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Ethnobotanical survey, anthelmintic effects and cytotoxicity of plants used for treatment of helminthiasis in the Central and Kara regions of Togo



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Abstract

Background: Traditional medicines are the main source of treatment of helminthiasis in endemic areas of Togo. The present study aimed to investigate the plants used by Traditional healers (THs) to treat helminth infections in endemic communities within the Central and Kara regions of Togo and to evaluate the anthelmintic activity of the three most cited plants.

Methods: An ethnobotanical survey was conducted from 19 to 24 June 2017 among traditional healers in the Central and Kara regions of Togo. The anthelmintic activity of the most cited plants namely *Aframomum melegueta* K. Schum, *Khaya senegalensis* A. Juss and *Xylopia aethiopica* A. Rich, was evaluated using microfilariae (Mf) of *Litomosoides sigmodontis*. The plants were evaluated for cytotoxicity according to the recommendation of NF EN ISO 10993-5 standard using the propidium iodide (PI) dye by flow cytometry on human peripheral blood mononuclear cells.

Results: A total of 197 THs were interviewed and 41 plant species were recorded. Leguminosae (14.6%) and Annonaceae (9.7%) families constitute the highest number of species cited for treatment of helminth infections. Afromomum melegueta was the most cited by the THs for the treatment of onchocerciasis (UV = 0.036) while X. aethiopica was associated with the treatment of schistosomiasis (UV = 0.061) and lymphatic filariasis (UV = 0.061). There was a great agreement among the THs regarding ethnomedicinal uses of plants to treat helminthiasis with ICF values ranging from 0.57 to 0.67. The anthelmintic assay yielded lethal doses values of 233 μ g/mL, 265 μ g/mL and 550 μ g/mL, respectively for X. aethiopica, A. melegueta and K. senegalensis. Afromomum melegueta and X. aethiopica presented no cytotoxicity, less than 20% death, whereas K. senegalensis induced moderate toxicity, 24 \pm 8% death.

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Conclusion: This study demonstrated the scientific rationale for the use of plants to treat helminthiasis in the Togolese traditional medicine. However, the use of *K. senegalensis* requires more caution since the plant is fairly toxic.

Trial Registration: NA

Keywords: Anthelmintic effects, Ethnopharmacology, Medicinal plants, Togo

Background

The emergence of resistance to anthelmintics makes it difficult to control helminth infections in endemic areas. One of the solution approaches is the search for new molecules and the development of effective therapies, affordable for low-income people, since the populations affected are leaving in developing countries. Many studies are already devoted to this, and even a new method of screening for filaricidal agents has recently been developed [1]. In this new approach, plants from the traditional pharmacopoeia have demonstrated proven anthelmintic effects [2].

Herbal medicines have been the source of many of the drugs prescribed today in modern medicine. Some examples are, aspirin from Salix alba [3], digitoxin from Digitalis [4], artemisinin from Artemisia annua [5]. Medicinal plants are precious resources in lowincome countries and more than 80% of African populations use them for health problems [6]. Therefore, the use of plant organs to heal is a question of culture and tradition in Africa [7, 8]. Understanding the properties and value of unprocessed raw medicinal plant materials is a national heritage for these countries [9]. It should be noted that for primary health care needs, a large part of the African population still turns to traditional medicine which is mainly based on herbal remedies. This is due in part to the preference and confidence of local healers over the health care system [10].

Chronic infections with helminths namely Onchocerca volvulus, Wuchereria bancrofti and Schistosoma haematobium induce diseases in endemic areas of Togo. The absence of vaccines, the constant exposure and the possibilities of reinfection with these helminths present a constant socio-economic problem and an increase in DALYs (disability-adjusted life years). Togo has an excellent biodiversity of medicinal plants used in traditional medicine for the treatment of many diseases. Thus, many traditional remedies have been developed by the practitioners of traditional medicine to treat helminth infections. However, scientific data on these herbal therapies are missing. The present study was initiated with the aim of documenting the plants usage in the treatment of helminth infections by traditional healers (TH) in endemic communities in the Central and Kara regions of Togo and in assessing their anthelmintic and cytotoxic effects in vitro.

Methods

Study area

The ethnobotanical survey was undertaken in the Central and Kara regions of Togo (Fig. 1). Togo is a West African country boarded in the North by the Republic of Burkina Faso, the Est. by the Republic of Benin, the West by the Republic of Ghana and the South by the Atlantic Ocean. From north to south the country is organized into five economic regions: the Savannah region, the Kara region, the Central region, the Plateaux region and the Maritime region. The Central and Kara regions belong to the tropical area with a dry season from October to March and a rainy season from April to September. The annual temperatures are between 20 and 39 °C, providing an excellent floristic biodiversity with numerous medicinal plants. The principal activities of the population are agriculture and trade [10].

Data collection

In total, 197 THs (136 from central region and 61 from Kara region) were interviewed from the 19th to 24th June 2017 using a structured questionnaire, after their informed consent and their agreement with a signature. The THs belong to the Tem tribe and Kabyè tribe, and all of them speak at least one local language, Kotokoli or Kabyè in which interviews were conducted. They were all members of the non-governmental organization (NGO) named "Centre d'Etude et de Recherche en Médecine Traditionnelle Appliquée du Togo" (CERME-TRA) (http://tg.viadeo.com/fr/profile/cermetra.ong). CERMETRA contributes to the training and counselling of the THs on patient management and environmental preservation, mainly protecting vulnerable and endangered plant species used in traditional medicine. For example, the harvest of the leaves of Pterocarpus erinaceus which is a species included in the red list of the IUCN (https://www.iucnredlist.org/species/62027797/6202 7800), is rigorously supervised by CERMETRA after the authorisation of the forestry services to avoid the removal of vital organs such as stem bark and roots. The main activity that threatens extinction *P. erinaceus* is the exploitation of its wood, it is not really the use in

