

Supplementary Information

Photoinduced β -Fragmentation of aliphatic alcohol derivatives for forging C–C bonds

Yiman Gao, Jie Liu, Cong Wei, Kui Zhang, Liangliang Song, Lingchao Cai*

Table of Contents

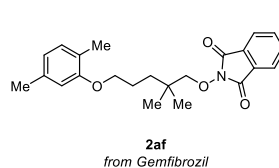
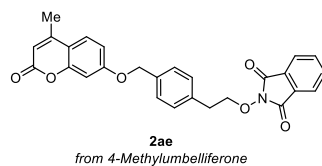
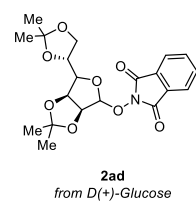
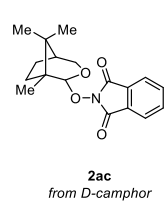
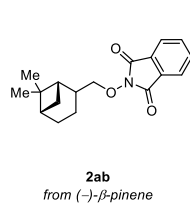
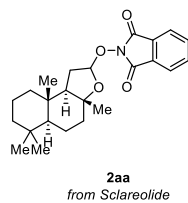
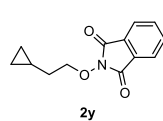
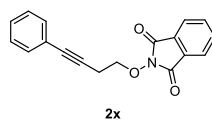
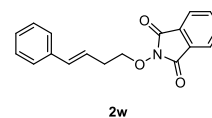
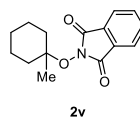
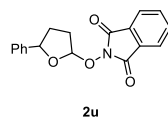
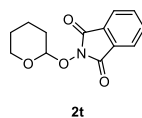
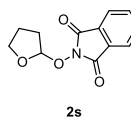
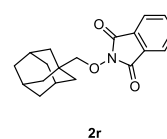
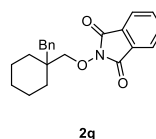
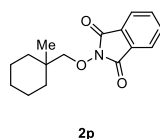
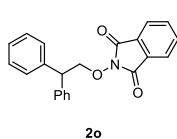
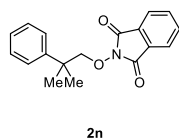
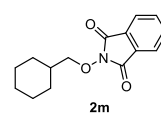
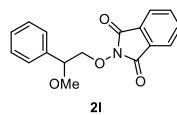
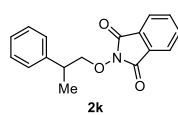
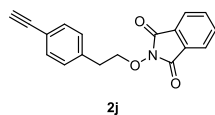
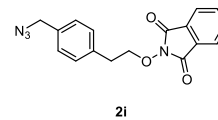
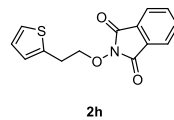
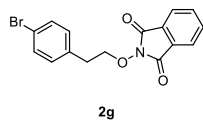
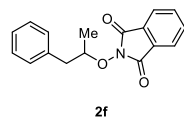
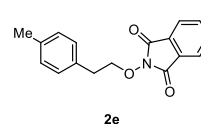
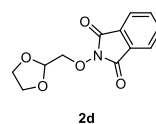
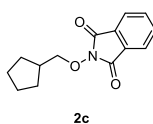
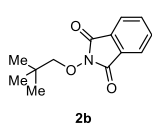
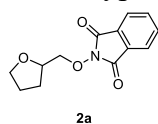
1. Supplementary Methods	3
1.1 General Information	3
2. Supplementary Discussion	4
2.1 Substrate Preparation	4
2.2 Experimental Procedures	16
2.2.1 General Procedure for the Synthesis of the products 3–62	16
2.2.2 Characterization Data for the product 3–62	17
3. Supplementary Notes	32
3.1 Reaction Profile General	32
3.2 Examining Functional Group Compatibility	33
3.3 In Vitro Antifungal Activities.	34
4. Supplementary Figures	38
4.1 NMR Spectra.....	38
5. Supplementary Reference	109

1. Supplementary Methods

1.1 General Information

All reactions were performed in dry solvents under an N₂ atmosphere and anhydrous conditions. DCM, THF, toluene, diethyl ether, and MeCN to be used in anhydrous reaction mixtures were dried by passage through activated alumina columns immediately prior to use. All other reagents were used as received from commercial sources. Reactions were monitored through thin layer chromatography (TLC) on 0.25-mm silica gel plates and visualized under UV light. Flash column chromatography (FCC) was performed using Flash silica gel (60-Å pore size, 40–63 µm). NMR spectra were recorded on Bruker Avance-400 or -600 instrument, calibrated to CD(H)Cl₃ as the internal reference (7.26 and 77.0 ppm for ¹H and ¹³C NMR spectra, respectively). ¹H NMR spectral data were reported in terms of chemical shift (δ, ppm), multiplicity, coupling constant (Hz), and integration. ¹³C NMR and ¹⁹F NMR spectral data were reported in terms of chemical shift (δ, ppm). The following abbreviations indicated the multiplicities: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad. High-resolution mass spectra were recorded using a SCIEX X500R LC-Q-TOF, ESI ion Source. The 15 W blue LED lamps were directly got from the supermarket. The crop pathogenic fungi, such as *Botrytis cinerea* (*B. cinerea*), *Sclerotinia sclerotiorum* (*S. sclerotiorum*), *Colletotrichum orbiculare* (*C. orbiculare*), *Thanatephorus cucumeris*(Frank)Donk (*T. cucumeris*), *Alternaria mali*, and *Ceratocystis paradoxa* (*C. paradoxa*), were purchased from the Agricultural Culture Collection of China, which were preserved in agar slants at 4 °C.

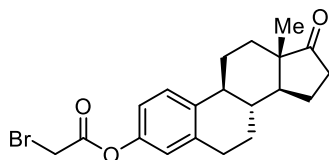
N-alkoxyphthalimides



2.1.1 Substrates 1 Preparation

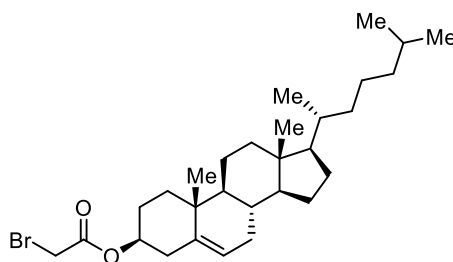
Esters, amides of *N*-aryl-substituted glycine,¹ *N*-aryl tetrahydroisoquinoline,² *N*-pyridyl-substituted glycine,³ *N*-(*p*-tolyl)-glycinitrile³⁰ were all prepared according to the previous reports.

Synthesis of *N*-aryl protected derivatives precursors



(8*R*,9*S*,13*S*,14*S*)-13-methyl-17-oxo-7,8,9,11,12,13,14,15,16,17-decahydro-6*H*-cyclopenta[*a*]phenanthren-3-yl 2-bromoacetate (1s-1)

The titled compound was synthesized following a literature procedure from estrone.⁴ Spectra are consistent with reported literature values.

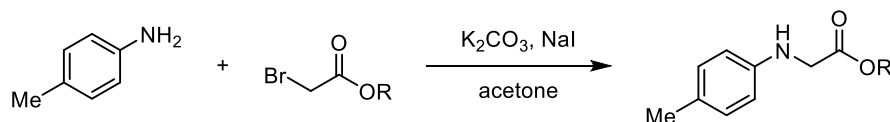


(3*S*,8*S*,9*S*,10*R*,13*R*,14*S*,17*R*)-10,13-dimethyl-17-((*R*)-6-methylheptan-2-yl)-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*-cyclopenta[*a*]phenanthren-3-yl 2-bromoacetate (1t-1)

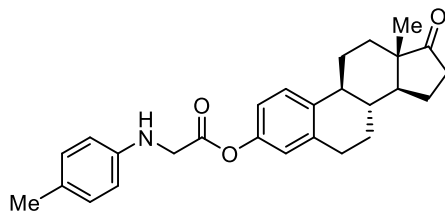
The titled compound was synthesized following a literature procedure from cholesterol.⁵ Spectra are consistent with reported literature values.

Synthesis of *N*-aryl protected derivatives

General method

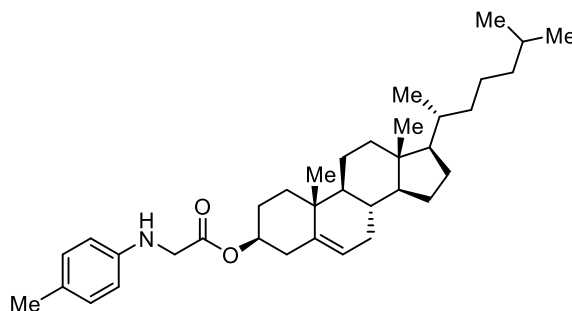


Following the literature procedure⁶ with slight modifications. The mixture of *p*-toluidine (1.1 g, 10 mmol), potassium carbonate (1.7 g, 12 mmol) and sodium iodide (1.8 g, 12 mmol) were treated with the corresponding bromoacetate-ester (11 mmol), in dry acetone as the solvent, at 60 °C for 12 h. To the resulting mixture, was cooled to room temperature, and filtered through a celite pad, then concentrated in vacuo, and subjected to flash chromatography to afford the *N*-aryl protected derivatives.



(8*R*,9*S*,13*S*,14*S*)-13-methyl-17-oxo-7,8,9,11,12,13,14,15,16,17-decahydro-6*H*-cyclopenta[*a*]phenanthren-3-yl *p*-tolylglycinate (1s**)**

Following the general method, the reaction of **1s-1** (390 mg, 1 mmol) afforded *N*-aryl protected derivatives **1s** as a yellow oil (0.17 g, 40% yield). ^1H NMR (400 MHz, CDCl_3) δ 7.31 (d, J = 8.4 Hz, 1H), 7.18 (d, J = 8.6 Hz, 1H), 7.06 (d, J = 7.5 Hz, 2H), 6.89 (d, J = 8.4 Hz, 1H), 6.84 (s, 1H), 6.64 (q, J = 8.5 Hz, 2H), 4.17 (s, 2H), 2.97–2.86 (m, 2H), 2.28 (s, 3H), 2.23–2.16 (m, 1H), 2.09–1.98 (m, 3H), 1.65–1.51 (m, 7H), 0.93 (s, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 221.0, 170.3, 153.7, 138.0, 131.9, 129.9, 126.5, 121.3, 118.5, 115.3, 113.4, 112.9, 50.5, 48.0, 46.6, 44.0, 38.4, 38.0, 35.9, 31.6, 26.5, 25.9, 21.6, 20.4, 13.9. HRMS (ESI, m/z) calcd for $\text{C}_{27}\text{H}_{32}\text{NO}_3$ ($\text{M}+\text{H}$) $^+$: 418.2377, found: 418.2372.

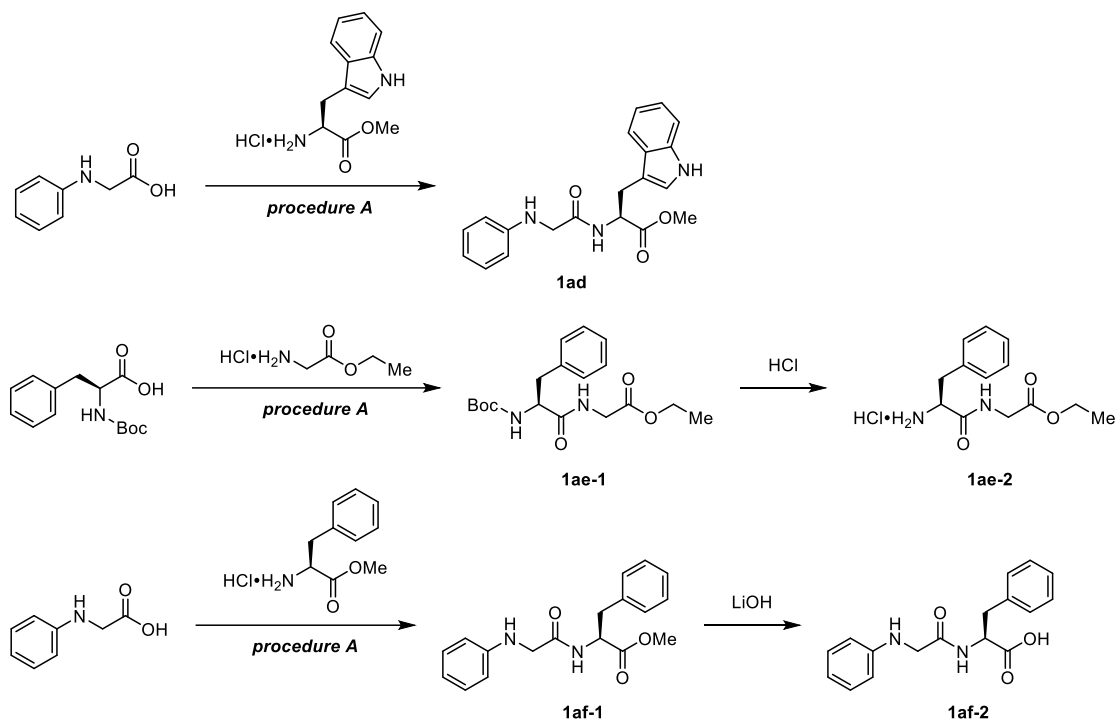


(3*S*,8*S*,9*S*,10*R*,13*R*,14*S*,17*R*)-10,13-dimethyl-17-((*R*)-6-methylheptan-2-yl)-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*-cyclopenta[*a*]phenanthren-3-yl *p*-tolylglycinate (1t**)**

Following the general method, the reaction of **1t-1** (506 mg, 1 mmol) afforded *N*-aryl protected derivatives **1t** as a white solid (0.35 g, 70% yield). m.p. 159–161 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.03 (d, J = 7.6 Hz, 2H), 6.56 (d, J = 7.6 Hz, 2H), 5.40 (s, 1H), 4.74 (d, J = 8.4 Hz, 1H), 4.17 (s, 1H), 3.88 (d, J = 4.2 Hz, 2H), 2.37 (d, J = 7.8 Hz, 2H), 2.27 (s, 3H), 2.07–1.95 (m, 2H), 1.94–1.80 (m, 3H), 1.69–1.32 (m, 7H), 1.24–0.99 (m, 17H), 0.94 (d, J = 6.2 Hz, 3H), 0.89 (d, J = 6.4 Hz, 6H), 0.70 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 170.7, 144.9, 139.4, 129.8, 127.4, 123.0, 113.2, 75.1, 56.7, 56.1, 50.0, 46.5, 42.3, 39.7, 39.5, 38.1, 36.9, 36.6, 36.2, 35.8, 31.9, 31.8, 28.2, 28.0, 27.7, 24.3, 23.8, 22.8, 22.6, 21.0, 20.4, 19.3, 18.7, 11.9. HRMS (ESI, m/z) calcd for $\text{C}_{36}\text{H}_{56}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 534.4306, found: 534.4298.

Synthesis of Peptides

Dipeptides and polypeptides were all prepared according to previous reports⁷ with slight modifications.



General Procedure A: methyl phenylglycyl-L-tryptophanate (**1ad**)

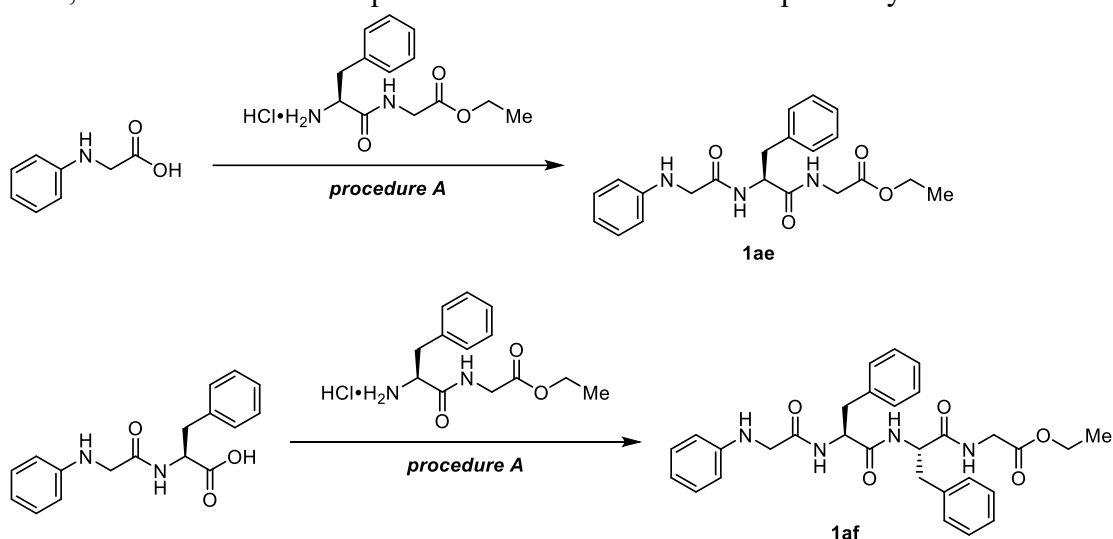
To a 100 mL round-bottom flask, *N*-phenyl glycine (3.5 mmol, 530 mg) and methyl *L*-tryptophanate hydrochloride (3.5 mmol, 891 g) were dissolved in 30 mL DCM. At 0 °C, Et₃N (7 mmol, 0.97 mL) was added and stirred for 5 min. HBTU (7 mmol, 2.65 g) was added. The reaction mixture was warmed to room temperature and stirred overnight. Subsequently, water was added to the round-bottom flask. The resulting mixture was extracted with DCM (20 mL x 3), and the combined organic layers were dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by column chromatography and the **1ad** was obtained as a pale-yellow solid. m.p. 164–165 °C; ¹H NMR (400 MHz, CDCl₃) δ 8.03 (s, 1H), 7.45 (d, *J* = 7.9 Hz, 1H), 7.31 (d, *J* = 8.1 Hz, 1H), 7.19 (q, *J* = 7.0 Hz, 4H), 7.07 (t, *J* = 7.4 Hz, 1H), 6.84 (t, *J* = 7.3 Hz, 1H), 6.60 (s, 1H), 6.54 (d, *J* = 7.7 Hz, 2H), 4.98 (m, *J* = 4.6 Hz, 1H), 3.74 (s, 2H), 3.69 (s, 3H), 3.29 (dq, *J* = 5.2, 16.2 Hz, 2H); ¹³C NMR (101 MHz, CDCl₃) δ 172.1, 170.5, 146.9, 136.1, 129.4, 127.4, 123.0, 122.2, 119.7, 119.0, 118.4, 113.3, 111.3, 109.5, 52.4, 52.2, 48.6, 27.5. HRMS (ESI, *m/z*) calcd for C₂₀H₂₂N₃O₃ (M+H)⁺: 352.1656, found: 352.1653.

ethyl *L*-phenylalanylglycinate hydrochloride (**1ae-2**)

Following the general procedure A, the reaction of glycine ethyl ester hydrochloride and *N*-(tert-butoxycarbonyl)-*L*-phenylalanine afforded the dipeptide **1ae-1** as a white solid. To a stirred solution of **1ae-1** (1 mmol, 350 mg) in dichloromethane (20 mL) was added 1N HCl (2 mL) and the resultant mixture was stirred for 60 min at room temperature. After completion of the reaction monitored by TLC, the reaction were dried over with Na₂SO₄, filtered and concentrated under reduced pressure. The desired product **1ae-2** was afforded as yellow solid, which without further purification and used in next step directly.

phenylglycyl-*L*-tryptophan (**1af-2**)

Following the general procedure A, the reaction of *N*-phenyl glycine and Methyl *L*-phenylalaninate hydrochloride afforded the dipeptide **1af-1** as a white solid. To a stirred solution of **1af-1** (1 mmol, 312 mg) in methanol (20 mL) was added 1N LiOH (2 mL) and the resultant mixture was stirred for 8 h at room temperature. After completion of the reaction monitored by TLC, the reaction was concentrated under reduced pressure to remove methanol, and then extracted with EtOAc (30 mL). The aqueous layers were collected and the pH was adjusted to 2-3 by 2N HCl. Following extraction with EtOAc (20 mL x 2), the combined organic layers were dried over with Na₂SO₄, filtered and concentrated under reduced pressure. The desired product **1af-2** was afforded as yellow solid, which without further purification and used in next step directly.



ethyl phenylglycyl-*L*-phenylalanylglycinate (**1ae**)

Following the general procedure A, the reaction of **1ae-2** afforded polypeptide **1ae** as a white solid. m.p. 135-136 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.29 (d, *J* = 7.2 Hz, 1H), 7.20 (s, 5H), 7.08 (s, 2H), 6.83 (t, *J* = 7.2 Hz, 1H), 6.75 (s, 1H), 6.54 (d, *J* = 7.8 Hz, 2H), 4.82 (q, *J* = 7.2 Hz, 1H), 4.19 (q, *J* = 7.1 Hz, 2H), 3.94 (m, *J* = 9.7 Hz, 2H), 3.76 (q, *J* = 17.6 Hz, 2H), 3.07 (t, *J* = 6.4 Hz, 2H), 1.28 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 171.2, 171.1, 169.5, 146.9, 136.2, 129.5, 129.2, 128.6, 127.0, 119.2, 113.3, 61.6, 53.8, 48.6, 41.3, 37.7, 14.1. HRMS (ESI, *m/z*) calcd for C₂₁H₂₆N₃O₄ (M+H)⁺: 384.1918, found: 384.1909.

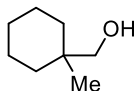
ethyl phenylglycyl-*L*-phenylalanyl-*L*-phenylalanylglycinate (**1af**)

Following the general procedure A, the reaction of **1ae-2** and **1af-2** afforded polypeptide **1af** as a white solid. m.p. 155–156 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.25–7.19 (m, 5H), 7.14 (d, *J* = 6.0 Hz, 3H), 7.07 (d, *J* = 7.2 Hz, 2H), 7.05 (d, *J* = 8.2 Hz, 1H), 6.97 (d, *J* = 5.6 Hz, 2H), 6.88 (s, 1H), 6.83 (t, *J* = 7.3 Hz, 1H), 6.54 (d, *J* = 8.0 Hz, 2H), 4.82 (q, *J* = 7.4 Hz, 1H), 4.77 (q, *J* = 7.1 Hz, 1H), 4.18 (q, *J* = 7.1 Hz, 2H), 4.01 (m, *J* = 7.9 Hz, 1H), 3.85 (m, *J* = 7.7 Hz, 1H), 3.70 (q, *J* = 16.2 Hz, 2H), 3.09 (m, *J* = 6.7 Hz, 1H), 2.96 (m, *J* = 4.3 Hz, 2H), 2.88 (m, *J* = 7.2 Hz, 1H), 1.28 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 171.3, 171.1, 170.9, 169.6, 147.0, 136.6, 136.0, 129.5, 129.4, 129.3, 128.6, 128.5, 127.0, 126.8, 119.0, 113.1, 61.5, 54.1, 54.0, 48.2,

41.3, 38.0, 37.8, 14.2. HRMS (ESI, m/z) calcd for $C_{30}H_{35}N_4O_5$ ($M+H$)⁺: 531.2602, found: 531.2605.

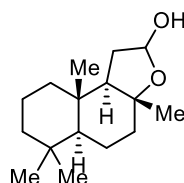
2.1.2 Substrates 2 Preparation

The *N*-alkoxyphthalimides **2a–d**, **2f**, **2k**, **2m–n**,⁸ **2l**,⁹ **2w**,¹⁰ **2x**,¹¹ **2s–u**,¹² **2v**,¹³ **2ad**¹⁴ were prepared using reported methods.



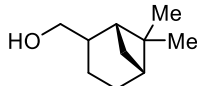
(1-methylcyclohexyl)methanol (1p-1)

The titled compound was synthesized following a literature procedure from 1-methylcyclohexanecarboxaldehyde.¹⁵ Spectra are consistent with reported literature values.



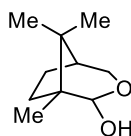
(3aR,5aS,9aS,9bR)-3a,6,6,9a-tetramethyldodecahydronaphtho[2,1-*b*]furan-2-ol (2aa-1)

The titled compound was synthesized following a literature procedure from sclareolide.¹⁶ Spectra are consistent with reported literature values.



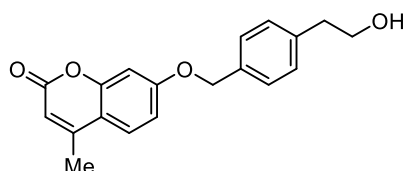
((1R,5R)-6,6-dimethylbicyclo[3.1.1]heptan-2-yl)methanol (2ab-1)

The titled compound was synthesized following a literature procedure from (1S)-(1)-beta-pinene.¹⁷ Spectra are consistent with reported literature values.



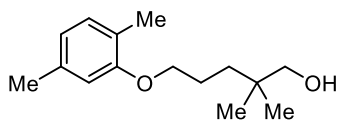
1,8,8-trimethyl-3-oxabicyclo[3.2.1]octan-2-ol (2ac-1)

The titled compound was synthesized following a literature procedure from *D*-camphor.¹⁸ Spectra are consistent with reported literature values.



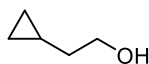
7-((4-(2-hydroxyethyl)benzyl)oxy)-4-methyl-2H-chromen-2-one (2ae-1)

The titled compound was synthesized following a literature procedure from 7-hydroxy-4-methyl-chromen-2-one.¹⁹ Spectra are consistent with reported literature values.



5-(2,5-dimethylphenoxy)-2,2-dimethylpentan-1-ol (2af-1)

The titled compound was synthesized following a literature procedure from gemfibrozil.²⁰ Spectra are consistent with reported literature values.

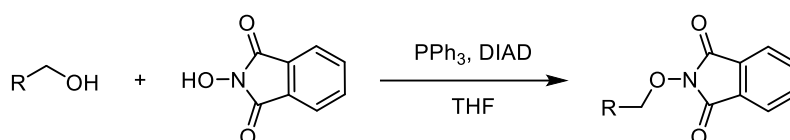


2-cyclopropylethan-1-ol (2y-1)

The titled compound was synthesized following a literature procedure from cyclopropylacetic acid.²¹ Spectra are consistent with reported literature values.

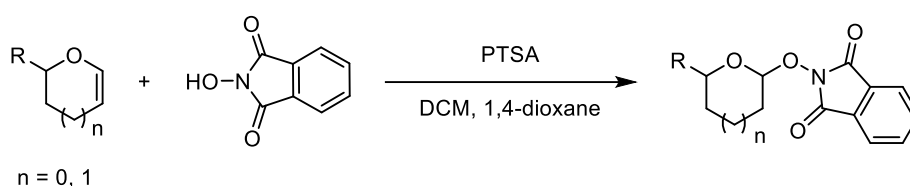
Synthesis of *N*-alkoxyphthalimides

Method A



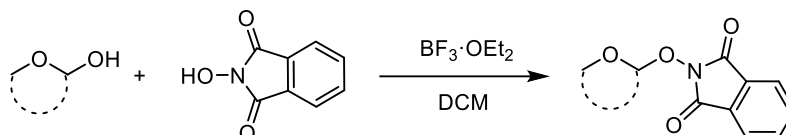
Following the literature procedure²² with slight modifications. To a solution of the alcohol (10.0 mmol), PPh₃ (3.15 g, 12.0 mmol), and *N*-hydroxyphthalimide (1.96 g, 12.0 mmol) in THF (30 mL) was added diisopropyl azodicarboxylate (2.4 mL, 12.0 mmol) over 10 min at room temperature. The resulting mixture was stirred for 3-24 h, taken up in EtOAc (20 mL), and washed with saturated NaHCO₃ (3 x 20 mL) and brine (2 x 30 mL). The organic layers were dried over anhydrous Na₂SO₄, concentrated in vacuo, and subjected to flash chromatography to afford the *N*-alkoxyphthalimides.

Method B

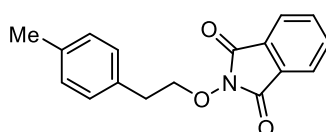


Following the literature procedure²³ with slight modifications. To a solution of the *N*-hydroxyphthalimide (1.5 g, 9.2 mmol) in DCM (30 mL) were added 1,4-dioxane (20 mL), PTSA (31.6 mg, 0.02 mmol) and dihydropyran or dihydrofuran (13.6 mmol) at room temperature. The reaction mixture was stirred for 3 h. To the resulting mixture, DCM (10 mL) and saturated Na₂CO₃ solution in H₂O (50 mL) was added. The aqueous layer was extracted with DCM (20 mL x 3), the combined organic layers were dried over anhydrous Na₂SO₄, concentrated in vacuo, and subjected to flash chromatography to afford the *N*-alkoxyphthalimides.

Method C

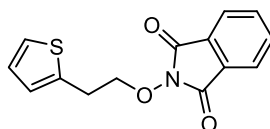


Following the literature procedure²⁴ with slight modifications. To a solution of hemiacetal (10 mmol) and *N*-hydroxyphthalimide (3.26 g, 20 mmol) in 70 mL wet DCM, then $\text{BF}_3 \cdot \text{OEt}_2$ (6.8 mL, 25 mmol) was added dropwise slowly at 0 °C. The reaction mixture was stirred for 5 h at room temperature. To the resulting mixture, DCM (10 mL) and saturated Na_2CO_3 solution in H_2O (50 mL) was added. The aqueous layer was extracted with DCM (20 mL x 3), the combined organic layers were dried over anhydrous Na_2SO_4 , concentrated in vacuo, and subjected to flash chromatography to afford the *N*-alkoxyphthalimides.



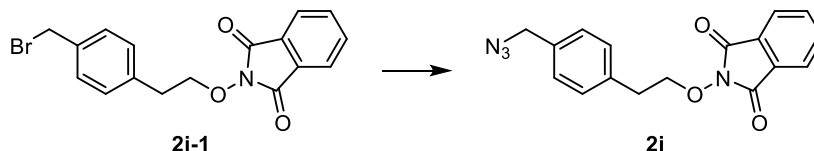
2-(4-methylphenethoxy)isoindoline-1,3-dione (**2e**)

Following the general method A, the reaction of 2-(p-Tolyl)ethanol (408 mg, 3 mmol) afforded *N*-alkoxyphthalimides **2e** as a white solid (0.67 g, 79% yield). m.p. 82–84 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.85 (m, 2H), 7.76 (m, 2H), 7.20 (d, $J = 7.4$ Hz, 2H), 7.13 (d, $J = 7.4$ Hz, 2H), 4.43 (t, $J = 7.3$ Hz, 2H), 3.13 (t, $J = 7.3$ Hz, 2H), 2.32 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.6, 136.2, 134.5, 133.6, 129.3, 128.9, 128.7, 123.5, 78.6, 34.2, 21.0. HRMS (ESI, m/z) calcd for $\text{C}_{17}\text{H}_{16}\text{NO}_3$ ($\text{M}+\text{H}$)⁺: 282.1125, found: 282.1125.



2-(2-(thiophen-2-yl)ethoxy)isoindoline-1,3-dione (**2h**)

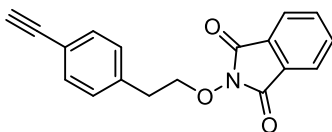
Following the general method A, the reaction of 2-Thiopheneethanol (1.28 g, 10 mmol) afforded *N*-alkoxyphthalimides **2h** as a white solid (1.53 g, 56% yield). m.p. 67–69 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.86 (m, 2H), 7.77 (m, 2H), 7.18 (d, $J = 4.8$ Hz, 1H), 6.98 (m, 2H), 4.46 (t, $J = 6.9$ Hz, 2H), 3.38 (t, $J = 7.1$ Hz, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.6, 138.7, 134.6, 128.9, 127.0, 125.8, 124.1, 123.6, 78.2, 29.0. HRMS (ESI, m/z) calcd for $\text{C}_{14}\text{H}_{12}\text{NO}_3\text{S}$ ($\text{M}+\text{H}$)⁺: 274.0533, found: 274.0537.



2-(4-(azidomethyl)phenethoxy)isoindoline-1,3-dione (**2i**)

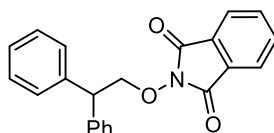
Following the general method A, the reaction of 2-(4-(bromomethyl)phenyl)ethanol afforded *N*-alkoxyphthalimides **2i-1**. The **2i** was synthesized following a literature procedure²⁵ from **2i-1** as a white solid (0.84 g, 60% yield). m.p. 112–114 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.85 (m, 2H), 7.77 (m, 2H), 7.36 (d, $J = 7.6$ Hz, 2H), 7.28 (d, $J =$

7.4 Hz, 2H), 4.46 (t, $J = 7.1$ Hz, 2H), 4.32 (s, 2H), 3.18 (t, $J = 7.2$ Hz, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.6, 137.1, 134.6, 133.8, 129.4, 128.9, 128.5, 123.6, 78.3, 54.5, 34.3. HRMS (ESI, m/z) calcd for $\text{C}_{17}\text{H}_{18}\text{N}_5\text{O}_3$ ($\text{M}+\text{NH}_4$) $^+$: 340.1404, found: 340.1406.



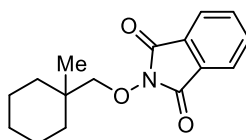
2-(4-ethynylphenethoxy)isoindoline-1,3-dione (**2j**)

Following the general method A, the reaction of 2-(4-ethynylphenyl)ethanol (292 mg, 2 mmol) afforded *N*-alkoxyphthalimides **2j** as a white solid (349 mg, 60% yield). m.p. 111–112 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.85 (m, 1H), 7.78 (m, 1H), 7.46 (d, $J = 7.6$ Hz, 2H), 7.30 (d, $J = 8.4$ Hz, 2H), 4.45 (t, $J = 7.0$ Hz, 2H), 3.17 (t, $J = 7.1$ Hz, 2H), 3.07 (s, 1H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.6, 137.8, 134.6, 132.3, 128.9 (2C), 123.6, 120.4, 83.5, 78.1, 77.0, 34.5. HRMS (ESI, m/z) calcd for $\text{C}_{18}\text{H}_{17}\text{N}_2\text{O}_3$ ($\text{M}+\text{NH}_4$) $^+$: 309.1233, found: 309.1242.



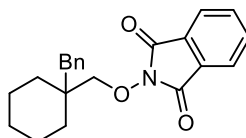
2-(2,2-diphenylethoxy)isoindoline-1,3-dione (**2o**)

Following the general method A, the reaction of 2,2-diphenyl ethanol (396 mg, 2 mmol) afforded *N*-alkoxyphthalimides **2o** as a white solid (500 mg, 73% yield). m.p. 155–156 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.82 (m, 2H), 7.75 (m, 2H), 7.42–7.30 (m, 8H), 7.23 (t, $J = 7.0$ Hz, 2H), 4.79 (d, $J = 7.4$ Hz, 2H), 4.65 (t, $J = 7.4$ Hz, 1H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.4, 140.5, 134.5, 128.8, 128.7, 128.3, 126.9, 123.5, 79.9, 49.8. HRMS (ESI, m/z) calcd for $\text{C}_{18}\text{H}_{17}\text{N}_2\text{O}_3$ ($\text{M}+\text{NH}_4$) $^+$: 361.1546, found: 361.1544.



2-((1-methylcyclohexyl)methoxy)isoindoline-1,3-dione (**2p**)

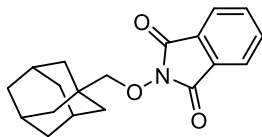
Following the general method A, the reaction of **2p-1** (256 mg, 2 mmol) afforded *N*-alkoxyphthalimides **2p** as a colorless oil (0.29 g, 51% yield). ^1H NMR (400 MHz, CDCl_3) δ 7.81 (m, 2H), 7.73 (m, 2H), 3.94 (s, 2H), 1.61–1.31 (m, 10H), 1.12 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.4, 134.3, 129.0, 123.3, 86.5, 34.5, 34.3, 26.2, 22.9, 21.6. HRMS (ESI, m/z) calcd for $\text{C}_{16}\text{H}_{22}\text{NO}_4$ ($\text{M}+\text{H}_2\text{O}+\text{H}$) $^+$: 291.1544, found: 291.1541.



2-((1-benzylcyclohexyl)methoxy)isoindoline-1,3-dione (**2q**)

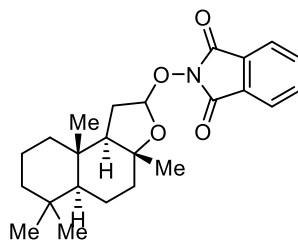
Following the general method A, the reaction of (1-benzylcyclohexyl)methanol (817 mg, 4 mmol) afforded *N*-alkoxyphthalimides **2q** as a white solid (0.84 g, 60% yield). m.p. 132–133 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.86 (m, 1H), 7.77 (m, 1H), 7.47 (d,

$J = 7.4$ Hz, 2H), 7.36 (t, $J = 7.3$ Hz, 2H), 7.26 (q, $J = 6.8$ Hz, 1H), 3.99 (s, 2H), 2.85 (s, 2H), 1.67–1.37 (m, 10H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.5, 137.7, 134.4, 131.1, 129.1, 127.9, 126.1, 123.4, 80.8, 42.1, 37.9, 32.6, 26.1, 21.6. HRMS (ESI, m/z) calcd for $\text{C}_{22}\text{H}_{24}\text{NO}_3$ ($\text{M}+\text{H}$) $^+$: 350.1751, found: 350.1749.



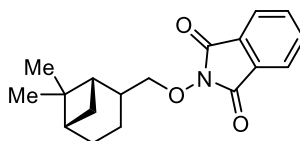
2-(((3*r*,5*r*,7*r*)-adamantan-1-yl)methoxy)isoindoline-1,3-dione (**2r**)

Following the general method A, the reaction of 1-Adamantanemethanol (831 mg, 5 mmol) afforded *N*-alkoxyphthalimides **2r** as a white solid (0.68 g, 44% yield). m.p. 139–140 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.83 (m, 2H), 7.75 (m, 2H), 3.80 (s, 3H), 2.04 (s, 12H), 1.75 (s, 1H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.5, 134.3, 129.1, 123.4, 88.2, 39.0, 37.0, 34.0, 28.0. HRMS (ESI, m/z) calcd for $\text{C}_{19}\text{H}_{22}\text{NO}_3$ ($\text{M}+\text{H}$) $^+$: 312.1594, found: 312.1599.



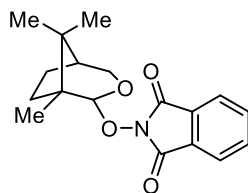
2-(((3*aR*,5*aS*,9*aS*,9*bR*)-3*a*,6,6,9*a*-tetramethyldodecahydronaphtho[2,1-*b*]furan-2-yl)oxy)isoindoline-1,3-dione (**2aa**)

Following the general method C, the reaction of **2aa-1** (250 mg, 1 mmol) afforded *N*-alkoxyphthalimides **2aa** as a white solid (0.14 g, 36% yield). m.p. 137–138 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.82 (m, 2H), 7.74 (m, 2H), 5.79 (t, $J = 5.5$ Hz, 1H), 2.30 (m, 1H), 2.10–1.98 (m, 2H), 1.84–1.65 (m, 2H), 0.95 (s, 3H), 1.53–1.35 (m, 5H), 1.29–0.99 (m, 5H), 0.88 (d, $J = 10.0$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.6, 134.3, 129.1, 123.3, 110.1, 84.1, 60.5, 57.0, 42.4, 39.7, 36.2, 33.5, 33.1, 28.9, 23.0, 21.0, 20.6, 18.3, 15.2. HRMS (ESI, m/z) calcd for $\text{C}_{24}\text{H}_{35}\text{N}_2\text{O}_4$ ($\text{M}+\text{NH}_4$) $^+$: 415.2591, found: 415.2596.



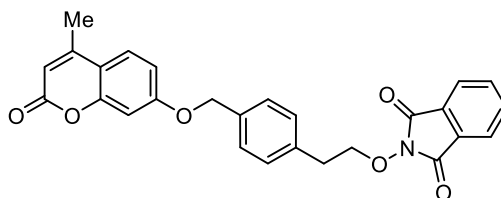
2-(((1*R*,5*R*)-6,6-dimethylbicyclo[3.1.1]heptan-2-yl)methoxy)isoindoline-1,3-dione (**2ab**)

Following the general method A, the reaction of **2ab-1** (926 mg, 6 mmol) afforded *N*-alkoxyphthalimides **2ab** as a white solid (0.90 g, 50% yield). ^1H NMR (400 MHz, CDCl_3) δ 7.84 (m, 2H), 7.76 (m, 2H), 4.19 (m, $J = 7.3$ Hz, 2H), 2.61 (m, 1H), 2.40 (m, 1H), 2.19–1.86 (m, 5H), 1.69 (m, $J = 6.3$ Hz, 1H), 1.23 (s, 3H), 1.01 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 163.5, 134.4, 128.9, 123.3, 83.3, 43.1, 41.1, 39.6, 38.4, 32.5, 27.7, 25.8, 23.2, 18.6. HRMS (ESI, m/z) calcd for $\text{C}_{18}\text{H}_{22}\text{NO}_3$ ($\text{M}+\text{H}$) $^+$: 300.1594, found: 300.1599.



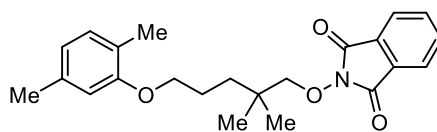
2-(((1S,2R,5S)-1,8,8-trimethyl-3-oxabicyclo[3.2.1]octan-2-yl)oxy)isoindoline-1,3-dione (2ac)

Following the general method C, the reaction of **2ac-1** (340 mg, 2 mmol) afforded *N*-alkoxyphthalimides **2ac** as a white solid (252 mg, 40% yield). m.p. 138–139 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.82 (m, 2H), 7.73 (m, 2H), 4.96 (s, 1H), 4.61 (d, *J* = 10.7 Hz, 1H), 3.45 (d, *J* = 10.7 Hz, 1H), 1.93–1.83 (m, 1H), 1.78–1.57 (m, 5H), 1.47 (s, 3H), 1.20 (s, 3H), 0.94 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 163.8, 134.2, 129.2, 123.3, 112.4, 67.8, 46.5, 45.7, 40.9, 35.4, 25.5, 24.5, 20.6, 15.4. HRMS (ESI, *m/z*) calcd for C₁₈H₂₁NO₄Na (M+Na)⁺: 338.1363, found: 338.1362.



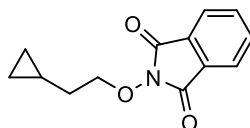
2-(4-(((4-methyl-2-oxo-2H-chromen-7-yl)oxy)methyl)phenyl)phenylthio)isoindoline-1,3-dione (2ae)

Following the general method A, the reaction of **2ae-1** (472 mg, 2 mmol) afforded *N*-alkoxyphthalimides **2ae** as a white solid (0.49 g, 65% yield). m.p. 196–200 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.88–7.83 (m, 2H), 7.77 (m, 2H), 7.52 (d, *J* = 8.8 Hz, 1H), 7.43–7.34 (m, 4H), 6.98–6.87 (m, 2H), 6.15 (s, 1H), 5.11 (s, 2H), 4.46 (t, *J* = 7.4 Hz, 2H), 3.19 (t, *J* = 7.3 Hz, 3H), 2.41 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 163.6, 161.7, 161.3, 155.2, 152.5, 137.2, 134.5, 134.2, 129.3, 128.9, 127.9, 125.6, 123.6, 113.8, 112.9, 112.1, 101.9, 78.4, 70.3, 34.4, 18.7. HRMS (ESI, *m/z*) calcd for C₂₇H₂₂NO₆ (M+H)⁺: 456.1442, found: 456.1450.



2-((5-(2,5-dimethylphenoxy)-2,2-dimethylpentyl)oxy)isoindoline-1,3-dione (2af)

Following the general method A, the reaction of **2af-1** (472 mg, 2 mmol) afforded *N*-alkoxyphthalimides **2af** as a colorless oil (0.49 g, 65% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.85 (m, 2H), 7.76 (m, 2H), 7.03 (d, *J* = 7.7 Hz, 1H), 6.68 (m, 2H), 4.00 (m, 4H), 2.35 (s, 3H), 2.22 (s, 3H), 1.93 (m, *J* = 2.8 Hz, 2H), 1.65 (m, *J* = 4.2 Hz, 2H), 1.15 (s, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 163.4, 157.1, 136.4, 134.4, 130.3, 129.3, 123.6, 123.4, 120.6, 112.1, 86.1, 68.4, 35.3, 34.4, 24.4, 24.1, 21.4, 15.8. HRMS (ESI, *m/z*) calcd for C₂₃H₂₈NO₄ (M+H)⁺: 382.2013, found: 382.2014.



2-(2-cyclopropylethoxy)isoindoline-1,3-dione (**2y**)

Following the general method A, the reaction of **2y-1** (517 mg, 6 mmol) afforded *N*-alkoxyphthalimides **2y** as a white solid (1.0 g, 72% yield). m.p. 63–65 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.85 (m, 2H), 7.76 (m, 2H), 4.29 (t, *J* = 6.9 Hz, 1H), 1.71 (q, *J* = 6.8 Hz, 1H), 0.89 (t, *J* = 6.3 Hz, 1H), 0.51 (d, *J* = 7.6 Hz, 1H), 0.13 (d, *J* = 4.6 Hz, 1H); ¹³C NMR (101 MHz, CDCl₃) δ 163.7, 134.4, 129.0, 123.5, 78.6, 33.1, 7.2, 4.3. HRMS (ESI, *m/z*) calcd for C₁₃H₁₄NO₃ (M+H)⁺: 232.0968, found: 232.0972.

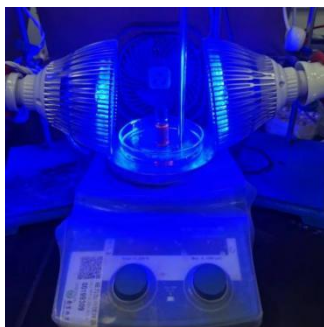
2.2 Experimental Procedures

2.2.1 General Procedure for the Synthesis of the products **3–62**

The *N*-alkoxyphthalimides (0.1 mmol), the esters, or amides of *N*-aryl-substituted glycine, or *N*-aryl tetrahydroisoquinoline, or peptides (0.2 mmol, 2.0 equiv.), 4CzIPN (5 mol%, 0.005 mmol, 4.0 mg), and DMSO (3 mL, 0.03 M) were added sequentially to a 4 mL clear-colored glass vial equipped with a magnetic stir bar. After bubbled with nitrogen gas for 5 minutes to remove oxygen, the vial was sealed and exposed to blue LEDs at 60 °C (Supplementary Figure 1). The reaction mixture was monitored by TLC until the starting material *N*-alkoxyphthalimides were consumed. Then, the reaction was quenched with water (2 mL), extracted with ethyl acetate, washed with brine, dried over anhydrous Na₂SO₄, concentrated in vacuo, and purified by column chromatography to yield the products **3–62**.

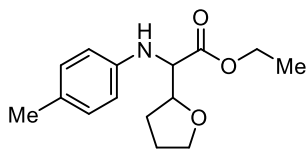
Details for the photoreactor setup

The reaction vial was placed in a 60 °C oil bath (the oil level height is 0.5 cm) at the center of a stir plate. Two parallel Blue LED lamps (E27PAR30/38, 15W, were purchased from <https://m.tb.cn/h.U2gWmW5?tk=UErD2y9tSM1>) were placed perpendicular to the sidewall of reaction vial so that the reaction vial can be equally exposed to the LEDs (at approximately 4 cm away from the light source, λ = 460–470 nm, 100–110 lm/W). A fan opposite to the reaction vial is always turned on during the reaction to offset the heat generated by the LED light.

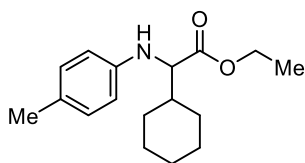


Supplementary Figure 1. Photoreactor setup

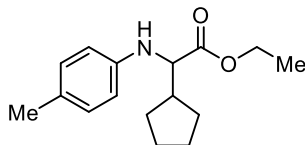
2.2.2 Characterization Data for the product 3–62.



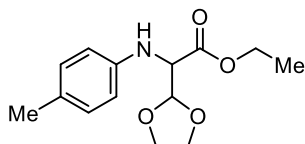
3: Yellow oil (24.3 mg, 92% yield); R_f = 0.40 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.00 (d, J = 7.9 Hz, 2H), 6.64 (d, J = 7.9 Hz, 1H), 6.59 (d, J = 7.9 Hz, 1H), 4.39–3.75 (m, 7H), 2.25 (s, 3H), 2.10–1.86 (m, 4H), 1.26 (t, J = 7.2 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.7, 172.4, 145.1, 144.6, 129.8, 129.7, 127.8, 127.6, 114.1, 113.9, 79.9, 79.4, 69.2, 68.7, 61.2 (2C), 61.1, 60.2, 28.4, 28.0, 26.0, 25.6, 20.4, 14.2 (2C). Spectra are consistent with reported literature²⁷ values.



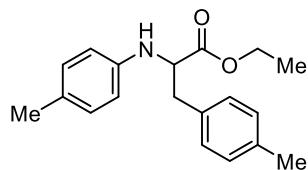
4: Yellow oil (24.5 mg, 89% yield); R_f = 0.67 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 6.99 (d, J = 8.0 Hz, 2H), 6.57 (d, J = 8.0 Hz, 2H), 4.18 (q, J = 7.1 Hz, 2H), 4.02 (s, 1H), 3.84 (d, J = 4.8 Hz, 1H), 2.24 (s, 3H), 1.87–1.68 (m, 6H), 1.29–1.15 (m, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.9, 145.2, 129.8, 127.3, 113.7, 62.5, 60.7, 41.3, 29.6, 29.2, 26.1, 20.4, 14.3. Spectra are consistent with reported literature¹ values.



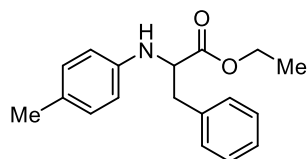
5: Yellow oil (46.4 mg, 89% yield); R_f = 0.62 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 6.99 (d, J = 7.9 Hz, 2H), 6.58 (d, J = 7.8 Hz, 2H), 4.17 (q, J = 7.1 Hz, 2H), 4.02 (s, 1H), 3.85 (d, J = 7.8 Hz, 1H), 2.25 (s, 3H), 1.92–1.42 (m, 9H), 1.25 (t, J = 7.2 Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 174.4, 145.1, 129.8, 127.5, 113.7, 61.3, 60.7, 43.2, 29.4, 29.1, 25.4, 25.1, 20.4, 14.3. Spectra are consistent with reported literature¹ values.



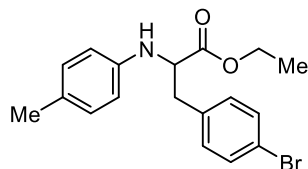
6: Yellow oil (30.7 mg, 60% yield); R_f = 0.32 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.00 (d, J = 7.7 Hz, 2H), 6.65 (d, J = 7.6 Hz, 2H), 5.38 (s, 1H), 4.35 (s, 1H), 4.25 (q, J = 7.6 Hz, 2H), 4.15–3.91 (m, 4H), 2.25 (s, 3H), 1.29 (t, J = 6.4 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 170.5, 144.7, 129.7, 127.9, 114.1, 103.5, 66.0, 65.6, 61.6, 60.7, 20.4, 14.2. Spectra are consistent with reported literature²⁸ values.



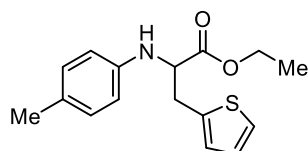
7: Yellow oil (21.7 mg, 73% yield); R_f = 0.68 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.11 (q, J = 7.2 Hz, 4H), 7.00 (d, J = 7.9 Hz, 2H), 6.56 (d, J = 7.8 Hz, 2H), 4.32 (t, J = 6.1 Hz, 1H), 4.14 (q, J = 6.7 Hz, 2H), 4.06 (s, 1H), 3.11 (t, J = 5.4 Hz, 2H), 2.34 (s, 3H), 2.26 (s, 3H), 1.21 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.4, 144.2, 136.5, 133.3, 129.8, 129.2 (2C), 127.6, 113.8, 61.0, 58.2, 38.3, 21.1, 20.4, 14.2. HRMS (ESI, m/z) calcd for $\text{C}_{19}\text{H}_{24}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 298.1802, found: 298.1794.



8: Yellow oil (13 mg, 46% yield); R_f = 0.79 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.29 (q, J = 7.3 Hz, 3H), 7.21 (d, J = 7.2 Hz, 2H), 7.01 (d, J = 8.1 Hz, 2H), 6.57 (d, J = 8.2 Hz, 2H), 4.34 (t, J = 6.3 Hz, 1H), 4.14 (dq, J = 2.2, 7.0 Hz, 2H), 4.13 (s, 1H), 3.14 (d, J = 6.3 Hz, 2H), 2.26 (s, 3H), 1.19 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.3, 144.1, 136.5, 129.8, 129.3, 128.5, 127.7, 126.9, 113.8, 61.0, 58.2, 38.7, 20.4, 14.1. HRMS (ESI, m/z) calcd for $\text{C}_{18}\text{H}_{22}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 284.1645, found: 284.1646.

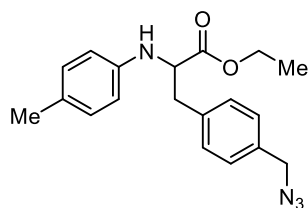


9: Yellow oil (13 mg, 54% yield); R_f = 0.55 (Petroleum ether/EtOAc, 5:1); ^1H NMR (600 MHz, CDCl_3) δ 7.44 (d, J = 8.1 Hz, 2H), 7.08 (d, J = 8.1 Hz, 2H), 7.02 (d, J = 8.0 Hz, 2H), 6.58 (d, J = 8.1 Hz, 2H), 4.33 (t, J = 6.2 Hz, 1H), 4.14 (q, J = 6.9 Hz, 2H), 3.10 (m, J = 6.4 Hz, 2H), 2.27 (s, 3H), 1.22 (t, J = 7.1 Hz, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 173.0, 143.8, 135.5, 131.5, 131.1, 129.9, 127.9, 120.9, 113.9, 61.2, 58.0, 38.0, 20.4, 14.2. HRMS (ESI, m/z) calcd for $\text{C}_{18}\text{H}_{20}\text{BrNO}_2$ ($\text{M}+\text{H}$) $^+$: 362.0750, found: 362.0744.

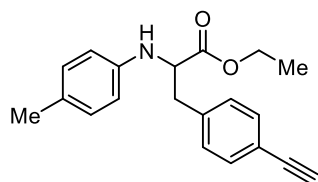


10: Yellow oil (23.7 mg, 82% yield); R_f = 0.58 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.19 (d, J = 4.7 Hz, 1H), 7.03 (d, J = 7.7 Hz, 2H), 6.96 (d, J = 2.8 Hz, 1H), 6.87 (s, 1H), 6.60 (d, J = 7.6 Hz, 2H), 4.36 (t, J = 5.7 Hz, 1H), 4.19 (q, J = 7.1 Hz, 2H), 3.39 (t, J = 7.3 Hz, 2H), 2.27 (s, 3H), 1.26 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.7, 143.9, 138.1, 129.9, 127.8, 126.8, 126.6, 124.8, 114.0, 61.3, 58.0,

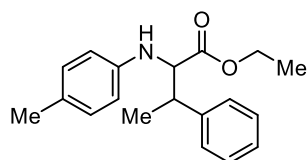
32.8, 20.5, 14.3, 14.2. HRMS (ESI, m/z) calcd for $C_{16}H_{20}NO_2S$ ($M+H$)⁺: 290.1209, found: 290.1211.



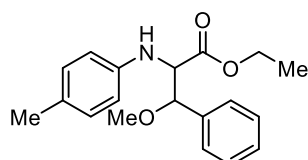
11: Yellow oil (28.7 mg, 85% yield); R_f = 0.36 (Petroleum ether/EtOAc, 5:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.25 (q, J = 10.4 Hz, 4H), 7.01 (d, J = 8.1 Hz, 2H), 6.57 (d, J = 8.4 Hz, 2H), 4.34 (d, J = 6.2 Hz, 3H), 4.13 (t, J = 7.2 Hz, 2H), 3.15 (q, J = 3.4 Hz, 2H), 2.26 (s, 3H), 1.19 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 173.2, 144.0, 136.8, 134.0, 129.9, 129.8, 128.4, 127.8, 113.9, 61.1, 58.1, 54.5, 38.4, 20.4, 14.2. HRMS (ESI, m/z) calcd for $C_{19}H_{23}N_4O_2$ ($M+H$)⁺: 339.1816, found: 339.1818.



