Review

The association of microbial infection and adaptive immune cell activation in Alzheimer's disease

Mathew Clement*

Division of Infection and Immunity, Systems Immunity University Research Institute, Cardiff University, Cardiff, UK *Correspondence: Mathew Clement, Division of Infection and Immunity, Systems Immunity University Research Institute, Cardiff University, Cardiff, UK. Email: clementm@cardiff.ac.uk

Summary

Alzheimer's disease (AD) is a progressive neurodegenerative disorder and the most common form of dementia. Early symptoms include the loss of memory and mild cognitive ability; however, as the disease progresses, these symptoms can present with increased severity manifesting as mood and behaviour changes, disorientation, and a loss of motor/body control. AD is one of the leading causes of death in the UK, and with an ever-increasing ageing society, patient numbers are predicted to rise posing a significant global health emergency. AD is a complex neuro-physiological disorder where pathology is characterized by the deposition and aggregation of misfolded amyloid-beta ($A\beta$)-protein that in-turn promotes excessive tau-protein production which together drives neuronal cell dysfunction, neuroinflammation, and neurodegeneration. It is widely accepted that AD is driven by a combination of both genetic and immunological processes with recent data suggesting that adaptive immune cell activity within the parenchyma occurs throughout disease. The mechanisms behind these observations remain unclear but suggest that manipulating the adaptive immune response during AD may be an effective therapeutic strategy. Using immunotherapy for AD treatment is not a new concept as the only two approved treatments for AD use antibody-based approaches to target $A\beta$. However, these have been shown to only temporarily ease symptoms or slow progression highlighting the urgent need for newer treatments. This review discusses the role of the adaptive immune system during AD, how microbial infections may be contributing to inflammatory immune activity and suggests how adaptive immune processes can pose as therapeutic targets for this devastating disease.

Keywords: Alzheimer's disease, immune system, neuroinflammation, immunotherapy, viruses

Abbreviations: ACE2: Angiotensin-Converting Enzyme 2; AD: Alzheimer's disease; ADCC: Antibody Dependent Cellular Cytotoxicity; ADRD: Alzheimer's Disease and Related Dementia; ALT: Alanine Aminotransferase; Apoe: Apolipoprotein E; APP: Amyloid Precursor Protein; ART: Anti-Retroviral Therapy; AST: Aspartate Aminotransferase; AB: amyloid-beta; BBB: Blood-Brain Barrier; CAR: Chimeric-Antigen-Receptor; CCL: Chemokine (C-C) Ligand; CCR: CC Chemokine Receptor; CDR3: Complementarity-Determining Regions; CNS: Central Nervous System; COVID: Coronavirus Disease; CSF: Cerebrospinal Fluid; CXCL-: Chemokine (C-X-C) Ligand; DAA: Direct-Acting Antivirals; Dtx: Diphtheria toxin; EBV: Epstein Barr Virus; EOAD: Early Onset AD; FDA: US Food and Drug administration; Foxp3: Forkhead Box Protein 3 gene; GWAS: Genome Wide Associated Studies; HAND: HIV-Associated Neurodegenerative Disorders; HCMV: Human Cytomegalovirus; HCV: Hepatitis C virus; HHV: Human Herpesviruses; HIV: Human Immunodeficiency Virus; HSV-1: Herpes Simplex Virus-1; IFITM3: Interferon Induced Transmembrane protein 3; IFN-: Interferons; IFNAR1/2: IFN1/2αβ- Receptor; IgM/G: Immunoglobulin-M/G; IL-: Interleukins; IP-10: Interferon-y Inducible Protein-10; ISGs: Interferon Stimulated Genes; LOAD: Late Onset AD; MAITs: Mucosal-Associated Invariant T-cells; MCI: Mild Cognitive Impairment; MCP-1: Monocyte Chemoattractant Protein-1; MHC-I/II: Major Histocompatibility Complex I/II; MR1: MHC Complex Class I-related molecules; MRI: Magnetic Resonance Imaging; MS: Multiple Sclerosis; NFTs: Neurofibrillary Tangles; NHS: National Health Service; NK: Natural Killer cells; PAMP: Pathogen-Associated Molecular Pattern; PBMC: Peripheral Blood Mononuclear Cells; PD-1: Programmed Cell Death Protein 1; PRRs: Pattern-Recognition Receptors; PSEN1/2: Presenilin1/2; Rag: Recombination Activating Gene; RANTES: Regulated on Activation, Normal T-cell Expressed and Secreted; RNA-seq: RNA-sequencing; SARS-CoV2: Severe Acute Respiratory Coronavirus 2; SPP1: Secreted Phosphoprotein 1 Osteopontin; TAP: Transporter associated with Antigen Presentation; TCRs: T-cell Receptors; Tdap: Tetanus, Diphtheria, Pertussis; T_{EMBA}: Effector Memory T-cells; T_{FH}: T Follicular Helper T-cells; TGF-β: Transforming Growth Factor-β; T_H cell: CD4* helper T-cells; TNF: Tumour Necrosis Factors; Tregs: Regulatory T-cells; TGG: TCR₃-chain; UK: United Kingdom; VZV: Varicella Zoster Virus; ₃δ: Gamma-Delta T-cells.

Introduction

Dementia is currently the leading cause of death in women and the second in men in the UK [1] and represents a significant global health burden. The most recent up-to-date evaluation of dementia sufferers in the UK stated that there are approximately 900, 000 patients with dementia [2]. Longterm projections forecast this to rise to 1 million by 2025 and to 1.6 million people by 2040 [2]. Alzheimer's disease (AD) is the most common form of dementia and global estimates suggest there are close to 50 million AD patients worldwide that are predicted to reach 152 million by 2050 [3]. This devastating impact on human life is further exacerbated by the lack of any highly effective treatment and poses a significant financial burden to the UK economy, commanding £34.7 billion of the NHS annual budget [2] with global costs estimated near \$1 trillion [3].

AD is a progressive neurodegenerative disease where pathology is driven by a combination of genetic, environmental, and immunological processes (reviewed in Ref. [3]). Neuropathology is defined by the presence of neuritic plaques



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and neurofibrillary tangles (NFTs) in the brain that have developed due to amyloid-beta accumulation (A β) and tau phosphorylation that leads to synaptic loss and dysfunction of neuronal cells, described below [3]. Early symptoms include loss of memory, language, visuospatial awareness, concentration, orientation, and mood. Late symptoms include delusions, hallucinations, personality loss, and loss of body/ motor control [4].

There is no current cure for AD, but recent exciting advances have been made that focus on new immunotherapybased anti-A β antibody treatments [5, 6]. The recent US Food and Drug Administration (FDA) approval of these treatments (discussed below) could not only potentially change the treatment outlook for AD patients but also highlight the key role that the immune system plays in the disease. This then raises the possibility that targeting other mechanisms within the immune system during AD could also be an effective way to treat disease.

It is widely accepted that both innate and adaptive immune cells are found within the central nervous system (CNS) in multiple neuropathological disorders including AD [7]. However, recent interest in the role of adaptive immune cells in AD and the emergence of the newly approved anti-A β immunotherapy has refocussed research attention on studying the key immunological mechanisms that contribute to pathology. This review discusses some of these recent findings and describes the inflammatory-mediated adaptive immune responses that are associated with viral, and to a lesser extent bacterial, infections during AD.

AD pathology

Neuropathology of AD can be classified into two main subsets: (i) an accumulation of NFTs, amyloid plaques, dystrophic neurites, neuropil threads, and other deposits found in the brains of AD patients; (ii) atrophy of the brain leading to synaptic and neuronal loss [3].

Amyloid plaques are extracellular deposits of A β that are synthesized as a result of proteolytic cleavage of amyloid precursor protein (APP) peptide by alpha-, beta-, or gammasecretases that gives rise to A β peptides of varying length (37–43 amino acids) [8]. In non-pathological conditions, A β 40 is the most predominant peptide produced as compared to other varying length A β peptides that include the longer amyloidogenic A β 42 peptide [9, 10]. However, genetic mutations or excessive overproduction of A β 42 peptide in combination with longer A β isoforms versus shorter isoforms increases A β monomer aggregate formation [10].

A β monomers can then develop into amyloid fibrils forming the aggregate plaques or develop into insoluble oligomers that can spread throughout the brain and accumulate in the parenchyma [11]. As plaques form, reactive microglia and astrocytes are recruited to these sites generating curvature and distortion of both axons and dendrites (dystrophic neurites) [10, 12]. This damage leads to an impairment of neuronal function and synaptic loss (see below) [10, 12].

NFTs are formed as a result of hyperphosphorylation of the tau protein that develops into filament-like structures that can become twisted throughout the parenchyma forming paired helical filaments [3]. These are not only found to accumulate mostly in axons but are also found in dendrites and result in the loss of cytoskeletal microtubules and tubulinassociated proteins [3]. NFT development can progress from a pre-tangle phase to more mature NFTs and then later extracellular tangles [3]. This results in neuronal loss through destabilization of the neuronal microtubules due to excessive accumulation of tau that is resistant to proteolysis [3, 13, 14]. A β also initiates a pathway that leads to tau-dependant synaptic dysfunction that directly correlates with the symptoms of progressive cognitive decline in AD patients [13, 14].

Synaptic loss arises due to dysfunction of neuronal cell processes [3]. This can include defective axonal transport, mitochondrial function, oxidative stress, and as described above, accumulation of A β and tau at synaptic sites [3]. Combined, these lead to brain atrophy through a progressive loss of pre-synaptic terminals, dendritic spines, and axonal function resulting in neuronal cell death [3]. Understanding the interplay of A β and tau with the immune system may help identify novel ways to treat AD [13, 14].

Genetic risk factors in AD

There are multiple genetic risk factors affecting the risk of developing AD that can be classified and subtyped based on the patient's age of disease onset and the method of inheritance [15]. In genetic terms, these are classified as Early Onset AD (EOAD) and Late Onset AD (LOAD) (reviewed in Ref. [15]). EOAD is commonly referred to as Familial AD where the individual receives one risk allele from either parent in an autosomal dominant pattern [16]. Approximately 35-60% of EOAD patients have first-degree relatives with dementia that include 10-15% autosomal dominant families within three generations or more [17]. EOAD is a rare form of AD and is caused by mutations in either APP, presenilin1 (PSEN1), or presenilin 2 (PSEN2) and typically develops between the ages of 30 and 60 [10]. Importantly, only 5–10% of EOAD patients can be explained by the pathogenic mutations within these three familial genes [15]. This suggests that non-A β pathways may contribute to AD pathology and are discussed below. LOAD, also referred to as Sporadic AD, is more polygenic and presents after the age of 65 with a genetic aetiology of up to 82% [15]. Multiple risk factors, including age, environmental factors, and multiple genetic variants, are also associated with LOAD [16].

