



CASE REPORT

A Case of Subcutaneous and Intranasal Phaeohyphomycosis Caused by *Microsphaeropsis arundinis* in an Immunocompromised Patient Misdiagnosed with Mucormycosis

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Microsphaeropsis arundinis is a dematiaceous fungus capable of causing soft tissue infections known as phaeohyphomycosis, mostly in immunocompromised individuals. These infections arise from the traumatic inoculation of fungal materials into the subcutis, and can spread to adjacent subcutaneous tissues or via the lymphatics in a sporotrichoid manner. A 76-year-old man presented with diffuse erythematous plaques and swelling on both forearms and dorsal hands, and rhinalgia. He had been undergoing treatment for hypertension, angina pectoris, and diabetes. Histopathologic examinations of the skin, painful nasal septum, and molecular identification using internal transcribed spacer regions confirmed a diagnosis of subcutaneous and intranasal phaeohyphomycosis caused by *M. arundinis*. The patient was treated with oral itraconazole for over 5 months, and no recurrence was observed until the time of writing this manuscript. We report a rare case of subcutaneous and intranasal phaeohyphomycosis caused by *M. arundinis* and propose that confirmation of the causative strains is necessary, as it could affect the prognosis and treatment of the disease. (*Ann Dermatol* 31(5) 571~575, 2019)

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-Keywords-

Fungal infection, *Microsphaeropsis arundinis*, Phaeohyphomycosis

INTRODUCTION

Coelomycete is an emerging group of pathogens capable of causing soft tissue infections in immunocompromised patients. These infections are classified as phaeohyphomycosis, and they are caused by dematiaceous fungi (pigmented hyphae)¹⁻³. *Microsphaeropsis arundinis*, a member of the class Coelomycetes, was first detected from the “giant reed” *Arundo donax* in Pakistan, and it usually inhabits terrestrial plant hosts and is ubiquitous in soil and fresh water environments³. Herein, we experienced a rare case of subcutaneous and intranasal phaeohyphomycosis caused by *M. arundinis*; we identified the fungal organism based on internal transcribed spacer (ITS) sequence analysis. To the best of our knowledge, this is the first such case in South Korea.

CASE REPORT

A 76-year-old Korean man presented with cutaneous lesions on both his forearms and dorsal hands. He was a farmer and was in close contact with soil for several decades. When he visited our clinic with enlarged skin lesions that had first appeared 6 months ago, he also complained of asymptomatic diffuse erythematous plaques and swelling with scales and pustules on both his forearms and dorsal hands (Fig. 1A). Subsequently, he reported having experienced rhinalgia a month ago. There

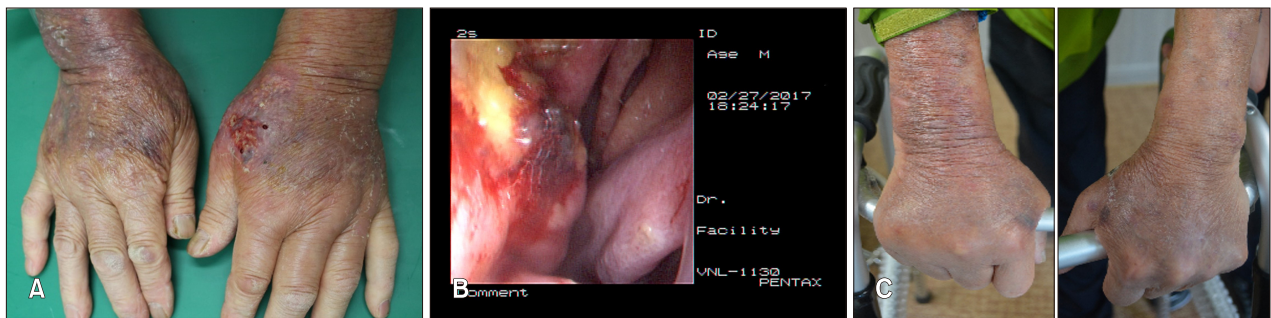


Fig. 1. (A) Diffuse purulent erythematous plaques and swelling with scales and pustules on both dorsal hands and forearms of the patient; (B) necrotic changes in the nasal septum with hemorrhage and exudate; (C) and completely recovered lesions after treatment with itraconazole for 5 months.

