



The C-Type Lectin Mincle: Clues for a Role in Crohn's Disease Adjuvant Reaction

Anje A. te Velde*

Tytgat Institute for Liver and Intestinal Research, Academic Medical Center, Amsterdam, Netherlands

Keywords: Mincle review, adjuvants, C-type lectin, Mycobacterium, Crohn's disease

The term adjuvant is predominantly used when discussing vaccines, but only mimics how normally infections activate the immune system to secure that an innate immune reaction induces dendritic cells (DCs) to become optimally stimulatory for T cells. The interaction with the pathogen determines the different signals that are needed for a DC to become fully operated and give the proper polarizing factors to the differentiating T cell. Optimal co-stimulation requires a signal that is provided by upregulated receptors (CD80 and CD86) on DCs, and the T cell polarizing signal is mediated by various soluble or membrane-bound factors, like IL-12 for Th1 cell polarization. These signals are provided by ligation of pattern recognition receptors (PRRs), such as toll-like receptors (TLRs) and C-type lectins that can sense infection through recognition of pathogen-associated molecular patterns (PAMPs) or various inflammatory tissue factors (1).

OPEN ACCESS

Edited by:

Roland Lang, University Hospital Erlangen, Germany

Reviewed by:

Zoltan Prohaszka, Semmelweis University, Hungary Trinath Jamma, University of Massachusetts Boston, United States

*Correspondence:

Anje A. te Velde a.a.tevelde@amc.nl

Specialty section:

This article was submitted to Molecular Innate Immunity, a section of the journal Frontiers in Immunology

Received: 31 May 2017 Accepted: 27 September 2017 Published: 23 October 2017

Citation:

te Velde AA (2017) The C-Type Lectin Mincle: Clues for a Role in Crohn's Disease Adjuvant Reaction. Front. Immunol. 8:1304. doi: 10.3389/fimmu.2017.01304 Resting macrophages have low major histocompatibility complex class II and co-stimulatory molecules expressed on their surface. Macrophages can take up microorganisms *via* receptors such as scavenger receptors, complement receptors, and C-type lectins for degradation in phagosomes resulting in peptides for presentation. Macrophages also continuously scavenge dead or dying cells. These are a rich source of self-antigens, so it is very important that they do not activate naïve T cells when there is no ongoing microbial infection.

In Crohn's disease (CD), a chronic inflammatory bowel disease (IBD), multiple factors have been described that contribute to disease pathogenesis (2). The exact etiology of IBDs still remains unknown, although it is thought that the diseases result from an excessive immune response directed against microbial or environmentally derived antigens that can be triggered by the disruption of the intestinal epithelial barrier integrity. The resulting inflammation is a very general reaction; a specific antigen mediating the inflammation has never been identified. The response is induced by the luminal microbiota, where microbial antigens as adjuvants stimulate the immune reaction. This results in activated innate (macrophages and neutrophils) and adaptive (Th1/Th17 and B lymphocytes) responses (3). In this respect, CD includes many characteristics of an immunologic adjuvant reaction.

Most proteins are poor immunogens when injected alone. Various substances that induce co-stimulatory, adjuvant, activity have been added in vaccines for a long time to induce appropriate antibody responses. Vaccines containing bacterial products, necessary for T cell responses, are very potent and therefore use in humans is limited. In recent years, the development of adjuvants that induce a strong cellular response has shifted from an empirical to a rational process based on knowledge of molecular mechanisms. A major breakthrough was the identification of the C-type lectin Mincle (macrophage-inducible C-type lectin) as one of the main receptors involved (4–6).

In this opinion article, I provide clues that the cellular adjuvant reaction that characterizes the pathophysiology of CD might be mediated by signaling *via* Mincle.

Mincle in Crohn's Disease Adjuvant Reaction

MINCLE

Mincle (also called clec4e or clecsf9) was first described as a downstream target of NF-IL6 (also named C/EBP-β), a transcription factor, in macrophages (7). They demonstrate that Mincle mRNA was strongly induced in response to several inflammatory stimuli, such as LPS, TNF- α , IL-6, and IFN- γ in murine macrophages. A few years later, Mincle was grouped together with macrophage C-type lectin (MCL), DC immunoreceptor, and Dectin-2 (DC-associated lectin-2) as type II-related C-type lectins (8). These genes were mapped in an arthritis susceptibility locus in a rat model and the first indication for an immune activating function of Mincle was proposed (9). Mincle serves as a receptor for various bacteria, fungi, and other molecules (listed in Figure 1A). Mincle signals *via* association with the FcRy chain that contains an activating receptor coupled with an immunoreceptor tyrosine-based activation motif, ultimately resulting in activation of NF- κ B (10).

The first suggestion that Mincle is a receptor for a cell wall component of *Mycobacterium tuberculosis*, the glycolipid trehalose-6,6'-dimycolate (TDM, also named cord factor), was made when it appeared to be involved in a characteristic process of mycobacterial infection: the formation of granulomas (4). Mycobacteria can persist in normal tissues (39); recruitment of Mincle by TDM coupled to immunoglobulin G-opsonized beads interferes with phagosome maturation (40). The activity of Mincle is mediated by a ligand binding site that is conserved in a wide range of mammalian species (41). The Th1/Th17 adjuvanticity of TDM and its synthetic analog trehalose-6,6'-dibehenate (TDB) including its molecular mechanism *via* Syk and Card9 was confirmed in several studies (5, 6, 42).

