

Therapeutic Effect of *Hedera helix* Alcoholic Extract Against Cutaneous Leishmaniasis Caused by *Leishmania major* in Balb/c Mice

Hossein Hooshyar^{1,2,*}; Safarali Talari¹; Fatemeh Feyzi¹

¹Department of Parasitology, School of Medicine, Kashan University of Medical Sciences, Kashan, IR Iran

²Anatomical Science Research Center, Kashan University of Medical Sciences, Kashan, IR Iran

*Corresponding author: Hossein Hooshyar, Department of Parasitology, School of Medicine, Kashan University of Medical Sciences, Kashan, IR Iran. Tel: +98-3615550021, Fax: +98-3615551112, E-mail: hooshyar4@yahoo.com

Received: December 3, 2012; Revised: January 23, 2013; Accepted: February 19, 2013

Background: Cutaneous leishmaniasis (CL) is common and endemic in many areas of Iran, caused by species of a protozoan parasite belonging to the genus *Leishmania*. There is not any effective vaccine against leishmaniasis; so, therapy is important for prevention and separation of disease. Herbal extract for treatment of CL is cost-effective, applicable topically to lesions, and can avoid the development of drug resistance.

Objectives: The aim of this study was to evaluate the *in vivo* activity of an alcoholic extract of *Hedera helix* (a native Iranian plant) on the experimental ulcer of zoonotic CL in Balb/c mice.

Materials and Methods: At least 5×10^6 promastigotes of *Leishmania major* (MHOM/64/IR/ER75) were inoculated subcutaneously in the tail base of Balb/c mice. Fifty six infected mice were distributed in four groups, two groups (16 mice for 20% alcoholic extract of *H. helix* and 13 for 70% extract) were used as experimental groups, one (15 mice) as placebo control (Control A), and one (12 mice) as negative control. Treatment effects of two concentrations were determined by comparison of placebo and nontreated groups via measuring the size of skin lesions and the number of parasitologically positive and negative mice after the therapy period.

Results: This study showed that the main lesion size did not decrease significantly, or the small lesions did not completely disappear after treatment by *H. helix* alcoholic extract. Amastigotes counts (mean \pm SD) of the skin lesions decreased in control A and 20% concentration groups, but in negative control and 70% concentration groups the number of parasites did not reduce.

Conclusions: The present study did not support the *in vivo* antileishmanial effect of *H. helix* extract. We recommend further studies using major components of *H. helix*, especially hederasaponin (saponin K10), to investigate the antileishmanial effect of this plant on *L. major*.

Keywords: Leishmania; Therapeutics; Plants; *Hedera*

1. Background

Leishmaniasis is endemic in 88 countries and about 350 million people are at risk. The prevalence of the disease is about 12 million worldwide (1). Clinical features of the disease include cutaneous leishmaniasis (CL), mucocutaneous leishmaniasis (MCL), and visceral leishmaniasis (VL) (2, 3). CL is a group of diseases caused by several species of a protozoan parasite belonging to the genus *Leishmania*, transmitted by sand fly insect vector. The disease usually causes skin lesions on the exposed region of the body and leaves permanent scars.

Annual incidence of CL is 1 to 1.5 million cases, of which 90% occurs in only seven countries: Afghanistan, Algeria, Brazil, Iran, Peru, Saudi Arabia and Syria (4). CL is a major health problem in many areas of Iran. This disease includes anthroponotic CL (ACL) caused by *Leishmania tropica* and zoonotic CL (ZCL) caused by *L. major* that are endemic in many different parts of Iran (3, 5). *L. major* is the etiological agent for ZCL, endemic in some regions,

especially rural areas of 17 out of 31 provinces of Iran (5, 6). It is estimated that about 80% of the cases reported in this country are of ZCL form (6). Systemic treatment of ZCL is intralésional or systemic administration of antimony compounds (sodium stibogluconate and meglumine antimonies) (4). These compounds are parenteral and associated with significant side effects (7). Other disadvantages of these drugs are requirement for intramuscular or intralésional injection every day for a long period, toxicity, and the recent resistance development in regions such as India (8).

During the last decades, many attempts have been made to obtain effective new compounds especially herbal extracts, for treatment of CL that would be economical, applicable topically to the lesions, and could avoid resistance development (9). Plants contain a wide variety of metabolites such as tannins, terpenoids, alkaloids and flavonoids, found to have antimicrobial properties

Implication for health policy/practice/research/medical education:

Cutaneous leishmaniasis is a common infection in many parts of Iran. Antileishmanial plant drugs could be used for treatment and control of the infection.

Copyright © 2014, Ahvaz Jundishapur University of Medical Sciences; Published by Kowsar Corp. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

(10). Natural extracts of different plants such as Thyme, Yarrow, *Euphorbia* spp., *Gossypium herbacium*, *Berberis vulgaris*, *Alkanna tinctoria* and *Peganum harmala* have been directly used on skin lesions as well as on the parasite in NNN medium (11-13).

