

Clinical Study

Genetic Variations of α -Methylacyl-CoA Racemase Are Associated with Sporadic Prostate Cancer Risk in Ethnically Homogenous Koreans

Sang-Jin Lee,¹ Jae Young Joung,² Hyekyoung Yoon,³ Jeong Eun Kim,²
Weon Seo Park,² Ho Kyung Seo,² Jinsoo Chung,² Jung-Ah Hwang,⁴ Seung-Hyun Hong,⁴
Seungyoon Nam,⁴ Sohee Park,³ Jeongseon Kim,^{5,6} Kang Hyun Lee,² and Yeon-Su Lee⁴

¹ Genitourinary Cancer Branch, National Cancer Center, Goyang 410-769, Republic of Korea

² Center for Prostate Cancer, National Cancer Center, 111 Jungbalsan-ro, Ilsandong-gu, Goyang, Gyeonggi-do 410-769, Republic of Korea

³ Cancer Biostatistics Branch, National Cancer Center, Goyang 410-769, Republic of Korea

⁴ Cancer Genomics Branch, National Cancer Center, 111 Jungbalsan-ro, Ilsandong-gu, Goyang, Gyeonggi-do 410-769, Republic of Korea

⁵ Molecular Epidemiology Branch, National Cancer Center, Goyang 410-769, Republic of Korea

⁶ Department of Biostatistics, Graduate School of Public Health, Yonsei University, Seoul 120-752, Republic of Korea

Correspondence should be addressed to Kang Hyun Lee; uroonco@ncc.re.kr and Yeon-Su Lee; yslee2@ncc.re.kr

Received 1 August 2013; Revised 7 October 2013; Accepted 9 October 2013

Academic Editor: Sue-Hwa Lin

Copyright © 2013 Sang-Jin Lee et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. To assess if the variants of (R)-alpha-methyl-CoA racemase (AMACR) gene would be associated with the risk of sporadic prostate cancer in ethnically homogenous Koreans. **Materials and Methods.** We enrolled 194 patients with prostate cancer and 169 healthy controls. A total of 17 single nucleotide polymorphisms of the AMACR gene were selected. The distribution of each genotype and haplotype was analyzed and their association with the incidence of prostate cancer was evaluated. Further, we detected AMACR expression in tumor with immunohistochemistry and analyzed its association with genotype regarding prostate cancer risk. **Results.** AG or GG genotype of rs2278008 (E277K) tended to lower prostate cancer risk. The minor G allele was found to be a significant allele that decreased the risk of prostate cancer (adjusted OR, 0.57; 95% CI, 0.35–0.93, P value = 0.025). In patients expression AMACR, AG or GG genotype was also significant genotype in terms of prostate cancer risk (adjusted OR, 0.47; 95% CI, 0.26–0.87, P value = 0.017). Further, [GGCGG] haplotype consisted of five coding SNPs of rs2278008, rs34677, rs2287939, rs10941112, and rs3195676 which decreased the risk of prostate cancer (P value = 0.047). **Conclusions.** Genetic variations of AMACR are associated with the risk of sporadic prostate cancer that underwent radical prostatectomy in Koreans.

1. Background

The detailed etiology of prostate cancer is still unclear; it is a very heterogeneous disease due to the involvement of various inherited genetic elements and environment factors, including a fatty diet. Many studies have sought to identify the risk factors of prostate cancer, mainly by a targeted gene approach, which has confirmed the effect of known carcinogenesis genes [1–4]. Recently, many studies including genome-wide association studies (GWAS) identified significant associations between lots of single nucleotide polymorphisms (SNPs) and prostate cancer [5–9]. Some studies

identified chromosome 5p13, the site of the gene encoding (R)-alpha-methyl-CoA racemase (AMACR), as the location of a prostate cancer susceptibility gene [10–12].

AMACR is catalytically involved in fatty acid oxidation, which converts (R)-alpha-methyl-branched-chain fatty acyl-CoA ester to the (S)-stereoisomer. AMACR is critical in prostate cancer cell progression; the downregulation of AMACR expression hampers the proliferation of the LAPC-4 androgen-responsive prostate cancer cells [13]. AMACR is abundantly expressed and is recognized as a standard tissue biomarker capable of a highly sensitive and specific diagnosis of prostate cancer [14–16]. An AMACR spliced variant was

reported capable of creating a novel transcript that is expressed with other forms of AMACR in prostate cancer [17]. These findings point to the value of AMACR and its variants in developing diagnostic biomarkers that will complement the diagnostic capability of PSA, while addressing the limitations of PSA, specifically its low specificity. The chromosomal region of AMACR has been validated as a susceptible locus for prostate cancer, including hereditary prostate cancer [10, 11]. Studies addressing the potential linkage between AMACR polymorphisms and sporadic prostate cancer risk in different populations, however, have not been consistent and have precluded any definitive conclusion [18–20]. The genetic heterogeneity of ethnically different study populations might have lead to these inconsistent results. This study was designed to assess whether genetic variations of AMACR were associated with sporadic prostate cancer development in Korean men, known to be an ethnically homogenous population [21], by investigating the impact of AMACR polymorphisms on the risk of prostate cancer and clinical features.

2. Materials and Methods

2.1. Study Subjects. The investigation was a hospital-based case-control study of prostate cancer. Both prostate cancer patients and controls were of the same Korean ethnic origin with all residents born in Korea. Any subject who had any relative with a past history of prostate cancer among their first-degree relatives was excluded. Patients who were histologically confirmed to have prostate cancer were enrolled at the National Cancer Center in Korea between January 2005 and February 2009. Controls were confirmed to be free of prostate cancer by determination of the blood PSA level and by digital rectal examination. Although the PSA level of three controls continually increased and exceeded 4 ng/mL, prostate needle biopsy conducted in the three subjects confirmed the absence of prostate cancer. All patients underwent radical prostatectomy and Gleason grading was determined with prostatectomy specimens. Demographic and clinical data were based on prostate cancer database in our institution. We obtained the written informed consent for participation in the study from all participants. This study was approved by the Institutional Review Board of the National Cancer Center in Korea (NCCNCS05-049).

2.2. Single Nucleotide Polymorphism (SNP) Selection and Genotyping. The target SNPs of AMACR were selected in the National Center for Biotechnology Information (NCBI) database version (<http://www.ncbi.nlm.nih.gov/projects/SNP/>). We collected the information of all coding SNPs in AMACR with heterozygosity > 0; a total of 17 SNPs were identified (see Supplemental Table 1 available online at <http://dx.doi.org/10.1155/2013/394285>). From these, four SNPs with a heterozygosity of 0.5 were excluded. Additional two SNPs (rs76184600 and rs117220551) were also excluded because of assay design failure to create suitable primers. Finally, primers for 11 SNPs were designed using DESINGER (Sequenom, CA, USA) software (Supplemental Table 2). Genomic DNA was prepared from 200 μ L of peripheral blood

using the QIAamp Blood Kit (QIAGEN, Valencia, CA, USA) according to the manufacturer's instructions. Genotyping was performed by the previously described method [1] and the resulting genotype data were obtained by Typer (version 4.0, Sequenom) and were subjected to statistical analysis.