Mincle protein is barely detectable on resting cells (4, 42). In experiments in various tissues in rhesus macaques, the frequencies of CD14⁺ gated cells that express Mincle in colon and ileum were low compared with bone marrow, liver, spleen, and lymph nodes (43). Induction of Mincle expression was shown to be induced by several pathogenic and non-pathogenic stimuli. Mincle was shown to be induced by TDM in the absence of Mincle protein expression *via* MCL (also called dectin-3) that was constitutively expressed in myeloid cells (44–46) through protein–protein interaction *via* its stalk region (47). C/EBP- β is the central hub in Mincle expression and connects TLR4 signals to TDB/TDM responsiveness through MyD88-dependent upregulation of Mincle (32, 48).

MINCLE AND CD

In literature, there is no direct link that connects Mincle to CD. There is, however, already a lot of information that links Mincle to other diseases. Most of these are also inflammatory-mediated diseases, such as rheumatoid arthritis (49, 50), allergic skin inflammation (28) and post-ischemic inflammation (51, 52), and other experimental inflammatory models (53–58).

Mincle has been shown to regulate numerous cellular responses including phagocytosis, endocytosis, respiratory burst, Nlrp3 inflammasome activation, NET formation, pro-inflammatory cytokine, and chemokine production and promotes Th1/Th17 responses [recently reviewed in Ref. (59, 60)]. These are all inflammatory reactions that have been described to play a role in CD. In **Figure 1A**, the different factors that are involved in Mincle signaling and also associated with CD are highlighted and discussed beneath.

As indicated, Mincle can act as a receptor for several different pathogens. The question is if these microorganisms have also been associated with CD. For Mycobacterium avium subspecies paratuberculosis, this is well known, it can be isolated from intestinal tissues and blood samples from CD patients at higher frequency than healthy persons (61). Treatment with antimycobacterial regimens in clinical trials achieved reversal of CD symptoms (62, 63). Also other bacteria linked to Mincle have been associated with CD: Listeria monocytogenes, Klebsiella pneumonia, Streptococcus pneumonia, Pneumocystis pneumonia, and Escherichia coli (64-70). A dysfunction in both a specialized form of autophagy, xenophagy, and HIF-1a was demonstrated to be involved in adherent invasive E. coli infections in CD (71). HIF-1α-induced inducible nitric oxide synthase produces nitric oxide (NO) that was shown to be upregulated in the inflamed mucosa in response to pro-inflammatory cytokines (72, 73).

In around 50% of CD patients' granulomas can be detected (74). Granulomas are linked to mycobacterium, and Mincle has been shown to be important (75). Several cytokines, including IL-1 and TNF- α , have been shown to promote the formation of granulomas (76). These cytokines can be secreted upon stimulation *via* Mincle after stimulation with TDM-mediated granuloma formation (4).

Toward fungal glycans, it has been demonstrated that human peripheral blood mononuclear cells (PBMCs) from CD patients show a hyperresponsiveness with a central role for Syk and Src signaling (77). PBMCs from patients with CD produce more IFN- γ and IL-17 upon exposure to *Candida* (78).

A well-known complication of CD is intestinal fibrosis. Recently, it was demonstrated that this was mediated *via* a PKCδmediated redox-dependent signaling process by accumulated advanced oxidation protein products (79).

CARD9 has been an autoimmune disease-associated gene, and differential expression of this gene might be a functional mechanism underlying observed GWAS signals (80). It coordinates Th17- and IL-22-producing cells in intestinal immune responses after epithelial injury in mice (81). Aberrant regulation of *CARD9*, either through genetic mutation (e.g., polymorphism) or activation by environmental triggers *via* Mincle, could contribute to pathological immune activation.

Crohn's disease patients treated with TNF blockers demonstrated an increased risk of opportunistic infections such as mycosis, aspergillosis, pneumocystosis, or cryptococcosis (82) and also Pityrosporum (Malassezia) folliculitis (83), and cutaneous lesions of Leishmaniasis (84).

About 50% of the world's population carry the *Helicobacter pylori* bacterium. In a meta-analysis, a negative association was found between *H. pylori* infection and CD. They conclude that *H. pylori* could exert an immunomodulatory effect in IBD (85) maybe by Mincle-mediated anti-inflammatory signaling (18).

Finally, danger/damage-associated molecular pattern (DAMP)-derived triggers from dead cells may contribute *via*