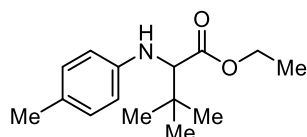
12: Yellow oil (20 mg, 65% yield); R_f = 0.54 (Petroleum ether/EtOAc, 5:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.43 (d, J = 7.8 Hz, 2H), 7.16 (d, J = 7.8 Hz, 2H), 7.01 (d, J = 7.7 Hz, 2H), 6.58 (d, J = 7.8 Hz, 2H), 4.33 (t, J = 6.0 Hz, 1H), 4.13 (t, J = 3.5 Hz, 2H), 3.14 (d, J = 5.9 Hz, 2H), 3.08 (s, 1H), 2.26 (s, 3H), 1.19 (t, J = 7.1 Hz, 3H); ^{13}C NMR (151 MHz, $CDCl_3$) δ 173.0, 143.9, 137.5, 132.2, 129.9, 129.4, 127.9, 120.7, 113.9, 83.5, 77.2, 61.2, 58.0, 38.5, 20.4, 14.2. HRMS (ESI, m/z) calcd for $C_{20}H_{22}NO_2$ ($M+H$)⁺: 308.1645, found: 308.1635.



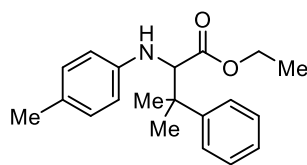
13: Yellow oil (18.5 mg, 65% yield); R_f = 0.62 (Petroleum ether/EtOAc, 5:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.37–7.23 (m, 10H), 6.97 (d, J = 7.9 Hz, 4H), 6.53 (t, J = 7.9 Hz, 6H), 4.22–4.12 (m, 4H), 4.08 (s, 1H), 3.96 (q, J = 6.8 Hz, 2H), 4.08 (s, 1H), 3.32 (m, J = 6.9 Hz, 1H), 3.20 (m, J = 6.9 Hz, 1H), 2.24 (s, 6H), 1.48 (d, J = 7.1 Hz, 3H), 1.43 (d, J = 7.0 Hz, 3H), 1.24 (t, J = 7.1 Hz, 3H), 1.0 (t, J = 7.12 Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 173.5, 173.2, 144.9, 144.8, 142.3, 141.9, 129.8, 129.7, 128.6, 128.4, 127.9 (2C), 127.7, 127.6, 127.1, 127.0, 114.1, 113.9, 63.6, 63.0, 61.0, 60.7, 43.3, 42.6, 20.4, 18.4, 17.5, 14.2, 13.9. HRMS (ESI, m/z) calcd for $C_{19}H_{24}NO_2$ ($M+H$)⁺: 298.1802, found: 298.1802.



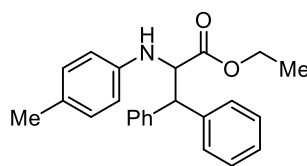
14: Yellow oil (22.8 mg, 73% yield); $R_f = 0.41$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.41–7.31 (m, 5H), 6.98 (d, $J = 8.0$ Hz, 1H), 6.92 (d, $J = 7.9$ Hz, 1H), 6.60 (d, $J = 7.9$ Hz, 1H), 6.44 (d, $J = 7.9$ Hz, 1H), 4.65 (dd, $J = 25.1, 5.1$ Hz, 1H), 4.30–4.05 (m, 3H), 3.36–3.30 (m, 3H), 2.22 (d, $J = 10.4$ Hz, 3H), 1.15 (dt, $J = 20.6, 7.2$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.9, 171.5, 144.8, 137.7, 137.4, 129.8, 129.6, 128.5, 128.4, 128.4, 128.3, 128.3, 127.7, 127.4, 127.1, 114.6, 114.0, 83.9, 83.8, 63.7, 63.2, 61.2, 61.0, 57.6, 57.4, 20.4, 20.4, 14.1, 14.0. HRMS (ESI, m/z) calcd for $\text{C}_{19}\text{H}_{24}\text{NO}_3$ ($\text{M}+\text{H}$) $^+$: 314.1751, found: 314.1750.



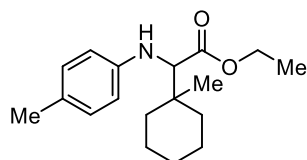
15: Yellow oil (15.6 mg, 63% yield); $R_f = 0.79$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.00 (d, $J = 7.96$ Hz, 2H), 6.61 (d, $J = 7.96$ Hz, 2H), 4.16 (q, $J = 7.06$ Hz, 2H), 4.09 (d, $J = 5.72$ Hz, 1H), 3.76 (s, 1H), 2.25 (s, 3H), 1.25 (t, $J = 7.13$ Hz, 3H), 1.08 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.5, 145.4, 129.8, 127.6, 114.1, 66.0, 60.5, 34.4, 26.8, 20.4, 14.3. HRMS (ESI, m/z) calcd for $\text{C}_{15}\text{H}_{24}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 250.1802, found: 250.1802.



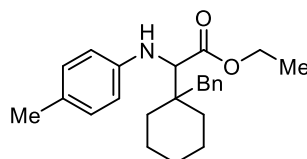
16: Yellow oil (14.4 mg, 46% yield); $R_f = 0.60$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.44 (d, $J = 7.9$ Hz, 2H), 7.35 (t, $J = 7.6$ Hz, 2H), 7.26 (t, $J = 7.9$ Hz, 1H), 6.97 (d, $J = 7.9$ Hz, 2H), 6.54 (d, $J = 7.9$ Hz, 2H), 4.14 (d, $J = 6.6$ Hz, 1H), 4.02–3.92 (m, 3H), 2.24 (s, 3H), 1.54 (d, $J = 11.9$ Hz, 6H), 1.03 (d, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.9, 145.5, 145.2, 129.7, 128.1, 127.7, 126.5 (2C), 114.2, 66.7, 60.5, 41.5, 25.7, 25.1, 20.4, 13.9. HRMS (ESI, m/z) calcd for $\text{C}_{20}\text{H}_{26}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 312.1958, found: 312.1960.



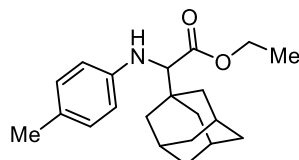
17: Yellow oil (12.6 mg, 35% yield); $R_f = 0.59$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.40–7.20 (m, $J = 7.7$ Hz, 10H), 7.00 (d, $J = 7.8$ Hz, 2H), 6.57 (d, $J = 7.8$ Hz, 2H), 4.74 (d, $J = 9.1$ Hz, 1H), 4.42 (d, $J = 9.2$ Hz, 1H), 3.94 (m, $J = 5.9$ Hz, 2H), 2.25 (s, 3H), 0.97 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.4, 144.3, 140.8, 140.0, 129.8, 128.7, 128.6, 128.5 (2C), 127.9, 127.1 (2C), 113.8, 61.1, 60.9, 54.3, 20.4, 13.8. HRMS (ESI, m/z) calcd for $\text{C}_{24}\text{H}_{26}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 360.1958, found: 360.1956.



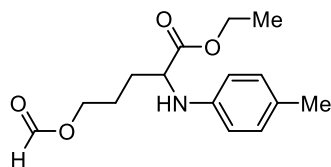
18: Yellow oil (12.4 mg, 43% yield); $R_f = 0.75$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 6.99 (d, $J = 7.8$ Hz, 2H), 6.61 (d, $J = 7.9$ Hz, 2H), 4.15 (q, $J = 7.0$ Hz, 2H), 4.09 (s, 1H), 4.06 (s, 1H), 3.92 (s, 1H), 2.25 (s, 1H), 1.25 (t, $J = 7.1$ Hz, 3H), 1.69–1.30 (m, 10H), 1.05 (s, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.5, 145.5, 129.8, 127.5, 114.1, 64.9, 60.5, 37.1, 34.9, 26.1, 21.8, 21.7, 20.4, 14.3. HRMS (ESI, m/z) calcd for $\text{C}_{18}\text{H}_{28}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 290.2115, found: 290.2120.



19: Yellow oil (12.8 mg, 35% yield); $R_f = 0.69$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.34–7.22 (m, 5H), 6.98 (d, $J = 7.9$ Hz, 2H), 6.50 (d, $J = 7.9$ Hz, 2H), 4.14 (q, $J = 7.1$ Hz, 2H), 4.02 (s, 1H), 2.99 (d, $J = 13.4$ Hz, 1H), 2.86 (d, $J = 13.4$ Hz, 1H), 2.25 (s, 3H), 1.79–1.32 (m, 10H), 1.22 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.8, 145.0, 138.2, 131.1, 129.8, 128.0, 127.6, 126.2, 114.3, 61.2, 60.5, 40.6, 31.6, 30.8, 26.0, 21.8, 21.5, 20.4, 14.3. HRMS (ESI, m/z) calcd for $\text{C}_{24}\text{H}_{32}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 366.2428, found: 366.2419.

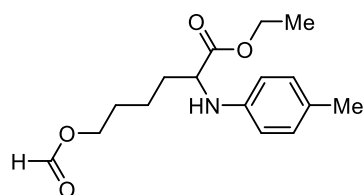


20: Yellow oil (20.5 mg, 31% yield); $R_f = 0.80$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 6.99 (d, $J = 8.0$ Hz, 2H), 6.60 (d, $J = 8.0$ Hz, 2H), 4.16 (q, $J = 7.1$ Hz, 2H), 4.10 (s, 1H), 3.64 (s, 1H), 2.25 (s, 3H), 2.05 (s, 3H), 1.85–1.56 (m, 15H), 1.26 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.0, 145.6, 129.7, 127.4, 114.0, 66.9, 60.5, 39.1, 36.9, 36.3, 28.4, 20.4, 14.4. HRMS (ESI, m/z) calcd for $\text{C}_{21}\text{H}_{30}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 328.2271, found: 328.2268.

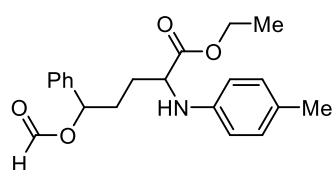


21: Yellow oil (15.9 mg, 69% yield); $R_f = 0.26$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 8.07 (s, 1H), 7.01 (d, $J = 7.9$ Hz, 2H), 6.57 (d, $J = 7.9$ Hz, 2H), 4.27–4.16 (m, 4H), 4.07 (s, 2H), 2.25 (s, 3H), 2.01–1.79 (m, 4H), 1.27 (t, $J = 7.0$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.9, 161.0, 144.4, 129.9, 127.8, 113.8, 63.4, 61.2, 56.6, 29.5, 24.8, 20.4, 14.2. HRMS (ESI, m/z) calcd for $\text{C}_{15}\text{H}_{21}\text{NO}_4$ ($\text{M}+\text{H}$) $^+$: 280.1543,

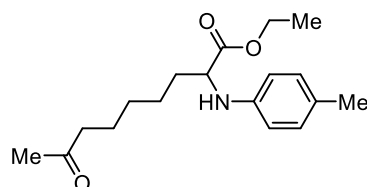
found: 280.1544.



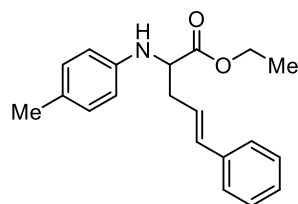
22: Yellow oil (21.2 mg, 72% yield); R_f = 0.30 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 8.07 (s, 1H), 7.00 (d, J = 7.8 Hz, 2H), 6.57 (d, J = 7.9 Hz, 2H), 4.27–4.14 (m, 4H), 4.05 (t, J = 6.0 Hz, 1H), 2.25 (s, 3H), 1.95–1.67 (m, 4H), 1.62–1.49 (m, 2H), 1.27 (t, J = 7.0 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 174.1, 161, 144.5, 129.8, 127.7, 113.7, 63.6, 61.1, 56.9, 32.6, 28.2, 22.0, 20.4, 14.3. HRMS (ESI, m/z) calcd for $\text{C}_{16}\text{H}_{24}\text{NO}_4$ ($\text{M}+\text{H}$) $^+$: 294.1700, found: 294.1696.



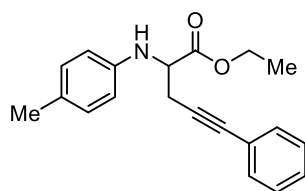
23: Yellow oil (26.8 mg, 78% yield); R_f = 0.52 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 8.12 (s, 1H), 7.33 (m, 5H), 7.00 (d, J = 7.9 Hz, 2H), 6.55 (d, J = 7.9 Hz, 2H), 5.91 (m, 1H), 4.19 (m, J = 3.5 Hz, 2H), 4.02 (m, 2H), 2.26 (s, 3H), 2.14–1.60 (m, 4H), 1.25 (m, J = 4.5 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.8, 160.3, 144.4 (2C), 139.4, 139.3, 129.9, 128.7, 128.4 (2C), 127.8 (2C), 126.6, 126.5, 113.9, 75.4, 75.3, 61.2, 56.8, 56.6, 32.3, 32.2, 29.0, 28.9, 20.4, 14.2. HRMS (ESI, m/z) calcd for $\text{C}_{21}\text{H}_{26}\text{NO}_4$ ($\text{M}+\text{H}$) $^+$: 356.1856, found: 356.1850.



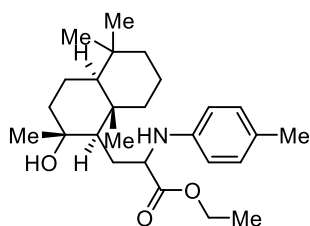
24: Yellow oil (7.1 mg, 23% yield); R_f = 0.45 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.01 (d, J = 7.6 Hz, 2H), 6.59 (d, J = 7.7 Hz, 2H), 4.28 (d, J = 6.6 Hz, 1H), 4.18 (d, J = 7.1 Hz, 2H), 4.10 (s, 1H), 2.25 (d, J = 5.4 Hz, 6H), 1.92–1.50 (m, 8H), 1.25 (t, J = 7.4 Hz, 3H), 0.90 (t, J = 6.0 Hz, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 209.7, 172.7, 144.3, 129.8, 128.0, 114.0, 61.4, 58.1, 55.1, 30.1, 29.7, 27.6, 22.8, 22.7, 20.4, 14.1, 13.8. HRMS (ESI, m/z) calcd for $\text{C}_{18}\text{H}_{28}\text{NO}_3$ ($\text{M}+\text{H}$) $^+$: 306.2064, found: 306.2060.



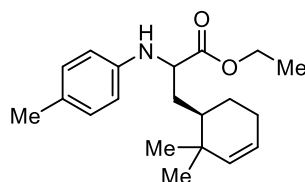
25: Yellow oil (23.8 mg, 77% yield); R_f = 0.54 (Petroleum ether/EtOAc, 5:1). Spectra are consistent with reported literature²⁶ values.



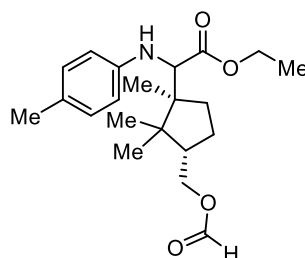
26: Yellow oil (20 mg, 65% yield); $R_f = 0.43$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.44–7.30 (m, 5H), 7.04 (d, $J = 7.6$ Hz, 2H), 6.64 (d, $J = 7.5$ Hz, 2H), 4.38–4.23 (m, 3H), 3.00 (d, $J = 5.3$ Hz, 2H), 2.27 (s, 3H), 1.31 (t, $J = 7.0$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.3, 143.9, 131.7, 129.9, 129.8, 128.2, 128.1, 128.0, 123.2, 114.2, 84.3, 83.6, 61.5, 55.9, 23.9, 20.4, 14.3, 14.2. HRMS (ESI, m/z) calcd for $\text{C}_{20}\text{H}_{22}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 308.1645, found: 308.1653.



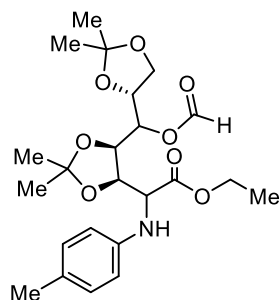
27: Yellow oil (23.8 mg, 57% yield); $R_f = 0.82$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 6.99 (d, $J = 7.8$ Hz, 2H), 6.61 (t, $J = 7.8$ Hz, 2H), 4.28–3.90 (m, 4H), 2.25 (s, 3H), 1.80–1.50 (m, 6H), 1.49–1.33 (m, 4H), 1.27–1.19 (m, 3H), 1.07 (s, 3H), 1.04–0.96 (m, 2H), 0.94–0.87 (m, 6H), 0.84 (s, 3H), 0.83–0.77 (m, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.9, 173.7, 145.6, 145.3, 129.8, 129.7, 127.6, 127.5, 114.2, 114.2, 65.3, 64.1, 60.4, 60.3, 56.0, 55.9, 50.1, 47.0, 41.9, 41.9, 40.6, 40.5 (2C), 38.3, 38.2, 34.8, 33.3 (2C), 33.1, 21.7, 21.6, 20.4, 19.3, 18.6, 18.3, 18.2, 18.1, 15.7, 15.4, 14.3, 8.7, 8.3. HRMS (ESI, m/z) calcd for $\text{C}_{26}\text{H}_{40}\text{NO}_2$ ($\text{M}-\text{H}_2\text{O}+\text{H}$) $^+$: 398.3054, found: 398.3044.



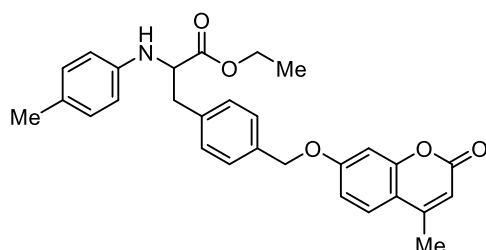
28: Yellow oil (16.7 mg, 53% yield); $R_f = 0.79$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.00 (d, $J = 7.8$ Hz, 2H), 6.61 (d, $J = 7.8$ Hz, 2H), 5.69 (s, 2H), 4.12 (m, 3H), 2.25 (s, 3H), 1.98 (m, 7H), 1.24 (t, $J = 7.1$ Hz, 3H), 1.07 (s, 1H), 1.04 (s, 2H), 0.95 (m, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.8, 145.4, 145.2, 129.8, 127.6, 127.6, 127.0, 126.9 (3C), 114.0 (2C), 62.5, 62.4, 60.5, 40.4, 39.6, 39.2, 39.1, 26.6 (2C), 26.4, 24.1, 23.7, 20.4, 20.3, 20.2 (2C), 20.0, 14.3 (2C). HRMS (ESI, m/z) calcd for $\text{C}_{20}\text{H}_{30}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 316.2271, found: 316.2270.



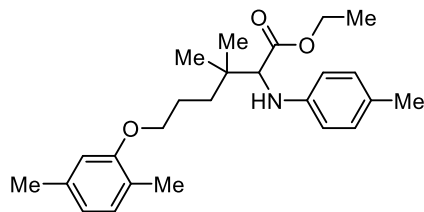
29: Yellow oil (17.7 mg, 50% yield); $R_f = 0.49$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 8.07 (m, 1H), 7.00 (m, 2H), 6.61 (m, 2H), 4.17 (m, 5H), 2.40–2.19 (m, 4H), 2.03–1.33 (m, 4H), 1.28–0.79 (m, 12H); ^{13}C NMR (101 MHz, CDCl_3) δ 174.0, 173.7, 173.5, 173.4, 161.2 (2C), 144.5, 144.3, 144.2, 129.9, 129.8 (2C), 128.0 (2C), 127.8 (2C), 114.3, 114.0, 113.9, 66.7, 66.4, 66.1, 66.0, 62.1, 62.0, 61.7, 61.4, 60.6 (2C), 60.5, 51.4, 50.9, 49.1, 48.5, 47.7, 47.6, 46.9, 46.4, 46.2, 46.1, 45.9, 44.7, 36.1, 35.8, 35.3, 31.9, 26.9 (2C), 26.4, 25.2, 24.4, 24.0, 23.9, 22.1, 20.7 (2C), 20.4, 19.8, 18.5, 18.2, 17.4, 16.9, 16.4, 14.3 (2C), 14.2, 14.1. HRMS (ESI, m/z) calcd for $\text{C}_{21}\text{H}_{32}\text{NO}_4$ ($\text{M}+\text{H}$) $^+$: 362.2326, found: 362.2329.



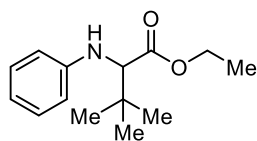
30: Yellow oil (33.4 mg, 74% yield); $R_f = 0.32$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 8.24 (m, 1H), 7.01 (m, 2H), 6.63 (m, 2H), 5.37–5.25 (m, 8H), 4.40–3.90 (m, 8H), 2.25 (s, 1H), 1.50–1.21 (m, 15H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.6, 171.5, 160.5, 160.1, 144.7, 143.9, 129.9, 129.8, 128.5, 128.2, 114.5, 114.1, 110.5, 110.1, 109.8, 109.5, 78.1, 77.7, 77.1, 76.6, 75.0, 74.9, 71.8, 71.5, 71.3, 71.0, 67.3, 67.0, 66.3, 66.0, 61.5 (2C), 61.4, 60.0, 56.3, 27.2, 27.0, 26.8, 26.6, 26.5, 25.4, 25.3, 20.4 (2C), 14.2. HRMS (ESI, m/z) calcd for $\text{C}_{23}\text{H}_{34}\text{NO}_8$ ($\text{M}+\text{H}$) $^+$: 452.2279, found: 452.2288.



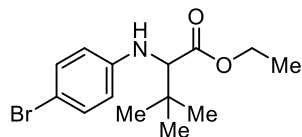
31: Yellow oil (33.4 mg, 74% yield); $R_f = 0.31$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.52 (d, $J = 8.8$ Hz, 1H), 7.37 (d, $J = 7.4$ Hz, 2H), 7.24 (d, $J = 7.5$ Hz, 2H), 7.00 (d, $J = 7.6$ Hz, 2H), 6.94 (d, $J = 8.8$ Hz, 1H), 6.90 (s, 1H), 6.56 (d, $J = 7.7$ Hz, 2H), 6.16 (s, 1H), 5.12 (s, 2H), 4.34 (t, $J = 6.1$ Hz, 1H), 4.14 (d, $J = 6.9$ Hz, 2H), 3.15 (t, $J = 6.1$ Hz, 2H), 2.42 (s, 3H), 2.25 (s, 3H), 1.19 (t, $J = 7.0$ Hz, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 173.2, 161.7, 161.3, 155.2, 152.6, 144.0, 136.8, 134.5, 129.9, 129.7, 128.0, 127.7, 125.6, 113.9, 113.8, 113.0, 112.1, 101.9, 70.2, 61.2, 58.1, 38.3, 20.4, 18.7, 14.2. HRMS (ESI, m/z) calcd for $\text{C}_{29}\text{H}_{30}\text{NO}_5$ ($\text{M}+\text{H}$) $^+$: 472.2119, found: 472.2115.



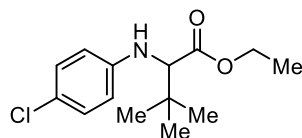
32: Yellow oil (34.2 mg, 86% yield); $R_f = 0.81$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.02 (m, 3H), 6.67 (m, 4H), 4.17 (q, $J = 7.1$ Hz, 2H), 3.95 (t, $J = 6.3$ Hz, 2H), 3.89 (s, 1H), 2.34 (s, 3H), 2.27 (s, 3H), 2.20 (s, 3H), 1.89 (m, $J = 5.9$ Hz, 2H), 1.62 (m, $J = 5.9$ Hz, 2H), 1.26 (t, $J = 7.1$ Hz, 3H), 1.11 (d, $J = 6.2$ Hz, 6H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.5, 157.0, 145.3, 136.5, 130.3, 129.8, 127.7, 123.7, 120.7, 114.2, 112.0, 68.3, 64.6, 60.6, 36.8, 36.1, 24.1, 24.0, 23.6, 21.4, 20.4, 15.8, 14.3. HRMS (ESI, m/z) calcd for $\text{C}_{25}\text{H}_{36}\text{NO}_3$ ($\text{M}+\text{H}$) $^+$: 398.2690, found: 398.2680.



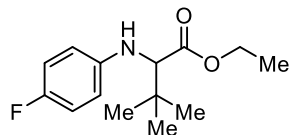
33: Yellow oil (15.9 mg, 68% yield); $R_f = 0.73$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.19 (t, $J = 7.5$ Hz, 2H), 6.75 (t, $J = 7.3$ Hz, 1H), 6.69 (d, $J = 7.9$ Hz, 2H), 4.17 (q, $J = 7.1$ Hz, 3H), 3.81 (s, 1H), 1.26 (t, $J = 7.2$ Hz, 3H), 1.10 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.4, 147.7, 129.3, 118.3, 113.8, 65.5, 60.6, 34.5, 26.8, 14.3. Spectra are consistent with reported literature⁷ values.



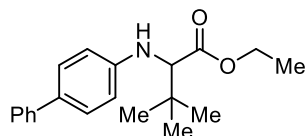
34: Yellow oil (16.6 mg, 53% yield); $R_f = 0.64$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.26 (d, $J = 7.9$ Hz, 2H), 6.56 (d, $J = 7.9$ Hz, 2H), 4.21 (s, 1H), 4.17 (q, $J = 7.0$ Hz, 2H), 3.74 (d, $J = 10.0$ Hz, 1H), 1.26 (t, $J = 7.2$ Hz, 3H), 1.08 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.0, 152.8, 146.7, 132.0, 115.4, 109.9, 65.6, 60.8, 34.5, 26.7, 14.3. HRMS (ESI, m/z) calcd for $\text{C}_{14}\text{H}_{21}\text{BrNO}_2$ ($\text{M}+\text{H}$) $^+$: 314.0750, found: 314.0750.



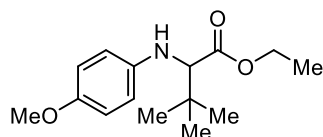
35: Yellow oil (16.3 mg, 61% yield); $R_f = 0.69$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.13 (d, $J = 8.0$ Hz, 2H), 6.61 (d, $J = 8.0$ Hz, 2H), 4.17 (q, $J = 6.9$ Hz, 3H), 3.74 (d, $J = 10.0$ Hz, 1H), 1.26 (t, $J = 7.5$ Hz, 3H), 1.08 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.1, 146.3, 129.1, 122.9, 115.0, 65.7, 60.7, 34.5, 26.8, 14.3. HRMS (ESI, m/z) calcd for $\text{C}_{14}\text{H}_{21}\text{ClNO}_2$ ($\text{M}+\text{H}$) $^+$: 270.1256, found: 270.1265.



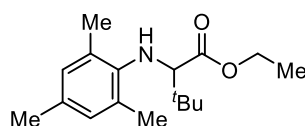
36: Yellow oil (27.2 mg, 54% yield); R_f = 0.72 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 6.89 (t, J = 8.5 Hz, 2H), 6.63 (dd, J = 4.41, 8.3 Hz, 2H), 4.17 (q, J = 4.3 Hz, 2H), 4.06 (d, J = 8.8 Hz, 1H), 3.70 (d, J = 9.1 Hz, 1H), 1.25 (t, J = 7.1 Hz, 3H), 1.09 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.3, 156.3 (d, J = 235.9 Hz), 144.0 (d, J = 1.6 Hz), 115.7 (d, J = 22.4 Hz), 115.7 (d, J = 22.4 Hz), 115.1 (d, J = 7.4 Hz), 66.6, 60.6, 34.3, 26.8, 14.3. ^{19}F NMR (377 MHz, CDCl_3) δ -126.5. HRMS (ESI, m/z) calcd for $\text{C}_{14}\text{H}_{21}\text{FNO}_2$ ($\text{M}+\text{H}$) $^+$: 254.1551, found: 254.1553.



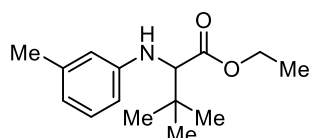
37: Yellow oil (31.0 mg, 50% yield); R_f = 0.58 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.58 (d, J = 7.6 Hz, 2H), 7.52–7.40 (m, 4H), 7.30 (t, J = 7.5 Hz, 1H), 6.79 (d, J = 8.0 Hz, 2H), 4.33 (d, J = 10.2 Hz, 1H), 4.22 (q, J = 7.1 Hz, 2H), 3.89 (d, J = 10.0 Hz, 1H), 1.30 (t, J = 7.1 Hz, 3H), 1.15 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.3, 147.2, 141.2, 131.3, 128.7, 128.0, 126.4, 126.2, 114.1, 65.5, 60.7, 34.6, 26.8, 14.4. HRMS (ESI, m/z) calcd for $\text{C}_{20}\text{H}_{26}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 312.1958, found: 312.1955.



38: Yellow oil (38.6 mg, 73% yield); R_f = 0.63 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 6.78 (d, J = 8.8 Hz, 2H), 6.66 (d, J = 8.7 Hz, 2H), 4.20–4.10 (m, 2H), 3.92 (d, J = 4.3 Hz, 1H), 3.76 (s, 3H), 3.69 (s, 1H), 1.24 (t, J = 7.1 Hz, 3H), 1.09 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.7, 152.8, 141.8, 115.6, 114.8, 67.1, 60.5, 55.7, 34.3, 26.8, 14.3. Spectra are consistent with reported literature²⁹ values.

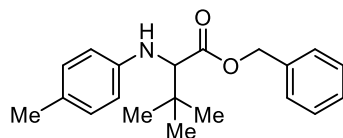


39: Yellow oil (16.3 mg, 55% yield); R_f = 0.78 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 6.77 (s, 2H), 4.00 (t, J = 6.7 Hz, 2H), 3.77 (s, 2H), 2.32 (s, 6H), 2.22 (s, 3H), 1.16 (s, 9H), 1.11 (t, J = 7.0 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 174.1, 141.4, 130.5, 129.7, 128.2, 67.6, 60.0, 34.4, 26.6, 20.5, 18.8, 14.1. HRMS (ESI, m/z) calcd for $\text{C}_{17}\text{H}_{28}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 278.2115, found: 278.2108.

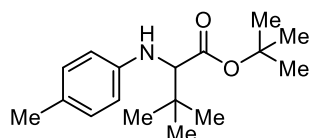


40: Yellow oil (13.5 mg, 27% yield); R_f = 0.68 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.08 (t, J = 7.6 Hz, 1H), 6.58 (d, J = 7.4 Hz, 1H), 6.54–6.46 (m, 2H), 4.18 (q, J = 7.1 Hz, 2H), 4.15 (s, 1H), 3.81 (d, J = 8.6 Hz, 1H), 2.29 (s, 3H), 1.27 (t, J = 7.2 Hz, 3H), 1.09 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.4, 147.7, 139.1, 129.2, 119.2, 114.7, 110.9, 65.5, 60.6, 34.5, 26.8, 21.6, 14.3. HRMS (ESI, m/z) calcd

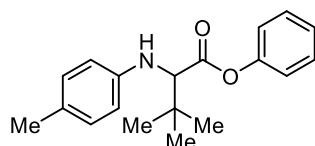
for $C_{15}H_{24}NO_2$ ($M+H$)⁺: 250.1802, found: 250.1798.



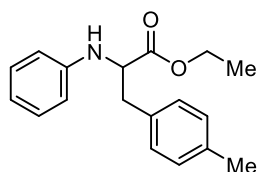
41: Yellow oil (14.3 mg, 46% yield); R_f = 0.69 (Petroleum ether/EtOAc, 5:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.40–7.29 (m, 5H), 7.19 (d, J = 8.0 Hz, 2H), 6.63 (d, J = 8.0 Hz, 2H), 5.51 (s, 2H), 4.10 (d, J = 7.5 Hz, 1H), 3.86 (d, J = 8.0 Hz, 1H), 2.28 (s, 3H), 1.10 (s, 9H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 173.5, 145.3, 135.6, 129.8, 128.5, 128.4, 128.3, 127.7, 114.2, 66.5, 66.1, 34.5, 26.8, 20.4. HRMS (ESI, m/z) calcd for $C_{20}H_{26}NO_2$ ($M+H$)⁺: 312.1958, found: 312.1956.



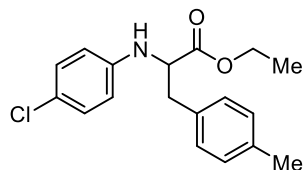
42: Yellow oil (30.6 mg, 55% yield); R_f = 0.66 (Petroleum ether/EtOAc, 5:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.00 (d, J = 7.8 Hz, 2H), 6.61 (d, J = 7.8 Hz, 2H), 4.06 (s, 1H), 3.65 (s, 1H), 2.26 (s, 3H), 1.49 (s, 9H), 1.09 (s, 9H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 172.7, 145.7, 129.7, 127.3, 114.1, 81.3, 66.5, 34.4, 28.1, 26.9, 20.4. HRMS (ESI, m/z) calcd for $C_{17}H_{28}NO_2$ ($M+H$)⁺: 278.2115, found: 278.2112.



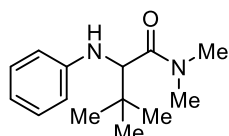
43: Yellow oil (11.3 mg, 38% yield); R_f = 0.52 (Petroleum ether/EtOAc, 5:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.37 (t, J = 7.6 Hz, 2H), 7.23 (t, J = 7.4 Hz, 1H), 7.05 (d, J = 7.8 Hz, 2H), 6.99 (d, J = 7.7 Hz, 2H), 6.72 (d, J = 7.9 Hz, 2H), 4.15 (s, 1H), 4.03 (s, 1H), 2.29 (s, 3H), 1.22 (s, 9H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 172.3, 150.5, 145.1, 129.9, 129.4, 128.0, 125.9, 121.5, 114.2, 66.1, 34.6, 26.9, 20.4. HRMS (ESI, m/z) calcd for $C_{19}H_{23}NO_2$ ($M+H$)⁺: 298.1802, found: 298.1799.



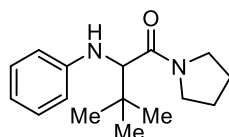
44: Yellow oil (16.4 mg, 58% yield); R_f = 0.61 (Petroleum ether/EtOAc, 5:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.21 (t, J = 7.5 Hz, 2H), 7.12 (q, J = 7.5 Hz, 4H), 6.78 (t, J = 7.2 Hz, 1H), 6.65 (d, J = 7.8 Hz, 2H), 4.37 (t, J = 6.1 Hz, 1H), 4.17 (q, J = 6.8 Hz, 2H), 3.14 (t, J = 6.7 Hz, 2H), 2.36 (s, 3H), 1.22 (t, J = 7.1 Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 173.2, 146.4, 136.6, 133.2, 129.4, 129.2 (2C), 118.4, 113.7, 61.1, 57.8, 38.2, 21.1, 14.2. HRMS (ESI, m/z) calcd for $C_{18}H_{22}NO_2$ ($M+H$)⁺: 284.1645, found: 284.1645.



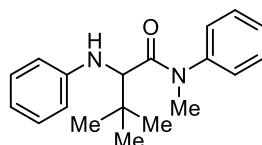
45: Yellow oil (18.1 mg, 57% yield); $R_f = 0.67$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (600 MHz, CDCl_3) δ 7.17–7.10 (m, 4H), 7.07 (d, $J = 8.0$ Hz, 2H), 6.55 (d, $J = 8.9$ Hz, 2H), 4.30 (t, $J = 6.2$ Hz, 1H), 4.17 (m, $J = 3.6$ Hz, 2H), 3.11 (m, $J = 7.5$ Hz, 2H), 2.35 (s, 3H), 1.22 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.0, 145.1, 136.7, 132.9, 129.3, 129.2 (2C), 122.9, 114.7, 61.3, 57.8, 38.0, 21.1, 14.2. HRMS (ESI, m/z) calcd for $\text{C}_{18}\text{H}_{21}\text{ClNO}_2$ ($\text{M}+\text{H}$) $^+$: 318.1256, found: 318.1247.



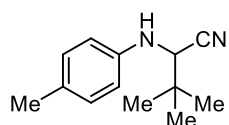
46: Yellow oil (20.4 mg, 44% yield); $R_f = 0.19$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.17 (t, $J = 7.6$ Hz, 2H), 6.75–6.65 (m, 3H), 4.54 (s, 1H), 4.22 (s, 1H), 3.13 (s, 3H), 2.95 (s, 3H), 1.09 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 173.1, 148.5, 129.3, 117.9, 114.0, 60.1, 38.2, 36.1, 35.6, 26.8. HRMS (ESI, m/z) calcd for $\text{C}_{14}\text{H}_{23}\text{N}_2\text{O}$ ($\text{M}+\text{H}$) $^+$: 235.1805, found: 235.1805.



47: Yellow oil (13.8 mg, 53% yield); $R_f = 0.20$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.16 (t, $J = 7.7$ Hz, 2H), 6.70 (m, $J = 6.5$ Hz, 3H), 4.54 (s, 1H), 4.00 (s, 1H), 3.52 (m, $J = 6.0$ Hz, 1H), 1.94 (m, 2H), 1.85 (t, 2H), 1.10 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 171.4, 148.1, 129.3, 118.0, 114.1, 63.0, 47.5, 45.7, 36.0, 26.8, 26.1, 24.1. HRMS (ESI, m/z) calcd for $\text{C}_{16}\text{H}_{25}\text{N}_2\text{O}$ ($\text{M}+\text{H}$) $^+$: 261.1962, found: 261.1960.

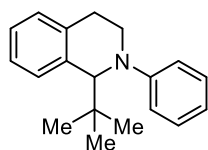


48: Yellow oil (16.3 mg, 55% yield); $R_f = 0.46$ (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.36 (s, 2H), 7.12 (t, $J = 7.08$ Hz, 2H), 6.96 (s, 2H), 6.71 (t, $J = 6.98$ Hz, 1H), 6.44 (d, $J = 7.56$ Hz, 2H), 3.26 (s, 3H), 0.95 (s, 9H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.9, 146.4, 143.3, 129.6, 129.2 (2C), 127.9, 118.9, 115.3, 62.5, 37.6, 36.3, 26.8. HRMS (ESI, m/z) calcd for $\text{C}_{19}\text{H}_{25}\text{N}_2\text{O}$ ($\text{M}+\text{H}$) $^+$: 297.1962, found: 297.1959.

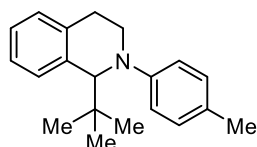


49: Yellow oil (17.8 mg, 88% yield); $R_f = 0.76$ (Petroleum ether/EtOAc, 5:1); ^1H NMR

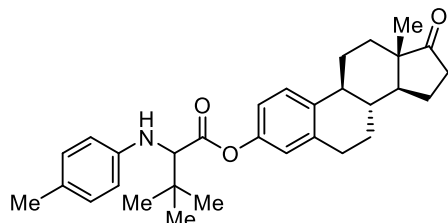
(600 MHz, CDCl₃) δ 7.09 (d, J = 8.1 Hz, 2H), 6.70 (d, J = 8.3 Hz, 2H), 3.92 (s, 1H), 2.30 (s, 3H), 1.21 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 143.2, 130.0, 129.5, 119.0, 114.6, 57.3, 34.7, 26.2, 20.5. HRMS (ESI, m/z) calcd for C₁₃H₁₉N₂ (M+H)⁺: 203.1543, found: 203.1541.



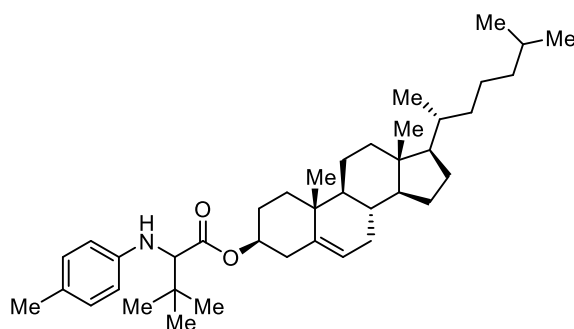
50: Yellow oil (13.5 mg, 51% yield); R_f = 0.85 (Petroleum ether/EtOAc, 5:1). Spectra are consistent with reported literature² values.



51: Yellow oil (15.6 mg, 56% yield); R_f = 0.83 (Petroleum ether/EtOAc, 5:1). Spectra are consistent with reported literature² values.

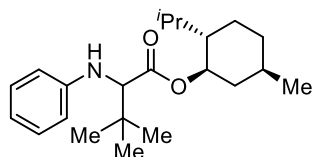


52: Yellow oil (6.1 mg, 13% yield); R_f = 0.36 (Petroleum ether/EtOAc, 5:1); ¹H NMR (400 MHz, CDCl₃) δ 7.26 (d, J = 8.7 Hz, 1H), 7.04 (d, J = 7.6 Hz, 2H), 6.76 (d, J = 8.4 Hz, 1H), 6.70 (d, J = 8.1 Hz, 3H), 4.01 (s, 1H), 2.89 (t, J = 3.6 Hz, 2H), 2.52 (q, J = 9.1 Hz, 1H), 2.45–2.36 (m, 1H), 2.28 (s, 3H), 2.19–1.93 (m, 2H), 1.70–1.41 (m, 10H), 1.20 (s, 9H), 0.92 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 220.9, 172.5, 148.3, 145.1, 138.0, 137.5, 129.9, 126.4, 121.4, 118.6 (2C), 114.3, 66.2, 50.4, 48.0, 44.1, 38.0, 35.9, 34.7, 31.5, 29.4, 26.9, 26.3, 25.7, 21.6, 20.4, 13.8. HRMS (ESI, m/z) calcd for C₃₁H₄₀NO₃ (M+H)⁺: 474.2930, found: 474.2996.

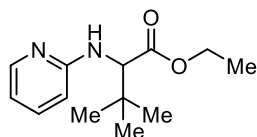


53: Pale solid (50.8 mg, 86% yield); m.p. 161–162 °C; R_f = 0.83 (Petroleum ether/EtOAc, 5:1); ¹H NMR (400 MHz, CDCl₃) δ 7.05–6.95 (m, 2H), 6.68–6.58 (m, 2H), 5.37 (s, 1H), 4.80–4.85 (m, 2H), 4.08 (s, 0.5H), 3.74 (s, 0.5H), 2.26 (s, 1H), 2.45–2.19 (m, 5H), 2.13–1.76 (m, 6H), 1.23–1.12 (m, 6H), 1.10 (s, 7H), 1.07–0.98 (m, 7H), 0.96–0.87 (m, 14H), 0.71 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 172.8, 145.5 (2C),

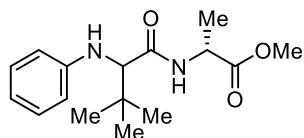
139.5 (2C), 129.7, 127.5, 122.8, 114.1 (2C), 74.4, 66.0 (2C), 56.7, 56.2, 50.0, 42.3, 39.7, 39.5, 38.2, 37.0, 36.9, 36.6, 36.2, 35.8, 34.4, 31.9 (2C), 28.2, 28.0, 26.9, 24.3, 23.9, 22.8, 22.6, 21.0, 20.4, 19.3, 18.7, 14.2, 11.9. HRMS (ESI, m/z) calcd for $C_{40}H_{64}NO_2$ ($M+H$)⁺: 590.4932, found: 590.4920.



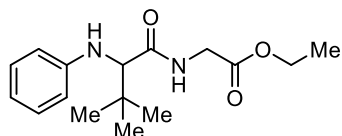
54: Yellow oil (12.8 mg, 37% yield); R_f = 0.85 (Petroleum ether/EtOAc, 5:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.16 (t, J = 7.7 Hz, 2H), 6.71 (m, J = 7.3 Hz, 3H), 4.66 (m, J = 5.4 Hz, 1H), 3.79 (s, 1H), 1.98–1.88 (m, 1H), 1.75–1.57 (m, 3H), 1.11 (s, 9H), 0.97–0.70 (m, 10H), 0.54 (q, J = 10.3 Hz, 3H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 173.2, 172.8, 147.7, 147.6, 129.2, 129.1, 118.6, 118.3, 114.5, 113.7, 75.1, 75.0, 66.1, 66.0, 46.9, 46.8, 40.9, 40.8, 34.3, 34.2, 33.9, 31.4, 26.9, 26.8, 25.9, 25.3, 22.9, 22.7, 22.0, 20.8, 15.7, 15.3. HRMS (ESI, m/z) calcd for $C_{22}H_{36}NO_2$ ($M+H$)⁺: 346.2741, found: 346.2738.



55: Yellow oil (7.1 mg, 30% yield); R_f = 0.48 (Petroleum ether/EtOAc, 3:1); 1H NMR (400 MHz, $CDCl_3$) δ 8.04 (d, J = 5.0 Hz, 1H), 7.50 (t, J = 7.7 Hz, 1H), 6.64 (t, J = 6.1 Hz, 1H), 6.52 (d, J = 8.4 Hz, 1H), 5.75 (s, 1H), 4.29 (d, J = 9.2 Hz, 1H), 4.20 (q, J = 7.0 Hz, 2H), 1.28 (t, J = 6.6 Hz, 3H), 1.11 (s, 9H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 172.4, 157.3, 138.9, 113.3, 108.5, 63.1, 60.9, 34.5, 26.8, 14.3. HRMS (ESI, m/z) calcd for $C_{13}H_{21}N_2O_2$ ($M+H$)⁺: 237.1598, found: 237.1591.

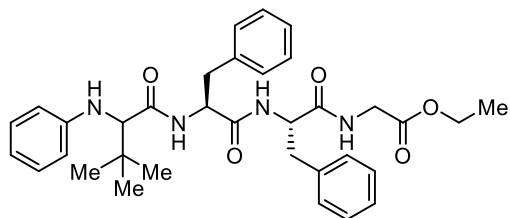


56: Pale solid (40.3 mg, 69% yield); m.p. 93–94 °C; R_f = 0.35 (Petroleum ether/EtOAc, 3:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.21 (t, J = 7.6 Hz, 1H), 7.02 (d, J = 6.2 Hz, 1H), 6.81 (t, J = 7.3 Hz, 1H), 6.65 (t, J = 9.6 Hz, 2H), 4.61 (m, J = 6.8 Hz, 1H), 4.08 (s, 1H), 3.74 (s, 1H), 3.66 (s, 2H), 3.43 (s, 1H), 1.37 (t, J = 14.2 Hz, 2H), 1.30 (t, J = 7.4 Hz, 2H), 1.14 (d, J = 6.7 Hz, 9H); ^{13}C NMR (101 MHz, $CDCl_3$) δ 173.0, 171.7, 147.4, 129.3, 129.3, 119.0, 114.0, 113.7, 68.4, 52.3, 47.9, 34.3, 27.2, 18.4. HRMS (ESI, m/z) calcd for $C_{16}H_{25}N_2O_3$ ($M+H$)⁺: 293.1860, found: 293.1860.

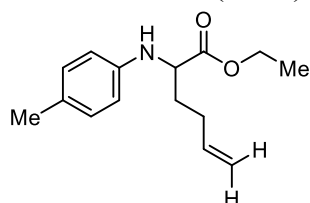


57: Pale solid (39.2 mg, 67% yield); m.p. 92–94 °C; R_f = 0.32 (Petroleum ether/EtOAc, 3:1); 1H NMR (400 MHz, $CDCl_3$) δ 7.21 (t, J = 7.7 Hz, 2H), 7.03 (s, 1H), 6.81 (t, J = 7.3 Hz, 1H), 6.66 (d, J = 7.9 Hz, 2H), 4.21 (t, J = 3.1 Hz, 1H), 4.18 (t, J = 7.1 Hz, 2H), 4.10 (s, 1H), 3.89 (d, J = 4.7 Hz, 1H), 3.84 (d, J = 4.7 Hz, 1H), 3.47 (d, J = 2.2 Hz, 1H),

1.5H), 3.72–3.59 (m, 0.5H), 3.49–3.36 (m, 1H), 3.31–2.87 (m, 1.5H), 1.28 (s, 3H), 1.13–0.83 (m, H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.6, 172.5, 171.0, 170.9, 169.3 (2C), 147.1, 146.9, 136.7, 136.0, 129.5, 129.4, 129.2 (2C), 128.7, 128.6, 127.0, 126.9, 119.1 (2C), 113.8, 113.6, 68.3 (2C), 61.6, 61.4, 53.8, 53.6, 41.4, 41.1, 37.6, 37.3, 34.2, 33.6, 27.2, 27.0, 14.2, 14.1. HRMS (ESI, m/z) calcd for $\text{C}_{25}\text{H}_{34}\text{N}_3\text{O}_4$ ($\text{M}+\text{H}$) $^+$: 440.2544, found: 440.2531.



61: Pale solid (50.8 mg, 86% yield); R_f = 0.31 (Petroleum ether/EtOAc, 3:1); m.p. 159–160 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.27–7.18 (m, 6H), 7.16–7.12 (m, 2H), 7.06–6.99 (m, 2H), 6.95–6.92 (m, 2H), 6.86–6.81 (m, 1H), 6.75–6.69 (m, 1H), 6.58 (d, J = 7.9 Hz, 1H), 6.32 (t, J = 5.1 Hz, 1H), 6.24 (d, J = 7.8 Hz, 1H), 4.72–4.56 (m, 2H), 4.24–4.18 (m, 2H), 4.04–3.96 (m, 1H), 3.89–3.80 (m, 1H), 3.45–3.33 (m, 1H), 3.05–2.98 (m, 2H), 2.97–2.81 (m, 2H), 1.29 (t, J = 7.1 Hz, 3H), 0.99 (s, 7H), 0.82 (s, 2H); ^{13}C NMR (101 MHz, CDCl_3) δ 172.9, 170.5, 170.4, 169.4, 136.3, 135.7, 130.3, 129.5, 129.3, 129.2, 129.0, 128.8, 128.7, 128.6, 128.5, 127.1, 127.0, 119.8, 113.9, 113.3, 68.2, 61.5, 54.5, 54.0, 41.3, 37.6, 37.5, 34.1, 27.2, 26.8, 14.2 (2C). HRMS (ESI, m/z) calcd for $\text{C}_{34}\text{H}_{43}\text{N}_4\text{O}_5$ ($\text{M}+\text{H}$) $^+$: 587.3228, found: 587.3217.



62: Yellow oil (9.3 mg, 19% yield); R_f = 0.77 (Petroleum ether/EtOAc, 5:1); ^1H NMR (400 MHz, CDCl_3) δ 7.00 (d, J = 8.1 Hz, 2H), 6.57 (d, J = 8.2 Hz, 2H), 5.91–5.78 (m, 1H), 5.08 (d, J = 17.4 Hz, 1H), 5.04 (d, J = 10.9 Hz, 1H), 4.19 (q, J = 7.11 Hz, 1H), 4.03 (br s, 1H), 2.25 (s, 3H), 2.00–1.89 (m, 1H), 1.89–1.79 (m, 1H), 1.27 (t, J = 7.0 Hz, 3H); ^{13}C NMR (101 MHz, CDCl_3) δ 174.3, 144.6, 137.3, 129.8, 127.6, 115.7, 113.8, 61.0, 56.5, 32.3, 29.7, 20.4. HRMS (ESI, m/z) calcd for $\text{C}_{15}\text{H}_{22}\text{NO}_2$ ($\text{M}+\text{H}$) $^+$: 248.1645, found: 248.1645.

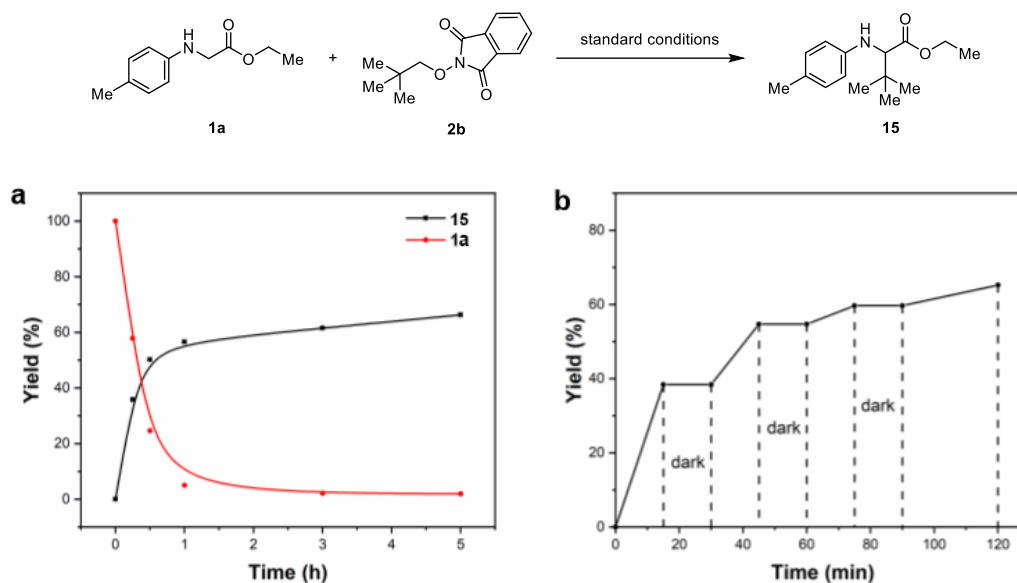
3. Supplementary Notes

3.1 Reaction Profile General

Time profile of the transformation with the light ON/OFF over time.

As can be seen in the time-course plot, the reaction proceeds rapidly within the first hour under the standard conditions and The yield of **15** continued to increase slowly over the next 4 hours (**Supplementary Figure 2**, a). From the profile of the reaction with the light ON/OFF over time, it was observed that the transformation progressed

smoothly under light, but no further conversion was observed when the light is turned off, which suggested that the transformation proceeds through a photoredox catalytic pathway rather than radical chain propagation (**Supplementary Figure 2, b**).

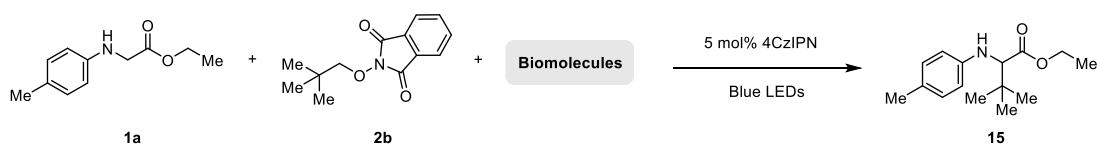



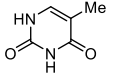
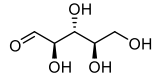
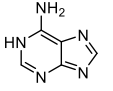
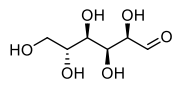
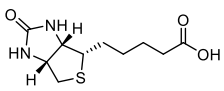
Standard conditions: 5 mol% 4CzIPN, 0.1 mmol **1a**, 2.0 equiv. **2b**, DMSO (0.03 M), 15 W Blue LEDs, 60 °C, 5 h

Supplementary Figure 2. Reaction profile. a Time course of the yield of **15** and consumption of **1a** during a reaction under the standard conditions. b Time course of the yield of **15** for during which the light was periodically turned on and off

3.2 Examining Functional Group Compatibility

In order to further prove the possibility of this reaction for biological application. We switched DMSO to a mixture H₂O/DMSO (1 : 4) solvent system and lowered the temperature to 37 °C. A broad of biomolecules were selected including nucleobase, amino acid, saccharide, biotin, protein and DNA. As shown in the following figure, all reactions worked and gave desired product in the presence of these biomolecules (entries 1–9).



Entry	Biomolecules	Yield ^a	Entry	Biomolecules	Yield ^a
1	none	63%	6	 L-Proline (1 equiv.)	29%
2	 Thymine (1 equiv.)	50%	7	 D-(-)-Ribose (1 equiv.)	86%
3	 Adenine (1 equiv.)	77%	8	 D-(+)-Glucose (1 equiv.)	68%
4	10 mg ml ⁻¹ bovine serum albumin	32%	9	 D-Biotin (1 equiv.)	60%
5	0.01 mg ml ⁻¹ pCMV-EGFP	72%			

Supplementary Figure 3. Examining functional group compatibility

^a Yields were determined by HPLC analysis.

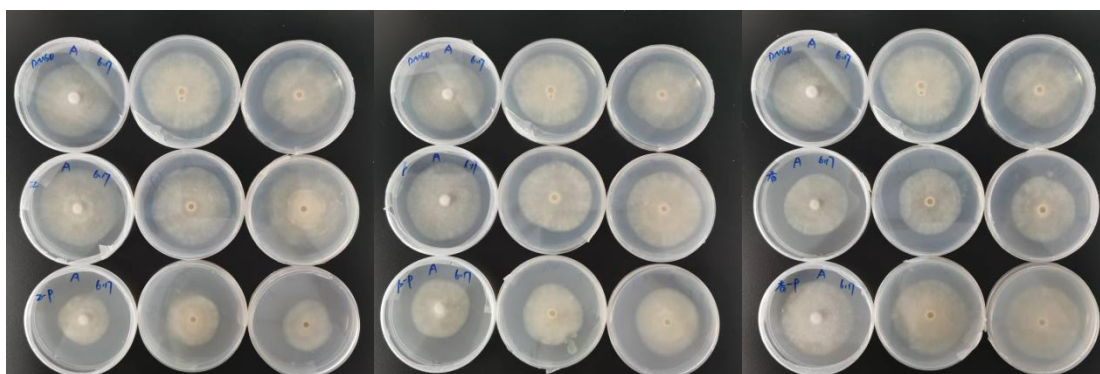
Experiment procedure: The glycine **1a** (0.2 mmol, 38.7 mg), *N*-alkoxyphthalimides **2b** (0.1 mmol, 23.3 mg), 4CzIPN (5 mol%, 0.005 mmol, 4.0 mg), and DMSO (2.4 mL), H₂O (0.6 mL), were added sequentially to a 4 mL clear-colored glass vial equipped with a magnetic stir bar. After bubbled with nitrogen gas for 5 minutes to remove oxygen, the vial was sealed and exposed to blue LEDs at 37 °C. The reaction mixture was monitored by TLC until the starting material *N*-alkoxyphthalimides were consumed. After the reaction was completed, 100 μL reaction mixture was diluted 100 times by sequential dilution and analyzed by HPLC with 30 μL.

