Evaluating the Role of Solubility in Oral Absorption of Poorly Water-Soluble Drugs Using Physiologically-Based Pharmacokinetic Modeling

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Poor aqueous solubility and dissolution of drug candidates drive key decisions on lead series optimization during drug discovery, on formulation optimization, and clinical studies planning during drug development. The interpretation of the *in vivo* relevance of early pharmaceutical profiling is often confounded by the multiple factors affecting oral systemic exposure. There is growing evidence that *in vitro* drug solubility may underestimate the true *in vivo* solubility and lead to drug misclassification. Based on 10 poorly water-soluble tyrosine kinase inhibitors, this paper demonstrates the use of physiologically-based pharmacokinetic (PK) analysis in combination with early clinical PK data to identify drugs whose absorption is truly limited by solubility *in vivo* and, therefore, expected to exhibit food effect. Our study supports a totality of evidence approach using early clinical data to guide decisions on conducting drug interaction studies with food and acid-reducing agents.

Study Highlights

WHAT IS THE CURRENT KNOWLEDGE ON THE TOPIC?

✓ The solubility of a drug candidate significantly impacts decisions in drug development. The Biopharmaceutics Classification System classification based solely on *in vitro* solubility can be conservative with respect to impact of solubility on a compound's absorption.

WHAT QUESTION DID THIS STUDY ADDRESS?

Deconvolution of the mechanisms underlying the gut bioavailability is hindered by parameter nonidentifiability. This research uses a combined *in vitro*, *in vivo*, and *in silico* analysis to understand the *in vivo* relevance of *in vitro*-measured solubility

In drug discovery, solubility of drug candidates in aqueous media is one of the pivotal physicochemical properties to optimize a chemical series. A candidate drug is commonly required to have solubility above 10 μ M to facilitate preclinical testing.¹ Low, highly variable oral bioavailability and less-than-dose-proportional exposure are among the consequences of poor drug solubility (**Table S1**). Therefore, pharmaceutical companies strive to increase the solubility of a drug candidate during lead optimization.^{1,2} An analysis of the early clinical data to understand the *in vivo* relevance for a better prediction of food and proton pump inhibitor effects on oral drug exposure.

WHAT DOES THIS STUDY ADD TO OUR KNOW-LEDGE?

A more precise classification based on a sound mechanistic understanding and totality of evidence is proposed for efficient drug development.

HOW MIGHT THIS CHANGE CLINICAL PHARMA-COLOGY OR TRANSLATIONAL SCIENCE?

☑ Identification of compounds that have sufficient *in vivo* solubility despite low *in vitro* solubility can support decisions on the need and timing of studies and help save valuable resources.

of *in vitro* solubility could be valuable in assessing the need for resource-intensive formulation development and/or clinical trials.

Based on the Biopharmaceutics Classification System (BCS),³ the solubility class threshold is determined using the highest strength that can be completely dissolved in 250 mL of an aqueous medium (pH 1–6.8 at 37°C). More recently, the Developability Classification System (DCS),⁴ recognized the need to differentiate between dissolution rate (IIa) and solubility (IIb) limitation. However, a precise classification of drug candidates may not be

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feasible at the end of lead optimization because the active dose range at this phase can only be estimated.⁵

Evidence suggests that oral bioavailability may not always depend on the solubility of poorly soluble drugs. First, several BCS II/IV drugs, such as naproxen, phenytoin, and diazepam, have an absolute bioavailability (F) > 90%.^{6,7} Second, although poorly water-soluble, lipophilic compounds are generally expected to show a better solubilization in gastrointestinal fluids in the presence of food and, thus, a better oral absorption in the fed state,⁹ poorly water-soluble anticancer drugs, such as imatinib and trametinib, show only a modest food effect, if any.¹⁰ Last, enabling formulations meant to improve the kinetic solubility of poorly water-soluble active ingredients do not always enhance oral bioavailability,¹¹ indicating that poor oral bioavailability may be caused by other factors.^{12,13}

Physiologically-based pharmacokinetic (PBPK) models are powerful tools that describe drug pharmacokinetic (PK) through the integration of PK mechanisms, compound data, and physiology.¹⁴ The objective of the current study is to conduct PBPK analysis of 10 poorly soluble anticancer drugs, as described by Peters,¹⁵ along with an analysis of gut bioavailability (fraction of the administered oral dose reaching the portal vein) and dose-exposure proportionality, to evaluate the *in vivo* relevance of *in vitro* solubility data in determining their oral absorption. Such an evaluation could the pave the way for better predicting the impact of food and acid-reducing agents on the exposure of poorly water-soluble drugs.

