GENOME SEQUENCES

Complete Genome Sequence of Rahnella aquatilis MEM40, a Plant Growth-Promoting Rhizobacterium Isolated from Rice Rhizosphere Soil, with Antagonism against Magnaporthe oryzae and Fusarium graminearum

Xiaowen Sun,a,b Wei Ma,a Yin Xu,a Xinmeng Jin,a [Hong Nia](https://orcid.org/0000-0002-4363-6987)

aState Key Laboratory of Biocatalysis and Enzyme Engineering, Hubei Collaborative Innovation Center for Green Transformation of Bio-resources, School of Life Sciences, Hubei University, Wuhan, People's Republic of China

^bState Key Laboratory of Agricultural Microbiology, Huazhong Agricultural University, Wuhan, People's Republic of China

ABSTRACT Rahnella aquatilis strain MEM40 is a plant growth-promoting rhizobacterium (PGPR) with antagonism against Magnaporthe oryzae and Fusarium graminearum that was isolated from rice rhizosphere soil in Hubei, China. Here, we report its complete genome sequence, which will increase our understanding of the mechanisms of plant growth promotion and biocontrol.

**Rahnella aquatilis is a Gram-negative facultative anaerobic nonpigmented rod-
Shaped bacterium belonging to the** *Enterobacteriaceae* **that has been found in a** variety of environments [\(1,](#page-1-0) [2\)](#page-1-1). This species can act as a plant growth-promoting rhizobacterium (PGPR) to provide nutrients for plant development [\(3,](#page-1-2) [4\)](#page-2-0) and can also be used as a biological control agent against some plant diseases, including grapevine crown gall [\(5\)](#page-2-1), apple fire blight [\(6\)](#page-2-2), and fruit storage rots [\(7\)](#page-2-3). Strain MEM40 was isolated from rice-wheat rotation soil (Hubei, China) using the 10-times dilution coating method on Pikovskaya solid medium. The single colony with a transparent zone was grown in Luria-Bertani medium overnight at 28°C. PCR was performed using the universal primers 27F (5'-AGAGTTTGATCCTGGCTCAG-3') and 1492R (5'-TACGGCTACCTTGTACG ACTT-3'). Phylogenetic analysis of the 16S rRNA gene identified MEM40 as an R. aquatilis strain. MEM40 possessed phosphate-solubilizing capacity [\(Fig. 1A\)](#page-0-0) and antag-

FIG 1 Phosphate-solubilizing and antifungal capacities of MEM40. (A) Phosphate-solubilizing capacity of MEM40 in Pikovskaya liquid medium [10.0 g glucose, 5.0 g Ca₃(PO₄)₂, 0.5 g (NH₄)₂SO₄, 0.2 g NaCl, 0.1 g MgSO₄·7H₂O, 0.2 g KCl, 0.5 g yeast extract, 0.002 g MnSO₄·H₂O, and 0.002 g FeSO₄·7H₂O, pH 7.0 to 7.4] (the maximum phosphate-solubilizing capacity reached 411.56 mg/liter). The inset shows a transparent zone of MEM40 on Pikovskaya solid medium. (B) Antifungal test of MEM40 against M. oryzae, the causal agent of rice blast disease [\(13\)](#page-2-4), and F. graminearum, the causal agent of wheat Fusarium head blight [\(14\)](#page-2-5), on peptonedextrose agar (PDA) medium (200.0 g potato, 15.0 g glucose, and 15.0 g agar, pH 7.0 to 7.4).

Citation Sun X, Ma W, Xu Y, Jin X, Ni H. 2020. Complete genome sequence of Rahnella aquatilis MEM40, a plant growth-promoting rhizobacterium isolated from rice rhizosphere soil, with antagonism against Magnaporthe oryzae and Fusarium graminearum. Microbiol Resour Announc 9:e00651-20. [https://doi.org/](https://doi.org/10.1128/MRA.00651-20) [10.1128/MRA.00651-20.](https://doi.org/10.1128/MRA.00651-20)

Editor Jason E. Stajich, University of California, Riverside

Copyright © 2020 Sun et al. This is an openaccess article distributed under the terms of the [Creative Commons Attribution 4.0](https://creativecommons.org/licenses/by/4.0/) [International license.](https://creativecommons.org/licenses/by/4.0/)

Address correspondence to Xiaowen Sun, [sunxiaowen525@sina.com,](mailto:sunxiaowen525@sina.com) or Hong Ni, [nh64@sina.com.](mailto:nh64@sina.com)

Received 16 June 2020 **Accepted** 17 August 2020 **Published** 10 September 2020

TABLE 1 CRISPR and GI positions of the MEM40 genome

onistic effects against Magnaporthe oryzae and Fusarium graminearum [\(Fig. 1B\)](#page-0-0). The complete genome sequence of MEM40 may be used in comparative genomics, functional gene searches, and engineering bacteria in the future.

