Colesevelam enhances the beneficial effects of brown fat activation on hyperlipidaemia and atherosclerosis development

Enchen Zhou^{1,2†}, Geerte Hoeke^{1,2†}, Zhuang Li^{1,2}, Arthur C. Eibergen ^{1,2}, Amber W. Schonk ^{1,2}, Martijn Koehorst ³, Renze Boverhof³, Rick Havinga⁴, Folkert Kuipers^{3,4}, Tamer Coskun⁵, Mariëtte R. Boon^{1,2}, Albert K. Groen^{3,4,6}, Patrick C.N. Rensen ^{1,2}, Jimmy F.P. Berbée^{1,2}, and Yanan Wang ^{1,2}*

¹Division of Endocrinology, Department of Medicine, Leiden University Medical Center, PO Box 9600, 2300 RC Leiden, The Netherlands; ²Einthoven Laboratory for Experimental Vascular Medicine, Leiden University Medical Center, Leiden, The Netherlands; ³Department of Laboratory Medicine, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands; ⁴Department of Pediatrics, University of Groningen, University Medical Center Groningen, Groningen, The Netherlands; ⁵Department of Diabetes/ Endocrine, Lilly Research Laboratories, Lilly Corporate Center, Indianapolis, IN, USA; and ⁶Department of Vascular Medicine, Academic Medical Center, Amsterdam, The Netherlands

Received 10 April 2019; revised 28 August 2019; editorial decision 16 September 2019; accepted 30 September 2019; online publish-ahead-of-print 7 October 2019

Time for primary review: 43 days

Aims

Brown fat activation accelerates the uptake of cholesterol-enriched remnants by the liver and thereby lowers plasma cholesterol, consequently protecting against atherosclerosis development. Hepatic cholesterol is then converted into bile acids (BAs) that are secreted into the intestine and largely maintained within the enterohepatic circulation. We now aimed to evaluate the effects of prolonged brown fat activation combined with inhibition of intestinal BA reabsorption on plasma cholesterol metabolism and atherosclerosis development.

Methods and results

APOE*3-Leiden.CETP mice with humanized lipoprotein metabolism were treated for 9 weeks with the selective β 3-adrenergic receptor (AR) agonist CL316,243 to substantially activate brown fat. Prolonged β 3-AR agonism reduced faecal BA excretion (-31%), while markedly increasing plasma levels of total BAs (+258%), cholic acid-derived BAs (+295%), and chenodeoxycholic acid-derived BAs (+217%), and decreasing the expression of hepatic genes involved in BA production. In subsequent experiments, mice were additionally treated with the BA sequestrant Colesevelam to inhibit BA reabsorption. Concomitant intestinal BA sequestration increased faecal BA excretion, normalized plasma BA levels, and reduced hepatic cholesterol. Moreover, concomitant BA sequestration further reduced plasma total cholesterol (-49%) and non-high-density lipoprotein cholesterol (-56%), tended to further attenuate atherosclerotic lesion area (-54%). Concomitant BA sequestration further increased the proportion of lesion-free valves (+34%) and decreased the relative macrophage area within the lesion (-26%), thereby further increasing the plaque stability index (+44%).

Conclusion

BA sequestration prevents the marked accumulation of plasma BAs as induced by prolonged brown fat activation, thereby further improving cholesterol metabolism and reducing atherosclerosis development. These data suggest that combining brown fat activation with BA sequestration is a promising new therapeutic strategy to reduce hyperlipidaemia and cardiovascular diseases.

Keywords

Brown adipose tissue • Bile acid metabolism • Cholesterol turnover • Hyperlipidaemia • Atherosclerosis

1. Introduction

Atherosclerosis represents the most common cause of cardiovascular diseases. A prominent risk factor for atherosclerosis is hyperlipidaemia,

i.e. high levels of low-density lipoprotein cholesterol (LDL-C) and trigly-cerides (TG) in the circulation. Currently, reducing circulating atherogenic lipoproteins with lipid-lowering medication, such as statins and PCSK9 inhibitors, remains the major strategy to prevent acute

^{*} Corresponding author. Tel: +31 71 52 68176; fax: +31 71 52 66881, E-mail: Y.Wang@lumc.nl

 $^{^\}dagger$ The first two authors contributed equally to this work.

[©] The Author(s) 2019. Published by Oxford University Press on behalf of the European Society of Cardiology. This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

cardiovascular events. However, only 30% of all cardiovascular events can be prevented by such treatment strategies, ¹ illustrating the need for new therapeutic strategies.

Brown fat is present in mammals as well as in (adult) humans and is an emerging target to combat hyperlipidaemia and atherosclerosis.^{2–4} Cold exposure, the best known physiological activator of brown fat, leads to the release of noradrenalin from sympathetic nerves that innervate brown fat. Noradrenalin binds to the β 3-adrenergic receptor (β 3-AR) on brown adipocytes, resulting in their activation to produce heat.⁵ As the β3-AR is highly expressed on brown and white adipocytes, and white fat does not substantially contribute to energy expenditure, the coldstimulated activation of brown fat inducing thermogenesis can be pharmacologically mimicked by selective β3-AR agonists such as CL316,243 compound, one of the most selective β 3-AR agonists available.⁶ Since heat generation is an energy consuming process, activated brown fat takes up large amounts of nutrients from the circulation, mainly TGderived fatty acids from TG-rich lipoproteins [TRLs; i.e. very-low-density lipoproteins (VLDL) and chylomicrons]. As a result, brown fat activation accelerates the formation and uptake of cholesterol-enriched lipoprotein remnants by the liver, thereby protecting from hyperlipidaemia and atherosclerosis development. In addition to reducing cholesterolenriched TRL remnant levels, β3-AR agonism also improves high-density lipoprotein (HDL) functionality as reflected by increased reverse cholesterol transport (RCT). 7,9 Collectively, these studies show that β 3-AR agonism increases cholesterol delivery towards the liver via both accelerating the clearance of cholesterol-enriched TRL remnants and improving HDL-mediated RCT.

Hepatic cholesterol turnover is mainly mediated by faecal excretion as bile acids (BAs) and, to a lesser extent, by faecal excretion of neutral sterols. 10 Hepatocytes synthesize primary BAs, i.e. cholic acid (CA) and chenodeoxycholic acid (CDCA), via the so-called classic pathway; while CDCA can also be synthesized via an alternative pathway. ¹¹ In mice, but not in humans, CDCA can be converted into more hydrophilic species, the so-called muricholic acids (MCAs).¹² Newly synthesized BAs are temporarily stored in the gallbladder and are secreted into the duodenum upon food ingestion to serve as detergents for absorption of nutrients. 13 By the enzymatic action of gut bacteria, part of the primary BAs are converted into secondary BAs. In the terminal ileum, 95% of BAs are reabsorbed by active transport with remaining BAs excreted in faeces. Reabsorbed BAs mainly circulate back through the portal vein to the liver completing one cycle of enterohepatic circulation. ¹⁴ BA synthesis is regulated by enterohepatic circulation of BAs via farnesoid X receptor (FXR), which inhibits transcription of genes in BA synthesis. 15 The enterohepatic circulation can be interrupted by BA sequestrants, which increases clearance of plasma (V)LDL-C by promoting conversion of hepatic cholesterol into BAs and up-regulation of hepatic LDL receptors. In fact, BA sequestrants such as Cholestyramine and Colesevelam have been proven to be effective to treat dyslipidaemia and prevent cardiovascular diseases. 16

Interestingly, both short-term and long-term activation of brown fat increases cholesterol delivery towards the liver. 7.17 Moreover, we previously showed that long-term activation of brown fat significantly increased hepatic cholesterol accumulation. 17 Since BA synthesis is the main pathway for hepatic cholesterol catabolism, short-term activation of brown fat indeed has been linked to increased BA production and increased faecal BA excretion. 18 However, how long-term brown fat activation influences BA metabolism and whether manipulation of BA metabolism on top of brown fat activation would lead to additional benefits on cholesterol metabolism and atherosclerosis development has

not been studied yet. Thus, the aim of the current study was to evaluate the effects of prolonged brown fat activation via $\beta 3$ -AR agonism on cholesterol and BA metabolism. In addition, we assessed whether inhibiting intestinal BA reabsorption beneficially influences the effects of prolonged $\beta 3$ -AR agonism on cholesterol turnover and atherosclerosis development. To this end, we treated hyperlipidaemic APOE*3-Leiden(E3L).CETP mice, a well-established model for human-like lipoprotein metabolism and atherosclerosis, 19,20 with or without the selective $\beta 3$ -AR agonist CL316,243 to activate brown fat for 9 weeks. In subsequent experiments, E3LCETP mice were treated with vehicle, CL316,243 alone, the BA sequestrant Colesevelam alone to inhibit intestinal BA reabsorption, or the combination of both for a period of 4 or 12 weeks.