3.3 In Vitro Antifungal Activities.

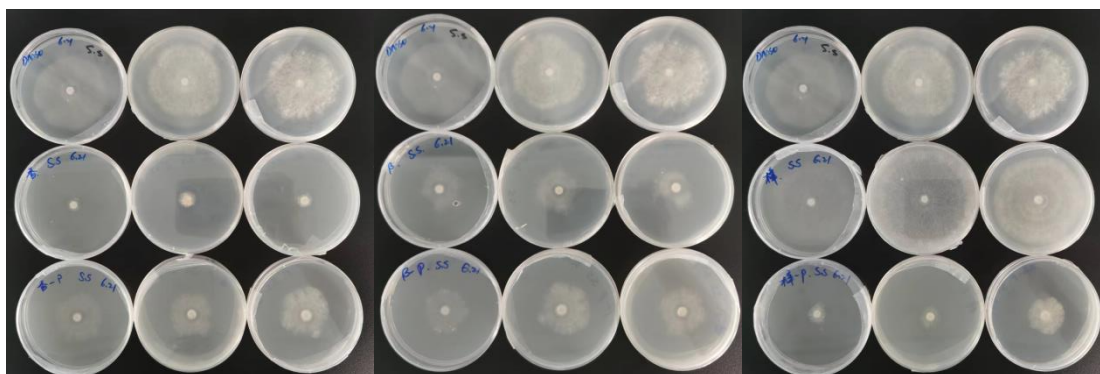
Each target compound was dissolved in DMSO to prepare the stock solution (10.0 g/L). The stock solution was added into the PDA medium, and the concentration of target compounds in the medium was 50.0 mg/L. Pure DMSO without the target compounds was utilized as the blank control, and boscalid was coassayed as the reference compound. Fresh dishes with a diameter of 5 mm were taken from the edge of the PDA-cultured fungi colonies and inoculated on the above three PDA media. Each treatment was tested for three replicates, and the antifungal effect was averaged. The relative

inhibitory rate I (%) of all the tested compounds was calculated through the equation: $I (\%) = [(C - T)/(C - 5)] \times 100$. In this equation, I is the inhibitory rate and C and T are the colony diameter of the blank control (mm) and treatment (mm), respectively.

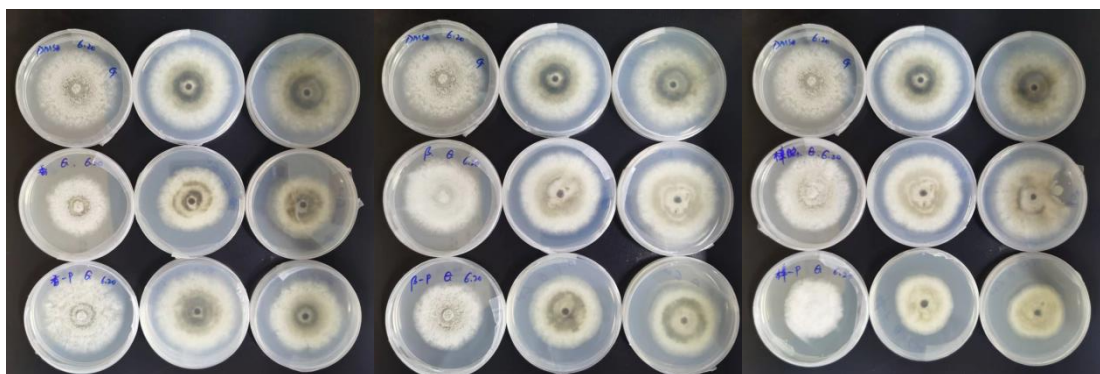
Mycelia growth of six crop pathogenic fungi after treating with the target compounds on PDA medium as illustrated in the figures (**Supplementary Figure 4. PDA medium**) below.



Supplementary Figure 4-1. PDA medium. *B. cinerea*



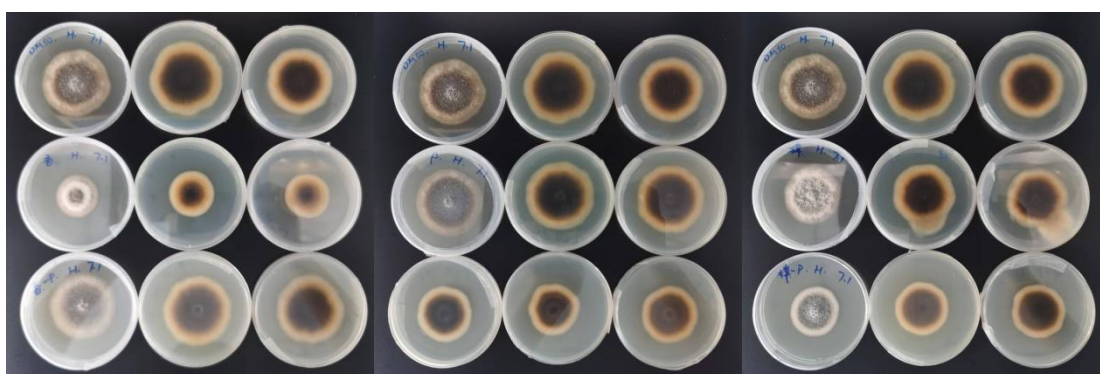
Supplementary Figure 4-2. PDA medium. *S. sclerotiorum*



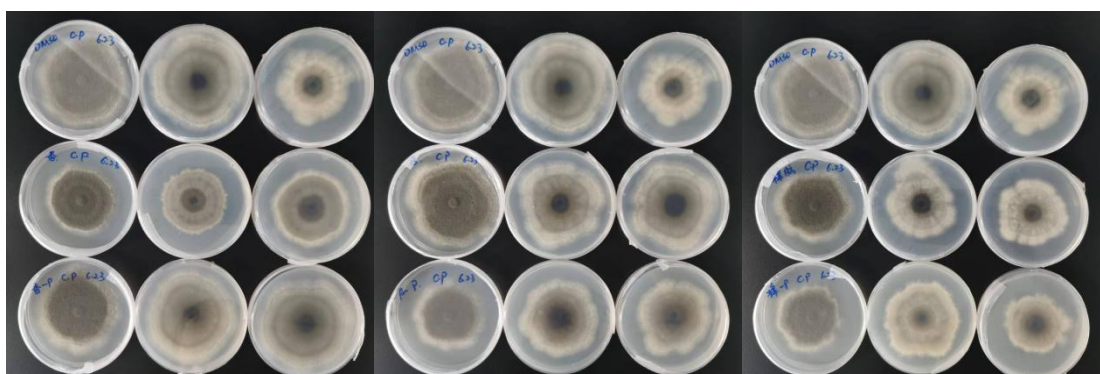
Supplementary Figure 4-3. PDA medium. *C. orbiculare*



Supplementary Figure 4-4. PDA medium. *T. cucumeris*



Supplementary Figure 4-5. PDA medium. *Alternariamali*



Supplementary Figure 4-6. PDA medium. *C. paradoxa*

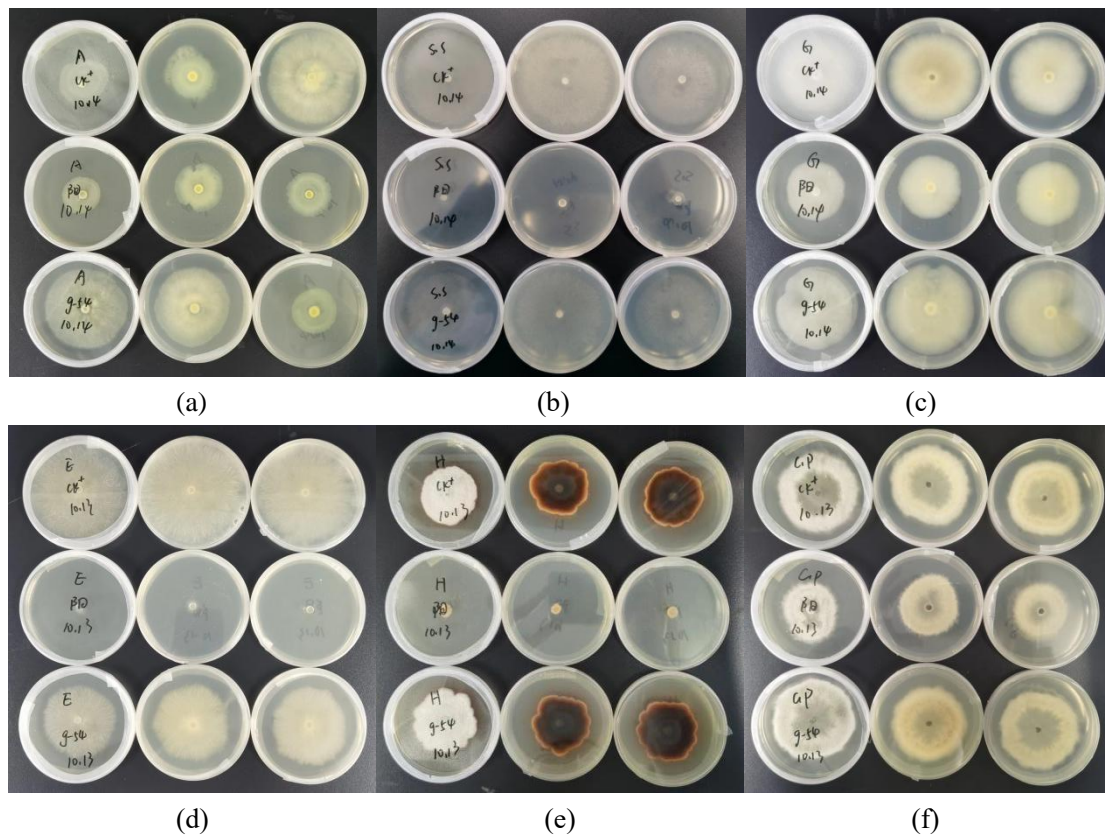
Supplementary Table 1. In vitro antifungal activities.

In Vitro Antifungal Activities of the target compound 54 at 50.0 mg/L

Compd	Inhibition rate (%) ^a					
	<i>B. cinerea</i>	<i>S. sclerotiorum</i>	<i>C. orbiculare</i>	<i>T. cucumeris</i>	<i>Alternariamali</i>	<i>C. paradoxa</i>
boscalid	36.8 ± 5.4	90.0 ± 0.7	27.7 ± 3.3	91.0 ± 1.7	89.7 ± 2.5	29.0 ± 0
54	-55.2 ± 6.2	8.3 ± 7.5	-0.6 ± 2.8	23.7 ± 1.6	-2.9 ± 2.7	-0.9 ± 2.1

^a Values are the mean ± standard deviation of three replicates.

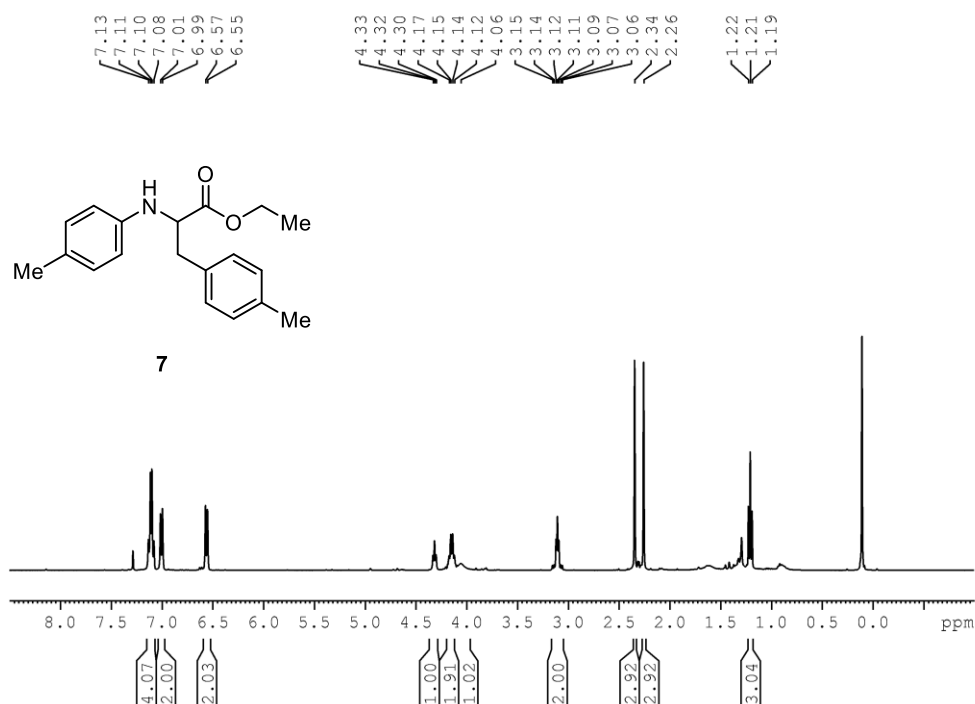
Mycelia growth of six crop pathogenic fungi after treating with the target compound **54** on PDA medium as illustrated in the figures (Supplementary Figure 5. PDA medium) below.



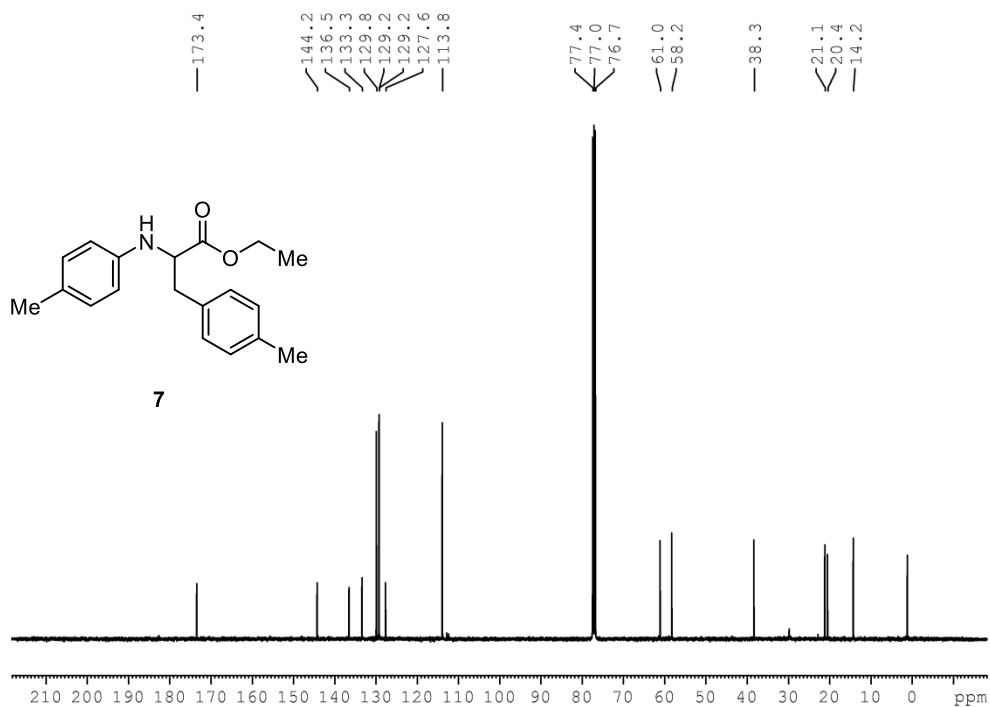
Supplementary Figure 5. PDA medium. (a) *B. cinerea*; (b) *S. sclerotiorum*; (c) *C. orbiculare*; (d) *T. cucumeris*; (e) *Alternaria mali*; (f) *C. paradoxa*.

4. Supplementary Figures

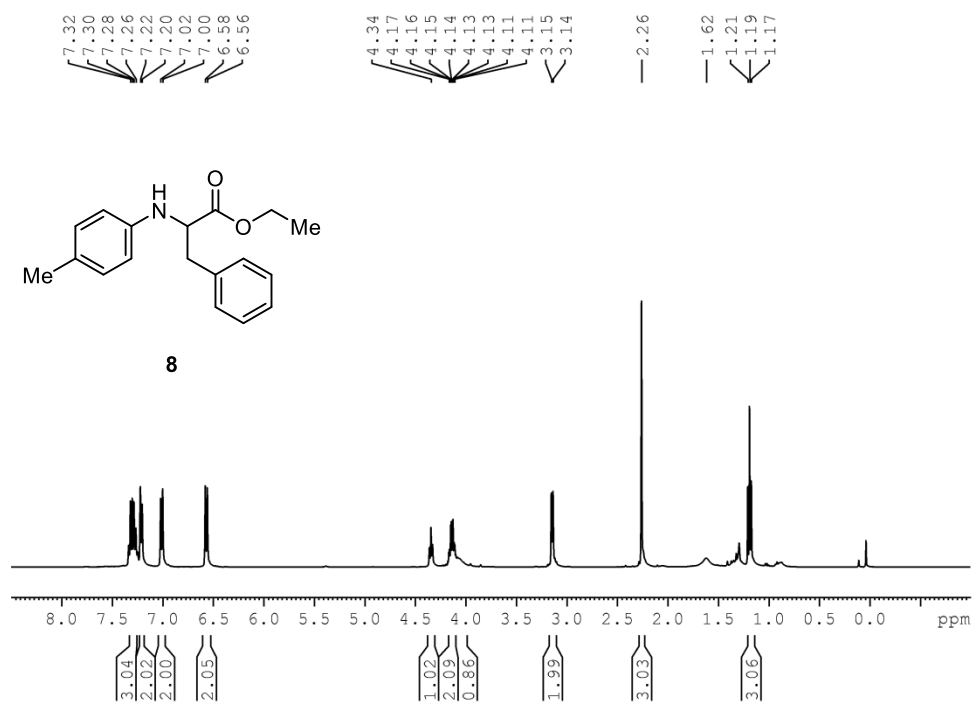
4.1 NMR Spectra



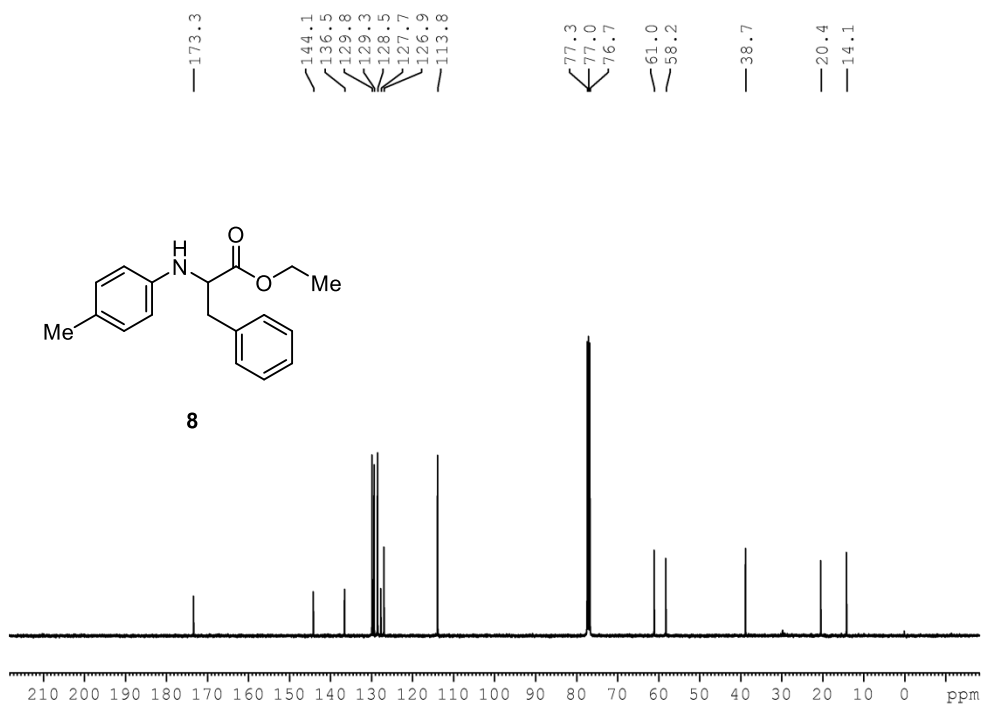
Supplementary Figure 6. ¹H NMR spectrum of compound 7



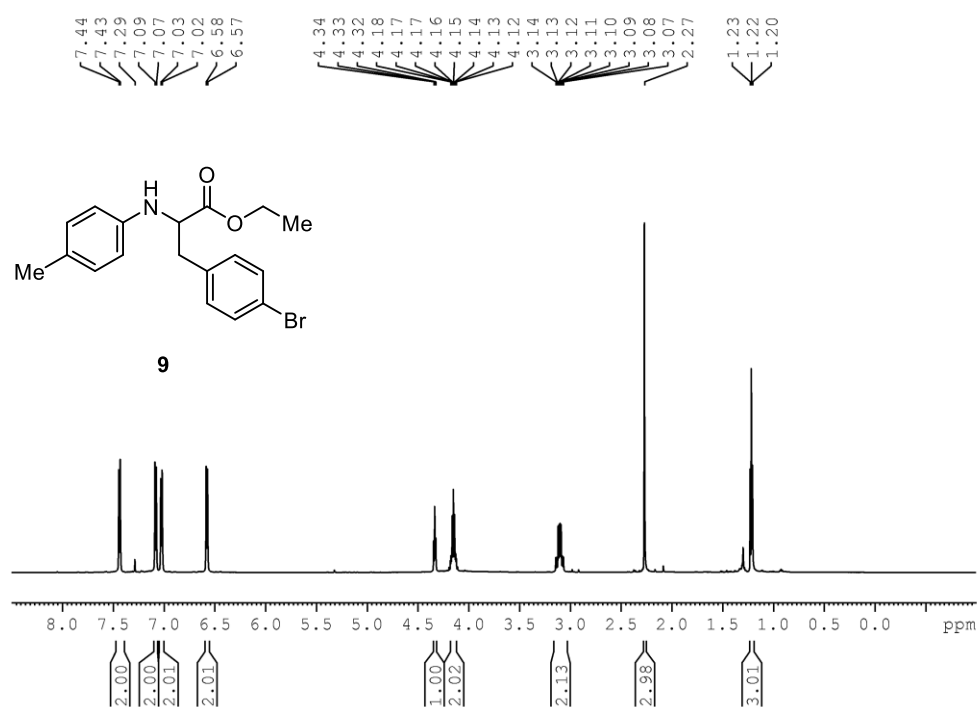
Supplementary Figure 7. ¹³C NMR spectrum of compound 7



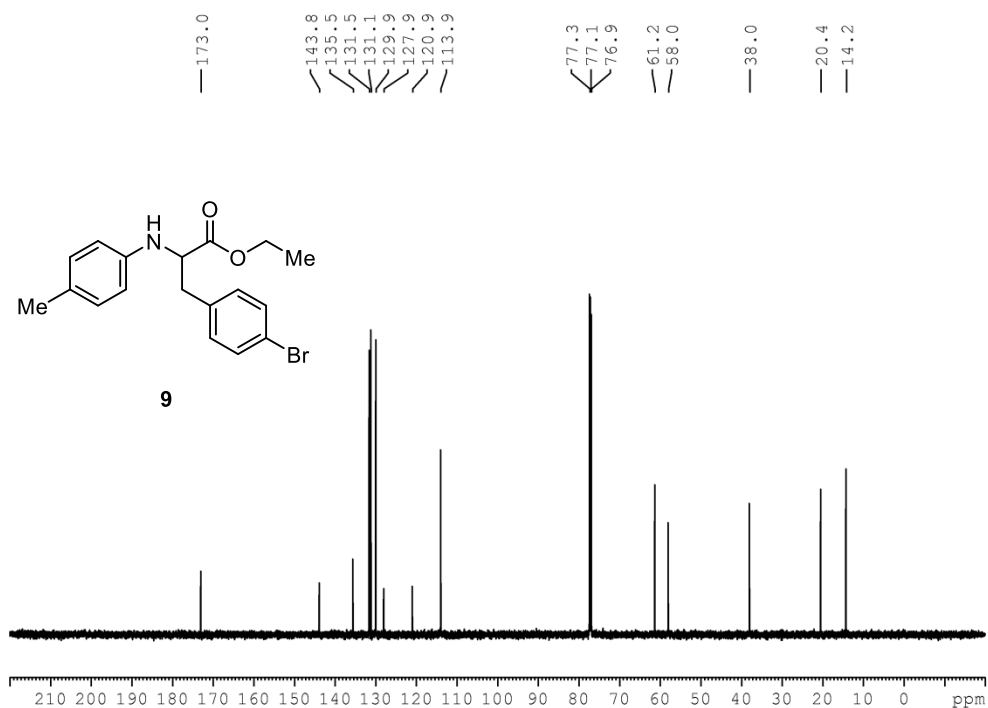
Supplementary Figure 8. ¹H NMR spectrum of compound 8



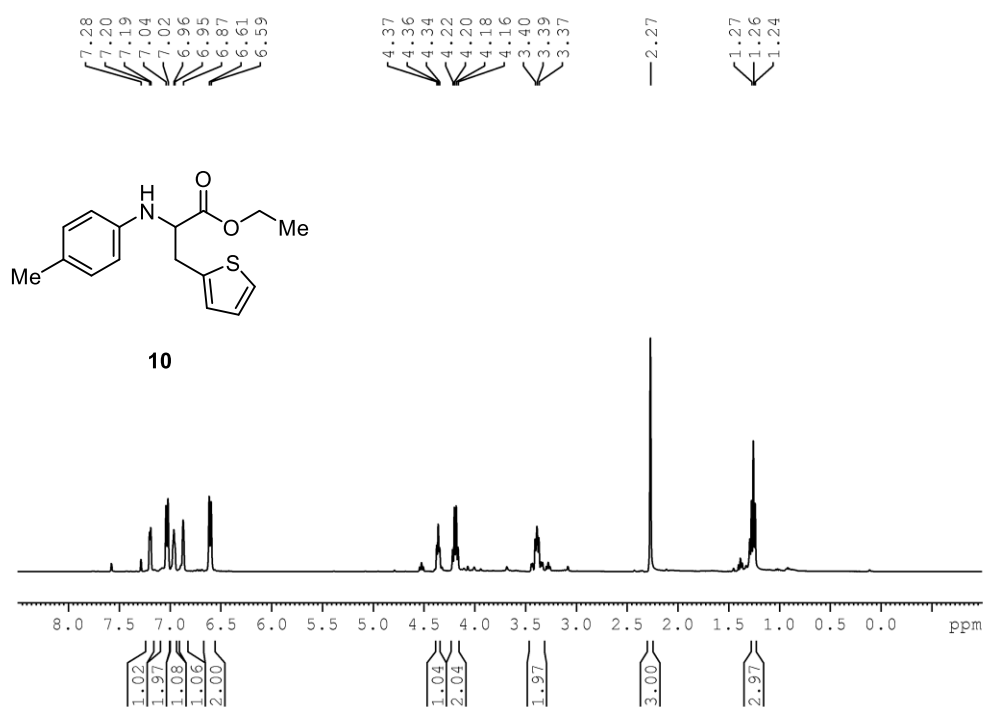
Supplementary Figure 9. ¹³C NMR spectrum of compound 8



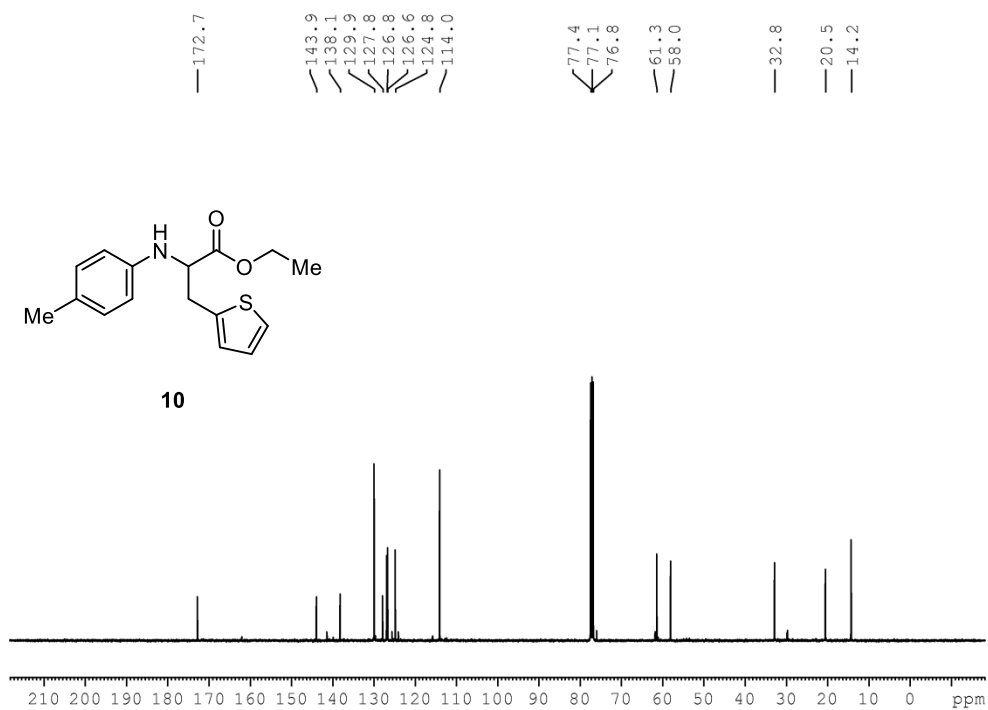
Supplementary Figure 10. ¹H NMR spectrum of compound 9



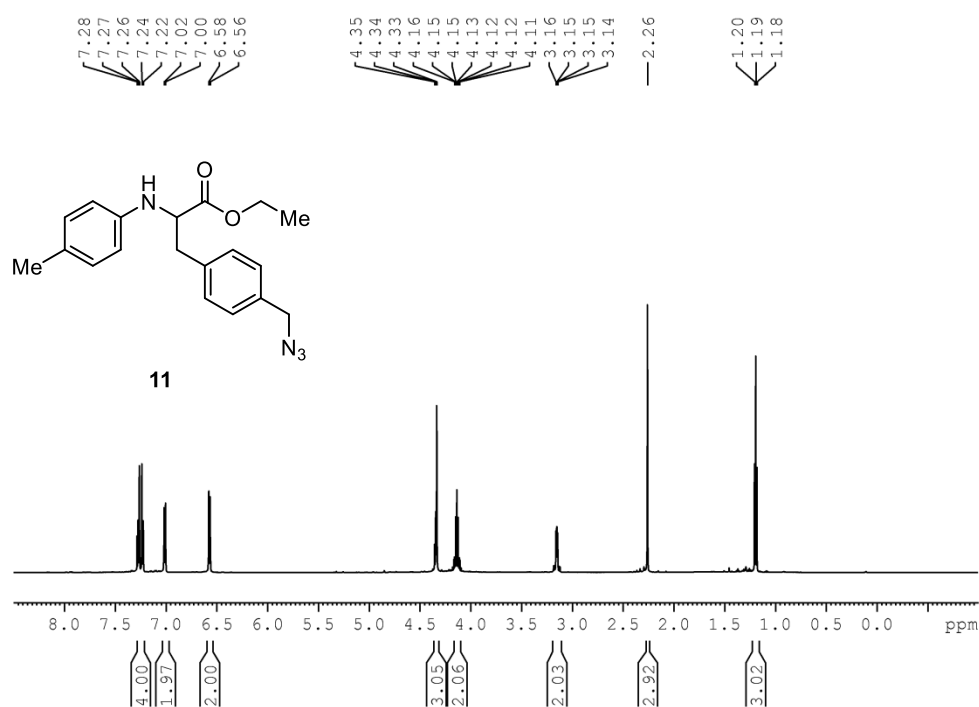
Supplementary Figure 11. ¹³C NMR spectrum of compound 9



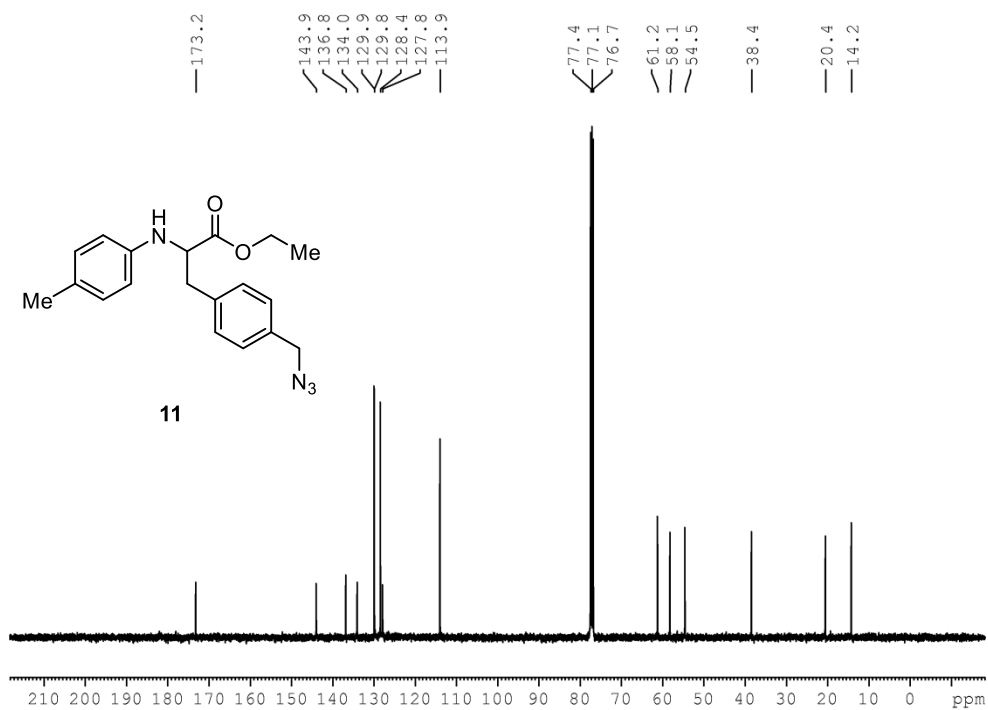
Supplementary Figure 12. ¹H NMR spectrum of compound 10



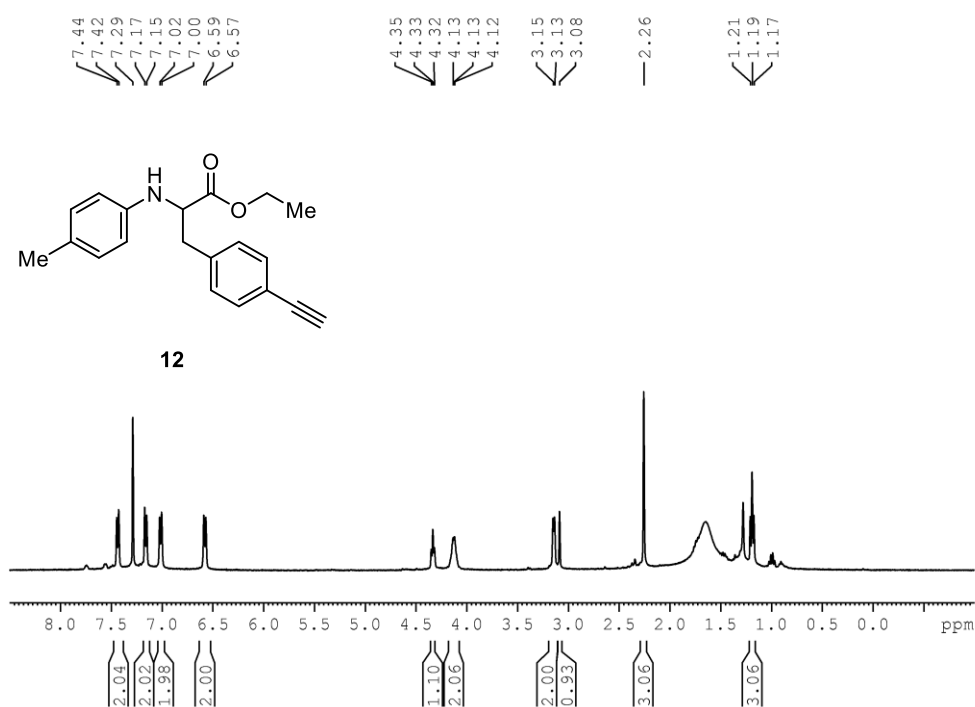
Supplementary Figure 13. ¹³C NMR spectrum of compound 10



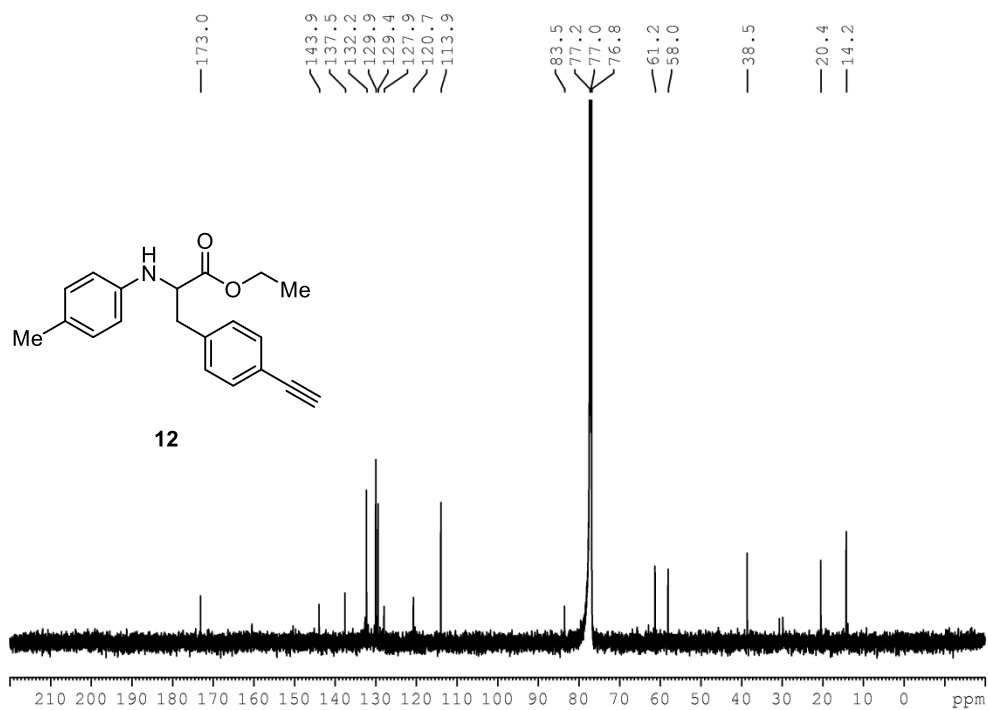
Supplementary Figure 14. ¹H NMR spectrum of compound 11



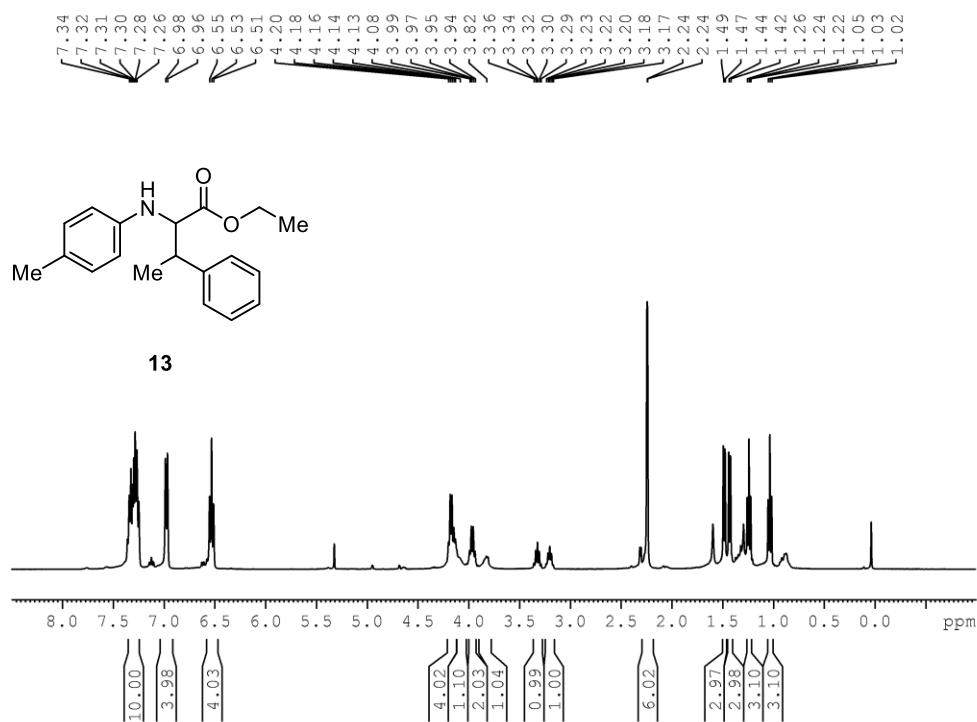
Supplementary Figure 15. ¹³C NMR spectrum of compound 11



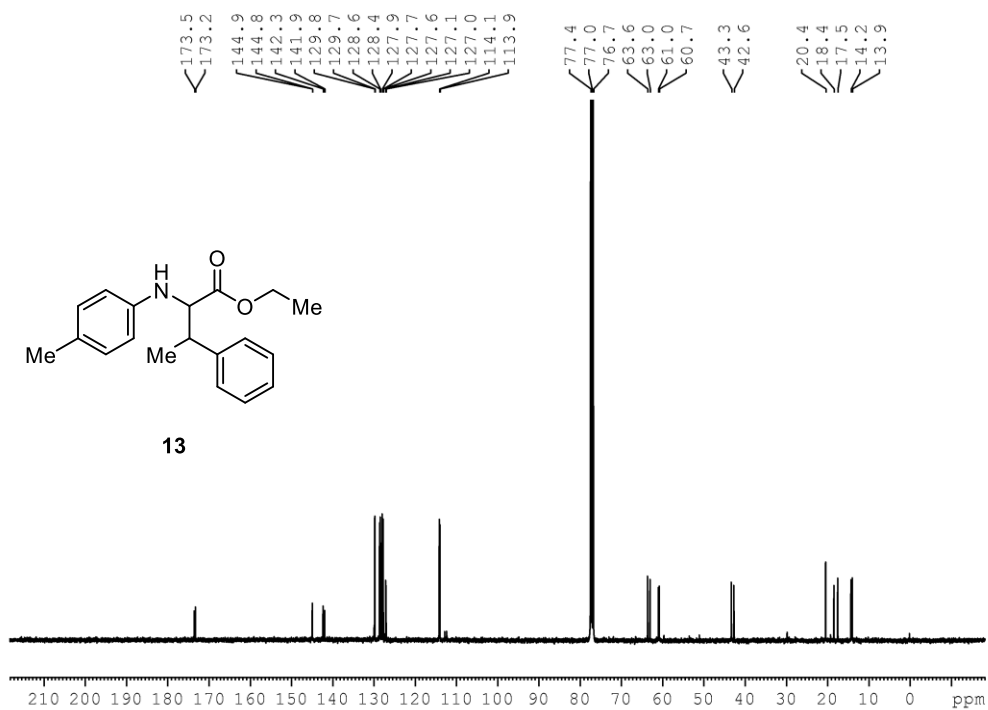
Supplementary Figure 16. ^1H NMR spectrum of compound 12



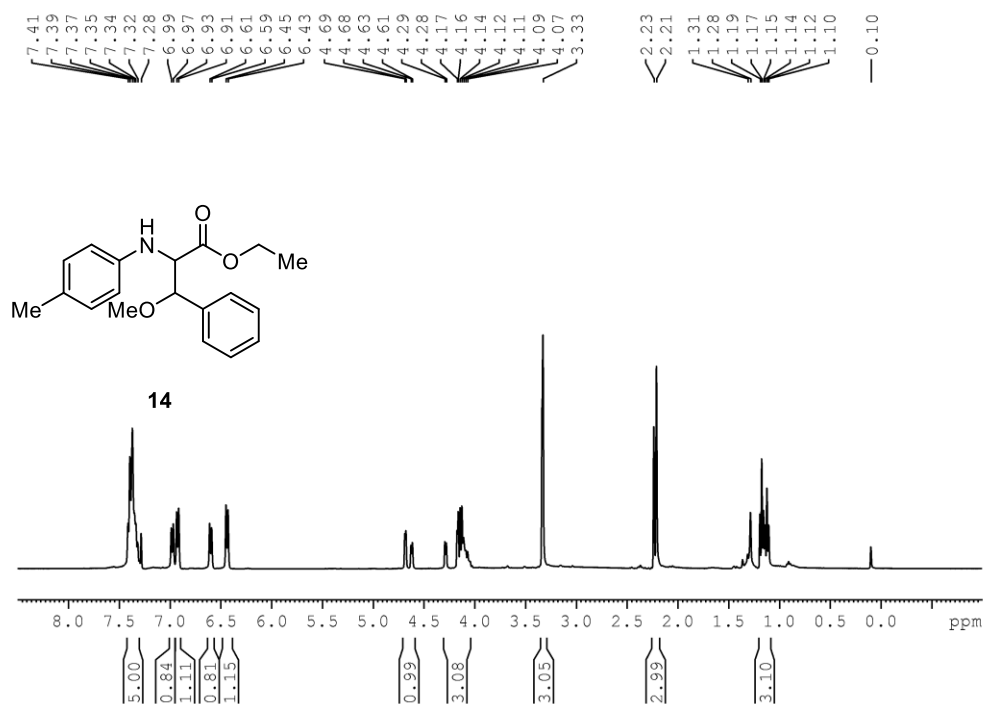
Supplementary Figure 17. ^{13}C NMR spectrum of compound 12



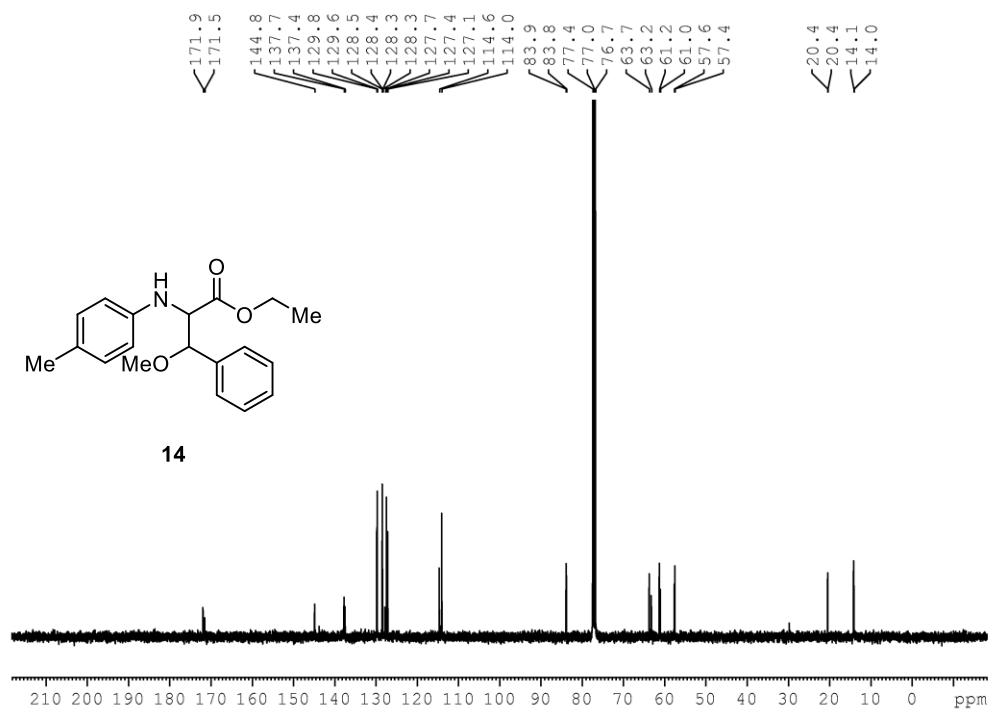
Supplementary Figure 18. ¹H NMR spectrum of compound 13



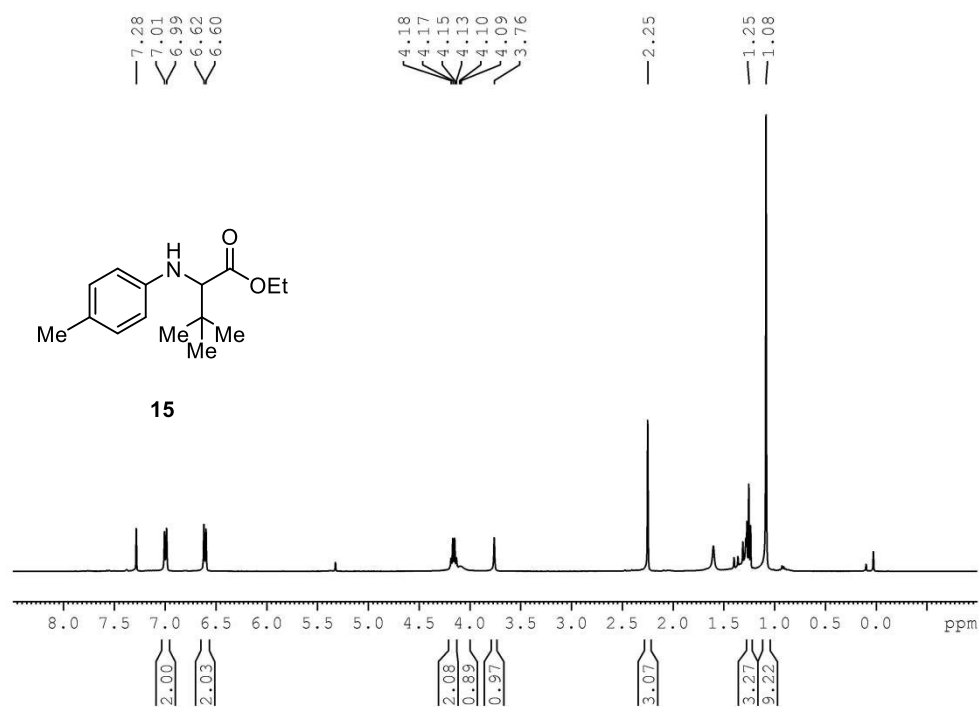
Supplementary Figure 19. ¹³C NMR spectrum of compound 13



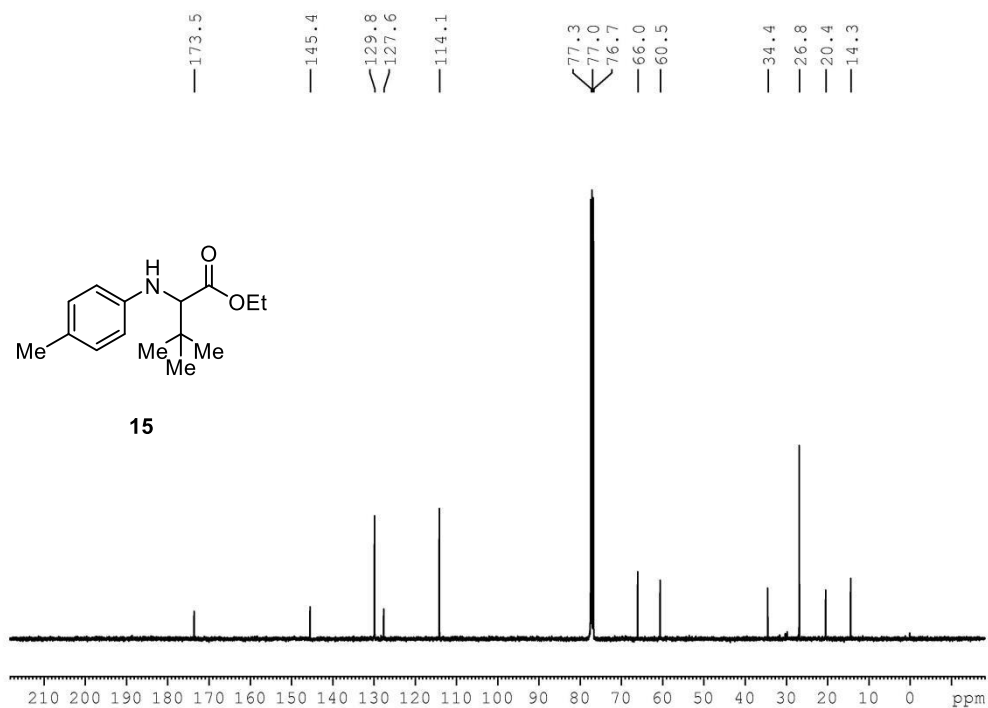
Supplementary Figure 20. ^1H NMR spectrum of compound 14



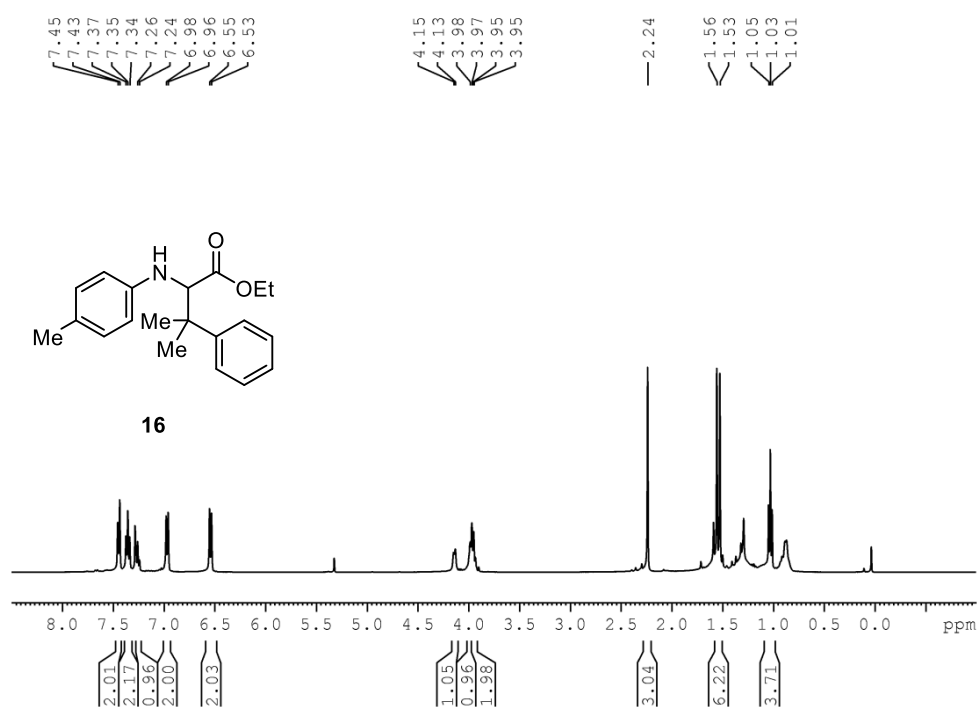
Supplementary Figure 21. ^{13}C NMR spectrum of compound 14



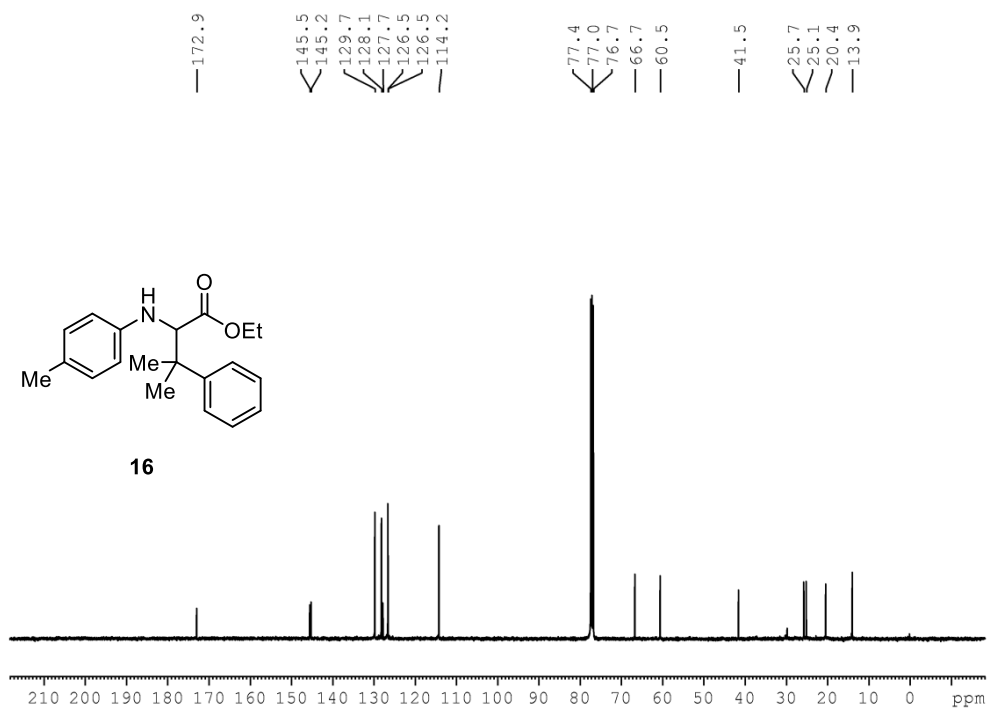
Supplementary Figure 22. ¹H NMR spectrum of compound 15