A single colony was grown in Luria-Bertani medium overnight at 28°C. Genomic DNA was extracted using a bacterial genomic DNA extraction kit (Solarbio, Beijing, China) and sheared into ~10-kb fragments with a gTUBE device (Covaris, USA). After single-strand overhang removal, DNA damage repair, end repair, and 3' adenylation, the fragments were linked with single-molecule real-time (SMRT) adaptors using the SMRTbell Express template preparation kit v2.0 (Pacific Biosciences, USA) and purified with AMPure PB beads. DNA concentration and library fragment size were measured with Qubit v3.0 and the Agilent 2100 bioanalyzer system. The library was sequenced with the Sequel sequencing kit v2.1, and sequencing data were processed using SMRT Link v5.0 [\(8,](#page-2-6) [9\)](#page-2-7). The read library contained 2,306,916,870 bp with an average subread length of 8,183 bp, a subread N_{50} value of 10,647 bp, and an average coverage of 459X. HGAP4 software (HGAP genome length, 5,500,000 bp; HGAP seed coverage, 30; HGAP seed length cutoff, –1; minimum confidence, 40; minimum coverage, 5) [\(10\)](#page-2-8) and Canu v1.6 (genome size, 5.5 Mbp; corrected error rate, 0.02) were used for assembly and self-correction. Minimus2 software (with the parameters overlap, 40 bp; conserr, 0.06; minid, 94; and maxtrim, 20 bp) was used for cyclization. The MEM40 genome consisted of a circular chromosome and two circular plasmids, and the genome size was 5,596,168 bp with a subread N_{50} value of 5,596,168 and G+C content of 52.2%. The publicly available sequence was annotated using NCBI PGAP. The clustered regularly interspaced short palindromic repeat (CRISPR) structures and genomic islands (GIs) were predicted using CRT [\(11\)](#page-2-9) and IslandPath [\(12\)](#page-2-10) software, respectively.

The circular chromosome consisted of 5,021,016 bp (G+C content, 52.3%) containing 4,467 protein-coding genes, 45 pseudogenes, 22 rRNA genes, 77 tRNA genes, 6 CRISPRs, and 11 GIs [\(Table 1\)](#page-1-3). The circular megaplasmid pMEM40-1 consisted of 525,842 bp (G+C content, 52.1%) containing 470 protein-coding genes, 8 pseudogenes, and 1 CRISPR [\(Table 1\)](#page-1-3). The circular plasmid pMEM40-2 consisted of 49,310 bp $(G+C$ content, 37.4%) containing 27 protein-coding genes and 3 pseudogenes.

Data availability. The genome assembly has been deposited in GenBank under the accession number [GCA_004328445.1.](https://www.ncbi.nlm.nih.gov/assembly/GCF_004328445.1/) The complete genome sequence of MEM40 has been deposited in GenBank under the main chromosome accession number [CP036490.1,](https://www.ncbi.nlm.nih.gov/nuccore/CP036490.1) and the sequences of the plasmids pMEM40-1 and pMEM40-2 have been deposited under accession numbers [CP036488.1](https://www.ncbi.nlm.nih.gov/nuccore/CP036488.1) and [CP036489.1,](https://www.ncbi.nlm.nih.gov/nuccore/CP036489.1) respectively. Sequence data have been deposited in the Sequence Read Archive under the accession number [SRP267417.](https://www.ncbi.nlm.nih.gov/sra/SRP267417)

ACKNOWLEDGMENT

This work was supported by grants from the National Key R&D Program of China (numbers 2018YFD0200500 and 2018YFD0200506).