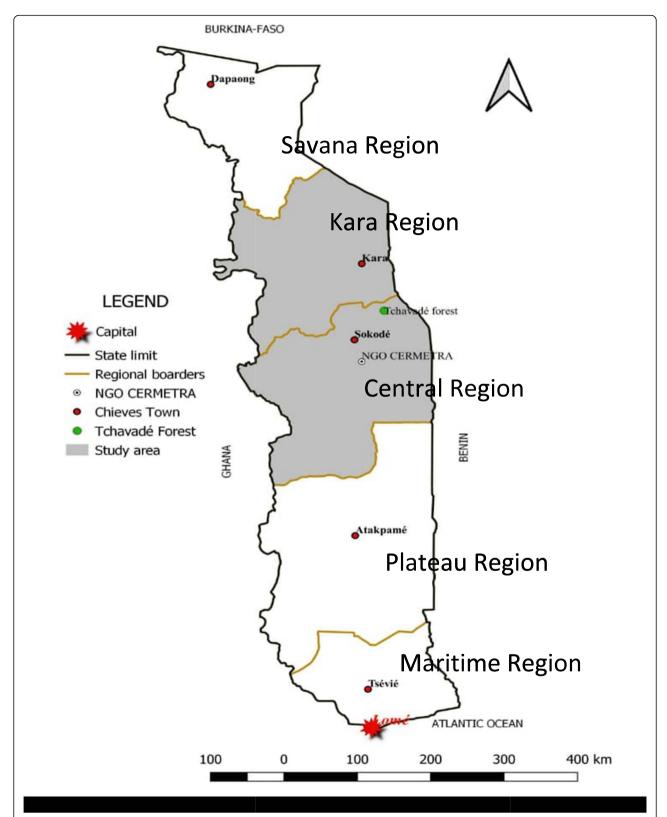


Fig. 1 Map of Togo showing the study area. The study was conducted in the Central and Kara regions of Togo. THs usually gather plant species for their medicines in the Tchavedè Forest. Various plant organs were collected for botanical identification

traditional medicine, which moreover focuses mainly on the leaves and not the vital organs. Actions carried out in the country therefore aim to limit the exploitation of its wood and reforestation. Thus, the University of Lomé through the Faculty of Sciences is very active in revitalizing the populations of P. erinaceus. The University is a partner of the "South Expert Plants Sustainable Development" (SEP2D) multilateral program. The challenge of this program is to strengthen interactions and partnerships in terms of plant biodiversity between research, teaching and society's demands. In the particular case of P. erinaceus, proposals have been made in order to provide the elements of responses essential to the adoption of sustainable silvicultural practices which should allow a rapid reconstitution of populations in West Africa. It specifically involves (i) analyzing the biophysical and socio-cultural factors of the multiplication and domestication of P. erinaceus in 3 countries in West Africa (Togo, Benin and Niger), (ii) studying the variability in the structural and technological characteristics of the species' wood in relation to environmental conditions, (iii) developing a viable and large-scale production strategy for P. erinaceus plants for reforestation in West Africa and (iv) strengthen the capacities of stakeholders to regenerate and sustainably manage stands of P. erinaceus. http://www.sep2d.org/projets-soutenus/rechercheoperationnelle/reconstitution-peuplement-pterocarpustogo-benin-niger.

After the interview, 41 of the most commonly plant species used by THs to treat helminthiasis were collected. Since the THs often referred to these plants in a colloquial manner, samples of all 41 plants were collected with members of CERMETRA (actually THs) in the Tchavadè forest. Verification and identification of all collected plant specimens were carried out at the Botanic Laboratory of the Faculty of Sciences at the University of Lomé. Plant taxonomy was confirmed on data available from the International Plant Names Index (IPNI) website: http://www.ipni.org/. Specimen of each plants was deposited at the herbarium of the University of Lomé.

Preparation of plants extracts

All the plants materials were collected from the Tchavadè forest at Sokodé in the central Region with Traditional healers. The grains of A. melegueta, bark of K. senegalensis and fruits of X. aethiopica were washed and air-dried in laboratory at room temperature. Plant materials were reduced to powder that was used for extraction. The extraction was performed by percolation of $100\,\mathrm{g}$ powder with $500\,\mathrm{mL}$ ethanol-water (70: 30) for $48\,\mathrm{h}$. The extract was then filtered with Whatman paper and the filtrate was evaporated until dry using a Rotary evaporator at $50\,\mathrm{^{\circ}C}$ under reduced pressure.

To perform cytotoxicity and anthelmintic tests, 100 mg/mL extracts were prepared by dissolving 1 g of dried extract in 10 mL distilled water. From this, serial dilutions were made and filtered using 0.45 μ m millipore adapted to a syringe.

Purification of peripheral blood microfilariae (mf)

Frozen Mf were obtained from the Institute for Medical Microbiology, Immunology and Parasitology, University Hospital Bonn (UKB), Bonn, Germany. The Mf for in vitro anti-microfilarial test were isolated from the peripheral blood of *L. sigmodontis*-infected cotton rats [11]. Blood was diluted with PBS in the ratio 1:2 and carefully loaded onto a 30-25% Percoll gradient (Sigma-Aldrich GmbH, Munich, Germany). After 30 min centrifugation (300 g) at room temperature without break, the recovered Mf were washed two times with non-supplemented RPMI-1640 medium (PAA, Linz, Austria), counted and frozen at -80 °C in freezing medium containing 6% DMSO and 15% fetal calf serum (FCS) (PAA). For this experiment, fresh aliquots of frozen Mf were thawed and controlled for Mf viability microscopically after 2h of pre-incubation at 37 °C in RPMI-1640 medium containing 10% FCS in order to revitalize the microfilariae. Only aliquots with more than 95% Mf viability were used.

Anthelmintic assay

For the assay, $100\,\mu\text{L/well}$ of suspension containing 75 microfilariae were grown in RPMI 1640 in a 96-well plate with plant extracts 200, 500 and $1000\,\mu\text{g/mL}$. The control was a subculture without plant extract. Albendazole (5 mg/mL) was used as reference drug as previously described [2]. After 7 days incubation at 37 °C in a humid atmosphere, the viability of the microfilariae was evaluated microscopically using trypan blue exclusion method. Living and dead Mf were counted for each concentration of drug. The concentration that induced 50% Mf death was considered as LD₅₀.

Cytotoxicity assay

The cytotoxicity assays was conducted according to the recommendation of NF EN ISO 10993-5 standard. Human peripheral blood mononuclear cells from healthy volunteers (n=13) were isolated using ficoll density gradient centrifugation method and treated with 200 µg/mL plant extracts for 24 h at 37 °C under 5% CO₂. Afterwards, cell pellets were harvested and stained with propidium iodide (PI) dye prior to cell acquisition using Cytoflex flow cytometer (Beckman Coulter, Brea, USA). From the lymphocytes gate, the percentage of cells expressing propidium iodide (PI) was assessed. Data were analysed using CytExpert 2.1 sofware (Beckman Coulter, Brea, USA).

Data analysis

A Microsoft Excel spreadsheet 2013 was used to perform simple calculations and determine plant frequencies. The relative importance of species was evaluated by the frequency in which it was mentioned by the THs and a "Used value" (UV) was calculated as follows: $UV = \Sigma U/n$ (ΣU is the total number of citations per species and n is the number of interviewed THs. The UV is useful in determining which plants have the best use and are most often indicated in the treatment of a disease [12]. The agreement (Informant consensus factor (ICF)) of the THs regarding the uses of medicinal plants to treat helminthiasis was calculated by the following formula: ICF=Nuc -Nt/(Nuc -1) where Nuc is the number of citations for the treatment of a given disease and Nt is the number of species used in the treatment of a given disease [13].