2.3. AMACR Expression in Prostate Cancer. At least three core tissue biopsies (each 3 mm in diameter) were taken from morphologically representative regions of each paraffin embedded prostate tumor and precisely arrayed using a custom built instrument. To identify AMACR protein expression, 4 μ m thick tissue sections were placed on glass slides for histological examination, one slide for staining with hematoxylin/eosin and the other slide for immunostaining with anti-AMACR antibody. On the basis of histological analysis, expression of AMACR protein in each patient was determined in a blinded fashion by a single experienced pathologist. Slides were scored by a semiquantitative method. The intensity of AMACR expression was graded on a score of 0 to 3+ (0 = no staining, 1+ = weakly positive, 2+ = positive, and 3+ = strongly positive).

2.4. Statistical Analyses. The demographic and clinical characteristics in the study population and controls were analyzed using a chi-square test for categorical variables and an independent *t*-test for continuous variables. Hardy-Weinberg equilibrium (HWE) was determined separately in cases and controls for each SNP. SNPs that deviated from the HWE in the control group were excluded (χ^2 test at *P* value < 0.05). The association between AMACR polymorphisms and prostate cancer risk was evaluated on the basis of genotypic frequencies of each SNP using an unconditional logistic regression model. The Cochran-Armitage trend test was also used to analyze the linear trend, as expressed by odds ratios (ORs). The haplotype estimation was carried out using the PHASE program (version 2.1; <http://stephenslab.uchicago.edu/software.html#phase>), which implements a Bayesian statistical method for reconstructing haplotypes. An unconditional logistic regression determined the association between AMACR haplotypes and prostate cancer risk, considering the haplotypic frequencies of each haplotype and the haplotype pair of each person. Additionally, the associations between genotypes and clinical parameters were determined using an unconditional logistic regression. To assess the power for detecting association, we used the power for genetic association analyses (PGA) [22]. All other statistical analyses were performed using SAS (version 9.1 SAS Institute, NC, USA). The reported *P* values are two-sided, and a significance level < 5% was considered statistically significant.

3. Results

3.1. Subjects Characteristics. Demographic and clinical characteristics of 194 prostate cancer patients and 169 controls are summarized in Table 1. Statistically significant differences between cases and controls were evident with respect to age, serum PSA level, and drinking status. Most of the prostate cancer patients displayed a high serum PSA level, with

TABLE 1: Demographic and clinical characteristics of the subjects.

Variables	Cases (<i>n</i> = 194)	Controls (<i>n</i> = 169)	<i>P</i> value ^a	
Age, years (median)	66.52 ± 7.16 (68.0)	60.13 ± 3.30 (61.0)	<0.001	
Serum PSA, ng/mL (median)	21.28 ± 28.15 (9.8)	1.47 ± 1.63 (1.0)	<0.001	
Serum PSA (ng/mL), no.			<0.001	
<4	11 (5.7%)	160 (94.7%)		
4–10 (≥4 to <10)	87 (44.9%)	8 (4.7%)		
10–20 (≥10 to <20)	43 (22.2%)	1 (0.6%)		
≥20	53 (27.3%)	0 (0.0%)		
BMI, kg/m ² (median)	24.35 ± 2.60 (24.1)	24.42 ± 2.34 (24.2)	0.800	
BMI, no.			0.388	
<25	127 (65.5%)	102 (61.1%)		
≥25	67 (34.5%)	65 (38.9%)		
Smoking, pack years (median)	21.93 ± 21.75 (18.3)	19.24 ± 20.60 (14.9)	0.264	
Smoking status, no.			0.672	
Never	52 (26.8%)	42 (24.9%)		
Ever	142 (73.2%)	127 (75.2%)		
Drinking status, no.			0.002	
Never	68 (35.1%)	34 (20.1%)		
Ever	126 (65.0%)	135 (79.9%)		
Hypertension, no.			0.502	
No	120 (61.9%)	109 (65.3%)		
Yes	74 (38.1%)	58 (34.7%)		
Family history of prostate cancer [†]			0.132	
No	186 (95.9%)	146 (98.6%)		
Yes	8 (4.1%)	2 (1.4%)		
Gleason score, no.				
2–6	99 (51.0%)			
7	62 (32.0%)			
8–10	33 (17.0%)			
pTstage, no.				
pT0	6 (3.1%)			
pT2a	37 (19.1%)			
pT2b	3 (1.6%)			
pT2c	89 (45.9%)			
pT3a	31 (16.0%)			
pT3b	28 (14.4%)			
Immunohistochemistry of AMACR	Serum PSA, ng/mL			
	<4	4–10 (≥4 to <10)	10–20 (≥10 to <20)	≥20
0 (no staining)	1 (2.6%)	10 (25.6%)	7 (18%)	21 (53.9%)
1+ (weakly positive)	0 (0%)	16 (47.1%)	9 (26.5%)	9 (26.5%)
2+ (positive)	5 (9.1%)	25 (45.5%)	12 (21.8%)	13 (23.6%)
3+ (strongly positive)	5 (7.7%)	36 (55.4%)	15 (23.1%)	9 (13.9%)
				0.004

PSA: prostate specific antigen; BMI: body mass index; no.: number; AMACR: (R)-alpha-methyl-CoA racemase.

^aPearson's χ^2 test for categorical variables and independent *t*-test for continuous variables; two-sided *P*-values.

[†]"Family" represents relatives with the exception of first-degree relative, excluding 21 persons with missing values in the control group.

the risk increasing with age, as expected. No significant differences were found in smoking status, body mass index, or hypertension (Table 1).

3.2. AMACR Polymorphisms and Prostate Cancer Risk. Genotype frequencies for all of the target SNPs of AMACR in this study are presented in Supplemental Table 3. From 11 cSNPs, one SNP failed during the data quality control process

because of high primer-dimer potential. Five cSNPs were revealed to be monomorphic for the Korean subjects. For the remaining five cSNPs (rs2278008, rs34677, rs2287939, rs10941112, and rs3195676), the association with prostate cancer was analyzed and is summarized in Table 2. One missense SNP (Glu277Lys), rs2278008, was found to be associated with prostate cancer risk. Individuals with the AG or GG genotype of rs2278008 revealed to have a lower risk compared to those

TABLE 2: Frequency of AMACR (5p13) polymorphisms and association with prostate cancer risk.