RESULTS

Biopharmaceutical properties

Table 1 shows the fasted state simulated intestinal fluid (FaSSIF) and simulated gastric fluid (SGF) solubility of the model drugs, permeability, and reported biopharmaceutical properties. The FaSSIF solubility of these compounds is generally < 1 mg/mL, except for imatinib. The measured solubility is consistent with literature, except for pazopanib whose measured solubility in SGF at pH 1.6 is higher than the value reported at pH 1.1.¹⁶

PBPK absorption models

PBPK modeling for all model drugs is exemplified by pazopanib (**Figure 1**). The PBPK simulation of oral PK profiles using clearance and distribution parameters derived from i.v. simulation, coupled with FaSSIF solubility and Caco-2 permeability, may not capture the observed exposure (**Figure 1b**). To capture the observed exposure, a 20-fold increase in the input solubility (**Figure 1c**) and reduced colonic absorption were needed (**Figure 1d**). PBPK simulation using a hypothetically high solubility results in maximum plasma concentration (C_{max}) and area under the plasma concentration-time curve (AUC) that are fourfold higher than the observed (**Table 2** and **Figure 1e**). This indicates that pazopanib exposure is limited by solubility. Poor sensitivity to a hypothetically high permeability shows that its exposure is not limited by permeability (**Figure 1f**).

The intrinsic microsomal clearance (CL_{int}) and a multiplicative factor (K_p) to scale the tissue distribution coefficients, derived from i.v. PBPK simulations are reported in **Table S2**. In the absence of

i.v. PK for vemurafenib and lapatinib, the volume of distribution for these two compounds were estimated (**Table S3**). Oral PBPK simulations of all model drugs under fasting conditions at doses used in their clinical food effect studies show that the simulated PK metrics are adequately captured (**Table 2**, **Figure S1**) except for erlotinib and M1, a development compound, whose exposures were overpredicted due to an intestinal loss of drug. Gastric emptying rate was decreased to capture the observed profiles of crizotinib, gefitinib, and imatinib. Colonic absorption of vemurafenib and pazopanib had to be decreased. Similar to the pazopanib case, the FaSSIF solubilities of dabrafenib, lapatinib, trametinib, and vemurafenib were increased to accurately predict their plasma exposure. The elimination slope of the predicted dabrafenib oral PK profile derived from the i.v. PK is steeper than the observed, suggesting prolonged absorption.¹⁷

Measured absolute bioavailability and calculated gut bioavailability

The gut bioavailability of vemurafenib and lapatinib cannot be determined in the absence of i.v. PK. All other compounds, including M1, have a relatively high gut bioavailability (> 0.75) at the dose tested, except for pazopanib and erlotinib (**Table S4**). Gut bioavailability > 1 can result from the use of mean values.

Effect of solubility and permeability on PK metrics based on PBPK model simulations

AUC and C_{max} ratios (hypothetical BCS class I-like solubility to best-fit solubility) from PBPK simulations are summarized in **Table 2**. Crizotinib, dabrafenib, gefitinib, imatinib, M1, and trametinib are insensitive to an increase in the input solubility. In contrast, the oral exposures of pazopanib and vemurafenib were sensitive to solubility with either measures of permeability (Caco-2 or calculated effective intestinal permeability (P_{eff})). For erlotinib, C_{max} was increased using the hypothetical BCS class I-like solubility but not the AUC, indicating that its rate of absorption depends on dissolution.

The use of Caco-2 and calculated $P_{\rm eff}$ led to similar results for all drugs except for the poorly permeable compounds vemurafenib and lapatinib, for which calculated $P_{\rm eff}$ in PBPK models led to a significant increase in exposure.

Clinical PK analysis on dose-exposure proportionality and food effect

Based on the clinical PK data presented in **Table 3**, systemic exposure increases in a dose-proportional or supra-dose-proportional manner for all compounds except for pazo-panib > 800 mg^{18,19} and M1. Even though the single-dose PK of crizotinib was reported to be less-than-dose-proportional,²⁰ the proportionality based on the data presented seems linear in our assessment.

A strong positive food effect on systemic exposure was only observed for lapatinib, vemurafenib, and pazopanib (**Table 3**). High intersubject variability prevents the detection of significant food effect on gefitinib exposure.²¹ The exposure of 150 mg erlotinib²² doubled under fed conditions in the first period but slightly decreased in the second period compared with