Hedera helix is a species of ivy, native to most of Eurasia, including Britain, south and east Mediterranean areas and Iran. It is an evergreen climbing plant, growing up to 20 - 30 m high where suitable surfaces (trees, cliffs, walls) are available, and also growing as ground cover where there are no vertical surfaces. Leaves of this plant are used in traditional medicine as bronchospasmodic, secretolytic and anti-inflammatory, also as an anthelmintic, and as an agent to reduce fever and cause diaphoresis. Some studies showed the effectiveness of leaves extract of this plant in fighting the bacterial, protozoal and fungal infections (14-16). Eguale et al. recently showed that hydro-alcoholic extracts of *H. helix* had antihelminthic activity against *Haemonchus contortus* *in vitro* and *in vivo* (17). *H. helix* is an endemic plant species growing in many regions. In Iran, this plant grows mainly in Kordestan and Shahrood cities as well as central, western, and eastern areas (18).

2. Objectives

The present study was an experimental research performed to determine and evaluate the effects of different concentrations of *H. helix* extract on Balb/c murine model infected by active promastigote of *L. major*.

3. Materials and Methods

This experimental and *in vivo* study was carried out on 90 Balb/c mice weighing approximately 30 g and 4 - 6 weeks of age (purchased from Pasteur Institute, Karaj, Iran). This study was performed according to the ethical standards and approved by ethics committee of deputy of research of Kashan University of Medical Sciences (no. 8304). The standard virulence strain of *L. major* (MHOM/64/IR/ER75) was obtained from the School of Public Health, Tehran University of Medical Sciences, cultured in RPMI-1640 (Gibco, Australia) supplemented with 15% inactivated fetal calf serum (FCS, Gibco, Australia). At least 5×10^6 promastigotes, harvested at the stationary phase of the culture, were inoculated subcutaneously in the tail base of the mice. After 40 days, nodules and ulcers appeared on 56 inoculated mice. Fifty six infected mice were distributed in four groups, two (16 mice for 20% extract and 13 for 70% extract) were used as experimental groups, one (15 mice) as placebo control (control A) and one (12 mice) as negative control. Leaves of *H. helix* were acquired from Kashan City, central Iran. Afterwards, the leaves were washed, dried, and extracted by Soxhlet apparatus and solved in ethanol 80% (percolation method) (19) in Barij Essence Pharmaceutical

Company.

Before using the plant extract for treatment, diameters of each lesion were measured using scale and vernier calipers. Impression smears were prepared from the lesions; the slide were fixed with absolute methanol, stained by Giemsa stain, and examined by light microscopy (1000x). Two concentrations (20% and 70%) of the *H. helix* extract were tested on the CL lesions of two groups of mice by rubbing on the local lesion two times a days, for 30 days. Control group A received ethanol 80% and negative control groups received no treatment. Treatment effects of two concentrations were determined through comparing the placebo and non-treated groups by measuring the size of the skin lesions and the number of parasitologically positive and negative mice after the therapy period. The results were analyzed as mean \pm SE (standard error of mean). Paired t-test, ANOVA and Fisher tests were used to compare the treated and untreated groups. Data were considered statistically significant at $P < 0.05$.

4. Results

This study showed that in both 20% and 70% concentrations of alcoholic extracts of *H. helix*, the mean lesion size increased at the end of the treatment. Sizes of lesions in both groups did not significantly increase in comparison with the control A groups (treated with ethanol 80%). Statistically significant increase in the size of ulcers was observed in negative control group compared with other three groups ($P < 0.03$). Results of this study showed that the main lesion size did not decrease significantly or small lesions did not completely disappear after treatment by *H. helix* alcoholic extracts (Table 1 and Figure 1). Amastigotes counts (mean \pm SD) of the skin lesions decreased in control A and 20% concentration groups, but the number of parasites did not reduce in negative control and 70% concentration groups. Direct smears of mice in the microscopic studies were positive after 1 month of treatment in all control and intervention groups.

Table 1. Therapeutic Effects of Different Concentrations of *H. helix* on the Average Diameters of CL Ulcers Induced by *L. major* in Balb/c Mice ^a

Ulcer Diameter	Before the Treatment, mm	After the Treatment, mm	Increase, %
Negative control	5.15 \pm 2.21	8.48 \pm 3.59	64.7
Control A	6.69 \pm 2.89	8.99 \pm 2.84	25.6
20% Extract	7.98 \pm 3.01	9.99 \pm 3.85	25.2
70% Extract	6.96 \pm 2.22	11.38 \pm 3.68	63.5

^a All data are presented in Mean \pm SD.

