



Draft Genome Sequence of *Sulfurospirillum* sp. Strain MES, Reconstructed from the Metagenome of a Microbial Electrosynthesis System

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A draft genome of *Sulfurospirillum* sp. strain MES was isolated through taxonomic binning of a metagenome sequenced from a microbial electrosynthesis system (MES) actively producing acetate and hydrogen. The genome contains the *nosZDFLY* genes, which are involved in nitrous oxide reduction, suggesting the potential role of this strain in denitrification.

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Microbial electrosynthesis is a recently discovered process whereby microorganisms consume electricity and CO_2 for the production of reduced end products, such as commodity chemicals and fuels. Electrochemical data have suggested direct and/or mediated electron transfer between the electrode and biocatalyst, but the specific metabolic pathways involved remain poorly understood. The draft genome described here was obtained from shotgun sequencing of an electroacetogenic mixedmicrobial community metagenome. The microbial electrosynthesis system (MES) was operated for >150 days with a cathode potential of -590 mV versus standard hydrogen electrode (SHE) (1, 2).

A dual sequencing approach was utilized for the aforementioned MES metagenome. Approximately 32 million and 900,000 reads were generated using the Illumina MiSeq platform (2 imes250-bp paired-end sequencing) and the Pacific Biosciences (PacBio RS) platform, respectively. The Illumina reads were quality trimmed with the CLC Genomics Workbench. The trimmed Illumina reads were utilized for subsequent genome assembly and error correction of the PacBio reads. The error-corrected PacBio reads and trimmed Illumina reads were assembled with Velvet, and the contigs were binned by phylogeny using BLASTn (3). The Sulfurospirillum-associated contig bin was used to map the raw metagenome reads. The resultant Sulfurospirillum-associated Illumina and PacBio reads were assembled using a combination of Velvet, CLC Genomics, and SPAdes (4, 5). The draft genome of this strain, thus named Sulfurospirillum sp. strain MES, is 2.67 Mbp (G+C content, 43.8%) contained in 130 contigs (61 >500 bp), with an N_{50} of 371,847 bp and the longest contig being 724,139 bp. The scaffolds were annotated with RAST (6), revealing a total of 2,691 features (2,655 protein-coding genes and 36 RNAs).

Phylogenetic analysis of the full-length 16S rRNA gene from the *Sulfurospirillum*-associated genome bin suggests that the *Sul*- furospirillum sp. strain MES is most closely related to the cultured Sulfurospirillum cavolei strain Phe91 (99% identical) and uncultured/enrichment culture clones associated with wastewateractivated sludge and petroleum reservoirs (7). An initial genomewide comparison to other Sulfurospirillum organisms with a draft or complete genome sequence suggests that Sulfurospirillum sp. strain MES is most closely related to Sulfurospirillum multivorans (81.6% and 78.0% identical at the nucleotide and amino acid levels, respectively). The members of Sulfurospirillum are heterotrophic denitrifiers capable of dissimilatory selenite and arsenate reduction and have been found in oil fields, aquifer sediments, and contaminated groundwater (8-10). Sulfurospirillum organisms have metabolic capabilities associated with reductive dehalogenation (10-12). These metabolisms are retained in the draft genome, which codes for the ability to utilize a wide variety of electron acceptors (e.g., arsenate, sulfur, and nitrate) and electron donors (e.g., formate and hydrogen [in the presence of acetate], pyruvate, fumarate, and lactate). Pangenomic analysis and comparative genomics are under way to analyze the other sequenced Sulfurospirillum genomes and to determine the importance of Sulfurospirillum sp. strain MES within the acetogenic MES. The aforementioned genome data provide new genetic insight into the Sulfurospirillum genus.

Nucleotide sequence accession numbers. The *Sulfurospirillum* sp. strain MES draft genome has been deposited at GenBank under the accession no. JSEC00000000. The version described in this paper is version JSEC00000000.

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REFERENCES

1. Marshall CW, Ross DE, Fichot EB, Norman RS, May HD. 2012. Electrosynthesis of commodity chemicals by an autotrophic microbial community. Appl Environ Microbiol 78:8412–8420. http://dx.doi.org/10 .1128/AEM.02401-12.

- Marshall CW, Ross DE, Fichot EB, Norman RS, May HD. 2013. Long-term operation of microbial electrosynthesis systems improves acetate production by autotrophic microbiomes. Environ Sci Technol 47: 6023–6029. http://dx.doi.org/10.1021/es400341b.
- 3. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. http://dx.doi.org/10.1016 /S0022-2836(05)80360-2.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. Genome Res 18:821–829. http://dx.doi .org/10.1101/gr.074492.107.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to singlecell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089 /cmb.2012.0021.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186 /1471-2164-9-75.
- 7. Tang Y-Q, Li Y, Zhao J-Y, Chi C-Q, Huang L-X, Dong H-P, Wu X-L.

2012. Microbial communities in long-term, water-flooded petroleum reservoirs with different *in situ* temperatures in the Huabei oilfield, China. PLoS One 7:e33535. http://dx.doi.org/10.1371/journal.pone.0033535.

- Hubert C, Voordouw G. 2007. Oil field souring control by nitratereducing *Sulfurospirillum* spp. that outcompete sulfate-reducing bacteria for organic electron donors. Appl Environ Microbiol 73:2644–2652. http: //dx.doi.org/10.1128/AEM.02332-06.
- Rowland HAL, Boothman C, Pancost R, Gault AG, Polya DA, Lloyd JR. 2009. The role of indigenous microorganisms in the biodegradation of naturally occurring petroleum, the reduction of iron, and the mobilization of arsenite from West Bengal aquifer sediments. J Environ Qual 38: 1598–1607. http://dx.doi.org/10.2134/jeq2008.0223.
- Sercu B, Jones ADG, Wu CH, Escobar MH, Serlin CL, Knapp TA, Andersen GL, Holden PA. 2013. The influence of *in situ* chemical oxidation on microbial community composition in groundwater contaminated with chlorinated solvents. Microb Ecol 65:39–49. http://dx.doi.org/10.10 07/s00248-012-0092-0.
- 11. Luijten MLGC, de Weert J, Smidt H, Boschker HTS, de Vos WM, Schraa G, Stams AJM. 2003. Description of Sulfurospirillum halorespirans sp. nov., an anaerobic, tetrachloroethene-respiring bacterium, and transfer of Dehalospirillum multivorans to the genus Sulfurospirillum as Sulfurospirillum multivorans comb. nov. Int J Syst Evol Microbiol 53:787–793. http://dx.doi.org/10.1099/ijs.0.02417-0.
- He J, Robrock KR, Alvarez-Cohen L. 2006. Microbial reductive debromination of polybrominated diphenyl ethers (PBDEs). Environ Sci Technol 40:4429–4434. http://dx.doi.org/10.1021/es052508d.