Parameters	Imatinib mesylate	M1	Crizotinib	Trametinib	Dabrafenib mesylate	Gefitinib	Erlotinib HCI	Lapatinib ditosylate	Vemurafenib	Pazopanib HCI
Measured solubility in FaSSIF pH 6.5 (mg/mL)	2 IA	0.0728	0.7430	N.A.	0.0037	0.0887	0.0124	0.0350	0.0054	0.0012
Measured solubility in SGF pH 1.6 (mg/mL)	N.A.	0.0213	N.A.	N.A.	0.0117	N.A.	0.1665	0.0055	0.0003	1.0779
Measured Caco-2 P _{app} passive (10 ⁻⁶ cm/ second)	25.90	14.70	15.35	15.44	12.45	10.41	40.43	0.21	0.11	28.46
cP _{eff} [10 ⁻⁴ cm/second]	2.61	6.19	2.22	1.07	2.38	5.23	3.58	1.84	2.54	0.85
pK _a values (basic)	8.07, 1.52 ³⁵	9.5, 2.8	9.4, 5.6 ²⁰	N.A.	2.2, 1.5 ³⁶	7.2, 5.4 ³⁷	5.42 ³⁸	4.6, 6.7 ³⁹	N.A.	6.4, 2.1 ¹⁸
pK _a values (acidic)	N.A.	N.A.	N.A.	N.A.	6.6 ³⁶	N.A.	N.A.	N.A.	7.9, 11.1 ⁴⁰	10.2 ¹⁸
Clinical dose in food effect study (mg) ^a	400	Clinical dose is yet to be established	250	0	150	250	150	1,500	096	800
BCS classification	11 ¹⁰	Likely IV	IV ²⁰	II ²⁴	11 ³⁶	11 ⁴¹	11 ³⁸	IV ⁴²	IV ⁴⁰	11 ¹⁸
Dose number	0.38	< 1	1.35	11.43	193.55	11.27	53.10	277.78	711.11	2,909.09
SLAD (mg) ^a	6,527.96	> dose in food effect study	985.72	0.45 ^b	4.41	227.43	24.18	23.75	8.20	0.55
DCS classification	lla	Likely IV	2	qII	qII	lla	qII	2	2	qII
Major metabolizing CYP enzymes	СҮРЗА4 ⁴³	CYP3A4 CYP2C8	СҮРЗА4 СҮРЗА5 ²⁰		СҮР2С8 СҮРЗА4 ³⁶	CYP3A4 CYP2D6 ⁴⁴	СҮРЗА4 ³⁸	СҮРЗА4 СҮРЗА5 ⁴²	CYP3A4 ⁴⁰	СҮРЗА4 ¹⁸
Dose number: dose number < BCS, Biopharmaceutical Class Classification System; FaSSIF 500 mL FaSSIF, solubility crite ^a Dose given as free base. ^b Ca	1 indicates that t iffication System; fasted state inte rion for DCS IIa/II, rounted using lite	the dose is soluble in Caco-2 P _{app} passive istinal fluid; N.A., no' Ib classification (Eq. rature solubility from	1 250 mL FaSSIF, , apparent passivvi t applicable; pK _a , i 3)). 1 the US Food and	solubility criterion e permeability dete acid dissociation c Drug Administrati	for BCS classifica armined in Caco-2 constant; SGF, sim on Clinical Pharma	tion (Eq. 2). assay; cP _{eff} , cald ulated gastric flu toology and Biop	ulated effective iid; SLAD, solubil	ntestinal permeat ty limited absorba /iew. ²⁴	ility; DCS, Developa ble dose (amount of	bility drug soluble in

Table 1 Summary of biopharmaceutical properties and metabolizing CYP enzymes



Figure 1 Physiologically-based pharmacokinetic (PBPK) simulations of pharmacokinetic profiles of pazopanib. (a) PBPK simulation of i.v. infusion of 5 mg pazopanib over 5 minutes to obtain the intrinsic clearance and multiplicative factor, for simultaneously scaling all tissue distribution coefficients. (b) Pazopanib 800 mg oral administration simulated with *in vitro* fasted state simulated intestinal fluid (FaSSIF) solubility and Caco-2 permeability. (c) Pazopanib 800 mg oral administration simulated with Caco-2 permeability and an input solubility that is 20-fold higher than FaSSIF solubility. (d) Pazopanib 800 mg oral administration simulated with Caco-2 permeability, an input solubility that is 20-fold higher than FaSSIF solubility and reduced colonic absorption. (e) Pharmacokinetic profile using hypothetical Biopharmaceutics Classification System class I-like solubility in the pazopanib PBPK model with good fit in (d). (f) Pharmacokinetic profile using hypothetically high permeability of 10*10⁻⁴ cm/second in the pazopanib PBPK model with good fit in d. [Colour figure can be viewed at wileyonlinelibrary.com]

fasted conditions. This anomaly is not further explained by the investigators.

studies. Therefore, oral exposure of these drugs is not expected to be solubility-limited.

DISCUSSION

In vitro solubility measurement within the biopharmaceutical framework (BCS and DCS)

Although the primary regulatory purpose of the BCS is guiding *in vivo* bioequivalence study waivers, its scientific rationale is used in other areas, including food effect predictions.⁹ As per the BCS specifications, all 10 model anticancer drugs have been classified as poorly soluble based on the drug substance.¹⁰ Of note, the dose number for imatinib is less than unity in our calculation (**Table 1**), which would qualify it as a BCS I drug. Under the DCS framework, imatinib, M1, crizotinib, and gefitinib have a solubility-limited absorbable dose (SLAD) higher than the clinical dose used in their food effect

Gut bioavailability

Gut bioavailability ≥ 0.75 indicates that systemic exposure of the most model compounds may not depend on the drug's solubility at the tested dose levels. Only erlotinib (0.61) and pazopanib (0.21) have a relatively low gut bioavailability pointing to an intestinal drug loss attributable to efflux, gut metabolism, and/or solubility limitation.