2. Methods

Detailed description of the Methods section is available in the Supplementary material online.

2.1 Animals and treatments

Hemizygous APOE*3-Leiden (E3L) mice were crossbred with homozygous human cholesteryl ester transfer protein (CETP) transgenic mice to generate heterozygous E3LCETP mice. At the age of 10–12 weeks, female mice were fed a Western-type diet (WTD; Altromin, Germany) containing 15% cacao butter, 1% corn oil, and 0.15% (w/w) cholesterol.

In a first experiment, ¹⁷ mice were randomized into two groups after a run-in period of 6 weeks on WTD. Mice were subsequently treated 5 days/week with the selective β 3-AR agonist CL316,243 (symbol: β ; Tocris Bioscience Bristol, UK; 20 μ g·mouse⁻¹) or vehicle (phosphate-buffered saline, symbol: –) by subcutaneous injections between 14:00 and 16:00 h for an additional 9 weeks.

In a second experiment, mice were randomized into two groups after a run-in period of 3 weeks on WTD and subsequently received WTD supplemented without or with 0.15% (w/w) Colesevelam (symbol: c; Genzyme Europe B.V., The Netherlands). After an additional run-in period of 3 weeks, mice in each treatment group were again randomized into two subgroups and additionally treated 5 days/week with vehicle or CL316,243 by subcutaneous injection for additional 4 weeks. This resulted in the following four treatment groups: (i) vehicle (–), (ii) CL316,243 (β), (iii) Colesevelam (c), and (iv) Colesevelam + CL316,243 (α).

In a third experiment, the set-up was similar to the second experiment, with the exception that mice were treated with CL316,243 or vehicle for 12 weeks.

Food intake and body weight were monitored weekly. Body composition (i.e. body fat and lean mass; EchoMRI-100; EchoMRI, Houston, TX, USA) was evaluated every 2 weeks. At the end of each experiment, mice were euthanatized by CO_2 suffocation and unconscious mice were perfused with ice-cold saline via cardiac perfusion, and various organs were isolated for further analysis.

These animal experiments were approved by the Animal Ethical Committee of Leiden University Medical Center, Leiden, The Netherlands (DEC 12252-02). All animal procedures were performed conform to the guidelines from Directive 2010/63/EU of the European Parliament on the protection of animals used for scientific purposes.

2.2 Faecal and plasma bile acid analysis

Faeces were collected over a 24-h period and dried at room temperature, weighed, and homogenized. BA composition was determined in an

aliquot of faeces by gas—liquid chromatography (GC). Plasma BA profile was measured using liquid chromatography tandem MS (LC-MS/MS).

2.3 Biliary bile acid collection and composition analysis

Mice were anaesthetized by intraperitoneal injection with Hypnorm (1 mL·kg⁻¹; Janssen Pharmaceuticals) and Diazepam (10 mg·kg⁻¹; Actavis). The bile duct was ligated and the gallbladder was cannulated to collect BAs. Hepatic bile was collected for 15 min and the average of bile flow per minute was calculated. BA compositions were determined in 5 μ L bile by GC as described above. Biliary cholesterol levels were determined using an enzymatic kit from Roche Diagnostics (Mannheim, Germany).

2.4 Gene expression analysis

Gene expression analysis was performed as described in the Supplementary material online. The primer sequences used are listed in Supplementary material online, *Table S1*.

2.5 Plasma lipid assays and lipoprotein profiles

Plasma was assayed for TG and total cholesterol (TC) using enzymatic kits from Roche Diagnostics (Mannheim, Germany) as described in the Supplementary material online. The distribution of TG and cholesterol over lipoproteins was determined in pooled plasma by fast-performance liquid chromatography (FPLC) using a Superose 6 column (GE Healthcare, Piscataway, NJ, USA).

2.6 Hepatic lipid content

Liver lipids were assayed as described in the Supplementary material online.

2.7 *In vivo* plasma decay and hepatic uptake of TG-rich lipoprotein-like particles

TRL-like particles (80 nm), double-labelled with glycerol tri[3 H]oleate ([3 H]TO) and [14 C]cholesteryl oleate ([14 C]CO), were prepared as described previously. 22 Mice were fasted for 4 h and injected (t=0) intravenously with 200 μ L of TRL-like particles (1 mg TG per mouse). Blood samples were taken from the tail vein at 2, 5, 10, and 15 min after injection to determine the plasma decay of [3 H]TO and [14 C]CO. After 15 min, livers were isolated and weighted, and 3 H- and 14 C-activity were quantified.

2.8 Atherosclerosis quantification

Hearts were collected, fixed in phosphate-buffered 4% formaldehyde, and embedded in paraffin. Four sections of the aortic root area with 50 µm intervals were used and stained with haematoxylin—phloxine—saffron for histological analysis. Lesions were categorized for lesion severity according to the guidelines of the American Heart Association adapted for mice²³ and classified as mild lesions (types 1–3) and severe lesions (types 4–5). Monoclonal mouse antibody M0851 against smooth muscle cell (SMC) actin was used to quantify the SMC area, Sirius Red staining was used to quantify the collagen area, and rat monoclonal antibody MAC3 was used to quantify macrophage area as described.¹⁷ Lesion area was determined with Image J Software (version 1.50i).

2.9 Statistical analysis

Differences between two groups were determined using the unpaired two-tailed Student's *t*-test. Differences between four groups were determined using one-way analysis of variance (ANOVA) with the LSD *post hoc* test, which however increases the alpha risk as it does not correct for multiple comparisons. The square root of the lesion area was transformed to linearize the relationship with the plasma TC exposure. Univariate regression of analyses was performed to test for significant correlations between atherosclerotic lesion area and plasma TC exposure. Multiple regression analysis was performed to predict the contribution of plasma TC exposures to the atherosclerotic lesion area. Probability values less than 0.05 were considered statistically significant. All statistical analyses were performed with the GraphPad Prism 7 for Windows.

3. Results

3.1 Prolonged β 3-AR agonism decreases faecal bile acid excretion and increases plasma bile acid levels

We previously fed female E3LCETP mice a WTD and treated them with the β 3-AR agonist CL316,243 (β) or vehicle (–) for 9 weeks, and observed that prolonged β3-AR agonism significantly increases liver TC levels.¹⁷ Since BA synthesis is the major route of hepatic cholesterol catabolism, we now analysed BAs level in faeces and plasma of this 9 weeks treatment study. 17 Notably, prolonged β 3-AR agonism reduced faecal total BA output into faeces (-31%; Figure 1A), which equals hepatic BA synthesis rate under steady-state conditions. While faecal CAderived BA secretion only tended to be reduced (-27%, P = 0.07; Figure 1B), faecal CDCA-derived BA secretion was significantly reduced (-35%; Figure 1C). The faecal excretion of secondary BAs was unaffected (Figure 1D). In plasma, total BA levels were markedly increased (+258%; Figure 1E), and this was due to an increase in both CA-derived BAs (+295%; Figure 1F) and CDCA-derived BAs (+217%; Figure 1G). β3-AR agonism also increased plasma secondary BA levels (+33%, Figure 1H), and the proportion of conjugated BAs (+55%, Supplementary material online, Figure S1A). Collectively, these data suggest that prolonged β3-AR agonism decreases faecal BA output related to stimulation of BA reuptake.

