

EML4-ALK translocation identification in RNA exosomal cargo (*ExoALK*) in NSCLC patients: a novel role for liquid biopsy

Pablo Reclusa^{1,2#}, Jean-François Laes^{3#}, Umberto Malapelle⁴, Anna Valentino⁵, Danilo Rocco⁶, Ignacio Gil-Bazo^{7,8}, Christian Rolfo^{1,9}

¹Phase I-Early Clinical Trials Unit, Oncology Department, Antwerp University Hospital (UZA) & Center for Oncological Research of Antwerp (CORE) Antwerp University, Antwerp, Belgium; ²Department of Surgical, Oncological and Stomatological Disciplines, University of Palermo, Palermo, Italy; ³OncoDNA SA, 6041 Gosselies, Belgium; ⁴Department of Public Health, University of Naples Federico II, Naples, Italy; ⁵Gene Expression and Molecular Genetics Laboratory, Institute of Biosciences and BioResources, National Council of Research, CNR, Naples, Italy; ⁶Oncology Unit, A.O.R.N. Vincenzo Monaldi – Ospedale dei Colli, Naples, Italy; ⁷Department of Oncology, Clínica Universidad de Navarra & Navarra Health Research Institute (IDISNA), Pamplona, Spain; ⁸Centro de Investigación Biomédica en Red de Cáncer (CIBERONC), Madrid, Spain; ⁹University of Maryland Marlene and Stewart Greenebaum Comprehensive Cancer Center, Baltimore, MD, USA

"These authors contributed equally to this work.

Correspondence to: Prof. Christian Rolfo, MD, PhD, MBA. University of Maryland Marlene and Stewart Greenebaum Comprehensive Cancer Center, Baltimore, MD, USA. Email: christian.rolfo@umm.edu.

Abstract: The introduction of druggable targets has significantly improved the outcomes of non-small cell lung cancer patients (NSCLC). *EML4-ALK* translocation represents 4–6% of the druggable alterations in NSCLC. With the approval of Crizotinib, first discovered drug for the *EML4-ALK* translocation, on first line treatment for patients with detected mutation meant a complete change on the treatment landscape. The current standard method for *EML4-ALK* identification is immunohistochemistry or FISH in a tumor biopsy. However, a big number of NSCLC patients have not tissue available for analysis and others are not suitable for biopsy due to their physical condition or the location of the tumor. Liquid biopsy seems the best alternative for identification in these patients that have no tissue available. Circulating free RNA has not been validated for the identification of this mutation. As a complementary tool, exosomes might represent a good tool for predictive biomarkers study, and due to their stability, they preserve the genetic material contained in them. Our group has described for the first time the translocation *EML4-ALK* in RNA isolated from exosomes derived from NSCLC patients using next generation sequencing.

Keywords: Exosomes; liquid biopsy; ALK translocation; non-small cell lung cancer (NSCLC); biomarkers

Submitted Oct 07, 2018. Accepted for publication Oct 17, 2018. doi: 10.21037/tcr.2018.11.35 **View this article at:** http://dx.doi.org/10.21037/tcr.2018.11.35

Introduction

Lung cancer treatment landscape has completely changed in the last decade. The introduction of druggable targets in the therapeutic scenario gave the opportunity to significantly improve the outcomes in the most lethal malignant disease. *EML4-ALK* translocation represents a story of success in drug development. The discovery of the aberration in tumors of patients with lung cancer in 2007 (1) soon translated into a rapid approval of the first ALK tyrosine kinase inhibitor (TKI), crizotinib (2), and the subsequent early arrival of second and third generation TKIs, all happening in roughly a decade. The current standard for the identification of the target necessarily implies identification of rearrangement in the tumor tissue either by FISH or immunohistochemistry. Unfortunately, despite the efforts, a big number of non-small cell lung cancer (NSCLC) patients have no tissue available for determination. The isolation of ctDNA is a novel approach, but still not validated for *EML4-ALK* translocation (3,4). Exosomes, new members of the liquid biopsy family, carrying genetic material represent an important tool for biomarkers discovery (5). Our group has identified, for the first time, *EML4-ALK* translocation in exosomes (*ExoALK*), using a next generation sequencing technique.

Translational Cancer Research, Vol 8, Suppl 1 January 2019



Figure 1 Schematic representation of ExoALK proof of concept study.

Material and methods

Patients selection

Patients with confirmed diagnosis of stage IIIB–IV NSCLC, naïve or under treatment with known *ALK* status were eligible for this proof of concept study. *EML4-ALK* translocation in tumor tissue was performed by FISH according to the clinical records of the patients. Patients gave their informed consent before the sample collection according to the institution regulation. Blood samples were collected only at a single point. Clinical data were collected from the electronic medical records. Samples and data from patients included in the study were provided by the Biobank of the University of Navarra (Reg#: B_0000612) and were processed following standard operating procedures approved by the Ethical and Scientific Committees. The trial was approved by the Antwerp University Hospital ethical committee n. 14/17/206.

Exosomal RNA isolation

The exosomal RNA cargo was extracted from 1 mL of plasma using ExoRNeasy Kit from QIAGEN with a modification of the original protocol to avoid loss of material and volume. This includes 3 centrifugations of 5' × 500 g, $15' \times 3,000$ g and $30' \times 10,000$ g instead of the filtration in 0.8 μ M (*Figure 1*).

