



Complete Genome Sequence of Mycobacteriophage Fulbright

Hari Kotturi,^a Umar Sahi,^a Cameron Kedy,^a DAhmed K. Ali^b

^aDepartment of Biology, University of Central Oklahoma, Edmond, Oklahoma, USA ^bDepartment of Biology, College of Science, Mustansiriyah University, Baghdad, Iraq

ABSTRACT Mycobacteriophage Fulbright was isolated from soil in central Oklahoma using *Mycobacterium smegmatis* mc²115. The genome of phage Fulbright is 42,396 bp long and contains 70 open reading frames (ORFs), with 33 having predicted functions and 37 having hypothetical proteins. It belongs to cluster N and shares 99% nucleotide identity with mycobacteriophage Phloss.

ycobacteriophages are bacteriophages that are capable of infecting a mycobacterial host (1). The genus *Mycobacterium* is composed of acid-fast and obligatory aerobic bacteria. Mycobacteria are ubiquitous microorganisms that were isolated from various soil types, such as those found on ranches, landfills, and boreal coniferous forests (2, 3). Mycobacteriophage Fulbright was isolated from a soil sample collected from the campus of the University of Central Oklahoma (UCO) (global positioning system [GPS] coordinates, 35.658889N, 97.474444W). The phage was isolated, purified, and propagated using the host, *M. smegmatis* mc²155, following the protocols described in the Science Education Alliance-Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES) manual (4). Briefly, the collected soil sample was enriched with M. smegmatis mc²155, incubated at 37°C for 24 h, filtered, and plated with the host bacteria. Three plague purifications were done to purify the phage for imaging and DNA extraction. Transmission electron microscopy images of the phage particle were taken by mounting sample on a carbon-stabilized, Formvar-coated copper grid stained with 1% uranyl acetate solution. Photographs were taken using the Hitachi H-7600 machine. Fulbright has a Siphoviridae morphology with an isometric head and a long, flexible noncontractile tail (Fig. 1). Phage genomic DNA was extracted from lysate following the SEA-PHAGES recommended protocols (4) using a Promega Wizard DNA cleanup kit. An Ultra II FS kit with dual-indexed barcoding (New England Biolabs) was used for generating the genomic libraries. The pooled libraries were run on an Illumina MiSeq system to yield single-end 150-base reads. Newbler v.2.9 and Consed v.29 were used with default parameters for assembling the genome and assessing the quality of the assembly (5). The approximate sequencing coverage of the phage genome is $2,501 \times$ with 749,000 reads used for the assembly. The assembled phage genome was annotated by students in the bioinformatics course at UCO in the spring of 2019 using the SEA-PHAGES-recommended parameters with DNAMaster v.5.0.2 (http://cobamide2.bio.pitt.edu/computer .htm), GeneMark v.3.25 (6), NCBI BLAST v.2.9.0 (7), Glimmer v.3.02 (8), HHpred v.3.2.0 (9), ARAGORN v.1.2.38 (10), and Phamerator (11).

The genome of phage Fulbright is 42,396 bp long with a 63% G+C content. It has a 3' sticky end with a 13-bp overhang which was determined as previously described (5). Genome analysis indicated that Fulbright has 70 open reading frames (ORFs), with 33 having predicted functions and 37 having hypothetical proteins. All predicted ORFs are transcribed in the forward direction, except ORFs 24, 25, 32 to 36, and 67. The predicted structural and assembly ORFs are organized on the left side, with nonstructural predicted ORFs on the genome's right side. These two regions are separated by lysis and immunity cassettes. Some of the structural predicted ORFs include ORF 6 (major

Citation Kotturi H, Sahi U, Kedy C, Ali AK. 2021. Complete genome sequence of mycobacteriophage Fulbright. Microbiol Resour Announc 10:e00123-21. https://doi.org/ 10.1128/MRA.00123-21.

Editor Simon Roux, DOE Joint Genome Institute

Copyright © 2021 Kotturi et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Hari Kotturi, hkotturi@uco.edu.

Received 1 February 2021 Accepted 18 February 2021 Published 18 March 2021



B1 #2 Print Mag: 130000x @ 51 mm 13:26 08/23/17

20 nm HV=80kV Direct Mag: 120000x X:Y: Core Facility for Imaging

FIG 1 Transmission electron microscopy (TEM) of Fulbright on a Formvar-coated copper grid stained with uranyl acetate, imaged using a Hitachi H-7600 machine.

capsid protein), ORF 13 (major tail protein), ORF 16 (tape measure protein), and ORFs 17 to 21 (minor tail proteins). The tail assembly chaperones (ORFs 14 and 15) have a -2 frameshift. Like other cluster N phages, Fulbright contains a predicted tyrosine integrase (ORF 35) and immunity repressor (ORF 36) transcribed in the reverse direction. A cluster includes phages with sequence similarity over 50% of their genomes (12). The lysis system is encoded by predicted ORF 27 (lysin A) and ORF 28 (holin) and lacks the lysin B gene. The genome lacks any tRNA genes, which is in line with other phages in this cluster. The phage genome shares 99% nucleotide identity with phage Phloss in the same cluster as that determined using BLASTn.