3.2 Prolonged β 3-AR agonism reduces the expression of genes involved in bile acid synthesis

To further reveal how β3-AR agonism regulates BA metabolism, hepatic mRNA expression of genes involved in BA metabolism was investigated. While β3-AR agonism only tended to reduce Cyp7a1, it significantly reduced Hsd3b7 (-57%) and Cyp8b1 (-40%) (Supplementary material online, Figure S1B), all of which are involved in the classical BA synthesis pathway. In addition, β3-AR agonism reduced Cyp27a1 (-53%) and Cyp7b1 (-38%) (Supplementary material online, Figure S1C), which are involved in the alternative BA synthesis pathway. This data is consistent with the reduced faecal BA excretion, implying a reduced hepatic BA synthesis rate under steady-state conditions. β3-AR agonism also tended to reduce Abcg5 (-31%, P = 0.06) and Bsep expression (-27%, P = 0.05) (Supplementary material online, Figure S1D), involved in excretion towards the bile of sterols and BAs, respectively. On the other hand, β3-AR agonism increased expression of Ost-β (+64%; Supplementary

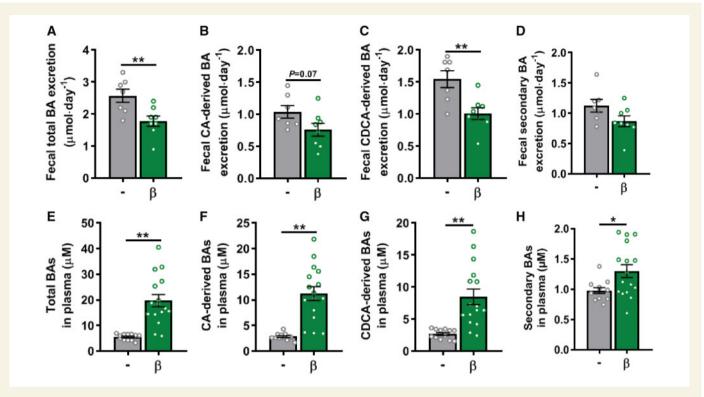


Figure 1 Prolonged β3-AR agonism decreases faecal bile acid excretion and increases plasma bile acid levels. *E3L.CETP* mice fed a WTD were treated with the β3-AR agonist CL316,243 (β) or vehicle (–) for 9 weeks. During the last week of treatment, faeces were collected, and BA species were assayed. Faecal excretion of (A) total bile acid (BAs), (B) cholic acid (CA)-derived BAs, (C) chenodeoxycholic acid (CDCA)-derived BAs, and (D) secondary BAs were calculated; n = 7-8 mice/group. Plasma was collected and (*E*) total BAs, (*F*) CA-derived BAs, (*G*) CDCA-derived Bas, and (*H*) secondary BAs were determined; n = 14-16 mice/group. Values are expressed as means \pm SEM. Differences were determined using the unpaired two-tailed Student's *t*-test. *P < 0.05, **P < 0.01 vs. vehicle (–).

material online, Figure S1E), involved in the basolateral BA secretion from the liver towards the systemic circulation. β 3-AR agonism tended to reduce Oatp1a1 expression (-60%, P=0.06) and significantly reduced Ntcp expression (-37%) (Supplementary material online, Figure S1F), involved in the uptake of reabsorbed BAs by the liver.

3.3 Bile acid sequestration reverses β 3-AR-mediated reduction of faecal bile acid output and normalizes elevated plasma bile acid levels

Because we observed that β 3-AR agonism decreases faecal BA excretion and increases plasma BAs, we next assessed whether inhibition of intestinal BA reabsorption, by using the BA sequestrant Colesevelam, would stimulate faecal BA loss and prevent the increase in plasma BAs during prolonged β 3-AR agonism. Mice were treated for 4 weeks with vehicle, the β 3-AR agonist alone, a low dose of the BA sequestrant alone (Colesevelam 0.15% in the WTD, w/w), or the combination of β 3-AR agonism and BA sequestration. β 3-AR agonism, Colesevelam, or the combination did not influence food intake (Supplementary material online, Figure S2A), body weight (Supplementary material online, Figure S2B), or body lean mass (Supplementary material online, Figure S2C). As expected, β 3-AR agonism tended to reduce body fat mass (P = 0.09; Supplementary material online, Figure S2D) and significantly reduced gonadal white adipose tissue weight (gWAT; Supplementary material online, Figure S2E). The combination of β 3-AR agonism and BA

sequestration significantly reduced body fat mass (Supplementary material online, Figure S2D) and gWAT weight (Supplementary material online, Figure S2E) as compared to vehicle. Liver weight was not significantly influenced by β 3-AR agonism alone or in combination with BA sequestration (Supplementary material online, Figure S2F).

Compared to vehicle, β 3-AR agonism alone, BA sequestration alone, and the combination of β 3-AR agonism and BA sequestration all increased bile flow (+43%, +33%, and +38% vs. vehicle, respectively; Figure 2A). The biliary BA secretion rate was not influenced by the different treatments (Figure 2B). Additionally, biliary cholesterol excretion rate was increased by β 3-AR agonism (+75% vs. vehicle), but not by BA sequestration or the combination of β 3-AR agonism and BA sequestration (Figure 2C).

Furthermore, although 4 weeks β 3-AR agonism did not significantly decrease faecal excretion of total BAs (*Figure 2D*) and CA-derived BAs (*Figure 2E*), excretion of CDCA-derived BAs was significantly decreased (-50% vs. vehicle; *Figure 2F*). BA sequestration on top of β 3-AR agonism strongly increased faecal excretion of total BAs (+91% vs. vehicle; +234% vs. β ; +47% vs. c; *Figure 2D*), CA-derived BAs (+201% vs. vehicle; +357% vs. β ; *Figure 2E*), and CDCA-derived BAs (+109% vs. β ; *Figure 2F*). In addition, BA sequestration on top of β 3-AR agonism markedly increased faecal secondary BA excretion (+122% vs. vehicle; +274% vs. β ; +40% vs. c; *Figure 2G*). Finally, although the β 3-AR agonism-induced increase in plasma BA levels was not as pronounced as after 9 weeks of treatment, concomitant BA sequestration normalized