In silico **PBPK** modeling probing the role of solubility in oral drug absorption

Hypothesis testing with PBPK modeling¹⁵ has been used to evaluate the role of *in vitro* solubility in oral absorption of

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	Observed	PK param	eters	PBPK sim	ulated PK per	parame meabilit	ters using Caco-2 y	РВРК	simulated	PK using	calculated P _{eff}	Caco- permeat	2 Dility	Calculate	d P _{eff}
	A U C ₀₋₂₄ (hour*µM)	С _{max} (µM)	T _{max} (hour)	AUC ₀₋₂₄ (hour*µM)	с _{max} (µM)	T _{max} (hour)	Solubility increase (mg/mL)	AUC ₀₋₂₄ (hour*µM)	C _{max} (µM)	T _{max} (hour)	Solubility increase (mg/mL)	AUC ₀₋₂₄ ratio	c _{max} ratio	AUC ₀₋₂₄ ratio	c _{max} ratio
Imatinib	28.91	3.51	1.51	36.4	2.78	1.6	None	36	2.65	2.24	None	1.0	1.0	1.0	1.0
M1	1.073	0.0621	œ	1.75	0.312	0.96	None	1.76	0.352	0.8	None	1.0	1.0	1.0	1.0
Crizotinib	3.635	0.311	5.05	3.36	0.261	3.6	None	3.34	0.257	4	None	1.0	1.0	1.0	1.0
Trametinib	0.1162	0.0136	1.5	0.0912	0.0052	2.16	0.0008 → 0.0069	0.0896	0.0054	1.76	0.0008 → 0.0416	1.1	1.7	1.0	1.0
Dabrafenib	25.38	3.99	2	18.6	3.89	2.48	0.0037 → 0.0616	18.4	3.52	2.72	$0.0037 \rightarrow 0.0616$	1.0	1.3	1.0	1.3
Gefitinib	2.549	0.161	3.72	2.24	0.142	3.82	None	2.27	0.145	3.42	None	1.0	1.0	1.0	1.0
Erlotinib	21.3	1.94	4.13	53.9	4.03	3.67	None	35.6	1.98	4.71	None	1.1	2.1	1.6	3.4
Lapatinib	18.07	1.46	3.96	15.3	1.18	2.88	0.0350 → 6.6044	22.7	1.4	3.92	0.0350 → 0.0943	1.0	1.0	4.6	11.0
Vemurafenib	94.71	6.41	4.08	109	5.91	4.7	$0.0054 \rightarrow 0.6124$	133	7.3	6.14	None	2.6	2.6	54.1	53.8
Pazopanib	843.6	47.3	4	889	50.7	5.44	$0.0012 \rightarrow 0.0237$	978	55.7	5.12	$0.0012 \rightarrow 0.2370$	3.9	4.0	2.7	2.7
AUC ₀₋₂₄ , area ur P effective int	nder the plasm estinal permea	a concentrat bility: PK. pf	tion time c Jarmacoki	urve from 0 to inetic: T . tim	24 hours; B e to reach t	CS, Biop he maxin	narmaceutical Classifica	tion System; on.	C _{max} , maxim	um plasn	ia concentration; PBPK, p	hysiologica	Ily-based	pharmaco	kinetic;

5 max' -2 Ś. 5 eff'

	Dose range tested (mg)	Dosing schedule	Increase in exposure	Food effect study dose (mg)	AUC fed/fasted ratio	C _{max} fed/ fasted ratio
Imatinib	25-1,000	N.A.	Dose proportional ⁴³	400	0.9245	0.89 ⁴⁵
M1	30-1,400	Steady-state	Less than proportional (food effect dose is within the linear region)	30	1.17	1.29
Crizotinib	50–300 50–200 200–300	Single dose Steady state Steady state	Less than proportional More than proportional More than proportional ²⁰	250	0.86 ⁴⁶	0.86 ⁴⁶
Trametinib	0.125–10	Single Dose	More than proportional (C _{max} proportional) ²⁴	2	0.897 ⁴⁷	0.30147
Dabrafenib	12–300	Single dose Steady state	Dose proportional Less than proportional ³⁶	150	0.70 ²³	0.49 ²³
Gefitinib	50–500 50–400 50–700	Single dose (HV) Steady state (pat.) Steady state (pat.)	Dose proportional Dose proportional More than proportional ⁴¹	250	1.37 ²¹	1.32 ²¹
Erlotinib	100-1,000	N.A.	Dose proportional ³⁸	150	1.97/0.93 ²²	1.57/1.15 ²²
Lapatinib	Approx. 600–1,800	Steady state	Dose proportional ⁴²	1,500	4.25 ⁴⁸	3.03 ⁴⁸
Vemurafenib	240-960	Single dose Steady state	Dose proportional Dose proportional ⁴⁰	960	4.7 ⁴⁹	2.5 ⁴⁹
Pazopanib	50–2,000	Single dose Steady state	Less than proportional Less than proportional ¹⁸	800	2.34 ⁵⁰	2.08 ⁵⁰

Table 3 Evaluation of dose-exposure proportionality for model compounds from the FDA's Clinical Pharmacology and Biopharmaceutics Reviews and food effect

AUC, area under the plasma concentration time curve; C_{max}, maximum plasma concentration; FDA, US Food and Drug Administration; HV, healthy volunteers; N.A., not applicable; pat., patients.

poorly water-soluble drugs. During model development, a 2 to 200-fold increase over the measured *in vitro* solubility was necessary to simulate the observed exposure of 5 extremely poorly water-soluble (solubility < 0.01 mg/mL) drugs (**Table 2**). The *in vitro* solubility in a defined solvent could underestimate the true *in vivo* solubility, which may be influenced by supersaturation (especially for weak bases), and formulation effects. For example, hydroxypropyl methylcellulose presented in the shell of dabrafenib capsules shows an inhibitory effect on precipitation of supersaturated dabrafenib solution,²³ or the presence of sodium dodecyl sulfate in trametinib²⁴ tablets may increase the apparent drug solubility.