Studies using large-scale genome-wide associated approaches (GWAS) and whole genome analyses have identified up to 75 risk loci with AD pathology with 42 new loci identified recently [18]. This study used 111, 326 AD patients and 677, 663 controls from 15 European country databanks [18]. Importantly, this study identified 22 immunerelated risk loci that associate with immune functions and placed these into functional processes using gene ontology [18]. The authors described these as 'tier 1' related genes signifying a greater likelihood of being the causal risk gene responsible for AD [18]. 21 of these immune risk loci genes are novel [18] and complement the already described 28 immune risk loci genes reviewed by Frost et al. [19]. These include importantly Apoe, Trem2, Cd33, Clu, Cr1, Plcg2, Abi3, and the Ms4a and Hla families [10, 19] among others, and their association with AD are well described by Frost et al. and will not be discussed at length here [19].

The *Apoe* gene encodes apolipoprotein E and was the first genetic risk factor found associated with LOAD and is the most significant risk loci to AD [10]. APOE has been well described and is shown to play a role in both innate and adaptive immune function and can regulate levels of A β [20]. The main function of APOE is the regulation of lipid transport and in the brain is expressed mostly by astrocytes and

oligodendrocytes; however, expression of APOE is greatly increased by reactive microglia during AD [10]. Importantly, individuals who carry the ε 4 allelic isoform of the gene (*Apo* ε 4) have a 3–4-fold increase in the likelihood of developing AD [20, 21]. It has also been shown in both mice and humans that *Apo* ε 4 carriers also have increased A β aggregates and tau accumulation [22–24], whereas carriers of the ε 2 allelic isoform (*APO* ε 2) can enhance A β clearance and have lower AD risk [25].

APOE4 has also been shown to promote blood-brain barrier (BBB) dysfunction, even in individuals who are cognitively intact [26, 27]. However, in cognitively impaired individuals, APOE4 can result in more severe BBB dysfunction even though no effect on cerebrospinal fluid (CSF) A β and tau levels are seen [26, 27]. This could suggest that in APOE4 carriers the BBB dysfunction may allow recruitment of immune cells into the brain driving neuroinflammation. APOE has also been shown to influence the function of T-cells during AD and is discussed below.

The immune system and AD

The brain was classically thought to be a site of immune privilege that was inaccessible to the systemic immune system. However, multiple studies have revealed a direct link between the brain and circulating immune cells [28–30]. The exposure of the brain parenchyma to the full force of the immune system can be further exacerbated through loss of homeostatic protective measures, including BBB damage and dysregulation, systemic and vascular inflammation or impaired meningeal lymphatic drainage, all of which are observed in AD [10]. This could promote infiltration and recruitment of systemic immune cells into the brain contributing, in the context of AD, to neuroinflammation and AD pathology [7].

Whilst the roles of both innate and adaptive arms of the immune system in AD have been well classified, this review will focus on recent work describing the neuroinflammatory properties of the adaptive immune system and the responses associated with microbial infections that contribute to AD pathology.

Adaptive immunity

One of the most described adaptive immune cells that contribute to AD pathology is T-cells (reviewed in Ref. [26, 31]. Multiple studies have shown the presence of T-cells in the leptomeninges and the hippocampus of brains collected post mortem from human AD patients [26, 32, 33]. This increase in T-cell presence is more prevalent in the CD8+ lineage as compared to the CD4⁺ lineage and is associated with more severe disease when found in the hippocampus [26, 32, 33]. This is also seen in AD mouse models where the increased prevalence of T-cells in the brain is associated with an increase of both A β and tau (reviewed in Ref. [26]). However, contrasting studies have described how the CD8+ T-cell infiltrate correlates only with an increase in tau accumulation but not A β formation [32] and may potentially be linked to overall systemic "inflammaging" (chronic inflammation when ageing) [34].

Mouse studies have now shown that the infiltrate of CD8⁺ T-cells into the brains of APP/PS1-21 AD mice can modulate neuronal and synapse-related gene expression [33]. This suggests that these infiltrating T-cells either directly interact with microglia or astrocytes, or neuroinflammatory properties mediated by cytokine/chemokine release are influencing neuronal cell function. This study went on to show that even when APP/PS1-21 AD mice were treated with depleting anti-CD8 antibodies, neither plaque formation nor cognition was altered [33]. This may suggest that CD8⁺ T-cell function alone may not be sufficient to drive pathology. However, more recent data now suggests that T-cells can directly interact with microglia and that depleting both T-cells and microglia can reduce tau-mediated neurodegeneration [35]. This interaction now suggests a dual pathological role of T-cells contributing to AD pathology, by directly interacting with the brain-resident microglia influencing their function, whilst also simultaneously contributing to proinflammatory cytokine release.

This pathogenic role of CD8⁺ T-cells in AD is also confirmed in studies that identified that this increased prevalence is restricted to the CD8⁺/CD45RA⁺ effector memory ($T_{\rm EMRA}$) lineage and that these are found in both the blood and CSF of human AD patients [36]. This study went further and also identified that these CD8⁺ T-cells found patrolling the CSF were clonally expanded and antigen experienced as they retained Epstein Barr Virus (EBV)-reactive T-cell receptors (TCRs) (discussed further below) [36].

However, there is conflicting evidence as to the role of T-cells in AD pathology. Marsh et al. took the approach of crossing Rag2--- and Il2ry--- mice with 5xFAD AD mice to generate an AD mouse model system that is deficient of T-, B-, and Natural Killer (NK) cells [37]. They went on to show that the depletion of immune cells within this AD mouse results in an increase in severe Aß pathology with enhanced neuroinflammation and decreased microglial activation [37]. They also showed that adoptive bone marrow transfer and restoration of T-, B-, and NK cells in these mice reduced Aß pathology and increased microglial activation [37]. However, this study has taken an approach to delete all immune cells including CD8+, CD4+, and Regulatory T-cells (Tregs) and does not discriminate between the pathological or protective roles of these different immune subsets during AD. Similar studies also crossed Rag2--- mice with APP/PS1AE9 AD mice (now only deficient in B- and T-cells) and showed that after adoptive bone marrow transfer, there is a significant reduction in A β levels [38]. These studies may point to discrepancies in the different AD mouse models used but also highlight that AD pathology can be significantly influenced by both T- and possibly B-cells. Direct antigenic stimulation of either T- or B-cells was not examined in these studies, so it is possible that activation of these cells via this route could drive an enhanced neuroinflammatory response exacerbating disease but requires further study.

It is also important to consider what effect CD4⁺ helper T-cells (T_H) have during AD either in combination with CD8⁺ T-cell function or in isolation. The pathological role of non-regulatory CD4⁺ T-cells during AD has now revealed that CD4⁺, T_H1, and T_H17 cells are directly contributing to pathology and are found within both human and mouse AD brains [31, 39]. Studies using APP/PS1-21 AD mice have shown that not only do the T_H1 and T_H17 cells enter the parenchyma but they also stimulate a range of proinflammatory cytokines including interleukins (IL-), tumour necrosis factors (TNF-), and interferons (IFN-) such as IL-6, IL-1 β , TNF- α , IFN- γ , and monocyte chemoattractant protein-1 (MCP-1), that, in turn, influences microglia and astrocyte function [19, 39] (reviewed in Ref. [26]). However, conflicting studies have shown that adoptive transfer of A β -specific T_H1 cells into

5xFAD AD mice leads to an increase in T-cell mediated activation of MHC-II⁺ microglia that display increased phagocytic activity of A β [40]. This study may support the theory that enhanced T-cell activation is advantageous during AD in helping clear A β plaques and may provide an effective therapeutic strategy.

The role of T_H17 cell involvement in AD pathology is less well described. However, T_H17 cells have been shown to induce neuroinflammation and neurodegeneration through the production of IL-17 and IL-22 in rat AD models [41]. This study identified that the release of proinflammatory cytokines mediates neuronal function through the Fas/FasL apoptotic pathway [41]. More recent studies went further and developed clonal A β -specific T_H1 and T_H17 effector T-cells (Teffs) in vitro. The authors adoptively transferred these cells into APP/PS1-21 AD mice and showed that the presence of these Aβ-reactive Teffs accelerated systemic and brain inflammation, impaired cognition and increased amyloid burden and microglial activation [31]. Taken together this suggests the potential to either directly target and suppress the T-cellmediated neuroinflammatory responses, the T-cell itself or block the Fas/FasL pathway. This may represent an innovative approach for the design of novel therapies for AD and contradicts the theory that increased T-cell activation in AD could be advantageous.

Tregs have also been implicated in AD development (reviewed in Ref. [42]). These CD4⁺ T-cells express the transcription factor forkhead box protein 3 gene (Foxp3) and typically regulate the immune system through the production of immunosuppressive cytokines including transforming growth factor- β (TGF- β) and IL-10 [43]. Foxp3⁺ Tregs have been found to be systemically elevated in 5xFAD AD mice and also in elderly AD patients [44]. In these human studies, patients with mild cognitive impairment (MCI) were shown to have increased expression of programmed cell death protein 1 (PD-1)-negative Tregs [44]. This may suggest that in contrast to the studies described above that show increased numbers of proinflammatory CD4+ and CD8+ effector T-cells in the brain during AD, there is an overall increase in immunosuppression mediated by Tregs that may be contributing to pathology [44]. Baruch et al. went further by suggesting that due to the significant upregulation of Tregs, it would be appropriate to enhance the hosts' immune system to counter the immunosuppressive environment [45]. Based on this strategy, Baruch et al. treated 5xFAD and APP/PS1-21 AD mice with anti-PD-1 monoclonal antibodies [46]. They showed that PD-1 treatment developed elevated CD4+ IFN-y responses that subsequently resulted in an overall decrease in A β plaque formation, rescued cognitive performance, and promoted monocyte recruitment into the brain [46]. The authors rightly state that this increased infiltrate of activated immune cells into the brain would need to be carefully monitored for any bystander damage [46, 47].

Another way to reduce Treg-mediated suppression has been performed in studies that deplete Tregs completely *in vivo* by crossing Foxp3-diphtheria toxin mice (Dtx) with 5xFAD AD mice [45]. These studies showed that when Tregs are depleted, 5xFAD AD mice not only displayed improved cognitive behaviour with reduced A β pathology, but also lead to an influx of macrophages and T-cells into the brain [45]. However, Treg-depleted mice will develop enhanced autoimmunity and non-specific T-cell activation [48], which in combination with enhanced immune cell infiltrate into the brain would need to be closely monitored [45] and would pose challenges if chosen as a treatment. IL-10 as mentioned above is an immunosuppressive cytokine and can be produced by multiple T-cell subsets including Tregs, and has been shown to be upregulated in the brains of AD patients [49]. Studies have also examined what effect depleting IL-10 in vivo has on AD pathology [49]. Using Il-10-/- mice crossed with APP/ PS1-21 AD mice, the authors observed a preservation of synaptic ability, mitigated cognitive decline, and cerebral amyloid accumulation [49]. This data may suggest a mechanism behind the observation that Treg depletion, and thus IL-10 depletion restores cognitive function supporting the hypothesis described by Baruch et al. above [45]. Depleting IL-10 would be a more manageable strategy rather than targeting whole cell lineages such as Tregs. However, it is not known if Tregs are the main producers of IL-10 during AD as another source of IL-10 during AD could be T_H2 cells [50]. Studies have shown that the adoptive transfer of WT T_{H}^2 cells into APP/PS1-21 AD mice can improve cognitive function and reduce AD pathology [51]. This study, therefore, suggests that immunosuppression elicited by IL-10 may be useful during AD. However, this may depend on whether the adoptively transferred T_H^2 cells in this study are actually releasing IL-10 as this may only arise if the T_{H}^2 cells are being activated. This highlights the therapeutic potential of targeting IL-10 in vivo in AD but suggests the direct impact of the cellular source of upregulated IL-10 needs further study. There are also ongoing studies that aim to enhance Treg function during AD to suppress immune activity that are discussed below.