Statistical analyses were performed using Graph Pad PRISM 5.02 software (GraphPad Software, La Jolla, USA). The χ^2 test was used for the comparison between groups and the difference was considered significant with a p-value< 0.05.

Results

Ethnobotanical study

Characteristics of traditional healers

A total of 197 THs were interviewed, among them 168 were men and 29 were women. 76.14% of the cohort were older than 40 years and 77.66% practiced traditional medicine for more than 10 years (Table 1). The majority of THs inherit the knowledge from their family according to the percentage in that category (77.16%). According to the ethnical affiliation, the interviewed THs belonged to *Tem* (82.65%), *Mina* (6.64%), *Moba* (8.67%) and *Gourma* (2.04%) tribes.

Main Helminthiasis treated by THs

THs claimed to treat onchocerciasis, lymphatic filariasis and schistosomiasis based on observed clinical symptoms and all interviewed THs were familiar with these three helminth infections. Figure 2a shows the frequencies in which THs had cases and treated the associated pathologies. 43.65% of THs had treated schistosomiasis, 28.93% had treated lymphatic filariasis and 17.26% had treated onchocerciasis.

The THs recognized the disease by the main characteristics of symptoms. Figure 2 shows the list and frequencies of symptoms used by the THs to diagnose onchocerciasis, lymphatic filariasis and schistosomiasis. Itching and eye disorders were the most common symptoms associated with *O. volvulus* infections (Fig. 2b). Hematuria was the most obvious symptom for *S. haematobium* infections with nearly 70% of THs associating this symptom with the infection (Fig. 2c). Lymphatic filariasis (LF) caused by *W. bancrofti* was diagnosed by "big foot" (49%) and "swelling of the foot" (36.84%).

Plants used for the treatment of Helminthiasis

A total of 41 plant species from various families were prominently mentioned by THs during the interviews. Most belonged to Leguminosae (14.6%) and Annonaceae (9.7%) families and are listed in Table 2.

According to the UV score, A. melegueta, X. aethiopica and K. senegalensis were the most frequently used for the treatment of onchocerciasis, schistosomiasis and lymphatic filariasis. A. melegueta was the most cited by the THs for the treatment of onchocerciasis (UV = 0.036) while X. aethiopica was associated with the treatment of schistosomiasis (UV = 0.061) and lymphatic filariasis (UV = 0.061). Moreover, A. melegueta, X. aethiopica and K. senegalensis were also more used for the treatment of inflammation (UV = 0.066, UV = 0.056and UV = 0.025 respectively, data not shown). The agreement among the THs for treating a given disease with plant materials was high with 0.57 for onchocerciasis, 0.66 for lymphatic filariasis and 0.61 schistosomiasis.

Roots, seeds, leaves and barks were the main plant's organs used by THs to prepare their medicinal recipes respectively, 35, 24, 21 and 14% (Fig. 3a). All the interviewed THs claimed to harvest plant materials during any season and at any time of the day.

For the formulation of medicinal recipes, powder (36.17%) and decoction (34.04%) were most cited (Fig. 3b). In addition, it was observed that most of the THs administrated the preparation by oral (51.06%) or topical (63.83%) routes.

Anthelmintic effects

In the anthelmintic assay, the microfilariae from the control subculture, without drug were not stained, while

Table 1 Characteristics of THs treating Helminthiasis

	Gender	Age N (%)		Experience I	N (%)		Origin of the	knowledge N	(%)
	N (%)	< 40 years	≥ 40 years	0–4 years	5–9 years	≥ 10 years	Heritage	Training	Calling
Men	168(85.28)	38(80.85)	130(8.67)	10(55.56)	22(84.62)	136(88.89)	133(67.51)	21(10.66)	14(07.10)
Women	29(14.72)	9(19.15)	20(13.33)	8(44.44)	4(15.38)	17(11.11)	19(9.64)	9(04.57)	1(0.51)
Total	197 (100)	47(23.86)	150(76.14)	18(9.14)	26(13.20)	153(77.66)	152(77.16)	30(15.22)	15(7.61)

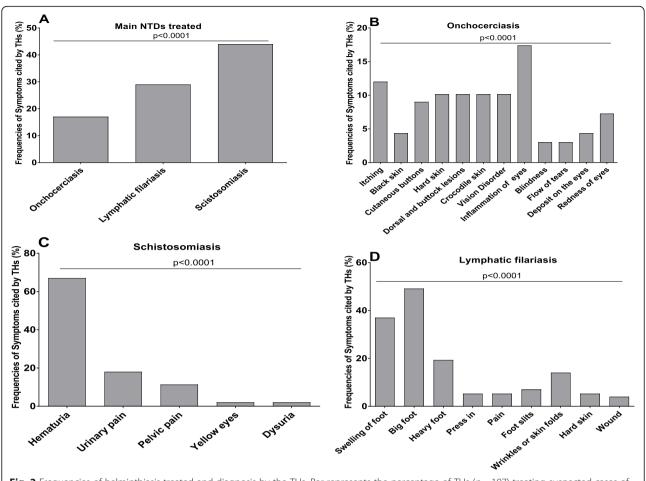


Fig. 2 Frequencies of helminthiasis treated and diagnosis by the THs. Bar represents the percentage of THs (*n* = 197) treating suspected cases of onchocerciasis, lymphatic filariasis, schistosomiasis, dermatitis and inflammation **a** and symptoms used by the THs (=197) for the diagnosis of the diseases: onchocerciasis **b**, schistosomiasis **c** and lymphatic filariasis **d**. Chi-square was used to compare differences between groups and *p* value was less than 0.0001

microfilariae from Albendazole, *A. melegueta*, *K. senegalensis* and *X. aethiopica*, subculture were stained blue in the presence of trypan blue (Fig. 4). This indicated that *A. melegueta*, *K. senegalensis* and *X. aethiopica* had induced microfilariae death. The concentration that induced the death of 50% of microfilariae was 233 μ g/mL for *X. aethiopica*, 265 μ g/mL for *A. melegueta* and 550 μ g/mL for *K. senegalensis* (Fig. 4).

Cytotoxicity

The cytotoxicity of the plants was evaluated by flow cytometry using PI staining after PBMCs were cultured with the plant extracts. According to the NF EN ISO 10993-5 standards classification, *A. melegueta* and *X. aethiopica* plant extracts were not cytotoxic. This was indicated by less than 20% of CD4 $^+$ PI $^+$ cells. It was however observed that *K. senegalensis* induced moderate cell toxicity (24 \pm 8% of CD4 $^+$ PI $^+$ cells) (Fig. 5).