SNP	Genotype	Cases (%)	Controls (%)	Crude OR (95 %CI)	P	Adjusted OR (95 %CI) ^a	P	P _{trend} [†]
rs2278008	A/A	145 (74.7)	109 (64.9)	Ref.		Ref.		0.0407
	A/G	47 (24.2)	56 (33.3)	0.63 (0.40–1.00)	0.0500	0.51 (0.29–0.91)	0.0220	
	G/G	2 (1.0)	3 (1.8)	0.50 (0.08–3.05)	0.4535	0.42 (0.06–2.99)	0.3892	
	A/G + G/G	49 (25.3)	59 (35.1)	0.62 (0.40–0.98)	0.0415	0.51 (0.29–0.89)	0.0178	
	A allele	337 (86.9)	274 (81.5)	Ref.		Ref.		
rs34677	G allele	51 (13.1)	62 (18.5)	0.67 (0.45–1.00)	0.0506	0.57 (0.35–0.93)	0.0251	0.8638
	G/G	143 (73.7)	123 (73.2)	Ref.		Ref.		
	G/T	46 (23.7)	43 (25.6)	0.92 (0.57–1.49)	0.7343	1.27 (0.66–2.42)	0.4761	
	T/T	5 (2.6)	2 (1.2)	2.15 (0.41–11.27)	0.3656	1.81 (0.25–12.93)	0.5559	
	G/T + T/T	51 (26.3)	45 (26.8)	0.97 (0.61–1.56)	0.9149	1.30 (0.70–2.44)	0.4069	
rs2287939	G allele	332 (85.6)	289 (86.0)	Ref.		Ref.		0.7474
	T allele	56 (14.4)	47 (14.0)	1.04 (0.68–1.58)	0.8644	1.30 (0.74–2.27)	0.3656	
	C/C	149 (76.8)	124 (73.8)	Ref.		Ref.		
	C/T	39 (20.1)	41 (24.4)	0.79 (0.48–1.30)	0.3587	0.66 (0.36–1.23)	0.1935	
	T/T	6 (3.1)	3 (1.8)	1.66 (0.41–6.79)	0.4777	0.78 (0.14–4.21)	0.7681	
rs10941112	C/T + T/T	45 (23.2)	44 (26.2)	0.85 (0.53–1.37)	0.5095	0.67 (0.37–1.22)	0.1938	0.2155
	C allele	337 (86.9)	289 (86.0)	Ref.		Ref.		
	T allele	51 (13.1)	47 (14.0)	0.93 (0.61–1.43)	0.7403	0.72 (0.43–1.22)	0.2268	
	G/G	69 (35.6)	76 (45.2)	Ref.		Ref.		
	G/A	96 (49.5)	66 (39.3)	1.60 (1.02–2.52)	0.0410	1.52 (0.86–2.70)	0.1514	
rs3195676	A/A	29 (14.9)	26 (15.5)	1.23 (0.66–2.29)	0.5164	1.24 (0.56–2.76)	0.5979	0.2955
	G/A + A/A	125 (64.4)	92 (54.8)	1.50 (0.98–2.28)	0.0616	1.44 (0.85–2.47)	0.1779	
	G allele	234 (60.3)	218 (64.9)	Ref.		Ref.		
	A allele	154 (39.7)	118 (35.1)	1.22 (0.90–1.65)	0.2055	1.21 (0.82–1.79)	0.3273	
	G/G	69 (35.9)	74 (44.0)	Ref.		Ref.		
rs10941112	G/A	94 (49.0)	68 (40.5)	1.48 (0.94–2.33)	0.0883	1.40 (0.79–2.48)	0.2553	0.2849
	A/A	29 (15.1)	26 (15.5)	1.20 (0.64–2.23)	0.5728	1.19 (0.53–2.66)	0.6729	
	G/A + A/A	123 (64.1)	94 (56.0)	1.40 (0.92–2.14)	0.1172	1.34 (0.78–2.29)	0.2849	
	G allele	232 (60.4)	216 (64.3)	Ref.		Ref.		
	A allele	152 (39.6)	120 (35.7)	1.18 (0.87–1.60)	0.2856	1.16 (0.79–1.72)	0.4404	

SNP: single nucleotide polymorphisms; OR: odds ratio; CI: confidence interval; AMACR: (R)-alpha-methyl-CoA racemase.

^aAdjusted for age, smoking quantity, alcohol drinking, and family history of prostate cancer.

[†]Cochran-Armitage trend test for the number of variant alleles.

with the AA genotype (adjusted OR, 0.51; 95% CI, 0.29–0.89, $P = 0.018$). Also, comparison of the allele frequencies showed a statistically significant higher frequency of G allele in the control group than the case group, indicating that the G allele was associated with a lower risk of prostate cancer than the A allele (adjusted OR, 0.57; 95% CI, 0.35–0.93, $P = 0.025$). One other cSNP (G175D), rs10941112, also showed a significant association for the GA genotype which was evident by an increased prostate cancer risk (OR, 1.60; 95% CI, 1.02–2.52, $P = 0.041$), although it was no more significant after adjustment. No significant evidence of prostate cancer association with the remaining SNPs was identified.

3.3. Association of the AMACR Polymorphism with Clinical Factors of Prostate Cancer.

Reclassifying the patients into

two groups according to their GS produced 95 patients (49.0%) with $GS \geq 7$ and 99 patients (51.0%) with $GS < 7$. Interestingly, for patients with $GS \geq 7$, the GA or AA genotype of rs10941112 and rs3195676 showed statistically significant association with increased risk of prostate cancer with an adjusted OR of 2.28 ($P = 0.010$) and 2.35 ($P = 0.008$), respectively, (Table 3). The same genotypes of these cSNPs were also associated with an increased risk of prostate cancer with statistical significance for the patients with pTstage \geq pT2c (adjusted OR 2.60, $P = 0.007$ for rs10941112 and an adjusted OR of 2.66, $P = 0.006$ for rs3195676).

3.4. AMACR Polymorphisms and Risk of Prostate Cancer with Amacr Expression.

For the case group, AMACR expression in tumor tissues was determined by immunostaining and

TABLE 3: Association between AMACR (5p13) polymorphisms and clinical factors in prostate cancer patients.

