



CKJ REVIEW

The dietary source of trimethylamine N-oxide and clinical outcomes: an unexpected liaison

Marie Evans ¹, Lu Dai ², Carla Maria Avesani¹, Karolina Kublickiene¹ and Peter Stenvinkel¹

¹Renal Unit, Department of Clinical Sciences and Technology (CLINTEC), Karolinska Institutet, Stockholm, Sweden and ²Aging Research Center, Department of Neurobiology, Care Sciences and Society, Karolinska Institutet, Stockholm, Sweden

Correspondence to: Marie Evans; E-mail: marie.evans@ki.se

ABSTRACT

The profile of gut microbiota can vary according to host genetic and dietary characteristics, and be influenced by disease state and environmental stressors. The uremic dysbiosis results in a loss of biodiversity and overgrowth of microorganisms that may cause elevation of metabolic solutes such as trimethylamine N-oxide (TMAO), inducing pathogenic effects on its host. In patients with chronic kidney disease (CKD), TMAO levels are elevated because of a decreased clearance and an increased production from the uremic gut dysbiosis with a disrupted intestinal barrier and elevated enzymatic hepatic activity. Dietary precursors of TMAO are abundant in animal-derived foods such as red meat, egg yolk and other full-fat dietary products. TMAO is also found naturally in fish and certain types of seafood, with the TMAO content highly variable according to the depth of the sea where the fish is caught, as well as processing and storage. Although evidence points towards TMAO as being an important link to vascular damage and adverse cardiovascular outcomes, the evidence in CKD patients has not been consistent. In this review we discuss the potential dietary sources of TMAO and its actions on the intestinal microbiome as an explanation for the divergent results. We further highlight the potential of a healthy diet as one feasible therapeutic opportunity to prevent gut dysbiosis and reduce uremic toxin levels in patients with CKD.

LAY SUMMARY

There is a link between the intestinal microbiota and human health. Patients with chronic kidney disease have an altered microbiota, with accumulation (because of decreased renal clearance) and increased production of toxins such as trimethylamine-N-oxide (TMAO). Elevated TMAO may induce cardiovascular and kidney damage. Dietary precursors of TMAO are found in animal-derived foods (red meat, egg, fish) and full-fat dietary products. In this review we discuss the potential dietary sources of TMAO, and its actions on the intestinal microbiome and association with worse clinical outcomes. We further highlight the potential of a healthy diet as one feasible therapeutic opportunity to prevent dysbiosis and reduce toxin levels in patients with chronic kidney disease.

Keywords: chronic kidney disease, diet, gut microbiome, trimethylamine-N-oxide, uremic toxin

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AN INDUSTRIALIZED MICROBIOTA

The human gut serves as the host for trillions of microorganisms, commonly referred to as the gut microbiome. These microorganisms constitute its own ecosystem, with physiological functions such as vitamin synthesis and immune system maturation, and they maintain functions for the intestinal barrier defence [1]. In this intricate process, the gut microbiome produces numerous metabolites, either derived directly from dietary compounds or generated through the complex host-microbiome interplay. The metabolome of the gut could thus be of three different types: entirely produced from the microbiome, produced from both the host and the microbiome, or produced from the microbiome and diet [2]. Among the metabolic compounds, the short-chain fatty acids such as propionate and acetate are involved in energy homeostasis, immune regulation, blood pressure control and maintaining the gut barrier defence [3]. One example of the metabolic effects of short-chain fatty acids is that experimental administration of these to humans stimulates the production of glucagon-like peptide 1 and results in lower weight gain [3]. Other important metabolites that interact with the gut microbiota are bile acids, which primarily are synthesized in hepatocytes from cholesterol. Dysbiosis of the gut impacts on bile acid metabolism leading to the accumulation of primary conjugated bile acids in the colon, which may exert pro-inflammatory effects on the intestinal epithelial cells and ultimately result in impaired insulin sensitivity and liver steatosis [1]. Thus, through various interactions with its host, the gut microbiome exerts its actions through regulating various metabolic pathways.