The role of other subsets of the adaptive immune system in AD are more poorly understood. However, recent studies have suggested their potential role in disease. These include CD4⁺ T follicular helper T-cells (T_{FH}), B-cells, NK cells and gamma-delta ($\gamma\delta$) T-cells.

T_{FH} cells are the main source of proinflammatory IL-21 [52]. Systemic increases of IL-21 have been shown in both AD and MCI patients; however, these studies have not identified whether this was solely due to T_{FH} production and to date, no studies have shown the presence of T_{FH} cells in an AD brain [52]. These patients also displayed increases in both $A\beta$ peptide-specific plasma IgM and IgG levels [53]. This is also seen in 5xFAD AD mice as compared to controls suggesting an IL-21 T_{FH}-mediated inflammatory response may be contributing to pathology [54]. IL-21 can also stimulate T_{FH} cells in an autocrine manner that could further exacerbate the presence and contribution of T_{FH} cells in AD [54, 55]. An excess production of IL-21 during an inflammatory response also mediates IL-17 release by T_H17 cells and thus could suggest that T_{FH} cells are an important immune mediator of excessive inflammation of other immune subsets in AD [54, 55].

B-cells are essential for the production of antibodies in mammalian biology [56]. The role of B-cells in AD is not fully understood; however, B-cells have been shown to be present in the brain parenchyma of 3xTG and APP/PS1-21 AD mice [57]. It was first thought that the presence of B-cells in the brain may produce immunoglobulins that were specific to, and interfere with A β plaque formation [57]. However, this study found that depletion of B-cells in AD mice by crossing 3xTG or APP/PS1-21 AD mice with J_HT mice (B-cell deficient) resulted in a lack of immunoglobulin deposition in the brain that subsequently reduced AD pathology. This result was also confirmed using anti-B-cell production of immunoglobulins, an event that happens in response to microbial infections, can contribute to AD pathology [57].

NK cells are another subset of immune cells that play a vital role in host defence and bridges the gap between the innate and adaptive immune response [58]. NK cells can kill an infected cell directly via production of cytokines or mediate antibody-dependent cellular cytotoxicity (ADCC) [58]. The role of NK cells in AD is poorly understood, but studies have shown that depletion of NK cells using anti-NK1.1 antibodies dramatically improves the cognitive function of 3xTG AD mice [59]. This depletion of NK cells was also associated with a decrease in microglial proliferative capacity and rescued inflammatory cytokine production [59]. It has also been shown that in human AD patients most systemic NK cell numbers contract; however, RNA-sequencing (RNA-seq) analysis has revealed an expansion of a unique NK cell subset expressing Cx3cr1, Tbx21, Myom2, Dusp1, and Zfp36l2 that may potentially drive NK-mediated AD pathology [60]. Together these studies imply that NK cells play an important role in AD pathology, but similarly to T-cells their effect may be restricted to proinflammatory cytokine release or ADCCmediated damage. It may be, therefore, useful to target for deletion the expanded NK subset described above or define the specific proinflammatory cytokines released by these cells.

 $\gamma\delta$ T-cells are less abundant than CD4⁺ T_H or CD8⁺ T-cells but they play an important role in anti-microbial defence but can also support wound healing and immune tolerance [61]. Using TCR deep sequencing of both human blood and brain samples from AD patients, analysis of the TCR revealed clonal diversity and somatic variability within the TCR γ -chain (TRG) and identified putative TCR clonotypes that were more specific in both the brain and blood [61]. Whilst this highlights that a biased $\gamma\delta$ TCR subset may contribute to disease, further studies are needed to elucidate the importance of these findings and whether any dominance of the TRG region or other TCR complementarity-determining regions (CDR3) can be used as markers of AD pathology [61].

There are other immune cells not discussed at length here that may potentially contribute to AD pathology including mucosal-associated invariant T-cells (MAITs). Interestingly, brain astrocytes and microglia can express functional MHC complex class I-related molecules (MR1) that present microbial antigens to MAIT cells [62]. This raises the possibility that microglial antigen recognition by MAITs, similar to that seen with T-cells [35], may contribute to AD pathology, but any mechanism for this has not been described and would need further study to dissect any link.

It is also worth considering what influence AD genetic risk loci factors have on T-cell function. T-cell activation is elevated in individuals that express *Apo*e4 (risk allele) as compared to those that express *Apo*e2 or *Apo*e3 [63]. This activation is also seen *in vivo* using *Apo*e^{-/-} mice that have increased levels of T_H1 and T_H17 cells in their brains with concurrent increases in proinflammatory cytokine release (IL-17, IFN- γ , TNF- α , IL-12, IL-1 β , and IL-6) [64]. This data show that APOE can modulate T_H1 and T_H17 proinflammatory-mediated cytokine responses [64] and is a key regulator of T-cell activation; however, *in vitro* studies suggest that APOE lipoproteins can actually inhibit T-cell activation by limiting inflammatory cytokine release [65, 66]. More work is needed to discover whether activated T-cells and neuroinflammation seen during AD are a consequence of the individual expressing *Apo*e4.

Cytokines, inflammation, and AD

As mentioned throughout, AD pathology can be exacerbated by the production of cytokines and chemokines that can drive neuroinflammation. These are released either through: (i) the activated systemic innate and/or adaptive immune response which then recruits to the brain; (ii) through the interaction of the immune cells with brain-resident cells; or (iii) released via aberrant brain-resident microglia and astrocytes themselves. These include those that are most well described, that are either proinflammatory, including IL-1 β/α , IL-6, IL-18, TNF- α , and IFN- γ ; and those that are anti-inflammatory, including IL-4, IL-10, and TGF- β [19, 67].

There are 15 cytokines that have been associated with AD with at least 23 cytokine polymorphisms associated with disease risk [67, 68]. Zheng *et al.* classified all cytokines associated with AD into three main conditions "(1) having polymorphisms that are significantly associated with AD, (2) having corresponding genotype/phenotype data, and (3) having previous records of the changed levels in AD patients" [67, 68]. Dysregulation of these cytokines can then contribute to excessive A β production in an inflammatory manner that subsequently leads to further increases in IL-1 β , IL-6, TNF- α , and IFN- γ production by glial cells, creating a vicious cycle of pathogenic events [67].

New data have suggested a role of the proinflammatory cytokine secreted phosphoprotein 1 Osteopontin (SPP1) in AD [69]. This study showed that SPP1 is upregulated by perivascular macrophages and is required by microglia for synaptic phagocytosis in the hippocampus of APP-NL-F AD mice [69]. The authors suggest that SPP1-microglia crosstalk mediates aberrant microglial activity during AD [69]. SPP1 release has also been shown to promote the survival of autoreactive T-cells in the brain of multiple sclerosis (MS) mouse models [70]. This may suggest that the microglia may also become activated by the presence of these autoreactive T-cells further exacerbating microglial synaptic phagocytosis, but would need further studies to confirm any association.

Recent studies have also suggested that IFN release plays a key role in A β -mediated pathology (reviewed in Ref. [71]). The IFN family have widespread anti-viral or immunemodulatory functions and is released upon pathogenassociated molecular pattern (PAMP) recognition (reviewed in Ref. [71]). Microglia and astrocytes both express patternrecognition receptors (PRRs) that can recognize PAMPs and their engagement results in transcription of cytokines (described above) including type-I IFNs [71]. There are three major classes of IFN: type-I IFNs comprising IFN- α and IFN- β , type-II IFN (IFN- γ) and type-III IFN (IFN- λ) [72]. Cell signalling is induced when type-I IFNs bind to the IFN- $\alpha\beta$ receptor (IFNAR1 and IFNAR2) and initiate transcription of interferon-stimulated genes (ISGs) [72].

Multiple studies have shown that ISGs are elevated in the brains of AD patients as compared to healthy controls [73, 74]. These findings were supported using *in vivo* 5xFAD AD mouse models where type-I IFN activation was shown to induce A β pathology in microglia and other neuronal cells [75]. The authors showed that blocking IFNAR signalling in 5xFAD AD mice reduces both microglial cell accumulation and synapse loss [74] and that specific deletion of IFNAR1 expressed on microglia can rescue memory and reduce synaptic defects [75]. IFNAR1 deletion on other neuronal cells has also been shown to restore synaptic terminals and decrease A β plaque

formation [75]. The importance of type-I IFN signalling in AD pathology was shown in studies using APP/PS1-21 AD mice where primary microglia isolated from the brains of mice with ablated type-I IFN signalling displayed increased phagocytic capacity to uptake $A\beta1-42$ [76].

GWAS studies have also shown that viral-mediated IFN responses are associated with an increase in microglialmediated tau pathology [77], and as discussed below, viral infections can play a critical role in exacerbating AD. More recent studies went further and used transcriptomic approaches to reveal polymorphisms in ISGs within the hippocampus in multiple mouse models of AD and compared these findings to human GWAS AD datasets [78]. These studies suggest that ISG polymorphisms may increase the overall risk of AD [78]. However, these require further study to describe the mechanisms underpinning their involvement and whether these are associated with microbial infection or microglia function. This is now of significant importance given that Interferon-induced transmembrane protein 3 (IFITM3), a key anti-viral restriction factor, is activated by inflammatory cytokines in neurons and astrocytes and binds gammasecretase subsequently upregulating A β production [79]. The authors showed that IFITM3 is upregulated in LOAD patients and that crossing Ifitm3--- mice with 5xFAD AD mice results in reduced gamma-secretase activity with reduced amyloid plaque formation [79]. This study suggests a key role for IFNinduced risk of neuroinflammation and exacerbation of AD pathology. Recent studies have suggested a mechanism behind this type-I IFN-mediated effect whereby pathogenic tau stimulates microglia to release type-I IFN that is mediated by cGAS-STING signalling in 5xFAD AD mice [80]. This study suggested that deletion of cGAS induces a myocyte enhancer factor 2c gene (*Mef2c*) network that promotes cognitive resilience and thus suggests that cGAS signalling may be an effective therapeutic target in AD [80]. Whilst this new study does reveal new mechanistic insight, and as described by Sandford et al. [71], it is still not fully understood how IFN production directs the immune system to exacerbate AD pathology and tauopathy and requires further study [71, 78].