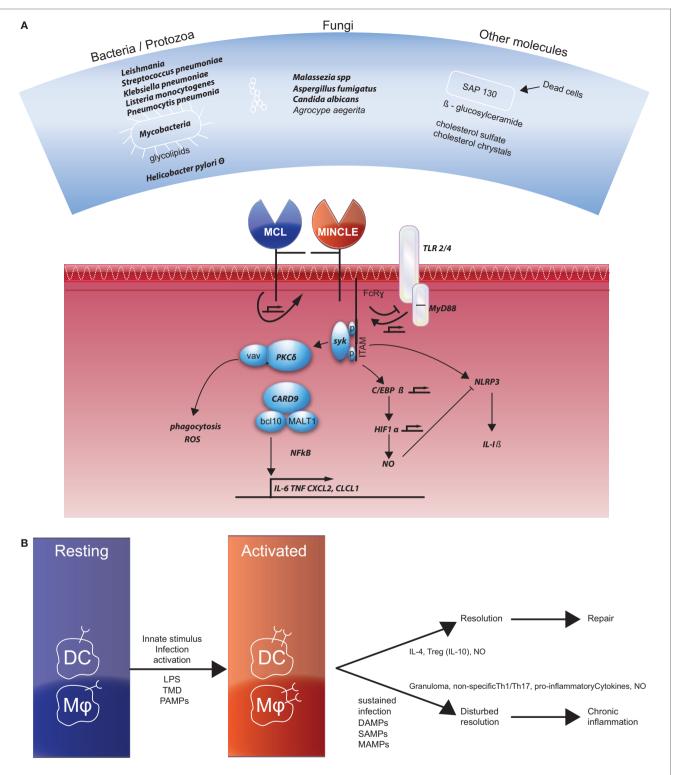


FIGURE 1 | **(A)** Overview of the ligands of Mincle (top panel) and the signaling pathways described in the current literature. Factors that were also documented in literature related to Crohn's disease are depicted in bold. References of the different ligands not mentioned in the text are for bacterial pathogen species (11–13), *Leishmania* (14), mycobacteria, and corynebacteria (15, 16), *Streptococcus pneumonia* (17), *Helicobacter pylori* (18), *Malassezia* (19, 20), *Aspergillus fumigatus* (21), *Candida albicans* (22, 23), SAP130 (10, 24, 25), β-glucosylceramide (26), cholesterol crystals (27), and cholesterol sulfate (28). References for the various signaling molecules not mentioned in the text are for TLR4/MyD88 (29), PKC (30), vav proteins (31), HIF1α (32) and NIrp3 (33, 34). **(B)** Resting dendritic cells (DCs) and macrophages also do not express Mincle, but they do express macrophage C-type lectin (MCL). Different activating signals induce Mincle expression. Two different pathways induce repair or chronic inflammation as indicated. References for activation and chronic inflammation are given in the text, for the resolution: Th2 skewing *via* IL-4 (35, 36), IL-10 (37), and NO (38).

Mincle to excessive and sustained inflammation in CD patients with active disease (86).

Taken together, numerous Mincle-related ligands and signaling molecules can be linked to CD.

HOW COULD MINCLE MEDIATE CD INFLAMMATION?

The efficacy by which Mincle handles microbes, microbial products, and damaged cells directs whether the outcome will be with suppression or with excess inflammation (see Figure 1B). These pathways can be polarized, but since CD is a chronic relapsing disease the activating signals and wound healing processes might also be present more or less in parallel. Mincle functions as a receptor for different bacteria and fungi, leading to proper immune responses that functions to eradicate pathogens (38, 82). Early response mediated via TLRs and MCL expressed on macrophages by a primary infectious stimulus (PAMP) results in the upregulation of Mincle expression. This leads to a sustained signaling process via the activating motif of the FcRy chain and the production of pro-inflammatory cytokines and finally a non-specific activation of Th1/Th17 response. In an appropriate immune response, the end product is the eradication of the infectious agent and resolution of the inflammation. Mincle stimulation can help by inducing antiinflammatory genes and genes involved in wound healing. In case of a sustained infection when pathogenic ligands are still present or because of tissue damage resulting in the presence of DAMPs or self-associated molecular patterns (SAMPs) continuous Mincle signaling remains. TLR- and Mincle co-dependent genes are enriched among genes required to handle persisting D/M/SAMP signals (38).

There are several pathways that counter-regulate Mincle on macrophages and DCs. Among them is the observed effect of IL-4 on Mincle expression of monocyte-derived DCs (87). This is, however, an artificial system, because these cells co-express surface markers (CD83 and DC-SIGN) that are not found to on the same cells in the *in vivo* situation (88).

In CD, there is evidence that a dysregulated macrophage function and a consecutive defective acute inflammatory response result in the impaired clearance of commensal bacteria. The persistence of the bacteria leads to a chronic granulomatous inflammation. Pathogenic infections may act as triggers or

REFERENCES

- Kapsenberg ML. Dendritic-cell control of pathogen-driven T-cell polarization. Nat Rev Immunol (2003) 3(12):984–93. doi:10.1038/nri1246
- Maloy KJ, Powrie F. Intestinal homeostasis and its breakdown in inflammatory bowel disease. *Nature* (2011) 474(7351):298–306. doi:10.1038/nature10208
- Kmiec Z, Cyman M, Slebioda TJ. Cells of the innate and adaptive immunity and their interactions in inflammatory bowel disease. *Adv Med Sci* (2017) 62(1):1–16. doi:10.1016/j.advms.2016.09.001
- Ishikawa E, Ishikawa T, Morita YS, Toyonaga K, Yamada H, Takeuchi O, et al. Direct recognition of the mycobacterial glycolipid, trehalose dimycolate, by C-type lectin Mincle. *J Exp Med* (2009) 206(13):2879–88. doi:10.1084/ jem.20091750

contributing factors for the chronic inflammation (2, 89) that is mediated by other stimuli of various nature, involving microbialassociated molecular patterns, DAMPs, or SAMPs, all described to be ligands of Mincle.

