


# Modulation of Biliary Cancer Chemo-Resistance Through MicroRNA-Mediated Rewiring of the Expansion of CD133+ Cells

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**BACKGROUND AND AIMS:** Changes in single microRNA (miRNA) expression have been associated with chemo-resistance in biliary tract cancers (BTCs). However, a global assessment of the dynamic role of the microRNome has never been performed to identify potential therapeutic targets that are functionally relevant in the BTC cell response to chemotherapy.

**APPROACH AND RESULTS:** High-throughput screening (HTS) of 997 locked nucleic acid miRNA inhibitors was performed in six cholangiocarcinoma cell lines treated with cisplatin and gemcitabine (CG) seeking changes in cell viability. Validation experiments were performed with mir-Vana probes. MicroRNA and gene expression was assessed by TaqMan assay, RNA-sequencing, and *in situ* hybridization in four independent cohorts of human BTCs. Knockout of microRNA was achieved by CRISPR-CAS9 in CCLP cells (MIR1249KO) and tested for effects on chemotherapy sensitivity *in vitro* and *in vivo*. HTS revealed that MIR1249 inhibition enhanced chemotherapy sensitivity across all cell lines. MIR1249 expression was increased in 41% of cases in human BTCs. In validation experiments, MIR1249 inhibition did not alter cell viability in untreated or dimethyl sulfoxide-treated cells; however, it did increase the CG effect. MIR1249

expression was increased in CD133+ biliary cancer cells freshly isolated from the stem cell niche of human BTCs as well as in CD133+ chemo-resistant CCLP cells. MIR1249 modulated the chemotherapy-induced enrichment of CD133+ cells by controlling their clonal expansion through the Wnt-regulator FZD8. MIR1249KO cells had impaired expansion of the CD133+ subclone and its enrichment after chemotherapy, reduced expression of cancer stem cell markers, and increased chemosensitivity. MIR1249KO xenograft BTC models showed tumor shrinkage after exposure to weekly CG, whereas wild-type models showed only stable disease over treatment.

**CONCLUSIONS:** MIR1249 mediates resistance to CG in BTCs and may be tested as a target for therapeutics. (HEPATOLOGY 2020;72:982-996).

**B**iliary tract cancers (BTCs) include cholangiocarcinoma (CCA) and gallbladder cancer, and their incidence is increasing worldwide.<sup>(1-4)</sup> Lack of effective radical treatments and rapid failure of palliative ones highlight the need for a better understanding of BTC biology and mechanisms of response to treatment.<sup>(2,5)</sup> Eighty percent of patients

*Abbreviations:* ABC, advanced biliary cancer; BTC, biliary tract cancer; CCA, cholangiocarcinoma; CG, cisplatin and gemcitabine; CRISPR-CAS9, CRISPR-associated protein-9 nuclease; CSC, cancer stem cell; CTRL, control; DMSO, dimethyl sulfoxide; FACS, fluorescence-activated cell sorting; FZD8, frizzled class receptor 8; gRNA, single-guide RNA; HTS, high-throughput screening; ISH, *in situ* hybridization; KO, knockout; LNA, locked nucleic acid; miRNA, microRNA; MIR, microRNA; TCGA, The Cancer Genome Atlas; WT, wild type.

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with BTCs present at an advanced stage, when treatment options are limited to chemotherapy with cisplatin and gemcitabine (CG).<sup>(5)</sup> Only 11% of patients gain a long-term benefit from chemotherapy, while primary resistance is detected in 20% of patients.<sup>(6)</sup> Most patients develop secondary resistance after an initial response or stabilization of the disease, which is responsible for a global median overall survival shorter than 12 months. Several mechanisms of chemo-resistance may act synergistically and drive cancer cells to escape biochemical inhibition or cell death caused by chemotherapy.<sup>(7)</sup> Acquisition of stemness features in cancer cells appears to be a driver of resistance that is common with other tumor types.<sup>(8,9)</sup>

MicroRNAs (miRNAs) are small noncoding RNAs controlling mRNA expression.<sup>(10)</sup> We and others demonstrated that miRNAs are aberrantly expressed in BTCs and promote biliary carcinogenesis.<sup>(11-19)</sup> Despite growing evidence that links single miRNAs with chemo-resistance, no comprehensive genome-wide

approach has been undertaken to date to assess the functional role of miRNAs in the cellular dynamics involved in drug response in BTCs. It is known that chemo-resistance is a peculiar feature of BTCs, which is responsible for the poor prognosis of these patients. Thus, in this study we have investigated the functional role of microRNA inhibitors in mediating drug response in chemotherapy-treated BTC cells, using a high-throughput approach that investigates the inducible role of miRNAs in response to cancer drugs.

## Experimental Procedures

### HUMAN TISSUES

The human BTC tissues were collected under approval of the Ethical Committee for Clinical Research at three independent institutions: the Humanitas Research Hospital (#21072014; cohort

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1), the Royal Marsden Hospital (CCR 4415; cohort 2), and the University Hospital of Padua (#0010416; cohort 3). The study protocols conformed to the ethical guidelines of the 1975 Declaration of Helsinki, as per ethical approval given by the institutional review board. Formalin-fixed paraffin-embedded tissues were retrieved and RNA was extracted from the tumor and the matched nontumor component after microscopic dissection using the Ambion RecoverAll kit (Thermo Fisher Scientific, Waltham, MA). Relapse-free survival was used as an endpoint of the study. Disease recurrence was defined as the presence of imaging-proven disease.

## HIGH-THROUGHPUT SCREENING

A human locked nucleic acid (LNA) miRNA inhibitor library (miRCURY LNA version 3; #190102-3) was purchased from Exiqon (Life Technologies, Paisley, United Kingdom). The library was distributed across fifteen 96-well plates (Greiner Bio-One, Frickenhausen, Germany) in a volume of 5  $\mu$ L in each well. Each plate included two negative controls (LNA negative A and LNA negative B from Exiqon) and positive controls (AllStars Hs positive cell death phenotype control, SI04381048; Qiagen, Manchester, United Kingdom). Fifteen microliters of transfecting solution with medium and Hiperfect (PN301705; Qiagen) was added to each well. Thirty microliters of cell solution was then added to each well to have a final concentration of 10,000 cells and 50 nM of miRNA inhibitors. A column with no cells ( $\times 8$ ) was added in one plate. Forty-eight hours later, 50  $\mu$ L of a combination of cisplatin (232120; Sigma-Aldrich, Gillingham, United Kingdom) and gemcitabine (1288463-200MG; Sigma-Aldrich) diluted in medium were added. Cisplatin was dissolved in sterile phosphate-buffered saline and stocked at a concentration of 1 mg/mL (3 mM). Gemcitabine was diluted in dimethyl sulfoxide (DMSO) and stocked at a concentration of 10 mg/mL (30 mM). Both stocks were then diluted in medium to achieve a final solution that would always contain less than 0.001% of DMSO. Cell viability was measured 72 hours later by CellTiter-Blue Assay (Promega, Madison, WI). The cell-viability measurement from each hit was normalized to that of the averaged negative controls across the respective plate. Each cell line was tested

in triplicate. Statistical significance ( $P \leq 0.05$ ) was determined by two-sided Student  $t$  test across three replicates.