The gut microbiota is individual and varies according to host genetic and dietary characteristics, disease state and environmental stressors such as medication. In chronic non-communicable diseases, the gut microbiome is less diverse. Those with a less rich microbacteria flora have been shown to have elevated insulin resistance, more obesity and dyslipidemia [4]. During the current era, an industrialized microbiota have emerged due not only to changes in eating habits but also to air pollution, microplastics and heat stress [5]. The obesity-associated microbiome usually presents with a reduction in *Bacteroides* species, along with an increase of Firmicutes phylum (e.g. *Clostridium*, *Lactobacillus*, *Bacillus*, *Ruminococcus* and *Enterococcus*) [6]. In chronic kidney disease (CKD), dysbiosis is common and referred to as “uremic dysbiosis.” The uremic dysbiosis results in an altered gut microbiome with an overgrowth of microorganisms which may cause pathogenic effects on its host [7]. These changes occur already in mild kidney dysfunction; *Roseburia*, which has been suggested to serve as a marker of a normal intestinal microbiome, are decreased already in early-stage CKD and become even less abundant in patients on dialysis [8]. This imbalance, which also may be exacerbated by iatrogenic causes such as medication with phosphate binders, proton pump inhibitors and antibiotics, could result in the accumulation of uremic retention products that may further impact on disease pathogenesis and clinical outcomes [9, 10]. The importance of the gut microbiota in CKD was first demonstrated in the 1960s in an experimental study where nephrectomized rats absent of microbiome were observed to live longer than nephrectomized rats with a preserved gut microbiota [11]. Since then, a number of uremic retention products, both protein-bound and soluble, and have been identified as being microbiota-derived and potentially responsible for several of the observed pathological effects in CKD patients [12].

Among the uremic retention products there has been a particular focus on generation of the free-water-soluble molecule trimethylamine N-oxide (TMAO), which has been associated with an increased risk of cardiovascular disease (CVD) and all-cause mortality [13, 14]. Initially thought to be a waste product, TMAO serves as a link to a number of disease conditions and their related pathogenetic mechanisms, including endothelial dysfunction [15], acute heart failure [16], foam cell formation [17], infarcted coronary arteries [18], decreased reverse cholesterol transport [19], inflammation [20, 21] and early vascular ageing [22].

THE LINK BETWEEN GUT MICROBIOTA, TMAO AND THE KIDNEY

Humans are not able to demethylate TMAO and >95% is excreted unchanged by the kidneys through tubular secretion or glomerular filtration [23]. As kidney function deteriorates, TMAO concentrations increase [24]. The median TMAO concentration is around 5.8 $\mu\text{M/L}$ in healthy volunteers but rises 13-fold in CKD stage 5 and remains high after 12 months on dialysis [24]. TMAO, which is a free-soluble low-molecular weight solute of 75 Da is cleared by extracorporeal dialysis to around 85% as opposed to other protein-bound gut-derived uremic toxins such as indoxyl sulfate [25]. The different mechanisms by which TMAO accumulates in CKD are summarized in Fig. 1. Supporting the evidence of a significant role of renal clearance of TMAO, it was observed that following a successful kidney transplantation, TMAO levels return to the levels of healthy adults [24, 26]. TMAO formation could be due to breakdown of food rich in the precursor of TMAO, trimethylamine (TMA). Intestinal bacteria could also produce TMA directly from dietary L-carnitine, phosphatidylcholine, choline or betaine. The TMA-lyase enzyme complex CutC/D converts choline to TMA [27], while L-carnitine and betaine are converted to TMA by CntA/B and YeaW, respectively [28]. The precursor TMA is subsequently transformed into TMAO by the liver-enzyme flavin-containing monooxygenase 3 (FMO3). The activity of FMO is increased in the uremic milieu, thus being at least partly responsible for the elevated TMAO formation associated with CKD [29]. Additionally, patients with CKD have been shown to harbor more of the TMAO-producing intestinal bacteria as opposed to healthy people. Dysbiosis results in the breakdown of the intestinal mucosa barrier through disruption of the enterocyte tight junctions [30]. Consequently, more of the uremic toxins and precursors, such as TMA, are leaked to the bloodstream, reaching the liver, and are converted to TMAO. These alterations are believed to occur in the beginning of the disease process. Even in children with CKD, serum levels of sCD14 and the tight junction protein Zo-1 were increased in those with reduced kidney function, indicating that the phenotype of the “leaky gut” is present at an early stage [31]. Thus, TMAO levels are elevated in CKD because of a decreased clearance and an increased production from the uremic gut dysbiosis with a disrupted intestinal barrier and elevated FMO3 activity in the liver.