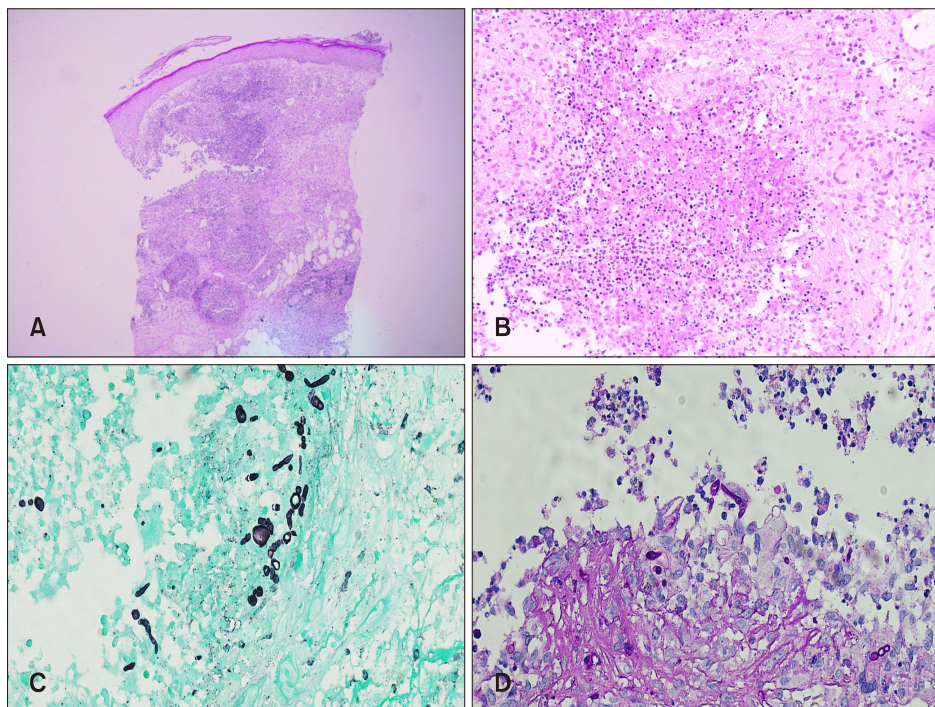


Fig. 2. (A, B) Dense infiltrates of inflammatory cells such as neutrophils, monocytes, histiocytes, and giant cells extending from the upper dermis through the subcutaneous fatty layer (H&E: A, ×40; B, ×200); (C, D) Gomori's methenamine silver (GMS) staining and periodic acid-Schiff (PAS) staining revealed fungal hyphae and spores (C: GMS, ×400; D: PAS, ×400).

was no fever or lymphadenopathy, and both respiratory and neurological examinations were unremarkable. The patient was under medication for hypertension, angina pectoris, and diabetes mellitus. He was suspected as having bacterial, atypical mycobacterial, or deep fungal infections, and a skin biopsy was subsequently performed from the lesion. Bacterial and fungal cultures were taken, and the cutaneous tuberculosis (TBC) polymerase chain reaction (PCR) test (BAG Health Care GmbH, Lich, Germany) was performed. In addition, the patient was referred to the otorhinolaryngology department for rhinalgia. Otorhinolaryngology examinations revealed necrotic changes accompanied by hemorrhage in the nasal septum (Fig. 1B); subsequently, a biopsy was performed at that site.