## MIR1249-KNOCKOUT GENERATION THROUGH CRISPR-CAS9

CCLP-1 cells were transfected using Lipofectamine 3000 reagent (Thermo Fisher Scientific), with the CRISPR vector pCAS-Guide-EF1a-GFP CRISPR Vector (GE100018; OriGene Technologies, Inc., Rockville, MD) expressing single-guide RNAs (gRNAs) containing the inserted target sequence for miR1249. Target sequences of the gRNA were as follows: gRNA3 5' CGTCGGTCGTGGTAGATAGG 3'; gRNA4 5' AATCTCGACCGGACCTCGAC 3'. Forty-eight hours later, green fluorescent protein-positive cells were sorted with a FACSaria-II (BD Biosciences, San Jose, CA) and maintained in culture. Genome editing was verified at day 17 using the Indel identification kit (Clontech Biotec, Mountain View, CA). Cells were enriched for the edited clones by performing serial dilution. Final assessment of the successful genome editing was performed by sequencing and real-time PCR.

## STATISTICAL ANALYSES

Statistical analyses were performed by GraphPad Prism 6 (La Jolla, CA). Results are expressed as mean  $\pm$  SD, unless indicated otherwise. Groups were compared with either a two-tailed Student  $t$  test (for analysis of two groups) or using one-way analysis of variance to compare multiple groups. Significance was accepted when  $P$  was less than 0.05.

CCLP-1 BTC XENOGRAFT MODEL. BTC xenograft tumours (WT N:20; MIR1249KO N:20) were established subcutaneously in 6-7 weeks female NOD-scid IL2Rgnull mice (Charles River Laboratories, Wilmington, MA, USA). The study was performed in accordance with UK Home Office regulations under the Animals Scientific Procedures Act 1986 and in accordance with UK National Cancer Research Institute guidelines and the ARRIVE guidelines. Animals were housed in specific pathogen-free rooms in autoclaved, aseptic microisolator cages with a maximum of five animals per cage. Please see supplementary material for more information.

## Results

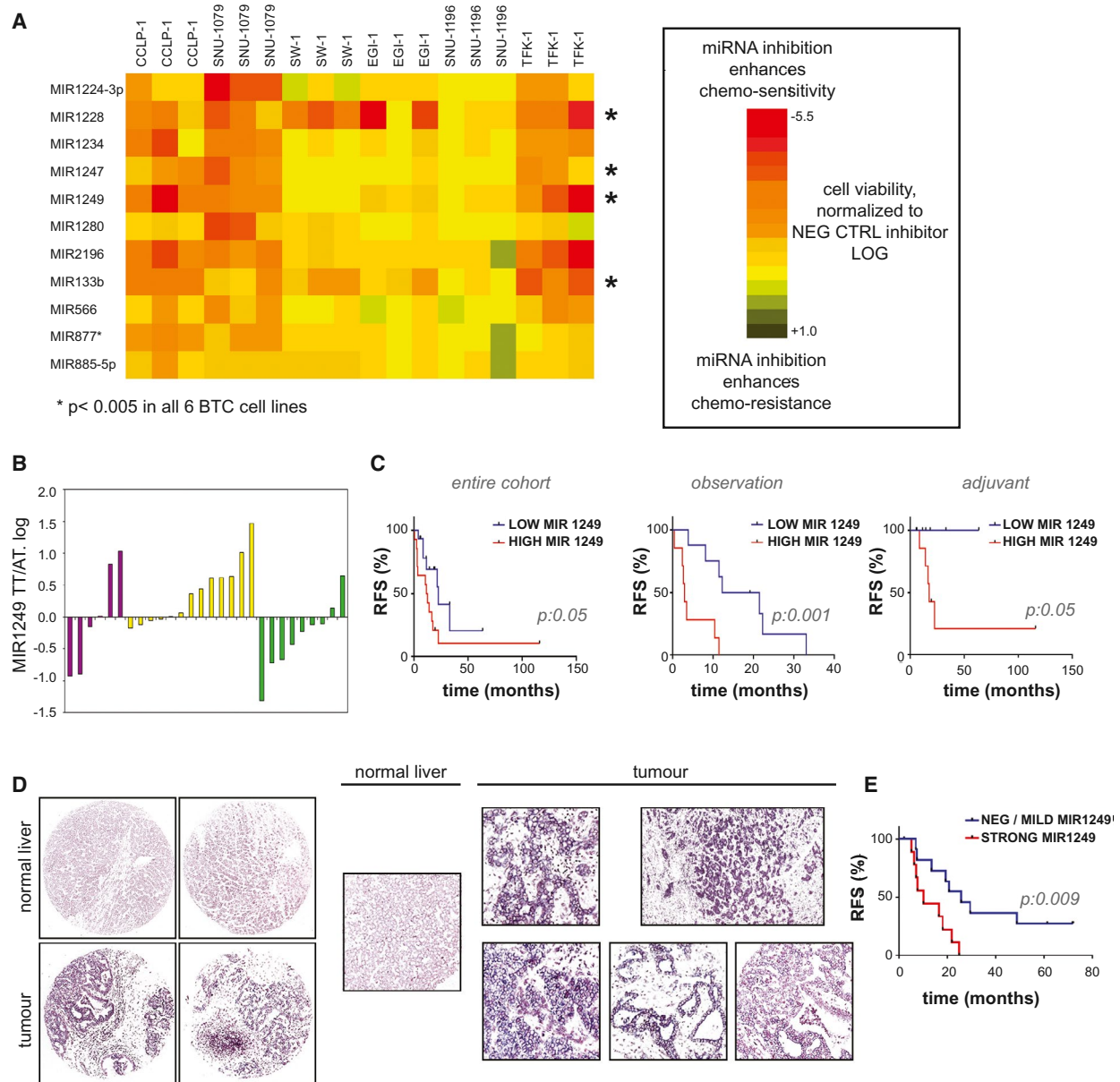
### HIGH-THROUGHPUT FUNCTIONAL STUDIES AND CHARACTERIZATION OF HUMAN CANCER TISSUES IDENTIFIED MIR1249 INHIBITION AS A CLINICALLY RELEVANT STRATEGY TO INCREASE CHEMO-SENSITIVITY IN HUMAN BTCs