It will be of potential interest to study the direct role of Mincle as a predominant activating C-type lectin receptor via a Syk/Card9-dependent signaling mechanism in CD. Genetic susceptibility, barrier defects, or bacterial handling, dysbiosis or infection, sustained innate immunity, and defective regulation are all layers of a multi-hit model of intestinal inflammation (2). They are combined with different homeostatic modules such as autophagy, ER stress, antimicrobial proteins, the microbiota, PRRs, cytokine modules, and regulatory T cells. Defective modules may predispose people to the development of chronic intestinal inflammation. Determination of the role of Mincle in these layers and modules will reveal if Mincle is an important receptor of mediator of the chronic nature of CD, which could be relevant for therapeutic intervention. Targeting Syk has been suggested as a treatment for allergic and autoimmune disorders (90). In rheumatoid arthritis, inhibition of Syk has been studied as a treatment option (91). Although there is no direct evidence on the role of Mincle in CD, the data on the expression and role of Mincle in health and disease reveal numerous potential starting points. The synergy and antagonisms of the various PRRs, whether these are C-type lectins or TLRs, and their differential regulation on cells of the innate immune system, macrophages, and DCs is an important topic to understand the endogenous adjuvant reaction that they might induce. There are probably multiple mechanisms and interactions that result in the observed pathogenic immune reaction that is the fundament of CD. Here, the surprising overlap between features of CD and the roles that Mincle plays in a (chronic) immune reaction might indicate that CD could be an adjuvant reaction induced by Mincle triggering.

AUTHOR CONTRIBUTIONS

The author confirms being the sole contributor of this work and approved it for publication.

ACKNOWLEDGMENTS

Dr. R. Snackers from Com-unicate is greatly acknowledged for drawing the figures.

- Schoenen H, Bodendorfer B, Hitchens K, Manzanero S, Werninghaus K, Nimmerjahn F, et al. Cutting edge: Mincle is essential for recognition and adjuvanticity of the mycobacterial cord factor and its synthetic analog trehalose-dibehenate. *J Immunol* (2010) 184(6):2756–60. doi:10.4049/ jimmunol.0904013
- Decout A, Silva-Gomes S, Drocourt D, Barbe S, Andre I, Cueto FJ, et al. Rational design of adjuvants targeting the C-type lectin Mincle. *Proc Natl Acad Sci U S A* (2017) 114(10):2675–80. doi:10.1073/pnas.1612421114
- Matsumoto M, Tanaka T, Kaisho T, Sanjo H, Copeland NG, Gilbert DJ, et al. A novel LPS-inducible C-type lectin is a transcriptional target of NF-IL6 in macrophages. *J Immunol* (1999) 163(9):5039–48.
- 8. Balch SG, Greaves DR, Gordon S, McKnight AJ. Organization of the mouse macrophage C-type lectin (Mcl) gene and identification of a

subgroup of related lectin molecules. *Eur J Immunogenet* (2002) 29(1):61–4. doi:10.1046/j.1365-2370.2002.00266.x

- Flornes LM, Bryceson YT, Spurkland A, Lorentzen JC, Dissen E, Fossum S. Identification of lectin-like receptors expressed by antigen presenting cells and neutrophils and their mapping to a novel gene complex. *Immunogenetics* (2004) 56(7):506–17. doi:10.1007/s00251-004-0714-x
- Yamasaki S, Ishikawa E, Sakuma M, Hara H, Ogata K, Saito T. Mincle is an ITAM-coupled activating receptor that senses damaged cells. *Nat Immunol* (2008) 9(10):1179–88. doi:10.1038/ni.1651
- Chinthamani S, Settem RP, Honma K, Kay JG, Sharma A. Macrophage inducible C-type lectin (Mincle) recognizes glycosylated surface (S)-layer of the periodontal pathogen *Tannerella forsythia*. *PLoS One* (2017) 12(3):e0173394. doi:10.1371/journal.pone.0173394