The findings raised a suspicion of mucormycosis. Hence, other fungal infections, including mucormycosis, were considered as differential diagnoses, and further examinations were performed. The patient was tested for aspergillus antigen and (1-3)- β -D-glucan. Invasive involvement of both arms as well as nasal invasion was observed; therefore, chest x-ray and bone scan were performed to check for more invasive lesions. Bone scan revealed cellulitis; however, chest x-ray showed no remarkable findings. Histopathologic examination of the skin showed dense infiltrates of inflammatory cells such as neutrophils, monocytes, histiocytes, and giant cells extending from the upper dermis through the subcutaneous fatty layer (Fig. 2A, B). Staining for acid-fast bacilli showed negative results; sim-

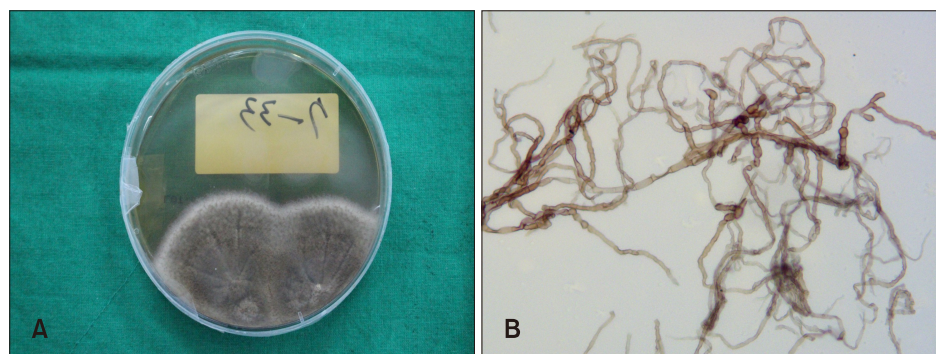


Fig. 3. (A) *Microsphaeropsis arundinis* colonies growing on Sabouraud dextrose agar at 30°C demonstrating gray to dark green colonies (14 days); (B) microscopic morphology of *M. arundinis* stained with lactophenol cotton blue showing pigmented, septate, and irregularly formed hyphae, with swollen segments.

ilarly, PCR for mycobacteria and bacterial culture showed negative results. Gomori's methenamine silver staining and periodic acid-Schiff staining showed scattered fungal spores and hyphae (Fig. 2C, D). The histopathologic findings of the nasal septal lesions were similar to the results of skin biopsy. Through consultation with the Department of Infectious Diseases, the blood tests revealed negative results for aspergillus antigen and positive results for (1-3)- β -D-glucan (100.0, positive >80). The cultures of the biopsy specimen and nasal septum tissue grew a dematiaceous (pigmented) fungus; microscopic examination showed short and thick irregularly shaped septate hyphae (Fig. 3). Owing to the absence of conidia, the organism could not be morphologically identified. Hence, gene sequencing analysis was performed for molecular identification of the organism. The sequence of the D1/D2 domain of ribosomal RNA gene regions and ITS1-5.8S-ITS2 regions, using primers and standard sequencing methods, showed identity with *M. arundinis* (100% sequence identity to *M. arundinis* cystathionine beta-synthase (CBS) 100243 [GenBank accession number JX496123.1] and 100% sequence identity to *M. arundinis* 0012 [GenBank accession number KY992587.1])⁴. Based on these findings, the patient was diagnosed with subcutaneous and intranasal phaeohyphomycosis caused by *M. arundinis* (Fig. 4).

The patient was initially administered fluconazole 200 mg and cefbuperazone 2 g daily. This was changed to itraconazole 200 mg/d and isoconazole topical cream after the presence of pathogenic organism was confirmed; subsequently, itraconazole was administered for more than 5 months. Thereafter, the lesion completely disappeared and the patient no longer complained of rhinalgia; there was no subsequent relapse of infection in cutaneous lesions (Fig. 1C).