Data availability. The complete genome sequence of phage Fulbright is available in GenBank under the accession number MK977708 with the NCBI SRA accession number SRX10061435.

ACKNOWLEDGMENTS

The University of Central Oklahoma is part of the Science Education Alliance-Phage Hunters Advancing Genomics and Evolutionary Science (SEA-PHAGES) program, supported by the Howard Hughes Medical Institute (HHMI).

We thank Eric M. Kelough for his contributions to this annotation. We thank Graham Hatfull, Debbie Jacobs-Sera, Dan Russell, Rebecca Garlena, Steve Cresawn, and Welkin Pope for guidance during the genome preparation and annotation. We thank Ben Fowler at the imaging core facility at Oklahoma Medical Research Foundation for his electron microscopy help.

Funding for this project was acquired through the University of Central Oklahoma, Office of Research and Sponsored Programs–RCSA Grant Program, and by the National Institute of General Medical Sciences of the National Institutes of Health under award number P20GM103447. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

REFERENCES

- Jacobs-Sera D, Marinelli LJ, Bowman C, Broussard GW, Guerrero Bustamante C, Boyle MM, Petrova ZO, Dedrick RM, Pope WH, Modlin RL, Hendrix RW, Hatfull GF, Science Education Alliance Phage Hunters Advancing Genomics and Evolutionary Science Sea-Phages Program. 2012. On the nature of mycobacteriophage diversity and host preference. Virology 434:187–201. https://doi.org/10.1016/j.virol.2012.09.026.
- Bardarov S, Kriakov J, Carriere C, Yu S, Vaamonde C, McAdam RA, Bloom BR, Hatfull GF, Jacobs WR Jr. 1997. Conditionally replicating mycobacteriophages: a system for transposon delivery to *Mycobacterium tuberculosis*. Proc Natl Acad Sci U S A 94:10961–10966. https://doi.org/10.1073/pnas.94.20.10961.
- Norby B, Fosgate GT, Manning EJ, Collins MT, Roussel AJ. 2007. Environmental mycobacteria in soil and water on beef ranches: association between presence of cultivable mycobacteria and soil and water physicochemical characteristics. Vet Microbiol 124:153–159. https://doi.org/10 .1016/j.vetmic.2007.04.015.
- 4. Jordan TC, Burnett SH, Carson S, Caruso SM, Clase K, DeJong RJ, Dennehy JJ, Denver DR, Dunbar D, Elgin SC, Findley AM, Gissendanner CR, Golebiewska UP, Guild N, Hartzog GA, Grillo WH, Hollowell GP, Hughes LE, Johnson A, King RA, Lewis LO, Li W, Rosenzweig F, Rubin MR, Saha MS, Sandoz J, Shaffer CD, Taylor B, Temple L, Vazquez E, Ware VC, Barker LP, Bradley KW, Jacobs-Sera D, Pope WH, Russell DA, Cresawn SG, Lopatto D, Bailey CP, Hatfull GF. 2014. A broadly implementable research course in phage discovery and genomics for first-year undergraduate students. mBio 5:e01051-13. https://doi.org/10.1128/mBio.01051-13.

- Russell DA. 2018. Sequencing, assembling, and finishing complete bacteriophage genomes. Methods Mol Biol 1681:109–125. https://doi.org/10 .1007/978-1-4939-7343-9_9.
- Besemer J, Borodovsky M. 2005. GeneMark: Web software for gene finding in prokaryotes, eukaryotes and viruses. Nucleic Acids Res 33:W451–W454. https://doi.org/10.1093/nar/gki487.
- Zhang Z, Schwartz S, Wagner L, Miller W. 2000. A greedy algorithm for aligning DNA sequences. J Comput Biol 7:203–214. https://doi.org/10 .1089/10665270050081478.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679. https://doi.org/10.1093/bioinformatics/btm009.
- Söding J, Biegert A, Lupas AN. 2005. The HHpred interactive server for protein homology detection and structure prediction. Nucleic Acids Res 33:W244–W248. https://doi.org/10.1093/nar/gki408.
- Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. https://doi.org/10.1093/nar/gkh152.
- Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. BMC Bioinformatics 12:395. https://doi.org/10.1186/1471-2105 -12-395.
- Hatfull G. 2014. Molecular genetics of mycobacteriophages. Microbiol Spectr 2:1–36. https://doi.org/10.1128/microbiolspec.MGM2-0032-2013.