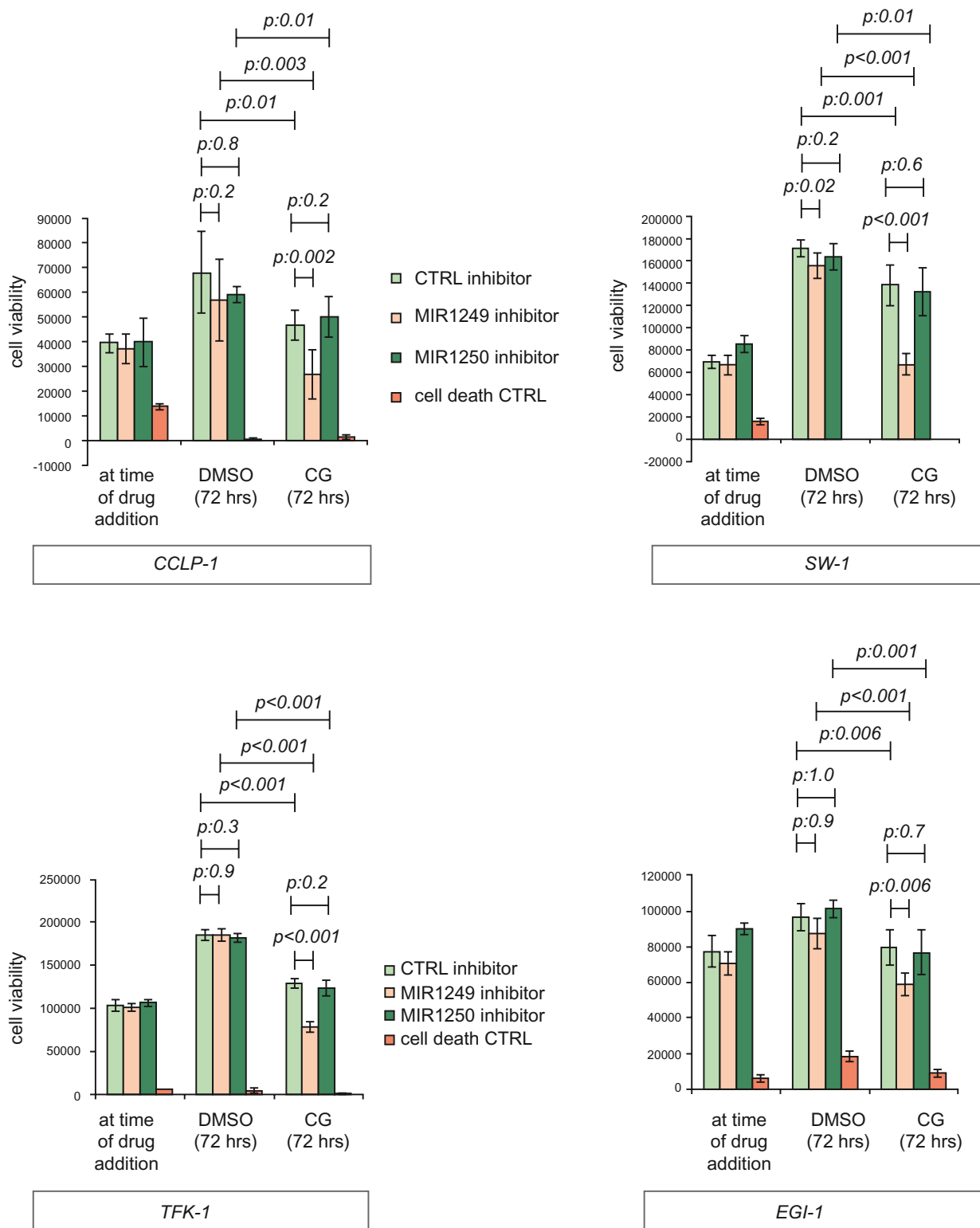
High-throughput screening (HTS) technologies were applied to screen a panel of six CG-treated BTC cell lines against a library of LNA miRNA inhibitors. The 50% growth inhibitory concentration for CG was derived for each cell line to define the concentration at which the combination of CG could induce cytotoxicity without reducing cell viability by more than 50%, to enable identification of sensitizers (Supporting Fig. S1). HTS was run in triplicate in each cell line (Supporting Table S1). Inhibition of MIR148a and members of the let-7 family reduced sensitivity in a number of cell lines, in line with published literature.<sup>(13,20)</sup> Eleven miRNA-inhibitors acted as sensitizers in all intrahepatic CCA cells, and four in all extrahepatic CCA cells ( $P < 0.05$ ) (Fig. 1A). Inhibitors of MIR1249, MIR133b, MIR1247, and MIR1228 decreased cell viability across all cell lines in comparison to control (CTRL) inhibitors. The tissue expression of these four short-listed miRNAs was determined in human BTCs by TaqMan assay to investigate the clinical relevance of these candidates. MIR133b, MIR1247, and MIR1228 expression was not increased in the tumor tissue in comparison to matched adjacent nontumor tissue (cohort 1,  $n = 29$ ) (Supporting Fig. S2A). Conversely, MIR1249 was overexpressed in the tumor compartment in comparison to paired nontumor tissue in 32% of cases (Fig. 1B). Interestingly, when the cohort was split according to median MIR1249 tumor expression, cases with high expression were associated with worse prognosis independently of adjuvant chemotherapy (Fig. 1C and Supporting Table S2). At multivariate analysis (considering T stage, N stage, adjuvant chemotherapy and MIR1249 tumor expression), adjuvant treatment (hazard ratio [HR] 0.70;  $P < 0.001$ ) and MIR1249 expression (HR

0.65;  $p: 0.004$ ) maintained an independent prognostic value. An increase of MIR1249 expression by TaqMan assay was observed in 53% of the cases of a separate cohort (cohort 2,  $n = 28$ ) (Supporting Fig. S2B). When cohorts 1 and 2 were pooled together, 41% of cases showed increased MIR1249 expression in the tumor. RNA-seq data confirmed overexpression of MIR1249 in the tumor tissue in comparison to paired normal tissue in 55% of cases (The Cancer Genome Atlas [TCGA] cohort,<sup>(21)</sup>  $n = 9$ ) (Supporting Fig. S2C). Kaplan-Meier analysis of the whole TCGA cohort showed that tumor MIR1249 expression was again associated with progression-free interval (Supporting Fig. S2D). When assessed by *in situ* hybridization (ISH), MIR1249 was strongly positive in 53% of the tumor cases and was statistically associated with lower survival outcome (cohort 3,  $n = 28$ ) (Fig. 1D-E and Supporting Table S3).