Discussion

This study aimed to document the use of plants in the treatment of helminthiasis, the survey revealed that male THs were more represented contrary to the reports of Gale et al. who found that women in Togo were more represented in complementary and alternative medicine [14]. The reason could be linked to cultural issues in the study region, since women retain a more traditional role as homeworkers. The findings however were in accordance with other study that showed the predominance of men among TH in Tem tribe [10]. The majority of THs were over 40 years old and had more than 10 years professional experience in traditional medicine. Most of them inherited their knowledge from their family confirming previous findings in this region [15]. To diagnose the disease, the THs rely on symptoms. Characteristic symptoms of lymphatic filariasis are hydrocele and lymphedema [16]. The diagnosis of urinary schistosomiasis should be especially suspected in cases of terminal hematuria and eosinophilia. It can be

Table 2 Diversity of plants used for the treatment of helminthiasis: administration route, parts and the mode of preparation

Species	Family	Local name	Voucher	Used parts	Formulation	Route	Use value		
			number				Onchocerciasis	Lymphatic filariasis	Schistosomiasis
Acacia sieberiana Tausch.	Leguminosae	Bovom	TOGO15381	Root	Powder	Oral		0.005	
Aframomum melegueta K.Schum.*	Zingiberaceae	Abaltchangai/ Kalmboa	TOGO15382	Seed	Decoction, Dough	Oral, Topical	0.030	0.036	0.025
Anacardium occidentale L.	Anacardiaceae	Atcha	TOGO15383	Bark	Decoction	Oral			0.005
Annona senegalensis Pers.	Annonaceae	Tchoyhodè/ Tchutchudè	TOGO15384	Root, Leaves, Bark	Decoction	Topical		0.015	0.005
Biophytum petersianum Klotzsch	Oxalidaceae	Kpirikpozo	TOGO15385	Whole plant	Decoction	Oral			0.005
Blighia sapida K.D.Koenig	Sapindaceae	Kpézou/ Kpèzéou	TOGO15386	Bark, Fruit, Seed, Leaves	Dough, Decoction	Oral, Topical	0.005		0.010
Bombax costatum Pellegr. & Vuillet	Bombacaceae	Folo	TOGO15387	Bark, Leaves	Decoction	Topical		0.005	0.005
Calotropis procera (Aiton) W.T.Aiton	Asclepiadaceae	Tchovow	TOGO15388	Leaves, Bark, Root, Sap	Powder	Topical		0.005	
Cyathula prostrata Blume	Amaranthaceae	Amatamata	TOGO15392	Leaves	Powder, Decoction	Topical			
Dychrostachys cinerea R.Vig.	Leguminosae	Sozozi	TOGO15393	Leaves	Dough, Decoction	Topical			
Erythrina senegalensis A.Rich.	Leguminosae	Gbingbintoukoloko/ Kpodjkpalo	TOGO15394	Root	Decoction	Oral	0.005		0.010
Flueggea virosa Baill.	Euphorbiaceae	Tchakatchaka	TOGO15397	Root, Leaves	Infusion, maceration, Decoction, Powder	Oral, Topical		0.005	0.005
Hannoa undulata Planch.	Simaroubaceae	Dgbéré	TOGO15398	Root, Bark	Infusion, Decoction	Oral, Topical		0.005	
Héliotrope indicum L.	Boraginaceae	Soudjondjon, Soukoudjo	TOGO15399	Root	Powder	Oral, Topical			0.010
Hexalobus monopetalus Engl. & Diels	Annonaceae	Barakoundou	TOGO15400	Root	Infusion, Decoction Powder	Oral, Topical			
Jatropha curcas L.	Euphorbiaceae	Sawkofolmo	TOGO15401	Leaves, Sap	Decoction	Oral	0.005		
Khaya senegalensis A.Juss.*	Meliaceae	Frémou/ Hèmo	TOGO15402	Bark, Root, Leaves	Dough	Topical, Oral	0.020	0.015	0.036
Kigelia africana (Lam.) Benth.	Bignoniaceae	Abiliou/ Limié	TOGO15403	Bark, Leaves, Root, Seed	Mashing	Oral, Topical	0.005	0.020	
<i>Landolphia hirsuta</i> (Hua) Pichon	Apocynaceae	Low	TOGO15404	Root	Powder	Oral			0.005
Lannea barteri Engl.	Anacardiaceae	Kpatandew	TOGO15405	Bark	Decoction, Infusion	Oral			0.015
Lawsonia inermis L.	Lythraceae	Lali	TOGO15406	Leaves	Powder	Oral			
<i>Lophira lanceolata</i> Tiegh. ex Keay	Ochnaceae	Kparakpara	TOGO15407	Leaves, Root	Dough	Topical	0.005		
Ocimum canum Sims en u	Lamiaceae	Kozossognna/ Hagzao, Kosonsong	TOGO15409	TOGO15409 Leaves, Root, Whole plant	Mashing	Ocular, Oral	0.010	0.005	0.005

Table 2 Diversity of plants used for the treatment of helminthiasis; administration route, parts and the mode of preparation (Continued)