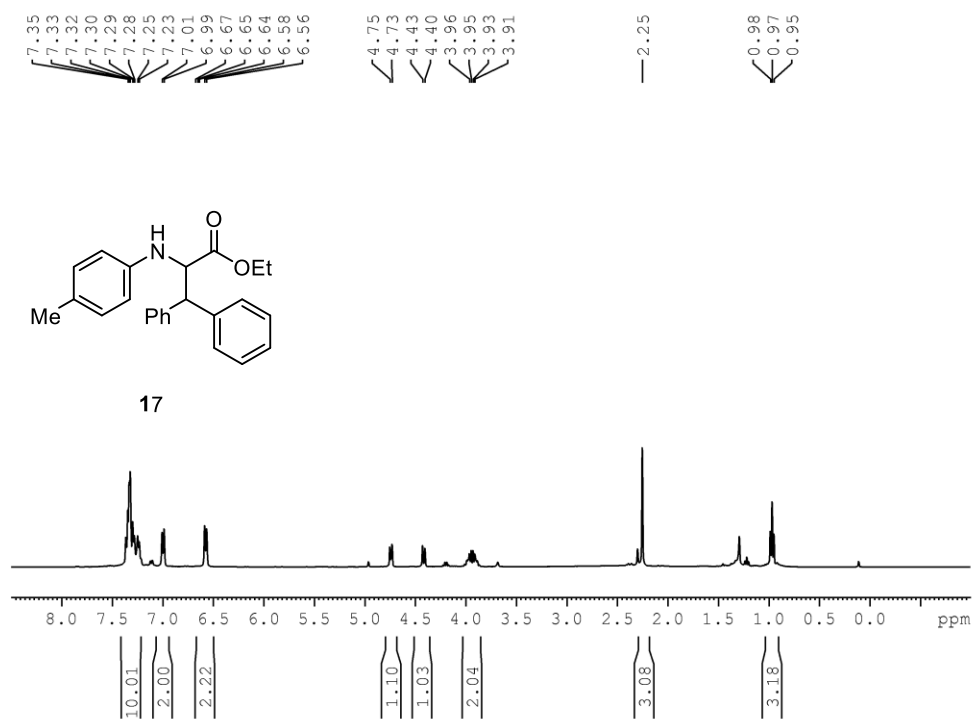
Supplementary Figure 23. ¹³C NMR spectrum of compound 15



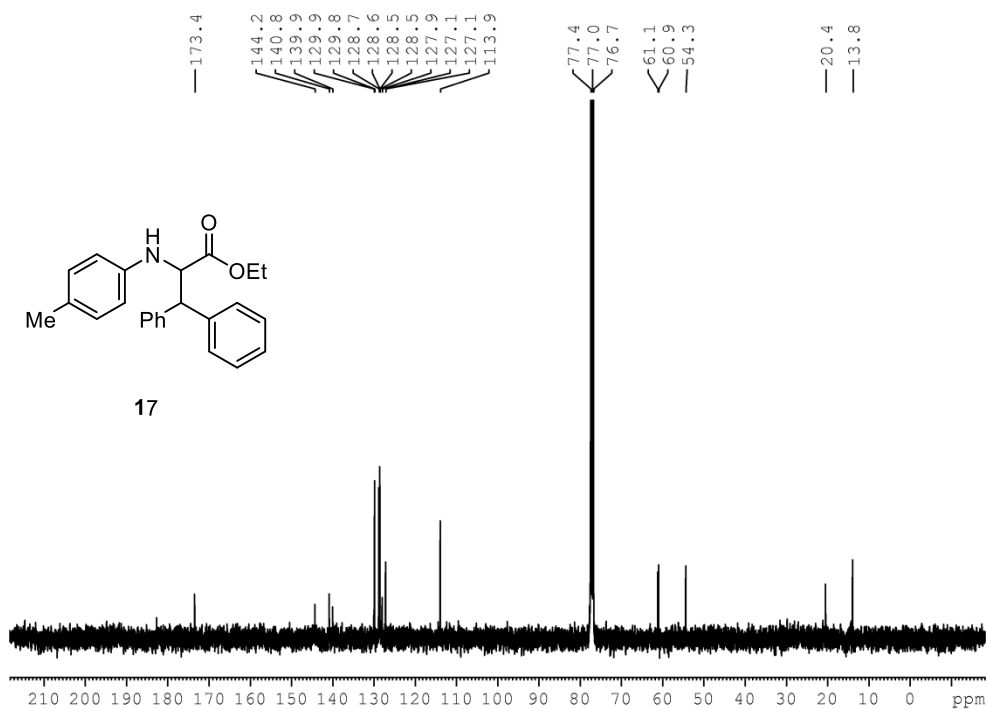
Supplementary Figure 24. ^1H NMR spectrum of compound 16



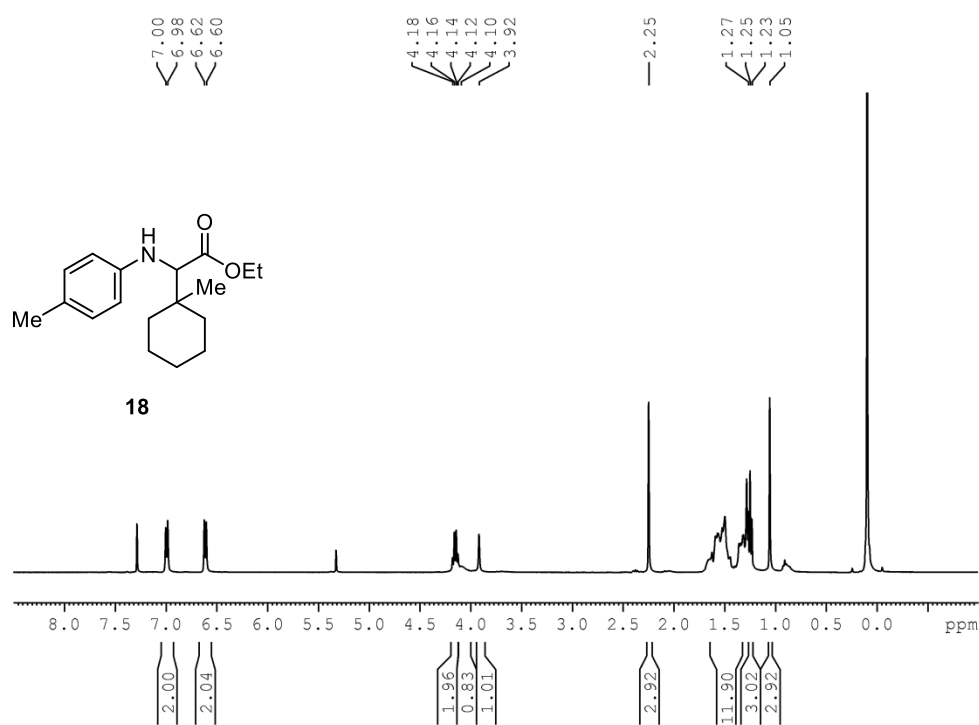
Supplementary Figure 25. ^{13}C NMR spectrum of compound 16



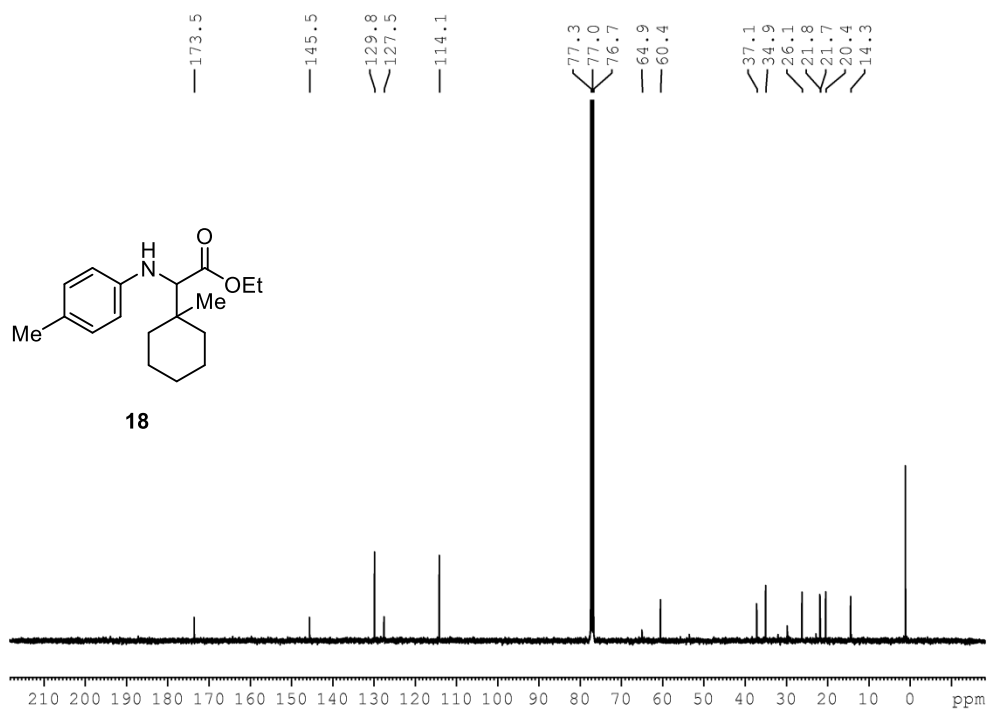
Supplementary Figure 26. ¹H NMR spectrum of compound 17



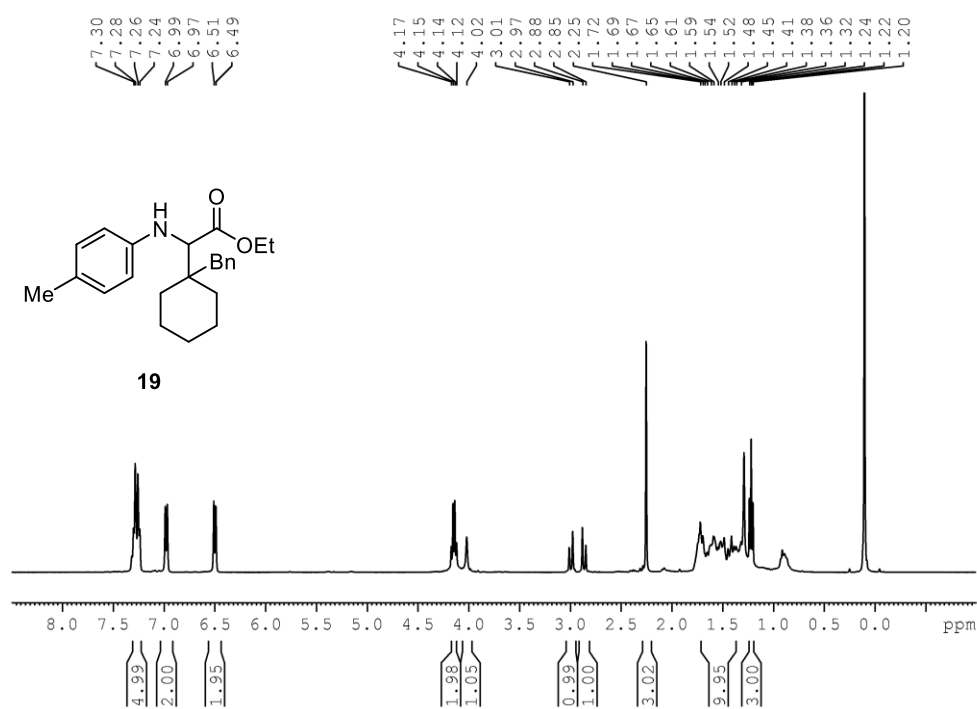
Supplementary Figure 27. ¹³C NMR spectrum of compound 17



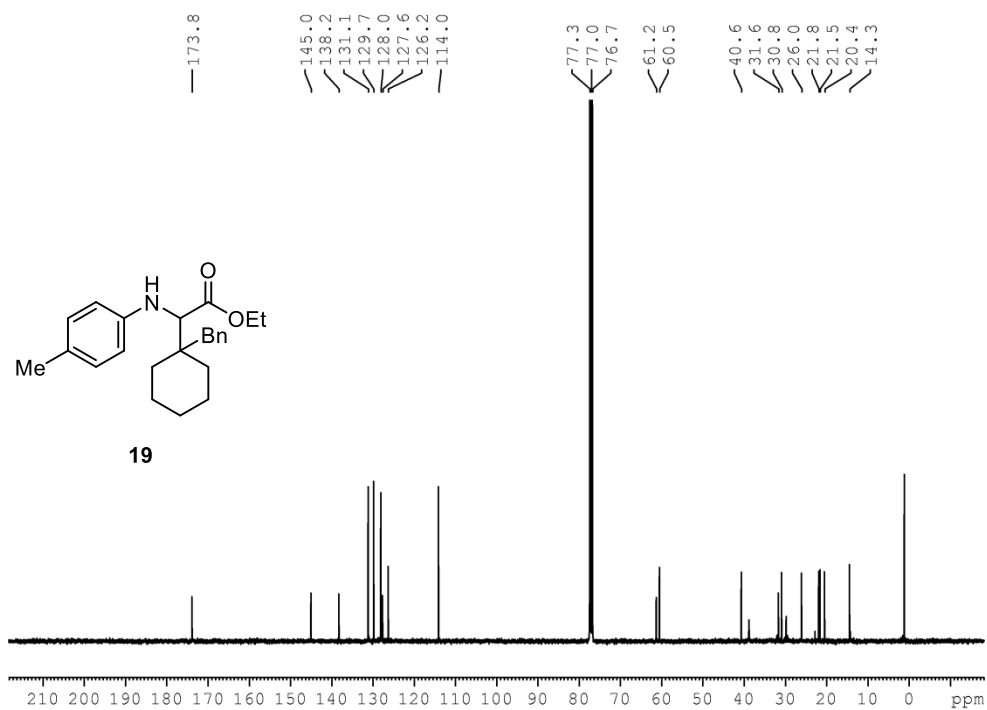
Supplementary Figure 28. ¹H NMR spectrum of compound **18**



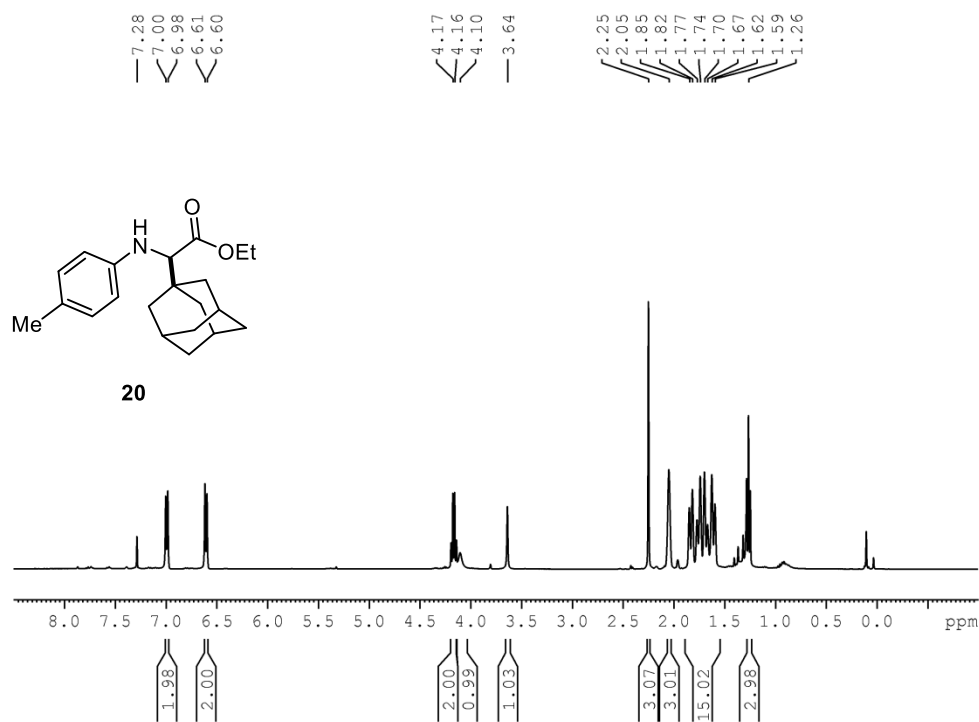
Supplementary Figure 29. ¹³C NMR spectrum of compound **18**



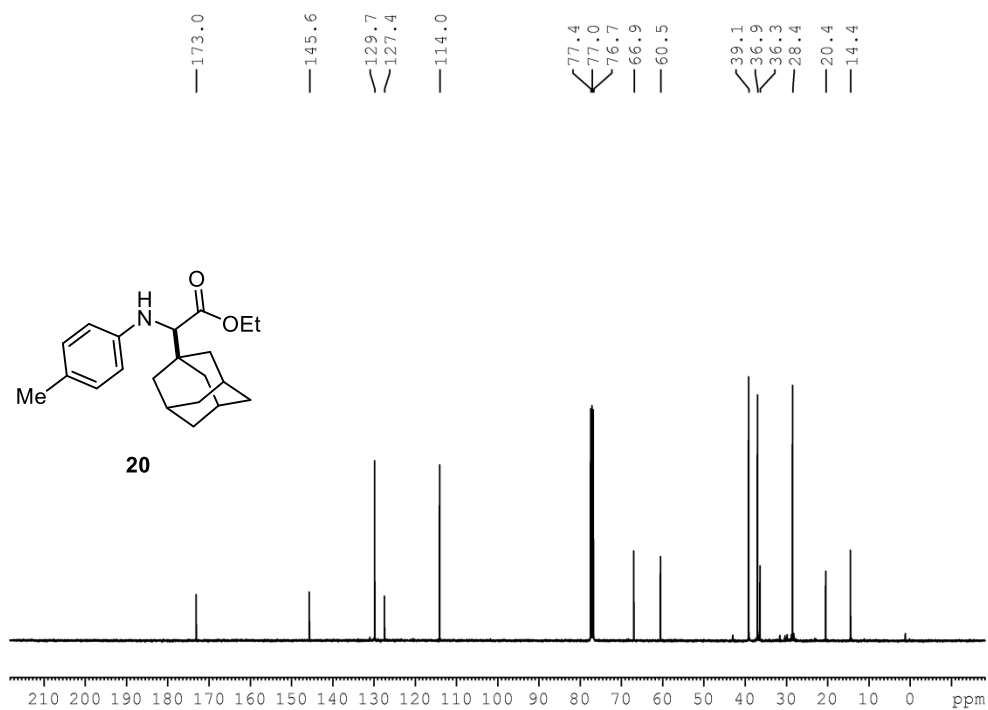
Supplementary Figure 30. ^1H NMR spectrum of compound 19



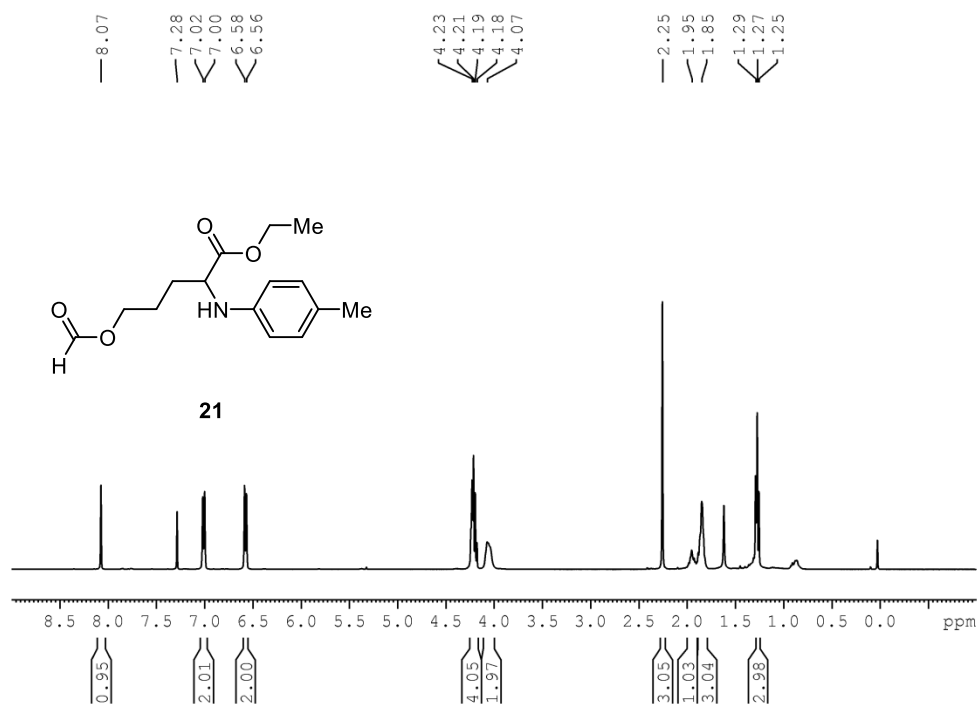
Supplementary Figure 31. ^{13}C NMR spectrum of compound 19



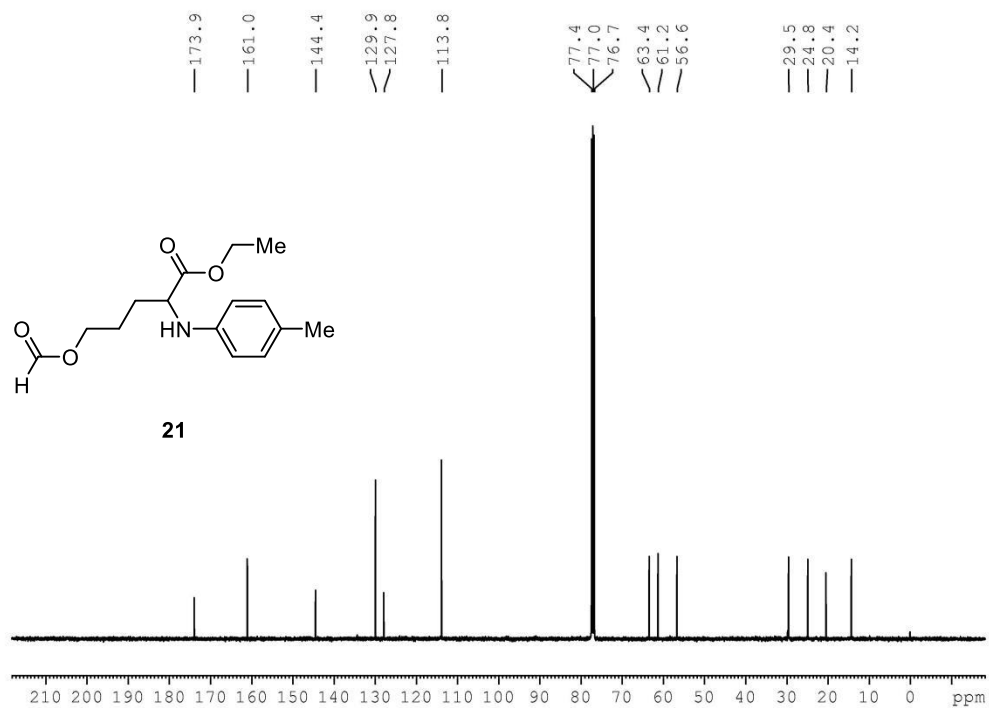
Supplementary Figure 32. ^1H NMR spectrum of compound **20**



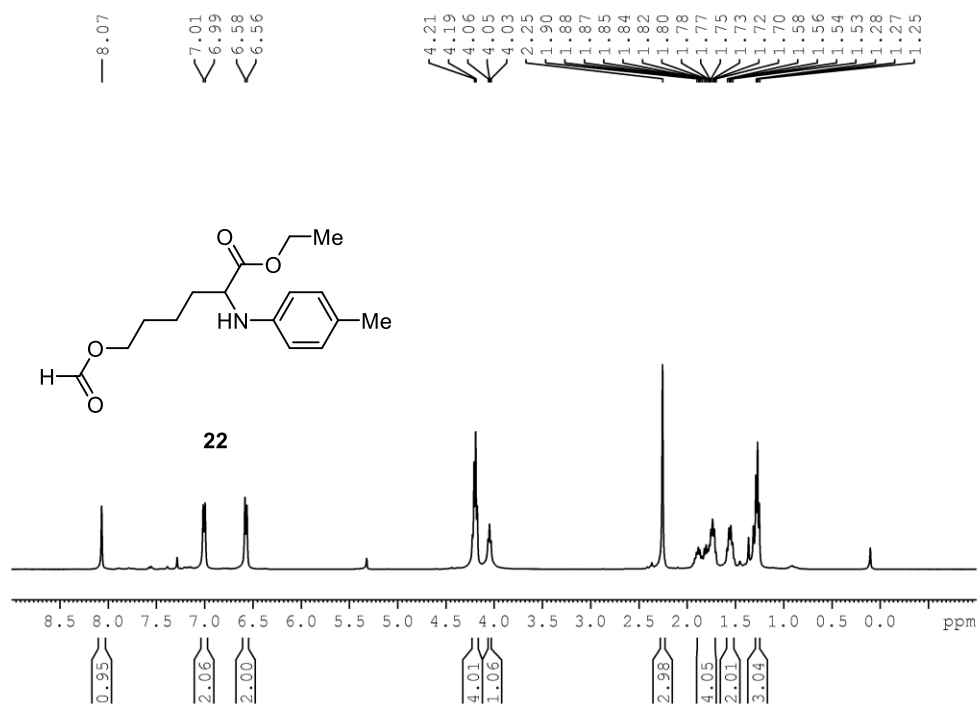
Supplementary Figure 33. ^{13}C NMR spectrum of compound **20**



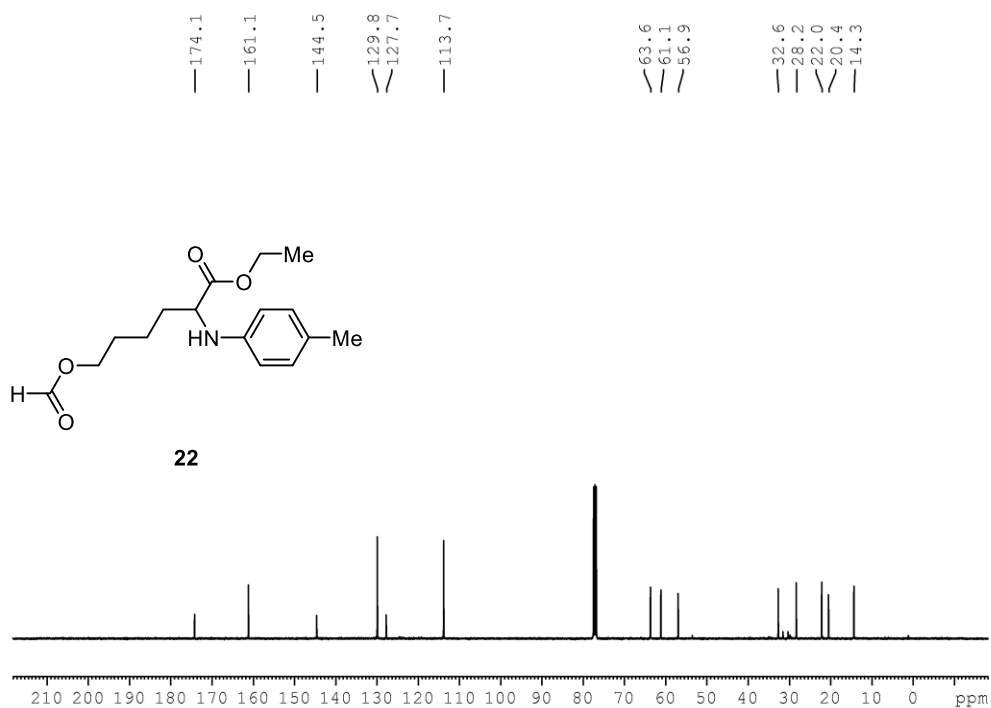
Supplementary Figure 34. ¹H NMR spectrum of compound 21



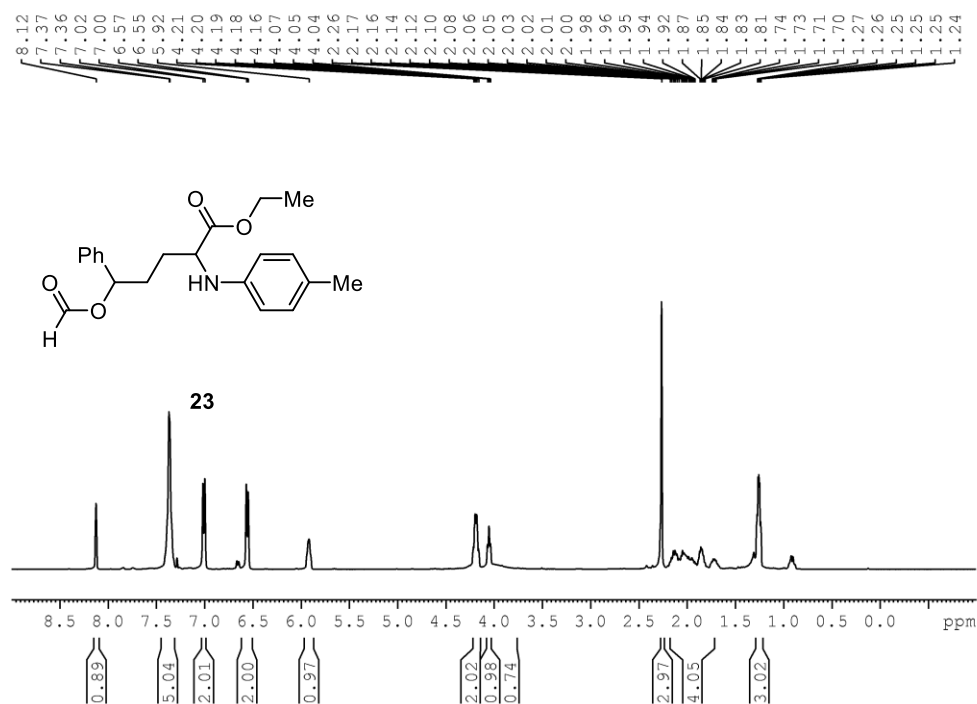
Supplementary Figure 35. ¹³C NMR spectrum of compound 21



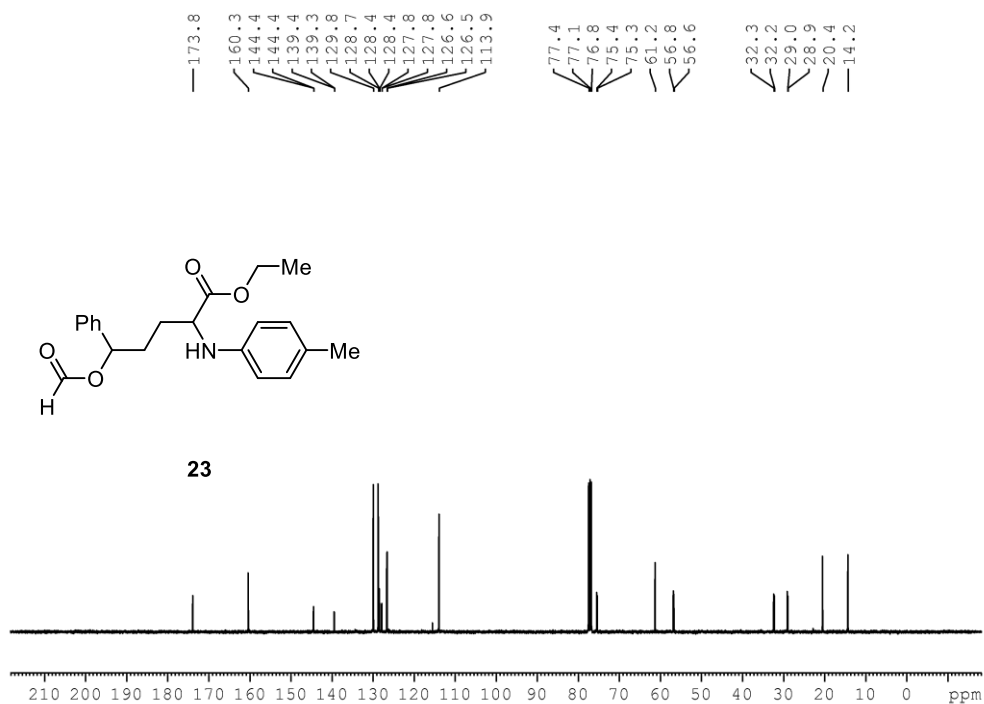
Supplementary Figure 36. ¹H NMR spectrum of compound 22



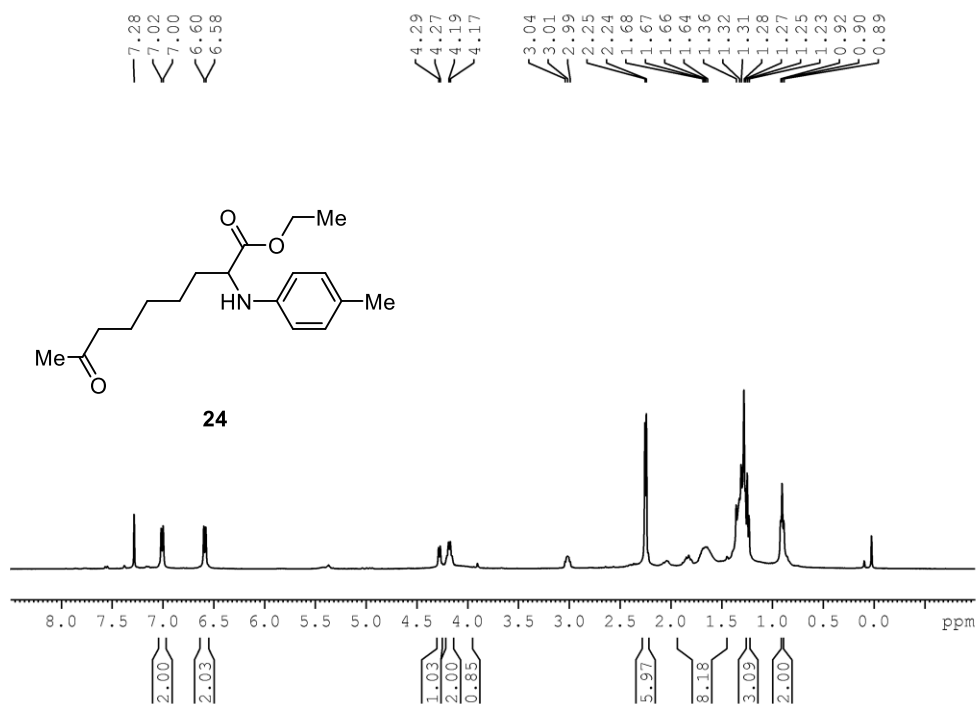
Supplementary Figure 37. ¹³C NMR spectrum of compound 22



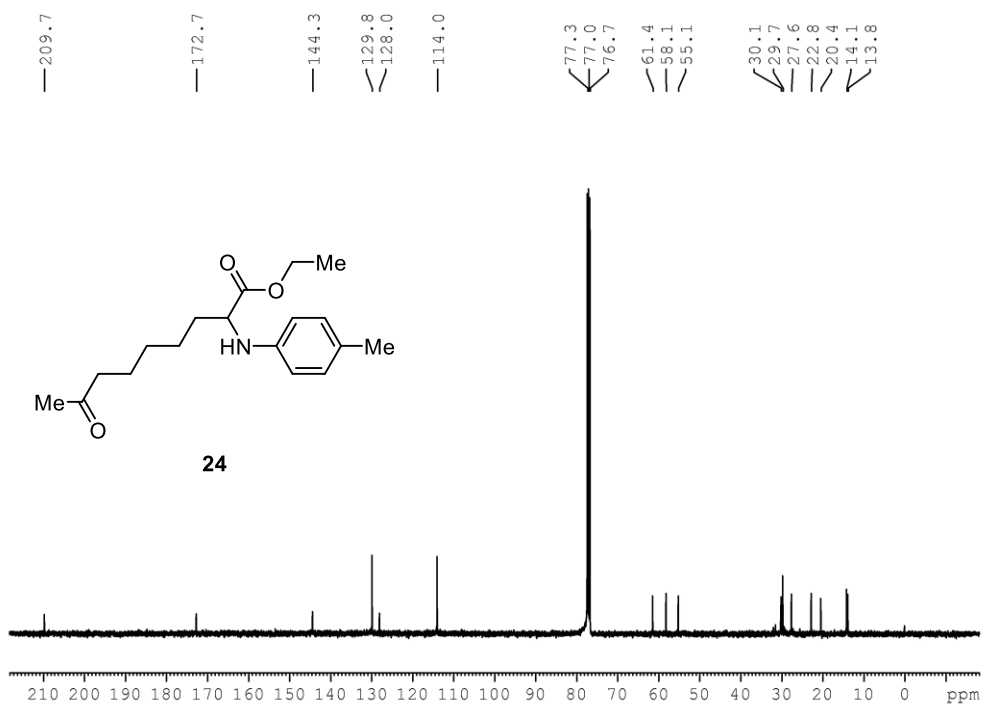
Supplementary Figure 38. ¹H NMR spectrum of compound 23



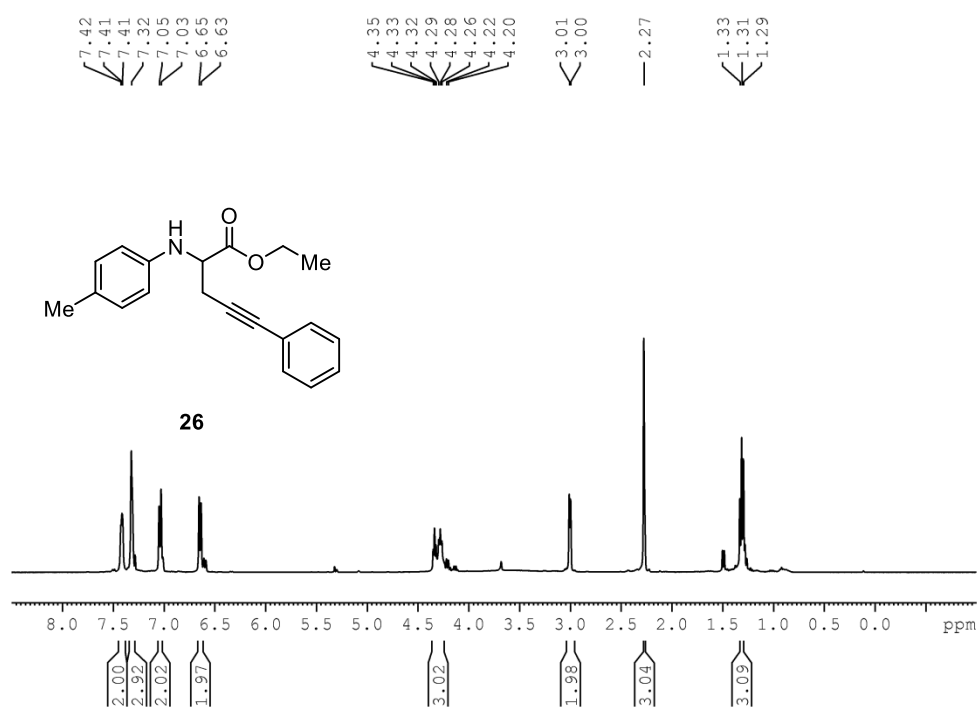
Supplementary Figure 39. ¹³C NMR spectrum of compound 23



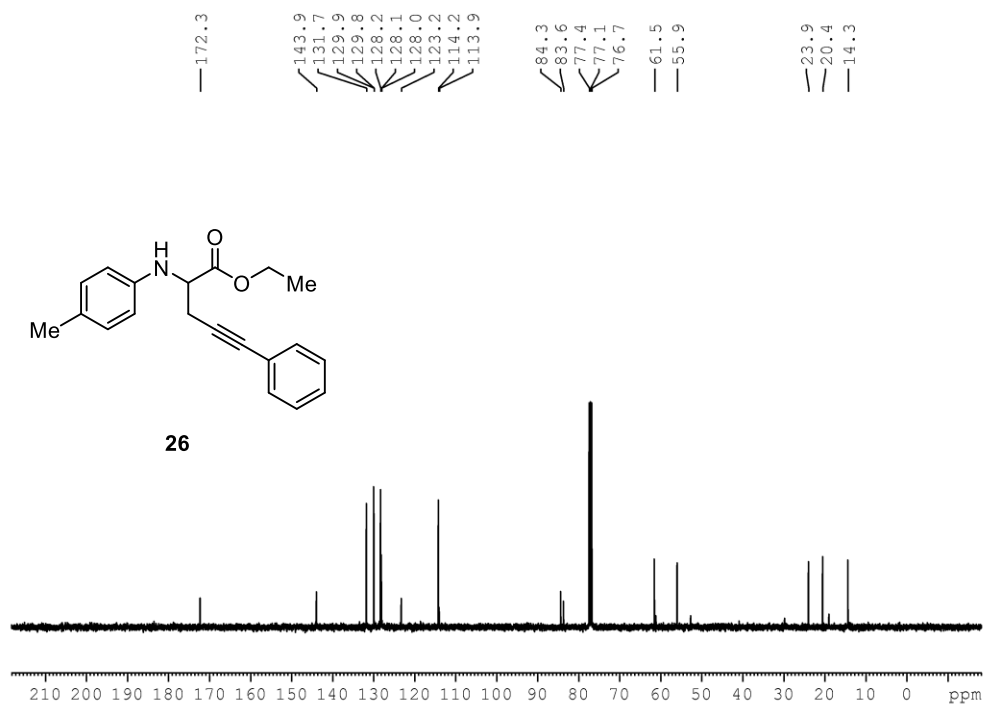
Supplementary Figure 40. ^1H NMR spectrum of compound 24



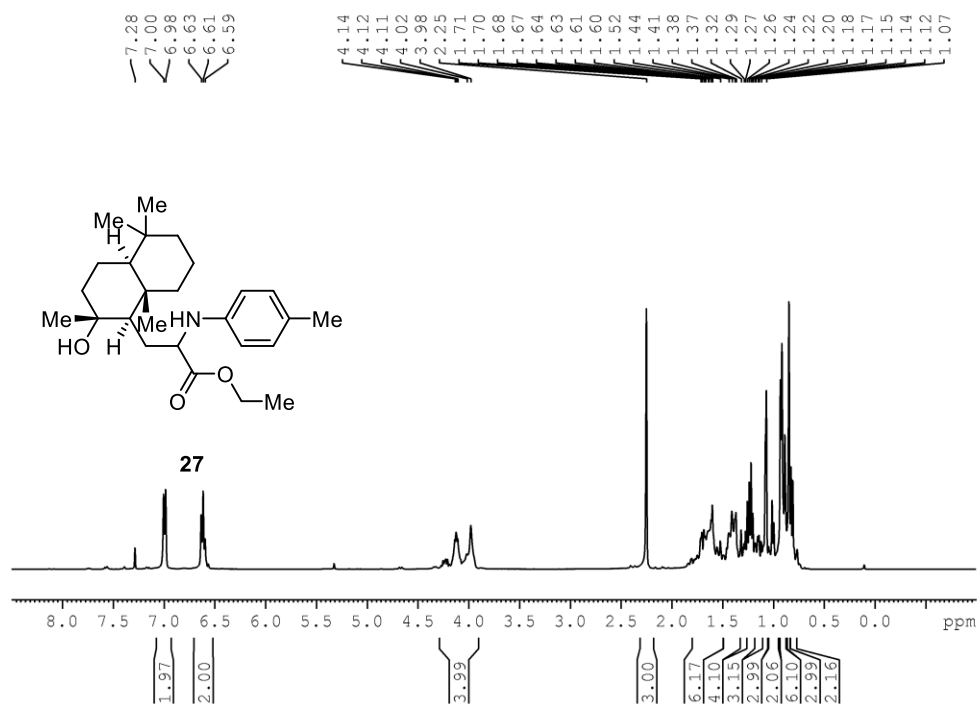
Supplementary Figure 41. ^{13}C NMR spectrum of compound 24



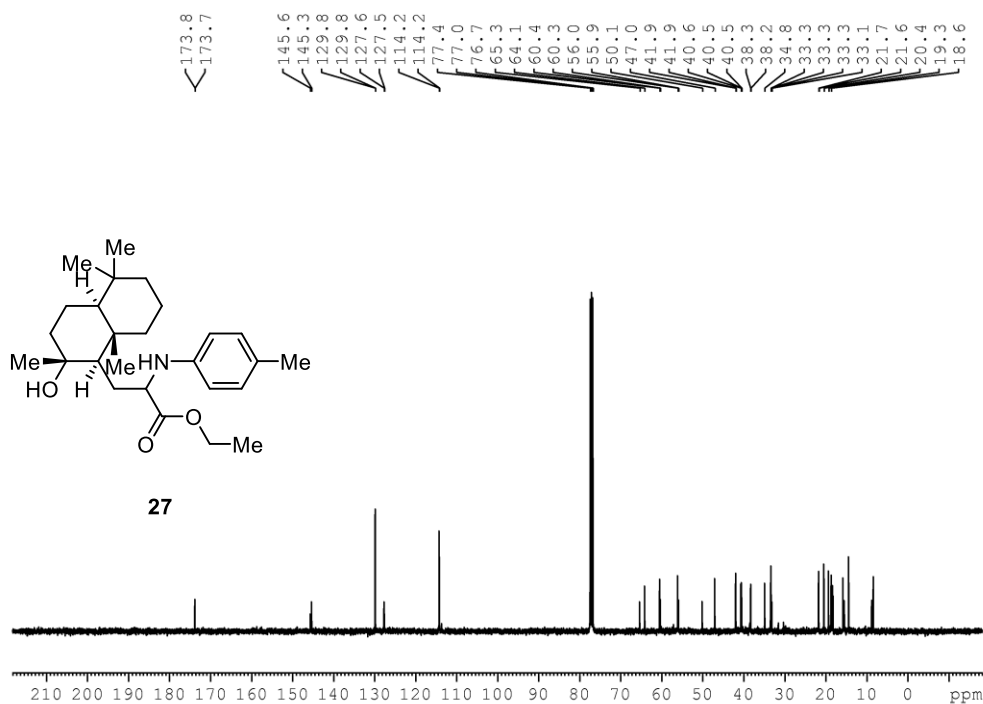
Supplementary Figure 42. ¹H NMR spectrum of compound 26



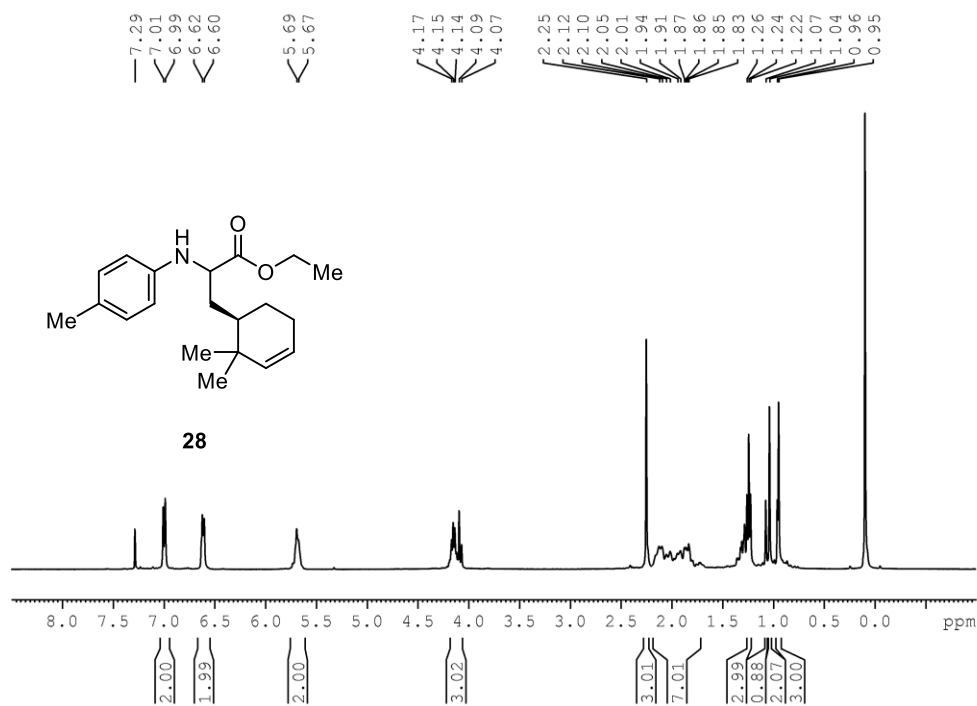
Supplementary Figure 43. ¹³C NMR spectrum of compound 26



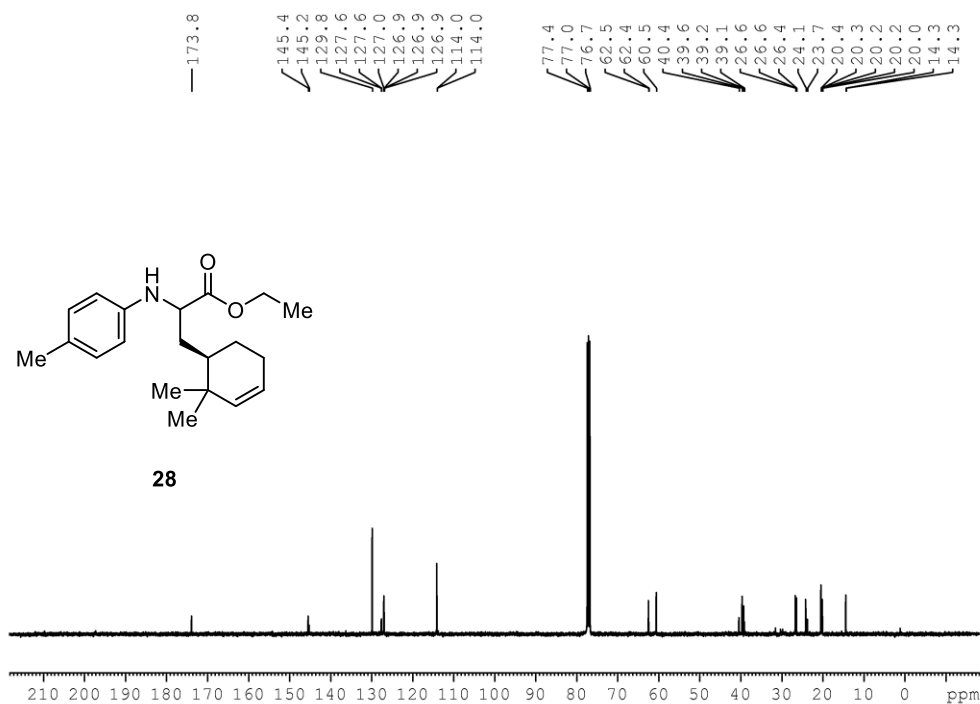
Supplementary Figure 44. ¹H NMR spectrum of compound 27



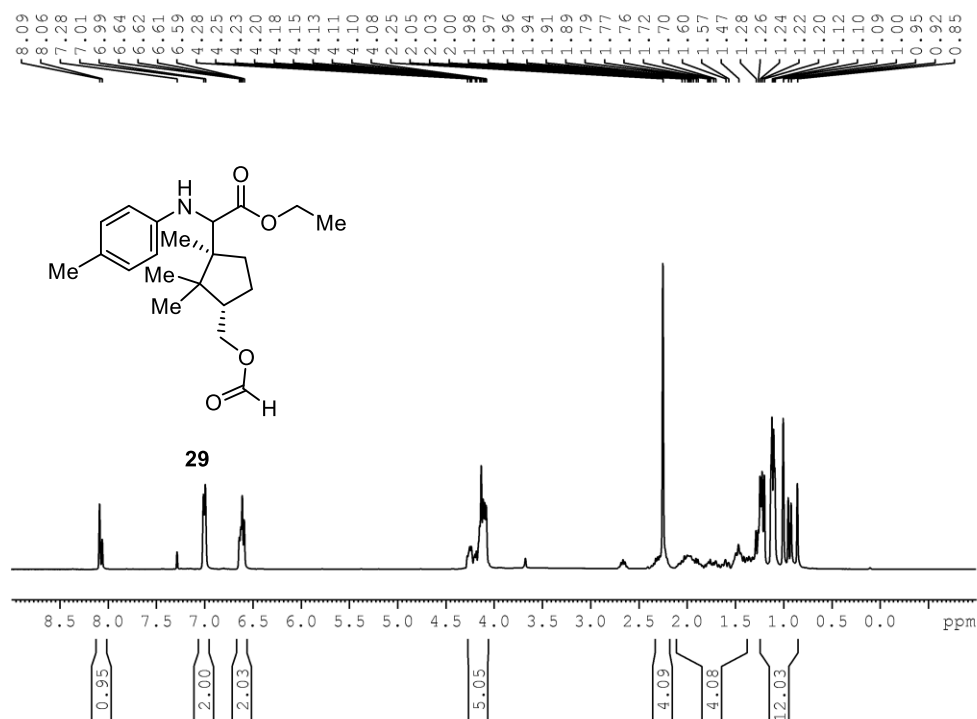
Supplementary Figure 45. ¹³C NMR spectrum of compound 27



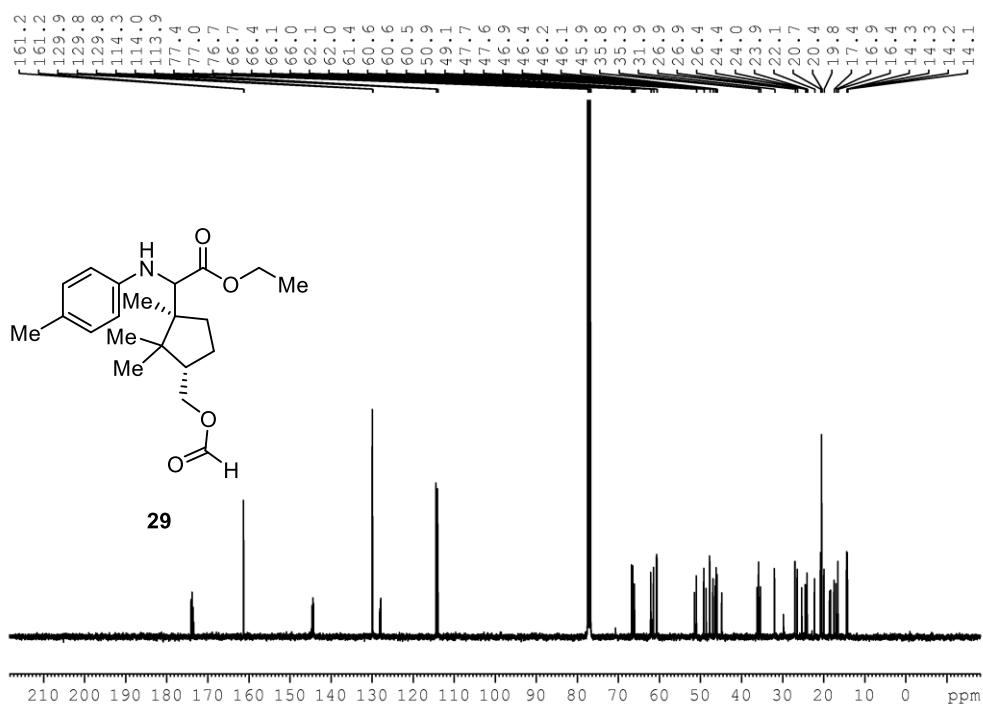
Supplementary Figure 46. ^1H NMR spectrum of compound 28



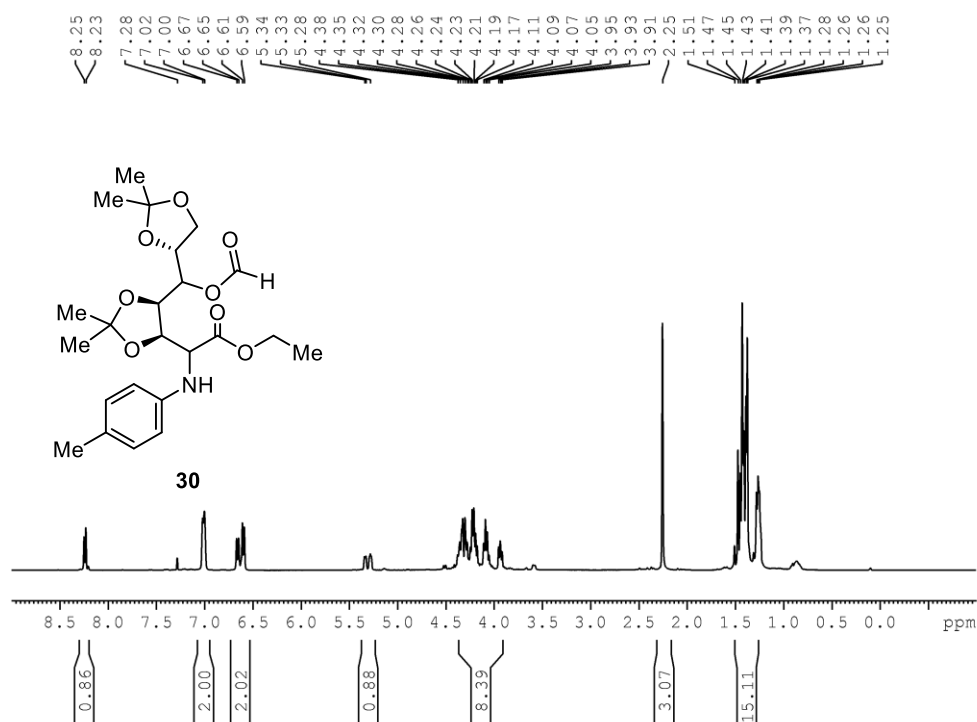
Supplementary Figure 47. ^{13}C NMR spectrum of compound 28