DIETARY SOURCES OF TMAO

Dietary precursors of TMAO are abundant in animal-derived foods such as red meat (beef, pork, lamb, veal, processed meat

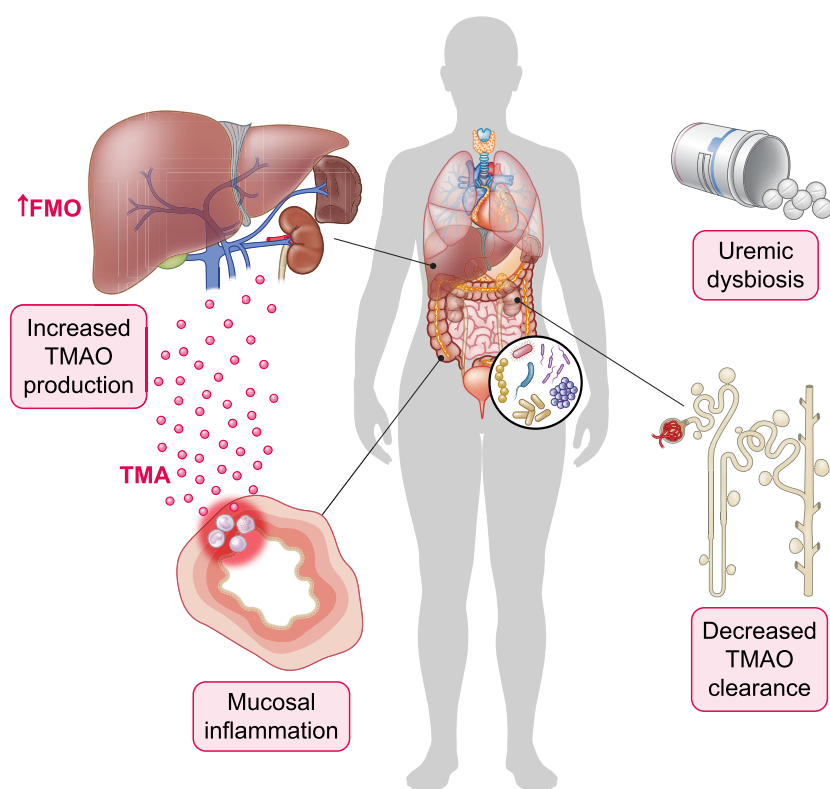
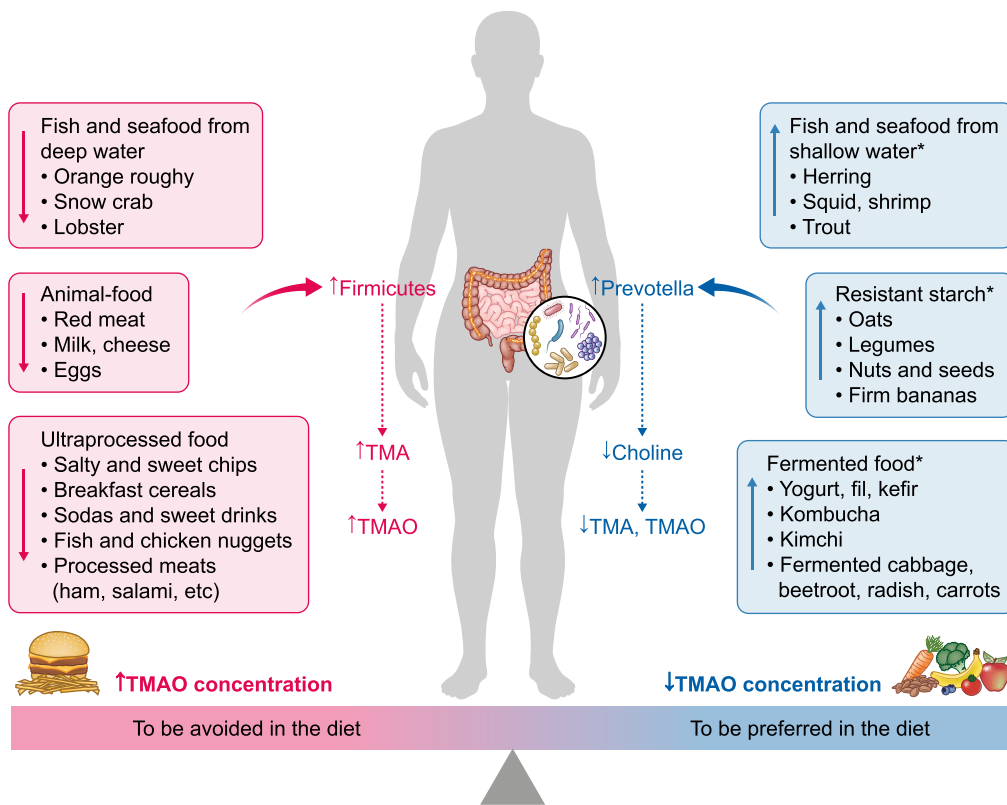