TOGO15410 Bark, Leaves, Root, Powder Oral, Topical Seed	Species	Family	Local name		Used parts	Formulation	Route	Use value		
ripolatosa (lacq.) RBit. Leguminosae Solo TOCO15411 Bark, Leaves, Rood. Powder Onal, Topical ris buildora (Benth, ex. in summinosae Leguminosae Limbré Limbré TOCO15411 Root, Leaves, Seed Powder Topical, Onal ris smuleirianus Luphorbiaceae Limbré Limbré TOCO15412 Root, Leaves, Seed Powder Oral FEAI rivennez Thorn. Prediction Upperaceae Djéyawa TOCO15413 Seed Powder Oral, Topical rivennez Thorn. Prediction Répédoute/ TOCO15413 Seed Powder Oral, Topical ris subressa Production Combretaceae Répédoute/ TOCO15413 Reoccition, Decoction Oral, Topical pub serinaceae Prod. Combretaceae Tomminosae ToCO15418 Root, Bark, Leaves, Bark Maceration, Decoction Oral, Topical pub serinaceae Poil. Faboceae ToTAT Bark TOCO15418 Root, Bark, Leaves, Bark Maceration, Infusion Oral, Topical pub serinaceae Poil. Faboceae Kichintrihin TOCO1542 Reaves				number				Onchocerciasis	Lymphatic filariasis	Schistosomiasis
specificate (Benth, as, Leguminosae) Kodoléyae TOGOIS411 Rou, Le, Ba, Fr Powder Topical, Oral Le, Ba, Fr betwoen Limbré Limbré Imbré Limbré TOGOIS413 Seed Powder, Decoction Topical briteanse Fhonn Piperaceae Déyawa TOGOIS413 Seed Powder, Decoction Topical dricara Taub. Leguminosae Rétécourée TOGOIS413 Seed Powder, Decoction Topical dricara Taub. Leguminosae Rétécourée TOGOIS415 Root, Bark, Leaves Powder Dough, Decoction Oral, Topical dricara Jaub. Leguminosae Sissnow/ Kizinja TOGOIS417 Raves, Root Decoction Oral, Topical dricaraceas Poir. Fabaceae Tern/ Tem TOGOIS419 Root, Leaves, Root Maceration, Infusion Oral, Topical dricaracears Poir. Rabitileo/ Kakayo TOGOIS421 Leaves, Mode plant Powder Oral, Topical dricaracears Poir. Alaberdesee Kafirhileo/ Kakayo TOGOIS422 Leaves, Whole plant Powder Oral, Topical	Parkia biglobosa (Jacq.) R.Br. ex G.Don	Leguminosae	Solo		Leaves, Root,	Powder	Oral, Topical		0:002	0.025
Hus muellerianus Euphorbiaceae Limbré Limbré Limbré TOGO15412 Root, Leaves, Seed Powder, Decoction Topical Fibell Timbré Limbré Limbré TOGO15413 Seed Powder, Decoction Topical adricand Taubn Leguminosae Kpalo TOGO15415 Root, Bark, Leaves Powder Oral Topical sistiede kostrofyl Harms Meliaceae Bisélecuée TOGO15415 Root Bowder Oral Topical pus erinaceus Poir Fabaceae Sissinow/ Kizzina TOGO15417 Leaves, Root Macration, Dough Oral, Topical pus erinaceus Poir Fabaceae Krightilor Kizzina TOGO15417 Root, Leaves, Bark Macration, Dough Oral, Topical pus erinaceus Poir Fabaceae Krightilor Kizzina TOGO15419 Root, Leaves, Bark Macration, Dough Oral, Topical pundus tarribdus (SM) Rubiaceae Krightilor Kizzina TOGO15421 Leaves, Bark Macration Oral, Topical pundus tarribdus (SM) Myriaceae Krightilor Kizzina TOGO15422 Leaves, Bark Macration <th< td=""><td><i>Pericopsis laxiflora</i> (Benth. ex Baker)Meeuwen</td><td>Leguminosae</td><td>Kodoléya</td><td></td><td>e, Ba, Fr</td><td>Powder</td><td>Topical, Oral</td><td></td><td></td><td>0.015</td></th<>	<i>Pericopsis laxiflora</i> (Benth. ex Baker)Meeuwen	Leguminosae	Kodoléya		e, Ba, Fr	Powder	Topical, Oral			0.015
incensor Houn. Piperaceae Djeyawa TOGO15413 Seed Powder, Decoction Topical adricona Taub. Leguminosae Kpalo TOGO15416 Root, Bark, Leaves Mashing Oral, Topical is suberosa Engl. & Combreaceae Sissinow/ Kizzina TOGO15416 Root Bark, Leaves Powder Oral, Topical pvs erinaceus Poir. Fabaceae Tem/ Tem TOGO15418 Root Maceration, Decoction, Oral Oral, Topical n.) Hook! Tomaraceae Tem/ Tem TOGO15418 Root Maceration, Infusion, Dough Oral, Topical n.) Hook! Rubiaceae Kidjithilo/ Kakayo TOGO15419 Root Maceration, Infusion Oral, Topical phola latifolius (SM) Rubiaceae Krdintchin TOGO1542 Leaves, Root Powder Oral, Topical na latifolius (SM) Mulaceae Nobacoudou/ TOGO1542 Leaves, Mole plant Powder Topical na accidentalis (L) Lien Mylaceae Nobacoudou/ TOGO1542 Leaves, Mhole plant Powder Topical <td>Phyllanthus muellerianus (Kuntze) Exell</td> <td>Euphorbiaceae</td> <td>Limbré Limbré</td> <td></td> <td>, Leaves, Seed</td> <td>Powder</td> <td>Oral</td> <td></td> <td>0.005</td> <td>0.010</td>	Phyllanthus muellerianus (Kuntze) Exell	Euphorbiaceae	Limbré Limbré		, Leaves, Seed	Powder	Oral		0.005	0.010
officeno Taub. Leguminosae Kpalo TOGO15416 Root, Bark, Leaves Mashing Oral, Topical edreto koschyi Harms Melaceee Brétécouré/ Hérétécoude TOGO15416 Root, Bark, Leaves Powder Oral, Topical pus erinaceus Poil. Fabaceae Termanido TOGO15416 Root Maceration, Decoction, Powder Oral, Topical coccinea (Schumach.) Connaraceae Termanido TOGO15419 Root, Bark, Leaves, Bark Maceration, Infusion, Dough Oral, Topical hyl Hook f. Rubiaceae Kidjithilof Kakayo TOGO15419 Root, Bark, Leaves, Bark Maceration, Infusion, Dough Oral, Topical rica longepedunculara Polygalaceae Forthintchin TOGO15421 Root, Bark, Leaves, Bark Movder Oral, Topical rica Burm f. Malvaceae Kichintchin TOGO15421 Leaves, Mole plant Powder Topical rica Burm f. Myrtaceae Kondouvu TOGO15422 Leaves, Whole plant Powder Topical I Melaceae Adjendjakbézou TOGO15428 Root, Leaves, Bark Maceration,	Piper guineense Thonn.	Piperaceae	Djéyawa			Powder, Decoction	Topical	0.010		0.015
cocle fol Roschyi Hamms Meliaceae Bistiebouréh TOGO15415 Root, Bark, Leaves Powder Oral, Topical pus emaceus Poir. Fabaceae Tem/ Tem TOGO15416 Root, Leaves, Root Maceration, Decoction, Oral, Topical Oral, Topical pus emaceus Poir. Fabaceae Tem/ Tem TOGO15418 Root, Leaves, Root Infusion, Dough Oral, Topical n) Hook f. Antole Park Robin plant, Fruit Maceration, Infusion, Decoction, Powder Oral, Topical coclinear (Schumach, StM.) Rubiaceae Fozi/ Bnbna TOGO15418 Root, Leaves, Bark Maceration, Infusion, Decoction, Powder Oral, Topical coal Jongepeduncular Polygalaceae Fozi/ Bnbna TOGO15421 Leaves, Root Powder Oral, Topical na Burm.f. Malvaceae Richintchin TOGO15421 Leaves, Root Powder Oral, Topical na conditional (L.) Lien Asealpiniaceae Krinintchin TOGO15421 Leaves, Root Decoction, Infusion Oral, Topical na connetice vall Marciaceae Anonaceae Anionaceae Adjendjakpézou	Prosopis africana Taub.	Leguminosae	Kpalo		. Bark, Leaves	Mashing	Oral		0.005	0.005