- Kottom TJ, Hebrink DM, Jenson PE, Nandakumar V, Wuthrich M, Wang H, et al. The interaction of pneumocystis with the C-type lectin receptor Mincle exerts a significant role in host defense against infection. *J Immunol* (2017) 198(9):3515–25. doi:10.4049/jimmunol.1600744
- Ostrop J, Lang R. Contact, collaboration, and conflict: signal integration of Syk-coupled C-type lectin receptors. *J Immunol* (2017) 198(4):1403–14. doi:10.4049/jimmunol.1601665
- Iborra S, Martinez-Lopez M, Cueto FJ, Conde-Garrosa R, Del Fresno C, Izquierdo HM, et al. *Leishmania* uses Mincle to target an inhibitory ITAM signaling pathway in dendritic cells that dampens adaptive immunity to infection. *Immunity* (2016) 45(4):788–801. doi:10.1016/j.immuni.2016.09.012
- van der Peet PL, Nagata M, Shah S, White JM, Yamasaki S, Williams SJ. Lipid structure influences the ability of glucose monocorynomycolate to signal through Mincle. Org Biomol Chem (2016) 14(39):9267–77. doi:10.1039/ C6OB01781A
- Tima HG, Al Dulayymi JR, Denis O, Lehebel P, Baols KS, Mohammed MO, et al. Inflammatory properties and adjuvant potential of synthetic glycolipids homologous to mycolate esters of the cell wall of *Mycobacterium tuberculosis. J Innate Immun* (2017) 9(2):162–80. doi:10.1159/000450955
- Behler-Janbeck F, Takano T, Maus R, Stolper J, Jonigk D, Tort Tarres M, et al. C-type lectin Mincle recognizes glucosyl-diacylglycerol of *Streptococcus pneumoniae* and plays a protective role in pneumococcal pneumonia. *PLoS Pathog* (2016) 12(12):e1006038. doi:10.1371/journal.ppat.1006038
- Devi S, Rajakumara E, Ahmed N. Induction of Mincle by *Helicobacter pylori* and consequent anti-inflammatory signaling denote a bacterial survival strategy. *Sci Rep* (2015) 5:15049. doi:10.1038/srep15049
- Yamasaki S, Matsumoto M, Takeuchi O, Matsuzawa T, Ishikawa E, Sakuma M, et al. C-type lectin Mincle is an activating receptor for pathogenic fungus, Malassezia. *Proc Natl Acad Sci U S A* (2009) 106(6):1897–902. doi:10.1073/pnas.0805177106
- Ribbing C, Engblom C, Lappalainen J, Lindstedt K, Kovanen PT, Karlsson MA, et al. Mast cells generated from patients with atopic eczema have enhanced levels of granule mediators and an impaired Dectin-1 expression. *Allergy* (2011) 66(1):110–9. doi:10.1111/j.1398-9995.2010.02437.x
- Zhao G, Xu Q, Lin J, Chen W, Cui T, Hu L, et al. The role of Mincle in innate immune to fungal keratitis. *J Infect Dev Ctries* (2017) 11(1):89–97. doi:10.3855/ jidc.7570
- Wells CA, Salvage-Jones JA, Li X, Hitchens K, Butcher S, Murray RZ, et al. The macrophage-inducible C-type lectin, Mincle, is an essential component of the innate immune response to *Candida albicans. J Immunol* (2008) 180(11):7404–13. doi:10.4049/jimmunol.180.11.7404
- Bugarcic A, Hitchens K, Beckhouse AG, Wells CA, Ashman RB, Blanchard H. Human and mouse macrophage-inducible C-type lectin (Mincle) bind Candida albicans. Glycobiology (2008) 18(9):679–85. doi:10.1093/glycob/ cwn046
- Gross O, Gewies A, Finger K, Schafer M, Sparwasser T, Peschel C, et al. Card9 controls a non-TLR signalling pathway for innate anti-fungal immunity. *Nature* (2006) 442(7103):651–6. doi:10.1038/nature04926
- Hara H, Ishihara C, Takeuchi A, Imanishi T, Xue L, Morris SW, et al. The adaptor protein CARD9 is essential for the activation of myeloid cells through ITAM-associated and toll-like receptors. *Nat Immunol* (2007) 8(6):619–29. doi:10.1038/ni1466
- Nagata M, Izumi Y, Ishikawa E, Kiyotake R, Doi R, Iwai S, et al. Intracellular metabolite beta-glucosylceramide is an endogenous Mincle ligand possessing immunostimulatory activity. *Proc Natl Acad Sci U S A* (2017) 114(16): E3285–94. doi:10.1073/pnas.1618133114