Figure 1: TMAO in patients with CKD. Created with BioRender.com.

and ham), egg yolk and other full-fat dietary products (whole milk, yogurt, cream cheese and butter) (Fig. 2). Consumption of food items rich in dietary precursors of TMAO will lead to processing by the gut microbiome [27] resulting in the release of TMA into the blood and further oxidation into TMAO by hepatic FMO3 [32]. A study on protein source (red meat versus white meat versus non-meat) found that subjects consuming approximately 220 g of steak/day for 1 month had higher TMAO levels accompanied by a reduced fractional renal excretion rate of TMAO [33]. In addition to TMAO derived from its dietary precursors, TMAO is found naturally in fish and certain types of seafood. Fish-source TMAO can bypass gut and liver metabolism and be absorbed directly into the blood stream. TMAO is also present in fish oil/krill oil supplements. A variety of internal and external factors, such as fish/seafood species, feeding quality, fishing zone and storage conditions, could affect the endogenous TMAO concentration in fish products and subsequently contribute to the variation in urinary and circulating TMAO after fish consumption [34, 35]. To date, population studies assessing the relation between diet and circulating TMAO have been inconclusive [36–38], possibly due to differences in dietary habits/culture and genetic heterogeneity in host gut microbial and FMO3 activity across populations. Other factors, such as the role of kidney excretion in the metabolic process could further complex the link between diet and TMAO. It is worth noting that the health effects of TMAO may vary with its dietary source, highlighting the relevance of other compounds in the food.