pus evinaceus Polit. Combretaceae Sissinow/ Kizizina TOGO15412 Roat Peccotion Decoction. Oral, Topical pus evinaceus Polit. Fabaceae Tem/ Tem TOGO15413 Roat Maceration, Decoction, Oral Oral, Topical pub serinaceus Polit. Caccinea (Schumach, Caccineae) Connaraceae Tohamalido TOGO15418 Roat Maceration, Decoction, Powder Oral, Topical phalus latifolius (SMI) Rubiaceae Kidjithilo/ Kakayo TOGO15419 Roat, Leaves, Bark, Leaves, Bark, Leaves, Bark, Leaves, Roat Maceration, Infusion, Oral, Topical Oral, Topical kcalennalis (L) Lien Caesalpiniaceae Ktchintchin TOGO15421 Leaves, Mole plants Powder Topical na anamaticum (L) Like Malvaceae Kanafourou TOGO15422 Leaves, Whole plants Powder Oral, Topical LMPerry Combretaceae Souwo/ Sintéou TOGO15422 Reaves, Bark Maceration Topical centrico de Benth. Meliaceae Adjendjakpézou TOGO15428 Roat, Leaves, Bark Infusion, Dough Topical chanaceae <td>Pseudocedrela kotschyi Harms</td> <td>Meliaceae</td> <td>Btétéouré/ Hététéoudè</td> <td></td> <td>, Bark, Leaves</td> <td>Powder</td> <td>Oral, Topical</td> <td></td> <td>0.015</td> <td>0.005</td>	Pseudocedrela kotschyi Harms	Meliaceae	Btétéouré/ Hététéoudè		, Bark, Leaves	Powder	Oral, Topical		0.015	0.005
pus eninaceus Poir. Fabaceae Tem/ Tem TOGO15418 Root. Maceration, Decoction, Infusion, Dowgher Oral Topical Infusion	Pteleopsis suberosa Engl. & Diels	Combretaceae	Sissinow/ Kizizina	TOGO15416 Root		Decoction	Oral, Topical	0.010		0.005
coccined (Schumach.) Hook.f. Connaraceae Trhamalido TOGO15419 Touch (Lavaes, Bark, and Infusion) Infusion, Doughh (Asia) Oral, Topical (Asia) TOGO15419 Touch (Lavaes, Bark, and Infusion) Maceration, Infusion Oral, Topical (Asia) Oral, T	Pterocarpus erinaceus Poir.	Fabaceae	Tem/ Tem	TOGO15417 Leav	es, Root	Maceration, Decoction, Infusion, powder	Oral		0.010	
phalus latifolius (SM)RubiaceaeKidjithilo/ KakayoTOGO15419Root, Leaves, Bark, PruitMaceration, Infusion, Oral, Topical Decoction, PowderOral, Topical Oral, Topical Oral, Topical Oral, Topical Oral, Topical Oral, Topical Oral, Decoction, PowderOral, Topical Oral, Topical Oral, Topical Oral, Topical Oral, Topical Oral, Decoction, InfusionOral, Topical Oral, Topical Oral, Topical Oral, Topical Oral, Topical Oral, Decoction, InfusionAccidentalis (L) LienMalvaceaeKrhintchinTOGO15422Leaves, RootPowderTopical Oral, Topical Oral, Topical Oral, Decoction, InfusionOral, Topical Oral, Decoction, InfusionL.M.Perry Ilia glaucescensCombretaceaeSouwo/ SimtéouTOGO15424Root, Leaves, BarkMacerationTopical Oral,	Rourea coccinea (Schumach. & Thonn,) Hook.f.	Connaraceae	Tchamalido			Infusion, Dough	Oral, Topical			0.015
tca longepedunculataPolygalaceaeFozi/ BnbnaTOGO15420Roott, Bark, Leaves, Moole plantsDough, Decoction, InfusionOral, Topicalvcidentalis (L) LienCaesalpiniaceaeKtchintchinTOGO15421Leaves, Moole plantPowderTopicalnaromaticum (L)MyrtaceaeKanafourouTOGO15422Leaves, Whole plantPowderOral, TopicalL.M.PerryMyrtaceaeKanafourouTOGO15424Root, Bark, Seed, BarkDecoction, InfusionOral, TopicalL.M.PerryCombretaceaeSouwo/ SimtéouTOGO15424Root, Leaves, BarkMacerationTopicalewetica VahlMeliaceaeAdjendjakpézouTOGO15425Root, Leaves, BarkInfusion, DoughOcular, Oral, Topicalchamae P.Beauv.AnnonaceaeSoozi/ Koékrabi, Togo15427FruitPoccoction, Infusion, Mutsion, PowderOral, Topical	Sarcocephalus latifolius (SM.) E.A.Bruce	Rubiaceae	Kidjithilo/ Kakayo		, Leaves, Bark, le plant, Fruit	Maceration, Infusion, Decoction, Powder	Oral, Topical	0.010	0.015	0.015
CaesalpiniaceaeKtchintchinTOGO15421Leaves, Mhole plantPowderTopicalMalvaceaeNbazoudou/ KpenzaloTOGO15422Leaves, Whole plantPowderOral, TopicalMyrtaceaeKanafourouTOGO15423Root, Bark, Seed, LeavesDecoction, InfusionOralCombretaceaeSouwo/ SimtéouTOGO15424Root, Leaves, BarkMacerationTopicalMeliaceaeAdjendjakpézouTOGO15425Root, Leaves, BarkInfusion, DoughTopicalAnnonaceaeSoozi/ Koékrabi, SooseTOGO15427FruitDecoction, Infusion, PowderOral, Topical	Securidaca longepedunculata Fresen.	Polygalaceae	Fozi/ Bnbna		, Bark, Leaves, le plants	Dough, Decoction, Infusion	Oral, Topical	0.015	0.036	0.005
Malvaceae Nbazoudou/ Kpenzalo TOGO15422 Leaves, Whole plant Powder Oral, Topical Myrtaceae Kanafourou TOGO15423 Root, Bark, Seed, Leaves Decoction, Infusion Oral Combretaceae Souwo/ Simtéou TOGO15424 Root, Leaves, Bark Infusion, Dough Topical Annonaceae Doumfodou TOGO15426 Root, Leaves, Bark Infusion, Dough Topical Annonaceae Doumfodou TOGO15426 Root Fruit Decoction, Infusion, Powder Oral, Topical	Senna occidentalis (L.) Lien	Caesalpiniaceae			es, Root	Powder	Topical			0.005
Myrtaceae Kanafourou TOGO15423 Root, Bark, Seed, Leaves Decoction, Infusion Oral Leaves Combretaceae Souwo/ Simtéou TOGO15424 Root, Leaves, Bark Maceration Topical Meliaceae Adjendjakpézou TOGO15425 Root, Leaves, Bark Infusion, Dough Topical Annonaceae Doumfodou TOGO15426 Root Root Maceration Topical Annonaceae Soozi/ Koékrabi, Soosie TOGO15427 Fruit Powder Oral, Topical	Sida acuta Burm.f.	Malvaceae	Nbazoudou/ Kpenzalo		es, Whole plant	Powder	Oral, Topical			0.010
Combretaceae Souwo/ Simtéou TOGO15424 Root, Leaves, Bark Maceration Topical Meliaceae Adjendjakpézou TOGO15425 Root, Leaves, Bark Infusion, Dough Topical Annonaceae Doumfodou TOGO15426 Root Maceration Topical Annonaceae Soozi/ Koékrabi, TOGO15427 Fruit Decoction, Infusion, Oral, Topical	Syzygium aromaticum (L.) Merr. & L.M.Perry	Myrtaceae	Kanafourou		, Bark, Seed, es	Decoction, Infusion	Oral	0.005		0.025
Meliaceae Adjendjakpézou TOGO15425 Root, Leaves, Bark Infusion, Dough Ocular, Oral, Topical Annonaceae Dough TOGO15426 Root Maceration Topical Annonaceae Soozi/ Koékrabi, Soossé TOGO15427 Fruit Decoction, Infusion, Oral, Topical	<i>Terminalia glaucescens</i> Planch. ex Benth.	Combretaceae	Souwo/ Simtéou		, Leaves, Bark	Maceration	Topical			
Annonaceae Doumfodou TOGO15426 Root Maceration Topical Annonaceae Soozi/ Koékrabi, TOGO15427 Fruit Decoction, Infusion, Oral, Topical Powder	Trichilia emetica Vahl	Meliaceae	Adjendjakpézou		, Leaves, Bark	Infusion, Dough	Ocular, Oral, Topical		0.015	0.005
Annonaceae Soozi/ Koékrabi, TOGO15427 Fruit Decoction, Infusion, Oral, Topical Powder	Uvaria chamae P.Beauv.	Annonaceae	Doumfodou			Maceration	Topical		0.005	0.005
	Xylopia aethiopica A.Rich.*	Annonaceae	Soozi/ Koékrabi, Soossé			Decoction, Infusion, Powder	Oral, Topical	0.020	0.061	0.061