The PBPK simulations of vemurafenib, lapatinib, and pazopanib show a significantly higher-than-observed exposure when a hypothetically high BCS class I-like solubility is used as input (**Table 2**). This suggests that the absorption of these compounds is limited by solubility. BCS class IV drugs, vemurafenib and lapatinib, exhibit significantly higher-than-observed exposure with a hypothetically high solubility as well as permeability, indicating a solubility-limited and/or permeability-limited exposure. On the contrary, the C_{max} and AUC ratios with hypothetically high solubility are close to one for imatinib, trametinib, crizotinib, dabrafenib, gefitinib, and M1 (**Table 2**), indicating that the exposures of these drugs are not solubility-limited.

The need to reduce the gastric emptying rate for crizotinib, imatinib, and gefitinib for better fit to the observed profile is possibly due to drug-induced delayed gastric emptying. The use of the generic PBPK model to identify delayed gastric emptying was already validated in the rat. $^{25}\,$

PBPK simulations of erlotinib show an intestinal drug loss that is likely due to gut metabolism or efflux. Erlotinib, a CYP3A substrate, is known to be metabolized in the gut.²⁶ With only 1% of the dose as parent drug in the feces after oral administration,²⁶ the possibility of transporter-mediated intestinal loss may be ruled out. The *in vitro* FaSSIF solubility was sufficient to explain the observed erlotinib PK profile but using a hypothetical BCS class I-like solubility enables a higher C_{max} , whereas AUC remains unchanged (**Figure S3c**), indicating a slow *in vivo* dissolution but nonsolubility limited absorption. Other model drugs are substrates of CYP3A4 as well (**Table 1**), but their exposures are probably not limited by gut metabolism, as evidenced by the good gut bioavailability (> 0.75) for most of these compounds and by the PBPK analysis.

Both the calculation of gut bioavailability as well PBPK analysis relies on i.v. PK data at doses leading to concentrations comparable to the oral administration. When such i.v. PK data are not available, PBPK analysis can still be carried out if elimination parameters can be estimated from well-characterized, single-dose oral PK profiles with < 20% AUC extrapolation.

Analysis of dose-exposure relationship

A less-than-dose-proportional exposure observed in dose-escalating PK studies can help identify solubility-limited drug absorption. However, identifying dose-exposure trends is confounded by high intersubject variability in clinical studies, insufficient number of subjects per dose group, or insufficient number of doses in the dose range of interest (**Figure S2**).

Identification of solubility-limited exposure and food effect predictions

Table 4 distinguishes drugs with solubility-limited absorption (shown in red) from those that are not (shown in green). On the right, the food effect for these drugs are also color-coded to distinguish drugs that show a positive food effect (red) from those that do not (green).

Although the BCS classification identifies all the 10 study compounds to have solubility-limited absorption, the DCS is less conservative and able to predict lack of food effect in crizotinib, gefitinib, M1, and imatinib. However, this a very small number to draw any comparative conclusions.

When phase I clinical data are available, calculation of gut bioavailability and PBPK analysis become possible. Apart from erlotinib, gut bioavailability and PBPK analysis show similar trends of solubility-limited absorption for the model drugs. Although a gut bioavailability < 1 can identify an intestinal drug loss, it cannot distinguish between mechanisms contributing to intestinal loss (e.g., solubility-limitation, transporter-mediated efflux, and gut metabolism). Hypothesis testing with PBPK analysis can distinguish among intestinal loss mechanisms, gut metabolism, and poor aqueous solubility, as exemplified by erlotinib (Figure S3). Another advantage of the PBPK approach over gut bioavailability method is that it can be applied even when i.v. PK data are not available, provided the oral PK profile is sufficiently well characterized. When PK data from the entire dose range of interest becomes available, it is possible to distinguish solubility limitation (less-thandose-proportional exposure) from gut metabolism and efflux (supra dose-proportional exposure), and to confirm the PBPK analysis. The drugs identified by PBPK to have solubility-limited absorption are also those that show a positive food effect. The extent of food effect correlates with the AUC and C_{\max} ratios (hypothetical BCS class I-like solubility to best fit) from PBPK simulations. It should be noted, however, that the "in vivo solubility" derived from the PBPK modeling represents a minimum solubility. True in vivo solubility at a given dose may be even higher.

As pointed out earlier, establishing a dose-exposure relationship is often challenged by high variability and insufficient dose groups, commonly encountered in oncology drug development. This is true for BCS IV vemurafenib and lapatinib, where the dose linearity seems to contradict the outcome from PBPK analysis. For both compounds, the dose range in the dose-escalation studies does not cover that tested in food effect studies well, especially in the upper end. In the case of M1, less-than-dose-proportional exposure indicates solubility-limitation at doses much higher than the food effect study, which is why no food effect was expected.

Our study demonstrates that if the oral absorption of a drug candidate can be accurately identified as solubility-limited, it is straightforward to predict the solubilization-driven food effect.