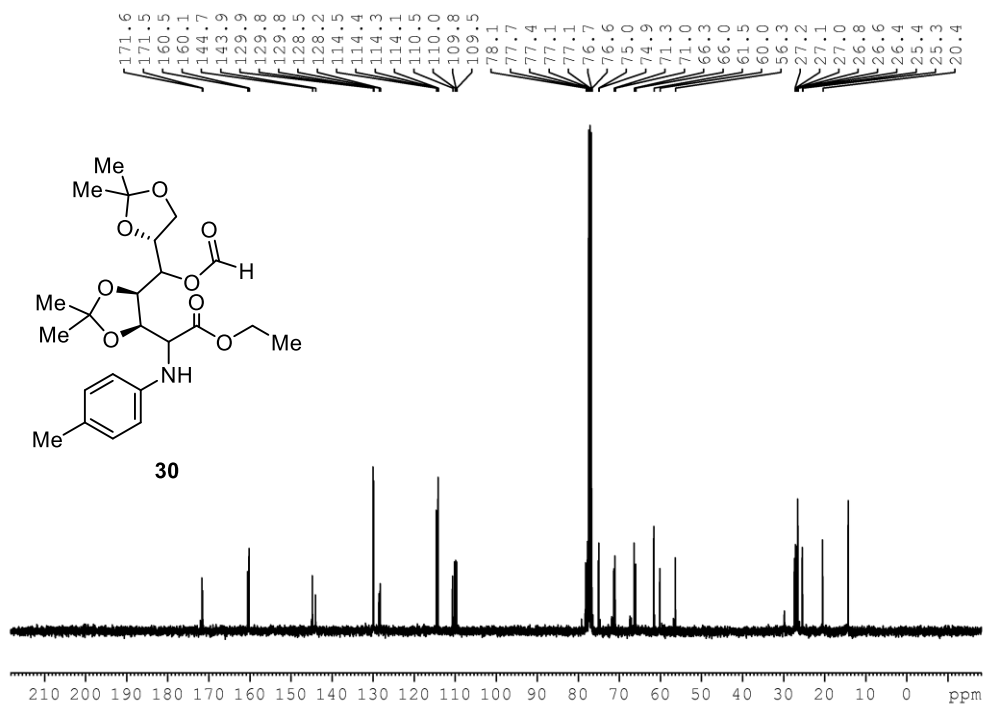
Supplementary Figure 48. ¹H NMR spectrum of compound 29



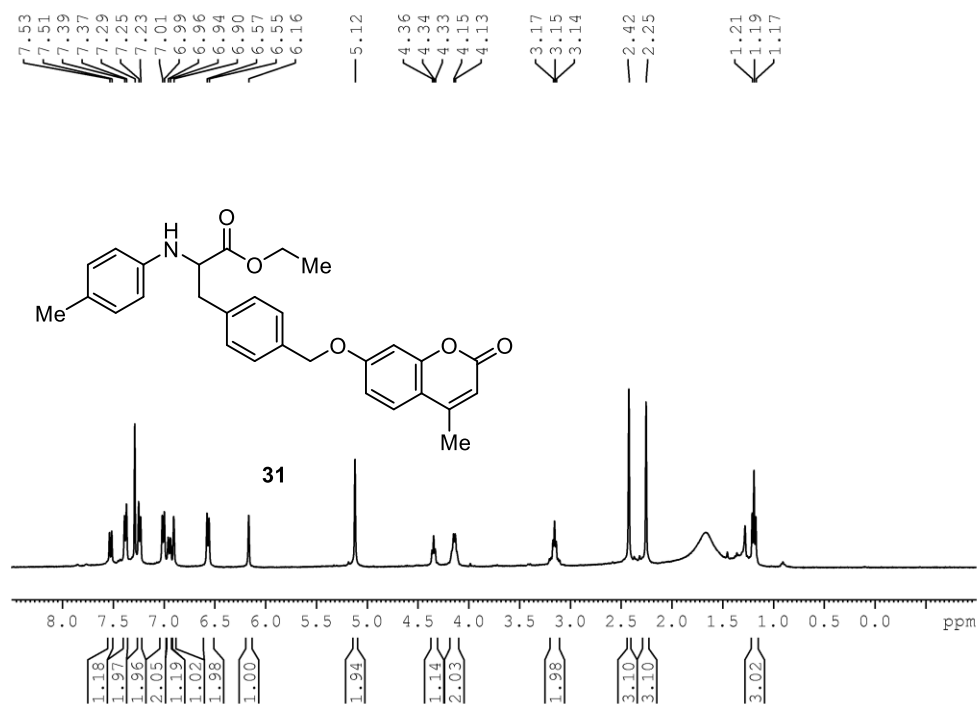
Supplementary Figure 49. ¹³C NMR spectrum of compound 29



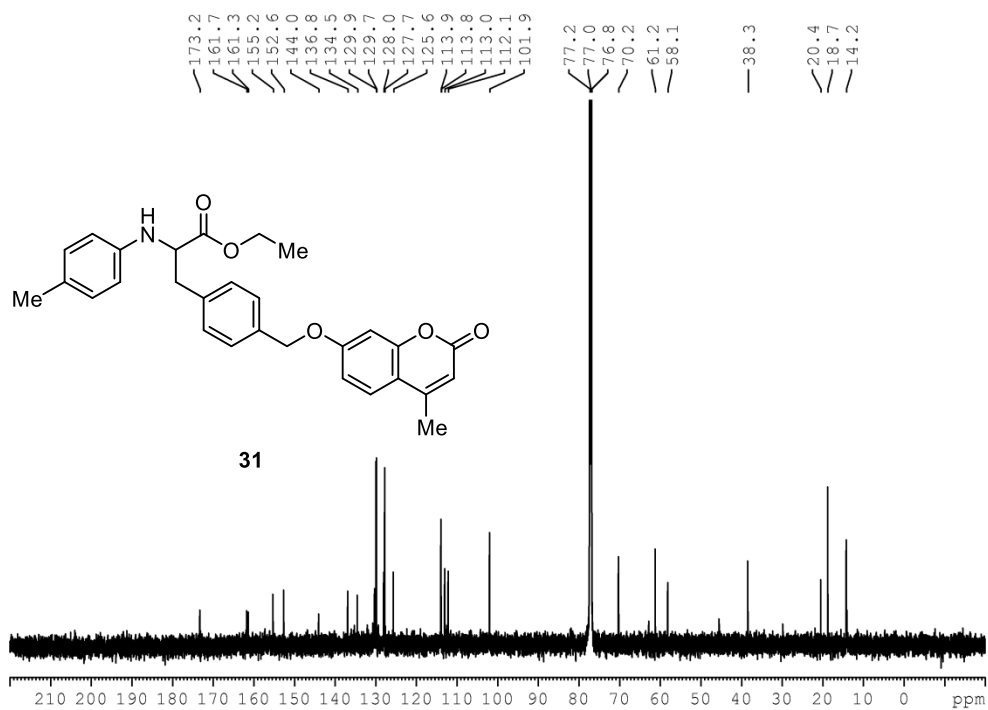
Supplementary Figure 50. ¹H NMR spectrum of compound 30



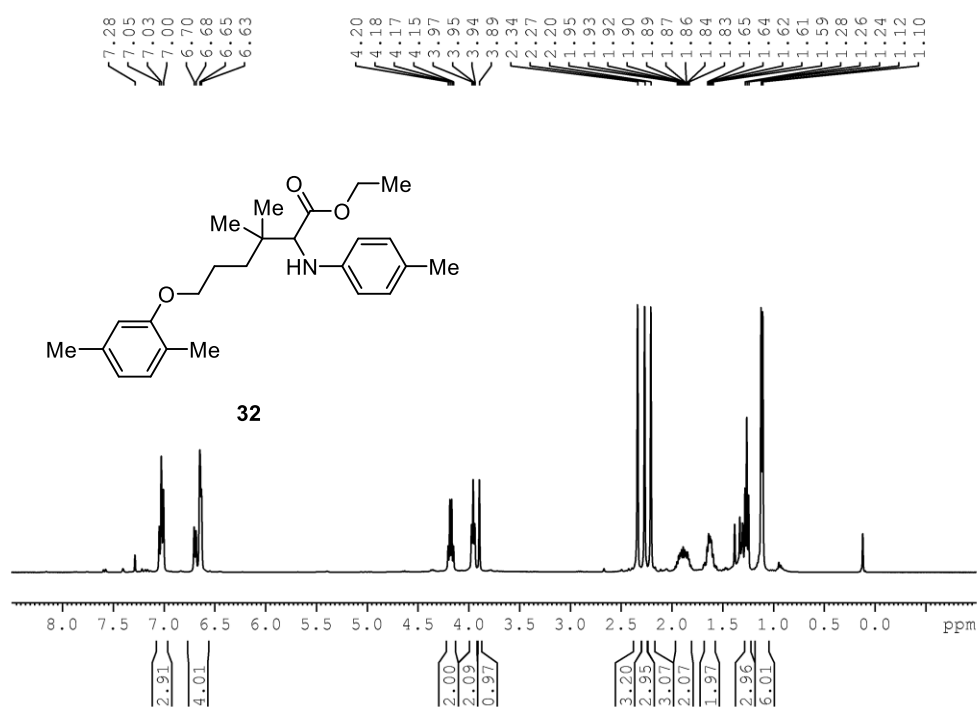
Supplementary Figure 51. ¹³C NMR spectrum of compound 30



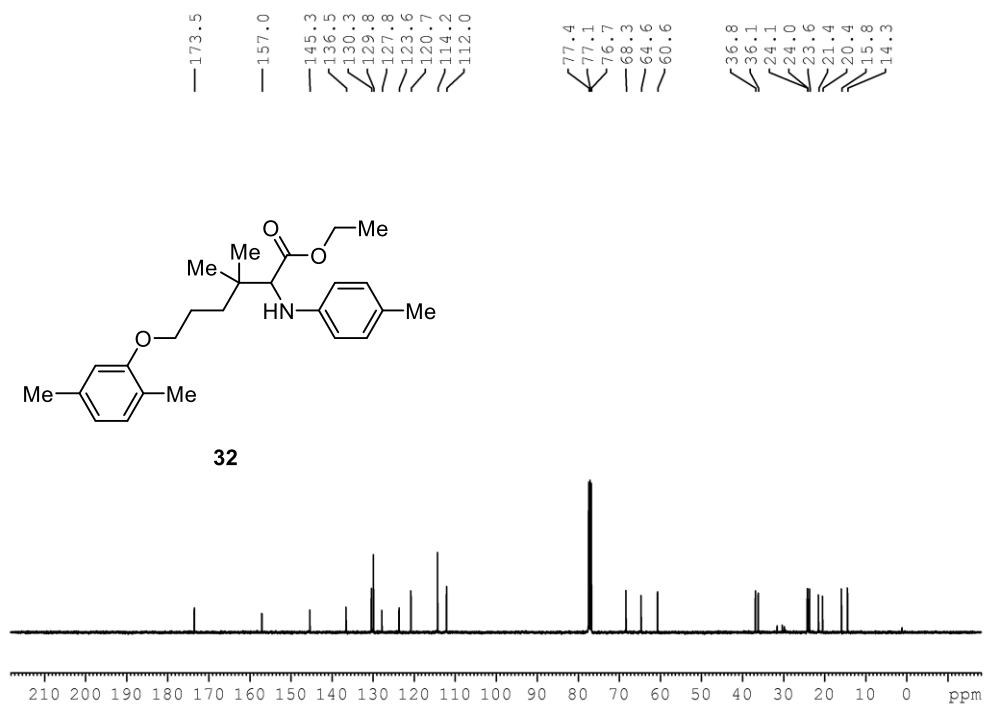
Supplementary Figure 52. ¹H NMR spectrum of compound 31



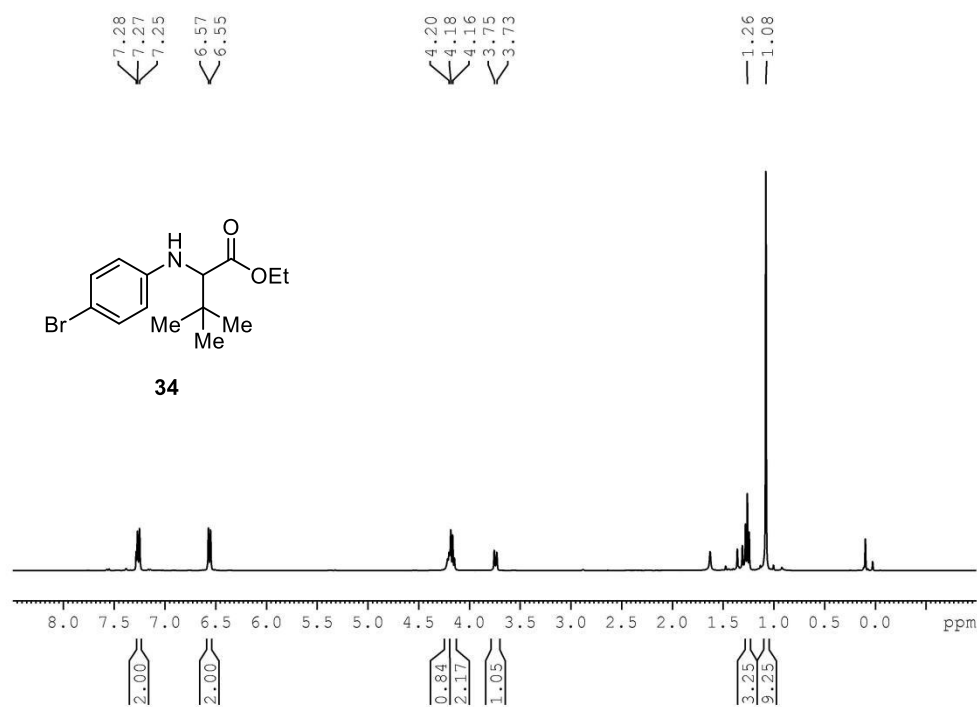
Supplementary Figure 53. ¹³C NMR spectrum of compound 31



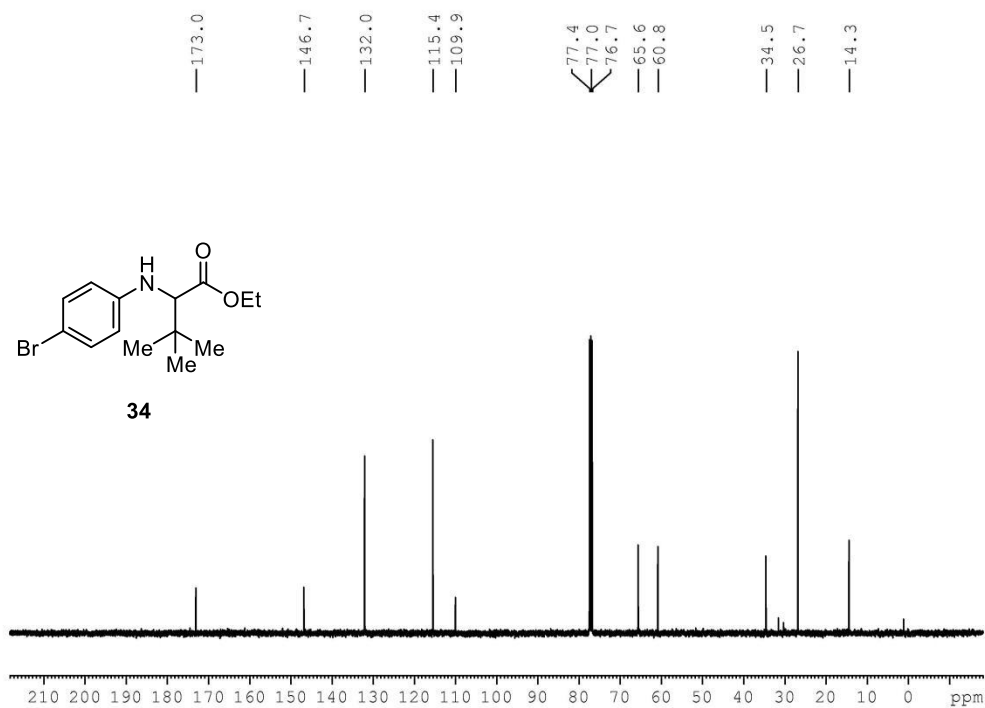
Supplementary Figure 54. ^1H NMR spectrum of compound 32



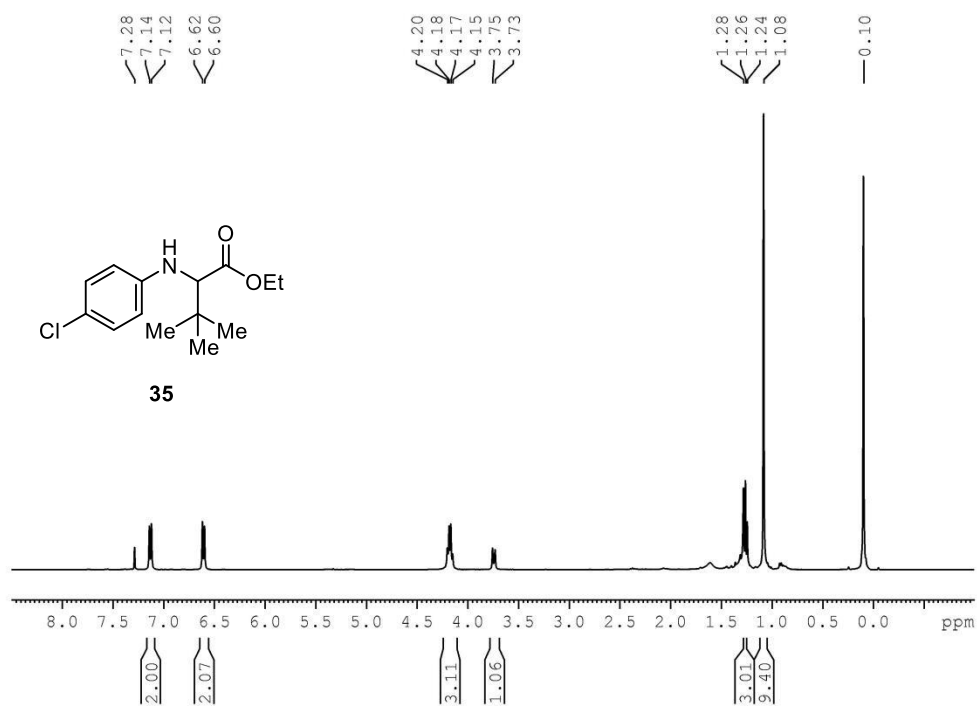
Supplementary Figure 55. ^{13}C NMR spectrum of compound 32



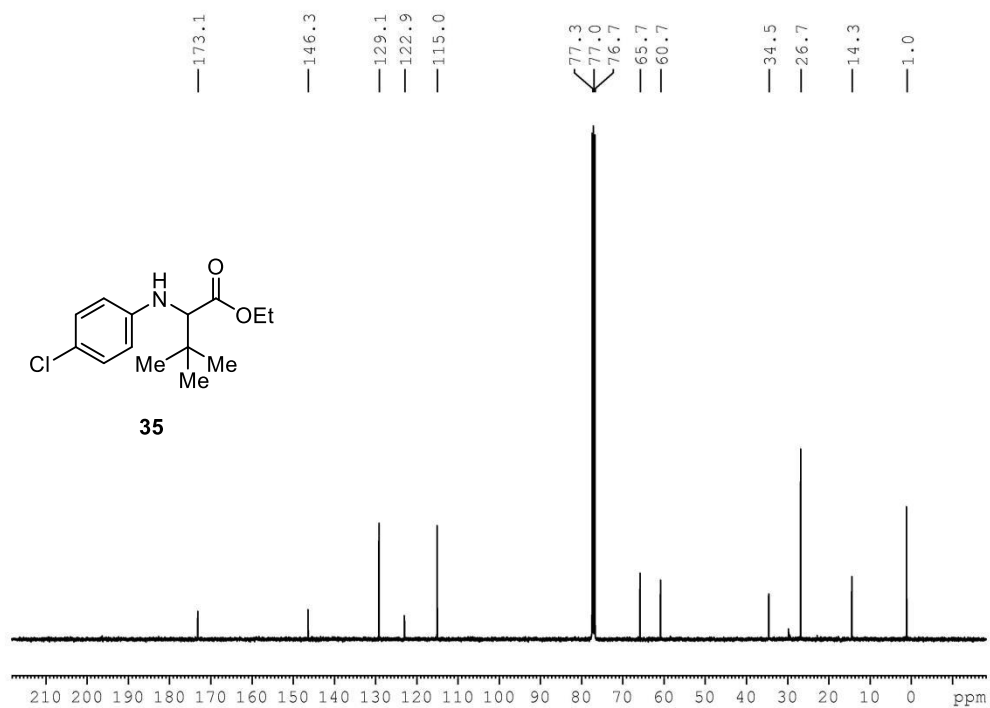
Supplementary Figure 56. ¹H NMR spectrum of compound 34



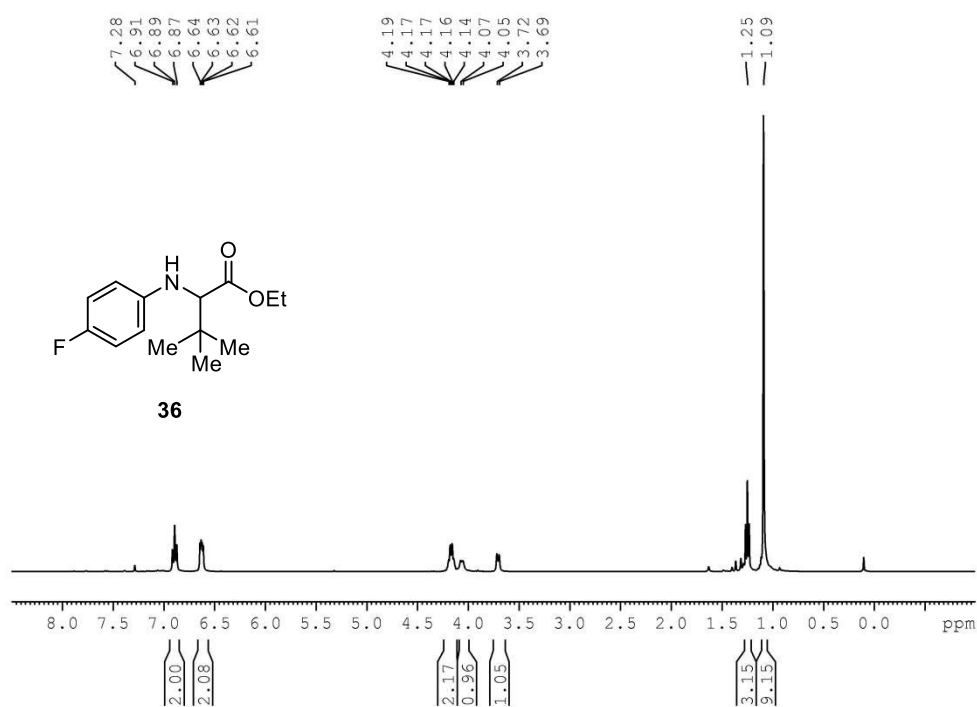
Supplementary Figure 57. ¹³C NMR spectrum of compound 34



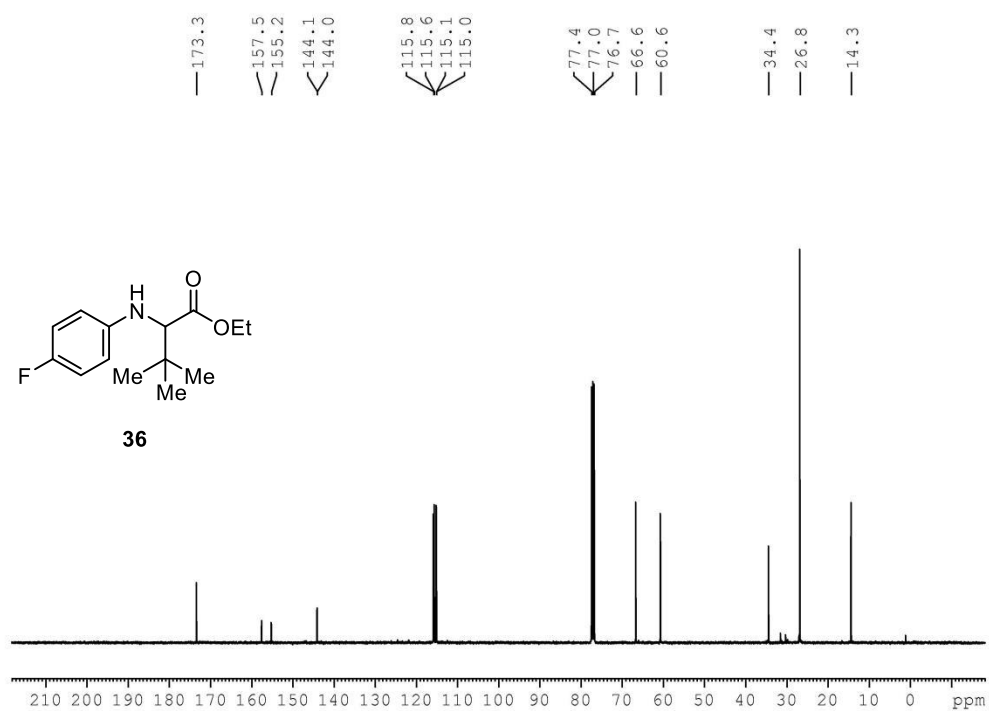
Supplementary Figure 58. ¹H NMR spectrum of compound 35



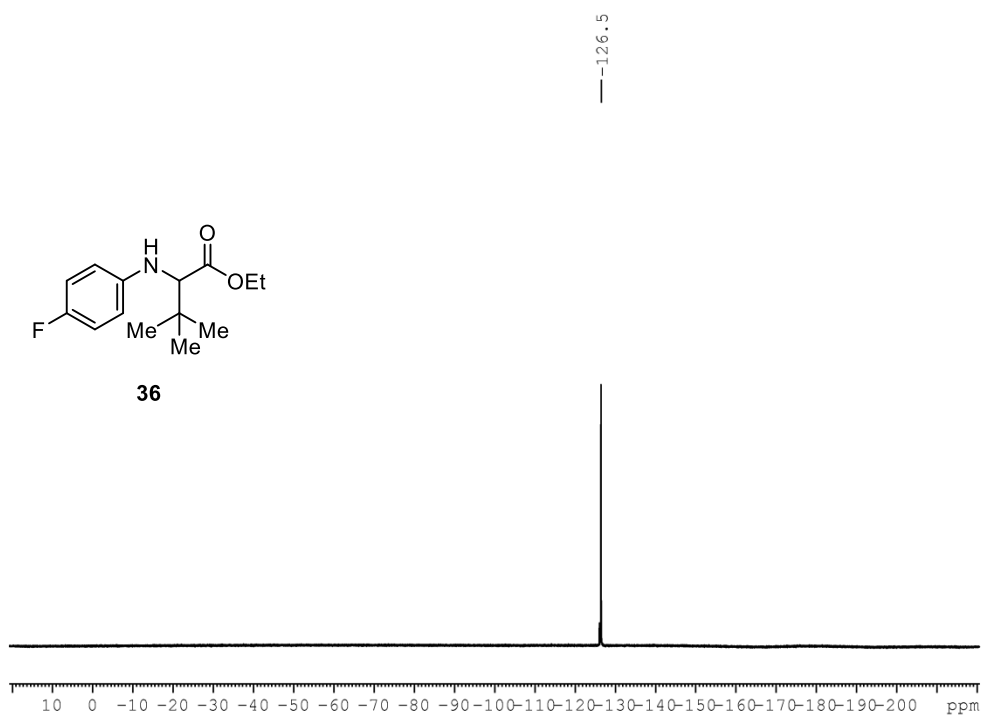
Supplementary Figure 59. ¹³C NMR spectrum of compound 35



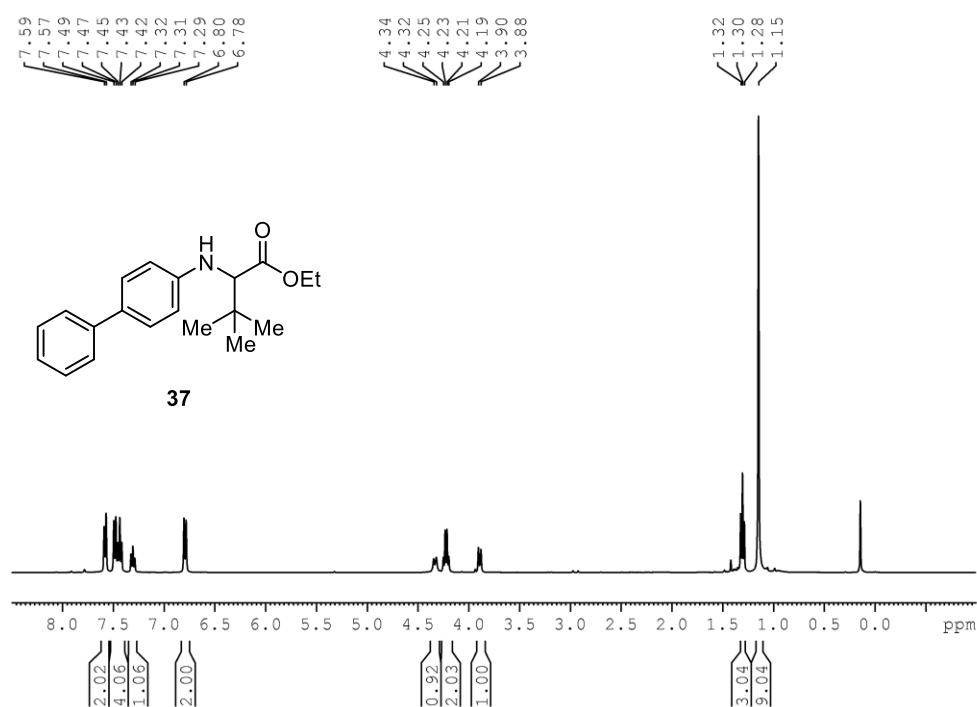
Supplementary Figure 60. ¹H NMR spectrum of compound 36



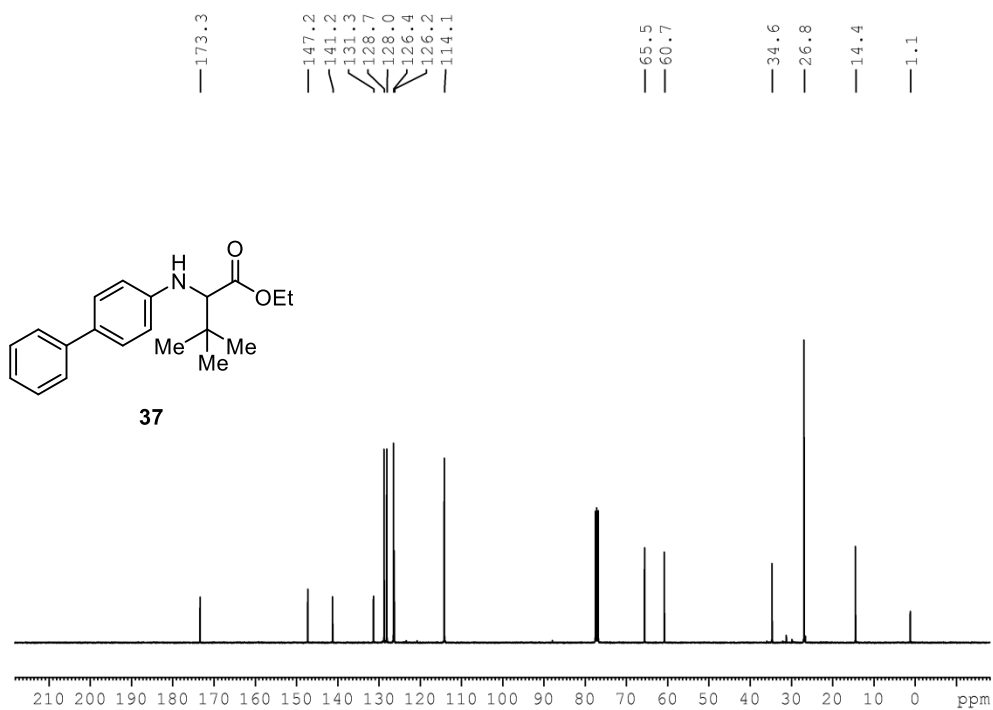
Supplementary Figure 61. ¹³C NMR spectrum of compound 36



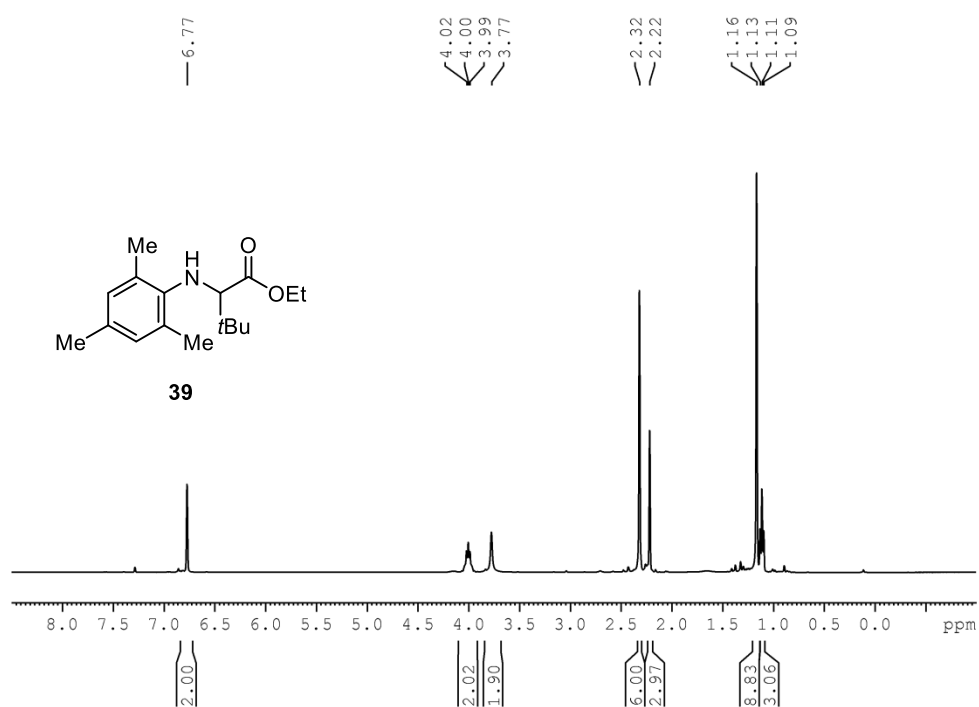
Supplementary Figure 62. ^{19}F NMR spectrum of compound 36



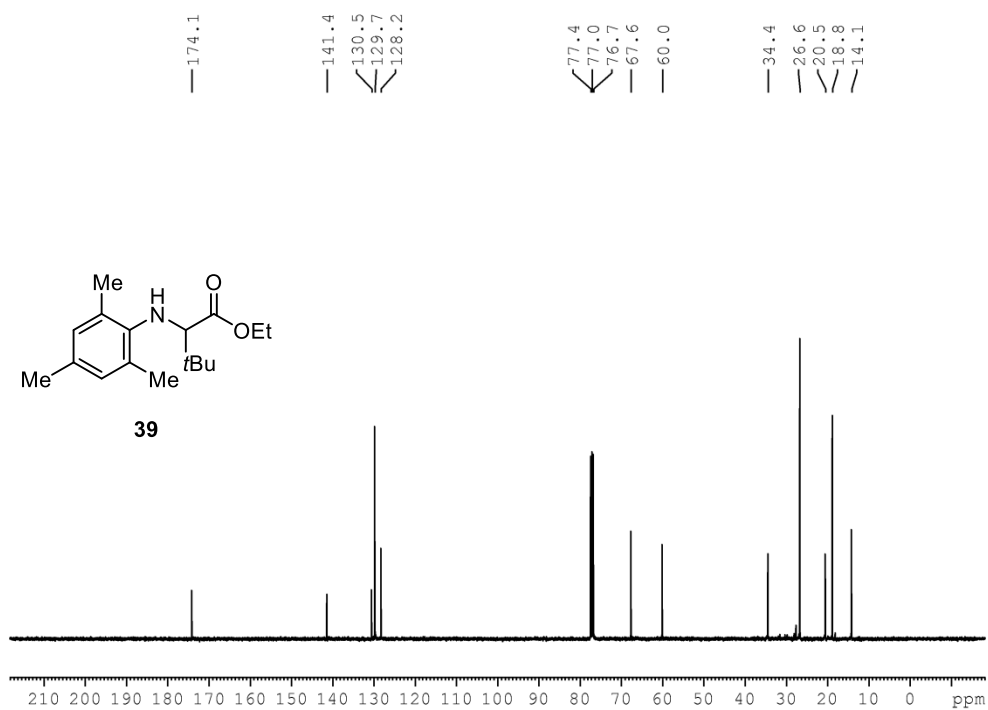
Supplementary Figure 63. ¹H NMR spectrum of compound 37



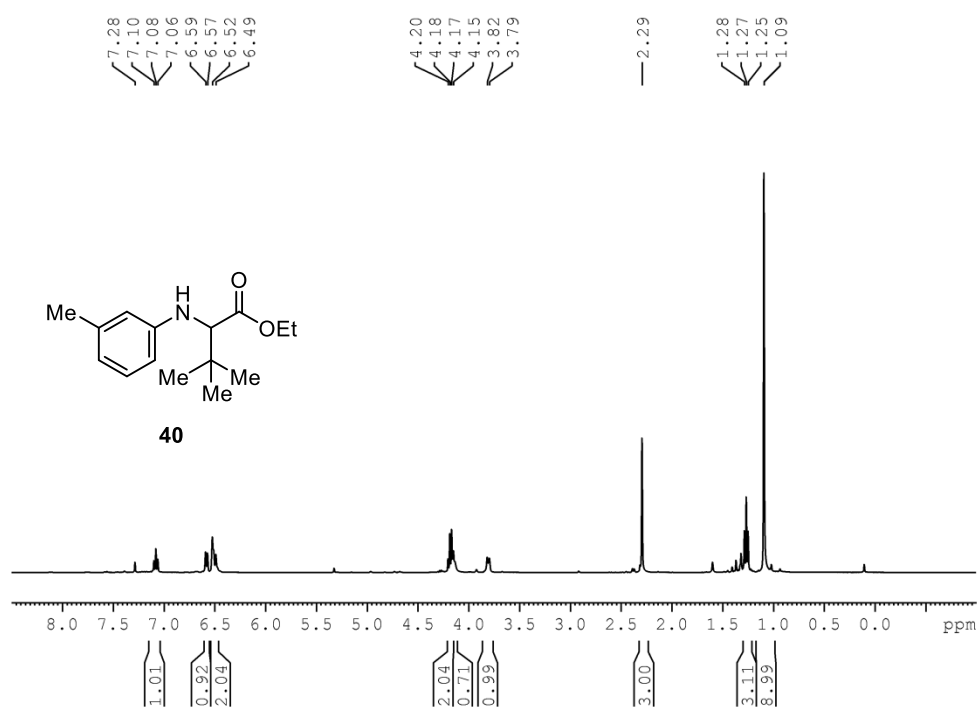
Supplementary Figure 64. ¹³C NMR spectrum of compound 37



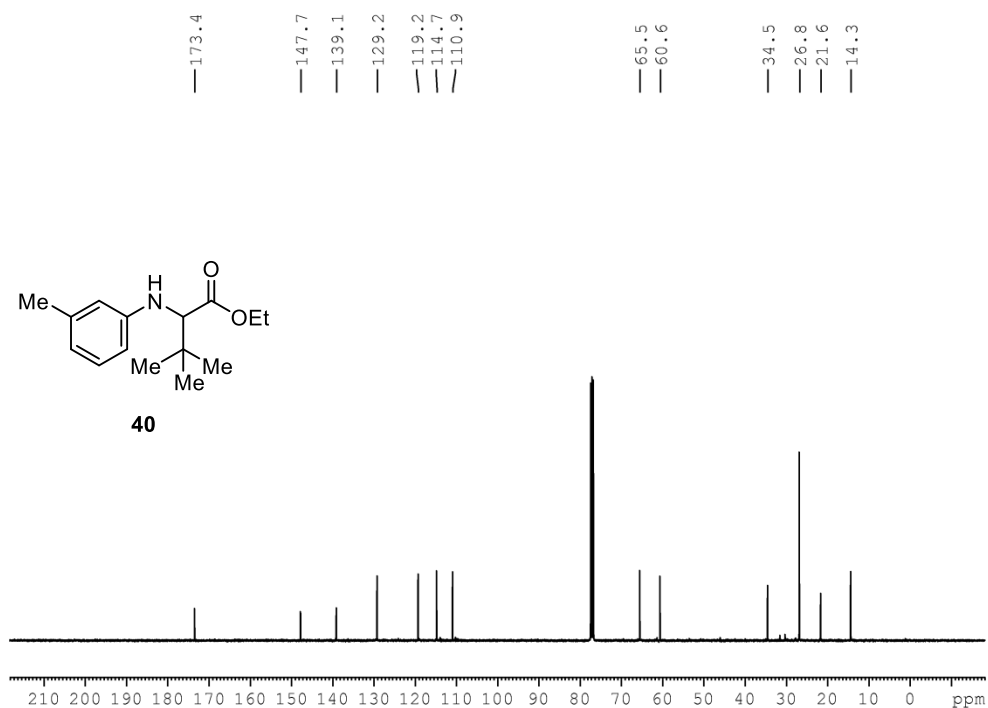
Supplementary Figure 65. ¹H NMR spectrum of compound 39



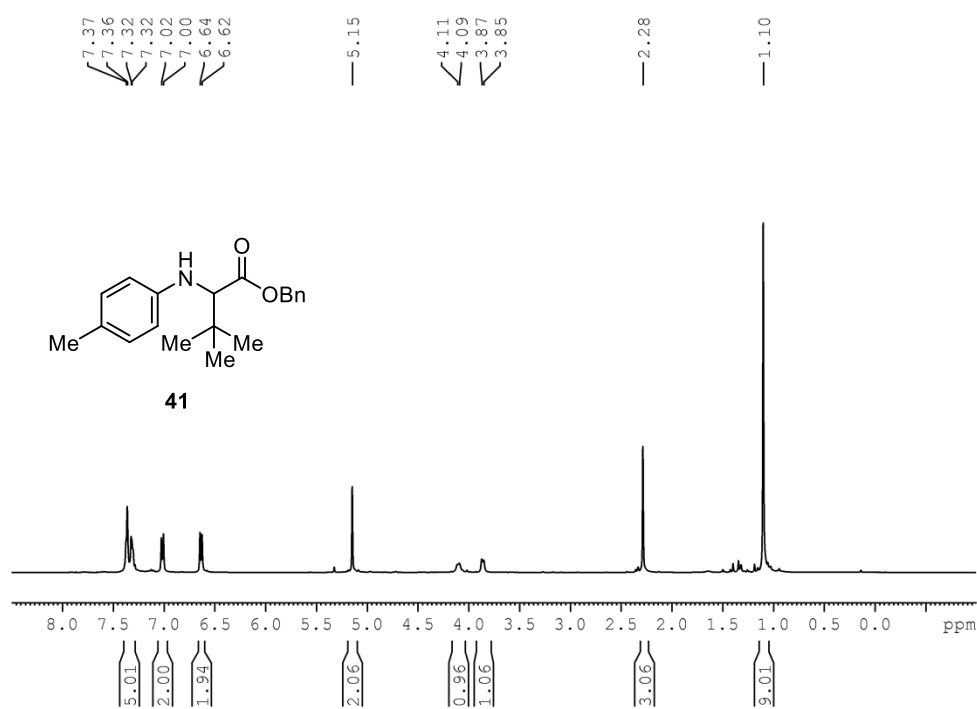
Supplementary Figure 66. ¹³C NMR spectrum of compound 39



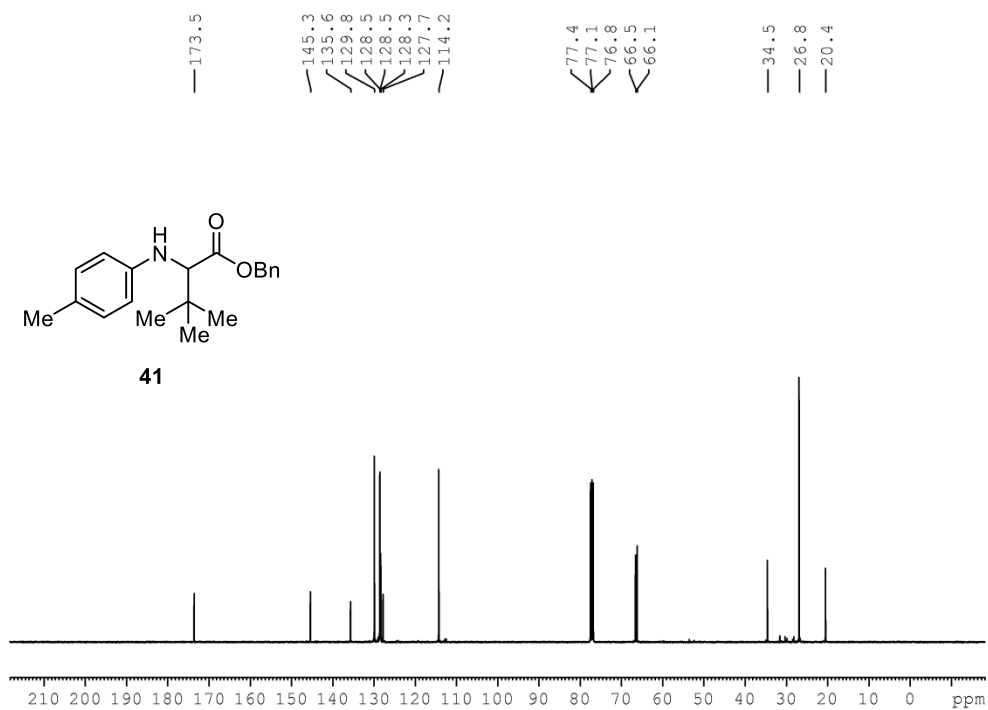
Supplementary Figure 67. ^1H NMR spectrum of compound 40



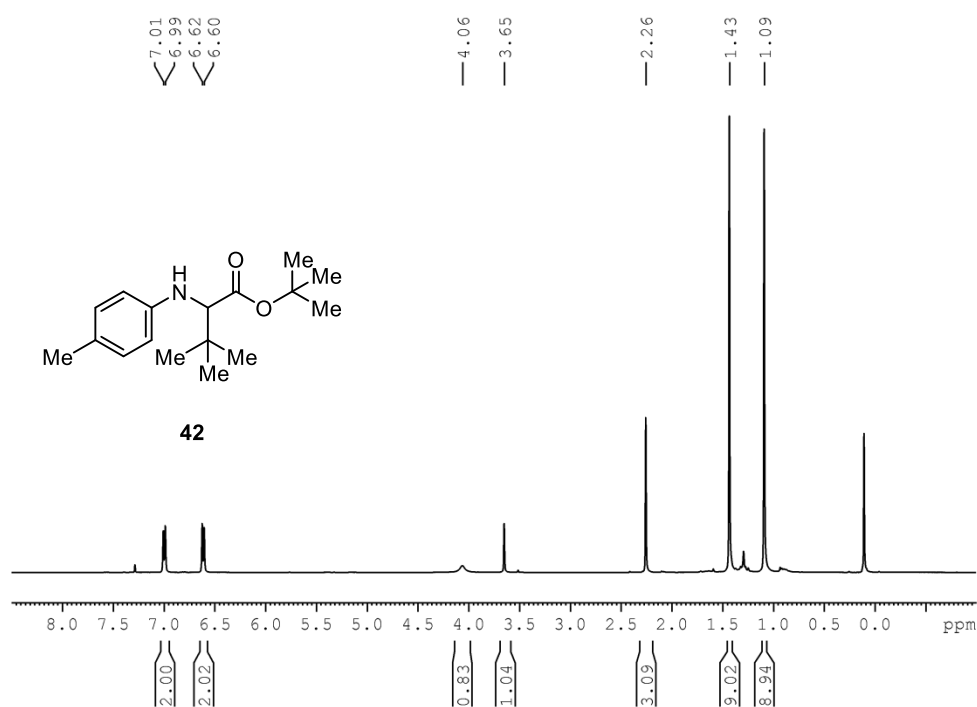
Supplementary Figure 68. ^{13}C NMR spectrum of compound 40



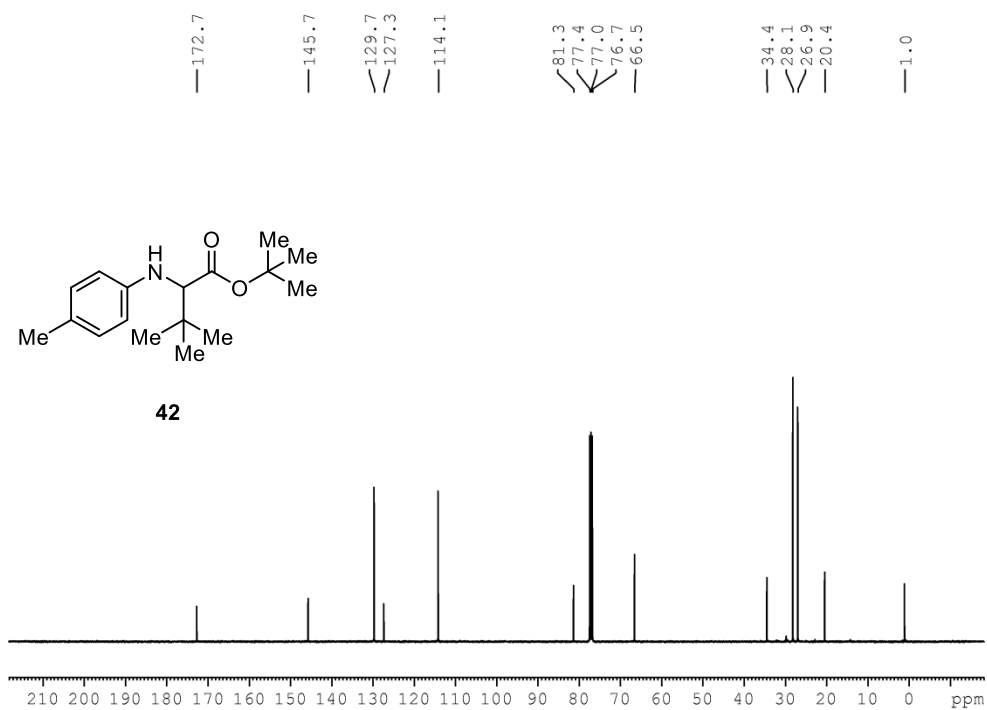
Supplementary Figure 69. ^1H NMR spectrum of compound 41



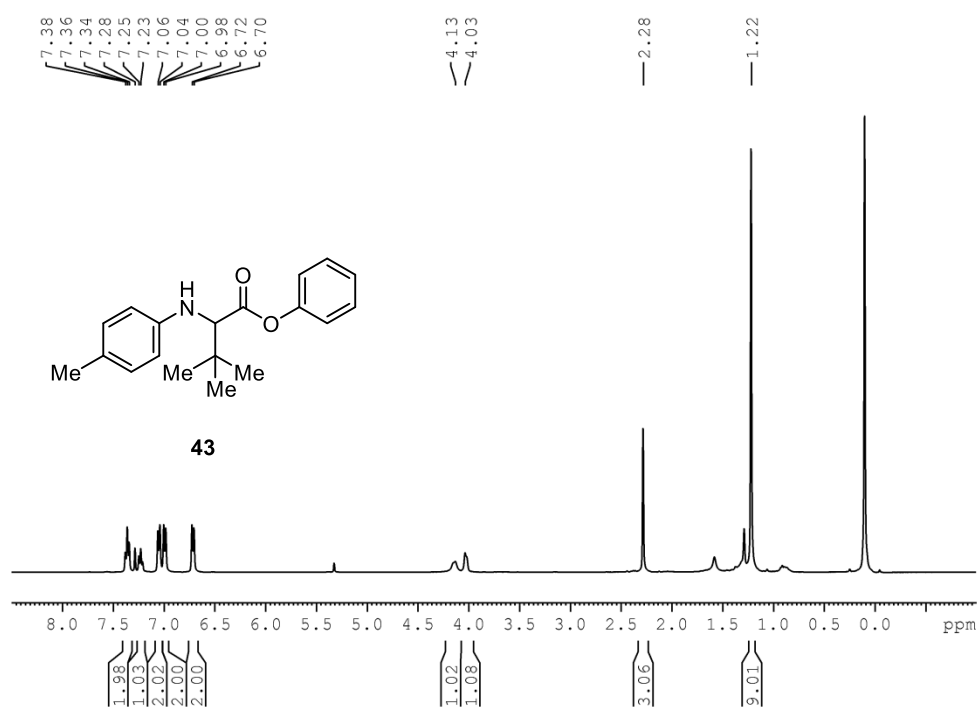
Supplementary Figure 70. ^{13}C NMR spectrum of compound 41



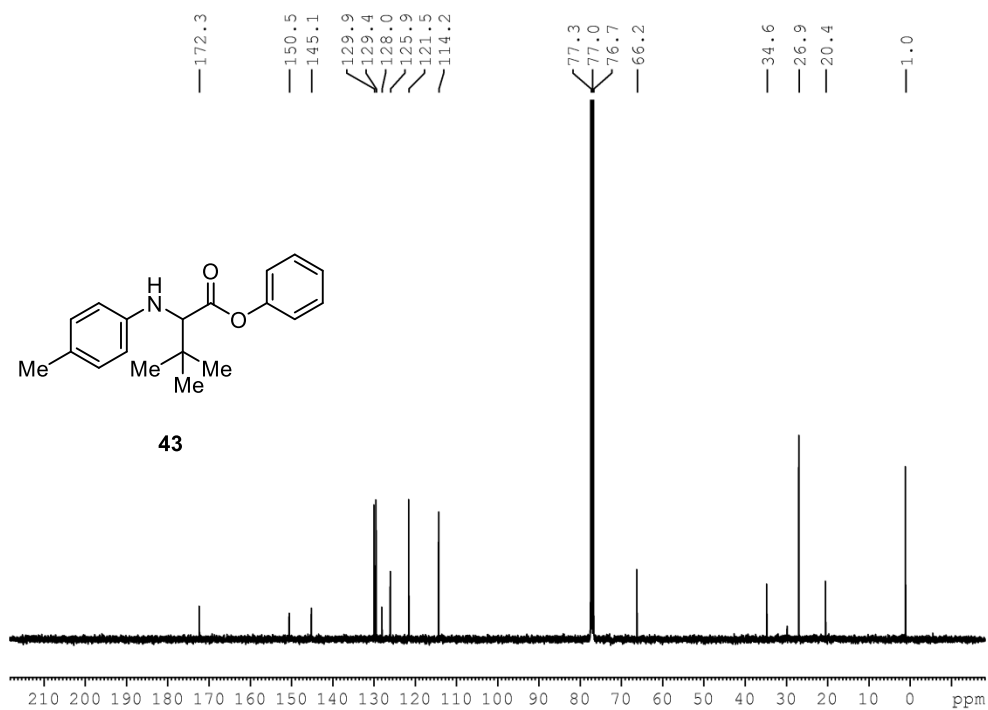
Supplementary Figure 71. ¹H NMR spectrum of compound 42