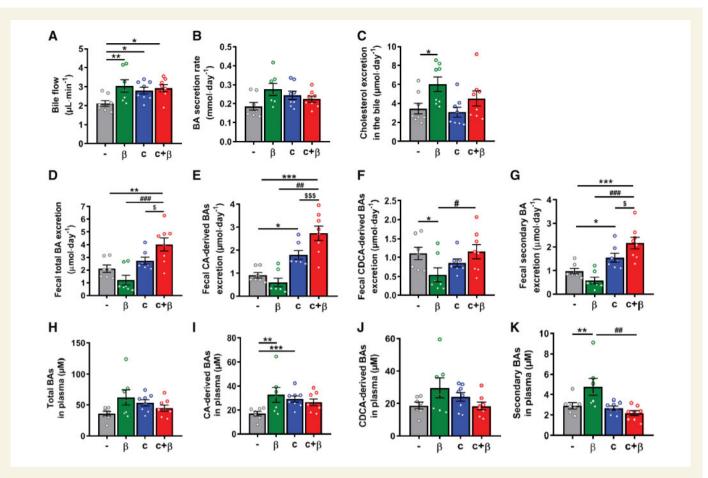


Figure 2 Bile acid sequestration reverses β3-AR induced reduction of faecal bile acid excretion and normalizes elevated plasma bile acid levels. *E3LCETP* mice fed a WTD were treated with vehicle (–), the β3-AR agonist CL316,243 (β), the bile acid (BA) sequestrant Colesevelam (c), or their combination (c + β) for 4 weeks. Bile duct cannulation was performed to collect the bile and (A) bile flow (μ L per minute) was determined. (B) BA secretion rate and (C) cholesterol excretion rate in the bile were determined. Faeces were collected to determine faecal (D) total BAs, (E) cholic acid (CA)-derived BAs, (F) chenodeoxycholic acid (CDCA)-derived BAs, and (G) secondary BAs. In plasma, (H) total BAs, (I) CA-derived BAs, (J) CDCA-derived BAs, and (K) secondary BAs were determined. N = 7-8 mice/group. Values are expressed as means \pm SEM. Differences between four groups were determined using one-way ANOVA with the LSD post hoc test. *P < 0.05, **P < 0.01, ***P < 0.01 vs. vehicle (–); *P < 0.05, **P < 0.01 vs. P < 0.01 vs. P < 0.05 (B); *P < 0.05 (B); *P < 0.05 (B); *P < 0.05 (B) *P

these plasma BA levels (*Figure 2H–J*). β 3-AR agonism clearly increased plasma secondary BA levels (+63% vs. vehicle), which was completely reversed by BA sequestration (-118% vs. c+ β ; *Figure 2K*). Taken together, these data indicate that inhibition of BA reabsorption by BA sequestration reverses β 3-AR agonism-induced reduction of faecal BA excretion, i.e. stimulates hepatic BA synthesis under these conditions, and normalizes β 3-AR agonism-mediated increased plasma BA levels.

3.4 Bile acid sequestration on top of β 3-AR agonism reverses hepatic cholesterol accumulation and further improves plasma cholesterol levels

As the BA sequestrant Colesevelam on top of β 3-AR agonism strongly increased faecal BA excretion and normalized plasma BA levels, we evaluated whether the addition of BA sequestration could also correct the β 3-AR agonism-induced hepatic cholesterol accumulation as shown previously ¹⁷ and further lower plasma lipids. We confirmed that β 3-AR agonism significantly increased hepatic TC levels (+26%). BA sequestration alone reduced hepatic TC levels as compared to vehicle (-41%)

(Figure 3A). Importantly, BA sequestration on top of β 3-AR agonism also largely reduced hepatic TC levels as compared to vehicle (-37%) and β 3-AR agonism alone (-50%) (Figure 3A), and to similar levels as BA sequestration alone. Hepatic TG and PL contents were not influenced by any of the treatments (Figure 3B and C).

Next, we assessed the effect of BA sequestration on top of β 3-AR agonism on plasma lipid levels. After 4 weeks of treatment, plasma TG levels were reduced by β 3-AR agonism (-52%) and tended to be reduced by BA sequestration alone (-33%, P = 0.07) as compared to vehicle. BA sequestration on top of β 3-AR agonism reduced plasma TG levels as compared to vehicle (-74%) and also as compared to BA sequestration alone (-62%) (*Figure 3D*). In addition, BA sequestration alone reduced plasma TC levels as compared to vehicle (-47%). BA sequestration on top of β 3-AR agonism also reduced plasma TC levels as compared to vehicle (-55%) and to β 3-AR agonism alone (-49%; *Figure 3E*).

Since cholesterol can be carried in plasma by either pro- or atherogenic lipoprotein classes, we also determined the distribution of cholesterol over plasma non-HDL and HDL. Plasma non-HDL-C levels tended to be reduced by β 3-AR agonism alone (-27%, P = 0.05) and were significantly reduced by BA sequestration alone (-55%) and BA sequestration

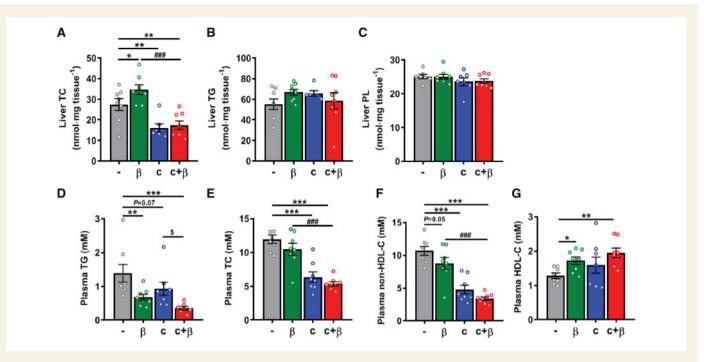


Figure 3 Bile acid sequestration on top of β3-AR agonism reverses hepatic cholesterol accumulation and further improves plasma cholesterol levels. *E3LCETP* mice fed a WTD were treated with vehicle (–), β3-AR agonist CL316,243 (β), bile acid sequestrant Colesevelam (c), or their combination (c + β). After 4 weeks treatment, mice were killed and liver samples were collected to evaluate hepatic (A) total cholesterol (TC), (B) triglyceride (TG), and (C) phospholipid (PL) levels. Blood samples were collected and plasma was assayed for (D) TG, (E) TC, (F) non-HDL-cholesterol (non-HDL-C), and (G) HDL-cholesterol (HDL-C) levels. N = 7-8 mice/group. Values are expressed as means ± SEM. Differences between four groups were determined using one-way ANOVA with the LSD post hoc test. *P < 0.05, **P < 0.01, ***P < 0.001 vs. vehicle (–); *##P < 0.001 vs. β3-AR agonist (β); *P < 0.05 vs. Colesevelam (c).

on top of β 3-AR agonism (-68%; Figure 3F) as compared to vehicle. Moreover, BA sequestration on top of β 3-AR agonism further reduced non-HDL-C levels as compared to β 3-AR agonism alone (-56%; Figure 3F). In addition, both β 3-AR agonism alone (+34%), and in combination with BA sequestration (+52%; Figure 3G) increased antiatherogenic HDL-C levels as compared to vehicle. Taken together, these findings indicate that BA sequestration on top of β 3-AR agonism reverses the β 3-AR agonism-induced hepatic cholesterol accumulation and further reduces plasma non-HDL-C levels.

3.5 Bile acid sequestration does not interfere with the β 3-AR agonism-induced plasma clearance and hepatic uptake of cholesterol-enriched TRL remnants

As β 3-AR agonism increases the formation and hepatic uptake of cholesterol-enriched TRL remnants ^{7,17} and BA sequestration on top of β 3-AR agonism further lowers plasma non-HDL-C levels, we next studied whether BA sequestration on top of β 3-AR agonism influenced the hepatic uptake of cholesterol-enriched TRL remnants. Hereto, we treated mice with the BA sequestrant Colesevelam on top of the β 3-AR agonist CL316,243 for 12 weeks. Similar as in the 4-week study, β 3-AR agonism alone and in combination with BA sequestration, but not BA sequestration alone, reduced body fat mass (-37% and -38%, respectively; Supplementary material online, *Figure S3A*) and gWAT weight (-55% and -61%, respectively; Supplementary material online, *Figure S3B*) as compared to vehicle. In agreement with previous studies, ^{7,17} we also observed that β 3-AR agonism induced substantial brown fat activation and

browning of WAT as evidenced from decreased lipid contents in BAT (Supplementary material online, Figure S3C) and subcutaneous WAT (scWAT, Supplementary material online, Figure S3D), while BA sequestration on top of β 3-AR agonism did not further add to these effects. The mRNA expression of genes related to intestinal BA reabsorption in the ileum is shown in Supplementary material online, Table S2. β 3-AR agonism significantly increased the mRNA expression of BA transporters Asbt and Ost- β . Furthermore, β 3-AR agonism markedly increased Shp and Fgf15 mRNA expression in the ileum, while the expression of these genes was reduced by BA sequestration, and normalized by the combination treatment.