	AG + GG (%)	AA (%)	OR (95% CI)	P	Adjusted OR (95% CI) ^a	P	OR (95% CI)	P	Adjusted OR (95% CI) ^a	P
Gleason score										
<7	29 (59.2)	70 (48.3)	Ref.	Ref.	Ref.	Ref.	30 (58.8)	168 (49.9)	Ref.	Ref.
≥7	20 (40.8)	75 (51.7)	0.64 (0.33–1.24)	0.1883	0.63 (0.32–1.24)	0.1845	21 (41.2)	169 (50.1)	0.70 (0.38–1.26)	0.2340
pTstage										
≤pT2b	12 (24.5)	34 (23.4)	Ref.	Ref.	Ref.	Ref.	12 (23.5)	80 (23.7)	Ref.	Ref.
≥pT2c	37 (75.5)	111 (76.6)	0.94 (0.44–2.01)	0.8822	1.00 (0.47–2.15)	0.9904	39 (76.5)	257 (76.3)	1.01 (0.51–2.02)	0.9739
rs34677	GT + TT (%)	GG (%)					T (%)	G (%)		
Gleason score										
<7	26 (51.0)	73 (51.0)	Ref.	Ref.	Ref.	Ref.	30 (53.6)	168 (50.6)	Ref.	Ref.
≥7	25 (49.0)	70 (49.0)	1.00 (0.53–1.90)	0.9933	1.06 (0.55–2.05)	0.8558	26 (46.4)	164 (49.4)	0.89 (0.50–1.57)	0.6811
pTstage										
≤pT2b	18 (35.3)	28 (19.6)	Ref.	Ref.	Ref.	Ref.	20 (35.7)	72 (21.7)	Ref.	Ref.
≥pT2c	33 (64.7)	115 (80.4)	0.45 (0.22–0.91)	0.0254	0.42 (0.21–0.87)	0.0193	36 (64.3)	260 (78.3)	0.50 (0.27–0.91)	0.0243
rs2287939	CT + TT (%)	CC (%)					T (%)	C (%)		
Gleason score										
<7	27 (60.0)	72 (48.3)	Ref.	Ref.	Ref.	Ref.	31 (60.8)	167 (49.6)	Ref.	Ref.
≥7	18 (40.0)	77 (51.7)	0.62 (0.32–1.23)	0.1716	0.52 (0.25–1.05)	0.0686	20 (39.2)	170 (50.4)	0.63 (0.35–1.16)	0.1372
pTstage										
≤pT2b	12 (26.7)	34 (22.8)	Ref.	Ref.	Ref.	Ref.	14 (27.5)	78 (23.1)	Ref.	Ref.
≥pT2c	33 (73.3)	115 (77.2)	0.81 (0.38–1.74)	0.5952	0.84 (0.38–1.81)	0.6492	37 (72.5)	259 (76.9)	0.80 (0.41–1.55)	0.5011
rs1094112	GA + AA (%)	GG (%)					A (%)	G (%)		
Gleason score										
<7	54 (43.2)	45 (65.2)	Ref.	Ref.	Ref.	Ref.	70 (45.5)	128 (54.7)	Ref.	Ref.
≥7	71 (56.8)	24 (34.8)	2.47 (1.34–4.53)	0.0037	2.28 (1.22–4.29)	0.0101	84 (54.5)	106 (45.3)	1.45 (0.96–2.18)	0.0751
pTstage										
≤pT2b	22 (17.6)	24 (34.8)	Ref.	Ref.	Ref.	Ref.	28 (18.2)	64 (27.4)	Ref.	Ref.
≥pT2c	103 (82.4)	45 (65.2)	2.50 (1.27–4.91)	0.0080	2.60 (1.30–5.21)	0.0071	126 (81.8)	170 (72.6)	1.69 (1.03–2.79)	0.0389
rs195676	GA + AA (%)	GG (%)					A (%)	G (%)		
Gleason score										
<7	54 (43.9)	45 (65.2)	Ref.	Ref.	Ref.	Ref.	70 (46.1)	128 (55.2)	Ref.	Ref.
≥7	69 (56.1)	24 (34.8)	2.40 (1.30–4.41)	0.0050	2.35 (1.26–4.41)	0.0076	82 (53.9)	104 (44.8)	1.44 (0.96–2.17)	0.0808
pTstage										
≤pT2b	22 (17.9)	24 (34.8)	Ref.	Ref.	Ref.	Ref.	28 (18.4)	64 (27.6)	Ref.	Ref.
≥pT2c	101 (82.1)	45 (65.2)	2.45 (1.24–4.82)	0.0095	2.66 (1.33–5.34)	0.0057	124 (81.6)	168 (72.4)	1.69 (1.02–2.78)	0.0408

pTstage: pathologic stage; OR: odds ratio; 95% CI: 95% confidence interval; AMACR: (R)-alpha-methyl-CoA racemase.
^a Adjusted for age, smoking quantity, alcohol drinking, and family history of prostate cancer.

TABLE 4: Association analysis of AMACR (5p13) polymorphisms with AMACR expressing prostate cancer risk.

SNP	Genotype	Cases (%)	Controls (%)	Crude OR (95%CI)	P	Adjusted OR (95%CI) ^a	P	P _{trend} [†]
rs2278008	A/A	120 (77.9)	109 (64.9)	Ref.		Ref.		0.0088
	A/G	33 (21.4)	56 (33.3)	0.54 (0.32–0.88)	0.0147	0.48 (0.26–0.89)	0.0207	
	G/G	1 (0.6)	3 (1.8)	0.30 (0.03–2.95)	0.3040	0.37 (0.04–3.92)	0.4091	
	A/G + G/G	34 (22.1)	59 (35.1)	0.52 (0.32–0.86)	0.0104	0.47 (0.26–0.87)	0.0167	
	A allele	246 (87.9)	274 (81.5)	Ref.		Ref.		
rs34677	G allele	34 (12.1)	62 (18.5)	0.61 (0.39–0.96)	0.0327	0.59 (0.34–1.02)	0.0598	
	G/G	113 (73.4)	123 (73.2)	Ref.		Ref.		0.7336
	G/T	36 (23.4)	43 (25.6)	0.91 (0.55–1.52)	0.7217	1.33 (0.66–2.67)	0.4217	
	T/T	5 (3.2)	2 (1.2)	2.72 (0.52–14.29)	0.2373	2.28 (0.29–8.15)	0.4354	
	G/T + T/T	41 (26.6)	45 (26.8)	0.99 (0.60–1.63)	0.9738	1.40 (0.71–2.73)	0.3300	
rs2287939	G allele	239 (85.4)	289 (86.0)	Ref.		Ref.		
	T allele	41 (14.6)	47 (14.0)	1.05 (0.67–1.66)	0.8169	1.35 (0.72–2.50)	0.3479	
	C/C	122 (79.2)	124 (73.8)	Ref.		Ref.		0.2589
	C/T	30 (19.5)	41 (24.4)	0.74 (0.44–1.27)	0.2763	0.72 (0.37–1.38)	0.3207	
	T/T	2 (1.3)	3 (1.8)	0.68 (0.11–4.13)	0.6728	0.50 (0.05–4.88)	0.5550	
rs10941112	C/T + T/T	32 (20.8)	44 (26.2)	0.74 (0.44–1.24)	0.2542	0.70 (0.37–1.33)	0.2775	
	C allele	247 (88.2)	289 (86.0)	Ref.		Ref.		
	T allele	33 (11.8)	47 (14.0)	0.82 (0.51–1.32)	0.4186	0.79 (0.44–1.42)	0.4274	
	G/G	58 (37.7)	76 (45.2)	Ref.		Ref.		0.4154
	G/A	74 (48.1)	66 (39.3)	1.47 (0.91–2.37)	0.1134	1.45 (0.79–2.66)	0.2322	
rs3195676	A/A	22 (14.3)	26 (15.5)	1.11 (0.57–2.15)	0.7601	1.08 (0.45–2.57)	0.8655	
	G/A + A/A	96 (62.3)	92 (54.8)	1.37 (0.88–2.13)	0.1688	1.35 (0.76–2.38)	0.3041	
	G allele	107 (38.2)	118 (35.1)	Ref.		Ref.		
	A allele	173 (61.8)	218 (64.9)	1.14 (0.82–1.59)	0.4271	1.12 (0.73–1.71)	0.6157	
	G/G	58 (37.9)	74 (44.0)	Ref.		Ref.		0.5202
rs3195676	G/A	73 (47.7)	68 (40.5)	1.37 (0.85–2.21)	0.1959	1.34 (0.73–2.46)	0.3503	
	A/A	22 (14.4)	26 (15.5)	1.08 (0.56–2.10)	0.8211	1.03 (0.43–2.46)	0.9503	
	G/A + A/A	95 (62.1)	94 (56.0)	1.29 (0.82–2.02)	0.2646	1.25 (0.71–2.22)	0.4360	
	G allele	106 (38.1)	120 (35.7)	Ref.		Ref.		
	A allele	172 (61.9)	216 (64.3)	1.11 (0.80–1.54)	0.5364	1.07 (0.70–1.64)	0.7577	

