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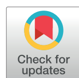


Corrigendum

# Corrigendum to "Gap junctional communication is involved in differentiation of osteoclasts from bone marrow and peripheral blood monocytes"

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In the original published version of this article, a typographical error was present in Methods section 2.6. The pixel size displayed incorrectly as "354 μm × 354 μm" instead of the correct value of "0.69 μm × 0.69 μm". The authors apologize for the mistake. The corrected version of section 2.6 is displayed below.

## 2.6. Analysis of the effects of AAP10 on GJC with Fluorescence Recovery After Photobleaching (FRAP)

Osteoblast lineage MC3T3-E1 cells were obtained from LGC Standards (Teddington, UK). The cells were cultured in α-MEM containing 10% FBS, 2 mM L-glutamine, 100 IU/ml penicillin and 100 μg/ml streptomycin (Sigma-Aldrich) in T25 cell culture flasks and passaged every 2−3 days. For the FRAP experiment, $3.95 \times 10^4$ cells/cm$^2$ were cultured in 4 compartment glass bottom cell culture dishes (Cellview; Greiner Bio-One, Kremsmünster, Austria) in 500 μl α-MEM per compartment. 100 μM AAP10 was added to the cultures once daily and finally after staining with calcein red-orange. The cells were cultured for 3 days, after which the media were removed and replaced with phenol red free D-MEM (Gibco/Thermo Fisher Scientific, Waltham, MA) containing 5 μM calcein red-orange, AM

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(Molecular Probes/Thermo Fisher Scientific). Staining was performed at +37 °C for 30 minutes after which the cells were washed 3 times with phenol red free D-MEM. FRAP experiments were performed with a Zeiss LSM 780 confocal microscope using a Plan-Apochromat 20x/0.8 objective (Carl Zeiss Microscopy GmbH, Germany). The excitation wavelength was 561 nm and emission was collected between 566 nm - 670 nm. The pixel size was set to 0.69 μm × 0.69 μm. The time-lapse imaging interval was set to 5 seconds and recovery of the fluorescence was followed for 280 seconds. All measurements were performed at +37 °C and 5% $CO_2$ maintained by an OkoLab bold line top stage incubator (OkoLab, Italy). Before analyzing the results, the fluorescence signals were corrected for acquisition bleaching and normalized by scaling the pre-bleach intensity values to 1, and the post-bleach values were scaled to 0.


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