- Kiyotake R, Oh-Hora M, Ishikawa E, Miyamoto T, Ishibashi T, Yamasaki S. Human Mincle binds to cholesterol crystals and triggers innate immune responses. *J Biol Chem* (2015) 290(42):25322–32. doi:10.1074/jbc. M115.645234
- Kostarnoy AV, Gancheva PG, Lepenies B, Tukhvatulin AI, Dzharullaeva AS, Polyakov NB, et al. Receptor Mincle promotes skin allergies and is capable of recognizing cholesterol sulfate. *Proc Natl Acad Sci U S A* (2017) 114(13):E2758–65. doi:10.1073/pnas.1611665114
- Brown GD. Sensing necrosis with Mincle. Nat Immunol (2008) 9(10): 1099–100. doi:10.1038/ni1008-1099
- Strasser D, Neumann K, Bergmann H, Marakalala MJ, Guler R, Rojowska A, et al. Syk kinase-coupled C-type lectin receptors engage protein kinase C-sigma to elicit Card9 adaptor-mediated innate immunity. *Immunity* (2012) 36(1):32–42. doi:10.1016/j.immuni.2011.11.015
- Roth S, Bergmann H, Jaeger M, Yeroslaviz A, Neumann K, Koenig PA, et al. Vav proteins are key regulators of Card9 signaling for innate antifungal immunity. *Cell Rep* (2016) 17(10):2572–83. doi:10.1016/j.celrep.2016.11.018
- Schoenen H, Huber A, Sonda N, Zimmermann S, Jantsch J, Lepenies B, et al. Differential control of Mincle-dependent cord factor recognition and macrophage responses by the transcription factors C/EBPbeta and HIF1alpha. *J Immunol* (2014) 193(7):3664–75. doi:10.4049/jimmunol.1301593
- Schweneker K, Gorka O, Schweneker M, Poeck H, Tschopp J, Peschel C, et al. The mycobacterial cord factor adjuvant analogue trehalose-6,6'dibehenate (TDB) activates the Nlrp3 inflammasome. *Immunobiology* (2013) 218(4):664–73. doi:10.1016/j.imbio.2012.07.029
- Desel C, Werninghaus K, Ritter M, Jozefowski K, Wenzel J, Russkamp N, et al. The Mincle-activating adjuvant TDB induces MyD88-dependent Th1 and Th17 responses through IL-1R signaling. *PLoS One* (2013) 8(1):e53531. doi:10.1371/journal.pone.0053531
- Sousa AO, Lee FK, Freiji R, Lagrange PH, Nahmias A. Human immunodeficiency virus infection alters antigen-induced cytokine responses in patients with active mycobacterial diseases. *J Infect Dis* (1998) 177(6):1554–62. doi:10.1086/515326.
- Wevers BA, Kaptein TM, Zijlstra-Willems EM, Theelen B, Boekhout T, Geijtenbeek TB, et al. Fungal engagement of the C-type lectin Mincle suppresses dectin-1-induced antifungal immunity. *Cell Host Microbe* (2014) 15(4):494–505. doi:10.1016/j.chom.2014.03.008
- Patin EC, Willcocks S, Orr S, Ward TH, Lang R, Schaible UE. Mincle-mediated anti-inflammatory IL-10 response counter-regulates IL-12 in vitro. *Innate Immun* (2016) 22(3):181–5. doi:10.1177/1753425916636671
- Lee WB, Kang JS, Choi WY, Zhang Q, Kim CH, Choi UY, et al. Minclemediated translational regulation is required for strong nitric oxide production and inflammation resolution. *Nat Commun* (2016) 7:11322. doi:10.1038/ ncomms11322
- Hunter RL, Hwang SA, Jagannath C, Actor JK. Cord factor as an invisibility cloak? A hypothesis for asymptomatic TB persistence. *Tuberculosis (Edinb)* (2016) 101S:S2–8. doi:10.1016/j.tube.2016.09.023
- Patin EC, Geffken AC, Willcocks S, Leschczyk C, Haas A, Nimmerjahn F, et al. Trehalose dimycolate interferes with FcgammaR-mediated phagosome maturation through Mincle, SHP-1 and FcgammaRIIB signalling. *PLoS One* (2017) 12(4):e0174973. doi:10.1371/journal.pone.0174973
- Rambaruth ND, Jegouzo SA, Marlor H, Taylor ME, Drickamer K. Mouse Mincle: characterization as a model for human Mincle and evolutionary implications. *Molecules* (2015) 20(4):6670–82. doi:10.3390/molecules20046670
- Ostrop J, Jozefowski K, Zimmermann S, Hofmann K, Strasser E, Lepenies B, et al. Contribution of MINCLE-SYK signaling to activation of primary human APCs by mycobacterial cord factor and the novel adjuvant TDB. *J Immunol* (2015) 195(5):2417–28. doi:10.4049/jimmunol.1500102
- Byrareddy SN, Little D, Mayne AE, Villinger F, Ansari AA. Phenotypic and functional characterization of monoclonal antibodies with specificity for rhesus macaque CD200, CD200R and Mincle. *PLoS One* (2015) 10(10):e0140689. doi:10.1371/journal.pone.0140689
- Miyake Y, Toyonaga K, Mori D, Kakuta S, Hoshino Y, Oyamada A, et al. C-type lectin MCL is an FcRgamma-coupled receptor that mediates the adjuvanticity of mycobacterial cord factor. *Immunity* (2013) 38(5):1050–62. doi:10.1016/j. immuni.2013.03.010
- 45. Zhao XQ, Zhu LL, Chang Q, Jiang C, You Y, Luo T, et al. C-type lectin receptor dectin-3 mediates trehalose 6,6'-dimycolate (TDM)-induced Mincle expression through CARD9/Bcl10/MALT1-dependent nuclear factor