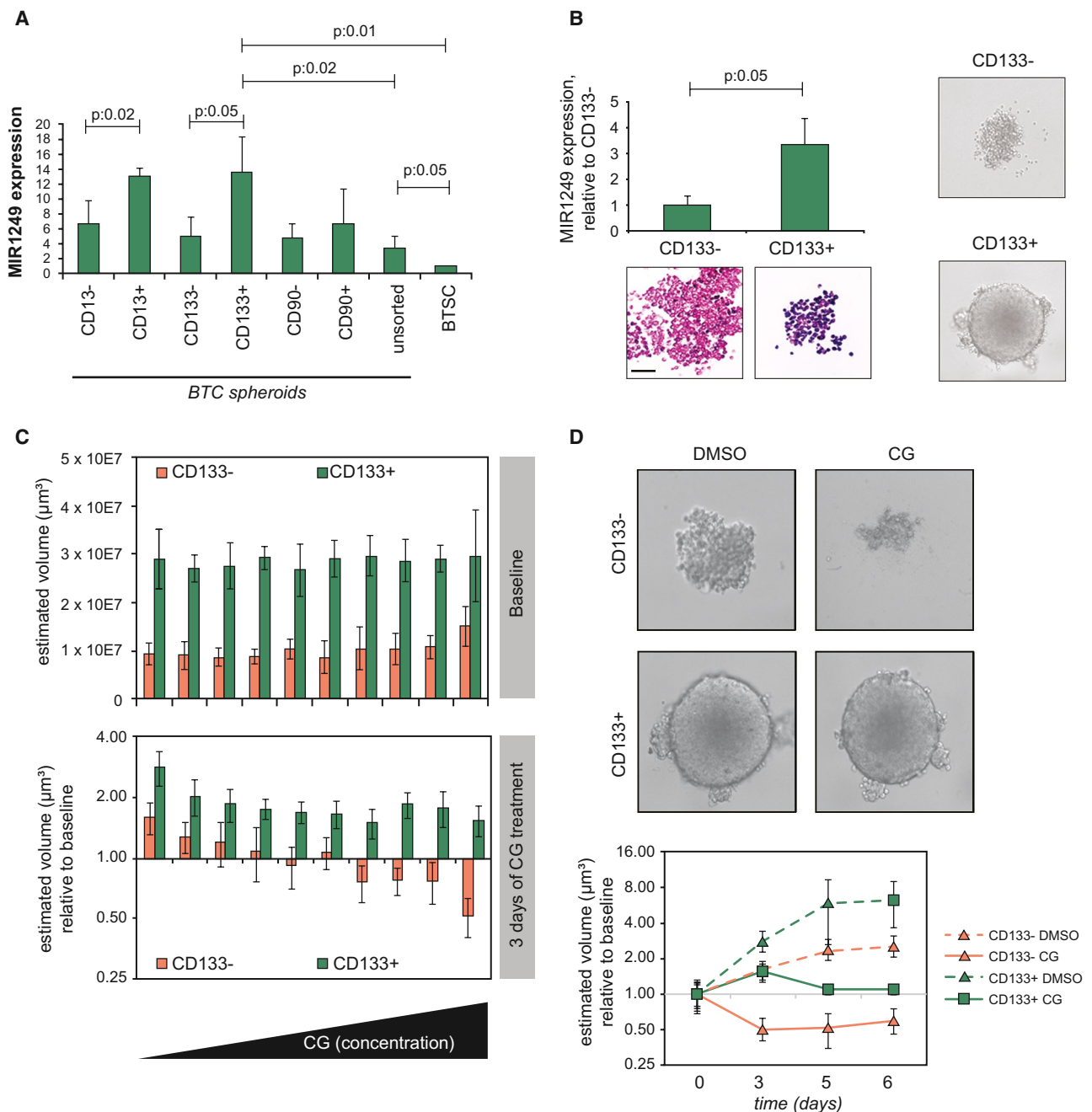
### VALIDATION FUNCTIONAL STUDIES IDENTIFIED THE INVOLVEMENT OF MIR1249 IN DRIVING A CHEMOTHERAPY-SPECIFIC REACTIVE RESPONSE IN CANCER CELLS

On the basis of the clinical and biological relevance of these data, MIR1249 was selected as a candidate for further studies. MIR1250 inhibitor (which exhibited no effect on any of the cells in the HTS) was included in the validation phase as a negative control, along with scrambled control (Supporting Fig. S2E). Using alternative probes for miRNA inhibition, the ability of MIR1249 inhibitor to enhance BTC cell response to CG chemotherapy was validated. Interestingly, we did not observe a cytotoxic effect for MIR1249 inhibitor in the absence of chemotherapy treatment (Fig. 2), suggesting that MIR1249 interferes with a chemotherapy-specific response. Indeed, MIR1249 expression was increased as a response to CG treatment in human CCA cells (Supporting Fig. S2F). Enrichment of resistant cells expressing stem cell markers is known to occur in response to chemotherapy treatment in a variety of cancers.<sup>(22,23)</sup> Thus, we hypothesized that MIR1249 inhibition can increase chemo-sensitivity by limiting the expansion of this resistant subpopulation. To verify our hypothesis, we assessed the expression of MIR1249 in human BTC spheroids generated from human cells freshly extracted from the stem cell