In line with the 4-week intervention, 12 weeks of β 3-AR agonism alone improved dyslipidaemia by reducing plasma TG (-35%; Figure 4A), mainly via reducing (V)LDL-TG (Figure 4B), TC (-31%; Figure 4C), and non-HDL-C (-45%; Figure 4D) levels as compared to vehicle, while increasing HDL-C levels (+52%; Figure 4E). The decrease in (V)LDL-C and increase in HDL-C by β 3-AR agonism alone and on top of BA sequestration was confirmed by FPLC (Figure 4F). As compared to β 3-AR agonism alone, BA sequestration on top of β 3-AR agonism did not influence plasma TG (Figure 4A), but further lowered plasma TC (-24%; Figure 4C) and tended to further reduce non-HDL-C levels (-32%, P = 0.06; Figure 4D). Next, the total plasma TC and non-HDL-C exposure during the treatment period were calculated. The combination treatment further reduced both the total plasma TC exposure (-18%; Figure 4G) and non-HDL-C exposure (-20%; Figure 4H) as compared to β 3-AR agonism alone.

After 12 weeks of treatment, we evaluated the plasma clearance and hepatic uptake of intravenously injected glycerol tri[³H]oleate (triolein,

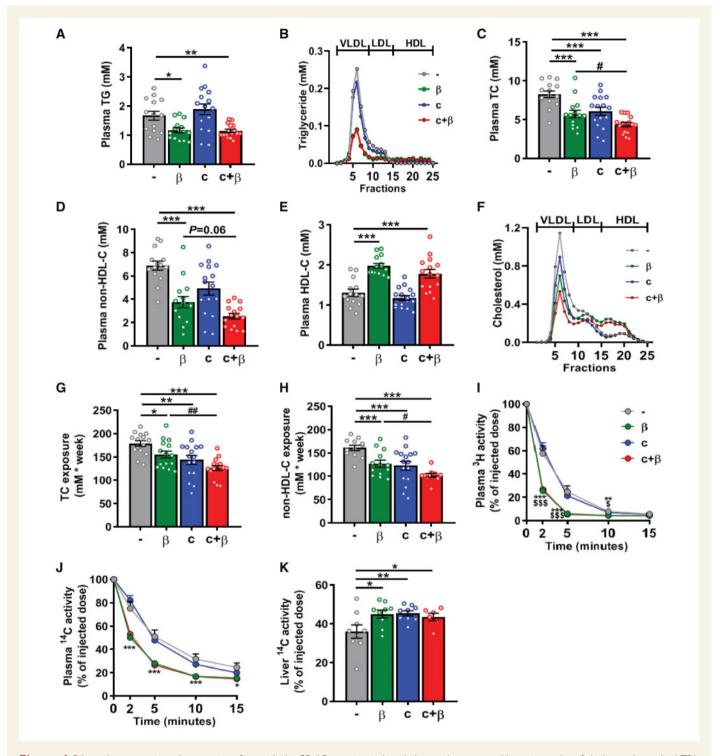


Figure 4 Bile acid sequestration does not interfere with the β3-AR agonism-induced plasma clearance and hepatic uptake of cholesterol-enriched TRL remnants. *E3LCETP* mice fed a WTD were treated with vehicle (–), the β3-AR agonist CL316,243 (β), the bile acid sequestrant Colesevelam (c), or their combination (c + β). After 12 weeks treatment, blood was collected to determine plasma (A) triglycerides (TG), (B) distribution of TG over lipoproteins, (C) total cholesterol (TC), (D) non-HDL-cholesterol (non-HDL-C), (E) HDL-cholesterol (HDL-C), and (F) distribution of TC over lipoproteins. (G) TC and (H) non-HDL-C exposure were calculated; n = 13-16 mice/group. Mice were injected intravenously with glycerol trig H] oleate and [14C] cholesteryl oleate-labelled lipoprotein-like particles. Plasma clearance of (f) H-activity and (f) 14C-activity, and (f) hepatic uptake of 14C-activity after 15 min were measured; n = 6-9 mice/group. Values are expressed as means ± SEM. Differences between four groups were determined using one-way ANOVA with the LSD post hoc test. *P < 0.05, **P < 0.01, ***P < 0.001 vs. vehicle (–); **P < 0.05, ***P < 0.05, ***P < 0.001 vs. Colesevelam (c).

TO) and [\$^4C\$]cholesteryl oleate (CO) double-labelled VLDL-mimicking particles. In line with previous studies, \$^{7.17}\$ \$\beta\$-AR agonism alone markedly accelerated the plasma clearance of [\$^3H\$]TO-derived activity (Figure 4I) and [\$^14C\$]CO (Figure 4J), and increased the hepatic uptake of the formed cholesterol-enriched TRL remnants (+25%, Figure 4K) as compared to vehicle. Additional BA sequestration did not further accelerate the plasma clearance of [\$^3H\$]TO-derived activity and [\$^14C\$]CO and also did not further increase the hepatic uptake of [\$^14C\$]CO (Figure 4I-K) as compared to \$^3-AR agonism alone.

3.6 Bile acid sequestration on top of β 3-AR agonism tends to further attenuate atherosclerosis development

To investigate if the beneficial effects of BA sequestration on top of β3-AR agonism on BA and cholesterol metabolism would translate in a further protection against atherosclerosis development, we evaluated the atherosclerotic lesion area in the root of the aortic arch after 12 weeks of treatment. As expected, β3-AR agonism alone decreased atherosclerotic lesion area throughout the aortic root (Figure 5A and B), resulting in lower mean atherosclerotic lesion area as compared to vehicle (-56%; Figure 5C). BA sequestration on top of β 3-AR agonism strongly attenuated atherosclerotic lesion area by -79% as compared to vehicle; and as compared to β 3-AR agonism alone tended to further reduce the atherosclerotic lesion area (-54%; P = 0.16) (Figure 5C). The total plasma TC exposure during the study strongly correlated with the square root (SQRT)-transformed lesion area (β = 2.14, R^2 = 0.40; P < 0.001; Figure 5D). Moreover, although atherosclerotic lesion severity was not significantly mitigated by any treatment (Supplementary material online, Figure S4A), β3-AR agonism increased the proportion of lesion-free valves as compared to vehicle (+122%), which was further increased by additional BA sequestration (+199% vs. vehicle; +34% vs. β ; Figure 5E). Proportions of SMC area and collagen area were not affected by any of the treatments (Figure 5F, G and Supplementary material online, Figure S4B), while BA sequestration on top of \(\beta 3-AR \) agonism further decreased the percentage of macrophage area within the lesion (-34% vs. vehicle; -26% vs. β; Figure 5H and Supplementary material online, Figure S4B) and increased the stability index defined by the ratio of stable markers (i.e. SMC area and collagen area) vs. the unstable marker (i.e. macrophage area) (+70% vs. vehicle; +44% vs. β ; Figure 51).

Taken together, BA sequestration in addition to $\beta 3\text{-}AR$ agonism tends to further reduce atherosclerosis development, an effect that is strongly related to its plasma cholesterol-lowering effect.