REFERENCES

- 1. Izard D, Gavini F, Trinel P, Leclere H. 1979. Rahnella Aquatilis, a new member of the Enterobacteriaceae. Ann Microbiol (Paris) 130:163–177. (In French.)
- 2. Yuan L, Li L, Zheng F, Shi Y, Xie X, Chai A, Li B. 2020. The complete

genome sequence of Rahnella aquatilis ZF7 reveals potential beneficial properties and stress tolerance capabilities. Arch Microbiol 202:483– 499. [https://doi.org/10.1007/s00203-019-01758-1.](https://doi.org/10.1007/s00203-019-01758-1)

3. Li L, Jiao Z, Hale L, Wu W, Guo Y. 2014. Disruption of gene pqqA or pqqB

reduces plant growth promotion activity and biocontrol of crown gall disease by Rahnella aquatilis HX2. PLoS One 9:e115010. [https://doi.org/](https://doi.org/10.1371/journal.pone.0115010) [10.1371/journal.pone.0115010.](https://doi.org/10.1371/journal.pone.0115010)

- 4. Vyas P, Joshi R, Sharma KC, Rahi P, Gulati A, Gulati A. 2010. Cold-adapted and rhizosphere-competent strain of Rahnella sp. with broad-spectrum plant growth-promotion potential. J Microbiol Biotechnol 20: 1724 –1734.
- 5. Chen F, Guo YB, Wang JH, Li JY, Wang HM. 2007. Biological control of grape crown gall by Rahnella aquatilis HX2. Plant Dis 91:957–963. [https://](https://doi.org/10.1094/PDIS-91-8-0957) [doi.org/10.1094/PDIS-91-8-0957.](https://doi.org/10.1094/PDIS-91-8-0957)
- 6. Abo-Elyousr A, Sallam M, Zeller W. 2011. Effect of acibenzolar-S-methyl and Rahnella aquatilis (Ra39) on fire blight of apple plants. Acta Hortic 896:511–518. [https://doi.org/10.17660/ActaHortic.2011.896.75.](https://doi.org/10.17660/ActaHortic.2011.896.75)
- 7. Navarta LG, Calvo J, Calvente V, Benuzzi D, Sanz MI. 2011. Freezing and freeze-drying of the bacterium Rahnella aquatilis BNM 0523: study of protecting agents, rehydration media and freezing temperatures. Lett Appl Microbiol 53:565–571. [https://doi.org/10.1111/j.1472-765X.2011.03150.x.](https://doi.org/10.1111/j.1472-765X.2011.03150.x)
- 8. Kingan SB, Heaton H, Cudini J, Lambert CC, Baybayan P, Galvin BD, Durbin R, Korlach J, Lawniczak MKN. 2019. A high-quality de novo genome assembly from a single mosquito using PacBio sequencing. Genes (Basel) 10:62. [https://doi.org/10.3390/genes10010062.](https://doi.org/10.3390/genes10010062)
- 9. Rhoads A, Au KF. 2015. PacBio sequencing and its applications. Genomics Proteomics Bioinformatics 13:278 –289. [https://doi.org/10.1016/j.gpb](https://doi.org/10.1016/j.gpb.2015.08.002) [.2015.08.002.](https://doi.org/10.1016/j.gpb.2015.08.002)
- 10. Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. [https://doi.org/10.1038/](https://doi.org/10.1038/nmeth.2474) [nmeth.2474.](https://doi.org/10.1038/nmeth.2474)
- 11. Bland C, Ramsey TL, Sabree F, Lowe M, Brown K, Kyrpides NC, Hugenholtz P. 2007. CRISPR recognition tool (CRT): a tool for automatic detection of clustered regularly interspaced palindromic repeats. BMC Bioinformatics 8:209. [https://doi.org/10.1186/1471-2105-8-209.](https://doi.org/10.1186/1471-2105-8-209)
- 12. Hsiao W, Wan I, Jones SJ, Brinkman FS. 2003. IslandPath: aiding detection of genomic islands in prokaryotes. Bioinformatics 19:418 – 420. [https://](https://doi.org/10.1093/bioinformatics/btg004) [doi.org/10.1093/bioinformatics/btg004.](https://doi.org/10.1093/bioinformatics/btg004)
- 13. Wilson RA, Talbot NJ. 2009. Under pressure: investigating the biology of plant infection by Magnaporthe oryzae. Nat Rev Microbiol 7:185–195. [https://doi.org/10.1038/nrmicro2032.](https://doi.org/10.1038/nrmicro2032)
- 14. Figueroa M, Hammond-Kosack KE, Solomon PS. 2018. A review of wheat diseases: a field perspective. Mol Plant Pathol 19:1523–1536. [https://doi](https://doi.org/10.1111/mpp.12618) [.org/10.1111/mpp.12618.](https://doi.org/10.1111/mpp.12618)