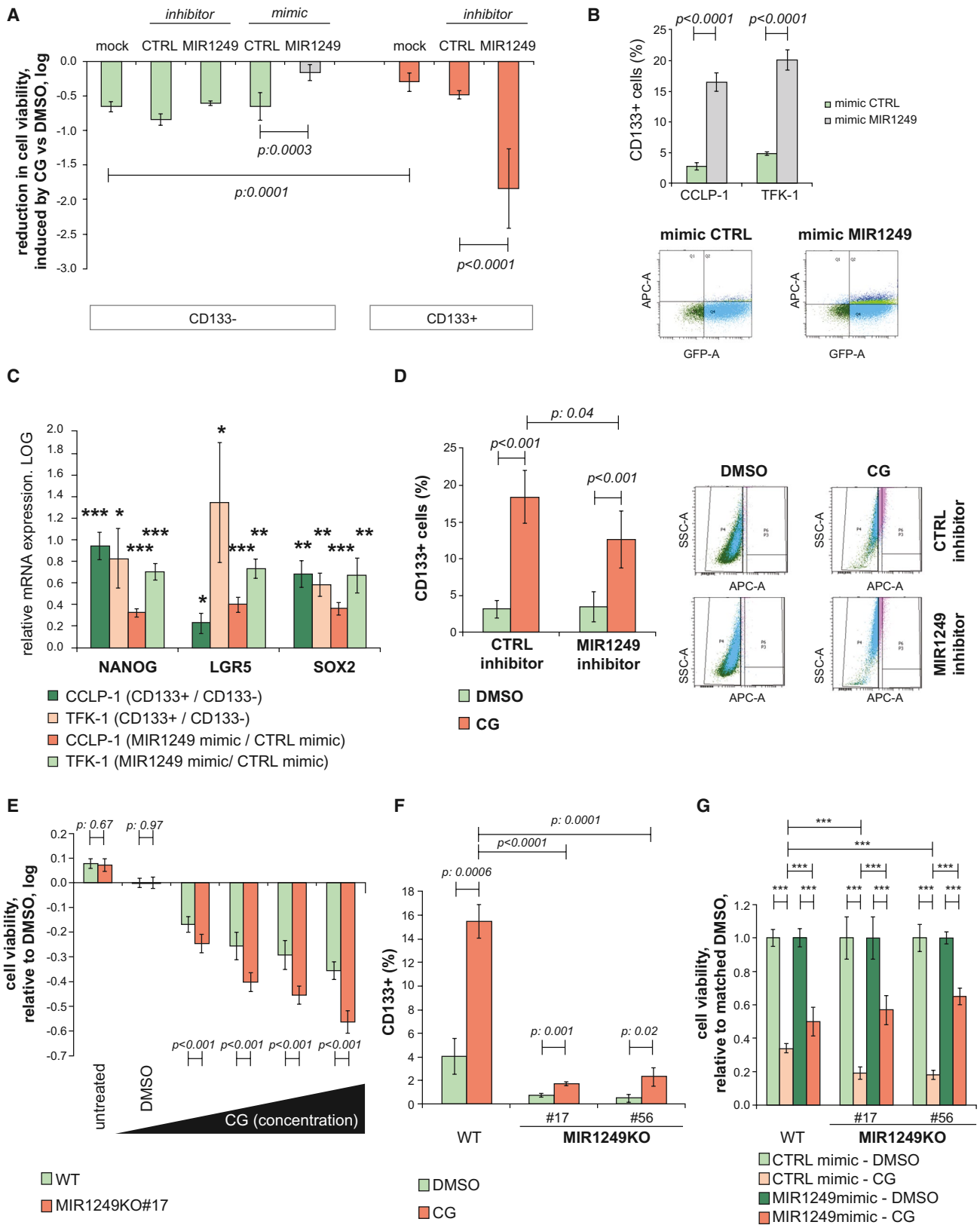




**FIG. 2.** MIR1249 inhibition decreases cell viability in chemotherapy-treated cells only. MIR1249 inhibition was achieved using mirVana inhibitory probes (Thermo Fisher Scientific). Cells were plated in 96-well plates and transfected with the indicated probes for 48 hours before being treated with DMSO or CG for a further 72 hours. Cell death CTRL was used as a positive control. Cell viability was assessed by CellTiter Blue assay (Promega, Madison, WI). Bars indicate the mean of six independent experiments  $\pm$  SD.



**FIG. 3.** Human CD133+ BTC cells hold high MIR1249 levels and are highly chemo-resistant. (A) CSC niche of human BTC samples was identified. Cells were then extracted, sorted by cell surface markers, and cultured in 3D spheroids. MIR1249 expression was assessed by TaqMan assay. Bars indicate three independent replicates  $\pm$  SD. (B) CCLP-1 cells were sorted by FACS for CD133 surface expression. MIR1249 was assessed both by TaqMan (bars indicate the mean of three independent experiments  $\pm$  SD) and by ISH (representative pictures are shown; scale bar: 100  $\mu\text{M}$ ). (C) When cultured in ultralow attachment plates, CD133+ cells formed large and defined 3D spheroids conversely to CD133- cells. (C) CCLP-1 cells were sorted for CD133 surface expression and cultured in ultralow attachment plates. After 5 days, baseline imaging of spheroids by Celigo showed reproducible spheroid formation across the plates, even though CD133- spheroids were smaller. After 3 days of CG treatment (scalar concentrations up to 3  $\mu\text{M}$  cisplatin and 30 nM gemcitabine), CD133- spheroids shrank in volume, whereas CD133+ spheroids did not, even at the highest CG concentration. Bars represent the mean of six replicates  $\pm$  SD. (D) Representative pictures of spheroids with and without CG treatment. Spheroids were monitored up to 6 days. The volume of CD133- spheroids decreased from baseline, whereas the volume of CD133+ spheroids was stabilized (see also Supporting Fig. S2). Abbreviation: BTSC, biliary tract stem cells.





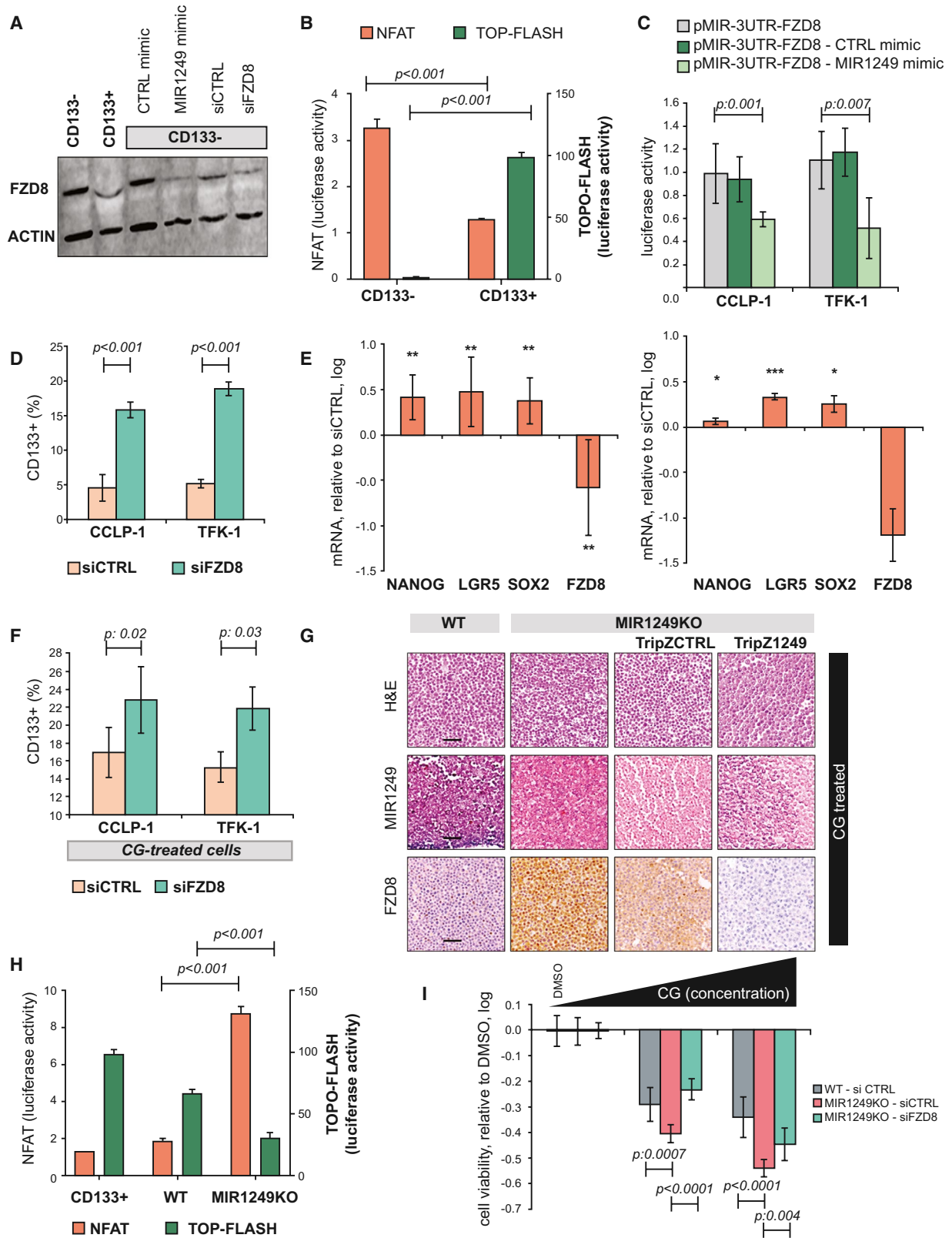
**FIG. 4.** MIR1249 inhibition prevents chemotherapy-induced enrichment of the CD133+ subclone. (A) CCLP-1 cells were transfected with indicated probes after being sorted for CD133 surface expression. Mock indicates the absence of any probe. Chemotherapy was added 48 hours after transfection, and cell viability was read 72 hours later by CellTiter Blue assay. Bars indicate the mean of six independent experiments  $\pm$  SD. (B) BTC cells were transfected and assessed for CD133+ by fluorescence-activated cell sorting (FACS). The population of CD133+ cells increased with enforced expression of MIR1249. Bars indicate the mean of three independent experiments  $\pm$  SD. Representative pictures of the FACS analysis are shown; CD133+ cells are identified by the double positivity of allophycocyanin axis and green fluorescent protein axis (right upper quadrant). (C) BTC cells were sorted for CD133 and collected for RNA extraction. Bars represent the log value of the indicated ratio with the relative control (positive values above the  $x$  axis indicate an increase in expression versus relative control). All markers are increased in CD133+ (vs. CD133-) cells and after transfection of unsorted cells with MIR1249 mimic (vs. mimic control). Bars indicate the mean of at least four independent replicates  $\pm$  SEM. \* $P$  < 0.05; \*\* $P$  < 0.01; \*\*\* $P$  < 0.001 versus relative control. (D) CCLP-1 cells were treated with DMSO or CG chemotherapy after transfection with CTRL or MIR1249 inhibitor and assessed for CD133+ by FACS. CG induced enrichment of CD133+ cells, and this enrichment was reduced in the case of transfection with MIR1249 inhibitor. Bars represent the mean of three independent experiments  $\pm$  SD. Representative pictures of FACS analysis are shown on the right. Pink dots represent CD133+ cells. (E) WT and MIR1249KO CCLP-1 cells were treated with DMSO and CG chemotherapy at scalar doses up to 5 M cisplatin and 50 nM gemcitabine. Bars represent the log of the ratio between CG-treated and DMSO-treated cells. Bars indicate the mean of six independent experiments  $\pm$  SD. (F) WT and two clones of MIR1249KO cells were treated with DMSO and CG chemotherapy and assessed for CD133 expression by FACS. Bars represent the mean of three independent experiments  $\pm$  SD. (G) Cells were transfected with CTRL mimic or MIR1249 mimic and treated with DMSO and CG chemotherapy. Bars represent the mean of six independent experiments  $\pm$  SD. \*\*\* $P$  < 0.001 (see also Supporting Fig. S3). Abbreviations: APC-A, allophycocyanin axis; GFP-A, green fluorescent protein axis; LGR5, leucine-rich repeat-containing G protein-coupled receptor 5; SOX2, SRY (sex determining region Y)-box 2.