SNP: single nucleotide polymorphisms; OR: odds ratio; CI: confidence interval.

^aAdjusted for age, smoking quantity, alcohol drinking, and family history of prostate cancer.

[†]Cochran-Armitage trend test for the number of variant alleles.

reclassified into two subgroups according to expression level: a negative group (score 0) versus a positive group (score 1+, 2+ or 3+). There were 39 cases (20.2%) in the negative group and 154 cases (79.8%) in the positive group (Table 1). Between variants of AMACR and intensity of AMACR expression, no significant relation was found except for rs2287939 (Supplemental Table 4). In the case group expressing AMACR protein, individuals with the AG or GG genotype of rs2278008 showed a decreased prostate cancer risk compared to those with the AA genotype (adjusted OR 0.47; 95% CI, 0.26–0.87, $P = 0.017$; Table 4 and supplemental figure).

Considering the clinical features, the AG or GG genotype of rs2278008 showed a significantly decreased risk of prostate cancer with GS ≥ 7 (adjusted OR 0.43; 95% CI, 0.19–0.99,

$P = 0.048$). Additionally, the GT or TT genotype of rs34677 was associated with a statistically significant decreased risk for patients with $\geq pT2c$ (adjusted OR 0.35; 95% CI, 0.15–0.79, $P = 0.012$). Also, the patients with GA or AA genotype of rs10941112 and rs3195676 revealed to have a significantly increased risk of developing unfavorable prostate cancer with $\geq pT2c$ (Table 5).

3.5. Haplotype Analysis of AMACR Polymorphism. The whole frequency distribution and association analysis of haplotypes for SNPs in AMACR are summarized in Table 6 (in Supplemental Table 5). When the haplotypes consisted of all five cSNPs, individuals with the 12345-5 [GGCGG] haplotype showed a significantly lower risk of prostate cancer compared

TABLE 5: Association between AMACR (5p13) polymorphisms and prognostic factors in prostate cancer patients in AMACR expressing subgroup.

rs2278008	AG + GG (%)	AA (%)	OR (95% CI)	P	Adjusted OR (95% CI) ^a	P	OR (95% CI)	P	Adjusted OR (95% CI) ^a	P
Gleason score										
<7	23 (67.6)	59 (49.2)					22 (64.7)	130 (52.8)		
≥7	11 (32.4)	61 (50.8)	0.46 (0.21-1.03)	0.0598	0.43 (0.19-0.99)	0.0482	12 (35.3)	116 (47.2)	0.61 (0.29-1.29)	0.1963
pTstage										
≤pT2b	7 (20.6)	28 (23.3)					7 (20.6)	53 (21.5)		
≥pT2c	27 (79.4)	92 (76.7)	1.17 (0.46-2.98)	0.7362	1.24 (0.48-3.18)	0.6530	27 (79.4)	193 (78.5)	1.06 (0.44-2.57)	0.8986
rs34677										
Gleason score										
<7	21 (51.2)	61 (54.0)					21 (51.2)	131 (54.8)		
≥7	20 (48.8)	52 (46.0)	1.12 (0.55-2.28)	0.7614	1.20 (0.57-2.53)	0.6249	20 (48.8)	108 (45.2)	1.16 (0.60-2.24)	0.6698
pTstage										
≤pT2b	15 (36.6)	20 (17.7)					14 (34.1)	46 (19.2)		
≥pT2c	26 (63.4)	93 (82.3)	0.37 (0.17-0.83)	0.0154	0.35 (0.15-0.79)	0.0119	27 (65.9)	193 (80.8)	0.46 (0.22-0.95)	0.0346
rs2287939										
Gleason score										
<7	19 (59.4)	63 (51.6)					19 (57.6)	133 (53.8)		
≥7	13 (40.6)	59 (48.4)	0.73 (0.33-1.61)	0.4360	0.62 (0.27-1.44)	0.2697	14 (42.4)	114 (46.2)	0.86 (0.41-1.79)	0.6865
pTstage										
≤pT2b	7 (21.9)	28 (23.0)					7 (21.2)	53 (21.5)		
≥pT2c	25 (78.1)	94 (77.0)	1.06 (0.42-2.72)	0.8972	1.07 (0.41-2.77)	0.8944	26 (78.8)	194 (78.5)	1.01 (0.42-2.47)	0.9743
rs10941112										
Gleason score										
<7	46 (47.9)	36 (62.1)					55 (51.4)	97 (56.1)		
≥7	50 (52.1)	22 (37.9)	1.78 (0.92-3.46)	0.0895	1.73 (0.87-3.45)	0.1165	52 (48.6)	76 (43.9)	1.21 (0.74-1.96)	0.4464
pTstage										
≤pT2b	15 (15.6)	20 (34.5)					15 (14.0)	45 (26.0)		
≥pT2c	81 (84.4)	38 (65.5)	2.84 (1.31-6.15)	0.0080	2.97 (1.35-6.52)	0.0066	92 (86.0)	128 (74.0)	2.16 (1.13-4.10)	0.0191
rs3195676										
Gleason score										
<7	46 (48.4)	36 (62.1)					55 (51.9)	97 (56.4)		
≥7	49 (51.6)	22 (37.9)	1.74 (0.90-3.39)	0.1019	1.71 (0.86-3.40)	0.1289	51 (48.1)	75 (43.6)	1.20 (0.74-1.95)	0.4635
pTstage										
≤pT2b	15 (15.8)	20 (34.5)					15 (14.2)	45 (26.2)		
≥pT2c	80 (84.2)	38 (65.5)	2.81 (1.30-6.08)	0.0089	2.94 (1.34-6.45)	0.0071	91 (85.8)	127 (73.8)	2.15 (1.13-4.09)	0.0197