Chemokine signalling is another major component of adaptive immune function. Mononuclear phagocytes such as monocytes and macrophages are some of the primary inducers of inflammation-mediated via chemokine release/signalling and have been shown to influence AD pathology (reviewed in Ref. [47]). Several chemokines such as CCL2 (MCP-1), CXCL8 (IL-8), CXCL10 (IP-10), and CCL5 (RANTES) are also produced in response to Aß peptide deposition and have been shown to regulate both microglial and astrocyte migration and the recruitment of other peripheral immune cells into the brain [67]. Activated microglia have also been shown to contribute to pathology mediated via a chemokine axis and can inhibit neuronal activity through expression of CCL-3/-4/-5 binding to neuronal CCR5 [81]. This study showed that genetic deletion of CCR5, whose expression is increased in mouse models of tauopathy, ameliorates tau pathology [81]. This suggests a paracrine signalling effect by activated microglia that influences neuronal function. Microbial infections are known to infect monocytes, macrophages, and microglia with some microbes exploiting chemokine receptors for viral entry [82]. AD pathology may, therefore, be induced as a side effect of neuroinflammation via infection-mediated chemokine release, similar to that seen with cytokines released during infection (discussed below).

Viral infection and AD

Recent data now highlights the potential role of viral infections in exacerbating cognitive decline and AD [83]. It is well known that many viruses are neurotropic and can access the brain driving neurological impairment including herpes simplex virus-1 (HSV-1), SARS-CoV-2, Polio, and West Nile Virus [84–86]. Further evidence suggests that human herpesviruses (HHV), which establish life-long chronic infections, can be found in the brains of deceased AD patients including HSV-1, HHV-6A, and HHV-7 [84, 87].

It was first suggested over 40 years ago that HSV-1 infection can induce encephalitis in the brain that displays similar properties to AD [88]. Later studies went on to confirm this discovery showing that HSV-1 genomic DNA and a functional HSV-1 genome is found in AD patient brains [89, 90]. Studies next went on to investigate the mechanisms behind these observations and identified that direct HSV-1 infection in primary neurons increases protein kinase A that mediates tau phosphorylation giving rise to dystrophic neurites and AD pathology [91, 92]. Lövheim *et al.* went on to state that HSV-1 plays an important role in early AD development and were able to detect the presence HSV-1-specific antibodies in plasma samples taken 6.6 years prior to the onset of dementia [93]. This, therefore, could suggest that HSV-1 circulating antibodies can potentially serve as risk biomarkers of AD in those patients who are also at genetic risk of developing AD.

More recent studies have examined what effect co-infection has on AD pathology and neuroinflammation [94]. Using human-induced neural stem cell cultures that were infected with quiescent HSV-1 and/or varicella-zoster virus (VZV), the authors showed that VZV infection alone displayed no typical characteristics of AD such as AB and tau accumulation but was able to induce gliosis and proinflammatory cytokine release [94]. The authors also showed that HSV-1 infection alone is enough to induce typical AD-like features, as stated before, and strikingly they showed that VZV infection of cells was able to reactivate HSV-1, which then drives A β and tau accumulation [94]. They went on to state that shingles induced by VZV infection could indirectly contribute to AD by promoting sufficient neuroinflammation to reactivate HSV-1 which then directly drives AD pathology [94]. Epidemiological studies have also shown that in patients over the age of 50 with an untreated active HSV-1 or VZV infection, the overall risk of dementia is increased 1.5-fold [94]. These studies have also shown that anti-viral medication could also lower the risk of dementia by 25% as compared to the untreated herpes-infected individuals [95].

Due to the very high seroprevalence of other herpes viruses within the human population, studies have shown that infection with the common β -herpesvirus human cytomegalovirus (HCMV) is also associated with AD [96-98]. HCMV infection results in increased localized viral-specific inflammation found in both the blood and brain of AD patients that is associated with worsening rates of cognitive decline and an increased accumulation of A β [96–98]. However, there are conflicting epidemiological studies that suggest CMV seroprevalence alone is not associated with AD pathology and that a co-infection hypothesis of CMV with HSV-1 is the driver of AD development and is similar to what is described above with HSV-1 and VZV [99]. Though chronic infection with HCMV results in viral latency, the virus is reactivated upon a host becoming immunocompromised or in a co-infection setting may reactivate by a host's redirection of the immune system to another active viral infection [100]. However, more studies are needed to confirm the mechanisms behind these findings. This co-infection hypothesis is further supported by a recent study showing that HSV-1 and SARS-CoV-2 infection in human CSF can result in amyloid aggregation of proteins known to be involved in AD, including APOE [101]. This may suggest that active site-specific viral replication within the CNS may be enough to trigger AD pathology.

This theory is supported by studies showing that EBV-specific TCRs are found to be both antigen experienced and clonally expanded within the CNS of AD patients [36]. Tiwari et al. support this theory and suggest that EBV-encoded proteins (e.g. BNLF-2A) can induce AD by interfering with antigen processing and presentation by inhibiting cellular TAP (transporter associated with antigen presentation) functions and downregulating MHC-I and MHC-II expression [102]. The authors showed that upon infection, this process can lead to an accumulation of neuronal cells and viral polypeptides that subsequently promote a build-up of oligomers and amyloidlike aggregates using in vitro Thioflavin-S fluorescence assays [102]. EBV infection is also known to be associated with a range of neurodegenerative conditions such as Parkinson's disease, viral-induced encephalitis, and Meningitis (reviewed in Ref. [103]). Recent large-scale epidemiological studies in over 10 million people show the risk of MS is increased 32-fold in EBV-infected individuals and suggest that EBV is the leading cause of MS [104]. The neurodegenerative events seen during infection may potentially arise due to systemic EBV-infected peripheral blood mononuclear cells (PBMC) crossing the BBB and replicating within brain endothelial cells promoting cytokine release and a loss of neurons [103, 105]. The constant systemic switch from latency to reactivation events associated with infection can, therefore, continually drive systemic stress further exacerbating BBB crossover and cognitive deficits seen with AD and other neurodegenerative diseases [103, 106]. Taken together, these studies highlight how herpes viruses that establish life-long infection increase the overall risk of developing AD. Future work must be performed in this area to identify the key viral-induced mechanisms behind these observations and whether anti-viral treatment could be useful in AD.

Recent attention has been extended to examine what effect the recent circulating pandemic SARS-CoV-2 virus has on AD pathology. Whilst all seven members of the *Coronaviridae* family have been shown to be neurotropic [107], the association of SARS-CoV-2 infection with AD is still in its infancy due to the recent emergence of the virus. However, it has been well described that cognitive deficits and dysfunction are observed in patients post-SARS-CoV-2 infection even after mild infections [108]. This neuroinflammatory nature of the infection has been shown to be driven by increased levels of CCL11 in the CSF and serum that leads to microglial activation and loss of neuronal function in mouse brains [109]. Whether these same viral-induced processes apply in exacerbating AD is not yet known, but they provide useful mechanistic insights to help inform future AD studies.

In the context of AD, recent epidemiological studies in over 6.2 million people >65 years of age, have suggested SARS-CoV-2 infection is associated with a nearly 2-fold increased risk of AD (0.35% (non-COVID-19) to 0.68% (COVID-19+) and these risks are significantly elevated within 360 days of infection, especially in people >85 years of age and in women [110]. Further studies have shown that within UK biobank cohorts, Apoe4 homozygotes were 2.31 times more likely to test positive for SARS-CoV-2 than Apoe3 homozygotes [111]. Studies that have used human-induced pluripotent stem cells have shown that APOE4 isogenic neurons and astrocytes observe a higher rate of SARS-CoV-2 infection compared to Apoe3 genotype with increased astrocytic apoptosis [112]. This is supported by studies that show $A\beta 42$ can bind with high affinity to the S1 subunit of SARS-CoV-2 and membrane-bound ACE2 (angiotensin-converting enzyme 2 receptor – S1 binds to ACE2 for viral entry) resulting in increased viral ingress and proinflammatory IL-6 production [113, 114]. ACE2 has also been shown to be significantly upregulated in the brains of AD patients irrespective of disease severity, gender, or age and has the potential to be potentially neuroprotective [115]. Furthermore, in vivo AD studies in non-SARS-CoV-2 infection models have shown that AB43 and A β 42, the two longer forms of A β , can be converted by ACE2 into the less toxic Aβ40 isoform and slow down Aβ42 aggregation in APP-transgenic J20 PDGF-APPSw AD mice [116]. Whilst this neuroprotective property of ACE may be useful in AD upon no infection, ACE2 upregulation may exacerbate AD upon SARS-CoV-2 infection with the virus exploiting this upregulated receptor for entry into endothelial cells in the brain to drive neuroinflammation. Combined, these studies demonstrate that SARS-CoV-2 infection can directly exploit the cellular factors that are associated with AD pathology.

The existence and now continued prevalence of SARS-CoV-2 circulating within the human population poses a risk to AD patients given the neurotropic nature of the virus. Studies that examine co-infection of SARS-CoV-2 with herpes virus family members that are associated with elevated risk of AD are now, therefore, of vital importance. This is due to the high level of seroprevalence of these viruses that are known to independently contribute to AD and where the likelihood of co-infection within humans remains high.

Epidemiological studies have also associated hepatitis C virus (HCV) and intestinal infections with AD [83] (reviewed in Ref. [117]). In bipolar patient cohorts that go on to potentially develop AD, 31% of AD patients were found to be HCV+ as compared to 16% of non-AD patients who were HCV+ [118]. These findings may suggest that an HCV-specific neuroinflammatory response similar to that seen with herpes virus infections is contributing to AD as no study has yet reported any HCV within the brains of deceased AD patients. Importantly, however, predictive markers of HCV liver cirrhosis (elevated aspartate aminotransferase [AST] to alanine aminotransferase [ALT] ratio) have been associated with AD diagnosis [119]. Here, AD patients were assessed for a correlation between HCV predictive markers with neuroimaging scores, cognitive ability, CSF biomarkers, brain function/atrophy, and amyloid accumulation and found an elevated AST to ALT ratio of 7.932 is associated with AD diagnosis [119]. More mechanistic studies have identified that APOE has also been shown to be heavily involved in HCV virion assembly and directly interacts with HCV envelope glycoproteins [120], but the role this mechanism plays in AD is not understood. This, therefore, points to a potential HCV-AD link, but this requires much further study. Due to the recent emergence and success of direct-acting antivirals (DAA) for the treatment of HCV, as mentioned above, it would be important to examine the influence of DAA treatment in patients who go on to develop AD.

The association of human immunodeficiency virus (HIV) and AD within humans is less well defined. However, there are similarities that exist between the cognitive dysfunctions seen during HIV infection and AD termed HIV-associated neuro-degenerative disorders (HAND) [121]. Though no large-scale epidemiological study has shown a direct link between HIV and AD [83], a recent meeting abstract has shown that HIV+ individuals had a higher prevalence of AD and related dementia (ADRD) [122]. The clinical scores of ADRD that the authors used to describe AD are not classified in this study; however, this data may now suggest a direct association of HIV infection with AD but requires more detailed analysis upon data availability [122].