(NF)-kappaB activation. J Biol Chem (2014) 289(43):30052-62. doi:10.1074/ jbc.M114.588574

- 46. Kerscher B, Wilson GJ, Reid DM, Mori D, Taylor JA, Besra GS, et al. Mycobacterial receptor, Clec4d (CLECSF8, MCL), is coregulated with Mincle and upregulated on mouse myeloid cells following microbial challenge. *Eur J Immunol* (2016) 46(2):381–9. doi:10.1002/eji.201545858
- Miyake Y, Masatsugu OH, Yamasaki S. C-type lectin receptor MCL facilitates Mincle expression and signaling through complex formation. *J Immunol* (2015) 194(11):5366–74. doi:10.4049/jimmunol.1402429
- Kerscher B, Dambuza IM, Christofi M, Reid DM, Yamasaki S, Willment JA, et al. Signalling through MyD88 drives surface expression of the mycobacterial receptors MCL (Clecsf8, Clec4d) and Mincle (Clec4e) following microbial stimulation. *Microbes Infect* (2016) 18(7–8):505–9. doi:10.1016/j. micinf.2016.03.007
- 49. Wu XY, Guo JP, Yin FR, Lu XL, Li R, He J, et al. Macrophage-inducible C-type lectin is associated with anti-cyclic citrullinated peptide antibodies-positive rheumatoid arthritis in men. *Chin Med J* (*Engl*) (2012) 125(17):3115–9.
- Nakamura N, Shimaoka Y, Tougan T, Onda H, Okuzaki D, Zhao H, et al. Isolation and expression profiling of genes upregulated in bone marrowderived mononuclear cells of rheumatoid arthritis patients. *DNA Res* (2006) 13(4):169–83. doi:10.1093/dnares/dsl006
- Suzuki Y, Nakano Y, Mishiro K, Takagi T, Tsuruma K, Nakamura M, et al. Involvement of Mincle and Syk in the changes to innate immunity after ischemic stroke. *Sci Rep* (2013) 3:3177. doi:10.1038/srep03177
- Arumugam TV, Manzanero S, Furtado M, Biggins PJ, Hsieh YH, Gelderblom M, et al. An atypical role for the myeloid receptor Mincle in central nervous system injury. J Cereb Blood Flow Metab (2017) 37(6):2098–111. doi:10.1177/0271678X16661201
- Saravanan C, Cao Z, Head SR, Panjwani N. Analysis of differential expression of glycosyltransferases in healing corneas by glycogene microarrays. *Glycobiology* (2010) 20(1):13–23. doi:10.1093/glycob/cwp133
- Jia R, Hashizume-Takizawa T, Du Y, Yamamoto M, Kurita-Ochiai T. Aggregatibacter actinomycetemcomitans induces Th17 cells in atherosclerotic lesions. *Pathog Dis* (2015) 73(3):ftu027. doi:10.1093/femspd/ftu027
- Sharma A, Steichen AL, Jondle CN, Mishra BB, Sharma J. Protective role of Mincle in bacterial pneumonia by regulation of neutrophil mediated phagocytosis and extracellular trap formation. J Infect Dis (2014) 209(11):1837–46. doi:10.1093/infdis/jit820
- Sharma A, Simonson TJ, Jondle CN, Mishra BB, Sharma J. Mincle-mediated neutrophil extracellular trap formation by regulation of autophagy. J Infect Dis (2017) 215(7):1040–8. doi:10.1093/infdis/jix072
- Greco SH, Torres-Hernandez A, Kalabin A, Whiteman C, Rokosh R, Ravirala S, et al. Mincle signaling promotes con A hepatitis. *J Immunol* (2016) 197(7):2816–27. doi:10.4049/jimmunol.1600598
- Zhou H, Yu M, Zhao J, Martin BN, Roychowdhury S, McMullen MR, et al. IRAKM-Mincle axis links cell death to inflammation: pathophysiological implications for chronic alcoholic liver disease. *Hepatology* (2016) 64(6):1978–93. doi:10.1002/hep.28811
- Brown GD, Crocker PR. Lectin receptors expressed on myeloid cells. *Microbiol Spectr* (2016) 4(5):MCHD-0036. doi:10.1128/microbiolspec. MCHD-0036-2016
- Mayer S, Raulf MK, Lepenies B. C-type lectins: their network and roles in pathogen recognition and immunity. *Histochem Cell Biol* (2017) 147(2): 223–37. doi:10.1007/s00418-016-1523-7
- Naser SA, Ghobrial G, Romero C, Valentine JF. Culture of *Mycobacterium avium* subspecies paratuberculosis from the blood of patients with Crohn's disease. *Lancet* (2004) 364(9439):1039–44. doi:10.1016/S0140-6736(04) 17058-X
- Alcedo KP, Thanigachalam S, Naser SA. RHB-104 triple antibiotics combination in culture is bactericidal and should be effective for treatment of Crohn's disease associated with *Mycobacterium paratuberculosis*. *Gut Pathog* (2016) 8:32. doi:10.1186/s13099-016-0115-3
- Borody TJ, Leis S, Warren EF, Surace R. Treatment of severe Crohn's disease using antimycobacterial triple therapy – approaching a cure? *Dig Liver Dis* (2002) 34(1):29–38. doi:10.1016/S1590-8658(02)80056-1
- Ebringer A, Rashid T, Tiwana H, Wilson C. A possible link between Crohn's disease and ankylosing spondylitis via *Klebsiella* infections. *Clin Rheumatol* (2007) 26(3):289–97. doi:10.1007/s10067-006-0391-2