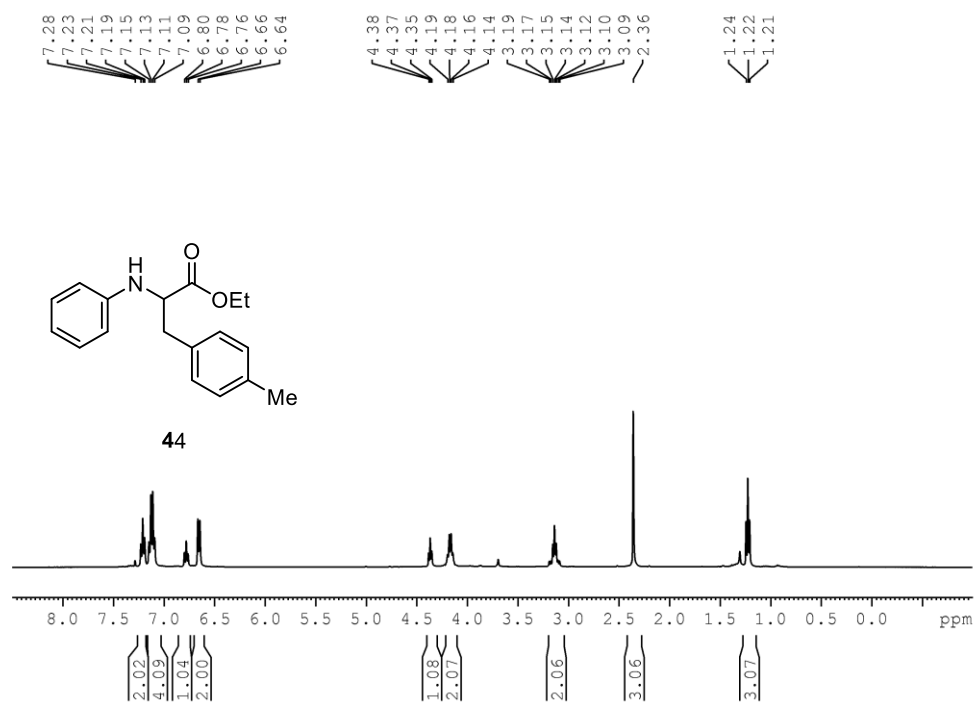
Supplementary Figure 72. ¹³C NMR spectrum of compound 42



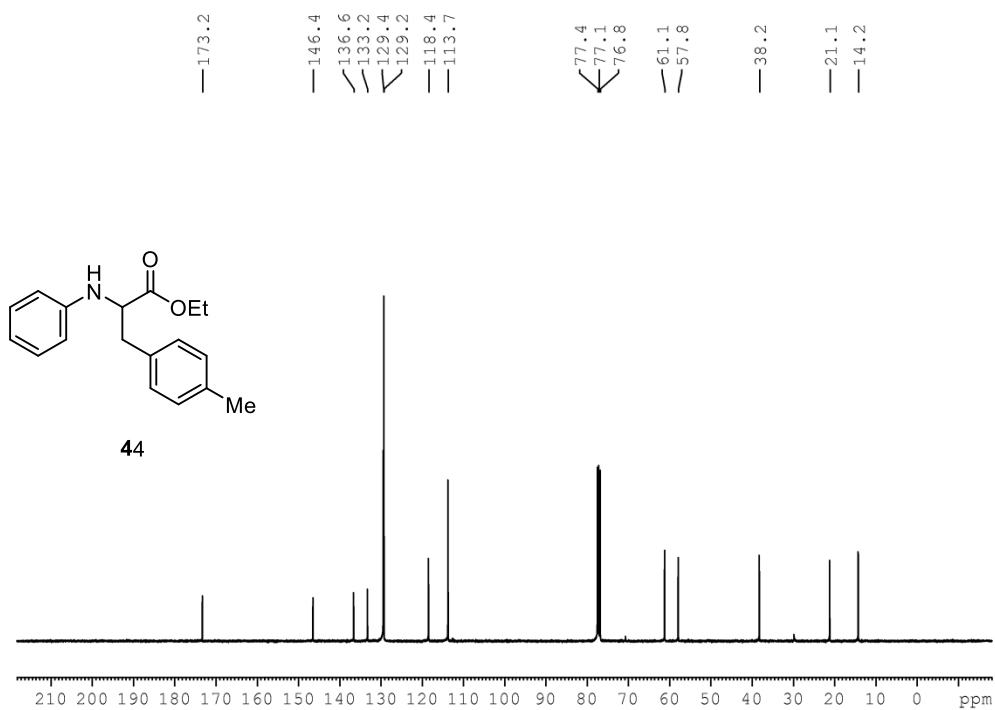
Supplementary Figure 73. ¹H NMR spectrum of compound 43



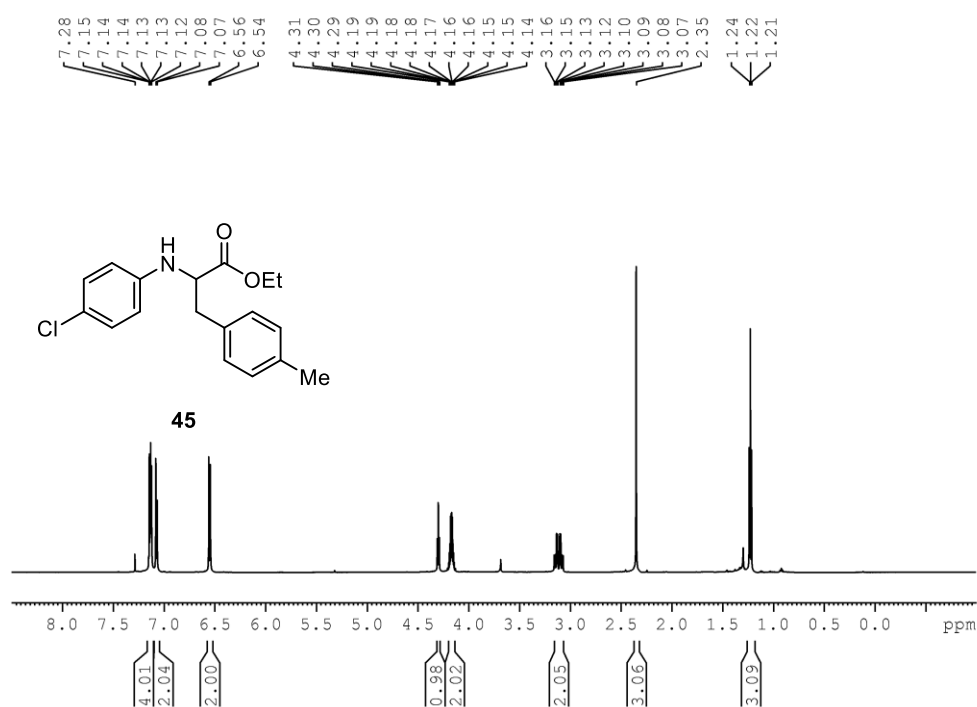
Supplementary Figure 74. ¹³C NMR spectrum of compound 43



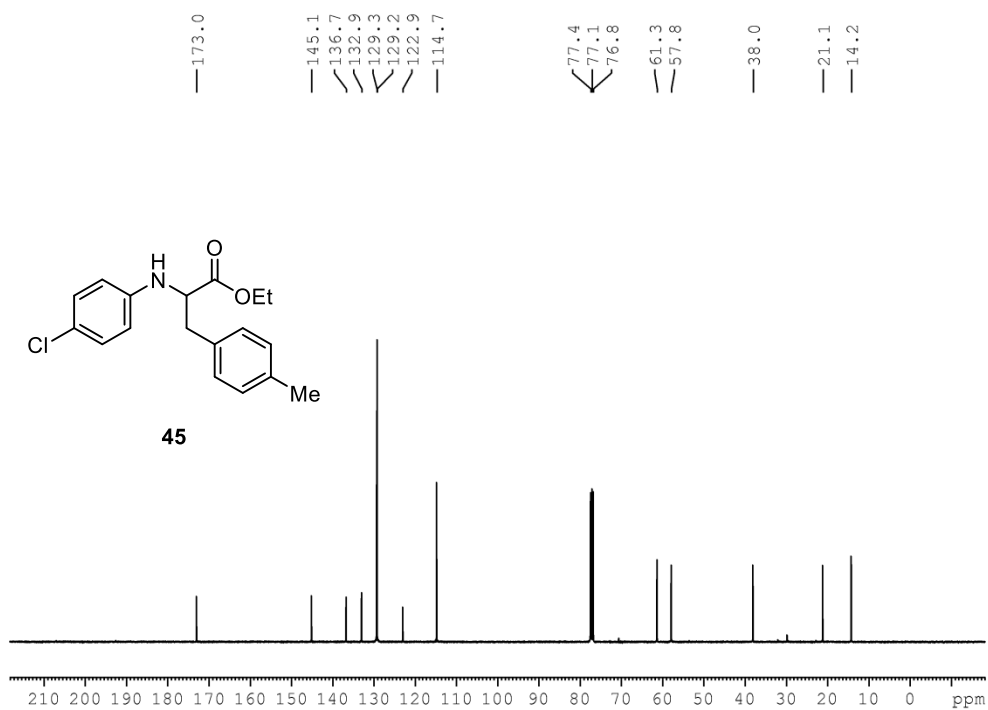
Supplementary Figure 75. ¹H NMR spectrum of compound 44



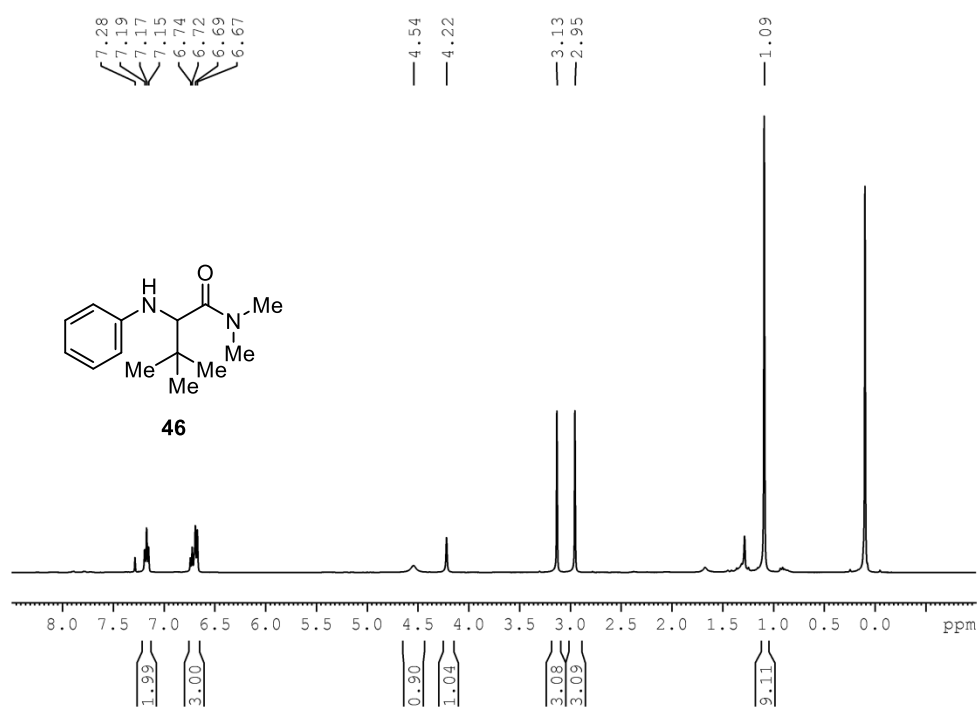
Supplementary Figure 76. ¹³C NMR spectrum of compound 44



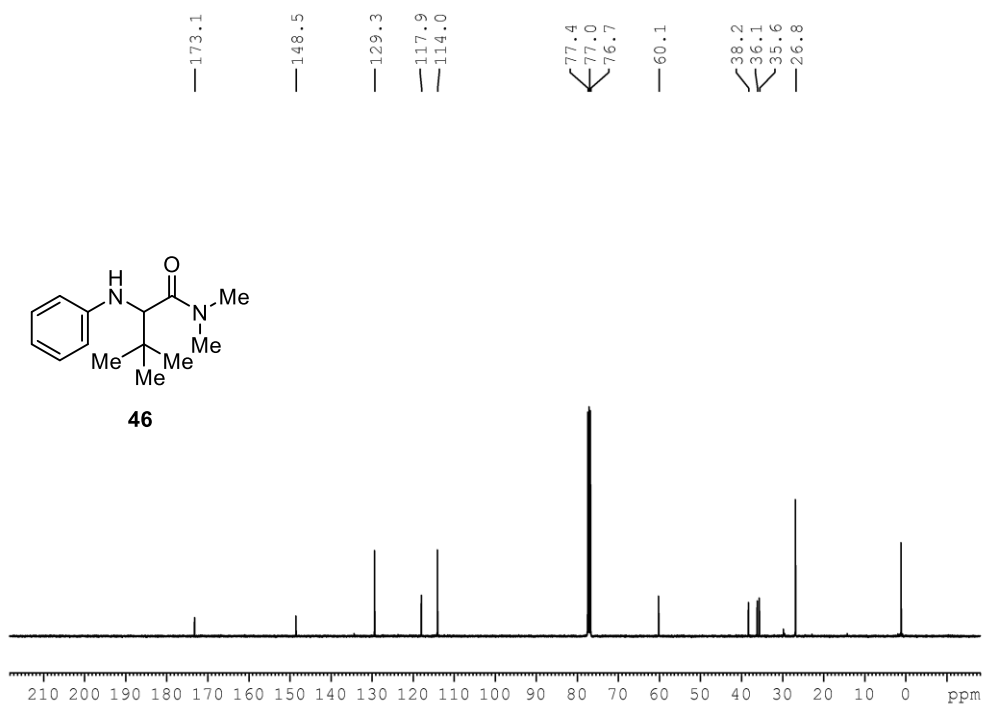
Supplementary Figure 77. ¹H NMR spectrum of compound 45



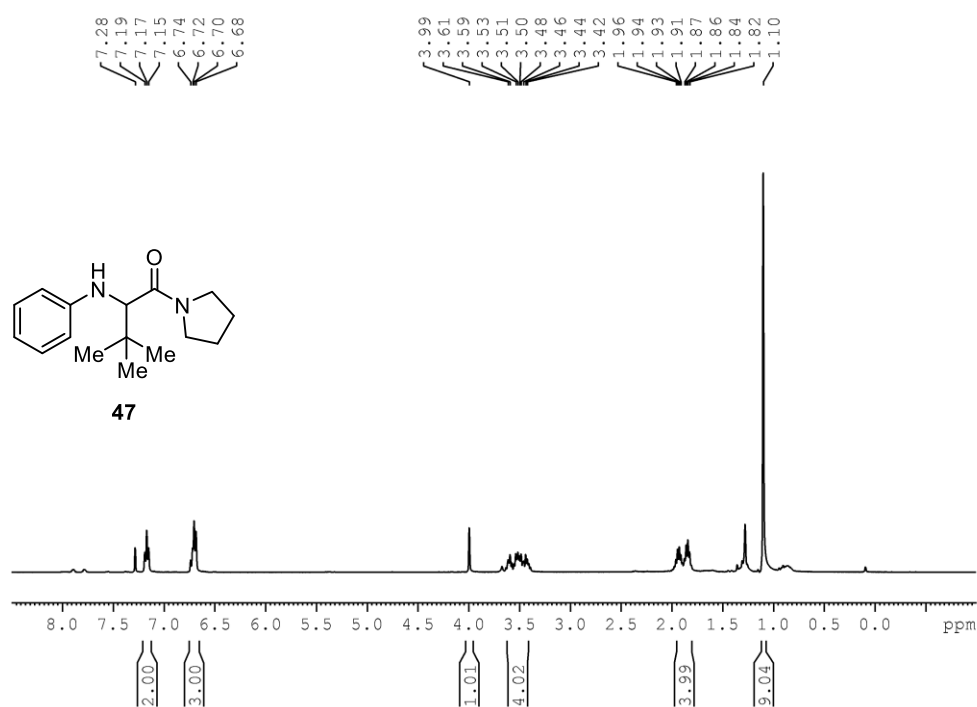
Supplementary Figure 78. ¹³C NMR spectrum of compound 45



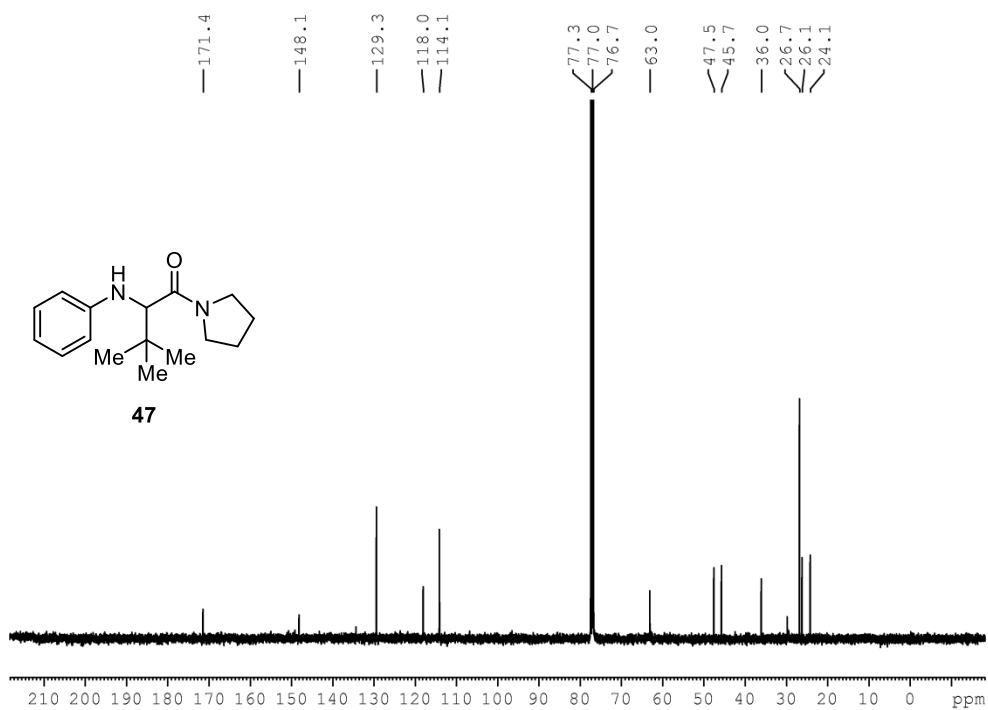
Supplementary Figure 79. ^1H NMR spectrum of compound 46



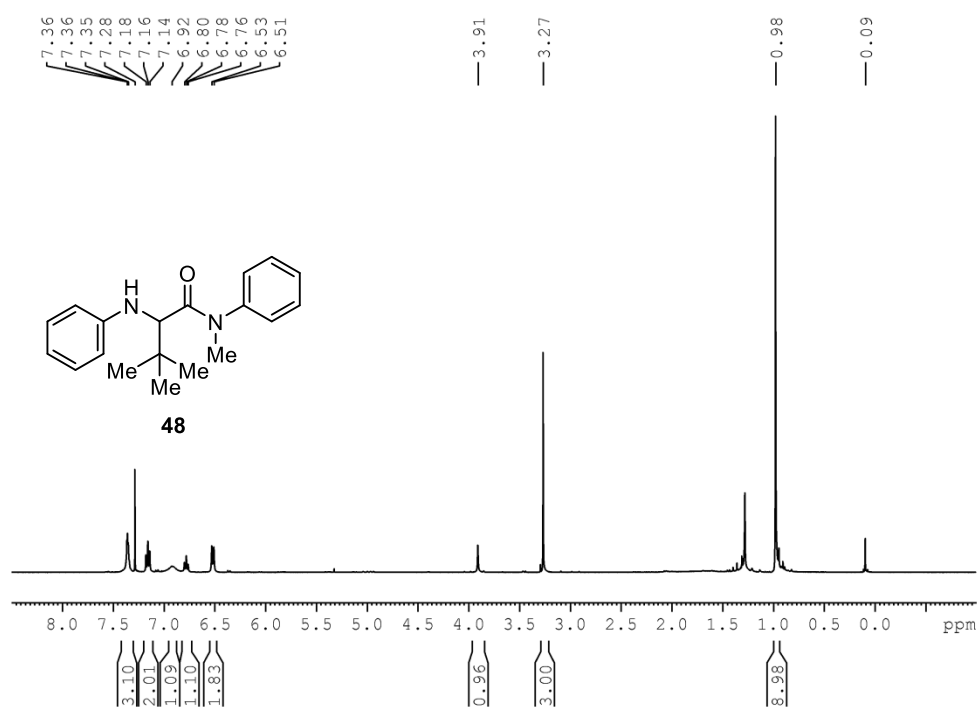
Supplementary Figure 80. ^{13}C NMR spectrum of compound 46



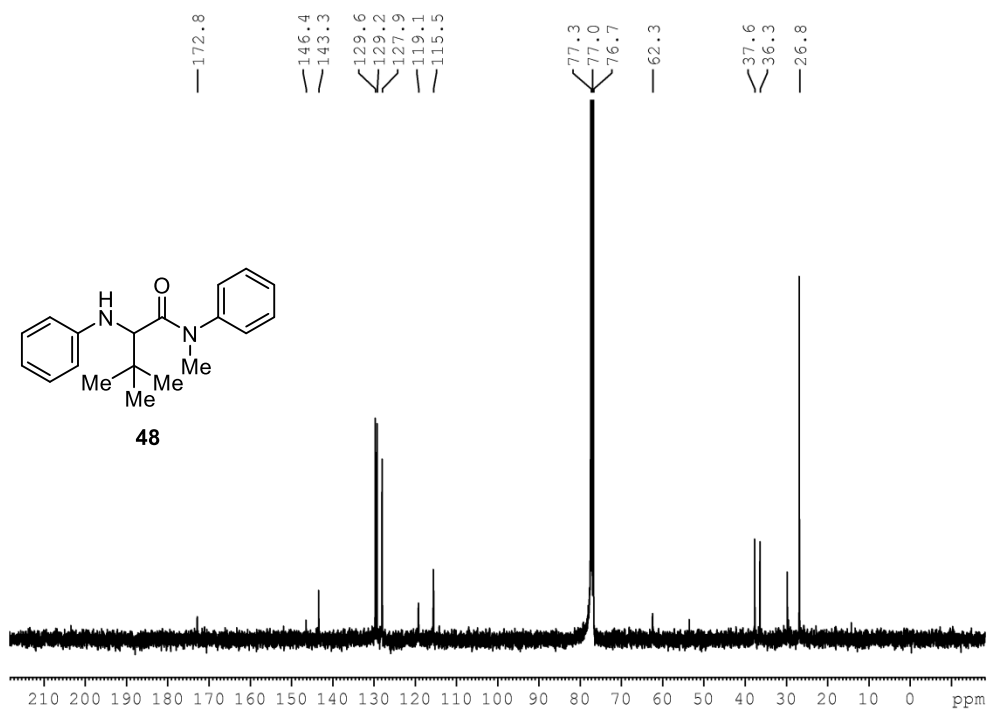
Supplementary Figure 81. ^1H NMR spectrum of compound 47



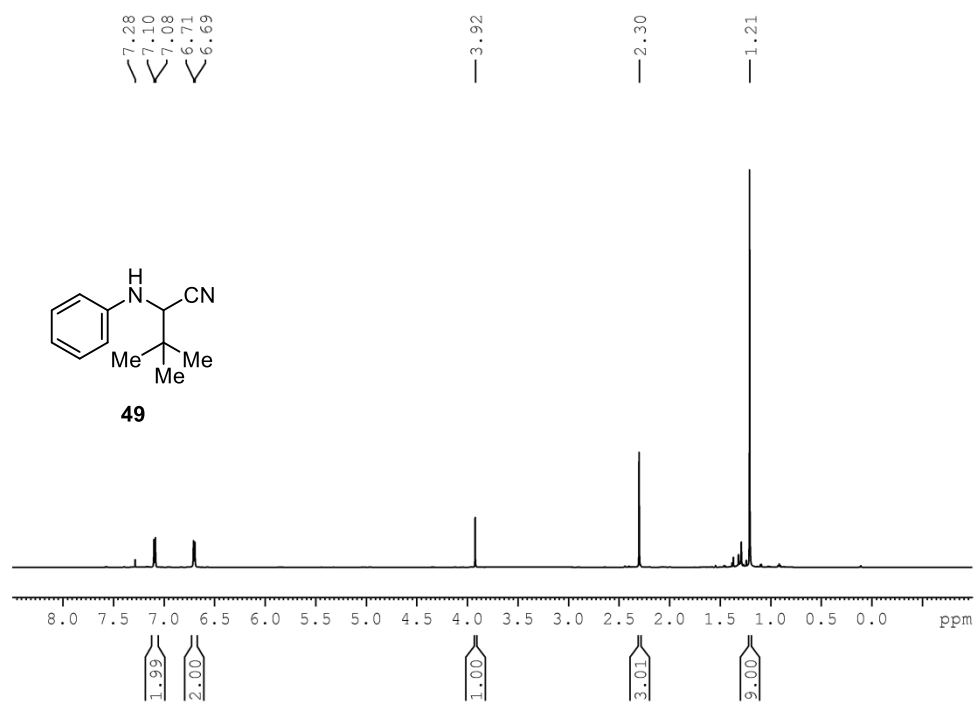
Supplementary Figure 82. ^{13}C NMR spectrum of compound 47



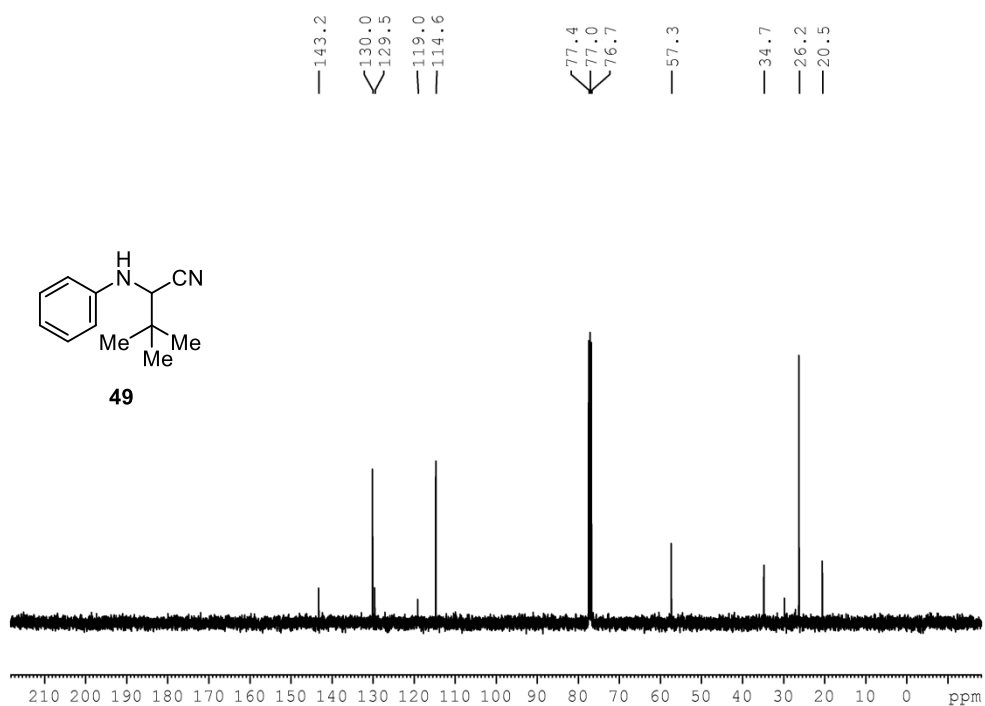
Supplementary Figure 83. ¹H NMR spectrum of compound 48



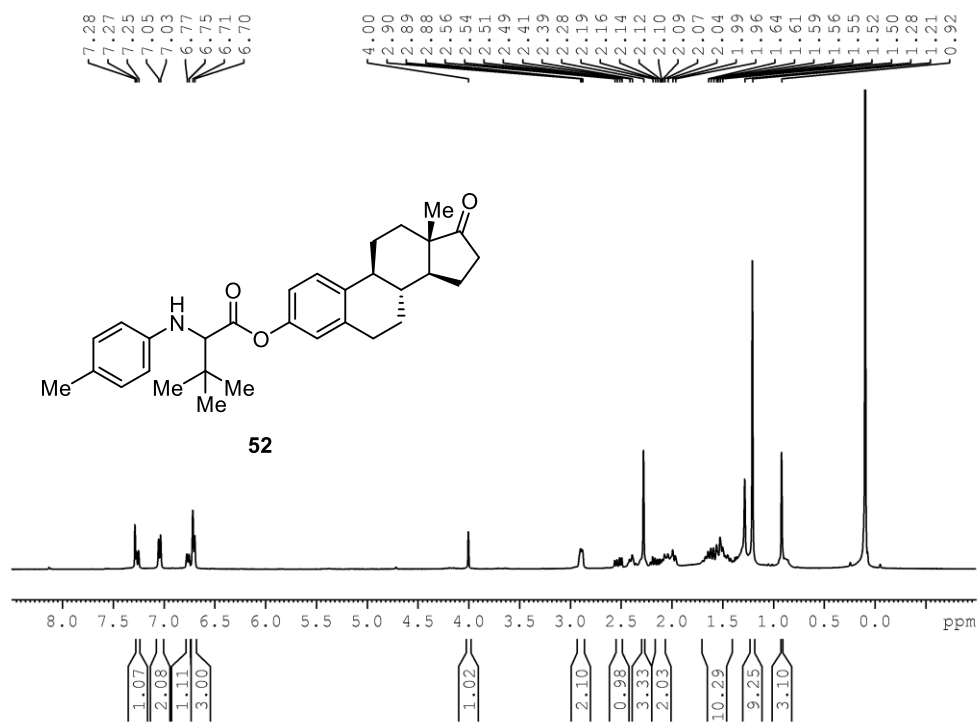
Supplementary Figure 84. ¹³C NMR spectrum of compound 48



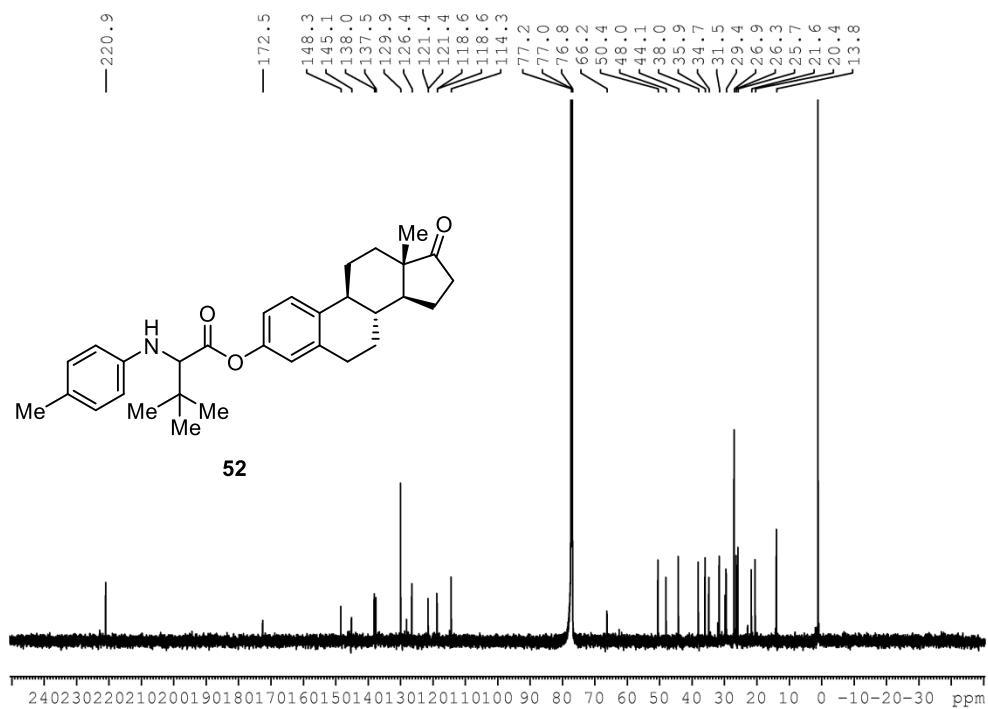
Supplementary Figure 85. ¹H NMR spectrum of compound **49**



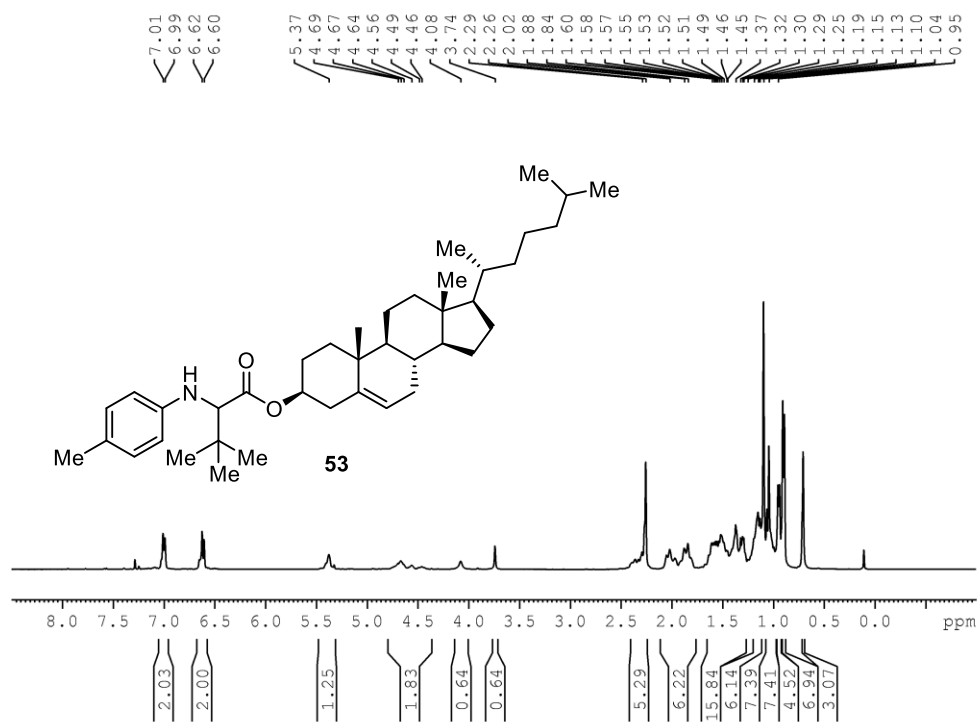
Supplementary Figure 86. ¹³C NMR spectrum of compound **49**



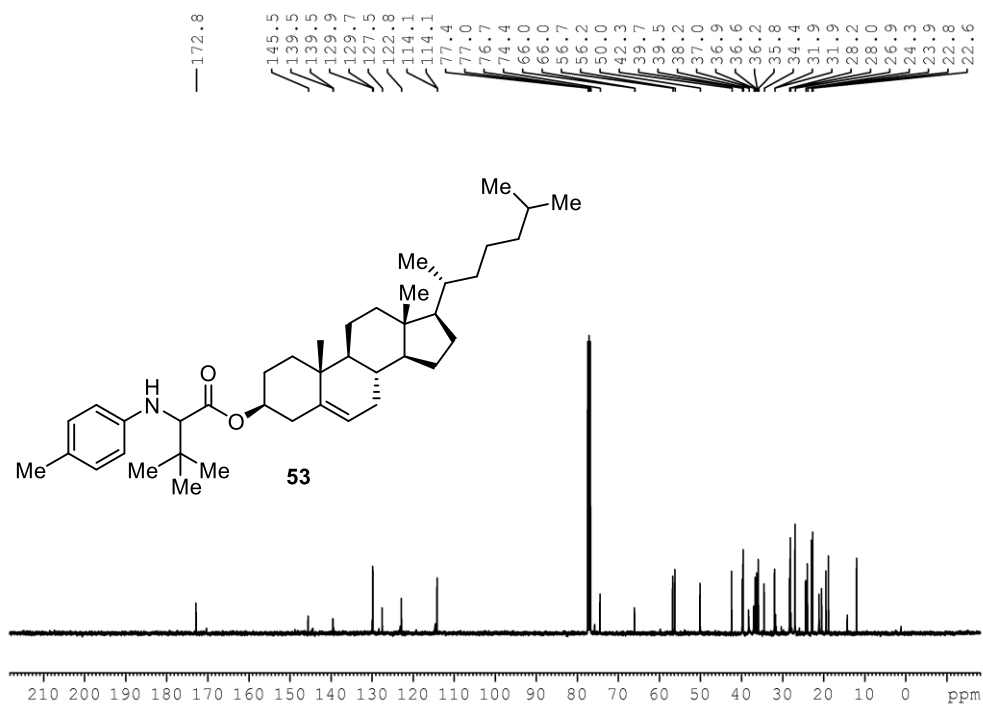
Supplementary Figure 87. ¹H NMR spectrum of compound 52



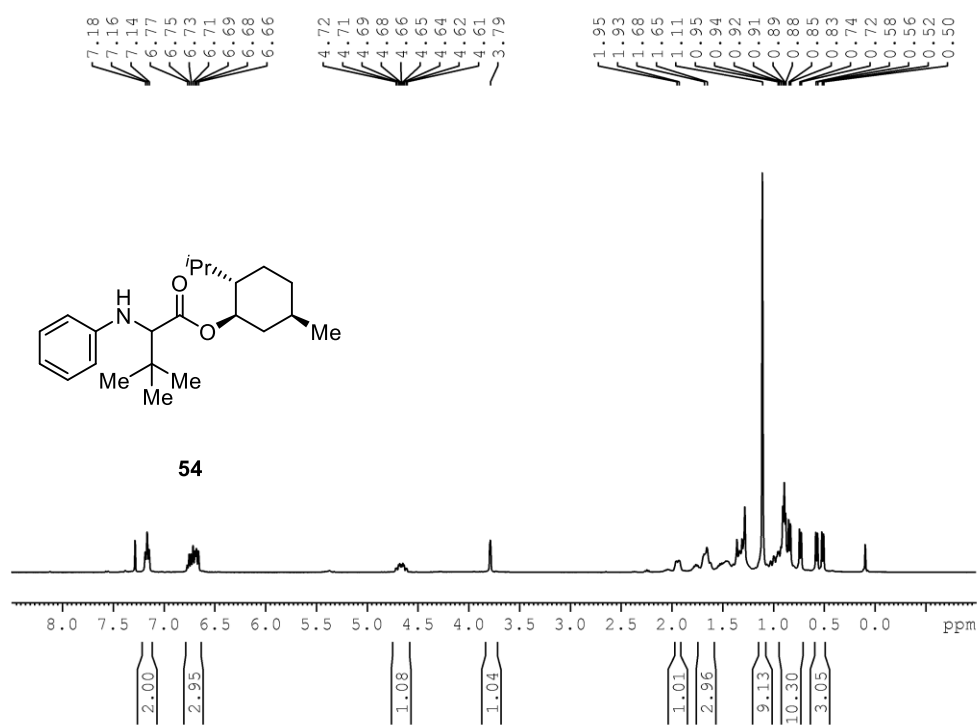
Supplementary Figure 88. ¹³C NMR spectrum of compound 52



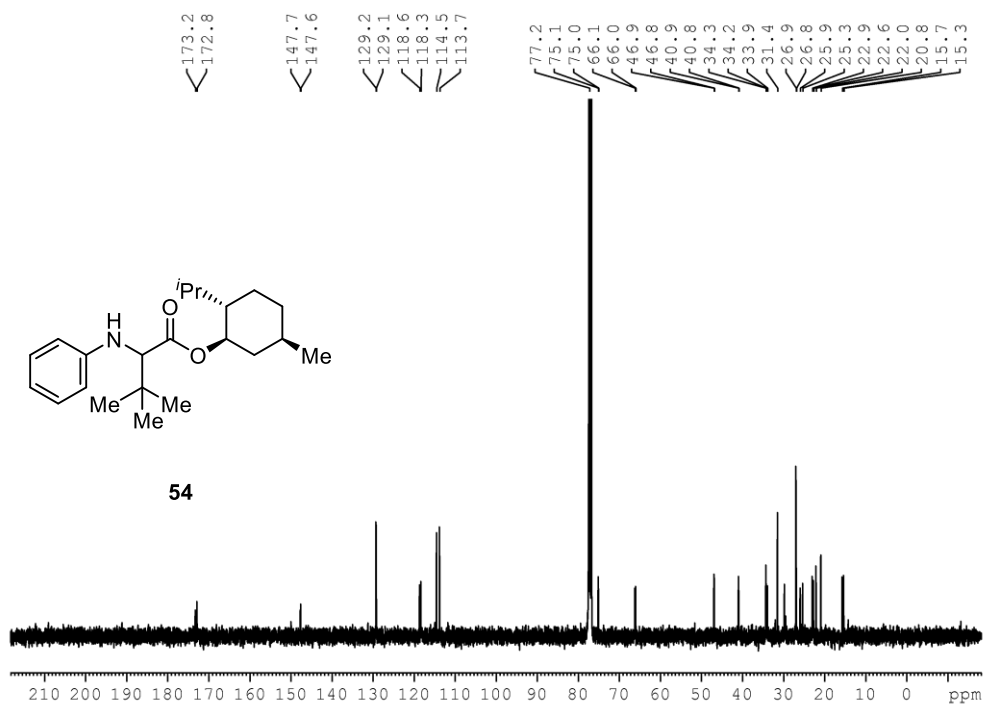
Supplementary Figure 89. ¹H NMR spectrum of compound 53



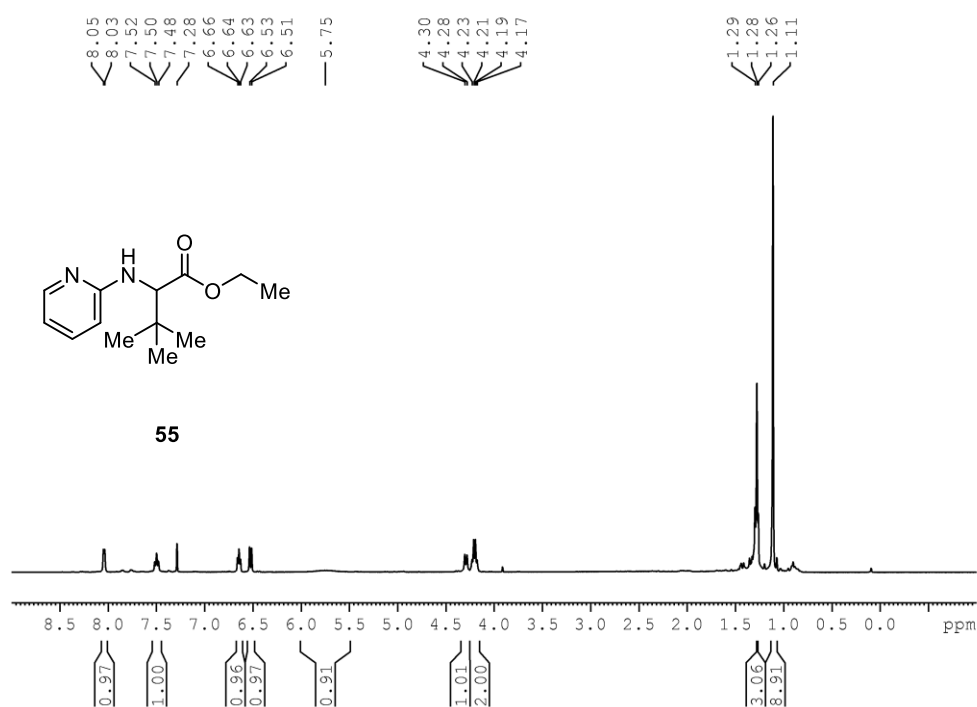
Supplementary Figure 90. ¹³C NMR spectrum of compound 53



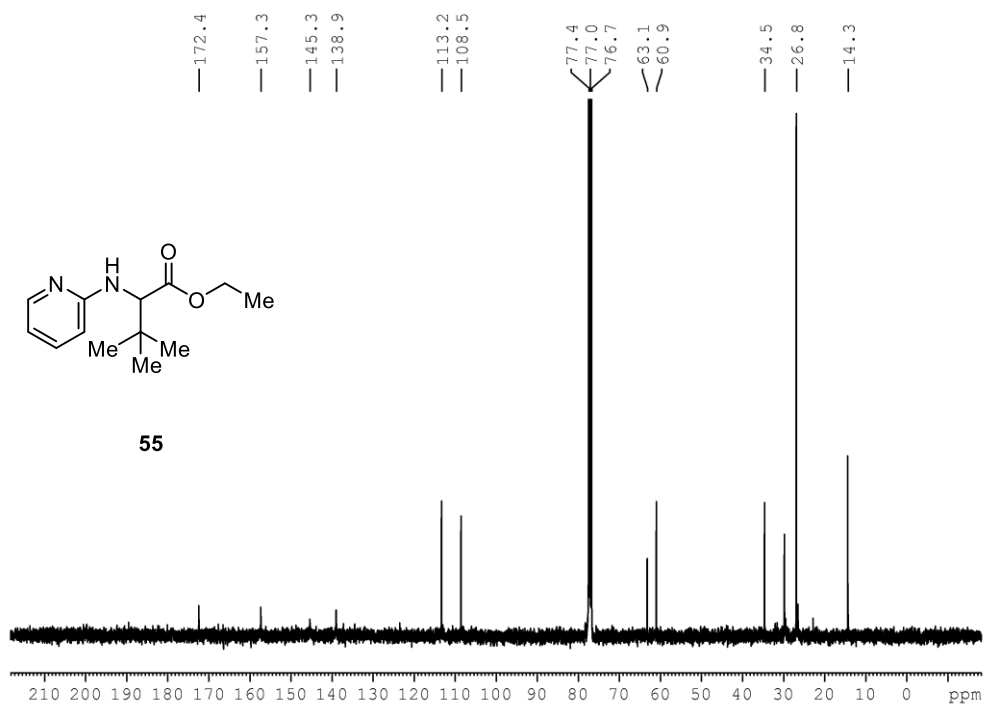
Supplementary Figure 91. ^1H NMR spectrum of compound 54



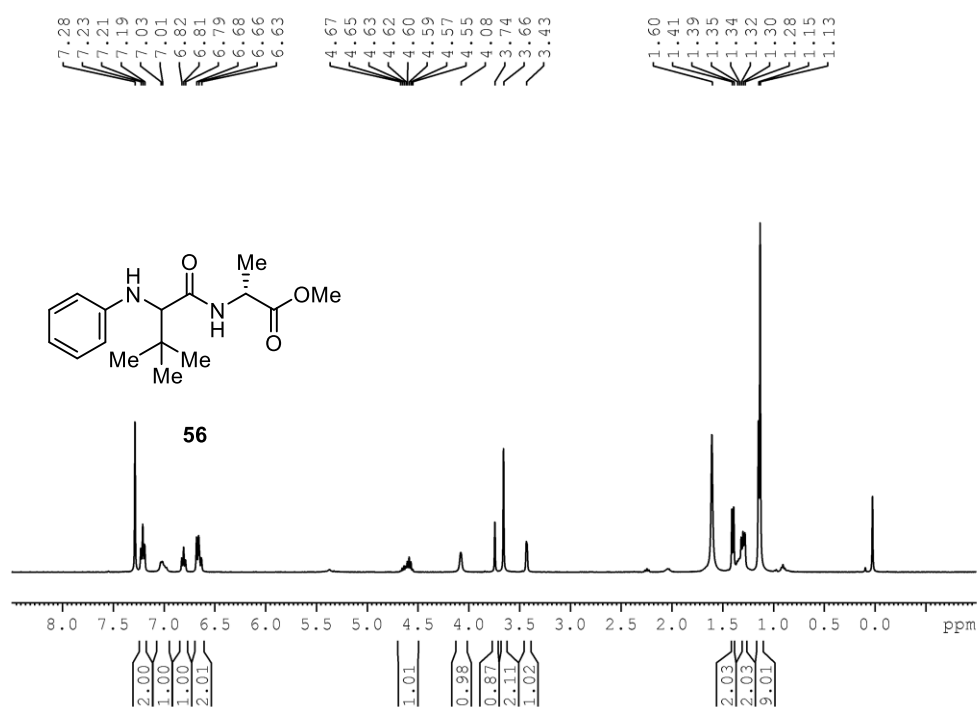
Supplementary Figure 92. ^{13}C NMR spectrum of compound 54



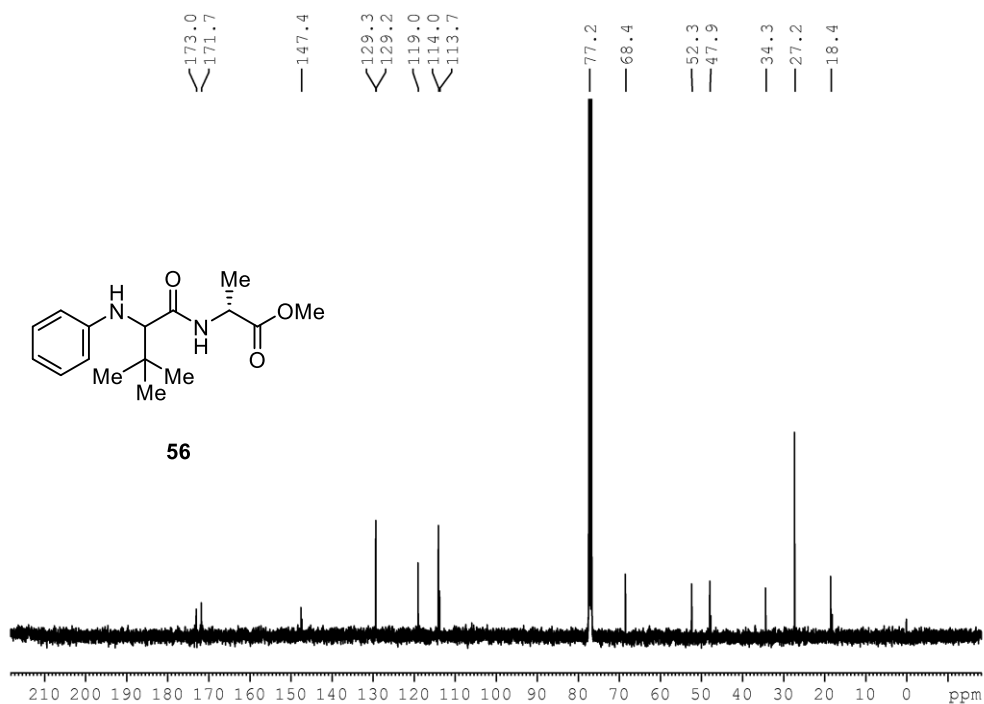
Supplementary Figure 93. ¹H NMR spectrum of compound 55



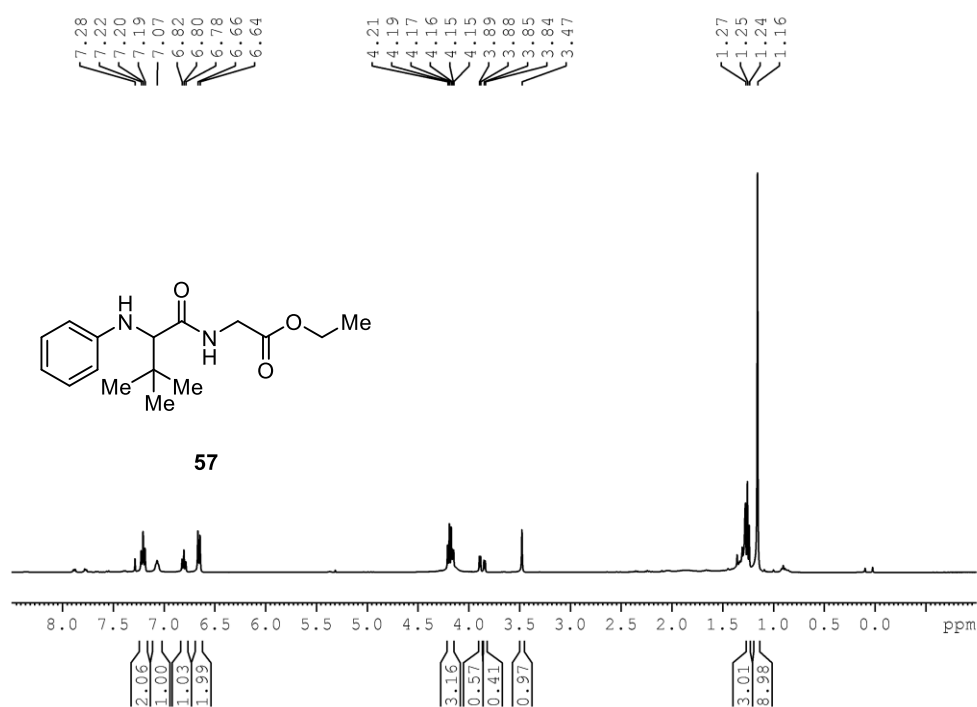
Supplementary Figure 94. ¹³C NMR spectrum of compound 55



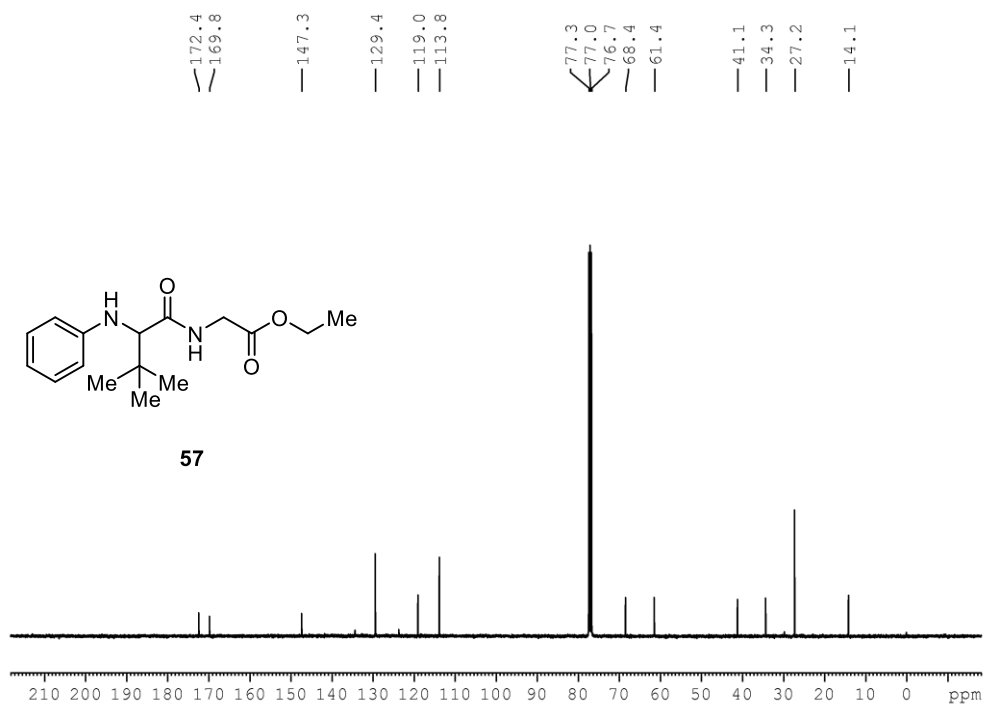
Supplementary Figure 95. ¹H NMR spectrum of compound 56



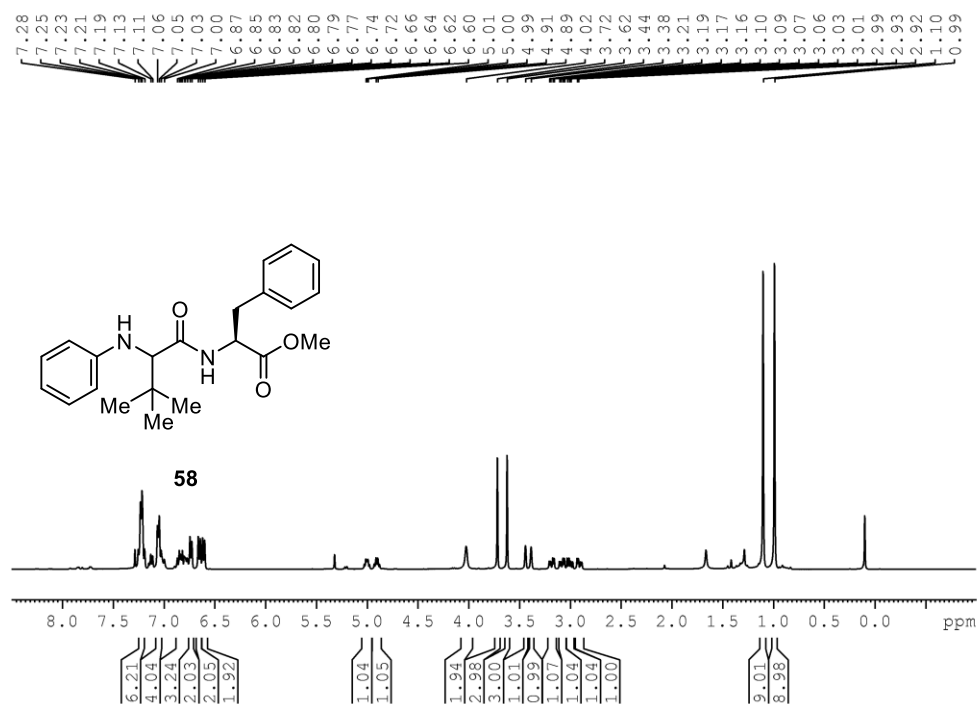
Supplementary Figure 96. ¹³C NMR spectrum of compound 56