There are also many similarities between the brain-specific pathways and mechanisms associated with HIV infection and AD, which have been extensively studied in mice (reviewed in Ref. [123]). Multiple murine studies have shown that HIV infection results in an increase in Aß synthesis and tau phosphorylation that is mediated by the HIV-specific Tat protein (reviewed in Ref. [123]). Importantly, in vivo studies show that expression of the lentiviral-vector derived Tat protein in the hippocampus of APP-PS1 AD mice increases Aβ1-42 synthesis and the overall size of amyloid plaques [124]. More recent studies went further to describe how the HIV-specific Gag polyprotein can also increase Aß expression and modulate APP metabolism. Whilst the increase in Aß expression was shown to be neurotoxic, the increase in APP was shown to sequester the Gag protein to restrict HIV-1 release [125]. This suggests the potential of $A\beta$ to act in an anti-microbial manner and is described further below.

These *in vivo* studies strongly suggest a direct link between HIV infection and AD; however, human studies remain less convincing. Similar levels of A β 1-42 have been shown in the CSF of both HAND and AD patients [126]; however, it is important to consider that the HAND patients would all have been undergoing anti-retroviral therapy (ART) meaning interpretation of this data is influenced by therapeutic treatment for infection and different ART treatments can result in 50–200% increase in A β production in mouse neuronal cells [127]. Studies have shown that any accumulation of amyloid aggregates in HIV patient brain samples begins prior to ART [128]. This again suggests the true function of overproduction of A β during HIV-1 infection is currently unknown as to its relationship with AD onset.

Human studies went on to show that in samples taken from the brains of HIV-infected individuals, the presence of accumulated amyloid was mostly found within neurons, whereas in AD, the neuritic plaques associated with AD pathology are mostly found outside of neurons [129]. This accumulation of A β may arise due to dysregulation of microglia that would normally remove any excessive A β [129]. The authors described how microglia can act as HIV reservoirs and that infection may activate microglia which then promotes a neuroinflammatory environment and failed removal of A β [121, 129].

These studies suggest a strong overlap between HIV+ HAND and AD. However, important studies are needed to examine in more detail whether there is a distinction between HAND and AD. As suggested in Ref. [121], Turner *et al.* have been actively recruiting patients to evaluate this very point and to build on machine learning techniques that distinguish between magnetic resonance imaging (MRI) morphometric differences in the brains of HAND and MCI AD patients [130]. Taken together the question of whether HIV can cause AD currently remains as a strong association rather than a definitive direct causation. As the HIV+ cohorts are now becoming aged individuals and HIV can prematurely age brains [131], this may create an opportunistic environment that would be suitable for AD development/onset.

Current epidemiological studies suggest that at least 45 different viral exposures can be significantly associated with neurodegenerative disorders, with some associated up to 15 years after infection [83]. This, therefore, raises the possibility that vaccination strategies aimed at viral infections may help reduce the onset of dementia and AD. Human studies have shown that vaccination to herpes zoster or a tetanus, diphtheria, and pertussis (Tdap) vaccine was associated with a 25% and 18% lower risk, respectively, for dementia than compared to no vaccine controls [132]. A recent epidemiological study went further and calculated that vaccination with Zostavax against VZV, lowered dementia occurrence by 19.9%, with the vaccine being found to be more effective in women under the age of 80 [133]. The researchers stated that this was a more natural experiment as the data were obtained in a non-biased way where other factors that can reduce the risk of dementia in the vaccinated group (such as healthier lifestyles) were eliminated using unique natural randomization [133]. These studies show exciting promise and suggest that anti-viral treatment and viral-based vaccines, of which there are multiple available for a range of different infections, including those mentioned above, need to be urgently extended to examine their influence in more detail on AD development. However, it is also possible that the host systemic anti-viral inflammatory response may stimulate the immune system to elicit cytokine-mediated neuroinflammation that contributes to AD pathology. There does, however, remain a lack of clear data regarding the viral-induced immune mechanisms that directly drive AD development and thus require further study.

Non-viral infections and AD

Whilst the association of bacterial and fungal infections with AD risk have gathered recent interest, less information exists examining their impact on AD as compared to viral infections. It has been shown that infection with gram-negative bacteria, including Porphyromonas gingivalis, the main cause of chronic periodontitis, can induce neuroinflammation increasing the risk of developing AD with elevated A\beta1-42 levels shown in Apoe^{-/-} mice brains [134]. This can induce lipopolysaccharide production that stimulates the release of proinflammatory cytokines at the site of infection, but also in the periphery, which may then be directed to the brain [135, 136]. The presence of Chlamydia pneumoniae, has also been found post mortem in the brains of deceased AD patients [137] and other bacteria have also been implicated in contributing to AD including Propionibacterium acnes and Helicobacter pylori (reviewed in Ref. [138]). Studies have also shown using APP/PS1-21 AD mice that polymicrobial activation can increase fibrillar Aß plaque formation in the hippocampus that activates astrocytes contributing to increased brainspecific inflammation [135]. Combined these studies support the hypothesis that similarly to some anti-viral responses, active anti-bacterial activity may be indirectly contributing to AD via a neuroinflammatory response (reviewed in Refs. [138, 139]).

Conflicting data suggests, however, that during AD, the presence of $A\beta$ can actually be protective against invading

pathogens [140]. The anti-microbial hypothesis of $A\beta$ is supported by *in vitro* studies whereby synthetic Aß peptide treatment exhibits potent anti-microbial activity towards eight common and clinically relevant microbial pathogens [141]. This protective hypothesis was later supported using transgenic human Aß expressing cell lines where overexpression of Aß increased host cell resistance to Candida albicans infection [142]. The authors went on to show that in 5xFAD AD mice, the presence of AB helped protect against Caenorhabditis elegans nematode infection and described how Salmonella typhimurium infection in the brain can induce an accelerated A β deposition that co-localizes with the bacteria [142]. They describe how amyloid oligomerization mediates the protective capacity of $A\beta$ and the presence of microbial cells within the cerebrum accelerates Aß deposition in 5xFAD AD mice [142]. The authors, therefore, suggest that $A\beta$ may have dual protective and pathological role during infection [142].

The anti-microbial role of $A\beta$ is also supported by studies where $A\beta42$ -overexpressing cells are shown to be resistant to killing by yeast and bacteria [143]. This presents an important finding in AD development. If as suggested by Frost *et al.* that APP and A β are involved in pathogen defence, then excessive A β production could be a by-product from innate immune cell activation supporting the hypothesis that infectious pathogens could play a vital role in AD development [19]. This raises the possibility that combination therapies that target the pathogen and the associated proinflammatory cytokine response may be a useful treatment regimen for AD.

Immunotherapy

There have been many studies that have used immunotherapy as a way to specifically target $A\beta$ to prevent plaque formation for effective treatment of AD [144]. These approaches used either synthetic $A\beta$ peptides (AN1792) to stimulate anti- $A\beta$ antibodies [145, 146], or use antibodies that target various regions of $A\beta$ [144]. However, these studies have largely failed due to a lack of efficacy and safety concerns. Further direct entry of antibodies into the brain poses a clinical challenge as well as the timings required to treat an individual recruited into a trial who may already have different levels of $A\beta$ throughout the cohort [47].

There have been more recent proof of concept approaches targeting Aß using anti-IgG1 antibody therapy (Gantenerumab and Lecanemab) which have been designated 'Breakthrough Therapies' by the US FDA [147, 148]. Both antibodies, (recently through Phase III trials), using between 850 and 2000 patients, showed reduction of Aβ plaques and slowed cognitive decline. Lecanemab is intended for earlystage AD and resulted in a 27% slowing in cognitive decline with reduced plasma AB42/40 ratio and reduced amyloid in the brain [149]. Gantenerumab has now failed to reach its key secondary endpoints in Phase III clinical trials and is currently suspended [150]. Donanemab is another anti-IgG1 A β antibody that has recently come through Phase III trials with promise [151]. The results of the trial are similar to the effects seen with Lecanemab and provide a 35% slowing of cognitive decline as compared to placebo [151]. Whilst Donanemab is not yet licensed, permissions are being actively sought for approval [151]. Until recently the only approved monoclonal antibody therapy to target A β was Aducanumab that lead to a reduction in amyloid; however, the approval of this drug is still debated [152–154]. Also, as mentioned above, due to successful Phase III trials, Lecanemab has recently been licensed

for use in human AD patients [6, 155]. However, it is important to consider that it is widely accepted that for the brain to respond to treatment and thus recover, amyloid plaques need to be completely removed from the brain [150]. It is important to note that Lecanemab treatment only results in a 27% slowing in cognitive decline in AD patients with the potential risk of side effects and consider whether any anti-amyloid IgG therapies are effective at removing most if not all, amyloid from human brains.

Based on the mostly failed approaches and more recently the mild benefits of 'breakthrough' drugs that target $A\beta$, it is, therefore, appropriate to also target the immune system for effective AD treatment utilizing the approaches that target various immune pathways as described throughout.

As described above Tregs can play an important immunemodulatory role that contributes to AD pathology. Recent studies have suggested that Tregs can potentially be neuroprotective and can suppress excessive microglia activation and inflammation seen in AD brains [156, 157]. These studies took an approach that examines what impact the adoptive transfer of *in vitro* expanded Tregs has on AD pathology using AD mice. Tregs were either isolated from mouse splenocyte cultures and expanded using AB1-42 peptide to ensure antigen-specific Tregs were expanded [157], or were derived from human PBMC cultures and expanded for 24 days [156]. Expanded mouse Tregs were transferred into 3xTG AD mice where a noted reduction in microglial activation was seen with an associated amelioration of cognitive impairment [157]. Human Tregs were transferred into 5xFAD AD mice that were crossed with Rag2 deficient mice. The immune activity in these mice is, therefore, restricted to the Treg compartment and resulted in a reduction of both amyloid burden and reactive glial cells but was also shown to reduce proinflammatory cytokine release [156]. These studies show promise, however, whilst targeting excessive inflammation may be useful therapeutically, Baruch *et al.* suggest that there is already an observed overall increase in Tregs in both AD patients and mice as compared to non-AD cohorts [45]. Contrastingly, they suggest that Treg depletion is more useful therapeutically in reversing cognitive impairment in mice [45]. They also suggest that activating T-cells in this context may be more useful than increasing the prevalence of Tregs (described above (page 4). There are also studies that are now examining the potential efficacy of chimeric-antigen-receptor (CAR)-Treg-based therapies in neurological disorders [158]. CARs are manufactured receptors that can provide a T-cell with the ability to target a specific surface target and activate the T-cell simultaneously upon target recognition [159]. Due to the different studies suggesting either a pathological or protective role of Tregs in AD, CAR-Treg therapies may potentially have promise yet more studies are needed to understand the mechanisms between modifying Tregs that either protect or exacerbate disease.

Other CAR T-cell-based therapies could therefore pose an exciting prospect for treating AD based on the theory suggested by Baruch *et al* [45]. CAR T-cells are undergoing rapid development and have mostly been investigated to improve T-cell function in cancer [159]. For effective CAR T-cell therapy the antigen or target needs to be defined and as stated above this remains complicated in AD. Studies have investigated whether commonly expressed proteins associated with AD pathology, such as A β and tau, could serve as T-cell antigens, but although strong T-cell responses against these antigens are detectable, there are no differences seen between AD patients and healthy controls [160]. In this study, the authors went on to describe that they believe there is not a key role for neuronal antigen-specific T-cells in AD [160]. However, this may be due to the antigens chosen within their study which may not be specific enough to result in observable change. More in-depth GWAS studies have discovered other specific antigens that could potentially serve as CAR targets. Here, novel-specific tau peptides have been shown to aggregate within AD patient cohorts that are dominant towards a specific patient's MHC restriction [161]. These have not been investigated using CAR T-cell therapy to date, but could serve as a more specific target.