4. Discussion

Activating brown fat is a promising strategy to combat hypercholesterolaemia by increasing the flux of lipoprotein-associated cholesterol towards the liver, thereby exerting atheroprotective effects. 7,9 The aim of this study was first to evaluate the effects of prolonged brown fat activation, via $\beta 3$ -AR agonism, on hepatic cholesterol turnover. Secondly, we aimed to assess the effects of BA sequestration on top of brown fat activation on hepatic cholesterol and BA metabolism as well as atherosclerosis development. We uncovered that the increased hepatic cholesterol content by prolonged $\beta 3$ -AR agonism as shown previously, 17 was accompanied by increased plasma BA levels and decreased faecal BA excretion. It is likely that more efficient BA reabsorption from the gut is mostly responsible for these effects, since biliary BA (representing both newly synthesized BAs and cycled BAs within the enterohepatic

circulation) output was actually increased under these conditions. Indeed, concomitant BA sequestration by Colesevelam markedly increased faecal BA excretion and lowered the hepatic cholesterol content. As a result, combining BA sequestration with $\beta 3\text{-}AR$ agonism further reduced plasma cholesterol levels and tended to further reduce atherosclerosis development and also increase plaque stability as compared to $\beta 3\text{-}AR$ agonism alone.

Previously, we observed that prolonged β3-AR agonism in mice increases the delivery of cholesterol to the liver via the uptake of cholesterol-enriched TRL remnants⁷ and HDL-C⁹ and this increased flux of cholesterol towards the liver results in a moderate hepatic cholesterol accumulation. ¹⁷ In fact, 4 weeks of β3-AR agonism already clearly increased the hepatic cholesterol level. In the current study, we further show that prolonged β 3-AR agonism decreased faecal BA excretion and increased plasma BA levels. Since there is negligible excretion of BAs via the urine and skin, hepatic BA synthesis from cholesterol equals faecal BA excretion under steady-state conditions to maintain BA pool size.²⁴ Our data therefore demonstrate that prolonged β3-AR agonism decreases hepatic BA synthesis in mice. This is supported by the reduced expression of genes involved in the classical and alternative BA synthesis pathways. The observation that the biliary BA secretion rate was not decreased after prolonged β 3-AR agonism, but rather tended to be increased, can be explained by the fact that the biliary BAs represent both newly synthesized BAs as well as BAs that are recycled within the enterohepatic circulation. β3-AR agonism increases BA reabsorption from the gut (i.e. increasing cycled BAs within the enterohepatic circulation), which is responsible for the overall increased biliary BA output. In line with our study, Baskin et al. 25 recently showed that the β 3-AR agonist mirabegron increased gallbladder size in humans, which could be caused by an increased BA-induced bile flow upon brown fat activation.

The current study shows that the effects of prolonged brown fat activation on BA metabolism partly differ from the effects of short-term brown fat activation. Previous observations by us and others with shortterm brown fat activation by means of β 3-AR agonism and cold exposure (i.e. 1 week) showed increased expression of genes related to BA synthesis 18 and increased faecal BA excretion. 9,18 This difference observed with treatment duration is likely explained by an initial transient induction of BA synthesis upon brown fat activation, that is driven by the increased hepatic influx of cholesterol, the main substrate for BA synthesis, ²⁶ and dependent on hepatic induction of Cyp7b1. ¹⁸ After prolonged brown fat activation, the higher concentration of BAs in the gut likely stimulate BA reabsorption from the gut to prevent BA loss from the body. Subsequently, both BAs in the gut, via induction of FGF15 production, and circulating BAs target the hepatic FXR pathway and inhibit BA synthesis via a well-established feedback mechanism. 15 Based on our findings, 4 weeks of β 3-AR agonism is sufficient to induce such an inhibitory feedback on BA synthesis. Collectively, available data indicate that brown fat activation initially increases BA synthesis and thereby faecal BA excretion, while prolonged brown fat activation, decreases hepatic BA synthesis and thereby induces hepatic cholesterol accumulation. It appears that under these conditions reabsorption of BAs from the gut becomes more efficient, which, possibly in combination with suppressed expression of hepatic BA uptake transporters, leads to elevated plasma levels of BAs. High hepatic cholesterol and BA levels may affect liver function by inducing liver inflammation. 27,28

Since prolonged β 3-AR agonism increased plasma levels of BAs, including secondary BAs, and decreased faecal BA excretion, we reasoned that prolonged β 3-AR agonism induces reabsorption of BAs from the gut. In fact, β 3-AR agonism clearly increased the expression of Asbt, the

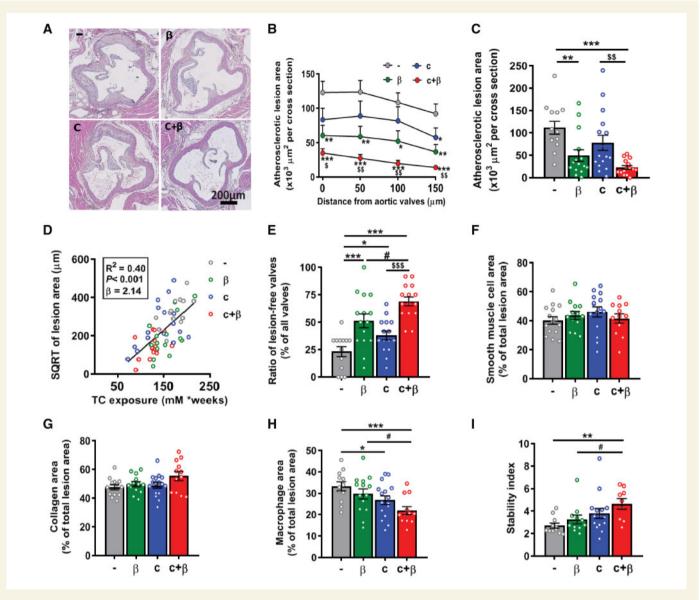


Figure 5 Bile acid sequestration on top of β3-AR agonism tends to further attenuate atherosclerosis development. *E3LCETP* mice fed a WTD were treated with vehicle (–), the β3-AR agonist CL316,243 (β), the bile acid sequestrant Colesevelam (c), or their combination (c + β) for 12 weeks. (A) Representative pictures of atherosclerotic lesions in aortic root area of each group are shown. (B) Plaque lesion area as a function of distance from the appearance of open valves and (*C*) mean atherosclerotic lesion area were calculated. (D) The square root (SQRT) of the mean atherosclerotic lesion area is plotted against the plasma total cholesterol (TC) exposure during the whole treatment period. (E) Ratio of the number of valves without any lesions divided by the total number of valves is shown. Relative areas of (*F*) smooth muscle cells, (*G*) collagen, and (*H*) macrophages within the lesion were determined. (*I*) The stability index was calculated as the ratio of stable markers (i.e. smooth muscle cell area and collagen area) per unstable marker (i.e. macrophage area). N = 13-16 mice/group. Values are expressed as means ± SEM. Differences between four groups were determined using one-way ANOVA with the LSD post hoc test. *P < 0.05, **P < 0.01, ***P < 0.01 vs. Colesevelam (c).

predominant transporter for the uptake of the luminal BAs, 29 and Ost- β , a basolateral BA transporter which plays a key role in BA efflux in the ileum. 30,31 In addition to increasing the expression of BA transporters, β 3-AR agonism increased plasma secondary BA levels. Intestinal anaerobic bacteria, i.e. *Eubacterium* and *Clostridium* are capable to deconjugate the liver-derived BAs and convert primary BAs into secondary BAs, such as LCA (omega-MCA in mice) and DCA, which has a high affinity to ASBT in the ileum. 33 The activity of those obligatory anaerobic bacteria would be largely impaired when faeces were collected and placed in the presence of oxygen. Thus, the effect of β 3-AR agonism on increasing

intestinal BA reabsorption may be attributed to upregulation of Asbt and Ost- β expression in the ileum as well as increased conversion of primary BAs into secondary BAs in the gut.