Awareness of the impact and limitation of measured solubility on predicting the oral drug absorption of a drug candidate in the early development phase is critical to saving valuable resources in drug development. Not all poorly water-soluble compounds defined under the biopharmaceutical framework have solubility-limited oral absorption. Our analysis demonstrates that deconvolution of the key mechanisms driving intestinal loss with PBPK analysis reliably identifies those poorly water-soluble drugs whose oral absorption is truly solubility-limited and are, therefore, likely to show a positive food effect. Deconvolution of mechanisms contributing to intestinal loss can further complement existing deconvolution methods for an improved *in vitro-in vivo* correlation.

Our study supports a totality of evidence approach using early clinical data to guide decisions on conducting drug interaction studies with food and acid-reducing agents (**Figure 2**). Further validation with a larger dataset could enhance confidence in the approach.

MATERIALS AND METHODS Materials

Dabrafenib mesylate and vemurafenib were purchased from Chem Shuttle (Hayward, CA), lapatinib ditosylate monohydrate and pazopanib hydrochloride (HCl) from Ark Pharm (Arlington Heights and Libertyville), erlotinib HCl from Activate Scientific (Prien-Chiemsee, Germany), imatinib mesylate from abcr GmbH (Karlsruhe, Germany), and trametinib *DMSO solvate from Asta Tech (Bristol, PA). Crizotinib and gefitinib were obtained from internal batches (Merck KGaA, Darmstadt, Germany). Simulated intestinal fluid powder (version 1) was purchased from Biorelevant.com (London, UK), and the water was purified by a Milli-Q water purification system (Merck KGaA). All other chemicals, solvents at the liquid chromatography grades, and high-performance liquid chromatography (HPLC) columns were obtained from Merck KGaA.

Model drug selection

Based on the clinical summary of oral anticancer drugs reported by Willemsen *et al.*, ¹⁰ crizotinib, dabrafenib mesylate, erlotinib HCl, gefitinib, imatinib mesylate, pazopanib HCl, and trametinib were selected as model drugs due to their poor aqueous solubility (BCS II or IV) based on the US Food and Drug Administration's (FDA's) reviews, availability of i.v. PK data in humans, and the absence of special handling requirements for safety reasons. M1, a development compound, which meets the above selection criteria, was also included in the analysis. Although the i.v. PK data are not available for vemurafenib and lapatinib ditosylate, they were included in the analysis due to the strong food effect observed in the clinical studies. The analysis was performed at clinically relevant doses.

Human in vivo PK data

The clinical PK data used in the current study are summarized in **Table S4**. The i.v. PK and absolute bioavailability studies were used for the calculation of the gut bioavailability. PBPK simulations were performed at doses used in food effect studies using the observed profiles in the fasted state. The dose-exposure proportionality information was obtained from single ascending dose studies.

In the absence of appropriate human *in vivo* PK data for erlotinib HCl and imatinib mesylate, closest substitutes were used. For erlotinib, the plasma clearance in healthy volunteers is not given.²⁷ Instead, the renal clearance in patients after 100 mg i.v.²⁸ was used to estimate the worst-case (i.e., lowest) gut bioavailability. For imatinib, because the plasma-concentration time profiles of the food studies are not available, the oral PK profiles in the absolute bioavailability study were used.²⁹

solubility-limited ϵ	exposure					
Compound	Dose number (BCS)	SLAD (DCS)	Gut bioavailability	PBPK modeling	Dose proportionality in single-dose PK studies	Food effect
Imatinib	=	_	lla			
M1	Likely IV	Likely IV				
Crizotinib	N	2				
Gefitinib	=	lla				
Trametinib	=	qII				
Dabrafenib	=	qII				
Erlotinib	=	qII				
Lapatinib	2	2	i.v. PK data required for the calculation is not available	solubility and/or permeability		
Vemurafenib	2	N		solubility and/or permeability		
Pazopanib	=	qII				
Every property/method	is color-coded for the drugs sele	cted in this study to disti	nguish solubility-limited drugs (red) from t	those that are not (green). On	the far right, the food effects for the	se drugs are also

Table 4 Heatmap summarizing the properties in the BCS and DCS frameworks along with properties/methods used in this analysis for the identification of

limitation of vemurafenib and lapatinib absorption is nonidentifiable), less than dose proportional increase of AUC and C_{max}, 2-fold positive food effect. Yellow – Anomalies: Food effect (~ 30%²¹ increase in AUC physiologically-based pharmacokinetic; PK, pharmacokinetic; SLAD, solubility limited absorbable dose (amount of drug soluble in 500 mL fasted state simulated intestinal fluid, solubility criterion for DCS IIa/IIb PBPK model not sensitive to increase in solubility, proportional, or more than dose proportional increase of AUC and C_{max} in the food effect dose range, absence of positive food effect (AUC and C_{max} fed/fasted gut pioavailability > 0.7 5, AUC, area under the plasma concentration time curve; BCS, Biopharmaceutical Classification System; C_{max}, maximum plasma concentration; DCS, Developability classification system; i.v., intravenous; BPPK, ratio < 1). Red – solubility-limited: Dose number > 1 (BCS II/IV), SLAD < clinical dose, gut bioavailability < 0.75, better exposure in PBPK model using increased input solubility (solubility and/or permeability and C_{max}) of gefitinib is confounded by a high intersubject variability, the food effect of erlotinib is inconsistent between the two periods of the 150 mg single-dose food effect study²² (BUS I/III), SLAU number illry limita 2 Z nat do not. Green -Irom mose Iood ellect urugs that show a positive aisunguisn 2 coded red or green classification (Eq. 3)). color ы́ы́

Dose number, dose number < 1 indicates that the dose is soluble in 250 mL FaSSIF, solubility criterion for BCS classification (Eq. 2); gut bioavailability, the fraction of the administered dose reaching the portal vein, deconvoluted from the absolute bioavailability using the systemic clearance.