niche of BTC samples, before and after selection for surface cell markers (Fig. 3A). MIR1249 expression was increased in BTC cells compared with noncancer biliary tract stem cells. CD133+ and CD13+ cells had increased expression of MIR1249 compared with CD133- and CD13 cells (Fig. 3A). CD133 has been consistently reported to be a marker of chemo-resistant cancer cells, which are enriched after treatment,<sup>(22-27)</sup> and CD133+ BTC cells were shown to be tumorigenic and express features of cancer stem cells (CSCs).<sup>(9,28)</sup> Therefore, we speculated that MIR1249 could affect chemo-resistance by inducing the expansion of CD133+ BTC cells. Indeed, an association between CD133 positivity and MIR1249 strong expression was observed in human BTC tissues (Supporting Fig. S2G). Increased MIR1249 expression was confirmed in CCLP-1 CD133+ cells sorted from 2D cultures both by TaqMan assays and ISH (Fig. 3B). CD133+ cells gave rise to spheroids, indicating their self-renewal properties (Fig. 3B-C), and were more resistant to CG chemotherapy when cultured in 2D (Supporting Fig. S2H) or 3D (Fig. 3C-D) in comparison to CD133- cells. Inhibition of MIR1249 in CD133+ CCLP-1 cells increased sensitivity in comparison to CTRL miRNA inhibitor, whereas MIR1249-enforced expression in CD133- cells reduced sensitivity to CG chemotherapy (Fig. 4A and Supporting Fig. S2I). To understand the potential of MIR1249 in controlling the expansion of CD133+ cells, we studied the fraction of CD133+ cells in the presence and absence

of MIR1249 modulation. Enforced expression of MIR1249 in BTC cells expanded the proportion of the CD133+ subpopulation (Fig. 4B), which had increased expression of stem cell markers (Fig. 4C). Indeed, CSC markers were increased in CD133+ versus CD133- cells, and after transfection with MIR1249 mimic compared with mimic control (Fig. 4C). Conversely, inhibition of MIR1249 reduced the CG-induced enrichment of CD133+ cells (Fig. 4D). To confirm the role of MIR1249 in the expansion of CD133+ cells, we generated a MIR1249 knockout (KO) CCLP-1 cell line using CRISPR-associated protein-9 nuclease (CRISPR-CAS9) technologies (Supporting Fig. S3A-C). MIR1249KO cells were more sensitive to CG treatment with a concentration-response effect (Fig. 4E) and showed impaired expansion of CD133+ cells (Supporting Fig. S3D and Fig. 4F), along with reduced expression of CSC markers (Supporting Fig. S3E) and lack of spheroid formation (Supporting Fig. S3F). Reintroduction of MIR1249 in MIR1249KO cells restored chemoresistance in CCLP-1 cells (Fig. 4G and Supporting Fig. S3G).