DISCUSSION

In 2004, Pendle et al.¹ described infections caused by *M.*

arundinis in patients with diabetes mellitus and chronic kidney disease. Since then, cutaneous infections caused by the same pathogen have been reported in the United States, Japan, and Australia, although it is still a rare opportunistic infection that occurs in immunocompromised patients^{2,3,5}. Especially in South Korea, phaeohyphomycosis due to *Exophiala* spp. has been reported in several cases⁶⁻⁸; however, to the best of our knowledge, this is the first case of phaeohyphomycosis caused by *M. arundinis*. In addition, it is a very rare case owing to the involvement of the nasal septum as well as the skin.

Phaeohyphomycosis, first named in 1974, can occur in various clinical forms from subcutaneous mycosis to deep organ infections⁹. Especially, the subcutaneous form tends to occur at the distal limbs, which might be easily exposed to trauma or infections⁶. *M. arundinis* infection can occur in the form of subacute onset on the limbs and can spread around by direct inoculation of mold-contaminated materials⁵. In this case, as there were no signs indicative of systemic involvement, the occurrence of phaeohyphomycosis in the nasal septum could be attributed to the direct contact of *M. arundinis* through the hands of the patient.

The histopathologic findings of phaeohyphomycosis are very typical, as observed in the present case, although identifying the causative pathogen is always challenging. The coelomycete fungi produce a blister-like fruiting structure called conidiomata, which differ from other fungi, and the size and shape of the conidia of *M. arundinis* differ from those of other coelomycete genera. However, the morphological diagnosis of *M. arundinis* is always challenging owing to poor sporulation^{10,11}. Therefore, species identification is often dependent on DNA sequencing using ITS, as in this case.

Phaeohyphomycosis is primarily treated by surgical excision alone, in cases with localized and small well-defined lesions. However, in case of wide or ambiguous cutaneous lesions, surgical treatment is difficult and systemic medications should be administered^{7,8}. In all the eight cases of *M. arundinis* infections reported so far, diffuse in-