Figure 2 Schematic illustration of the proposed methods to assess solubility limited absorption behavior through a mechanistic analysis of human clinical pharmacokinetic data. When pharmacokinetic data from one i.v. and one oral dose are available, the gut bioavailability can be easily calculated to assess whether there is a loss of drug in the gut. Using a physiologically-based pharmacokinetic (PBPK) model, it can be analyzed, if this loss of drug is related to insufficient solubilization. When pharmacokinetic data from ascending oral doses or food effect study become available, the outcomes of the PBPK modeling approach can be confirmed by the manner exposure behaves after increasing the dose or food intake.

Thermodynamic solubility measurement

The thermodynamic solubility of the model compounds in their marketed salt forms (in case of vemurafenib amorphous co-precipitate from ground market product), was measured in FaSSIF pH 6.5 (containing 3 mM sodium taurocholate and 0.75 mM phospholipids) and SGF pH 1.6. An excess amount of drug substance was weighed into an Erlenmeyer flask with 10 mL of the media. The flask was incubated in a shaking water bath (250 movements per minute) at 37°C for 24 hours. Samples were taken after 24 hours and centrifuged at 20,817 g and 37°C for 5 minutes. The supernatant was diluted with an organic solvent and analyzed by HPLC. If possible, a largely universally applicable HPLC method using a gradient of eluent A (water + 0.1% formic acid) and eluent B (acetonitrile + 0.1% formic acid) and Chromolith Performance RP18e 100 × 3 mm column with a low flow of 0.85 mL/minute was used. Otherwise, an alternative method was applied using a Chromolith High Resolution RP18e 100×4.6 mm column, a flow rate of 3 mL/minute, and a gradient of eluent A (1,900 mL water + 100 mL acetonitrile + 2 mL trifluoroacetic acid) and eluent B (1,900 mL acetonitrile + 100 mL water + 2 mL trifluoroacetic acid).

Effective human intestinal permeability from apparent permeability in Caco-2 cells

Permeability across a TC7 Caco-2 monolayer on a microporous polycarbonate membrane filter was measured bidirectionally (i.e., apical (A) \rightarrow basolateral (B) and B \rightarrow A) in a 24-well plate and up to 5 compounds per well. The apparent passive permeability (geometric mean of $P_{app} A \rightarrow B$ and B \rightarrow A) was measured in the presence of cyclosporin A that inhibits P-glycoprotein. Hank's balanced salt solution (pH 7.4) was used as the reservoir for apical and basolateral matrices. The compounds were added as dimethyl sulfoxide stock solutions resulting in a final drug concentration of 1 µM. The apical and basolateral drug concentrations after the reaction were quantified via liquid-chromatography mass spectrometry. A factor of 25 was used to scale the measured Caco-2 permeability to the effective human intestinal permeability.

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Calculated effective human intestinal permeability

The effective human intestinal permeability was calculated, using log P, the polar surface area and the number of hydrogen bond donors, according to Eq. $1.^{30}$ The input values used for the calculation are shown in **Table S5**.

$$\log P_{\rm eff} = -3.061 + 0.190 \text{CLOGP} - 0.010 \text{PSA} - 0.246 \text{HBD}$$
(1)

BCS/DCS classification

The BCS classification presented in literature¹⁰ is summarized in **Table 1**. Additionally, the dose number (D_o) was calculated to confirm whether the highest clinical dose (M_0) can be completely dissolved in 250 mL intestinal fluid according to Amidon *et al.*³ with Eq. 2 using the thermodynamic solubility in FaSSIF (C) and volume (V_0) of 250 mL.

$$D_0 = \frac{M_0 / V_0}{C_s}$$
(2)

The compounds were further classified based on the DCS classification by Butler *et al.*,⁴ calculating the SLAD using Eq. 3 with the thermodynamic solubility in FaSSIF and volume (V) of 500 mL, where M_p is equal to A_n (Eq. 4)³ for high permeability compounds (calculated $P_{eff} > 1 \times 10^{-4}$ cm/s) and equal to 1 for others. A_n was calculated from Eq. 4 using a tube radius R of 1 cm³ and residence time (t_{res}) of 3.32 hours.⁴

$$SLAD = S_{si} * V * M_p \tag{3}$$

$$A_n = \frac{P_{\text{eff}}}{R} * t_{\text{res}} \tag{4}$$

PBPK modeling

A generic whole-body PBPK model built in MATLAB software (version R2017a; The MathWorks, Natick, MA) has been used for a line-shape analysis of the observed i.v. and oral PK profiles at doses used in their

clinical food effect studies, as shown in **Figure 3**. The *in vitro* microsomal clearance generally tends to underpredict the human clearance.³¹ The clearance along with the distribution parameters were estimated by optimizing CL_{int} and K_p factors to best fit the i.v. PK profiles. These parameters are uncorrelated and have unique influences on the line shape. Hence, confidence in the estimated parameters is high.^{13,25} This principle of obtaining CL_{int} and K_p is similar to estimating CL_{int} using retrograde calculation and K_p factor in Simcyp (https://www.certara.com/) in a top-down analysis.