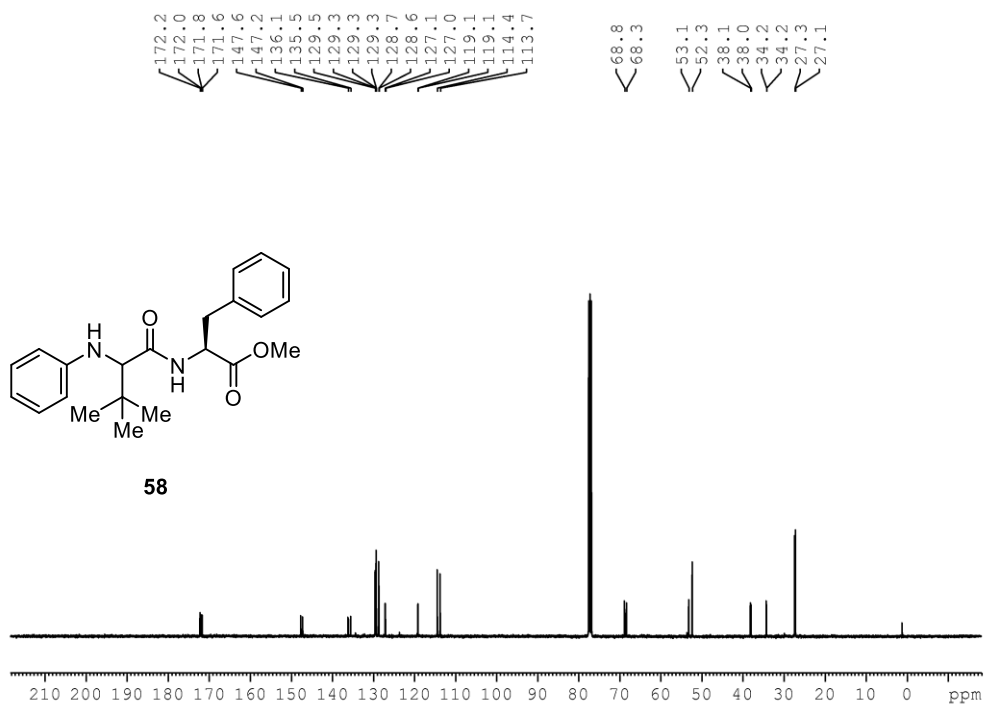
Supplementary Figure 97. ¹H NMR spectrum of compound 57



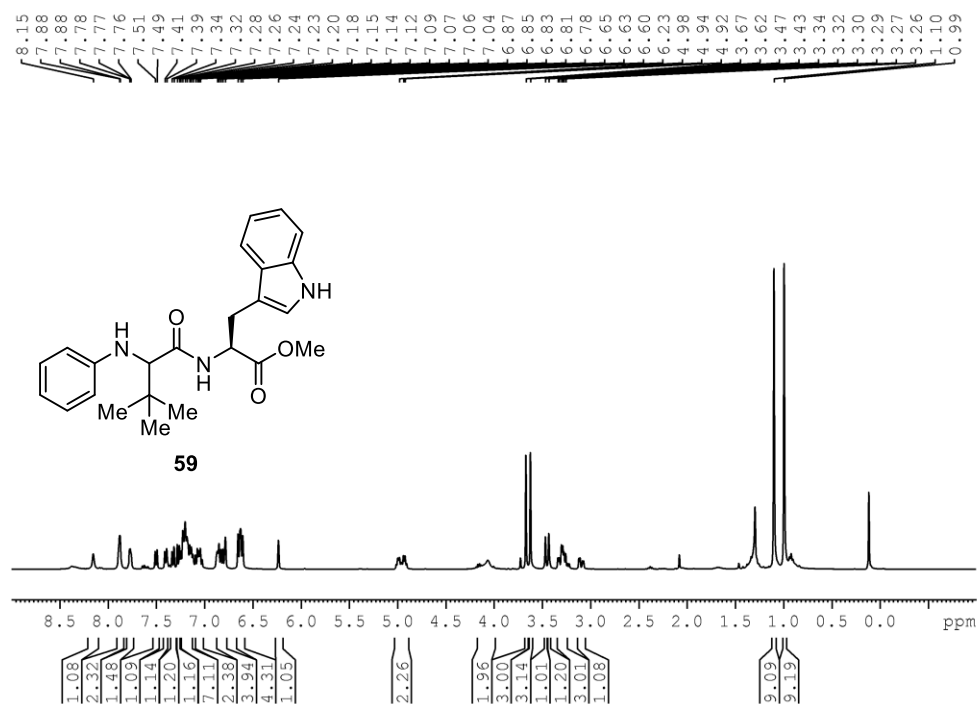
Supplementary Figure 98. ¹³C NMR spectrum of compound 57



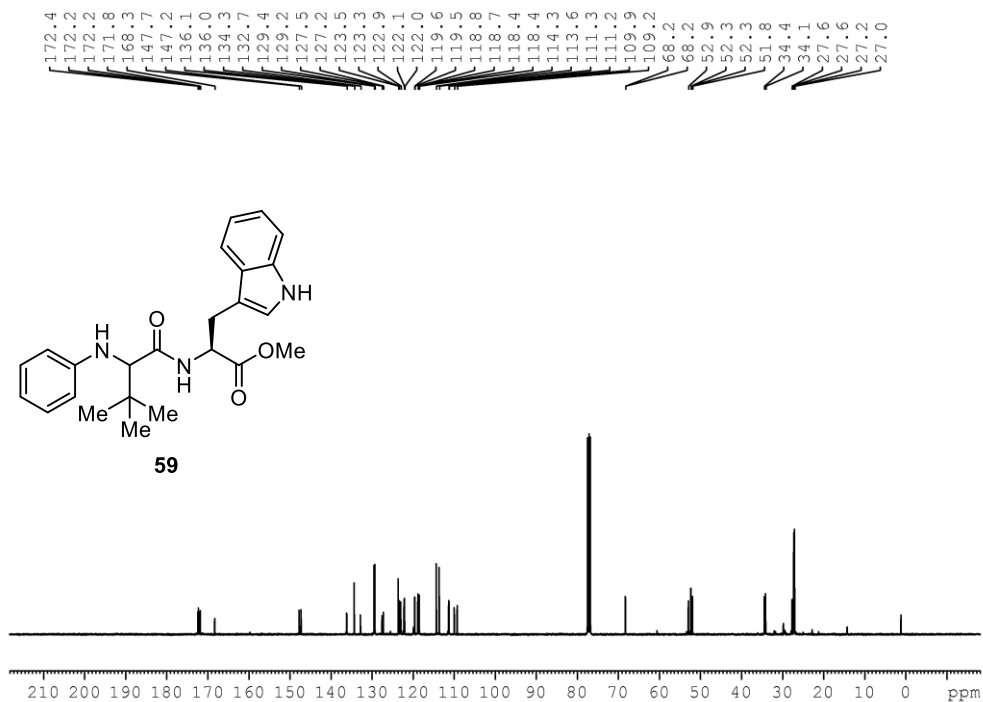
Supplementary Figure 99. ¹H NMR spectrum of compound 58



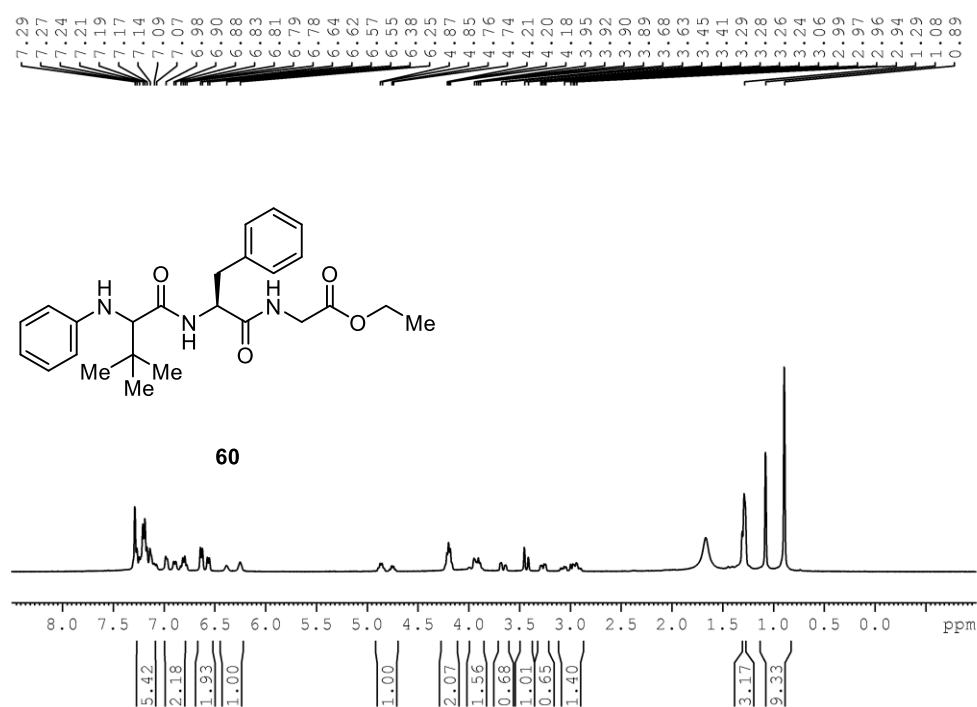
Supplementary Figure 100. ¹³C NMR spectrum of compound 58



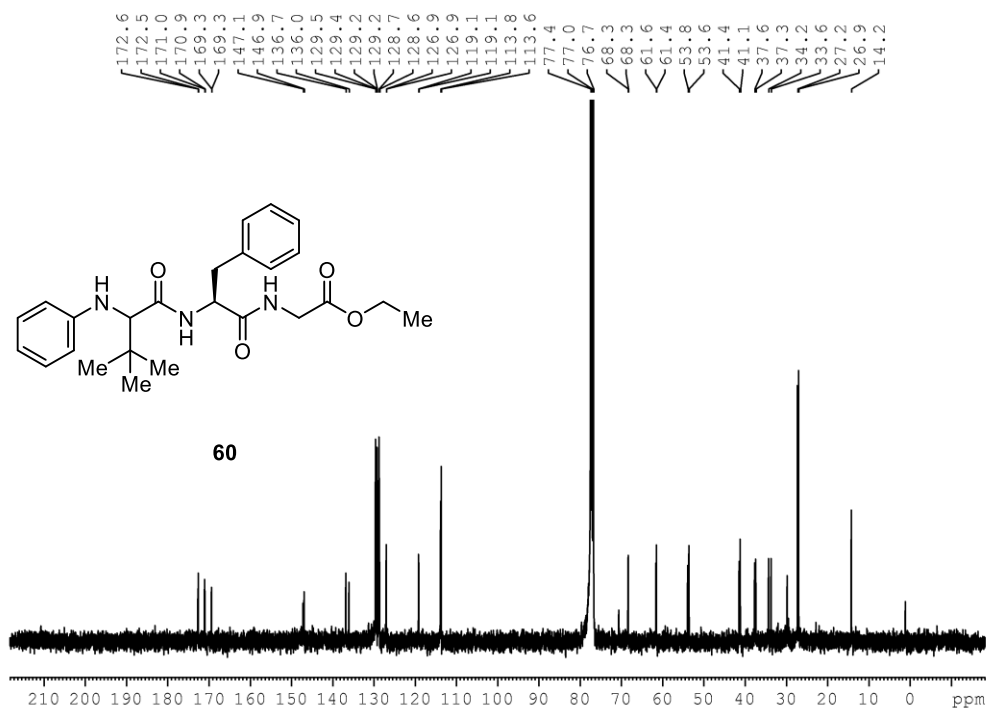
Supplementary Figure 101. ¹H NMR spectrum of compound 59



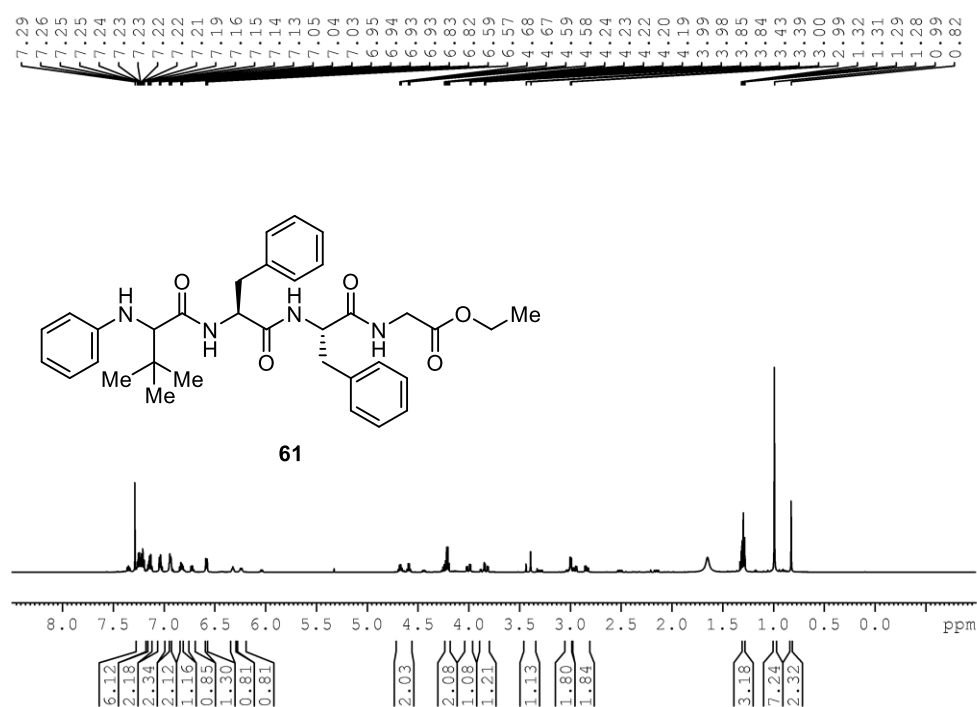
Supplementary Figure 102. ¹³C NMR spectrum of compound 59



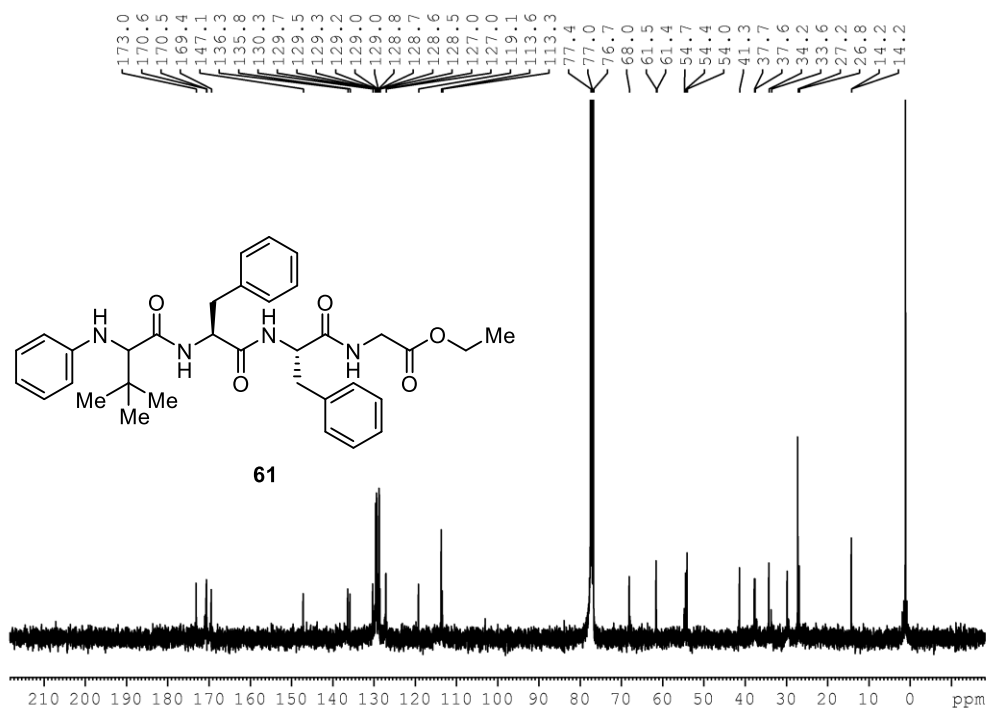
Supplementary Figure 103. ¹H NMR spectrum of compound 60



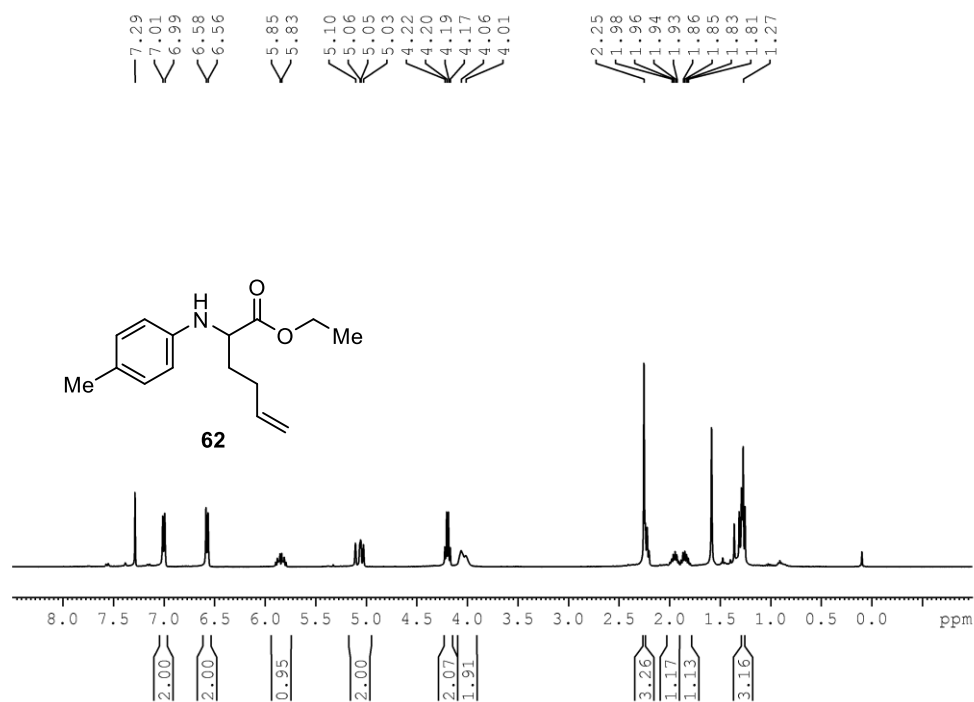
Supplementary Figure 104. ¹³C NMR spectrum of compound 60



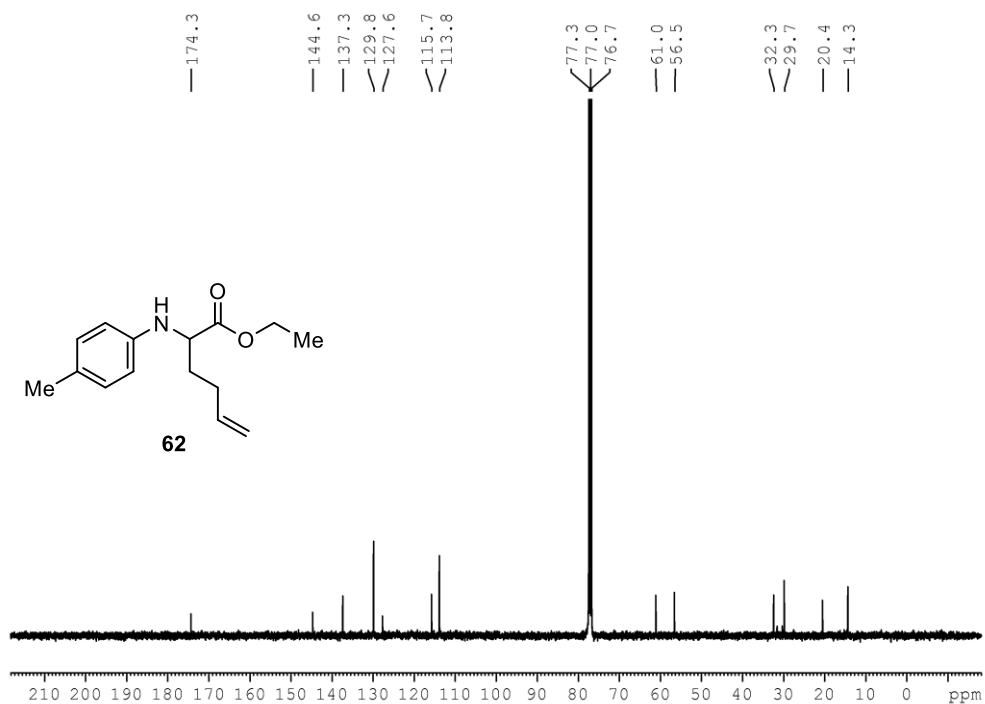
Supplementary Figure 105. ¹H NMR spectrum of compound 61



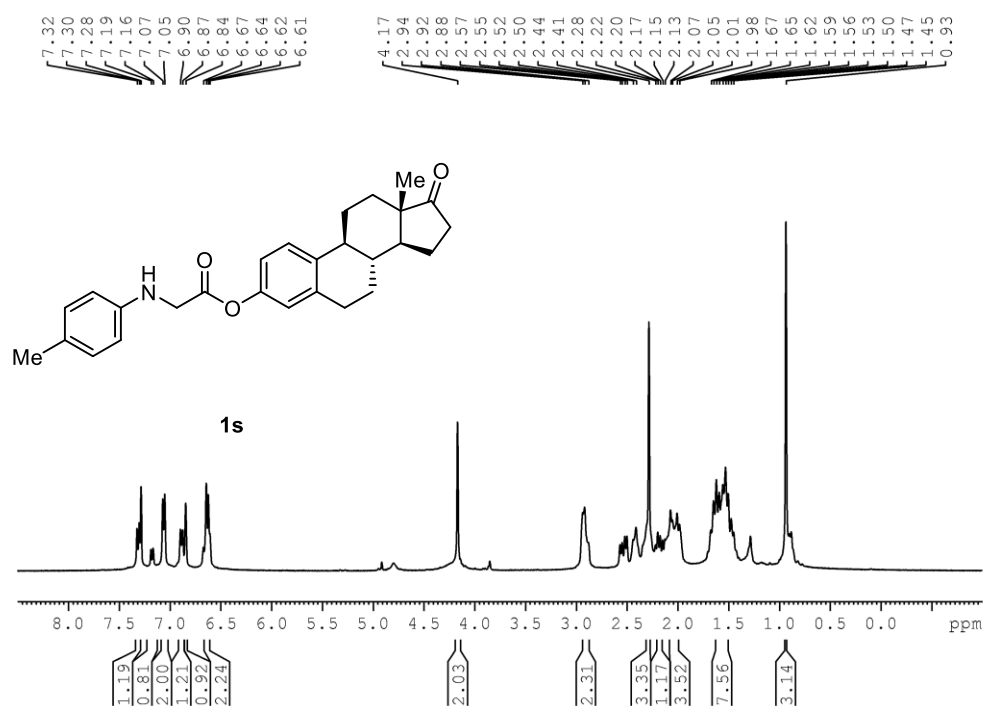
Supplementary Figure 106. ¹³C NMR spectrum of compound 61



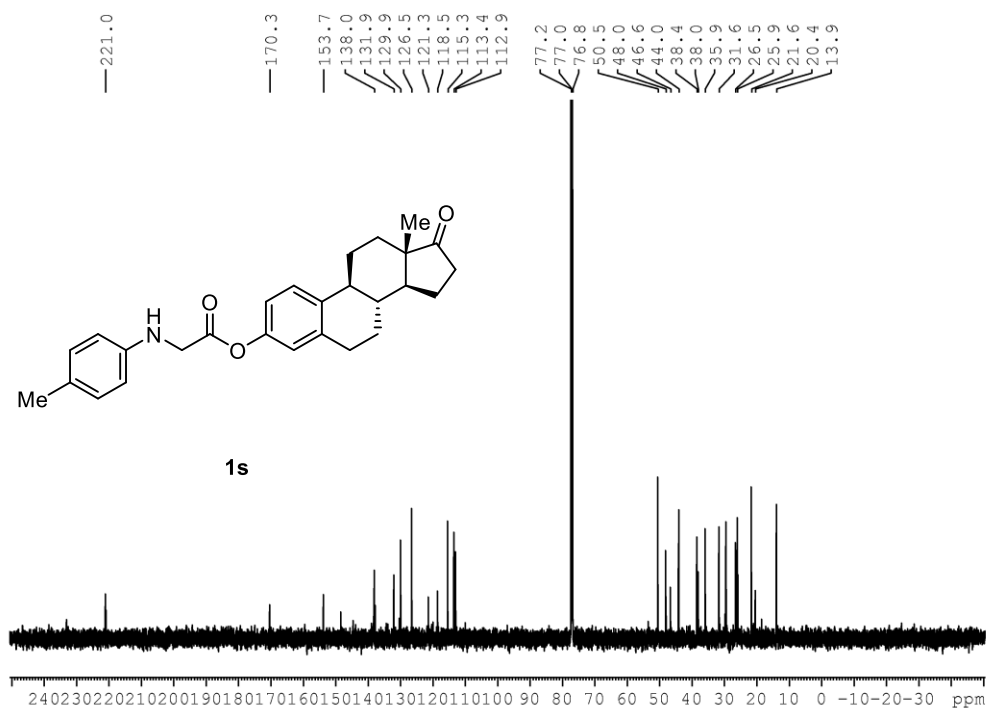
Supplementary Figure 107. ¹H NMR spectrum of compound 62



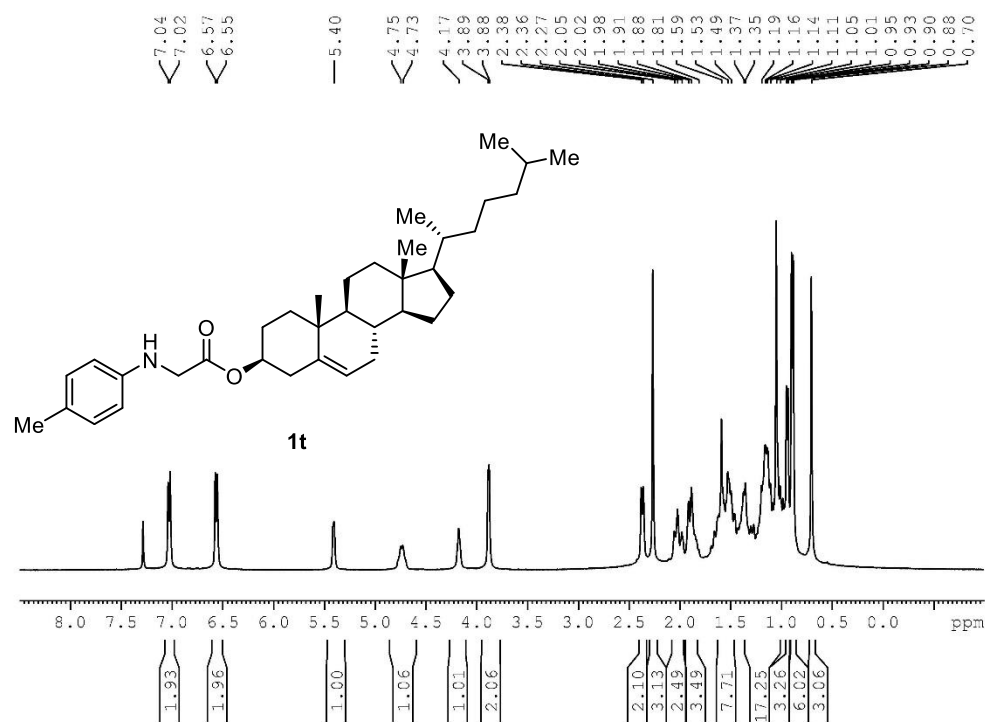
Supplementary Figure 108. ¹³C NMR spectrum of compound 62



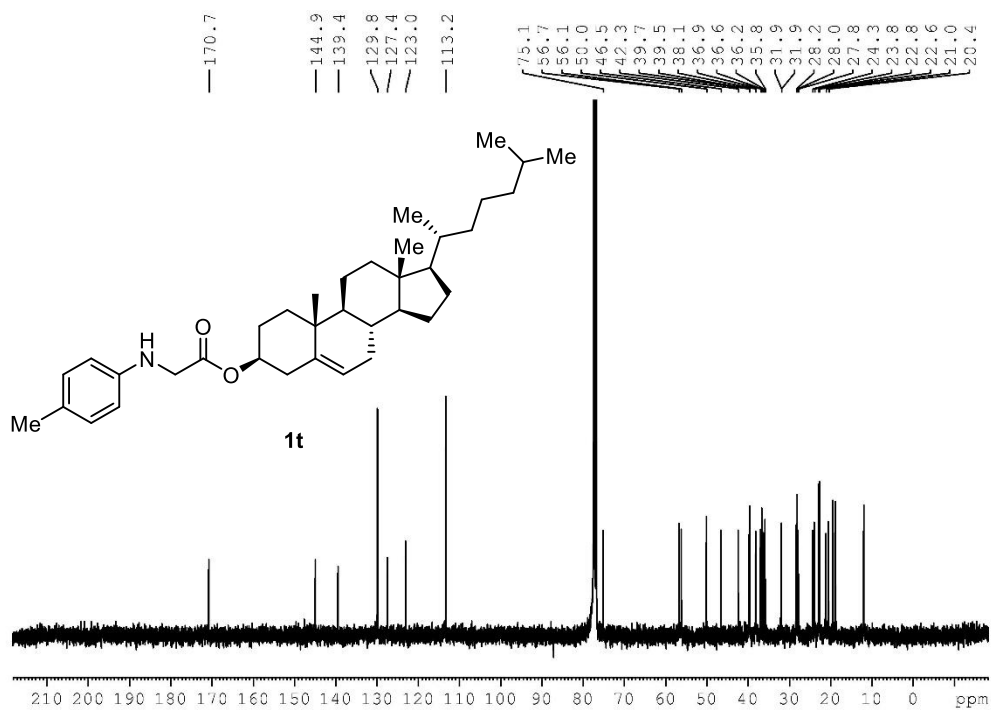
Supplementary Figure 109. ¹H NMR spectrum of compound 1s



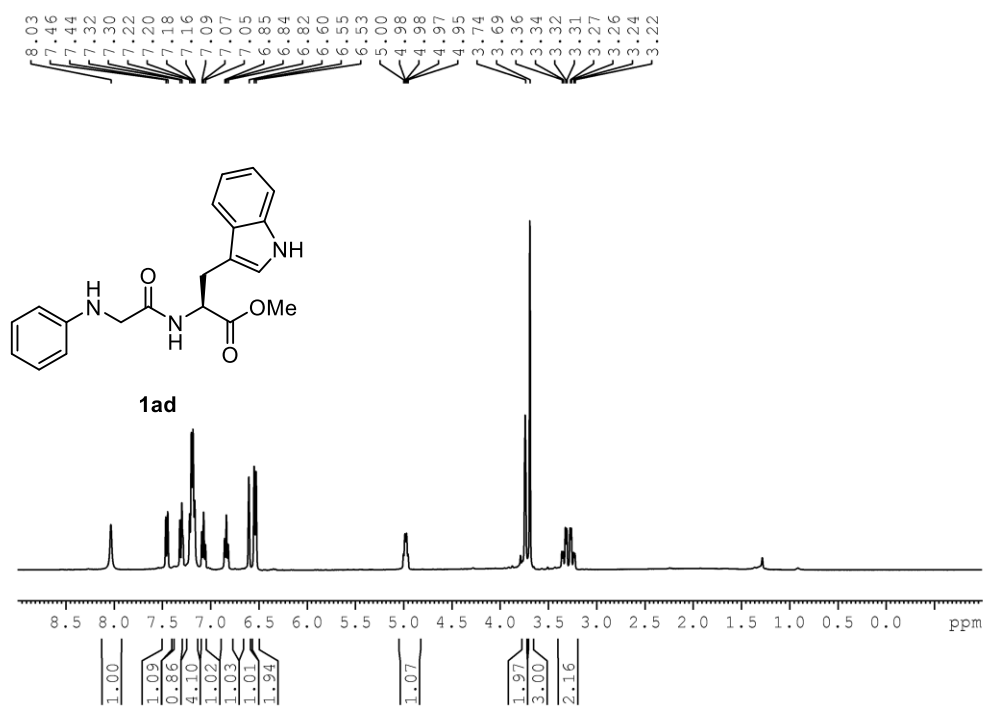
Supplementary Figure 110. ¹³C NMR spectrum of compound 1s



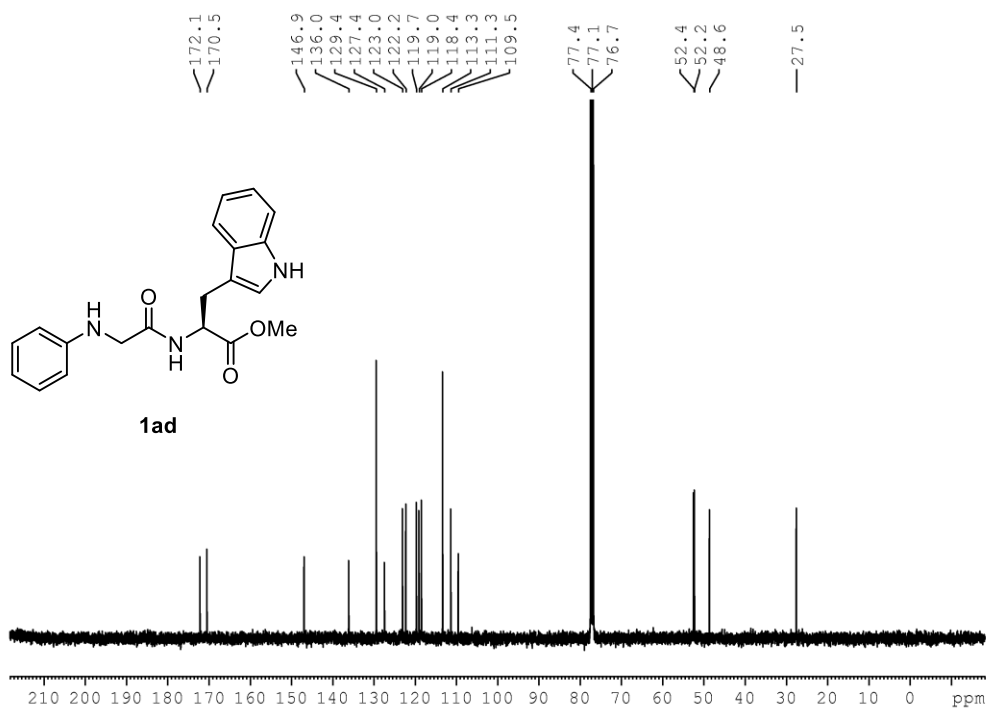
Supplementary Figure 111. ¹H NMR spectrum of compound 1t



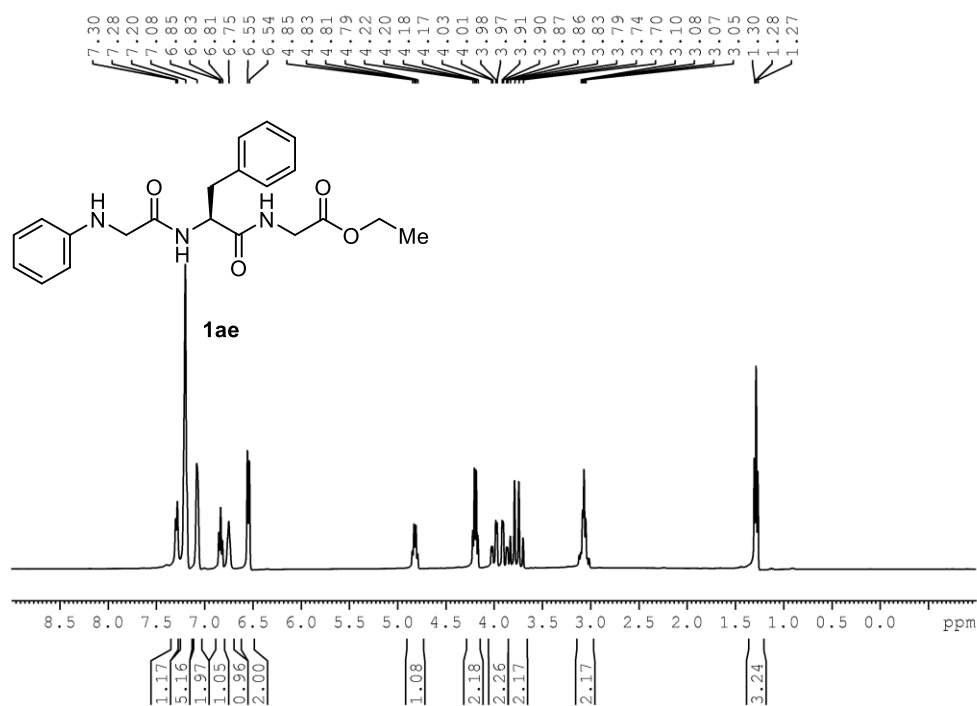
Supplementary Figure 112. ¹³C NMR spectrum of compound 1t



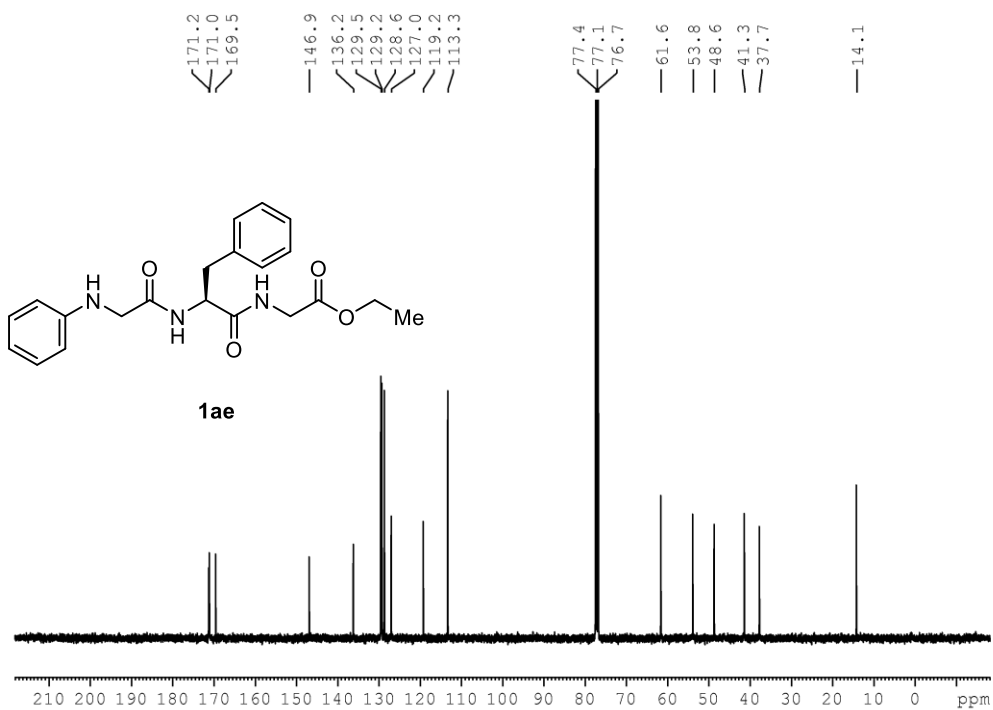
Supplementary Figure 113. ¹H NMR spectrum of compound 1ad



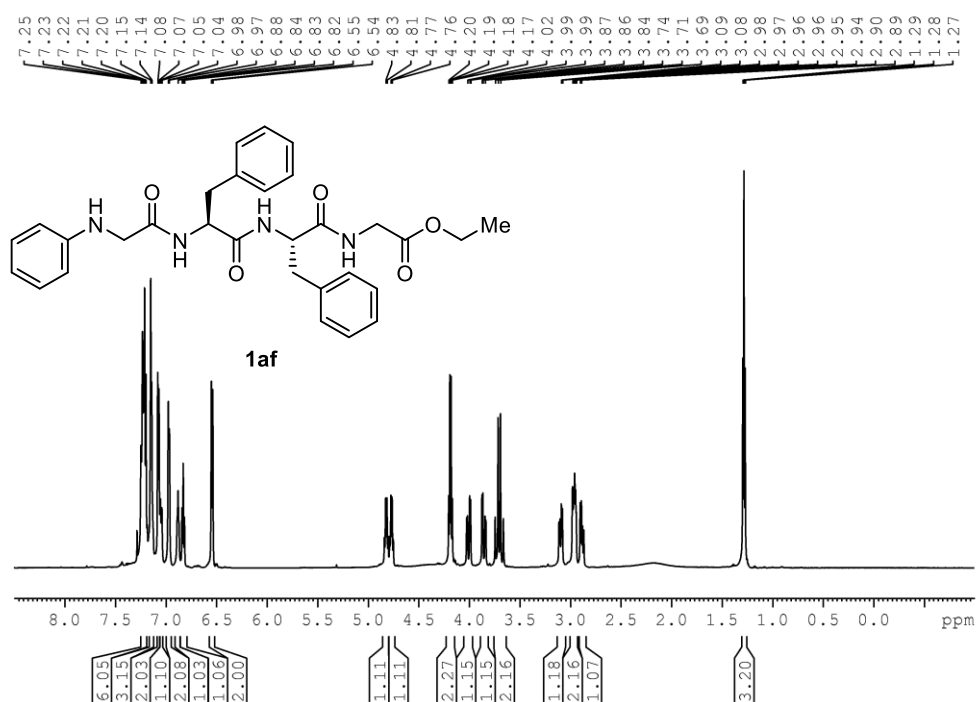
Supplementary Figure 114. ¹³C NMR spectrum of compound 1ad



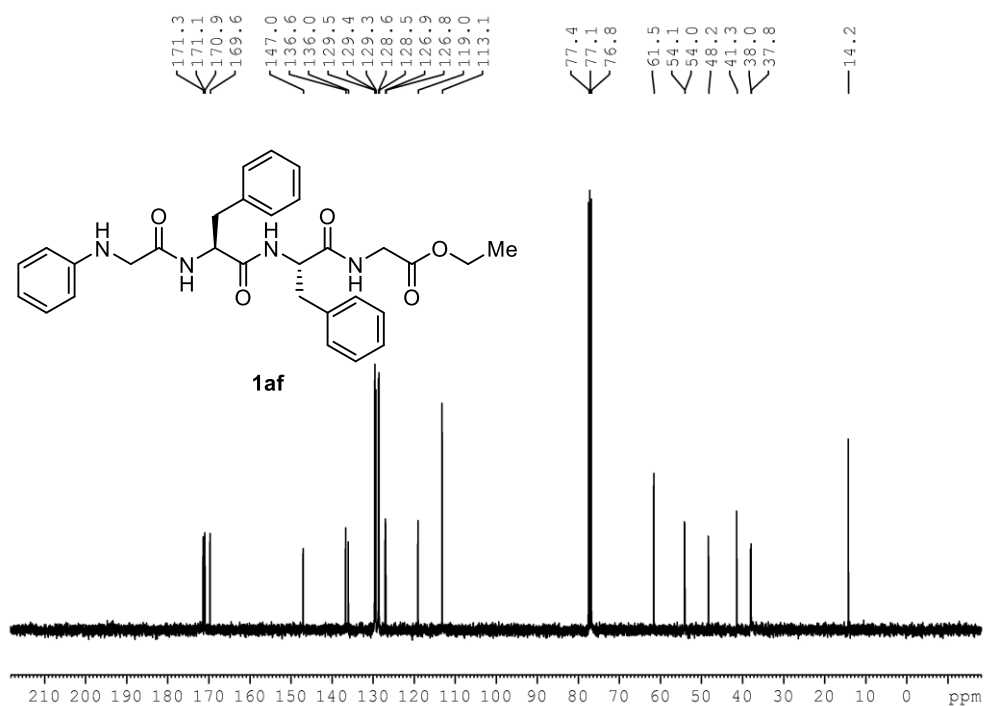
Supplementary Figure 115. ¹H NMR spectrum of compound 1ae



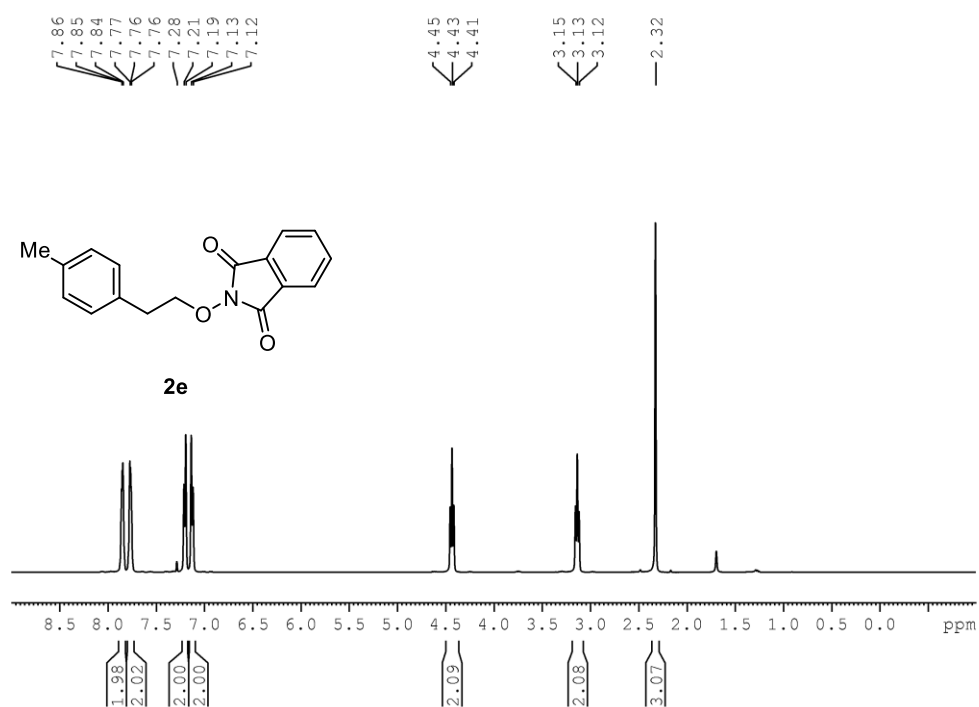
Supplementary Figure 116. ¹³C NMR spectrum of compound 1ae



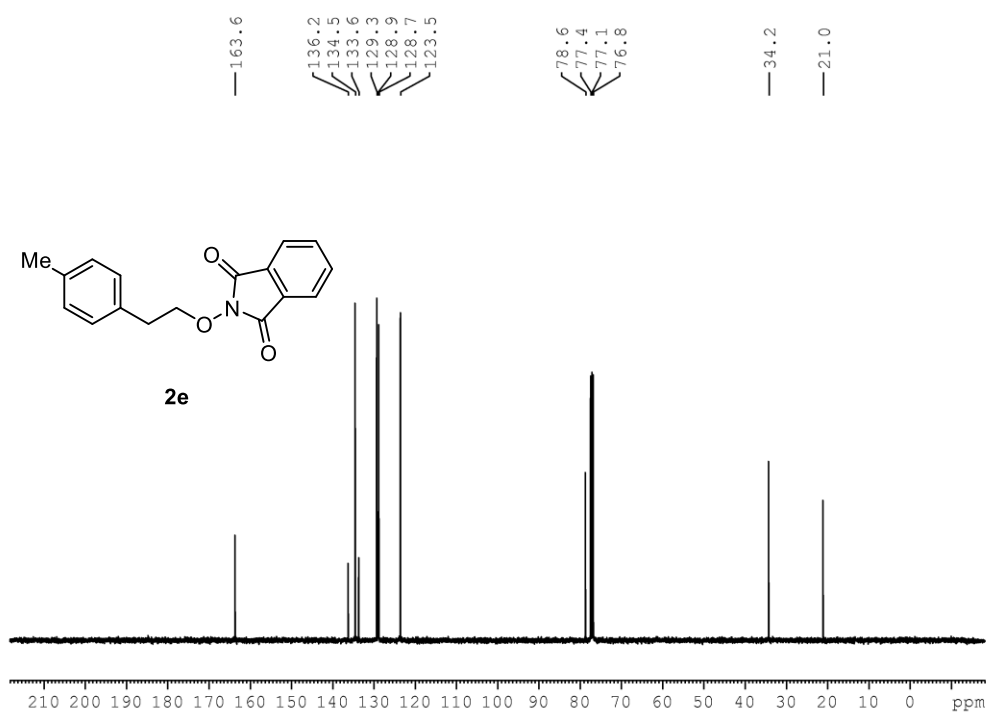
Supplementary Figure 117. ¹H NMR spectrum of compound 1af



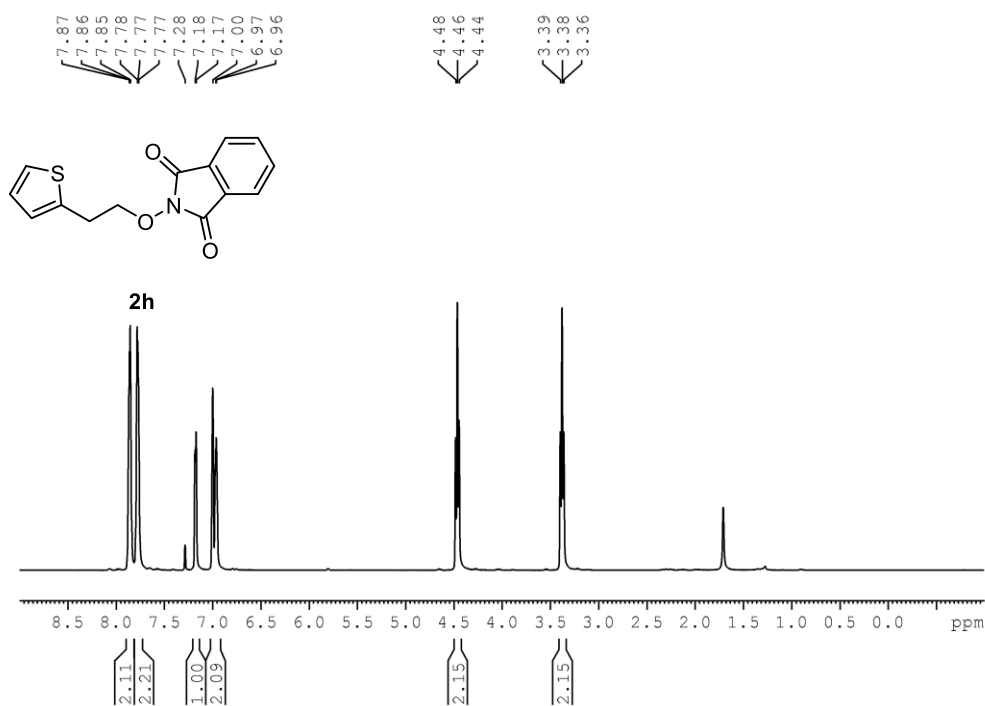
Supplementary Figure 118. ¹³C NMR spectrum of compound 1af



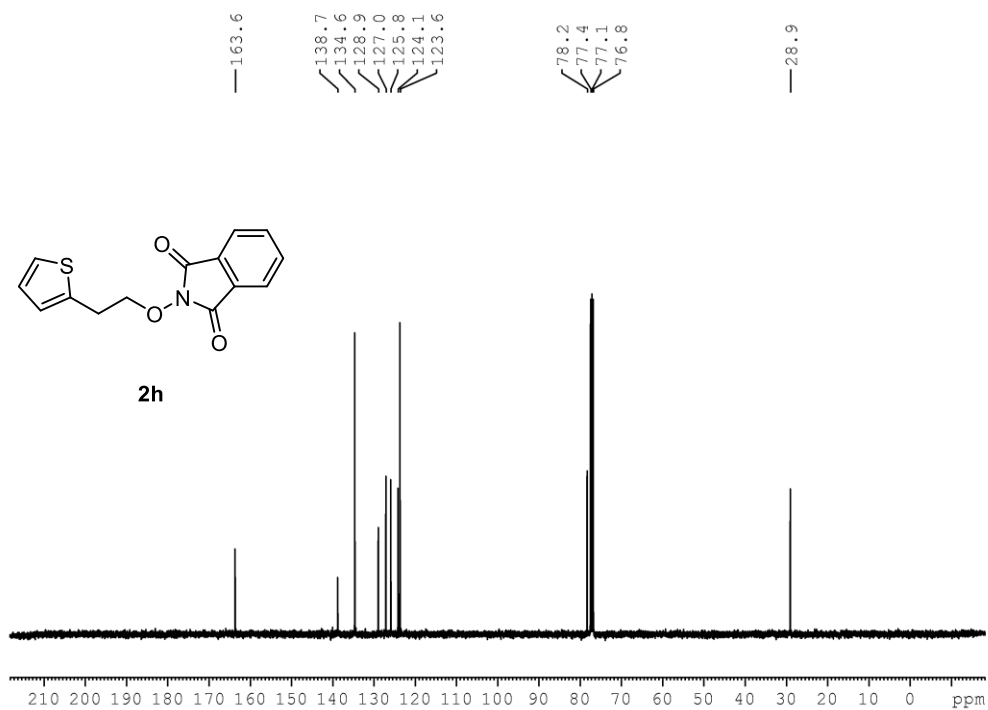
Supplementary Figure 119. ¹H NMR spectrum of compound **2e**



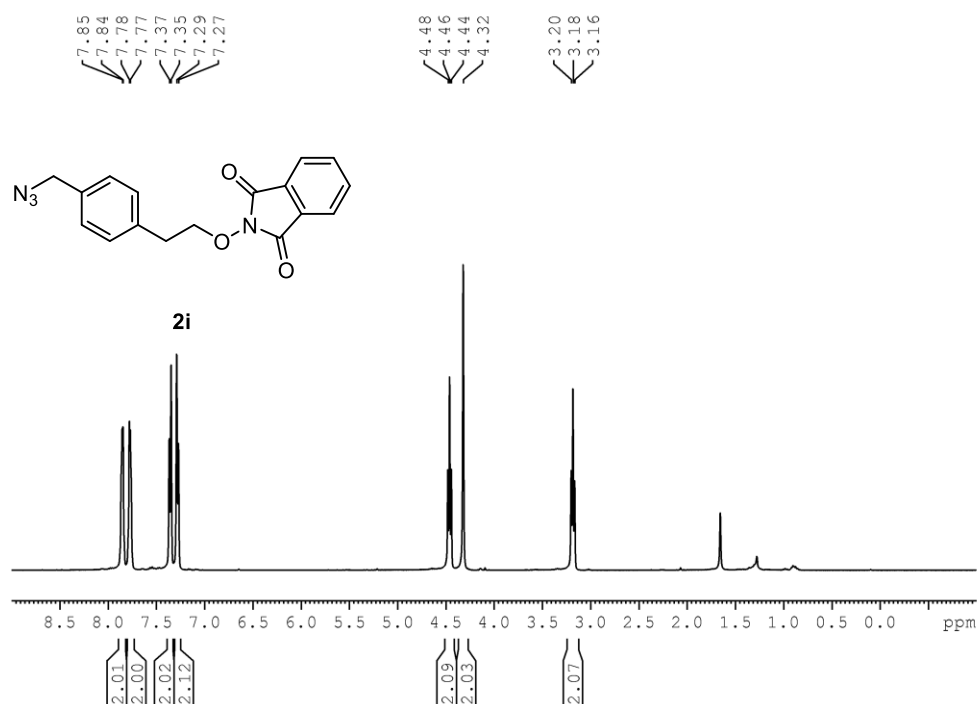
Supplementary Figure 120. ¹³C NMR spectrum of compound **2e**



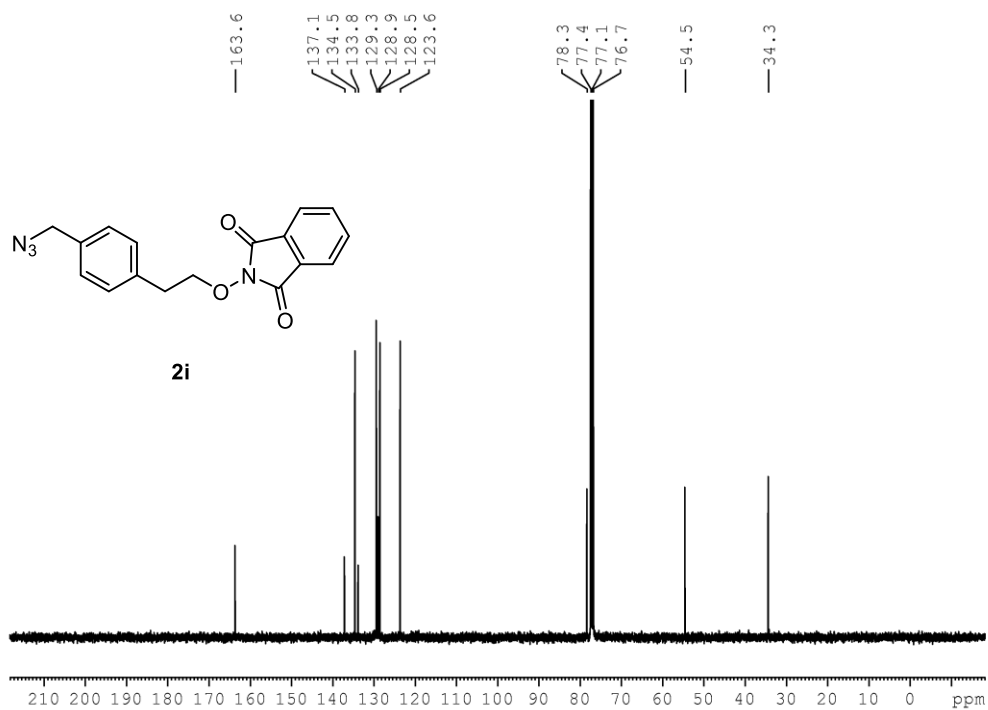
Supplementary Figure 121. ¹H NMR spectrum of compound 2h