pTstage: pathologic stage; OR: odds ratio; 95% CI: 95% confidence interval.
^a Adjusted for age, smoking quantity, alcohol drinking, and family history of prostate cancer.

TABLE 6: AMACR (5p13) haplotypes and their association with prostate cancer risk.

Haplotype*	Cases (%)	Controls (%)	Crude OR (95% CI)	P value	Adjusted OR(95% CI) ^a	P value
12345-1[AGCAA]	147 (37.9)	116 (34.3)	Ref.		Ref.	
12345-2[AGCGG]	116 (29.9)	103 (30.5)	0.89 (0.62–1.27)	0.5208	0.95 (0.60–1.50)	0.8205
12345-3[ATCGG]	56 (14.4)	47 (13.9)	0.94 (0.60–1.49)	0.7919	1.14 (0.62–2.10)	0.6743
12345-4[GGTGG]	33 (8.5)	39 (11.5)	0.67 (0.40–1.13)	0.1306	0.59 (0.31–1.13)	0.1108
12345-5[GGCGG]	11 (2.8)	20 (5.9)	0.43 (0.20–0.94)	0.0348	0.41 (0.17–1.00)	0.0489
12345-6[AGTGG]	18 (4.6)	9 (2.7)	1.58 (0.68–3.64)	0.2849	0.97 (0.35–2.74)	0.9603
12345-7[GGCAA]	7 (1.8)	3 (0.9)	1.84 (0.47–7.28)	0.3839	1.54 (0.28–8.36)	0.6190
Haplotype pair						
12-2 [GG]						
0 copies	145 (74.7)	110 (65.1)	Ref.		Ref.	
1 or 2 copies	49 (25.3)	59 (34.9)	0.63 (0.40–0.99)	0.0455	0.51 (0.29–0.90)	0.0199
14-3 [GG]						
0 copies	153 (78.9)	112 (66.3)	Ref.		Ref.	
1 or 2 copies	41 (21.1)	57 (33.7)	0.53 (0.33–0.84)	0.0074	0.43 (0.24–0.77)	0.0047
15-3 [GG]						
0 copies	150 (77.3)	112 (66.3)	Ref.		Ref.	
1 or 2 copies	44 (22.7)	57 (33.7)	0.58 (0.36–0.92)	0.0197	0.47 (0.27–0.84)	0.0104
145-3 [GGG]						
0 copies	150 (77.3)	112 (66.3)	Ref.		Ref.	
1 or 2 copies	44 (22.7)	57 (33.7)	0.58 (0.36–0.92)	0.0197	0.47 (0.27–0.84)	0.0104
1245-4 [GGGG]						
0 copies	151 (77.8)	113 (66.9)	Ref.		Ref.	
1 or 2 copies	43 (22.2)	56 (33.1)	0.57 (0.36–0.92)	0.0199	0.48 (0.27–0.86)	0.0132
1345-4 [GCGG]						
0 copies	182 (93.8)	148 (87.6)	Ref.		Ref.	
1 or 2 copies	12 (6.2)	21 (12.4)	0.46 (0.22–0.98)	0.0428	0.40 (0.17–0.95)	0.0379
12345-5 [GGCGG]						
0 copies	183 (94.3)	149 (88.2)	Ref.		Ref.	
1 or 2 copies	11 (5.7)	20 (11.8)	0.45 (0.21–0.96)	0.0400	0.41 (0.17–0.99)	0.0472

Haplotypes with total frequencies of less than 1 percent were excluded in table.

OR: odds ratio; CI: confidence interval; AMACR: (R)-alpha-methyl-CoA racemase.

*1: rs2278008; 2: rs34677; 3: rs2287939; 4: rs10941112; 5: rs3195676.

^aAdjusted for age, smoking quantity, alcohol drinking, and family history of prostate cancer.

to the subjects with the most frequent haplotype 12345-1 [AGCAA] (adjusted OR, 0.41; 95% CI, 0.17–1.00, $P = 0.049$). Furthermore, individuals with one or more copies of the [GGCGG] tended to have a significantly decreased risk of prostate cancer compared to those without [GGCGG] haplotype (adjusted OR, 0.41; 95% CI, 0.17–0.99, $P = 0.047$). Interestingly, when we considered the haplotype pair of each individual, the haplotype pair harboring the G allele of SNP rs2278008 revealed a lowered risk in combination with the G allele of rs34677, rs1094112, and rs3195676 (Table 6).

4. Discussion

AMACR is a key enzyme in the β -oxidation catabolic pathway of fatty acids and is known to be upregulated in several cancers including prostate cancer [23–26]. Sequence variants of AMACR have been previously investigated to find their association with prostate cancer risk [10–12, 27, 28]. Results, however, were inconsistent: One study suggested

that variants in AMACR gene were associated with familial, but not sporadic, prostate cancer [10] and other subsequent studies reported no association between sporadic prostate cancer and AMACR gene variants [18, 19] (<http://dceg.cancer.gov/research/how-we-study/genomic-studies/cgems-summary/>). In the subgroup analysis of one of these studies, a tendency toward decreased risk in homozygous white carriers of the variant alleles for both rs3195676 and rs10941112 was evident [19]. Thus, genetic studies about the association of genetic polymorphisms with prostate cancer risk might show different results depending on the ethnic background of the subjects. The use of Korean men, who are reported as an ethnically homogenous population [21], may deliver useful conclusion for the association of AMACR polymorphisms with prostate cancer risk, by reducing the possible influences related to genetic heterogeneity. Here, we found statistically significant association of AMACR polymorphisms with prostate cancer risk using single ethnic Koreans without any family history of immigration from other countries. Further,

we analyzed the risk of prostate cancer in relation with AMACR polymorphism in cases of AMACR expression.