FISH AND TMAO—A MATTER OF DEPTH

A Swedish group were the first to report that TMA and urinary TMAO levels were associated with intake of fish [39]. A subsequent study showed that compared with a group consuming red meat, the group consuming fish had 4–6 times higher urinary TMAO levels [40]. In another study of 9694 healthy people, it was reported that TMAO levels were associated with kidney function, being male and fish intake [41]. Based on such studies, it has been suggested that TMAO could serve as a potential biomarker of cod and salmon intake [42]. Fish commonly contain *Aeromonas salmonicida*, a bacterial species that is responsible for the production of TMA, which causes the unpleasant “fishy odour.” The content of TMA increases with spoilage and during storage of chilled fish fillets [43]. Taken together, fish may be a rich source of both TMA and TMAO, and fish consumption associates with urinary and plasma TMAO concentrations [36]. As fish is considered a healthy choice of food and a significant source of lipid bio-actives possessing cardiovascular health benefits, the link between TMAO and fish intake is counterintuitive. However, understanding the function of TMAO in nature may help explain this “fish paradox.” In nature, TMAO protects proteins and acts as an osmolyte that counteracts the effects of destabilizers such as low temperature, high urea and high hydrostatic pressure. In deep sea waters the weight of the water pushes water molecules into proteins and distorts them. Without the protection of TMAO life would not be possible in deep oceans. Thus, to habituate deep oceans, marine fish need to build up a protective muscular content of TMAO. A study by Yancey et al. [44] in



*The quantities of these food items have to be individualized in the diet according to the nutritional status and clinical condition, considering serum potassium and phosphate.

Figure 2: Dietary measures to lower TMAO levels.

the Hadal snailfish (the second-deepest fish recorded) showed a strong linear correlation between the muscular content of TMAO and depth. In general, deep-sea fish species contain high TMAO levels whereas freshwater and shallow-living seafood, such as farm-raised salmon, shrimp, trout and clams, are low or even absent in TMAO [45]. To test the effects of a single fish meal on circulating TMAO levels, a study was conducted in 10 healthy controls. The study showed that whereas a meal consisting of shrimp and canned tuna did not result in elevated circulating TMAO levels, wild salmon and especially fish sticks resulted in a major increase in serum TMAO. Deep-sea fish like cod and Alaska pollock are the main ingredients in fish sticks and a significant source of TMAO. Although serum TMAO levels had returned to baseline the next day the marked increase in TMAO, especially following a meal of fish sticks, is worrisome [45]. It is likely that when kidney function is compromised, TMAO levels may accumulate with time if TMAO-rich fish is consumed on a regular basis. To test this hypothesis, the effects of a single and regular meals with fish low or rich in TMAO should be conducted in CKD patients. Beside depth, water temperature also affects TMAO levels in fish. A study from New Zealand not only confirmed the link between depth and TMAO but also showed that the TMAO content was lower in the summer months when water was warmer [43]. As TMAO accumulate in deep-sea fish this offers clues for specific nutritional recommendations in patients with a reduced clearance of TMAO. This is a research area in which more work needs to be done with a huge potential for

clinical impact in patients with reduced renal function. A comparative study of TMAO in different animal species has strengthened the link between eating habits and outcome and survival advantage in the animal kingdom [46].

TMAO AND THE LINK TO CARDIOVASCULAR DAMAGE

Several epidemiological studies have shown an association between higher TMAO levels and increased cardiovascular risk [17, 47, 48]. The initial human study that suggested TMAO as a mediator of CVD, utilized a plasma metabolomic screening to detect three metabolites of phosphatidylcholine (choline, betaine, and TMAO). These three metabolites were independently found to be predictive of incident CVD events such as heart attack, stroke, and cardiovascular death over a 3-year follow-up period in 1876 subjects who were undergoing cardiac disease evaluation [17]. Subsequently, the researchers expanded their study and discovered that increased plasma levels of L-carnitine, a precursor to TMAO, also predicted cardiovascular events when elevated in association with TMAO over a 3-year follow-up period [19].