There are risks associated with activating T-cells in the brain using CAR T-cell therapy for AD. The neuronal toxicity associated with CAR T-cell therapy has been well described and can include encephalopathy, headaches, and tremors among others [159]. Depending on the CAR T-cell approach taken, those that would intend to induce activated T-cells, albeit to specific neuronal targets, could pose a serious danger to the patient. Those that intend to suppress immune activity to reduce inflammation, including CAR Tregs, could also pose risk as activated Tregs also produce perforins, granzyme B and IL-2 that could damage cells within the brain further exacerbating pathology [158]. These approaches, therefore, suggest that activating T-cells to harness their ability to differentiate or activate to various degrees could be useful for therapy. However, a more recent study went further to state that T-cells are the predominant drivers of AD pathology [35]. Here, the authors found that T-cells (notably CD8+ T-cells) are markedly increased in areas of the brain with increased tau pathology but not amyloid deposition in AD mice. They suggest that the T-cell infiltrate that contributes to tauopathy is driven by T-cell microglia interactions and that T-cell and microglia depletion blocks tau-mediated neurodegeneration [35]. This suggests that enhancing T-cell function may not be useful therapeutically but would point to therapies that reduce T-cell function may be more important.

Based on the immunotherapy approaches discussed above, it may be useful to design studies that target both A β in combination with immune modulator/s to maximize any potential success of an immunotherapeutic approach for AD. It is also important to consider the timing of these treatments and to determine whether the efficacy of treatment changes during early, mid, or late disease. These approaches may significantly impact disease progression and pose an exciting avenue for treatment moving forward.

Closing remarks

AD represents one of the greatest risks to global public health as humans now live longer. Without any highly effective therapies to appropriately treat the disease with strong efficacy, it is clear that the healthcare and economic burdens will become overwhelming. The studies that have been discussed here highlight the link between immune system-mediated inflammation in response to microbial infections with AD pathology and neurodegeneration. While therapeutic studies have mostly focused on targeting A β , these results have only elicited partial improvements and thus an effective treatment remains elusive. The highly influential role that the immune system plays in AD has been well described throughout and research surrounding this area is gathering pace. However, the key mechanisms behind these observations still require further study and indeed whether these mechanisms are influenced by an individual's increased genetic risk of disease. Whether the adaptive immune system is eliciting its effect via direct interactions with brain-resident cells altering their function to drive pathology, or whether this is due to neuroinflammation released as a result of an increased antimicrobial response, or whether it is a combination of both, has still not been comprehensively addressed. However, the data discussed here strongly highlights the significant contribution that the adaptive immune system, particularly T-cells, plays in AD pathology. Together, identifying new prognostic markers of disease, related to and derived from the immune system, will be crucial in developing novel therapies that will alleviate the symptoms of this debilitating disease and prevent neurodegeneration in AD.

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Ethical approval

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Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Office for National Statistics. *Deaths*. https://www.ons.gov.uk/ peoplepopulationandcommunity/birthsdeathsandmarriages/ deaths. (4 September 2023, date last accessed).
- Alzheimer's Society. *Dementia UK report*. https://www.alzheimers. org.uk/about-us/policy-and-influencing/dementia-scale-impactnumbers. (4 September 2023, date last accessed).
- Breijyeh Z, Karaman R. Comprehensive review on Alzheimer's disease: causes and treatment. *Molecules* 2020, 25, 5789. doi:10.3390/ molecules25245789.
- 4. Alzheimer's Society. *Symptoms of Alzheimer's disease*. https://www. alzheimers.org.uk/about-dementia/types-dementia/alzheimersdisease-symptoms.
- Perneczky R, Jessen F, Grimmer T, Levin J, Flöel A, Peters O, et al. Anti-amyloid antibody therapies in Alzheimer's disease. *Brain* 2023, 146, 842–9. doi:10.1093/brain/awad005.
- Mullard A. FDA approves second anti-amyloid antibody for Alzheimer disease. Nat Rev Drug Discov 2023, 22, 89. doi:10.1038/ d41573-023-00004-0.

- Rustenhoven J, Kipnis J. Brain borders at the central stage of neuroimmunology. *Nature* 2022, 612, 417–29. doi:10.1038/ s41586-022-05474-7.
- Trambauer J, Fukumori A, Kretner B, Steiner H. Analyzing amyloid-beta peptide modulation profiles and binding sites of gamma-secretase modulators. *Enzymology at the Membrane Interface: Intramembrane Proteases*. Vol. 584. 2017, 157–83. doi:10.1016/bs.mie.2016.10.013.
- Murphy MP, LeVine H. Alzheimer's disease and the amyloid-beta peptide. J Alzheimers Dis 2010, 19, 311–23. doi:10.3233/JAD-2010-1221.
- Chen X, Holtzman DM. Emerging roles of innate and adaptive immunity in Alzheimer's disease. *Immunity* 2022, 55, 2236–54. doi:10.1016/j.immuni.2022.10.016.
- Condello C, Yuan P, Schain A, Grutzendler J. Microglia constitute a barrier that prevents neurotoxic protofibrillar Abeta42 hotspots around plaques. *Nat Commun* 2015, 6, 6176. doi:10.1038/ ncomms7176.
- Tsai J, Grutzendler J, Duff K, Gan W-B. Fibrillar amyloid deposition leads to local synaptic abnormalities and breakage of neuronal branches. *Nat Neurosci* 2004, 7, 1181–3. doi:10.1038/nn1335.
- 13. Bloom GS. Amyloid-beta and tau: the trigger and bullet in Alzheimer disease pathogenesis. JAMA Neurol 2014, 71, 505–8.
- Busche MA, Hyman BT. Synergy between amyloid-beta and tau in Alzheimer's disease. *Nat Neurosci* 2020, 23, 1183–93. doi:10.1038/ s41593-020-0687-6.
- Cacace R, Sleegers K, Van Broeckhoven C. Molecular genetics of early-onset Alzheimer's disease revisited. *Alzheimers Dement* 2016, 12, 733–48. doi:10.1016/j.jalz.2016.01.012.
- Rubin L, Ingram LA, Resciniti NV, Ashford-Carroll B, Leith KH, Rose A, et al. Genetic risk factors for Alzheimer's disease in racial/ ethnic minority populations in the U.S.: a scoping review. *Front Public Health* 2021, 9, 784958. doi:10.3389/fpubh.2021.784958.
- Hoogmartens J, Cacace R, Van Broeckhoven C. Insight into the genetic etiology of Alzheimer's disease: a comprehensive review of the role of rare variants. *Alzheimers Dement (Amst)* 2021, 13, e12155. doi:10.1002/dad2.12155.
- Bellenguez C, Küçükali F, Jansen IE, Kleineidam L, Moreno-Grau S, Amin N, et al.; EADB. New insights into the genetic etiology of Alzheimer's disease and related dementias. *Nat Genet* 2022, 54, 412–36. doi:10.1038/s41588-022-01024-z.
- Frost GR, Jonas LA, Li YM. Friend, foe or both? Immune activity in Alzheimer's disease. *Front Aging Neurosci* 2019, 11, 337. doi:10.3389/fnagi.2019.00337.
- Canter RG, Penney J, Tsai LH. The road to restoring neural circuits for the treatment of Alzheimer's disease. *Nature* 2016, 539, 187– 96. doi:10.1038/nature20412.
- 21. Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, et al. Gene dose of apolipoprotein-E type-4 allele and the risk of Alzheimers-disease in late-onset families. *Science* 1993, 261, 921–3. doi:10.1126/science.8346443.
- Morris JC, Roe CM, Xiong C, Fagan AM, Goate AM, Holtzman DM, et al. APOE predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. *Ann Neurol* 2010, 67, 122– 31. doi:10.1002/ana.21843.
- 23. Shi Y, Yamada K, Liddelow SA, Smith ST, Zhao L, Luo W, et al.; Alzheimer's Disease Neuroimaging Initiative. ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy. *Nature* 2017, 549, 523–7. doi:10.1038/nature24016.
- 24. Yoshiyama Y, Higuchi M, Zhang B, Huang S-M, Iwata N, Saido TC, et al. Synapse loss and microglial activation precede tangles in a P301S tauopathy mouse model. *Neuron* 2007, 53, 337–51. doi:10.1016/j.neuron.2007.01.010.
- 25. Verghese PB, Castellano JM, Garai K, Wang Y, Jiang H, Shah A, et al. ApoE influences amyloid-beta (Abeta) clearance despite minimal apoE/Abeta association in physiological conditions. *Proc Natl Acad Sci U S A* 2013, 110, E1807–16.
- 26. Dai L, Shen Y. Insights into T-cell dysfunction in Alzheimer's disease. *Aging Cell* 2021, 20, e13511.