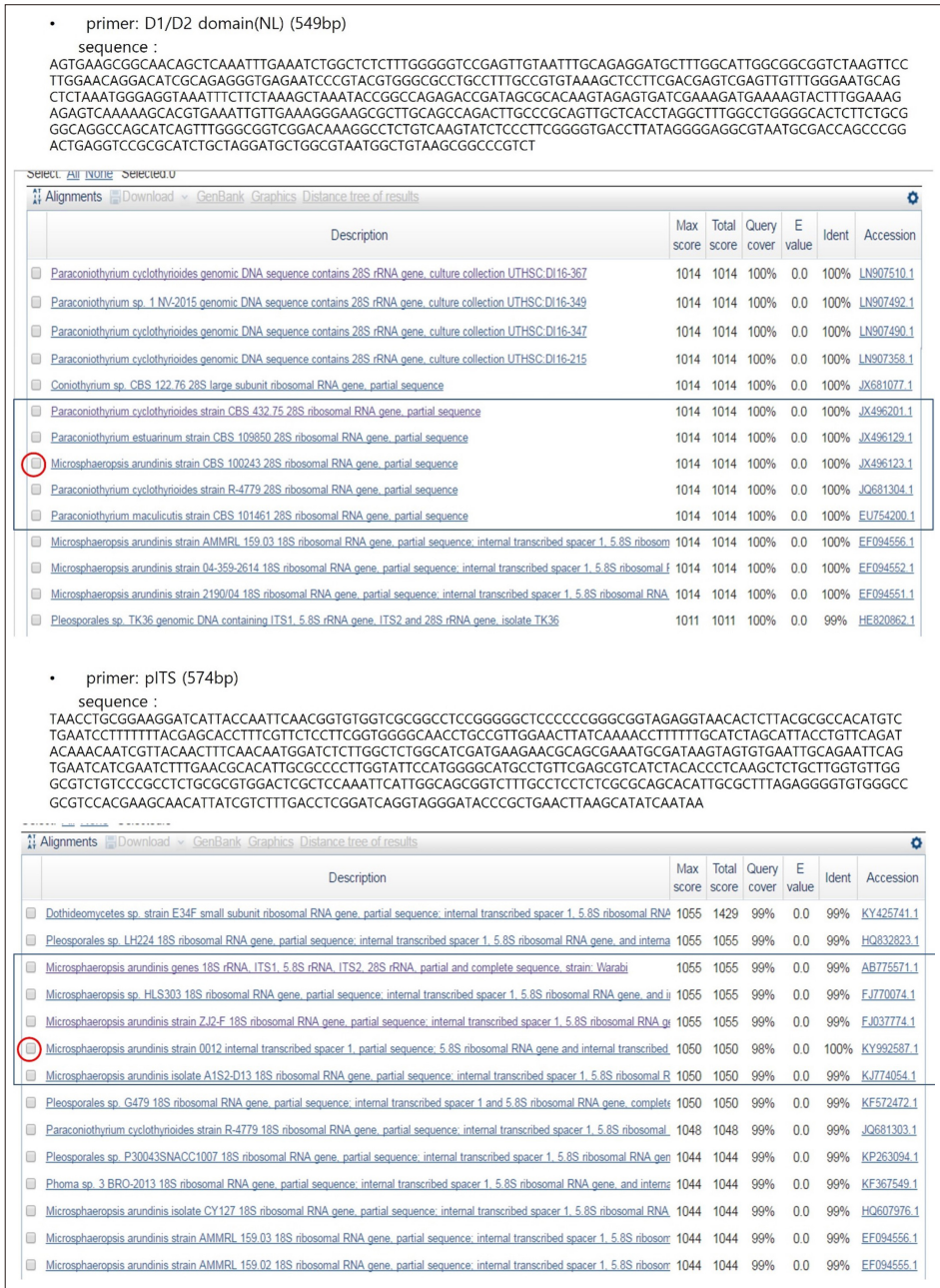


Fig. 4. The sequence of the D1/D2 domain of ribosomal RNA gene regions and ITS1-5.8S-ITS2 regions showed identity with *Microsphaeropsis arundinis* (100% sequence identity to *M. arundinis* CBS 100243 [GenBank accession number JX496123.1] and 100% sequence identity to *M. arundinis* 0012 [GenBank accession number KY992587.1]).

fection was observed and no surgical excision was performed; treatment involved use of antifungal agents and thermotherapy. This suggests that *M. arundinis* infection may demonstrate a diffuse form rather than localized or solitary nodular and cystic subcutaneous appearance. Triazole antifungal agents have been considered as the first-line therapy for phaeohyphomycosis. Besides, clinical data related to non-azole antifungal therapy for *M. arundinis* infection are limited, and the minimum inhibitory concentration of itraconazole for *M. arundinis* has been reported as ≤ 0.25 mg/L^{2,5,12}. Therefore, oral itraconazole was used for treatment of phaeohyphomycosis in this case, although

the lack of susceptibility test was considered as a limitation. To the best of our knowledge, this is a rare case of subcutaneous and intranasal phaeohyphomycosis caused by *M. arundinis*, and our patient was also immunocompromised, undergoing treatment for diabetes mellitus, similar to the patients in previous reports. He might have been misdiagnosed with mucormycosis based on the findings of nasal septal lesion biopsy and ENT examinations, although the pathogen was finally confirmed using gene sequencing analysis. Positive (1-3)- β -D-glucan in blood is indicative of invasive or disseminated fungal infections. This

test can be helpful for the detection of *Aspergillus*, *Candida*, and *Pneumocystis jirovecii*, which do not produce (1,3)- β -D-glucan¹³. This could help to rule out mucormycosis. Clinically, subcutaneous phaeohyphomycosis manifests mainly in nodular or cystic form in the limbs of immunosuppressed patients, whereas primary cutaneous mucormycosis is relatively rare, and malodorous brown or black necrotic lesions with an erythematous border are mainly observed¹⁴. Clinical differential diagnosis and gene sequencing analysis were used to diagnose phaeohyphomycosis in our case. Efforts should be made to identify the pathogenic strains that can influence the prognosis and treatment of this infectious disease.