Colesevelam is a BA sequestrant that reduces the reabsorption of BAs from the gut, with a preference for relatively hydrophobic species like deoxycholic acid (DCA; i.e. the main secondary BA derived from CA). Likely via this mechanism, Colesevelam particularly increased faecal secondary BA excretion. Interruption of the enterohepatic circulation by a very low dosage of Colesevelam is not complete and increased hepatic synthesis and sufficient BAs within the circulation resulted in a

similar biliary BA secretion rate and actually a slightly increased bile flow as compared to vehicle, which corroborates previous studies. 34,35 Most importantly and fully in line with our expectations, Colesevelam strongly decreased levels of both hepatic and plasma cholesterol, explained by the fact that BA elimination is by far the most important contributor to cholesterol turnover. 26 The $\beta 3$ -AR agonism-induced BA reabsorption is likely effectively inhibited by BA sequestration, as Colesevelam on top of $\beta 3$ -AR agonism markedly increased faecal excretion of total BAs and secondary BAs. As a consequence of prevention of reabsorption, plasma BA levels, in particular secondary BAs were normalized upon BA sequestration on top of $\beta 3$ -AR agonism. The fact that BA sequestration on top of $\beta 3$ -AR agonism still lowered hepatic cholesterol to similar levels as reached by BA sequestration alone indicates that the effects of BA sequestration on hepatic cholesterol levels and BA synthesis is stronger than the effects of $\beta 3$ -AR agonism.

Preclinical studies showed that both β 3-AR agonism alone ^{7,17} and the BA sequestrant Colesevelam alone 35 not only reduce plasma cholesterol levels but also atherosclerosis development. Importantly, we now show that BA sequestration on top of β 3-AR agonism further reduces plasma non-HDL-C levels, tended to further reduce atherosclerosis development and further increased plaque stability as evidenced by reduced macrophage area versus SMC and collagen area within the lesion. This finding is highly relevant from a clinical perspective. In humans, the β3-AR agonist Mirabegron increases brown fat activity and resting energy expenditure³⁶ and high brown fat activity is associated with a reduced risk of cardiovascular disease events.³⁷ In addition, BA sequestrants attenuate coronary heart disease³⁸ and coronary artery lesions in humans.³⁹ Based on our findings, we speculate that combining conventional lipid-lowering by BA sequestration with brown fat activation may further improve dyslipidaemia and reduce atherosclerosis development in clinic.

In conclusion, prolonged $\beta3\text{-AR}$ agonism promotes BA reabsorption from the gut, resulting in elevated plasma BA levels, suppressed hepatic BA synthesis and elevated hepatic cholesterol content. Concomitant BA sequestration on top of $\beta3\text{-AR}$ agonism increases faecal BA excretion, normalizes plasma BA levels, reverses the $\beta3\text{-AR}$ agonism-induced hepatic cholesterol accumulation, further lowers plasma non-HDL-C levels and tends to further lower atherosclerosis development. These data suggest that combining conventional BA sequestration with brown fat activation via $\beta3\text{-AR}$ agonism could be a new therapeutic strategy to further reduce dyslipidaemia and attenuate atherosclerosis development.

Supplementary material

Supplementary material is available at Cardiovascular Research online.

Authors' contributions

E.Z. and G.H.: study concept and design; acquisition of data; analysis and interpretation of data; drafting of the manuscript; edited and revised manuscript; Z.L., A.C.E., A.W.S., R.H.H., M.K., R.B., and F.K.: acquisition of data, edited and revised manuscript; T.C., M.R.B, and J.F.P.B.: study concept and design, edited and revised manuscript; A.K.G. and P.C.N.R.: study concept and design, obtained funding, study supervision, edited and revised manuscript; Y.W.: study concept and design; analysis and interpretation of data; drafting of the manuscript; edited and revised manuscript; obtained funding.

Acknowledgements

The authors thank Elsbeth J. Pieterman (TNO-Metabolic Health Research, Gaubius Laboratory, Leiden, The Netherlands) for technical assistance on the fast-performance liquid chromatography.

Conflict of interest: none declared.

Funding

This work was supported by the Netherlands Organization for Scientific Research-NWO (VENI grant 91617027 to Y.W.); the Netherlands Organisation for Health Research and Development-ZonMW (Early Career Scientist Hotel grant 435004007 to Y.W.); EU research funding (FP7-HEALTH-305707 to A.K.G.); the Netherlands Cardiovascular Research Initiative: an initiative with support of the Dutch Heart Foundation (CVON-GENIUS-2); and the Netherlands Heart Foundation (2009T038 to P.C.N.R.).

References

- Jukema JW, Cannon CP, de Craen AJ, Westendorp RG, Trompet S. The controversies of statin therapy: weighing the evidence. J Am Coll Cardiol 2012;60:875–881.
- Ouellet V, Labbe SM, Blondin DP, Phoenix S, Guerin B, Haman F, Turcotte EE, Richard D, Carpentier AC. Brown adipose tissue oxidative metabolism contributes to energy expenditure during acute cold exposure in humans. J Clin Invest 2012;122: 545–552
- Labbe SM, Caron A, Bakan I, Laplante M, Carpentier AC, Lecomte R, Richard D. In vivo measurement of energy substrate contribution to cold-induced brown adipose tissue thermogenesis. FASEB J 2015;29:2046–2058.
- Oikonomou EK, Antoniades C. The role of adipose tissue in cardiovascular health and disease. Nat Rev Cardiol 2019;16:83–99.
- Hoeke G, Kooijman S, Boon MR, Rensen PC, Berbee JF. Role of brown fat in lipoprotein metabolism and atherosclerosis. Circ Res 2016;118:173–182.
- Cannon B, Nedergaard J. Brown adipose tissue: function and physiological significance. Physiol Rev 2004;84:277–359.
- Berbee JF, Boon MR, Khedoe PP, Bartelt A, Schlein C, Worthmann A, Kooijman S, Hoeke G, Mol IM, John C, Jung C, Vazirpanah N, Brouwers LP, Gordts PL, Esko JD, Hiemstra PS, Havekes LM, Scheja L, Heeren J, Rensen PC. Brown fat activation reduces hypercholesterolaemia and protects from atherosclerosis development. *Nat Commun* 2015;6:6356.
- Khedoe PP, Hoeke G, Kooijman S, Dijk W, Buijs JT, Kersten S, Havekes LM, Hiemstra PS, Berbee JF, Boon MR, Rensen PC. Brown adipose tissue takes up plasma triglycerides mostly after lipolysis. J Lipid Res 2015;56:51–59.
- Bartelt A, John C, Schaltenberg N, Berbee JFP, Worthmann A, Cherradi ML, Schlein C, Piepenburg J, Boon MR, Rinninger F, Heine M, Toedter K, Niemeier A, Nilsson SK, Fischer M, Wijers SL, van Marken Lichtenbelt W, Scheja L, Rensen PCN, Heeren J. Thermogenic adipocytes promote HDL turnover and reverse cholesterol transport. Nat Commun 2017;8:15010.
- Chiang JY. Regulation of bile acid synthesis: pathways, nuclear receptors, and mechanisms. J Hepatol 2004;40:539–551.
- Bjorkhem I, Araya Z, Rudling M, Angelin B, Einarsson C, Wikvall K. Differences in the regulation of the classical and the alternative pathway for bile acid synthesis in human liver. No coordinate regulation of CYP7A1 and CYP27A1. J Biol Chem 2002;277: 26804—26807
- de Boer JF, Bloks VW, Verkade E, Heiner-Fokkema MR, Kuipers F. New insights in the multiple roles of bile acids and their signaling pathways in metabolic control. Curr Opin Lipidol 2018;29:194–202.
- 13. Houten SM. Homing in on bile acid physiology. Cell Metab 2006;4:423–424.
- Li T, Chiang JY. Bile acid signaling in metabolic disease and drug therapy. Pharmacol Rev 2014;66:948–983.
- 15. Chiang JY. Bile acids: regulation of synthesis. J Lipid Res 2009;50:1955–1966.
- The Lipid Research Clinics Coronary Primary Prevention Trial results. I. Reduction in incidence of coronary heart disease. JAMA 1984;251:351–364.
- 17. Hoeke G, Wang Y, van Dam AD, Mol IM, Gart E, Klop HG, van den Berg SM, Pieterman EH, Princen HMG, Groen AK, Rensen PCN, Berbée JFP, Boon MR. Atorvastatin accelerates clearance of lipoprotein remnants generated by activated brown fat to further reduce hypercholesterolemia and atherosclerosis. Atherosclerosis 2017;267:116–126.
- 18. Worthmann A, John C, Ruhlemann MC, Baguhl M, Heinsen FA, Schaltenberg N, Heine M, Schlein C, Evangelakos I, Mineo C, Fischer M, Dandri M, Kremoser C, Scheja L, Franke A, Shaul PW, Heeren J. Cold-induced conversion of cholesterol to bile acids in mice shapes the gut microbiome and promotes adaptive thermogenesis. Nat Med 2017;23:839–849.
- 19. van Vlijmen BJ, van den Maagdenberg AM, Gijbels MJ, van der Boom H, HogenEsch H, Frants RR, Hofker MH, Havekes LM. Diet-induced hyperlipoproteinemia and