## MIR1249 DRIVES CLONAL EXPANSION OF CD133+ CELLS BY REWIRING THE Wnt PATHWAY ACTIVATION

To identify the mechanisms through which MIR1249 mediates chemo-resistance, we



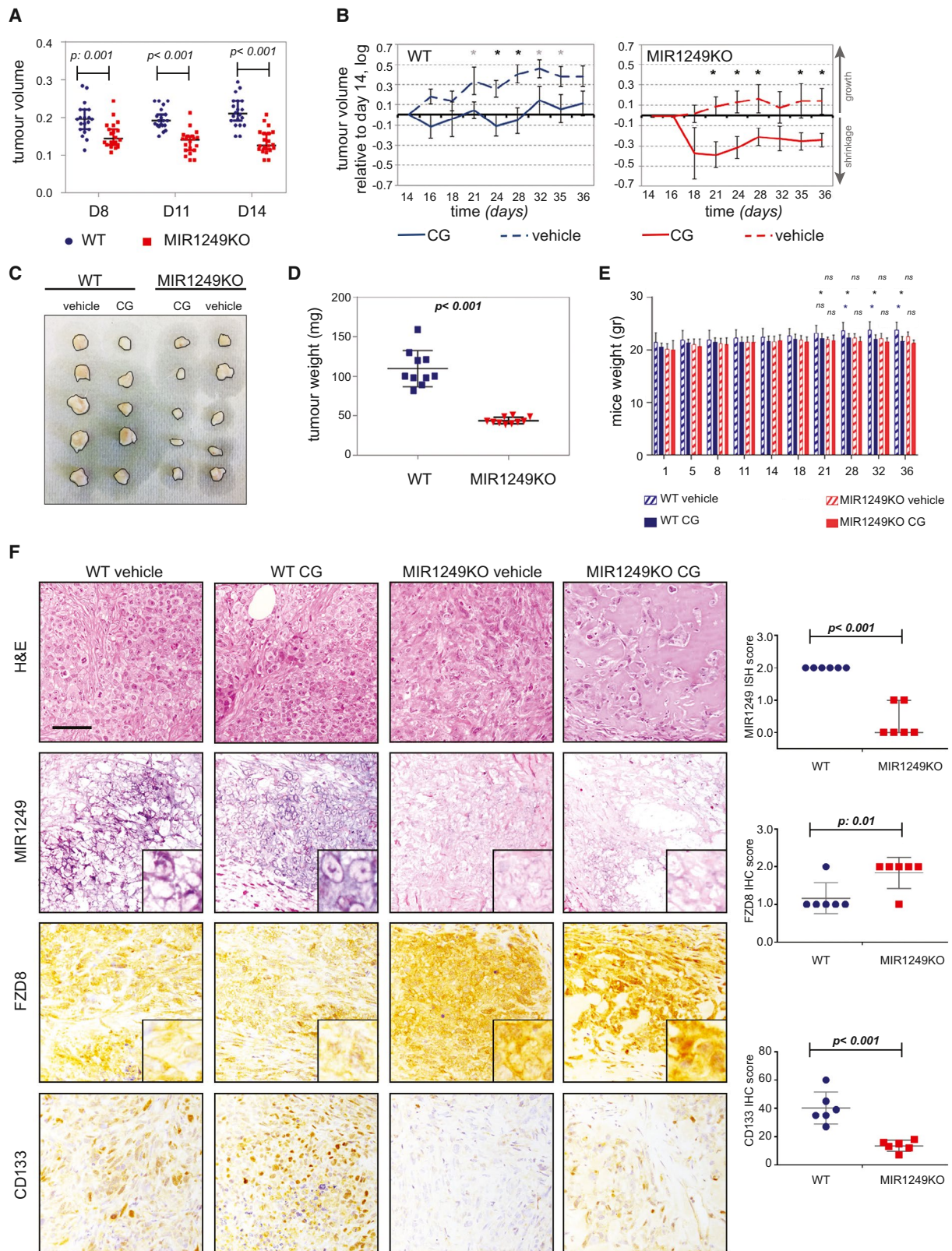
**FIG. 5.** MIR1249 activates the Wnt pathway by acting on FZD8. (A) CCLP-1 cells were sorted for CD133 expression and processed for western blot analysis (left side). CD133<sup>-</sup> cells were also transfected with the indicated probes for 48 hours before proceeding to western blot (right side). (B) CCLP-1 cells were sorted and transfected with nuclear factor of activated T cells (NFAT) and TOP-Flash vector for 48 hours before luciferase activity was recorded. Bars indicate the mean of six independent experiments  $\pm$  SD. Two-way analysis of variance  $< 0.05$ . (C) BTC cells were transfected with pMIR-3UTR-FZD8 vector  $\pm$  MIR1249 mimic, and the luciferase activity was recorded. Bars indicate the mean of six independent experiments  $\pm$  SD. (D) BTC cells were transfected for 48 hours, and positivity for CD133 was assessed by FACS. Bars represent the mean of three independent experiments  $\pm$  SD. (E) CCLP-1 (left) and TFK-1 (right) cells were transfected with small interfering FZD8 or small interfering CTRL for 48 hours and collected for mRNA expression by TaqMan assay. Bars represent the log value of the ratio between siFZD8 and siCTRL. Bars indicate the mean of three independent experiments  $\pm$  SD. \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ . (F) CG-treated BTC cells were transfected for 48 hours, and positivity for CD133 was assessed by FACS. Bars represent the mean of three independent experiments  $\pm$  SD. (G) CCLP-1 cells were infected with the indicated vectors and treated with CG chemotherapy before being fixed in formalin and embedded in paraffin for immunohistochemistry and ISH (see also Supporting Table S4). Scale bars: 100  $\mu$ m. (H) Cells were transfected with NFAT or TOP-Flash vectors, and luciferase activity was recorded after 48 hours. CD133<sup>+</sup> cells were added as controls. Bars indicate the mean of three independent experiments  $\pm$  SD. (I) CCLP-1 cells were transfected for 48 hours and treated with DMSO or increasing doses of CG (up to 5  $\mu$ M cisplatin and 50 nM gemcitabine). Bars represent the log value of the ratio between CG-treated and DMSO-treated cells. Bars indicate the mean of six independent experiments  $\pm$  SD. Abbreviations: LGR5, leucine-rich repeat-containing G protein-coupled receptor 5; siCTRL, small interfering CTRL; siFZD8, small interfering FZD8; SOX2, SRY (sex determining region Y)-box 2.

characterized the gene-expression profiles of chemotherapy-treated CCLP-1 cells after inhibition of MIR1249. Pathway analysis of the deregulated genes showed that MIR1249 inhibition induced changes in the same pathways that were deregulated by chemotherapy. We noticed an enrichment of deregulated genes in the Wnt pathway in both comparisons (chemotherapy vs. vehicle; MIR1249 inhibition vs. no inhibition), suggesting that MIR1249 inhibition may act on the Wnt pathway to restore chemotherapy sensitivity in BTC cells (Supporting Fig. S3H,I). In line with this hypothesis, Wnt deregulation was previously found to drive proliferation of chemo-refractory CSCs.<sup>(29,30)</sup> PANTHER pathway analysis of the predicted targets of MIR1249 based on DIANA software showed an enrichment of Wnt signaling (fold enrichment 1.41;  $p:9.1E-03$ ). *In silico* analyses revealed, among others, frizzled class receptor 8 (FZD8) as a potential mRNA target of MIR1249 (Supporting Fig. S3L). Previous evidence has suggested that FZD8 can act as a negative regulator of the canonical Wnt pathway by activating the noncanonical Wnt/Ca<sup>++</sup> signaling.<sup>(31-34)</sup> Thus, we hypothesized that MIR1249 mediates the expansion of CD133<sup>+</sup> cells by acting on FZD8. Indeed, FZD8 was significantly reduced in CD133<sup>+</sup> in comparison to CD133<sup>-</sup> cells and was associated with inactivation of the noncanonical Wnt and activation of the canonical Wnt pathway (Fig. 5A,B). FZD8 protein expression was reduced in CD133<sup>-</sup> cells transfected with MIR1249 mimic in comparison to CTRL mimic (Fig. 5A), and a luciferase

reporter test confirmed a direct interaction between MIR1249 and the 3' untranslated region (UTR) of FZD8 (Fig. 5C). In human BTC samples (cohort 3) there was a significant inverse relation between MIR1249 and FZD expression (Fisher exact test;  $P = 0.004$ ; Supporting Fig. S3N). Inhibition of FZD8 recapitulated the phenotype induced by MIR1249 mimic (Fig. 5D-F). MIR1249KO cells had increased activation of the noncanonical Wnt pathway (Fig. 5G-H, Supporting Fig. S3M, and Supporting Table S4), and FZD8 inhibition in these cells partially increased resistance to CG (Fig. 5I).