- Montagne A, Nation DA, Sagare AP, Barisano G, Sweeney MD, Chakhoyan A, et al. APOE4 leads to blood-brain barrier dysfunction predicting cognitive decline. *Nature* 2020, 581, 71–6. doi:10.1038/s41586-020-2247-3.
- Da Mesquita S, Louveau A, Vaccari A, Smirnov I, Cornelison RC, Kingsmore KM, et al. Functional aspects of meningeal lymphatics in ageing and Alzheimer's disease. *Nature* 2018, 560, 185–91. doi:10.1038/s41586-018-0368-8.
- 29. Herisson F, Frodermann V, Courties G, Rohde D, Sun Y, Vandoorne K, et al. Direct vascular channels connect skull bone marrow and the brain surface enabling myeloid cell migration. *Nat Neurosci* 2018, 21, 1209–17. doi:10.1038/s41593-018-0213-2.
- 30. Shaked I, Porat Z, Gersner R, Kipnis J, Schwartz M. Early activation of microglia as antigen-presenting cells correlates with T cell-mediated protection and repair of the injured central nervous system. J Neuroimmunol 2004, 146, 84–93. doi:10.1016/j.jneuroim.2003.10.049.
- Machhi J, Yeapuri P, Lu Y, Foster E, Chikhale R, Herskovitz J, et al. CD4+ effector T cells accelerate Alzheimer's disease in mice. *J Neuroinflammation* 2021, 18, 272. doi:10.1186/s12974-021-02308-7.
- 32. Merlini M, Kirabali T, Kulic L, Nitsch RM, Ferretti MT. Extravascular CD3+ T cells in brains of Alzheimer disease patients correlate with tau but not with amyloid pathology: an immunohistochemical study. *Neurodegener Dis* 2018, 18, 49–56. doi:10.1159/000486200.
- 33. Unger MS, Li E, Scharnagl L, Poupardin R, Altendorfer B, Mrowetz H, et al. CD8(+) T-cells infiltrate Alzheimer's disease brains and regulate neuronal- and synapse-related gene expression in APP-PS1 transgenic mice. *Brain Behav Immun* 2020, 89, 67–86. doi:10.1016/j.bbi.2020.05.070.
- 34. Franceschi C, Bonafè M, Valensin S, Olivieri F, De Luca M, Ottaviani E, et al. Inflamm-aging: an evolutionary perspective on immunosenescence. Ann N Y Acad Sci 2000, 908, 244–54. doi:10.1111/j.1749-6632.2000.tb06651.x.
- 35. Chen X, Firulyova M, Manis M, Herz J, Smirnov I, Aladyeva E, et al. Microglia-mediated T cell infiltration drives neurodegeneration in tauopathy. *Nature* 2023, 615, 668–77. doi:10.1038/s41586-023-05788-0.
- 36. Gate D, Saligrama N, Leventhal O, Yang AC, Unger MS, Middeldorp J, et al. Clonally expanded CD8 T cells patrol the cerebrospinal fluid in Alzheimer's disease. *Nature* 2020, 577, 399–404. doi:10.1038/s41586-019-1895-7.
- 37. Marsh SE, Abud EM, Lakatos A, Karimzadeh A, Yeung ST, Davtyan H, et al. The adaptive immune system restrains Alzheimer's disease pathogenesis by modulating microglial function. *Proc Natl Acad Sci U S A* 2016, 113, E1316–25. doi:10.1073/pnas.1525466113.
- 38. Spani C, Suter T, Derungs R, Ferretti MT, Welt T, Wirth F, et al. Reduced beta-amyloid pathology in an APP transgenic mouse model of Alzheimer's disease lacking functional B and T cells. *Acta Neuropathol Commun* 2015, 3, 71. doi:10.1186/s40478-015-0251-x.
- 39. Browne TC, McQuillan K, McManus RM, O'Reilly J-A, Mills KHG, Lynch MA. IFN-gamma production by amyloid beta-specific Th1 cells promotes microglial activation and increases plaque burden in a mouse model of Alzheimer's disease. *J Immunol* 2013, 190, 2241–51. doi:10.4049/jimmunol.1200947.
- Mittal K, Eremenko E, Berner O, Elyahu Y, Strominger I, Apelblat D, et al. CD4 T cells induce a subset of MHCII-expressing microglia that attenuates alzheimer pathology. *iScience* 2019, 16, 298–311. doi:10.1016/j.isci.2019.05.039.
- 41. Zhang J, Ke K-F, Liu Z, Qiu Y-H, Peng Y-P. Th17 cell-mediated neuroinflammation is involved in neurodegeneration of abeta1-42-induced Alzheimer's disease model rats. *PLoS One* 2013, 8, e75786. doi:10.1371/journal.pone.0075786.
- 42. Guo L, Li X, Gould T, Wang Z-Y, Cao W. T cell aging and Alzheimer's disease. *Front Immunol* 2023, 14, 1154699. doi:10.3389/fimmu.2023.1154699.
- 43. Zhao H, Liao X, Kang Y. Tregs: where we are and what comes next?. *Front Immunol* 2017, 8, 1578. doi:10.3389/fimmu.2017.01578.

- 44. Saresella M, Calabrese E, Marventano I, Piancone F, Gatti A, Calvo MG, et al. PD1 negative and PD1 positive CD4+ T regulatory cells in mild cognitive impairment and Alzheimer's disease. J Alzheimers Dis 2010, 21, 927–38. doi:10.3233/JAD-2010-091696.
- 45. Baruch K, Rosenzweig N, Kertser A, Deczkowska A, Sharif AM, Spinrad A, et al. Breaking immune tolerance by targeting Foxp3(+) regulatory T cells mitigates Alzheimer's disease pathology. *Nat Commun* 2015, 6, 7967. doi:10.1038/ncomms8967.
- 46. Baruch K, Deczkowska A, Rosenzweig N, Tsitsou-Kampeli A, Sharif AM, Matcovitch-Natan O, et al. PD-1 immune checkpoint blockade reduces pathology and improves memory in mouse models of Alzheimer's disease. *Nat Med* 2016, 22, 135–7. doi:10.1038/nm.4022.
- 47. Jevtic S, Sengar AS, Salter MW, McLaurin JA. The role of the immune system in Alzheimer disease: etiology and treatment. *Ageing Res Rev* 2017, 40, 84–94. doi:10.1016/j.arr.2017.08.005.
- Nystrom SN, Bourges D, Garry S, Ross EM, van Driel IR, Gleeson PA. Transient Treg-cell depletion in adult mice results in persistent self-reactive CD4(+) T-cell responses. *Eur J Immunol* 2014, 44, 3621–31. doi:10.1002/eji.201344432.
- 49. Guillot-Sestier MV, Doty KR, Gate D, Rodriguez J, Leung BP, Rezai-Zadeh K, et al. II10 deficiency rebalances innate immunity to mitigate Alzheimer-like pathology. *Neuron* 2015, 85, 534–48. doi:10.1016/j.neuron.2014.12.068.
- Rasquinha MT, Sur M, Lasrado N, Reddy J. IL-10 as a Th2 cytokine: differences between mice and humans. *J Immunol* 2021, 207, 2205–15. doi:10.4049/jimmunol.2100565.
- 51. Cao C, Arendash GW, Dickson A, Mamcarz MB, Lin X, Ethell DW. Abeta-specific Th2 cells provide cognitive and pathological benefits to Alzheimer's mice without infiltrating the CNS. *Neurobiol Dis* 2009, 34, 63–70. doi:10.1016/j.nbd.2008.12.015.
- Krishnaswamy JK, Alsén S, Yrlid U, Eisenbarth SC, Williams A. Determination of T follicular helper cell fate by dendritic cells. *Front Immunol* 2018, 9, 2169. doi:10.3389/fimmu.2018.02169.
- 53. Agrawal S, Abud EM, Snigdha S, Agrawal A. IgM response against amyloid-beta in aging: a potential peripheral protective mechanism. *Alzheimers Res Ther* 2018, 10, 81. doi:10.1186/s13195-018-0412-9.
- Baulch JE, Acharya MM, Agrawal S, Apodaca LA, Monteiro C, Agrawal A. Immune and inflammatory determinants underlying Alzheimer's disease pathology. *J Neuroimmune Pharmacol* 2020, 15, 852–62. doi:10.1007/s11481-020-09908-9.
- Spolski R, Leonard WJ. Interleukin-21: a double-edged sword with therapeutic potential. *Nat Rev Drug Discov* 2014, 13, 379–95. doi:10.1038/nrd4296.
- 56. Somers V, Dunn-Walters DK, van der Burg M, Fraussen J. Editorial: new insights into b cell subsets in health and disease. Front Immunol 2022, 13, 854889. doi:10.3389/fimmu.2022.854889.
- 57. Kim K, Wang X, Ragonnaud E, Bodogai M, Illouz T, DeLuca M, et al. Therapeutic B-cell depletion reverses progression of Alzheimer's disease. *Nat Commun* 2021, 12, 2185. doi:10.1038/s41467-021-22479-4.
- Pierce S, Geanes ES, Bradley T. Targeting natural killer cells for improved immunity and control of the adaptive immune response. *Front Cell Infect Microbiol* 2020, 10, 231. doi:10.3389/ fcimb.2020.00231.
- Zhang Y, Fung ITH, Sankar P, Chen X, Robison LS, Ye L, et al. Depletion of NK cells improves cognitive function in the Alzheimer disease mouse model. *J Immunol* 2020, 205, 502–10. doi:10.4049/ jimmunol.2000037.
- 60. Qi C, Liu F, Zhang W, Han Y, Zhang N, Liu Q, et al. Alzheimer's disease alters the transcriptomic profile of natural killer cells at single-cell resolution. *Front Immunol* 2022, 13, 1004885. doi:10.3389/fmmu.2022.1004885.
- 61. Aliseychik M, Patrikeev A, Gusev F, Grigorenko A, Andreeva T, Biragyn A, et al. Dissection of the human T-cell receptor gamma gene repertoire in the brain and peripheral blood identifies age- and Alzheimer's disease-associated clonotype profiles. *Front Immunol* 2020, 11, 12. doi:10.3389/fimmu.2020.00012.

- 62. Priya R, Brutkiewicz RR. Brain astrocytes and microglia express functional MR1 molecules that present microbial antigens to mucosal-associated invariant T (MAIT) cells. J Neuroimmunol 2020, 349, 577428. doi:10.1016/j.jneuroim.2020.577428.
- 63. Bonacina F, Coe D, Wang G, Longhi MP, Baragetti A, Moregola A, et al. Myeloid apolipoprotein E controls dendritic cell antigen presentation and T cell activation. *Nat Commun* 2018, 9, 3083. doi:10.1038/s41467-018-05322-1.
- 64. Wei J, Zheng M, Liang P, Wei Y, Yin X, Tang Y, et al. Apolipoprotein E and its mimetic peptide suppress Th1 and Th17 responses in experimental autoimmune encephalomyelitis. *Neurobiol Dis* 2013, 56, 59–65. doi:10.1016/j.nbd.2013.04.009.
- 65. Kelly ME, Clay MA, Mistry MJ, Hsieh-Li HM, Harmony JA. Apolipoprotein E inhibition of proliferation of mitogen-activated T lymphocytes: production of interleukin 2 with reduced biological activity. *Cell Immunol* 1994, 159, 124–39. doi:10.1006/ cimm.1994.1302.
- Macy M, Okano Y, Cardin AD, Avila EM, Harmony JA. Suppression of lymphocyte activation by plasma lipoproteins. *Cancer Res* 1983, 43(5 Suppl):2496s–502s.
- Domingues C, da Cruz e Silva OAB, Henriques AG. Impact of cytokines and chemokines on Alzheimer's disease neuropathological hallmarks. *Curr Alzheimer Res* 2017, 14, 870–82.
- Zheng C, Zhou XW, Wang JZ. The dual roles of cytokines in Alzheimer's disease: update on interleukins, TNF-alpha, TGF-beta and IFN-gamma. *Transl Neurodegener* 2016, 5, 7. doi:10.1186/ s40035-016-0054-4.
- 69. De Schepper S, Ge JZ, Crowley G, Ferreira LSS, Garceau D, Toomey CE, et al. Perivascular cells induce microglial phagocytic states and synaptic engulfment via SPP1 in mouse models of Alzheimer's disease. *Nat Neurosci* 2023, 26, 406–15. doi:10.1038/s41593-023-01257-z.
- Hur EM, Youssef S, Haws ME, Zhang SY, Sobel RA, Steinman L. Osteopontin-induced relapse and progression of autoimmune brain disease through enhanced survival of activated T cells. *Nat Immunol* 2007, 8, 74–83. doi:10.1038/ni1415.
- Sanford SAI, McEwan WA. Type-I interferons in Alzheimer's disease and other tauopathies. *Front Cell Neurosci* 2022, 16, 949340. doi:10.3389/fncel.2022.949340.
- McNab F, Mayer-Barber K, Sher A, Wack A, O'Garra A. Type I interferons in infectious disease. *Nat Rev Immunol* 2015, 15, 87– 103. doi:10.1038/nri3787.
- Taylor JM, Minter MR, Newman AG, Zhang M, Adlard PA, Crack PJ. Type-1 interferon signaling mediates neuro-inflammatory events in models of Alzheimer's disease. *Neurobiol Aging* 2014, 35, 1012– 23. doi:10.1016/j.neurobiolaging.2013.10.089.
- 74. Roy ER, Wang B, Wan Y-W, Chiu G, Cole A, Yin Z, et al. Type I interferon response drives neuroinflammation and synapse loss in Alzheimer disease. *J Clin Invest* 2020, 130, 1912–30. doi:10.1172/ JCI133737.
- 75. Roy ER, Chiu G, Li S, Propson NE, Kanchi R, Wang B, et al. Concerted type I interferon signaling in microglia and neural cells promotes memory impairment associated with amyloid beta plaques. *Immunity* 2022, 55, 879–894.e6. doi:10.1016/j. immuni.2022.03.018.
- Moore Z, Mobilio F, Walker FR, Taylor JM, Crack PJ. Abrogation of type-I interferon signalling alters the microglial response to Abeta(1-42). *Sci Rep* 2020, 10, 3153. doi:10.1038/s41598-020-59917-0.
- 77. Rexach JE, Polioudakis D, Yin A, Swarup V, Chang TS, Nguyen T, et al. Tau pathology drives dementia risk-associated gene networks toward chronic inflammatory states and immunosuppression. *Cell Rep* 2020, 33, 108398. doi:10.1016/j.celrep.2020.108398.
- 78. Salih DA, Bayram S, Guelfi S, Reynolds RH, Shoai M, Ryten M, et al. Genetic variability in response to amyloid beta deposition influences Alzheimer's disease risk. *Brain Commun* 2019, 1, fcz022. doi:10.1093/braincomms/fcz022.
- 79. Hur JY, Frost GR, Wu X, Crump C, Pan SJ, Wong E, et al. The innate immunity protein IFITM3 modulates gamma-secretase