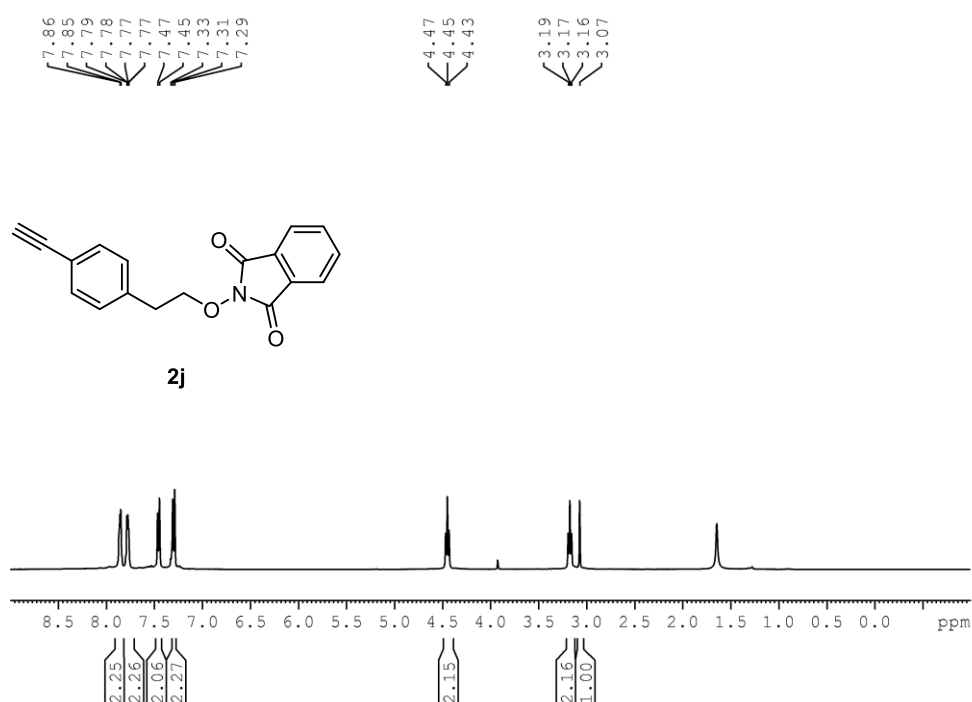
Supplementary Figure 122. ¹³C NMR spectrum of compound 2h



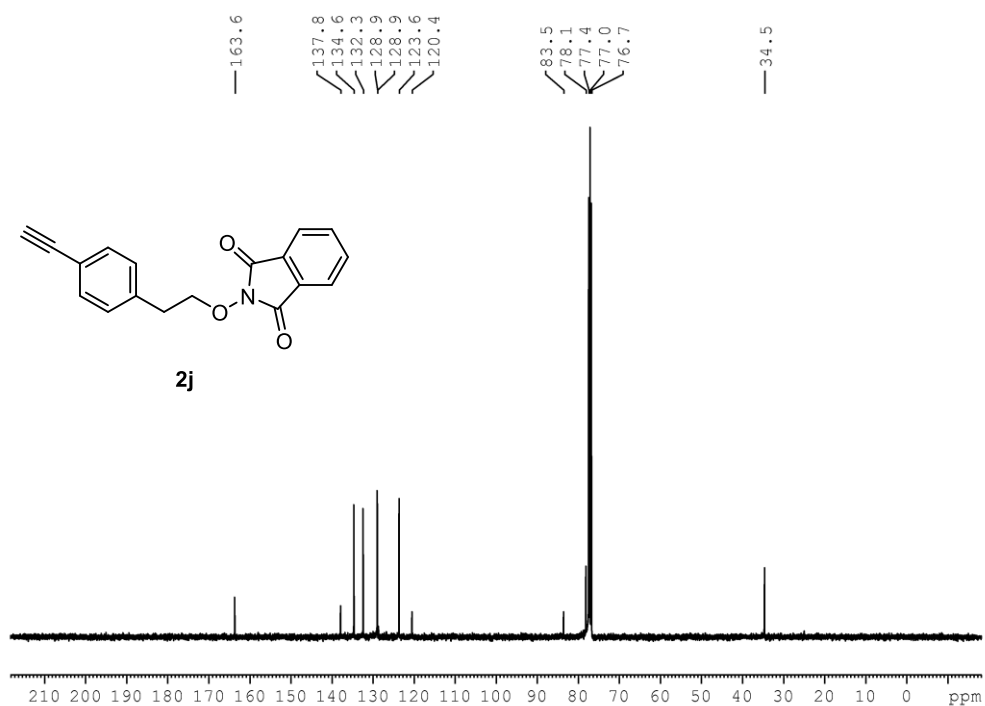
Supplementary Figure 123. ¹H NMR spectrum of compound 2i



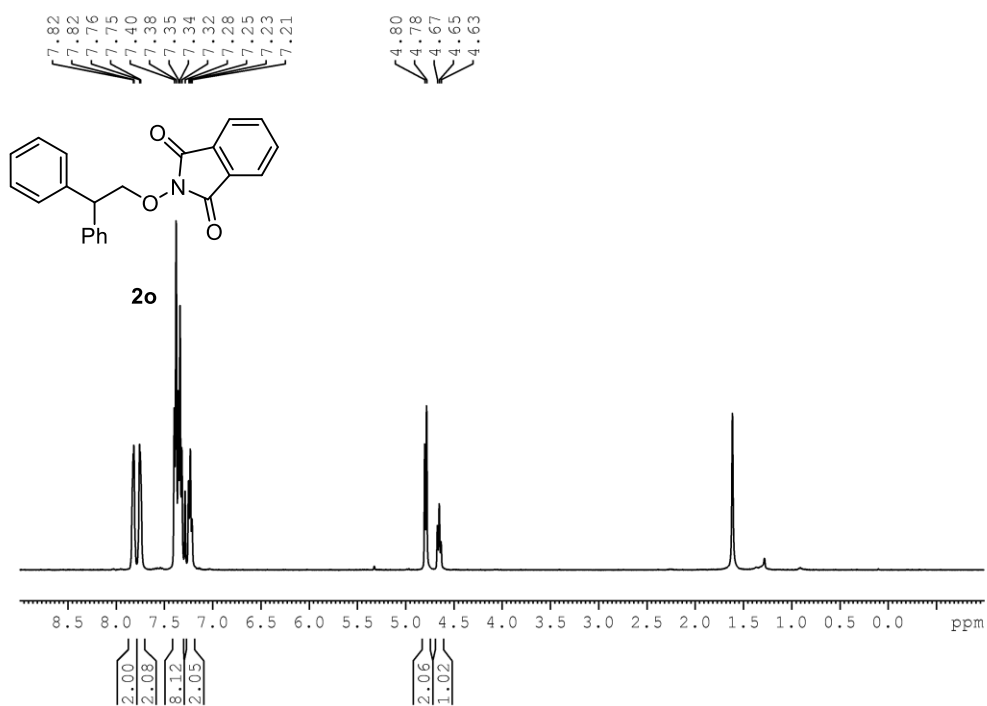
Supplementary Figure 124. ¹³C NMR spectrum of compound 2i



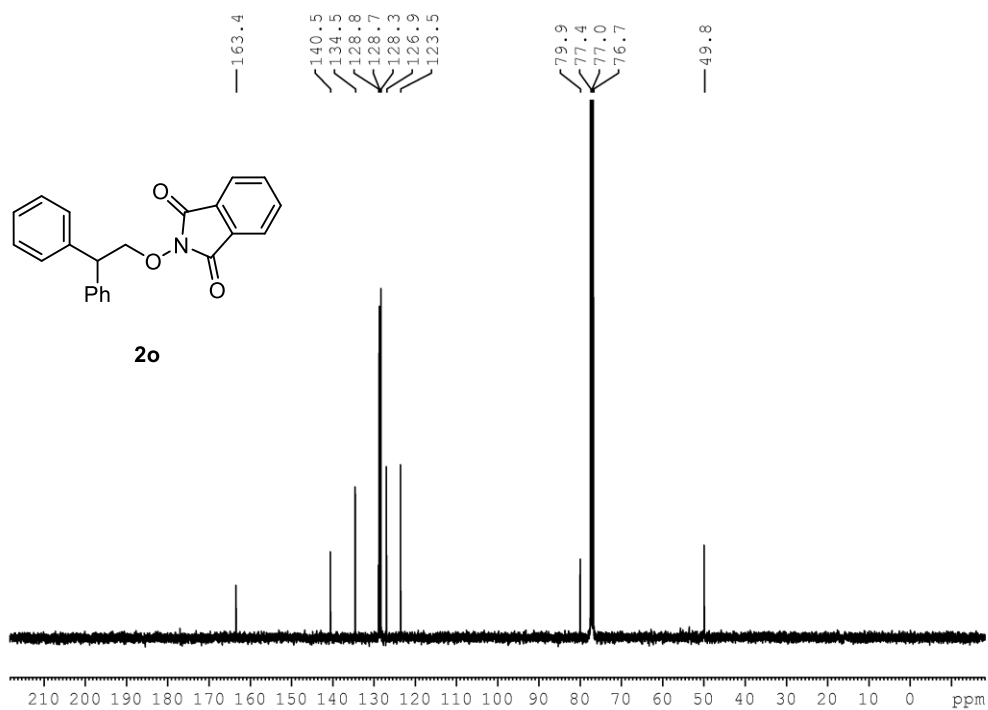
Supplementary Figure 125. ^1H NMR spectrum of compound **2j**



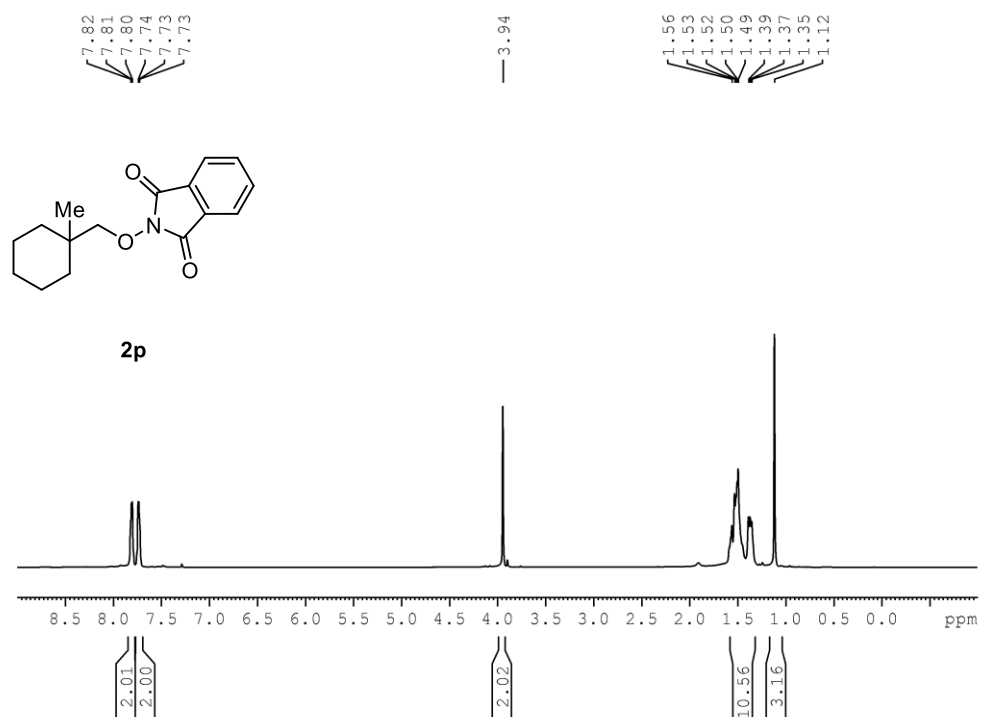
Supplementary Figure 126. ^{13}C NMR spectrum of compound **2j**



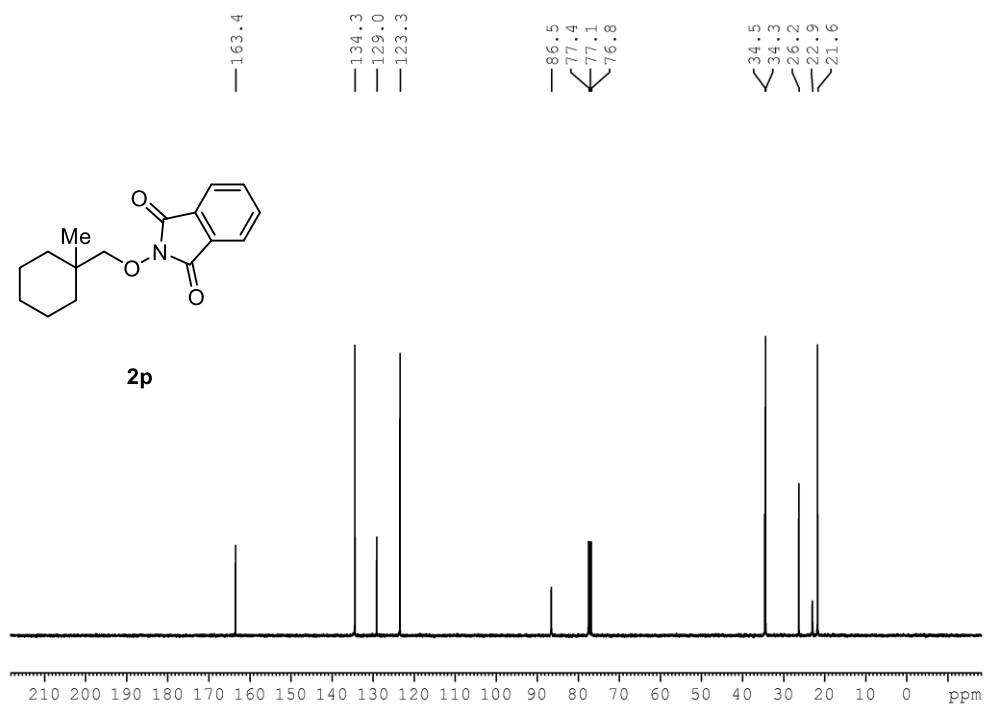
Supplementary Figure 127. ¹H NMR spectrum of compound 2o



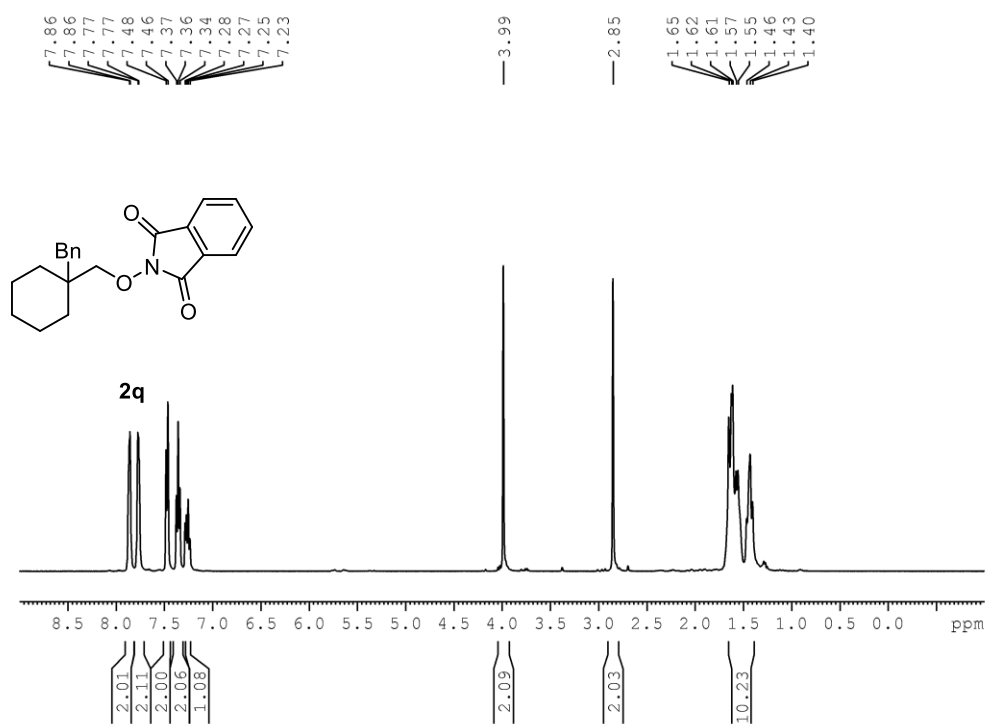
Supplementary Figure 128. ¹³C NMR spectrum of compound 2o



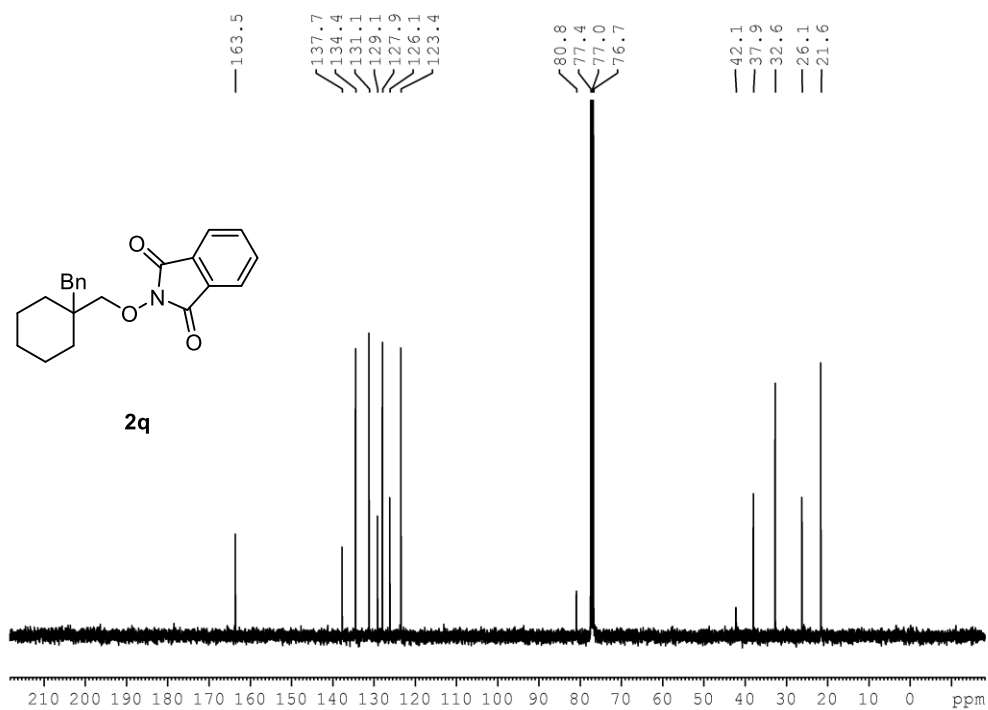
Supplementary Figure 129. ^1H NMR spectrum of compound **2p**



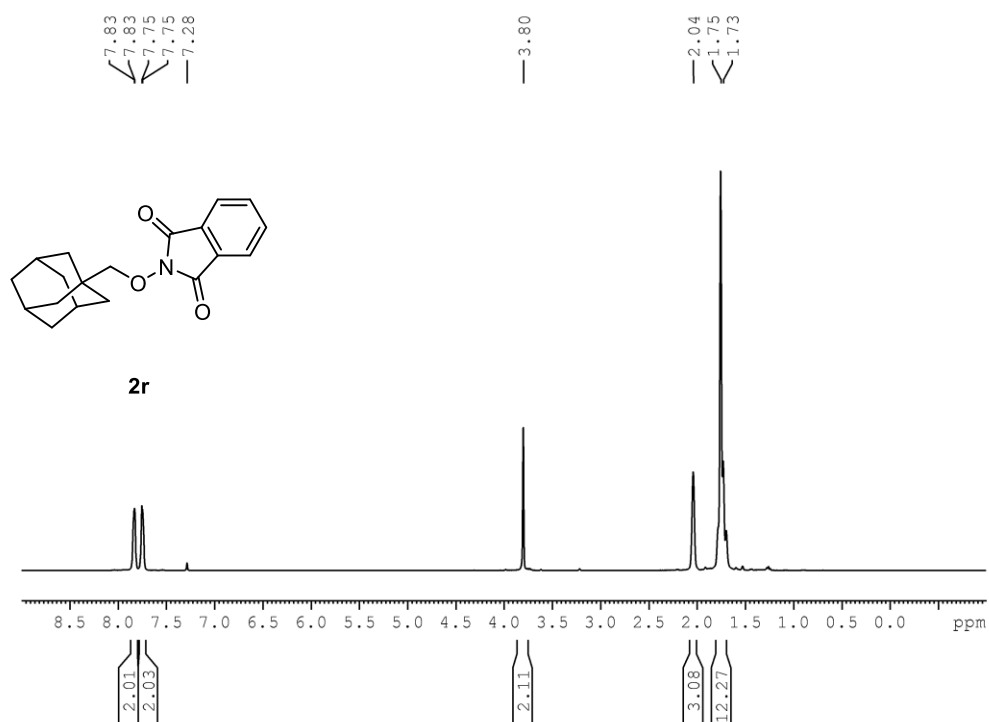
Supplementary Figure 130. ^{13}C NMR spectrum of compound **2p**



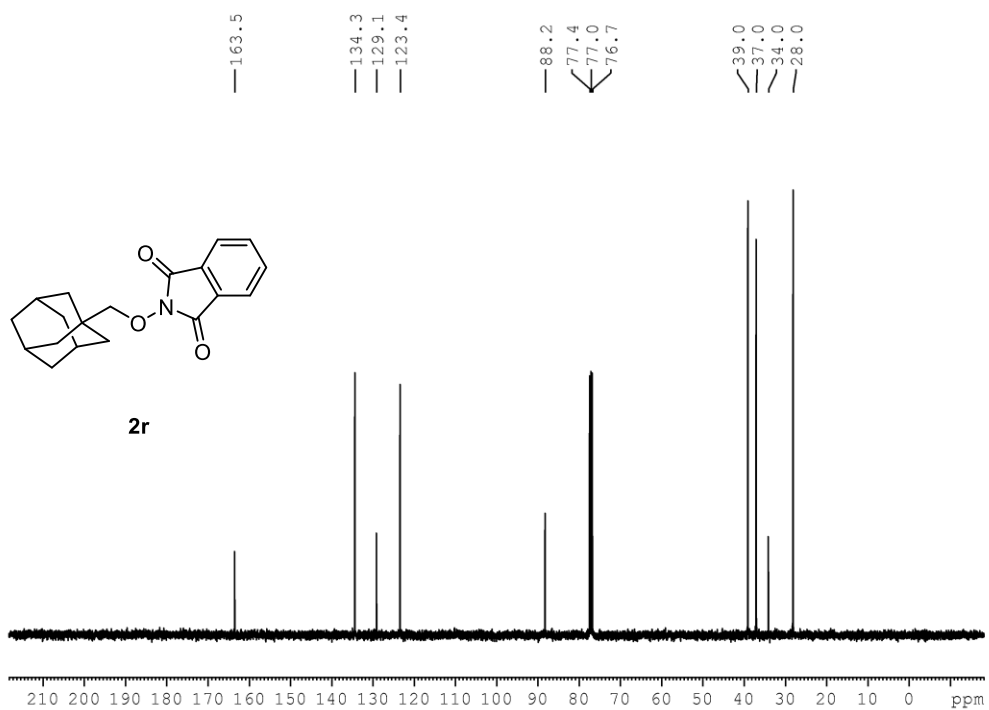
Supplementary Figure 131. ^1H NMR spectrum of compound 2q



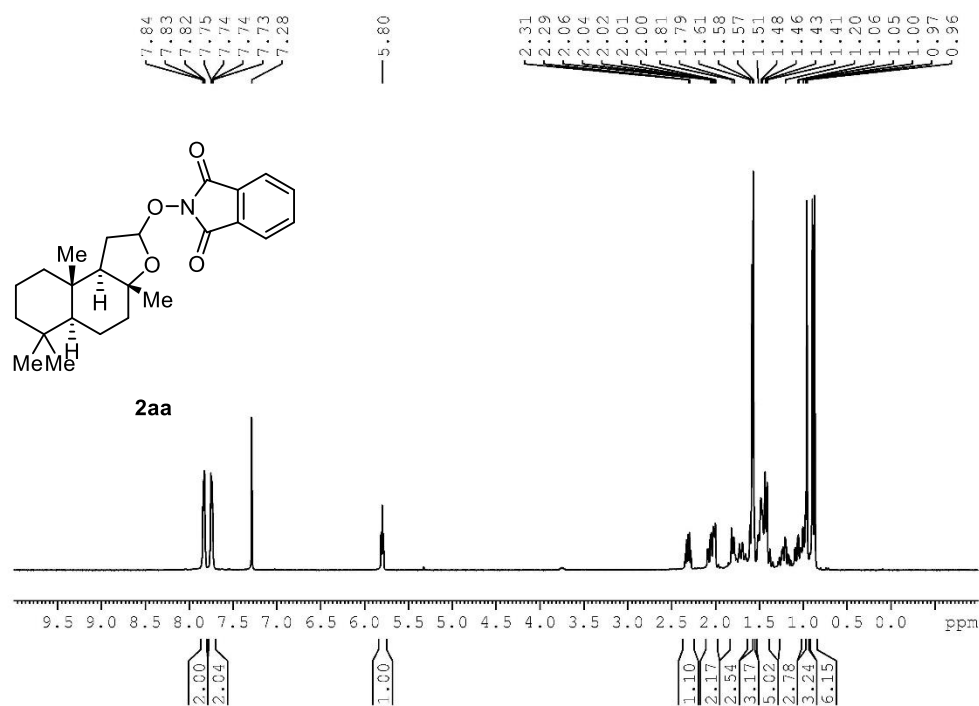
Supplementary Figure 132. ^{13}C NMR spectrum of compound 2q



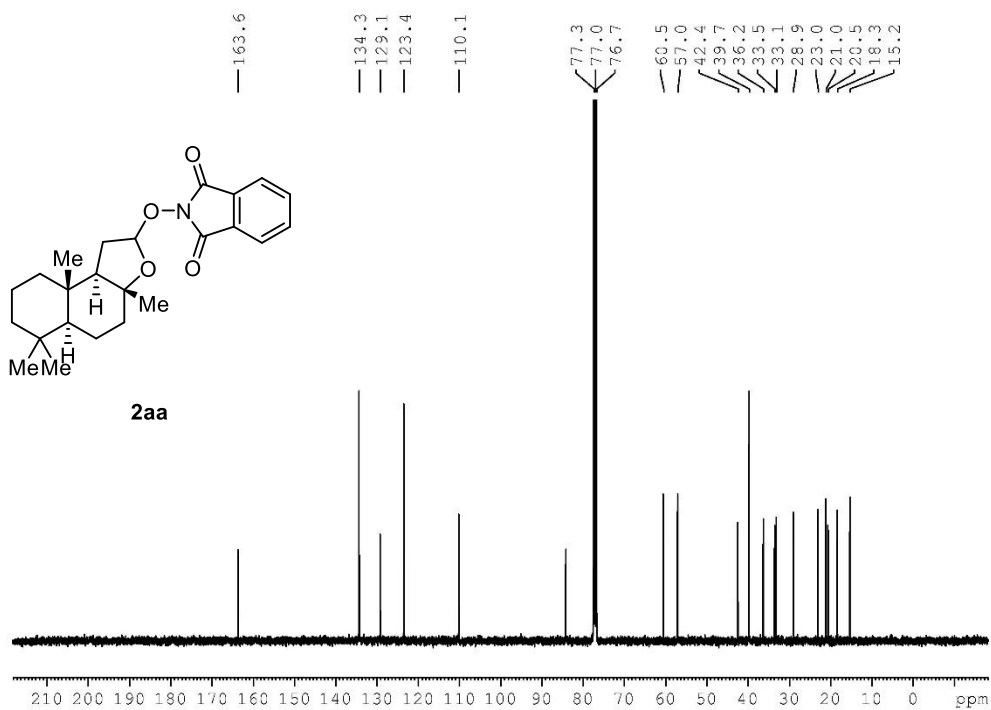
Supplementary Figure 133. ^1H NMR spectrum of compound **2r**



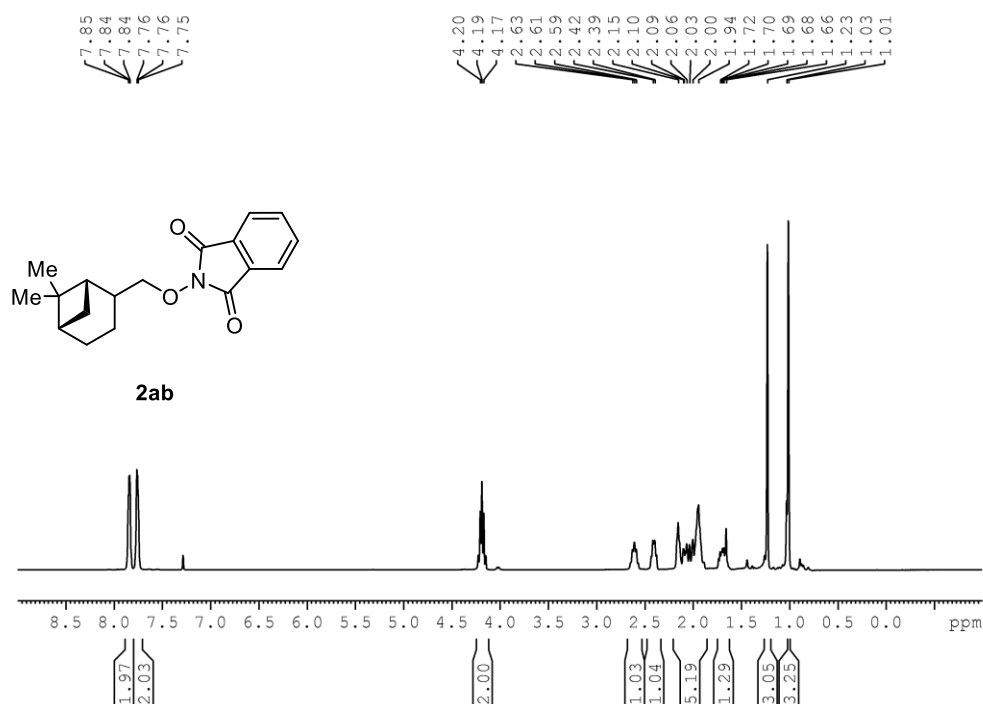
Supplementary Figure 134. ^{13}C NMR spectrum of compound **2r**



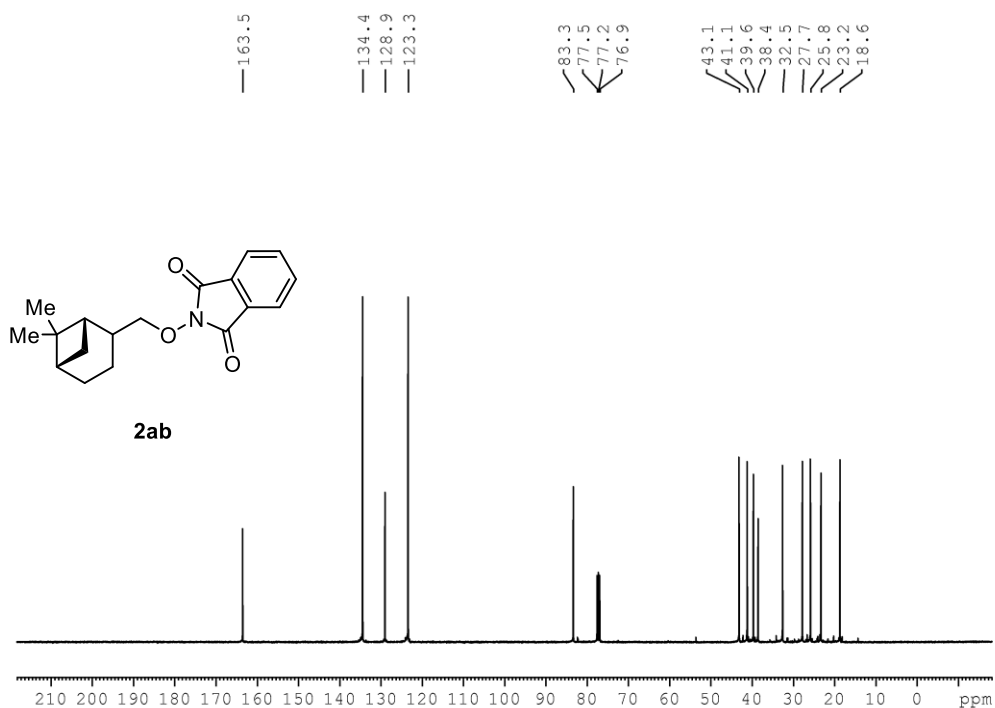
Supplementary Figure 135. ^1H NMR spectrum of compound 2aa



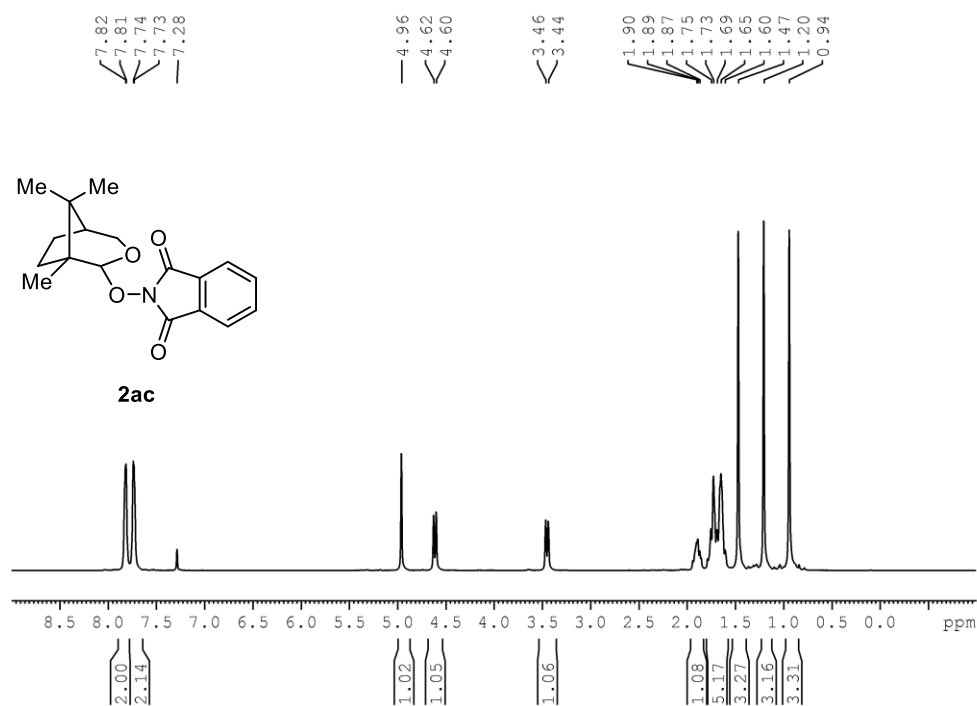
Supplementary Figure 136. ^{13}C NMR spectrum of compound 2aa



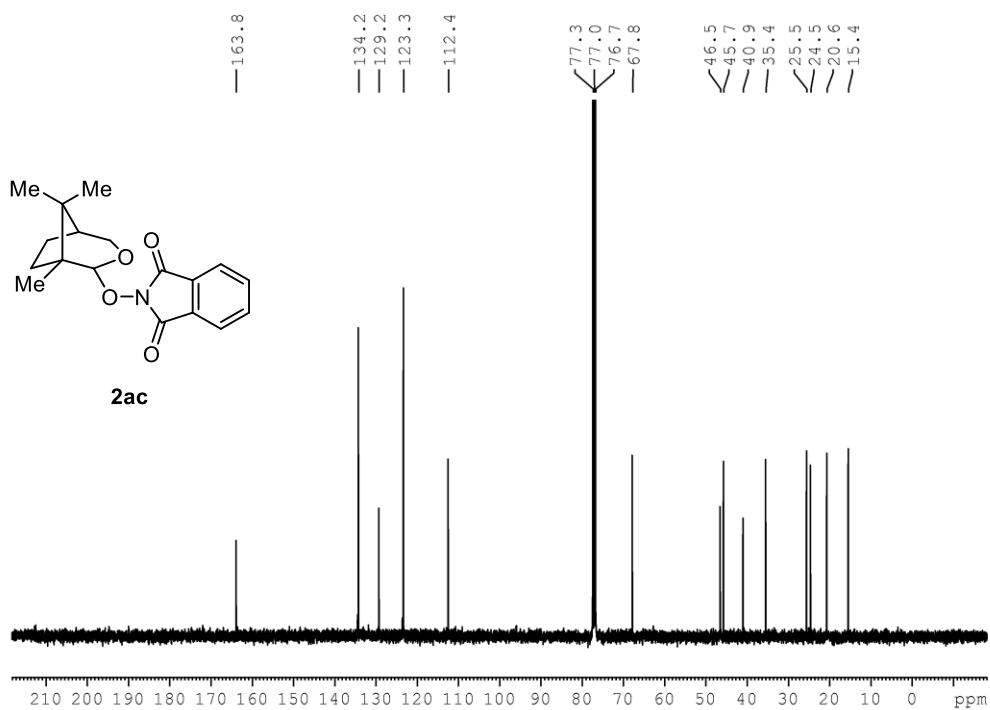
Supplementary Figure 137. ¹H NMR spectrum of compound 2ab



Supplementary Figure 138. ¹³C NMR spectrum of compound 2ab



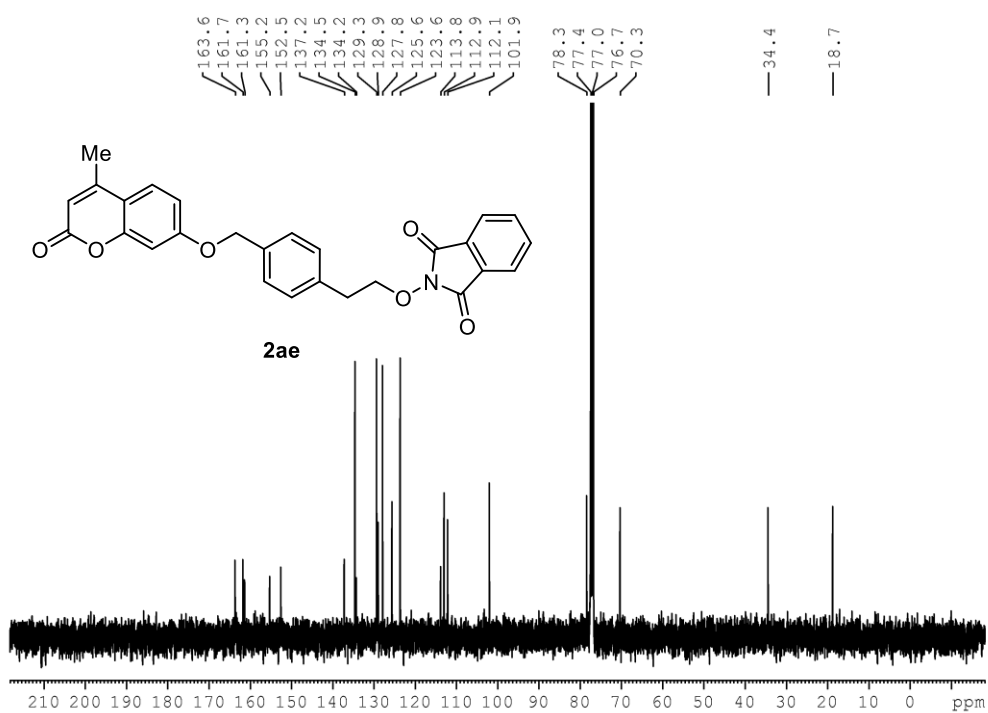
Supplementary Figure 139. ¹H NMR spectrum of compound 2ac



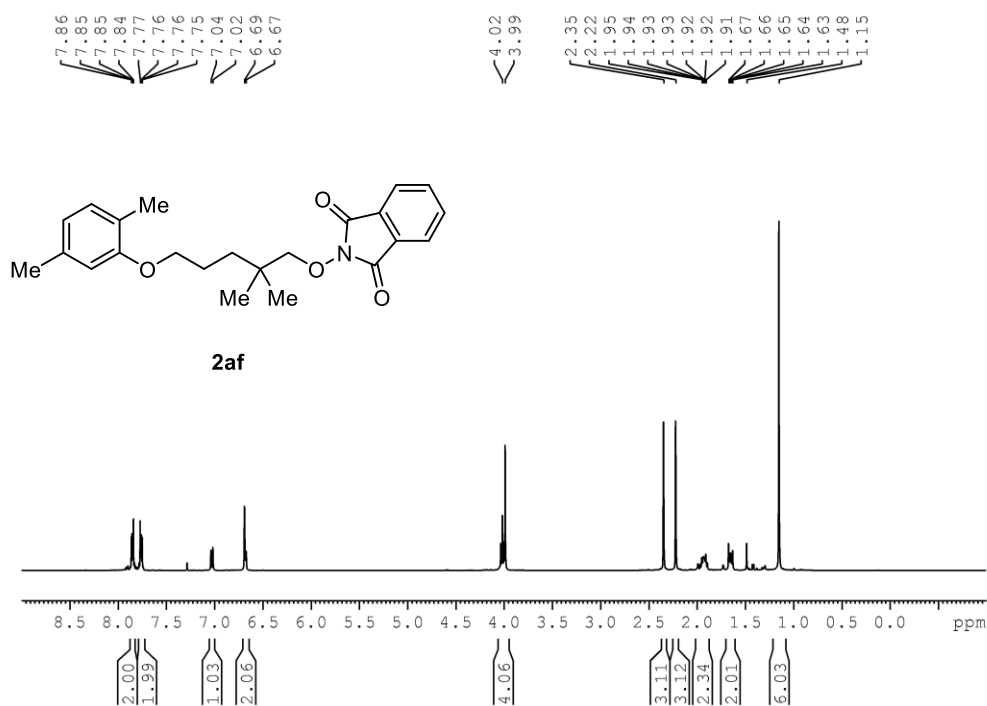
Supplementary Figure 140. ¹³C NMR spectrum of compound 2ac



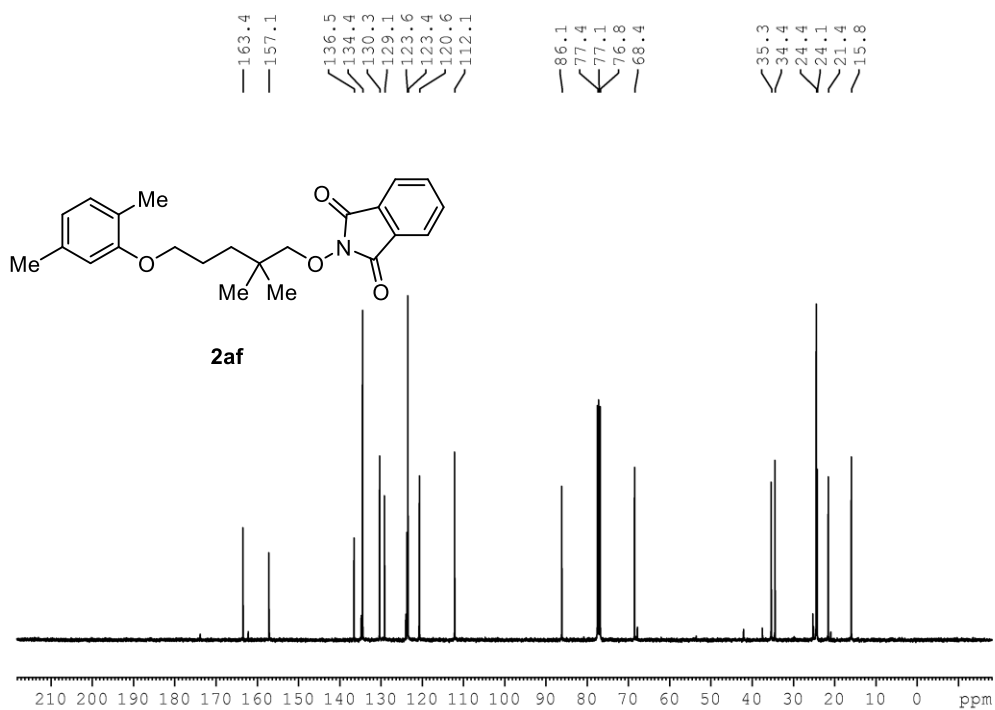
Supplementary Figure 141. ¹H NMR spectrum of compound 2ae



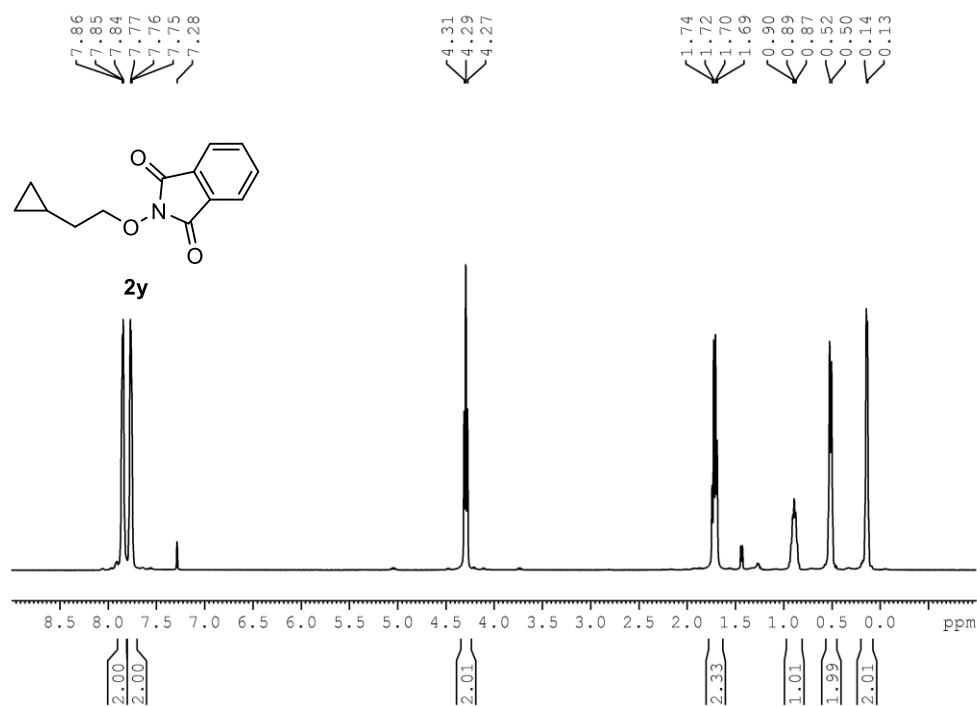
Supplementary Figure 142. ¹³C NMR spectrum of compound 2ae



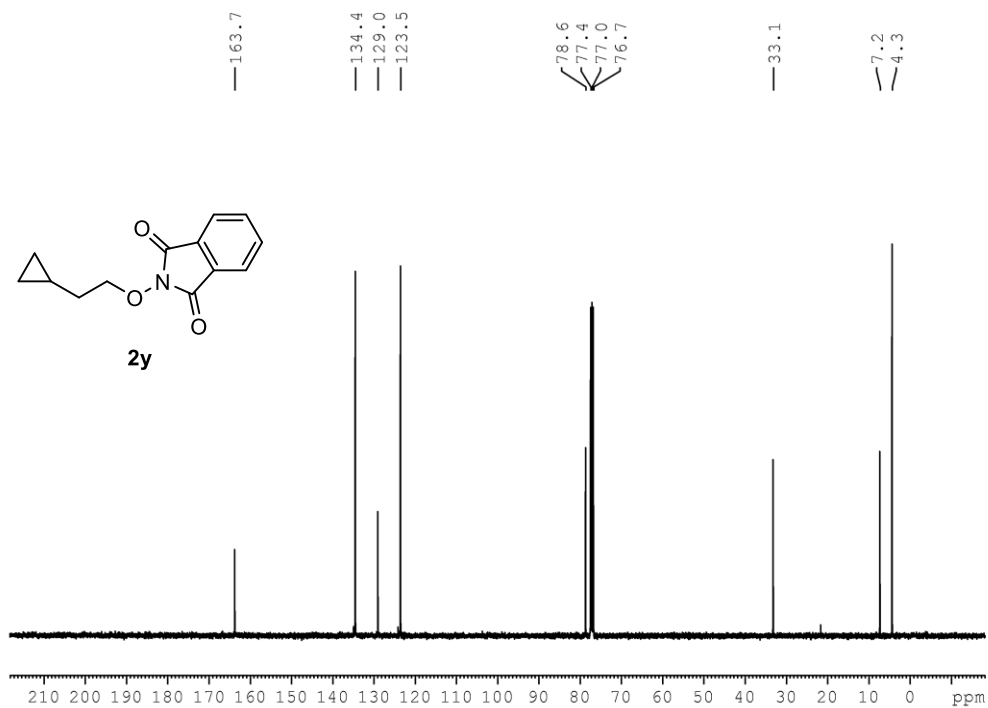
Supplementary Figure 143. ¹H NMR spectrum of compound 2af



Supplementary Figure 144. ¹³C NMR spectrum of compound 2af



Supplementary Figure 145. ^1H NMR spectrum of compound **2y**



Supplementary Figure 146. ^{13}C NMR spectrum of compound **2y**

5. Supplementary Reference

1. Wang, J. *et al.* Visible-light promoted α -alkylation of glycine derivatives with alkyl boronic acids. *Chem. Commun.* **57**, 1959–1962 (2021).
2. Ren, L. & Cong, H. Visible-light-driven decarboxylative alkylation of C-H bond catalyzed by dye-sensitized semiconductor. *Org. Lett.* **20**, 3225–3228 (2018).
3. Zhang, W. *et al.* Mesoionic pyrido[1,2-a]pyrimidinones: a novel class of insecticides inhibiting nicotinic acetylcholine receptors. *Bioorganic Med. Chem. Lett.* **26**, 5444–5449 (2016).
4. LaRochelle, M. C., Thomas, J. L. & Strickler, R. C. Reactivation of human placental 17 β , 20 α -hydroxysteroid dehydrogenase: affirmation of affinity labeling principles. *Steroids* **43**, 209–217 (1984).
5. Zhu, X., Lin, Y., Sun, Y., Beard, M. C. & Yan, Y. Lead-halide perovskites for photocatalytic α -alkylation of aldehydes. *J. Am. Chem. Soc.* **141**, 733–738 (2019).
6. Jerezano, A. V *et al.* Iodine-mediated one-pot synthesis of indoles and 3-dimethylaminoindoles via annulation of enamines. *Arkivoc* **2014**, 18–53 (2013).
7. Wang, C. *et al.* Visible-light-driven, copper-catalyzed decarboxylative C(sp³)-H alkylation of glycine and peptides. *Angew. Chem. Int. Ed.* **57**, 15841–15846 (2018).
8. Zhang, J. *et al.* Donor-acceptor complex enables alkoxyl radical generation for metal-free C(sp³)-C(sp³) cleavage and allylation/alkenylation. *Angew. Chem. Int. Ed.* **129**, 12793–12797 (2017).
9. Lei, X., Feng, J., Guo, Q., Xu, C. & Shi, J. Base-promoted formal [3 + 2] cycloaddition of α -halohydroxamates with carbon disulfide to synthesize polysubstituted Rhodanines. *Org. Lett.* **24**, 2837–2841 (2022).
10. Xu, P., Qian, B., Hu, B. & Huang, H. Palladium-catalyzed tandem hydrocarbonylative lactamization and cycloaddition reaction for the construction of bridged polycyclic lactams. *Org. Lett.* **24**, 147–151 (2022).
11. Xu, X., Liu, Y. & Park, C. M. Rhodium(III)-catalyzed intramolecular annulation through C-H activation: Total synthesis of (\pm)-antofine, (\pm)-septicine, (\pm)-tylophorine, and rosettain. *Angew. Chem. Int. Ed.* **51**, 9372–9376 (2012).
12. Shi, J. L., Wang, Z., Zhang, R., Wang, Y. & Wang, J. Visible-light-promoted ring-opening alkynylation, alkenylation, and allylation of cyclic hemiacetals through β -scission of alkoxy radicals. *Chem. A Eur. J.* **25**, 8992–8995 (2019).
13. Shu, C., Madhavachary, R., Noble, A. & Aggarwal, V. K. Photoinduced fragmentation borylation of cyclic alcohols and hemiacetals. *Org. Lett.* **22**, 7213–7218 (2020).
14. Cong, F., Lv, X. Y., Day, C. S. & Martin, R. Dual catalytic strategy for forging sp²-sp³ and sp³-sp³ architectures via β -scission of aliphatic alcohol derivatives. *J. Am. Chem. Soc.* **142**, 20594–20599 (2020).
15. Haibach, M. C., Stoltz, B. M. & Grubbs, R. H. Catalytic reduction of alkyl and aryl bromides using propan-2-ol. *Angew. Chem. Int. Ed.* **56**, 15123–15126 (2017).
16. Dixon, D. D., Lockner, J. W., Zhou, Q. & Baran, P. S. Scalable, divergent synthesis of meroterpenoids via ‘borono- sclareolide’. *J. Am. Chem. Soc.* **134**, 8432–8435 (2012).
17. Braun, J. C. & Fisher, G. S. Hydroboration of β -pinene. *Tetrahedron Lett.* **1**, 9–11 (1960).
18. Dhoro, F., Kristensen, T. E., Stockmann, V., Yap, G. P. A. & Tius, M. A. Asymmetric

- amine-intercepted Nazarov cyclization. *J. Am. Chem. Soc.* **129**, 7256–7257 (2007)
19. Ritter, C. *et al.* Bioorthogonal enzymatic activation of caged compounds. *Angew. Chem. Int. Ed.* **54**, 13440–13443 (2015).
 20. Ong, D. Y. *et al.* Controlled reduction of carboxamides to alcohols or amines by zinc hydrides. *Angew. Chem. Int. Ed.* **58**, 4992–4997 (2019).
 21. Maercker, A. & Theysohn, W. Versuche zur Umlagerung von 2-cyclopropyl-äthyl-anionen. *Justus Liebigs Ann. Chem.* **759**, 132–157 (1972).
 22. Xia, Q. *et al.* Visible light-driven α -alkylation of *N*-aryl tetrahydroisoquinolines initiated by electron donor-acceptor complexes. *Org. Lett.* **22**, 7290–7294 (2020).
 23. Nguyen, H. N. *et al.* Synthesis of 4-substituted chlorophthalazines, dihydrobenzoazepinediones, 2-pyrazolylbenzoic acid, and 2-pyrazolylbenzohydrazide via 3-substituted 3-hydroxyisoindolin-1-ones. *J. Org. Chem.* **77**, 3887–3906 (2012).
 24. Kang, T., Kim, H., Kim, J. G. & Chang, S. Synthesis of 1,2-amino alcohols via catalytic C-H amidation of sp^3 methyl C-H bonds. *Chem. Commun.* **50**, 12073–12075 (2014).
 25. Dubey, N., Sharma, P. & Kumar, A. Clay-supported Cu(II) catalyst: an efficient, heterogeneous, and recyclable catalyst for synthesis of 1,4-disubstituted 1,2,3-triazoles from alloxan-derived terminal alkyne and substituted azides using click chemistry. *Synth. Commun.* **45**, 2608–2626 (2015).
 26. Xuan, J. *et al.* Redox-neutral α -allylation of amines by combining palladium catalysis and visible-light photoredox catalysis. *Angew. Chem. Int. Ed.* **54**, 1625–1628 (2015).
 27. Qiao, J. *et al.* Direct, site-selective and redox-neutral α -C–H bond functionalization of tetrahydrofurans via quantum dots photocatalysis. *Angew. Chem. Int. Ed.* **60**, 27201–27205 (2021).
 28. Zeng, H. *et al.* Site-specific functionalization of 1,3-dioxolane with imines: a radical chain approach to masked α -amino aldehydes. *J. Org. Chem.* **83**, 5256–5266 (2018).
 29. Halland, N. & Jørgensen, K. A. Intermolecular addition of alkyl radicals to imines in the absence and in the presence of a Lewis acid. *J. Chem. Soc. Perkin Trans.1* 1290–1295 (2001).
 30. Kitoun, C. *et al.* Phosphine-mediated bioconjugation of the 3'-end of RNA. *Org. Lett.* **22**, 8034–8038 (2020).