In our study, rs2278008 (E277K) was associated with prostate cancer risk in ethnically homogenous Koreans with statistical significance. To our knowledge, this is the first report of a protective effect of rs2278008 of the AMACR gene on the development of sporadic prostate cancer. Although, rs2278008 SNP of AMACR was reported previously with the significantly different genotype frequencies between hereditary prostate cancer groups and controls, this tendency was not shown when compared with a sporadic prostate cancer group [10]. In our study, the GG or AG genotype of rs2278008 was more frequently observed in the control group when compared with the sporadic prostate cancer group. Furthermore, in subjects expressing AMACR, the association of rs2278008 with prostate cancer risk also showed statistical significance. With respect to the association of the gene variant with clinical factors of prostate cancer, recent data as part of the Michigan Prostate Cancer Genetics Project demonstrated a significant linkage between markers on 5p13-q11 and prostate cancer aggressiveness, as defined by GS [29]. These observations suggest that this region of the genome may harbor sequence variations associated with prostate cancer risk and the extent of tumor differentiation may be considered as a predictor for prognosis. We also found genetic variations of AMACR related to the histologic grade of prostate cancer. Patients with a specific sequence variant of rs2278008 and rs34677 tended to show a decreased risk of developing unfavorable prostate cancer with GS ≥ 7 or \geq pT2c. On the contrary, patients with the GA or AA genotype of rs10941112 and rs3195676 displayed a significantly increased risk of developing unfavorable prostate cancer. Based on this result, the genetic polymorphism of AMACR might influence the development of prostate cancer for patients expressing AMACR. Additionally, haplotype analysis revealed the combinatorial impact of individual SNPs clearly. The protective effect of rs2278008 may strongly influence cancer susceptibility with the risk decreased by the effect of rs34677, rs10941112, or rs3195676, since the G allele of rs2278008 was mostly revealed as statistically significant and the protective effects in any combination with G allele of rs34677, rs10941112, or rs3195676, but protective effects were not present with the others. This is the first report ever for the analysis of the combinatorial effect of AMACR SNPs on the prostate cancer susceptibility in haplotype.

Unlike other cancers, prostate cancer cells use fatty acid as their main energy source instead of glucose [30–32]. AMACR is required to enter into the fatty acid oxidation, resulting in energy production for the cancer cells [30, 31]. Combining our discovery (the protective role of rs2278008 in AMACR), the missense mutation by the minor allele of the sequence variant might disrupt or decrease the enzyme (AMACR) activity, subsequently leading to unfavorable energy supply in the cancer cells. The speculation can explain the lower prostate cancer risk for the missense mutation-harboring patients, which needs to be further studied. In addition, three-dimensional structural data implicate that the substitution encoded by rs1094112 (G175D) may directly impact the stability of the AMACR protein backbone [33]. Otherwise,

no studies have investigated the impact of genetic variations on characteristics of prostate cancer by changing functional structure.

Despite the new findings that the sequence variants of AMACR are related to the risk of sporadic prostate cancer in the ethnically homogenous population of Korean men, caution should be taken when interpreting our findings due to some limitations. Firstly, because of the relatively small number of cases and controls in this study, the power of this study is limited. Particularly, stratified analyses with respect to clinical factors of prostate cancer might be underpowered. By using PGA method, the SNPs in our study were calculated about 5%~ about 43% power at an σ value of 5%, although it has low power, and statistically significant finding was detected. Instead of each value for statistical power, we just presented wide range of statistical power because of many subgroups including clinical factors of prostate cancer. Secondly, the patients were older and consumed less alcohol than the controls, and the number of patients having a family history of prostate cancer (not first-degree relatives) was more than the controls, although it was not statistically significant. To minimize the effect of the different distribution in cases and controls, we adjusted the age, smoking quantity, alcohol drinking, and family history of prostate cancer in statistical analysis. Furthermore, our primary interest lay in the genotype and its association with prostate cancer risk, and it is unlikely that the genotype is affected by age. Thus, we believe that the bias possibly caused by different age distributions among cases and controls would minimally influence for the results of our study.

5. Conclusions

We concluded that the genetic variations of AMACR were associated with the risk of sporadic prostate cancer treated with radical prostatectomy in ethnically homogenous population of Korean men. Our investigation of the relationship between genotype frequencies, haplotype pair, and the expression of AMACR protein in tumors with clinical features demonstrates that sequence variants of AMACR may play an important role in the development of prostate cancer.

Abbreviations

AMACR:	(R)-Alpha-methyl-CoA racemase
SNP:	Single nucleotide polymorphism
GWAS:	Genome-wide association study
HWE:	Hardy-Weinberg Equilibrium
CIs:	Confidence intervals
LD:	Linkage disequilibrium.

Conflict of Interests

The authors declare that they have no conflict of interests.

Authors' Contribution

Sang-Jin Lee and Jae Young Joung equally contributed as first authors.

Acknowledgments

This study was supported by the Korean National Cancer Center Grants (nos. 1210110 and 0910221). All experiments were performed at the Genomics Core Facility in the National Cancer Center. This treatise was also supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (20110010731).