The relationship between cardiovascular outcomes, mortality and TMAO has also been demonstrated in CKD patients [18, 26, 49]. In a cross-sectional sample, serum TMAO independently predicted the number of infarcted coronary arteries [18]. In another cross-sectional analysis of 220 patients with estimated

glomerular filtration rate <45 mL/min/1.73 m² who underwent coronary angiography, elevated serum TMAO levels were associated with the severity of coronary arterial disease after adjusting for traditional cardiovascular risk factors [26]. Over a subsequent follow-up over 4 years, every 10-μM increase in baseline TMAO was associated with a 19% increase in total mortality [26]. One study included 521 CKD3 patients and found that those in the highest quartile of TMAO had a 1.9-fold greater risk of 5-year all-cause mortality compared with those with the lowest quartile [49]. However, the evidence of the detrimental effect of TMAO has not been consistent. In a large study of Canadian CKD patients, circulating TMAO levels independently predicted CVD events over a 3-year follow-up period in CKD3 but not in CKD4 [50]. Furthermore, in another study, those with prior cerebrovascular disease had lower TMAO levels [51] and a European follow-up study did not find an association between the development of coronary artery disease and plasma TMAO [52]. The reasons for these inconsistencies are not clear, but one alternative hypothesis is that only glomerular function was accounted for in the analyses showing a positive correlation between TMAO and CVD. This opened the possibility of residual confounding from tubular renal function [53].

TMAO EFFECTS ON THE VASCULAR SYSTEM

The mechanisms by which TMAO exerts its effects on the cardiovascular system have gradually been revealed. Predominantly, the consequences of TMAO are linked to vascular inflammation [15], platelet hyperactivity, calcification and atherosclerosis [54]. Experimental evidence indicates that TMAO may directly contribute to the development of atherosclerosis and lead to cardiovascular events by disrupting lipid handling and macrophage function, as well as causing vascular inflammation and platelet activation, which could result in thrombosis [55]. Studies using atherosclerosis-prone apolipoprotein E knockout mice (ApoE^{-/-}) supplemented with choline or TMAO showed increased plasma TMAO levels and larger aortic atherosclerotic plaques with higher macrophage content compared with wild-type mice [17]. In ApoE^{-/-} mice, elevated TMAO reduced the reverse cholesterol transport [19] and caused development of cholesterol-laden foam cells [17]. Proinflammatory mediators, such as cyclo-oxygenase-2, E-selectin and intracellular adhesion molecule-1, were upregulated in the aortic tissue of low-density lipoprotein receptor knockout mice who received dietary choline or intraperitoneal TMAO [56]. *In vitro* studies showed that TMAO may increase leukocyte adhesion to endothelial cells in an nuclear factor κB-dependent manner [56]. Furthermore, enhanced platelet activation and adhesion was observed following intraperitoneal TMAO injections in an *in vivo* carotid artery injury model [57]. TMAO also exacerbates the development of atherosclerotic plaques, which may ultimately lead to reduced blood flow or increased arterial stiffness [17]. In addition, TMAO was shown to impair endothelial signalling [15], an early event in the development of atherosclerosis [58], and an animal experiment showed that supplementation with TMAO impaired endothelium-dependent dilatation via reduced contribution of hyperpolarizing factor-type contribution [59].

Several studies have showed a link between TMAO and vascular ageing via upregulation of a prooxidative environment, further strengthening the suggestion that TMAO-deteriorated endothelial function is the link between TMAO and CVD [60, 61]. TMAO may also inhibit protein function or limit degradation of key enzymes or signalling proteins, leading to potentially harmful downstream effects [62]. TMAO promotes

atherosclerosis by inhibiting reverse cholesterol transport, a process that removes excess cholesterol from arterial walls [55]. Moreover, elevated TMAO promotes endothelial cell senescence [60]. TMAO also inhibits autophagy, causing the accumulation of damaged proteins and the impairment of cellular function [63]. Moreover, TMAO reduces the activity of a key enzyme, CYP3A4, responsible for metabolizing various drugs, including statins, which are commonly used to lower cholesterol levels [64]. TMAO can also cross the blood-brain barrier and affect brain function, leading to cognitive impairment and dementia [65].