- Liu Y, van Kruiningen HJ, West AB, Cartun RW, Cortot A, Colombel JF. Immunocytochemical evidence of *Listeria*, *Escherichia coli*, and *Streptococcus* antigens in Crohn's disease. *Gastroenterology* (1995) 108(5):1396–404. doi:10.1016/0016-5085(95)90687-8
- 66. Miranda-Bautista J, Padilla-Suarez C, Bouza E, Munoz P, Menchen L, Marin-Jimenez I. *Listeria monocytogenes* infection in inflammatory bowel disease patients: case series and review of the literature. *Eur J Gastroenterol Hepatol* (2014) 26(11):1247–52. doi:10.1097/MEG.00000000000188
- Rashid T, Ebringer A, Wilson C. The role of *Klebsiella* in Crohn's disease with a potential for the use of antimicrobial measures. *Int J Rheumatol* (2013) 2013;610393. doi:10.1155/2013/610393
- Mazzarella G, Perna A, Marano A, Lucariello A, Aufiero VR, Melina R, et al. Pathogenic role of associated adherent-invasive *Escherichia coli* in Crohn's disease. *J Cell Physiol* (2017) 232(10):2860–8. doi:10.1002/jcp.25717
- Shea KM, Edelsberg J, Weycker D, Farkouh RA, Strutton DR, Pelton SI. Rates of pneumococcal disease in adults with chronic medical conditions. *Open Forum Infect Dis* (2014) 1(1):ofu024. doi:10.1093/ofid/ofu024
- Long MD, Farraye FA, Okafor PN, Martin C, Sandler RS, Kappelman MD. Increased risk of pneumocystis jiroveci pneumonia among patients with inflammatory bowel disease. *Inflamm Bowel Dis* (2013) 19(5):1018–24. doi:10.1097/MIB.0b013e3182802a9b
- Mimouna S, Bazin M, Mograbi B, Darfeuille-Michaud A, Brest P, Hofman P, et al. HIF1A regulates xenophagic degradation of adherent and invasive *Escherichia coli* (AIEC). *Autophagy* (2014) 10(12):2333–45. doi:10.4161/15548627.2014.984275
- Dhillon SS, Mastropaolo LA, Murchie R, Griffiths C, Thoni C, Elkadri A, et al. Higher activity of the inducible nitric oxide synthase contributes to very early onset inflammatory bowel disease. *Clin Transl Gastroenterol* (2014) 5:e46. doi:10.1038/ctg.2013.17
- Soufli I, Toumi R, Rafa H, Touil-Boukoffa C. Overview of cytokines and nitric oxide involvement in immuno-pathogenesis of inflammatory bowel diseases. *World J Gastrointest Pharmacol Ther* (2016) 7(3):353–60. doi:10.4292/wjgpt. v7.i3.353
- 74. Wolfkamp SC, Te Velde AA, Weersma RK, Ponsioen CY, Stokkers PC. Is there a role for Crohn's disease-associated autophagy genes ATG16L1 and IRGM in formation of granulomas? *Eur J Gastroenterol Hepatol* (2010) 22(8):933–7. doi:10.1097/MEG.0b013e32833775e6
- Lang R. Recognition of the mycobacterial cord factor by Mincle: relevance for granuloma formation and resistance to tuberculosis. *Front Immunol* (2013) 4:5. doi:10.3389/fimmu.2013.00005
- Sato IY, Kobayashi K, Yamagata N, Shikama Y, Kasama T, Kasahara K, et al. Modulation of granuloma formation in vitro by endogenous mediators. *Immunopharmacology* (1991) 21(2):73–82. doi:10.1016/ 0162-3109(91)90010-V
- Baram L, Cohen-Kedar S, Spektor L, Elad H, Guzner-Gur H, Dotan I. Differential stimulation of peripheral blood mononuclear cells in Crohn's disease by fungal glycans. *J Gastroenterol Hepatol* (2014) 29(12):1976–84. doi:10.1111/jgh.12701
- 78. Jyonouchi H, Geng L, Cushing-Ruby A, Monteiro IM. Aberrant responses to TLR agonists in pediatric IBD patients; the possible association with increased production of Th1/Th17 cytokines in response to *Candida*, a luminal antigen. *Pediatr Allergy Immunol* (2010) 21(4 Pt 2):e747–55. doi:10.1111/j.1399-3038.2009.00923.x
- 79. Xu X, Sun S, Xie F, Ma J, Tang J, He S, et al. Advanced oxidation protein products induce epithelial-mesenchymal transition of intestinal epithelial cells via a PKC delta-mediated, redox-dependent signaling pathway. *Antioxid Redox Signal* (2017) 27(1):37–56. doi:10.1089/ars.2015.6611
- Fairfax BP, Humburg P, Makino S, Naranbhai V, Wong D, Lau E, et al. Innate immune activity conditions the effect of regulatory variants upon monocyte gene expression. *Science* (2014) 343(6175):1246949. doi:10.1126/ science.1246949
- Sokol H, Conway KL, Zhang M, Choi M, Morin B, Cao Z, et al. Card9 mediates intestinal epithelial cell restitution, T-helper 17 responses, and control of bacterial infection in mice. *Gastroenterology* (2013) 145(3):591–601e3. doi:10.1053/j.gastro.2013.05.047
- Pilmis B, Puel A, Lortholary O, Lanternier F. New clinical phenotypes of fungal infections in special hosts. *Clin Microbiol Infect* (2016) 22(8):681–7. doi:10.1016/j.cmi.2016.05.016

- Nasir A, El Bahesh E, Whitten C, Lawson A, Udall JN Jr. Pityrosporum folliculitis in a Crohn's disease patient receiving infliximab. *Inflamm Bowel* Dis (2010) 16(1):7–8. doi:10.1002/ibd.20928
- Marcoval J, Penin RM, Sabe N, Valenti-Medina F, Bonfill-Orti M, Martinez-Molina L. Cutaneous leishmaniasis associated with anti-tumour necrosis factor-alpha drugs: an emerging disease. *Clin Exp Dermatol* (2017) 42(3):331–4. doi:10.1111/ced.13061
- Castano-Rodriguez N, Kaakoush NO, Lee WS, Mitchell HM. Dual role of *Helicobacter* and *Campylobacter* species in IBD: a systematic review and meta-analysis. *Gut* (2017) 66(2):235–49. doi:10.1136/gutjnl-2015-310545
- Mueller C. Danger-associated molecular patterns and inflammatory bowel disease: is there a connection? *Dig Dis* (2012) 30(Suppl 3):40-6. doi:10.1159/000342600
- Hupfer T, Schick J, Jozefowski K, Voehringer D, Ostrop J, Lang R. Stat6-Dependent inhibition of Mincle expression in mouse and human antigenpresenting cells by the Th2 cytokine IL-4. *Front Immunol* (2016) 7:423. doi:10.3389/fimmu.2016.00423
- te Velde AA, van Kooyk Y, Braat H, Hommes DW, Dellemijn TA, Slors JF, et al. Increased expression of DC-SIGN+IL-12+IL-18+ and CD83+IL-12-IL-18dendritic cell populations in the colonic mucosa of patients with Crohn's disease. *Eur J Immunol* (2003) 33(1):143–51. doi:10.1002/immu.200390017

- Smith AM, Rahman FZ, Hayee B, Graham SJ, Marks DJ, Sewell GW, et al. Disordered macrophage cytokine secretion underlies impaired acute inflammation and bacterial clearance in Crohn's disease. *J Exp Med* (2009) 206(9):1883–97. doi:10.1084/jem.20091233
- Wong BR, Grossbard EB, Payan DG, Masuda ES. Targeting Syk as a treatment for allergic and autoimmune disorders. *Expert Opin Investig Drugs* (2004) 13(7):743–62. doi:10.1517/13543784.13.7.743
- Nijjar JS, Tindell A, McInnes IB, Siebert S. Inhibition of spleen tyrosine kinase in the treatment of rheumatoid arthritis. *Rheumatology (Oxford)* (2013) 52(9):1556–62. doi:10.1093/rheumatology/ket225

Conflict of Interest Statement: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 te Velde. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.