- atherosclerosis in apolipoprotein E3-Leiden transgenic mice. J Clin Invest 1994;**93**: 1403–1410
- de Haan W, de Vries-van der Weij J, van der Hoorn JW, Gautier T, van der Hoogt CC, Westerterp M, Romijn JA, Jukema JW, Havekes LM, Princen HM, Rensen PC. Torcetrapib does not reduce atherosclerosis beyond atorvastatin and induces more proinflammatory lesions than atorvastatin. *Circulation* 2008;**117**:2515–2522.
- Westerterp M, van der Hoogt CC, de Haan W, Offerman EH, Dallinga-Thie GM, Jukema JW, Havekes LM, Rensen PC. Cholesteryl ester transfer protein decreases high-density lipoprotein and severely aggravates atherosclerosis in APOE*3-Leiden mice. Arterioscler Thromb Vasc Biol 2006;26:2552–2559.
- 22. Rensen PC, van Dijk MC, Havenaar EC, Bijsterbosch MK, Kruijt JK, van Berkel TJ. Selective liver targeting of antivirals by recombinant chylomicrons—a new therapeutic approach to hepatitis B. *Nat Med* 1995;**1**:221–225.
- Wong MC, van Diepen JA, Hu L, Guigas B, de Boer HC, van Puijvelde GH, Kuiper J, van Zonneveld AJ, Shoelson SE, Voshol PJ, Romijn JA, Havekes LM, Tamsma JT, Rensen PCN, Hiemstra PS, Berbée JFP. Hepatocyte-specific IKKbeta expression aggravates atherosclerosis development in APOE*3-Leiden mice. Atherosclerosis 2012; 220:362–368
- Hofmann AF, Hagey LR. Key discoveries in bile acid chemistry and biology and their clinical applications: history of the last eight decades. J Lipid Res 2014;55:1553–1595.
- 25. Baskin AS, Linderman JD, Brychta RJ, McGehee S, Anflick-Chames E, Cero C, Johnson JW, O'Mara AE, Fletcher LA, Leitner BP, Duckworth CJ, Huang S, Cai H, Garraffo HM, Millo CM, Dieckmann W, Tolstikov V, Chen EY, Gao F, Narain NR, Kiebish MA, Walter PJ, Herscovitch P, Chen KY, Cypess AM. Regulation of human adipose tissue activation, gallbladder size, and bile acid metabolism by a β3-adrenergic receptor agonist. Diabetes 2018;60:875–881.
- Kuipers F, Bloks VW, Groen AK. Beyond intestinal soap-bile acids in metabolic control. Nat Rev Endocrinol 2014;10:488–498.
- Allen K, Jaeschke H, Copple BL. Bile acids induce inflammatory genes in hepatocytes: a novel mechanism of inflammation during obstructive cholestasis. Am J Pathol 2011; 178:175–186.
- Tall AR, Yvan-Charvet L. Cholesterol, inflammation and innate immunity. Nat Rev Immunol 2015:15:104–116.

- Kullak-Ublick GA, Stieger B, Meier PJ. Enterohepatic bile salt transporters in normal physiology and liver disease. Gastroenterology 2004;126:322–342.
- Dawson PA, Hubbert M, Haywood J, Craddock AL, Zerangue N, Christian WV, Ballatori N. The heteromeric organic solute transporter alpha-beta, Ostalpha-Ostbeta, is an ileal basolateral bile acid transporter. J Biol Chem 2005;280:6960–6968.
- Rao A, Haywood J, Craddock AL, Belinsky MG, Kruh GD, Dawson PA. The organic solute transporter alpha-beta, Ostalpha-Ostbeta, is essential for intestinal bile acid transport and homeostasis. *Proc Natl Acad Sci USA* 2008;**105**:3891–3896.
- 32. Wahlstrom A, Sayin SI, Marschall HU, Backhed F. Intestinal crosstalk between bile acids and microbiota and its impact on host metabolism. *Cell Metab* 2016;**24**:41–50.
- 33. Craddock AL, Love MW, Daniel RW, Kirby LC, Walters HC, Wong MH, Dawson PA. Expression and transport properties of the human ileal and renal sodium-dependent bile acid transporter. *Am J Physiol* 1998;**274**:G157–169.
- 34. Herrema H, Meissner M, van Dijk TH, Brufau G, Boverhof R, Oosterveer MH, Reijngoud DJ, Muller M, Stellaard F, Groen AK, Kuipers F. Bile salt sequestration induces hepatic de novo lipogenesis through farnesoid X receptor- and liver X receptor alpha-controlled metabolic pathways in mice. Hepatology 2010;51:806–816.
- 35. Meissner M, Wolters H, de Boer RA, Havinga R, Boverhof R, Bloks VW, Kuipers F, Groen AK. Bile acid sequestration normalizes plasma cholesterol and reduces atherosclerosis in hypercholesterolemic mice. No additional effect of physical activity. *Atherosclerosis* 2013;228:117–123.
- Cypess AM, Weiner LS, Roberts-Toler C, Franquet Elia E, Kessler SH, Kahn PA, English J, Chatman K, Trauger SA, Doria A, Kolodny GM. Activation of human brown adipose tissue by a beta3-adrenergic receptor agonist. *Cell Metab* 2015;21:33–38.
- Takx RA, Ishai A, Truong QA, MacNabb MH, Scherrer-Crosbie M, Tawakol A. Supraclavicular brown adipose tissue ¹⁸F-FDG uptake and cardiovascular disease. *Nucl Med* 2016;57:1221–1225.
- The Lipid Research Clinics Coronary Primary Prevention Trial results. II. The relationship of reduction in incidence of coronary heart disease to cholesterol lowering. *JAMA* 1984:251:365–374.
- Brensike JF, Levy RI, Kelsey SF, Passamani ER, Richardson JM, Loh IK, Stone NJ, Aldrich RF, Battaglini JW, Moriarty DJ. Effects of therapy with cholestyramine on progression of coronary arteriosclerosis: results of the NHLBI Type II Coronary Intervention Study. *Circulation* 1984;69:313–324.

Translational perspective

Current therapeutic strategies are unable to prevent the majority of cardiovascular disease (CVD)-relating morbidities and mortalities, illustrating the need for new therapeutic strategies. Brown fat has been shown as an emerging target to combat hyperlipidaemia and atherosclerosis. Here, we showed that prolonged brown fat activation promotes bile acid (BA) reabsorption, resulting in elevated plasma BA and hepatic cholesterol content, both of which are reversed by additional BA sequestration. Importantly, combining BA sequestration with brown fat activation further lowers plasma cholesterol and reduces atherosclerosis development, indicating the combination therapy as a new therapeutic strategy to treat hyperlipidaemia, and ultimately CVD.