TMAO AND THE LINK TO KIDNEY DISEASE PROGRESSION

In addition to the effects on vascular smooth muscle cells and endothelium, TMAO induces structural kidney damage [49]. Animal studies show that a high-fat diet or dietary supplementation with choline or TMAO induces tubulointerstitial fibrosis and promote the expression of kidney injury markers and pro-fibrotic genes [66]. TMAO activates renal fibroblasts and causes fibroblast proliferation [67]. In support of this finding, pharmacological inhibition of the TMA production in mice has been related to a lesser kidney injury and fibrosis [68]. Furthermore, TMAO may exert its actions directly on renal tubular cells through decreasing the protein expression of megalin, an effect that could be reversed by the antiproteinuric drugs candesartan and dapagliflozin [69].

DIETARY SOURCES OF TMAO AND CARDIOVASCULAR OUTCOMES IN CKD

So far inconsistencies exist with respect to the association between TMAO and CVD in CKD. One putative reason for the difference in effect of TMAO on clinical outcomes in CKD patients is that the elevated TMAO levels may arise from different dietary sources. Red meat as an important source of TMAO, has negative effects on both gut microbiota and host health [70]. Red meat also reduces the ability of kidneys to excrete TMAO [33]. Due to its choline content, intake of egg may also increase systemic TMAO levels. However, a study conducted in healthy controls showed that an intake of four eggs per day for 28 days did not increase circulating TMAO levels [71]. As previously reported, fish could also be a dietary source of TMAO [40], but fish intake (especially that rich in ω-3 fatty acids) is associated with lower cardiovascular event rate and lower risk of death in high risk cohorts or people with previously known vascular disease [72]. Examples of fish that are high in ω-3 levels include herring, mackerel, sable, salmon, tuna, anchovy, trout and sardine. To investigate the hypothesis that different dietary sources of TMAO could interact with the TMAO-mortality association, we performed an analysis in a European cohort of 737 patients with CKD stage 4–5 where we investigated the association between mortality, TMAO and 3-carboxy-4-methyl-5-propyl-2-furanpropionate (CMPF). CMPF is a metabolite of furan fatty acids, which is found predominantly in fish and fish oils, and considered as a biomarker of fish intake [73]. Long-chain ω-3 fatty acids are suggested to be the precursors of CMPF [74]. We found that, after extensive adjustments, ln-TMAO was positively associated with mortality, whereas ln-CMPF was negatively associated [75]. When we combined levels of TMAO and CMPF, we observed that compared with patients with low levels of both TMAO and CMPF, those with low TMAO levels and high CMPF levels had a lower mortality, while those with high TMAO

levels and low CMPF levels had higher mortality rate in the unadjusted model, albeit outside the significance level in the adjusted analysis. Patients with high levels of both TMAO and CMPF did not have any different association with mortality compared with patients with low levels, suggesting that the concomitant high CMPF levels may counteract an unfavorable association between TMAO and mortality. CMPF was further associated with an overall lower risk of start in kidney replacement therapy. Our interpretation is that an overall high fish intake, as suggested by high CMPF levels from fish predominantly rich in ω -3 fatty acids, also may result in higher TMAO levels that may be less harmful, as compared with high TMAO levels from red meat or fish low in ω -3 fatty acids but high in TMAO. A proper clinical trial with different dietary protein sources, including fish with different TMAO contents, is needed to confirm this hypothesis. As a recent study showed that a higher intake of plant-based protein, but not animal or dairy protein, was related to a lower risk of frailty [76] we believe that the renal community should pay much more attention to the impact of different sources of protein in relation to progression of kidney disease and the uremic phenotype.