References

- [1] J. Y. Joung, Y.-S. Lee, S. Park et al., "Haplotype analysis of prostate stem cell antigen and association with prostate cancer risk," *Journal of Urology*, vol. 185, no. 6, pp. 2112–2118, 2011.
- [2] J. Y. Joung, S. Park, H. Yoon et al., "Association of common variations of 8q24 with the risk of prostate cancer in Koreans and a review of the Asian population," *BJU International*, vol. 110, no. 6, part B, pp. E318–E325, 2012.
- [3] S. Perner, F. H. Schmidt, M. D. Hofer, R. Kuefer, and M. Rubin, "TMPRSS2-ETS gene fusion in prostate cancer," *Urologe A*, vol. 46, no. 7, pp. 754–760, 2007.
- [4] J. P. Struwing, P. Hartge, S. Wacholder et al., "The risk of cancer associated with specific mutations of BRCA1 and BRCA2 among Ashkenazi Jews," *The New England Journal of Medicine*, vol. 336, no. 20, pp. 1401–1408, 1997.
- [5] Z. Kote-Jarai, A. Amin Al Olama, D. Leongamornlert et al., "Identification of a novel prostate cancer susceptibility variant in the KLK3 gene transcript," *Human Genetics*, vol. 129, no. 6, pp. 687–694, 2011.
- [6] S. L. Zheng, J. Sun, F. Wiklund et al., "Cumulative association of five genetic variants with prostate cancer," *The New England Journal of Medicine*, vol. 358, no. 9, pp. 910–919, 2008.
- [7] A. M. Levin, M. J. Machiela, K. A. Zuhlke, A. M. Ray, K. A. Cooney, and J. A. Douglas, "Chromosome 17q12 variants contribute to risk of early-onset prostate cancer," *Cancer Research*, vol. 68, no. 16, pp. 6492–6495, 2008.
- [8] J. Gudmundsson, P. Sulem, D. F. Gudbjartsson et al., "Genome-wide association and replication studies identify four variants associated with prostate cancer susceptibility," *Nature Genetics*, vol. 41, no. 10, pp. 1122–1126, 2009.
- [9] M. B. Ishak and V. N. Giri, "A systematic review of replication studies of prostate cancer susceptibility genetic variants in high-risk men originally identified from genome-wide association studies," *Cancer Epidemiology Biomarkers and Prevention*, vol. 20, no. 8, pp. 1599–1610, 2011.
- [10] S. L. Zheng, B.-L. Chang, D. A. Faith et al., "Sequence variants of α -methylacyl-CoA racemase are associated with prostate cancer risk," *Cancer Research*, vol. 62, no. 22, pp. 6485–6488, 2002.
- [11] A. M. Levin, K. A. Zuhlke, A. M. Ray, K. A. Cooney, and J. A. Douglas, "Sequence variation in α -methylacyl-CoA racemase and risk of early-onset and familial prostate cancer," *Prostate*, vol. 67, no. 14, pp. 1507–1513, 2007.
- [12] J. L. Wright, M. L. Neuhauser, D. W. Lin et al., "AMACR polymorphisms, dietary intake of red meat and dairy and prostate cancer risk," *Prostate*, vol. 71, no. 5, pp. 498–506, 2011.
- [13] S. Zha, S. Ferdinandusse, S. Denis et al., " α -Methylacyl-CoA Racemase as an Androgen-Independent Growth Modifier in Prostate Cancer," *Cancer Research*, vol. 63, no. 21, pp. 7365–7376, 2003.
- [14] C. Prior, F. Guillen-Grima, J. E. Robles et al., "Use of a combination of biomarkers in serum and urine to improve detection of prostate cancer," *World Journal of Urology*, vol. 28, no. 6, pp. 681–686, 2010.
- [15] L. Häggarth, C. Hägglöf, S. J. Jaraj et al., "Diagnostic biomarkers of prostate cancer," *Scandinavian Journal of Urology and Nephrology*, vol. 45, no. 1, pp. 60–67, 2011.
- [16] B. Ouyang, B. Bracken, B. Burke, E. Chung, J. Liang, and S.-M. Ho, "A duplex quantitative polymerase chain reaction assay based on quantification of α -methylacyl-CoA racemase transcripts and prostate cancer antigen 3 in urine sediments improved diagnostic accuracy for prostate cancer," *Journal of Urology*, vol. 181, no. 6, pp. 2508–2514, 2009.
- [17] J. N. Mubiru, A. J. Valente, and D. A. Troyer, "A variant of the alpha-methyl-acyl-CoA racemase gene created by a deletion in exon 5 and its expression in prostate cancer," *Prostate*, vol. 65, no. 2, pp. 117–123, 2005.
- [18] S. Lindström, S. L. Zheng, F. Wiklund et al., "Systematic replication study of reported genetic associations in prostate cancer: strong support for genetic variation in the androgen pathway," *Prostate*, vol. 66, no. 16, pp. 1729–1743, 2006.
- [19] S. E. Daugherty, Y. Y. Shugart, E. A. Platz et al., "Polymorphic variants in α -methylacyl-CoA racemase and prostate cancer," *Prostate*, vol. 67, no. 14, pp. 1487–1497, 2007.
- [20] L. M. FitzGerald, R. Thomson, A. Polanowski et al., "Sequence variants of α -methylacyl-CoA racemase are associated with prostate cancer risk: a replication study in an ethnically homogeneous population," *Prostate*, vol. 68, no. 13, pp. 1373–1379, 2008.
- [21] M. A. Abdulla, I. Ahmed, A. Assawamakin et al., "Mapping human genetic diversity in Asia," *Science*, vol. 326, no. 5959, pp. 1541–1545, 2009.
- [22] I. Menashe, P. S. Rosenberg, and B. E. Chen, "PGA: power calculator for case-control genetic association analyses," *BMC Genetics*, vol. 9, article 36, 2008.
- [23] S. Ferdinandusse, S. Denis, P. T. Clayton et al., "Mutations in the gene encoding peroxisomal α -methylacyl-CoA racemase cause adult-onset sensory motor neuropathy," *Nature Genetics*, vol. 24, no. 2, pp. 188–191, 2000.
- [24] J. Xu, J. A. Stolk, X. Zhang et al., "Identification of differentially expressed genes in human prostate cancer using subtraction and microarray," *Cancer Research*, vol. 60, no. 6, pp. 1677–1682, 2000.
- [25] J. Luo, D. J. Duggan, Y. Chen et al., "Human prostate cancer and benign prostatic hyperplasia: Molecular dissection by gene expression profiling," *Cancer Research*, vol. 61, no. 12, pp. 4683–4688, 2001.
- [26] J. Luo, S. Zha, W. R. Gage et al., " α -methylacyl-CoA racemase: a new molecular marker for prostate cancer," *Cancer Research*, vol. 62, no. 8, pp. 2220–2226, 2002.
- [27] C. Zhang, R. Montironi, G. T. MacLennan et al., "Is atypical adenomatous hyperplasia of the prostate a precursor lesion?" *Prostate*, vol. 71, no. 16, pp. 1746–1751, 2011.
- [28] S. Minner, M. Enodien, H. Sirma et al., "ERG status is unrelated to PSA recurrence in radically operated prostate cancer in the absence of antihormonal therapy," *Clinical Cancer Research*, vol. 17, no. 18, pp. 5878–5888, 2011.
- [29] S. L. Slager, K. E. Zarfes, W. M. Brown et al., "Genome-wide linkage scan for prostate cancer aggressiveness loci using families from the University of Michigan prostate cancer genetics project," *Prostate*, vol. 66, no. 2, pp. 173–179, 2006.

- [30] Y. Liu, "Fatty acid oxidation is a dominant bioenergetic pathway in prostate cancer," *Prostate Cancer and Prostatic Diseases*, vol. 9, no. 3, pp. 230–234, 2006.
- [31] G. Zadra, C. Photopoulos, and M. Loda, "The fat side of prostate cancer," *Biochim Biophys Acta*, vol. 1831, no. 10, pp. 1518–1532, 2013.
- [32] A. Schulze and A. L. Harris, "How cancer metabolism is tuned for proliferation and vulnerable to disruption," *Nature*, vol. 491, no. 7424, pp. 364–373, 2012.
- [33] P. Yue and J. Moulton, "Identification and analysis of deleterious human SNPs," *Journal of Molecular Biology*, vol. 356, no. 5, pp. 1263–1274, 2006.