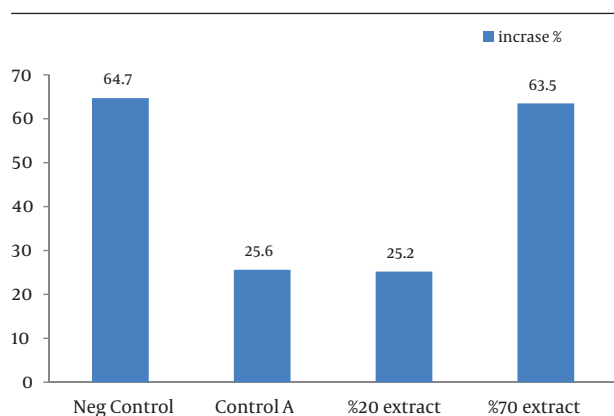


Figure 1. Diameter Increase Percentage of CL Ulcers of Mice Induced by *L. major*, Treated by Different Concentrations of *H. helix*

5. Discussion

The purpose of this study was to examine the effect of alcoholic *H. helix* extract on Balb/c murine model mice, infected by active promastigote of *L. major*. According to the results of this study, two concentrations of *H. helix* extract (20% and 70%) did not have any significant positive correlation with decrease of *L. major* ulcer diameter in Balb/c murine model. That was possibly due to several factors such as effectlessness of major active components of *H. helix* on this parasite. The second possible explanation for no significant effect of this plant extract on mice ulcer could be that *Leishmania* is an intracellular parasite and some factors such as genetic of the reservoir and the efficacy of cell-mediated immunity can hamper the influences of *H. helix* major components on the infected cells. This was in agreement with the previous research stating that leaves of *H. helix* contain the “emetine” compound, which is an amoebicidal alkaloid, and also triterpene saponins, which are effective against liver flukes, molluscs, intestinal parasites and fungal infections, all of which are extracellular parasites (14, 15).

Our findings were in disagreement with those of Talari et al. in which, 100 and 50 mg/mL of *H. helix* extract could kill all promastigotes of *L. major* *in vitro* (18). This difference of findings may be due to different methods and concentration of the plant extract in the two studies. The previous research used *in vitro* approach, eliminating the effect of immunoresponse of live hosts. In other words, promastigotes of *Leishmania* are extra cellular parasite, while amastigotes are obligate intracellular parasites.

Majester-Savornin showed that an extract of *H. helix* contains 60% saponic complex (CS 60), bidesmosides hederasaponin B, C and D (saponin K10), their corresponding monodesmosides α -, β -, δ -hederin, and hederagenin. CS 60 and bidesmosides have shown no effects. Monodesmosides were found to be as effective on promastigotes of *L. infantum* and *L. tropica*. Against amastigote forms,

only hederagenin exhibited a significant activity (20). However, further studies are required to investigate the activity of hederagenin on *L. major* amastigotes. Some studies have shown that some of Iranian and foreign medical plants have antileishmanial effects. For example, a study conducted by Manjili et al. showed that some Iranian medical plant including *Caesalpinia gilliesii*, *Satureia hortensis*, *Carum copticum* heirm, and *Thymus migricus*, displayed high antileishmanial activity and were toxic against this parasite both *in vitro* and *in vivo* (21).

A recent study by Gharavi et al. indicated that garlic extract could destroy the amastigotes of *L. major* due over-expression of iNOS and IFN γ genes in macrophages (22). Another study showed that the mixture of thyme and yarrow alcoholic extracts had proper effects on treatment of cutaneous lesions of *L. major* in Balb/c infected mice (13). However, my previous study showed the *in vitro* antileishmanial effect of *H. helix* extract, as a definite yield, but the present study did not support this result and had contradiction with others studies. We recommend further studies using the major components of *H. helix* especially hederagenin, hederasaponin B, C and D (saponin K10), to investigate the antileishmanial effects of this plant on *L. major*.

Acknowledgements

This study was supported by the research affairs vice chancellor of Kashan University of Medical Sciences.

Authors' Contribution

The authors are grateful to Dr. Mohebali in the Department of Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, and also Dr. Zahiri in the Department of Parasitology, Kashan University of Medical Sciences for technical support, and Dr. Naderi for his helpful suggestions on this manuscript.

Financial Disclosure

The authors declared no conflict of interest.

Funding/Support

None declared.

References

- Berman JD. Human leishmaniasis: clinical, diagnostic, and chemotherapeutic developments in the last 10 years. *Clin Infect Dis.* 1997;24(4):684-703.
- Postigo JA. Leishmaniasis in the World Health Organization Eastern Mediterranean Region. *Int J Antimicrob Agents.* 2010;36 Suppl 1:S62-5.
- Salman SM, Rubeiz NG, Kibbi AG. Cutaneous leishmaniasis: clinical features and diagnosis. *Clin Dermatol.* 1999;17(3):291-6.
- Khatami A, Firooz A, Gorouhi F, Dowlati Y. Treatment of acute Old World cutaneous leishmaniasis: a systematic review of the randomized controlled trials. *J Am Acad Dermatol.* 2007;57(2):e1-335. e29.