Next generation sequencing

The fusions were analyzed using the Ion Ampliseq RNA Fusion Lung Cancer panel (Thermo Fisher Scientific, Waltham, USA). The primers used for amplification were then partially digested by the Pfu enzyme. The product of digestion was ligated with corresponding barcoded adapters and purified using Ampure Beads (Agilent Genomics Inc). The product of purification was amplified for 5 more cycles and subsequently purified using Ampure Beads. Ten pM of each library was loaded into the IonChef system (Thermo Fisher Scientific, Waltham, USA) for the emulsion polymerase chain reaction (PCR) and then loaded in the chip.

The quality of the data was assessed using the Torrent suite software (Thermo Fisher Scientific, Waltham, USA) associated with the sequencing machine. The minimum read length for fusion detection was set to 50 bp (base pair). We consider that under this value the reads were too short to overlap targeted fusion breakpoint. In addition, to assess the detection quality, at least 20,000 reads were required per sample (*Figure 1*).

Results

A total of 19 patients with plasma samples were included in the study, 16 of them harboring *EML4-ALK* translocations in tissue. In 1 mL of plasma, the RNA concentration ranged from 0.5 to 121 mg/mL. In total, 17 patients were finally eligible for extracellular vesicles analysis. In nine patients, we were able to identify the translocation in the exosomal RNA. The concordance between tissue and exosomes was 64% (9 out of 14 patients). All three patients being negative for the fusion gene in tissue resulted also negative in the *ExoALK* analysis, representing a specificity of 100% (*Table 1*). No correlation was found between the RNA yield and the type of sample or its clinical variables.

Discussion

Here we demonstrate for the first time that EML4-ALK

Exosomal RNA (<i>ExoALK</i>) (n=17)	Tissue (patients n=19)	
	Positive	Negative
Positive	9	0
Negative	5	3
Degraded RNA	2	-
a	1000/	

Table 1 Concordance between tissue and ExoALK

Sensitivity =64%; Specificity =100%.

translocation detection in exosomes of NSCLC patients is feasible, with a high sensitivity and specificity. The exosomal analysis represents an opportunity for patients therapeutic selection with a minimal invasive procedure, easy to perform and with an affordable cost. The development of new techniques with a direct detection of the fusion by capture instead of by amplification would be the key to improve the sensitivity of *ExoALK* in the future.

Acknowledgments

The authors acknowledge OncoDNA for the free research collaboration, Dr. Maxime Lienard from OncoDNA for the contribution in exosomal cargo analysis and Prof. Patrick Pauwels from Molecular Pathology Unit at Antwerp University Hospital for the data analysis. Human samples and data from patients included in the study were provided by Biobank@UZA (Antwerp, Belgium; ID: BE71030031000), the AORN Vincenzo Monaldi, Department of Oncology, Naples, Italy and were processed following standard operating procedures approved by the Ethical and Scientific Committees. We particularly acknowledge the patients for their participation and the Biobank of the University of Navarra (Reg#: B_0000612) for its collaboration.

Funding: P Reclusa received a scholarship grant from University of Palermo.

Footnote

Provenance and Peer Review: The article was commissioned by the editorial office, *Translational Cancer Research* for the series "Targeted Therapy and Non-Small Cell Lung Cancer: A New Era?". The article has undergone external peer review.

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/tcr.2018.11.35). The series "Targeted Therapy and Non-Small Cell Lung Cancer: A New Era?" was commissioned by the editorial office without any funding or sponsorship. UM and CR served as the unpaid Guest Editors of the series. The authors

have no other conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee(s) and with the Helsinki Declaration (as revised in 2013). The trial was approved by the Antwerp University Hospital ethical committee n. 14/17/206. Informed consent was obtained from the patients.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: https://creativecommons.org/licenses/by-nc-nd/4.0/.

References

- Soda M, Choi YL, Enomoto M, et al. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. Nature 2007;448:561-6.
- Solomon BJ, Mok T, Kim DW, et al. First-line crizotinib versus chemotherapy in ALK-positive lung cancer. N Engl J Med 2014;371:2167-77.
- Nilsson RJ, Karachaliou N, Berenguer J, et al. Rearranged EML4-ALK fusion transcripts sequester in circulating blood platelets and enable blood-based crizotinib response monitoring in non-small-cell lung cancer. Oncotarget 2016;7:1066-75.
- Dagogo-Jack I, Brannon AR, Ferris LA, et al. Tracking the Evolution of Resistance to ALK Tyrosine Kinase Inhibitors Through Longitudinal Analysis of Circulating Tumor DNA. JCO Precis Oncol 2018;2018. doi: 10.1200/PO.17.00160.
- Rolfo C, Castiglia M, Hong D, et al. Liquid biopsies in lung cancer: The new ambrosia of researchers. Biochim Biophys Acta 2014;1846:539-46.

Cite this article as: Reclusa P, Laes JF, Malapelle U, Valentino A, Rocco D, Gil-Bazo I, Rolfo C. *EML4-ALK* translocation identification in RNA exosomal cargo (*ExoALK*) in NSCLC patients: a novel role for liquid biopsy. Transl Cancer Res 2019;8(Suppl 1):S76-S78. doi: 10.21037/tcr.2018.11.35