Because the i.v. PK data of vemurafenib and lapatinib ditosylate were not available, the clearance was estimated from the elimination rate constant (k_e) (Eq. 5) based on the slope of the elimination phase in the oral PK profiles (i.e., the linear part of the log PK profiles at time points well beyond the absorption phase). This is based on the assumption that the volume of distribution per kilogram body weight, when corrected for plasma protein binding, is the same across different species,³² as shown in **Table S3**.

$$k_e = \frac{Cl}{V_{ss,\mu}} \tag{5}$$

Clearance and volume of distribution along with FaSSIF solubility and effective permeability were used to simulate oral PK profiles. The input parameters are shown in **Table S2**. Effective permeability was derived from apparent permeability in Caco-2 cells or calculated from structural properties. Under the assumption of linear PK, the difference between the observed and simulated AUC can generate an understanding of the mechanisms underlying exposure (**Figure 3**). A simulated profile that cannot match the much steeper upswing of the observed profile and is characterized by a poor sensitivity to permeability, identifies an *in vivo* solubility that is greater than the measured solubility used as input.

The solubility value that best captures the observed profile represents the least *in vivo* solubility. The actual *in vivo* solubility could be higher. For BCS class IV drugs and those with borderline permeability, a higher permeability could have also achieved a similar good fit to the observed exposure.

PBPK simulations repeated with a hypothetically high solubility (dose/250 mL, analog BCS class I criteria, but at least 1 mg/mL) can identify any solubility-limited drug exposure. If the AUC or C_{max} ratio of exposure simulated by PBPK model using hypothetical BCS class I-like solubility to exposure derived from good fit is 1 or close to 1, exposure is not solubility-limited. A significantly higher ratio identifies solubility-limited exposure.

For drugs that induce gastric emptying delay, it may be necessary to reduce the gastric emptying rate in order to match the observed profile (**Figure 3**). When the observed profile is characterized by a significantly lower AUC compared with the simulated profile, gut metabolism or efflux could be responsible for the loss of the compound in the intestine (**Figure 3**). These two intestinal loss mechanisms cannot be distinguished.

Estimation of gut bioavailability

Oral bioavailability is a product of the fraction absorbed into the enterocytes (F_a), the fraction escaping intestinal metabolism (F_g), and the fraction escaping hepatic metabolism (F_h) (Eq. 6).³⁴ The product of $F_a \times F_g$,



Figure 3 Schematic illustration of the physiologically-based pharmacokinetic (PBPK) modeling approach. Cl_{int} , intrinsic microsomal clearance; K_p factor, multiplicative factor, to scale the tissue distribution coefficients; logP, decadic logarithm of the partition coefficient; PK, pharmacokinetic; pKa, acid dissociation constant.

the gut bioavailability, represents the fraction of the administered dose reaching the portal vein.

When PK data from i.v. and single-dose oral studies are available, the gut bioavailability can be calculated according to Eq. 7, using a liver blood flow (Q) of 90 L/hour,³³ assuming that the systemic clearance (Cl) is driven only by hepatic metabolism.

$$F = F_a \times F_g \times F_b = F_a \times F_g \times \left(1 - \frac{Cl}{Q}\right) \tag{6}$$

Rearranging Eq. 6 leads to Eq. 7:

Gut bioavailability =
$$F_a \times F_g = \frac{F}{1 - (Cl/Q)}$$
 (7)

Analysis of dose-exposure PK linearity and food effect

The dose-exposure linearity of the model drugs based on dose-escalating studies obtained from the FDA's Clinical Pharmacology and Biopharmaceutics Reviews and the reported food effect are summarized in **Table 3**.

SUPPORTING INFORMATION

Supplementary information accompanies this paper on the *Clinical Pharmacology & Therapeutics* website (www.cpt-journal.com).

Figure S1. Simulated and observed intravenous and oral (Caco-2 permeability) profiles.

Figure S2. Dose proportionality of AUC and C_{max} .

Figure S3. Hypothesis testing with physiologically based pharmacokinetic (PBPK) modeling to identify that the loss of erlotinib in the gut is mediated by gut metabolism and not due to solubility-limitation.

Table S1. Potential issues arising from poor drug solubility and their consequences.

Table S2. Summary of input parameters for the physiologically based pharmacokinetic simulations.

Table S3. Preclinical data to estimate the human clearance and volume of distribution of lapatinib and vemurafenib.

Table S4. Clinical pharmacokinetic data analyzed to assess solubility-limited absorption.

 Table S5.
 Input parameters for calculation of the effective permeability of the 10 model compounds.

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CONFLICT OF INTEREST

C.F., K.W., M.S., H.B., H.D., and S.-A.P. are employees of Merck Healthcare KGaA, Darmstadt, Germany. D.S. is employed by Merck KGaA.

AUTHOR CONTRIBUTIONS

C.F., D.S., S.A.P., and H.D. wrote the manuscript. S.-A.P. designed the research. C.F. performed the research. All authors analyzed the data.

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