DIETARY THERAPEUTIC OPPORTUNITIES

Dietary composition can modulate the gut microbiota and consequently affect microbiota-derived TMA and TMAO levels. The potential strategies to normalize TMAO levels in patients with CKD are summarized in Fig. 2. Since TMAO is generated from the gastrointestinal metabolism of foods containing choline, lecithin and L-carnitine, and these in turn are mostly present in red meat, deep water fish, eggs, milk and cheese, a dietary pattern compatible with a plant-based diet is an attractive and optimal opportunity to decrease TMAO levels [77]. Diets rich in animal protein with saturated fat are related to higher abundance of the bacteria phylum Firmicutes, which is associated with TMA production [19], while vegetarian diets, rich in oligosaccharide, are associated with bacteria genus *Prevotella* (related to a reduction in choline availability for TMA synthesis) [78]. In studies with healthy individuals following plant-based diets (Mediterranean, vegetarian and vegan diets) the TMAO levels decrease, while animal-based diets had the opposite effect [77].

Beyond the restricted intake of precursors of TMA/TMAO from red meat, a plant-based dietary pattern can decrease TMAO levels due to a synergistic effect with other dietary components, such as high phytochemicals and fiber intake, which can promote commensal microbial growth and ameliorate dysbiosis [79]. Even though there is a need to investigate the influence of plant-based diets in altering the TMAO in CKD, several studies with specific dietary interventions from vegetable sources already point in this direction. For example, interventional studies using resistant starch (a type of dietary fiber that can act as a substrate for microbial fermentation and improve the integrity of the intestinal epithelial barrier) is of great potential [80]. A meta-analysis that included eight crossover or parallel-designed randomized controlled trials that lasted for >4 weeks aiming to study the effect of resistant starch in CKD (including dialysis) showed that resistant starch promoted a reduction in the uremic toxin serum indole phenol sulfate, phosphate and interleukin-6 in patients on dialysis [81]. Under the same rationale of foods to improve the gut microbiota health, the consumption of fermented food increases the microbiome diversity and can ameliorate postprandial TMAO response [82]. Sources of fermented

food include yogurt, fil, kefir, kombucha and kimchi, but there are plenty of recipes for fermenting vegetables, using cabbage, beetroot, radish, turnip and carrots, that can be more culturally appealing for incorporating into the diet.

In addition to the dietary changes suggested above, it may be beneficial to replace fish and seafood from deep waters (orange roughy, snow crab, lobster, cod) by those from shallow waters (mackerel, barracouta, squid, herring, salmon, trout, clams, grey mullet and shrimp) [83], although studies are yet to prove this. Tuna is an exception, as it is fish from deep water, but has low TMAO levels [83]. Finally, the increasing consumption in the Western dietary pattern of so-called ultra-processed foods (UPF) requires attention. UPF encompass industrialized foods that have undergone heavy industrial processing (food heating and Maillard reaction), with the addition of sugar, salt, artificial non-caloric sweeteners, saturated fat and trans-fat, and of food additives to change the color and enhance taste [84]. Examples of UPF include salty and sweet chips, breakfast cereals, sugary products, soft drinks and processed juices, and commercialized ready-to-eat meals, such as the fish nuggets and others [84]. In mice it was shown that heated food (such as UPF) contributed to a decrease in the intestinal epithelial barrier with translocation of lipopolysaccharide from the gut to the systemic circulation, with an increase in colonization of Firmicutes in the gut, which could lead to an enhanced rise in TMAO levels [85]. However, as of yet this is of a speculative nature and requires confirmation.

CONCLUDING REMARKS

In patients with kidney dysfunction, several fundamentally different mechanisms contribute to increased TMAO levels linked to high CVD burden. Dietary changes could be an important and modifiable factor that offer potential beneficial effects on the gut microbiota and uremic retention solute concentrations. Understanding the way different dietary sources of TMAO may induce the detrimental effect of TMAO on host health is vital in order to be able to improve the outcomes of our CKD patients.

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CONFLICT OF INTEREST STATEMENT

None of the authors has any conflicts of interest to declare in relation to this manuscript.

DATA AVAILABILITY STATEMENT

No new data were generated or analysed in support of this research.

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