This table presents the used values (UV), administration route, formulation of drugs from each species of plants and the scientific, local name of each plant for the treatment of onchocerciasis, lymphatic filariasis, schistosomiasis, dermatitis and inflammation. * show the most cited plants by the THs for the treatment of helminthiasis and selected for in vitro screening

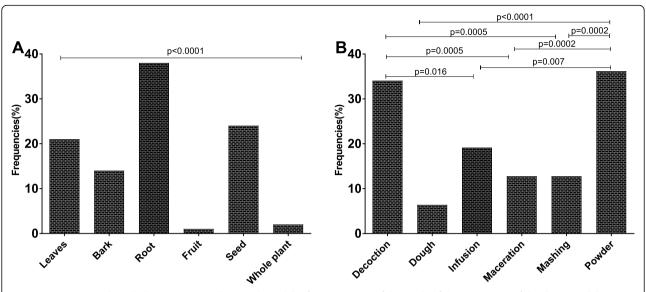


Fig. 3 Preparation mode and plant's organs used. Bars indicated the frequencies (%) of the mode of the preparation of the drug **a** and the parts of the plants used **b**. Chi-square was used to compare differences between groups and p value was less than 0.0001

considered that the interviewed THs have a basic know-ledge of the target diseases because they can recognize most of the specific symptoms [17].

Some plant families seem to stand out in any pharmacopoeia. A study on antimalarial plants in the Maritimes region of the same country revealed that out of 52 antimalarial plants species, Rubiaceae and Rutaceae were the most used to combat malaria [18]. In a study conducted in the central plate of Burkina Faso, the following families Caesalpiniaceae, Poaceae, Mimosaceae and Fabaceae have been ranked amongst the richest in species citations [19]. These medicinal plants were distributed among 28 families, the largest proportion belonging to the families Fabaceae and Anacardiaceae. Telefo et al. also identified 46 plant species belonging to 26 families, the largest number of species recorded in Asteraceae and Acanthaceae [20]. In this study, 41 species were recorded and the largest number of species belonged to Leguminosae and Annonacea. The preference for their use may be related to their accessibility, as they are common and grow more in this area. According to Heinrich et al., when the consensus factor of informants is high, it reflects a good knowledge of medicinal plants, a collective knowledge of their uses, but also an exchange of information between THs [21]. In the study the ICF was high meaning that there was a great agreement among the THs regarding the use of these plants for the treatment of helminthiasis. Many authors studied the anthelmintic properties of plants in Africa [22]. Several plants used in the treatment of helminthiasis were found active in in vitro screenings. For examples Ceratonia siliqua extract was shown to ameliorate Schistosoma *mansoni*-induced liver fibrosis, while Verbascum sinaiticum and Commiphora swynnertonii exerted trypanocidal activity [23, 24]. Anthelmintic effects of A. meleon *helminth parasites*, were observed by Akinsanya et al. [25]. The ethanolic extract of A. melegueta also known as "grain of paradise" has antiinflammatory properties by inhibiting the activity of cyclooxygenase-2 (COX-2) enzyme [26]. A. melegueta seeds are also used in Africa to treat diarrhoea and gastroenteritis [27]. In southern Nigeria, Benin and Togo, it is employed in divine practices [28]. Xylopia aethiopica was mostly used for the treatment of filariasis and schistosomiasis [29]. The plant is also used commonly in Nigeria by traditional herbalists to treat gastrointestinal helminth parasites [30]. Ademola et al. suggested the use of K. senegalensis extract in anthelmintic therapy in veterinary practice [31]. Antitrypanosomal activity of K. senegalensis was also investigated by some authors [32]. In the present study, in vitro anthelmintic activities of the three plants was investigated using microfilariae of Litomosoides sigmodontis at different concentrations of the plant extracts. The mortality of Mf was dose-dependent and X. aethiopica had more effect on the microfilariae indicated by the lowest LD₅₀. These data justify the use of the cited plants by the THs for the treatment of helminthiasis. Further studies on the effect of these plants on parasite paralysis and mobility should be carried on.

Fruit extracts are used to treat coughs, bronchitis, dysentery rheumatism and malaria [33–35]. The study showed that root, leaves and seed were the parts of the plants most cited by the THs for the treatment of helminthiasis. Previous studies have shown that leaves and roots were mostly used for the treatment of asthma [36].