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CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

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REFERENCES

1. Pendle S, Weeks K, Priest M, Gill A, Hudson B, Kotsiou G, et al. Phaeohyphomycotic soft tissue infections caused by the coelomycetous fungus *Microsphaeropsis arundinis*. *J Clin Microbiol* 2004;42:5315-5319.
2. Asahina A, Kobayashi M, Nakano K, Saito I, Yarita K, Kamei K, et al. Deep cutaneous infection with *Microsphaeropsis arundinis*: report of two Japanese cases. *Acta Derm Venereol* 2015;95:855-857.
3. Hall MR, Brumble LM, Mayes MA, Snow JL, Keeling JH. Cutaneous *Microsphaeropsis arundinis* infection initially interpreted as squamous cell carcinoma. *Int J Dermatol* 2013;52:84-86.
4. Kim MN, Shin JH, Sung H, Lee K, Kim EC, Ryoo N, et al. *Candida haemulonii* and closely related species at 5 university hospitals in Korea: identification, antifungal susceptibility, and clinical features. *Clin Infect Dis* 2009;48:e57-e61.
5. Crawford SJ, Chen SC, Halliday C, Rangan GK, Gottlieb T, Reid AB. *Microsphaeropsis arundinis* skin and soft tissue infection in renal transplant recipients: three case reports and a review of the literature. *Transpl Infect Dis* 2015;17:915-920.
6. Lee KC, Kim MJ, Chae SY, Lee HS, Jang YH, Lee SJ, et al. A case of phaeohyphomycosis caused by *Exophiala lecaniicorni*. *Ann Dermatol* 2016;28:385-387.
7. Ryu TH, Kwon IH, Choi JE, Ahn HH, Kye YC, Seo SH. A case of subcutaneous phaeohyphomycosis caused by *Exophiala oligosperma* showing multiple cysts. *Korean J Dermatol* 2017;55:259-263.
8. Suh MK, Kwon SW, Kim TH, Sun YW, Lim JW, Ha GY, et al. A case of subcutaneous phaeohyphomycosis caused by *Exophiala jeanselmei*. *Korean J Dermatol* 2005;43:124-127.
9. Ajello L, Georg LK, Steigbigel RT, Wang CJ. A case of phaeohyphomycosis caused by a new species of *Phialophora*. *Mycologia* 1974;66:490-498.
10. Sutton DA. Coelomycetous fungi in human disease. A review: clinical entities, pathogenesis, identification and therapy. *Rev Iberoam Micol* 1999;16:171-179.
11. Stchigel AM, Sutton DA. Coelomycete fungi in the clinical lab. *Curr Fungal Infect Rep* 2013;7:171-191.
12. Chowdhary A, Meis JF, Guarro J, de Hoog GS, Kathuria S, Arendrup MC, et al. ESCMID and ECMM joint clinical guidelines for the diagnosis and management of systemic phaeohyphomycosis: diseases caused by black fungi. *Clin Microbiol Infect* 2014;20 Suppl 3:47-75.
13. Ostrosky-Zeichner L, Alexander BD, Kett DH, Vazquez J, Pappas PG, Saeki F, et al. Multicenter clinical evaluation of the (1 \rightarrow 3) beta-D-glucan assay as an aid to diagnosis of fungal infections in humans. *Clin Infect Dis* 2005;41:654-659.
14. Perusquía-Ortiz AM, Vázquez-González D, Bonifaz A. Opportunistic filamentous mycoses: aspergillosis, mucormycosis, phaeohyphomycosis and hyalohyphomycosis. *J Dtsch Dermatol Ges* 2012;10:611-621; quiz 621-622.