### **IN VIVO VALIDATION OF THE MIR1249-DEPENDENT CHEMO-RESISTANCE IN MURINE TUMORS BEARING DISRUPTION OF MIR1249**

MIR1249KO cells had reduced *in vivo* tumorigenicity, confirming the role of MIR1249 in driving the expansion of CSCs (Fig. 6A and Supporting Fig. S4A). Chemotherapy sensitivity was increased in mice bearing MIR1249KO tumor xenografts. Indeed, the weekly combination of CG could induce tumor shrinkage in MIR1249KO xenografts, while causing only tumor stabilization in wild-type (WT) xenografts (Fig. 6B-D and Supporting Table S5). The chemotherapy schedule was well-tolerated with only minimal changes in weights for CG-treated mice at the end of the treatment course. No differences in weights were observed between WT and MIR1249KO CG-treated



**FIG. 6.** Lack of MIR1249 increases sensitivity to chemotherapy *in vivo*. (A) WT or MIR1249KO CCLP-1 cells were injected subcutaneously in the flank of NSG (nonobese diabetic scid gamma) mice (n = 20 each) and monitored for growth by caliper (see also Supporting Fig. S4). (B) At day 14, mice were randomized to be treated with a weekly combination of intraperitoneal gemcitabine (150 mg/kg) and cisplatin (2 mg/kg) or vehicle alone for 3 weeks before being sacrificed. Data are presented normalized to baseline pretreatment tumor size (day 14). Black \* indicates *P* value < 0.05; gray \* indicates *P* value between 0.05 and 0.085 (see also Supporting Table S5). (C) Representative pictures of five explanted tumors per group. (D) Explanted tumors of CG-treated mice were weighted before being stored for analyses. Error bars indicate the mean with SEM. (E) Mice were weighted periodically over the course of treatment. Bars represent the mean of 10 mice  $\pm$  SD. \**P* < 0.05. (F) Representative pictures of MIR1249 ISH staining and immunohistochemistry staining for the indicated proteins performed on the explanted tumors, along with quantitative analysis. Scale bars: 100  $\mu$ m. Three mice per group were analyzed (for a total of 12), and CG-treated and vehicle-treated tumors were grouped to assess the differences between KO and WT. Abbreviations: H&E, hematoxylin and eosin; ns, not significant.

mice, suggesting that the drug exposure was comparable among the two groups (Fig. 6E). In addition, no differences in liver and kidney toxicity were observed between WT and MIR1249KO mice when treated with CG (Supporting Fig. SF4B-C). To confirm the role of MIR1249 in driving the expansion of CD133+ cells through FZD8, we assessed the protein expression in explanted tumors from MIR1249KO versus WT xenograft (either treated with CG and vehicle) and observed that MIR1249KO tumors showed a lack of MIR1249 expression and reduced expression of CD133, with increase in FZD8 expression (Fig. 6F).

## Discussion

Therapeutic development for BTCs remains an unmet need. Chemotherapy does represent the main backbone for advanced biliary cancer (ABC) treatment, even though the response rate observed in patients with ABC does not exceed 25%. Strategies aimed at improving the efficacy of chemotherapy might prove beneficial to a large proportion of patients with ABC. The field of miRNA-based therapeutics has recently expanded and entered the phase of clinical investigation.<sup>(10,35,36)</sup> The ability of miRNAs to target multiple pathways is attractive, as it may prevent the onset of compensatory pathways.

Data from the MesomiR1 trial have shown the feasibility of a therapeutic approach based on miRNA replacement in human cancer patients.<sup>(37)</sup> However, this approach holds two major limitations: (1) toxicity related to the immuno-stimulatory effects of encapsulating delivery systems and (2) off-target effects induced by a disproportionately high level of miRNA in the cellular system. Conversely, an approach based on the inhibition of miRNAs would reduce the risk of off-target effects by affecting the physiological level of

a miRNA rather than introducing a perturbation that affects cellular homeostasis. Recent technologies have enabled chemical modifications of anti-miRNAs (i.e., the addition of LNA) that increase their stability,<sup>(38)</sup> allowing the clinical investigation of this therapeutic strategy in cancer patients (NCT02580552). In these studies, we have identified MIR1249 as a miRNA that drives the emergence of chemo-resistance by acting on the CD133+ cell population. Although we usually observe tumor stabilization with chemotherapy in patients with BTCs, our data suggest that the addition of MIR1249 inhibition to CG can increase tumor responses *in vivo*. It is known that partial responses are associated with prolonged life expectancy in patients with BTCs; therefore, we speculate that treatment with MIR1249 inhibitors might prove beneficial to affect the survival of patients with BTCs by preventing primary resistance and delaying the onset of secondary resistance. It has already been shown that the canonical Wnt/ $\beta$ -catenin signaling mediates self-renewal of stem cells, whereas noncanonical Wnt signaling pathways is involved in the maintenance of stem cells, cell plasticity, and inhibition of the canonical Wnt signaling cascade,<sup>(29)</sup> supporting our data that MIR1249 can drive maintenance and expansion of CSCs through regulation of the noncanonical Wnt pathway. With regard to FZD8, so far, data in the literature are contradictory, with some reports showing the capacity of FZD8 to stimulate malignant transformation of cancer cells,<sup>(39)</sup> and others showing its involvement in reducing tumor initiating capacity.<sup>(32)</sup> In line with our data that showed low expression of FZD8 in CSCs, analysis of the TCGA data sets showed unfavorable prognosis in the cases of pancreatic cancers with low FZD8 expression. Nonetheless, attempts to therapeutically inhibit FZD8 have failed so far due to lack of therapeutic index, suggesting that FZD8 may not be involved in promoting cancer growth.

Finally, in this study the combination of CG was given in a weekly schedule to BTC mouse models in an attempt to better mimic the schedule used in the ABC-02 trial, which licensed the combination CG for standard clinical practice. We noticed good tolerance of this schedule and suggest that this regimen be used in future *in vivo* BTC modeling, to increase the clinical relevance of preclinical findings.

In conclusion, we have provided evidence for a potential target to be considered in future therapeutic development. Our data suggest that MIR1249 is involved in the chemo-resistance in all different subtypes of CCA; therefore, we suggest testing the MIR1249 inhibitor in a trial including ABCs. In addition, it may be speculated that this mechanism may be shared with other cancer types as well, which may warrant investigation in other solid tumors.

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## Supporting Information

Additional Supporting Information may be found at [onlinelibrary.wiley.com/doi/10.1002/hep.31094/supinfo](http://onlinelibrary.wiley.com/doi/10.1002/hep.31094/supinfo).