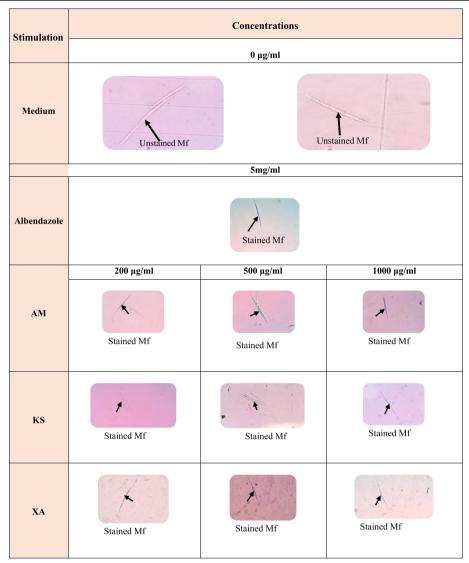


Fig. 4 Anthelmintic effect of *Xylopia aetiopica* (XA), *Aframomum melegueta* (AM) and *Khaya senegalensis* (KS) on microfilariae (Mf) of *Litomosoides sigmodontis*. Mf (n = 75/well) of *Litomosoides sigmodontis* were cultured in absence of plant extracts (0 μg/ml, medium) or in presence of AM (green line), KS (blue line) and XA (red line) at different concentrations (200 μg/ml, 500 μg/ml and 1000 μg/ml). After 7 days culture, the viability of microfilariae were evaluated using typan blue. Graph shows the percentage of dead Mf for each concentration. The concentration that induced 50% of death was designed lethal dose 50 (LD50)

A similar study showed that to treat liver disease, the most used parts of this plant were the leaves and roots, thus extracts of this plant can aid in a broad spectrum of symptoms [37].

The cytotoxicity of the three studied plants by the THs to treat helminthiasis was evaluated on human peripheral mononuclear blood cells (PBMCs) by flow cytometry. This method has the advantage to directly show the toxicity of the plant for human cells but an in vivo evaluation could indicate exactly which vital organs are damaged. *Afromomum melegueta* and *X. aethiopica* were not cytotoxic at 200 µg/mL but *K. senegalensis* revealed moderate toxicity with cell

mortality above 20% at the same concentration. Many studies were performed on the cytotoxicity of the three extracts. Sahar et al, showed that K. senegalensis would be toxic to human liver, breast and colon cancer cells with IC₅₀ of 61.1, 79.7, and 61 μ g/mL respectively and sesquiterpens occurring in the plant would be responsible for its toxicity [38]. On the other hand, Idoh et al. revealed that A. melegueta has a hepatoprotective effect on rats [29]. Apart from its hepatoprotective property, A. melegueta would also have anti-apoptotic properties [39]. Similarly, volatile oil from X. aethiopica was found to be non-toxic to human epidermal cells line [40].

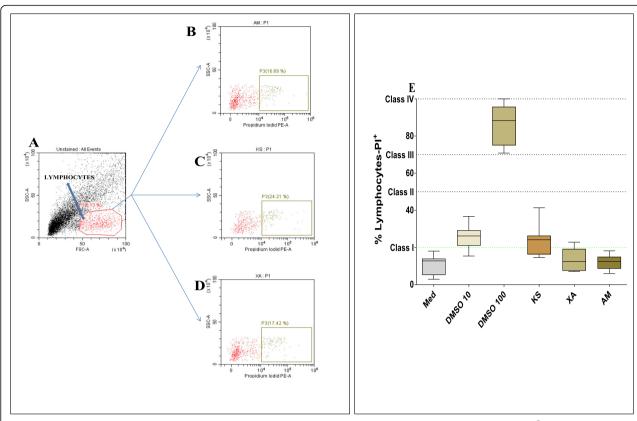


Fig. 5 Cytotoxicity of the most frequently used plants by the THs for the treatment of helminthiasis: Human PBMCs (2×10^5 cells/well) were left alone (Med) or stimulated with 200 μg/ml of AM, KS, XA, DMSO 100 and 10% for 24 h. Cells were stained with propidium iodide dye (Pl) and acquired by flow cytometry. **a:** lymphocytes gate, **b, c,** and **d** are Pl⁺ cells in presence of AM, KS and XA respectively. (E) Box whiskers (tukey) with outliers show the percentage of lymphocytes expressing Pl (n = 13). P values were determined by Mann-Whitney U-test. The NF EN ISO 10993-5 standards classifications were indicated by class I (not cytotoxic), class II (moderate cytotoxicity), class III (Benign cytotoxicity) and class IV (severe cytotoxicity)

Many studies have demonstrated the anthelmintic effects of total phenolic and flavonoids compounds [41–44]. However, it would be difficult to say with accuracy, that these chemical groups are responsible for the activity observed in the present study. Thus, further bioguided fractionation of each plant should be undertaken to identify the active principles.

Conclusion

This study demonstrated that THs had knowledge about the treatment of helminthiasis based on plants materials and highlighted the main plants used in the Central and Kara region of Togo. The anthelmintic and cytotoxicity effects of these commonly used plants were delineated. The benefits of listening to THs in such endemic areas is paramount to unveil potential new sources for fighting helminthiasis in general and moreover, contribute to the identification of new molecules for the treatment of symptoms and conditions arising from chronic helminth infections.

Abbreviations

AM: Aframomum melegueta; CD4: Cluster of Differenciation Antigen 4; CERMETRA: Centre d'Etude et de Recherche en Médecine Traditionnelle Appliquée du Togo; CO₂: Carbon Dioxide; COX-2: Cyclooxygenase-2; DALYs: Disability-Adjusted Life Years; DMSO: Dimethyl Sulfoxide; EN: European Norm; FCS: Fetal Calf Serum; IC50: The half maximal inhibitory concentration; ICF: Informant Consensus Factor; IPNI: International Plant Names Index; ISO: International Organization for Standardization; KS: Khaya senegalensis; LD50: Median Lethal Dose; LF: Lymphatic filariasis; Mf: Microfilariae; NF: Norme Française; NGO: Non-Governmental Organization; Nt: Number of species used in the treatment of a given disease; Nuc: Number of citations for the treatment of a given disease; PBMCs: Peripheral Blood Mononuclear Cells; PBS: Phosphate-buffered saline; PH: Propidium lodide; RPMI: Roswell Park Memorial Institute medium; THs: Traditional Healers; UV: Used Value; XA: Xylopia aethiopica; CBRS: Comité de Bioéthique pour la Recherche en Santé

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Authors' contributions

EA, GK, AHA, PET and OMA contributed to design the survey tool, carried out the survey and in vitro tests. GK, MR, LEL, AH and SDK designed the study protocol, analyzed, interpreted and validated all the data. KB, YA and TT contributed to the identification of the plants. GK, SDK and LEL wrote the paper which was then read and approved by all other authors.

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Availability of data and materials

The datasets during and/or analysed during the current study available from the corresponding author on reasonable request.

Ethics approval and consent to participate

The study has the authorisation code N°043/2016/MSPS/CAB/SG/DPLET/CBRS from the ethical board "Comité de Bioéthique pour la Recherche en Santé (CBRS)" of the Ministry of Health in Togo. The THs and healthy blood donors gave their informed consent and certifying their agreement with a signature (signed forms are available).

Consent for publication

Not Applicable.

Competing interests

The authors declare that they have no competing interests.

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