5. Mehrabani D, Motazedian MH, Asgari Q, Hatam GR, Owji SAA, Oryan A. Leishmania major in Tatera indica in Estahban, Southern Iran: Microscopy, culture, isoenzyme and PCR. *Pak J Med Sci*. 2011;**27**(4):7348.
6. Yaghoobi-Ershadi M. Phlebotomine Sand Flies (Diptera: Psychodidae) in Iran and their Role on Leishmania Transmission. *J Arthropod Borne Dis*. 2012;**6**(1):1-7.
7. Van Voorhis WC. Therapy and prophylaxis of systemic protozoan infections. *Drug*. 1990;**40**(2):176-202.
8. Berman J. Current treatment approaches to leishmaniasis. *Curr Opin Infect Dis*. 2003;**16**(5):397-401.
9. Rezayat M. Nanosilver in the treatment of localized cutaneous leishmaniasis caused by Leishmania major (MRHO/IR/75/ER): an in vitro and in vivo study. *DARU J Pharm Sci*. 2009;**17**(4).
10. Ghaffarifar F, Jorjani O, Mirshams M, Miranbaygi MH, Hosseini ZK. Photodynamic therapy as a new treatment of cutaneous leishmaniasis. *East Mediterr Health J*. 2006;**12**(6):902-8.
11. Fata A, Rakltshandeh H, Berenji F, Jalalianfard A. Treatment of cutaneous Leishmaniasis in murine model by alcoholic extract of Berberis vulgaris. *Iran J Parasitol*. 2006;**1**(1):39-42.
12. Yousefi R, Ghaffarifar F, Dalimi A. The effect of Alkanna tinctoria and Peganum harmala extracts on Leishmania major. *Iran J Parasitol*. 2009;**4**:40-7.
13. Shirani-Bidabadi L, Mahmoudi M, Saberi S, Zolfaghari-Baghbaderani A, Nilforoushzadeh MA, Abdoli H, et al. The effectiveness of mix extracts of Thyme, Yarrow and Propolis on Cutaneous Leishmaniasis: a comparative study in animal model (Balb/c). *Tehran Univ Med J*. 2009;**66**(11).
14. Timon-David P, Julien J, Gasquet M, Balansard G, Bernard P. [Research of antifungal activity from several active principle extracts from climbing-ivy: Hedera helix L (author's transl)]. *Ann Pharm Fr*. 1980;**38**(6):545-52.
15. Mshvildadze V, Favel A, Delmas F, Elias R, Faure R, Decanosidze G, et al. Antifungal and antiprotozoal activities of saponins from Hedera colchica. *Pharmazie*. 2000;**55**(4):325-6.
16. Delmas F, Di Giorgio C, Elias R, Gasquet M, Azas N, Mshvildadze V, et al. Antileishmanial activity of three saponins isolated from ivy, alpha-hederin, beta-hederin and hederacolchiside A1, as compared to their action on mammalian cells cultured in vitro. *Planta Med*. 2000;**66**(4):343-7.
17. Eguale T, Tilahun G, Debella A, Feleke A, Makonnen E. Haemonchus contortus: in vitro and in vivo anthelmintic activity of aqueous and hydro-alcoholic extracts of Hedera helix. *Exp Parasitol*. 2007;**116**(4):340-5.
18. Talari A, Hooshyar H, Sayaah M. In vitro effect of various concentrations of Hedera Helix extract and Glucantime on cutaneous Leishmaniasis. *Pejouhesh J*. 1997;**21**(3):46-54.
19. Mohebbali M, Chenari A, Nazari M. The effect of cassia fistula extracts on leishmania major ulcers in laboratory mice. *Pajoohandeh J*. 1999;**4**(1):9-14.
20. Majester-Savornin B, Elias R, Diaz-Lanza AM, Balansard G, Gasquet M, Delmas F. Saponins of the ivy plant, Hedera helix, and their leishmanicidal activity. *Planta Med*. 1991;**57**(3):260-2.
21. Kheiri Manjili H, Jafari H, Ramazani A, Davoudi N. Anti-leishmanial and toxicity activities of some selected Iranian medicinal plants. *Parasitol Res*. 2012;**111**(5):2115-21.
22. Gharavi MJ, Nobakht M, Khademvatan SH, Bandani E, Bakhshayesh M, Roozbehani M. Iran J Parasitol The Effect of Garlic extract on expression of INF γ and Inos genes in macrophages infected with Leishmania major. *Iran J Parasitol*. 2011;**6**(3):74-81.