin X, Dong H, Wang Y. Silencing of RAD51AP1 suppresses epithelial-mesenchymal transition and

- metastasis in non-small cell lung cancer. *Thorac Cancer*. 2019; 10(9):1748-1763.
11. Chudasama D, Bo V, Hall M, et al. Identification of cancer biomarkers of prognostic value using specific gene regulatory networks (GRN): a novel role of RAD51AP1 for ovarian and lung cancers. *Carcinogenesis*. 2018;39(3):407-417.
 12. Mathe A, Wong-Brown M, Morten B, et al. Novel genes associated with lymph node metastasis in triple negative breast cancer. *Sci Rep*. 2015;5:15832.
 13. Pathania R, Ramachandran S, Mariappan G, et al. Combined inhibition of DNMT and HDAC blocks the tumorigenicity of cancer stem-like cells and attenuates mammary tumor growth. *Cancer Res*. 2016;76(11):3224-3235.
 14. Martinez I, Wang J, Hobson KF, Ferris RL, Khan SA. Identification of differentially expressed genes in HPV-positive and HPV-negative oropharyngeal squamous cell carcinomas. *Eur J Cancer*. 2007;43(2):415-432.
 15. Wang Q, Tan Y, Fang C, et al. Single-cell RNA-seq reveals RAD51AP1 as a potent mediator of EGFRvIII in human glioblastomas. *Aging (Albany NY)*. 2019;11(18):7707-7722.
 16. Kauffmann A, Rosselli F, Lazar V, et al. High expression of DNA repair pathways is associated with metastasis in melanoma patients. *Oncogene*. 2008;27(5):565-573.
 17. Redmer T, Walz I, Klinger B, et al. The role of the cancer stem cell marker CD271 in DNA damage response and drug resistance of melanoma cells. *Oncogenesis*. 2017;6(1):e291.
 18. Henson SE, Tsai SC, Malone CS, et al. Pir51, a Rad51-interacting protein with high expression in aggressive lymphoma, controls mitomycin C sensitivity and prevents chromosomal breaks. *Mutat Res*. 2006;601(1-2):113-124.
 19. Song H, Xia SL, Liao C, et al. Genes encoding Pir51, Beclin 1, RbAp48 and aldolase b are up or down-regulated in human primary hepatocellular carcinoma. *World J Gastroenterol*. 2004; 10(4):509-513.
 20. Xie S, Jiang X, Zhang J, et al. Identification of significant gene and pathways involved in HBV-related hepatocellular carcinoma by bioinformatics analysis. *PeerJ*. 2019;7:e7408.
 21. Wang M, Gong Q, Zhang J, et al. Characterization of gene expression profiles in HBV-related liver fibrosis patients and identification of ITGBL1 as a key regulator of fibrogenesis. *Sci Rep*. 2017; 7:43446.
 22. Yildiz G, Arslan-Ergul A, Bagislar S, et al. Genome-wide transcriptional reorganization associated with senescence-to-immortality switch during human hepatocellular carcinogenesis. *PLoS One*. 2013;8(5):e64016.
 23. Wang HW, Hsieh TH, Huang SY, et al. Forfeited hepatogenesis program and increased embryonic stem cell traits in young hepatocellular carcinoma (HCC) comparing to elderly HCC. *BMC Genomics*. 2013;14(1):736.
 24. Melis M, Diaz G, Kleiner DE, et al. Viral expression and molecular profiling in liver tissue versus microdissected hepatocytes in hepatitis B virus-associated hepatocellular carcinoma. *J Transl Med*. 2014;12(1):230.
 25. Shi M, Chen MS, Sekar K, Tan CK, Ooi LL, Hui KM. A blood-based three-gene signature for the non-invasive detection of early human hepatocellular carcinoma. *Eur J Cancer*. 2014;50(5): 928-936.
 26. Lim HY, Sohn I, Deng S, et al. Prediction of disease-free survival in hepatocellular carcinoma by gene expression profiling. *Ann Surg Oncol*. 2013; 20(12):3747-3753.
 27. Cerami E, Gao J, Dogrusoz U, et al. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. *Cancer Discov*. 2012;2(5):401-404.
 28. Gao J, Aksoy BA, Dogrusoz U, et al. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal*. 2013;6(269):p11.
 29. Yang Z, Lu Y, Xu Q, Tang B, Park CK, Chen X. HULC and H19 played different roles in overall and disease-free survival from hepatocellular carcinoma after curative hepatectomy: a preliminary analysis from gene expression omnibus. *Dis Markers*. 2015; 2015:191029.
 30. Nguyen TTT, Park EM, Lim YS, Hwang SB. Nonstructural protein 5A impairs DNA damage repair: implications for hepatitis C virus-mediated hepatocarcinogenesis. *J Virol*. 2018;92(11).
 31. Bartkova J, Horejsi Z, Koed K, et al. DNA damage response as a candidate anti-cancer barrier in early human tumorigenesis. *Nature*. 2005;434(7035):864-870.
 32. Gorgoulis VG, Vassiliou LV, Karakaidos P, et al. Activation of the DNA damage checkpoint and genomic instability in human precancerous lesions. *Nature*. 2005;434(7035):907-913.
 33. Sarasin A, Kauffmann A. Overexpression of DNA repair genes is associated with metastasis: a new hypothesis. *Mutat Res*. 2008; 659(1-2):49-55.
 34. Pires E, Sung P, Wiese C. Role of RAD51AP1 in homologous recombination DNA repair and carcinogenesis. *DNA Repair (Amst)*. 2017;59:76-81.
 35. Klein HL. The consequences of Rad51 overexpression for normal and tumor cells. *DNA Repair (Amst)*. 2008;7(5):686-693.
 36. Obama K, Satoh S, Hamamoto R, Sakai Y, Nakamura Y, Furu-kawa Y. Enhanced expression of RAD51 associating protein-1 is involved in the growth of intrahepatic cholangiocarcinoma cells. *Clin Cancer Res*. 2008;14(5):1333-1339.
 37. Sankaranarayanan P, Schomay TE, Aiello KA, Alter O. Tensor GSVD of patient- and platform-matched tumor and normal DNA copy-number profiles uncovers chromosome arm-wide patterns of tumor-exclusive platform-consistent alterations encoding for cell transformation and predicting ovarian cancer survival. *PLoS One*. 2015;10(4):e0121396.
 38. Halazonetis TD, Gorgoulis VG, Bartek J. An oncogene-induced DNA damage model for cancer development. *Science*. 2008; 319(5868):1352-1355.
 39. Parpys AC, Kratz K, Speed MC, Leung SG, Schild D, Wiese C. RAD51AP1-deficiency in vertebrate cells impairs DNA replication. *DNA Repair (Amst)*. 2014; 24:87-97.
 40. Parpys AC, Zhao W, Sharma N, et al. NUCKS1 is a novel RAD51AP1 paralog important for homologous recombination and genome stability. *Nucleic Acids Res*. 2015;43(20):9817-9834.