in Alzheimer's disease. Nature 2020, 586, 735-40. doi:10.1038/ s41586-020-2681-2.

- Udeochu JC, Amin S, Huang Y, Fan Li, Torres ERS, Carling GK, et al. Tau activation of microglial cGAS-IFN reduces MEF2Cmediated cognitive resilience. *Nat Neurosci* 2023, 26, 737–50. doi:10.1038/s41593-023-01315-6.
- Festa BP, Siddiqi FH, Jimenez-Sanchez M, Won H, Rob M, Djajadikerta A, et al. Microglial-to-neuronal CCR5 signaling regulates autophagy in neurodegeneration. *Neuron* 2023, 111, 2021–2037.e12. doi:10.1016/j.neuron.2023.04.006.
- 82. Tang Y, Chaillon A, Gianella S, Wong LM, Li D, Simermeyer TL, et al. Brain microglia serve as a persistent HIV reservoir despite durable antiretroviral therapy. *J Clin Invest* 2023, 133, e167417. doi:10.1172/JCI167417.
- Levine KS, Leonard HL, Blauwendraat C, Iwaki H, Johnson N, Bandres-Ciga S, et al. Virus exposure and neurodegenerative disease risk across national biobanks. *Neuron* 2023, 111, 1086–1093. e2. doi:10.1016/j.neuron.2022.12.029.
- 84. Itzhaki RF. Overwhelming evidence for a major role for herpes simplex virus type 1 (HSV1) in Alzheimer's disease (AD): underwhelming evidence against. *Vaccines (Basel)* 2021, 9, 679. doi:10.3390/vaccines9060679.
- Steardo L, Steardo L, Zorec R, Verkhratsky A. Neuroinfection may contribute to pathophysiology and clinical manifestations of COVID-19. *Acta Physiol (Oxf)* 2020, 229, e13473. doi:10.1111/ apha.13473.
- 86. van den Pol AN. Viral infections in the developing and mature brain. *Trends Neurosci* 2006, 29, 398–406. doi:10.1016/j. tins.2006.06.002.
- 87. Readhead B, Haure-Mirande J-V, Funk CC, Richards MA, Shannon P, Haroutunian V, et al. Multiscale analysis of independent Alzheimer's cohorts finds disruption of molecular, genetic, and clinical networks by human herpesvirus. *Neuron* 2018, 99, 64–82.e7. doi:10.1016/j.neuron.2018.05.023.
- Ball MJ, Nuttall K, Warren KG. Neuronal and lymphocytic populations in human trigeminal ganglia: implications for ageing and for latent virus. *Neuropathol Appl Neurobiol* 1982, 8, 177–87. doi:10.1111/j.1365-2990.1982.tb00273.x.
- Wozniak MA, Mee AP, Itzhaki RF. Herpes simplex virus type 1 DNA is located within Alzheimer's disease amyloid plaques. J Pathol 2009, 217, 131–8. doi:10.1002/path.2449.
- 90. Jamieson GA, Maitland NJ, Wilcock GK, Yates CM, Itzhaki RF. Herpes simplex virus type 1 DNA is present in specific regions of brain from aged people with and without senile dementia of the Alzheimer type. J Pathol 1992, 167, 365–8. doi:10.1002/ path.1711670403.
- Wozniak MA, Frost AL, Itzhaki RF. Alzheimer's disease-specific tau phosphorylation is induced by herpes simplex virus type 1. J Alzheimers Dis 2009, 16, 341–50. doi:10.3233/JAD-2009-0963.
- Zambrano A, Solis L, Salvadores N, Cortés M, Lerchundi R, Otth C. Neuronal cytoskeletal dynamic modification and neurodegeneration induced by infection with herpes simplex virus type 1. J Alzheimers Dis 2008, 14, 259–69. doi:10.3233/jad-2008-14301.
- Lovheim H, Gilthorpe J, Adolfsson R, Nilsson L-G, Elgh F. Reactivated herpes simplex infection increases the risk of Alzheimer's disease. *Alzheimers Dement* 2015, 11, 593–9. doi:10.1016/j.jalz.2014.04.522.
- 94. Cairns DM, Itzhaki RF, Kaplan DL. Potential involvement of varicella zoster virus in Alzheimer's disease via reactivation of quiescent herpes simplex virus type 1. J Alzheimers Dis 2022, 88, 1189–200. doi:10.3233/JAD-220287.
- 95. Lopatko Lindman K, Hemmingsson E-S, Weidung B, Brännström J, Josefsson M, Olsson J, et al. Herpesvirus infections, antiviral treatment, and the risk of dementia-a registry-based cohort study in Sweden. *Alzheimers Dement (N Y)* 2021, 7, e12119. doi:10.1002/trc2.12119.
- 96. Barnes LL, Capuano AW, Aiello AE, Turner AD, Yolken RH, Torrey EF, et al. Cytomegalovirus infection and risk of Alzheimer

disease in older black and white individuals. J Infect Dis 2015, 211, 230–7. doi:10.1093/infdis/jiu437.

- Lurain NS, Hanson BA, Martinson J, Leurgans SE, Landay AL, Bennett DA, et al. Virological and immunological characteristics of human cytomegalovirus infection associated with Alzheimer disease. J Infect Dis 2013, 208, 564–72. doi:10.1093/infdis/jit210.
- Westman G, Berglund D, Widén J, Ingelsson M, Korsgren O, Lannfelt L, et al. Increased inflammatory response in cytomegalovirus seropositive patients with Alzheimer's disease. *PLoS One* 2014, 9, e96779. doi:10.1371/journal.pone.0096779.
- 99. Lovheim H, Olsson J, Weidung B, Johansson A, Eriksson S, Hallmans G, et al. Interaction between cytomegalovirus and herpes simplex virus type 1 associated with the risk of Alzheimer's disease development. J Alzheimers Dis 2018, 61, 939–45. doi:10.3233/JAD-161305.
- Clement M, Humphreys IR. Cytokine-mediated induction and regulation of tissue damage during cytomegalovirus infection. *Front Immunol* 2019, 10, 78. doi:10.3389/fimmu.2019.00078.
- 101. Christ W, Kapell S, Mermelekas G, Evertsson B, Sork H, Bazaz S, et al. SARS-CoV-2 and HSV-1 induce amyloid aggregation in human CSF. *bioRxiv* 2022, 2022.09.15.508120.
- 102. Tiwari D, Singh VK, Baral B, Pathak DK, Jayabalan J, Kumar R, et al. Indication of neurodegenerative cascade initiation by amyloidlike aggregate-forming EBV proteins and peptide in Alzheimer's disease. ACS Chem Neurosci 2021, 12, 3957–67. doi:10.1021/ acschemneuro.1c00584.
- 103. Zhang N, Zuo Y, Jiang L, Peng Y, Huang X, Zuo L. Epstein-Barr virus and neurological diseases. *Front Mol Biosci* 2021, 8, 816098. doi:10.3389/fmolb.2021.816098.
- 104. Bjornevik K, Cortese M, Healy Brian C, Kuhle J, Mina MJ, Leng Y, et al. Longitudinal analysis reveals high prevalence of Epstein-Barr virus associated with multiple sclerosis. *Science* 2022, 375, 296–301. doi:10.1126/science.abj8222.
- 105. Tiwari D, Mittal N, Jha HC. Unraveling the links between neurodegeneration and Epstein-Barr virus-mediated cell cycle dysregulation. *Curr Res Neurobiol* 2022, 3, 100046. doi:10.1016/j.crneur.2022.100046.
- 106. Carbone I, Lazzarotto T, Ianni M, Porcellini E, Forti P, Masliah E, et al. Herpes virus in Alzheimer's disease: relation to progression of the disease. *Neurobiol Aging* 2014, 35, 122–9. doi:10.1016/j. neurobiolaging.2013.06.024.
- 107. Morgello S. Coronaviruses and the central nervous system. J Neurovirol 2020, 26, 459–73. doi:10.1007/s13365-020-00868-7.
- 108. Venkataramani V, Winkler F. Cognitive deficits in long covid-19. N Engl J Med 2022, 387, 1813–5. doi:10.1056/ NEJMcibr2210069.
- 109. Fernandez-Castaneda A, Lu P, Geraghty AC, Song E, Lee M-H, Wood J, et al. Mild respiratory COVID can cause multi-lineage neural cell and myelin dysregulation. *Cell* 2022, 185, 2452–2468 e16.
- 110. Wang L, Davis PB, Volkow ND, Berger NA, Kaelber DC, Xu R. Association of COVID-19 with new-onset Alzheimer's disease. J Alzheimers Dis 2022, 89, 411–4. doi:10.3233/JAD-220717.
- 111. Kuo CL, Pilling LC, Atkins JL, Masoli JAH, Delgado J, Kuchel GA, et al. APOE e4 genotype predicts severe COVID-19 in the UK Biobank Community Cohort. J Gerontol A Biol Sci Med Sci 2020, 75, 2231–2. doi:10.1093/gerona/glaa131.
- 112. Wang C, Zhang M, Garcia G, Tian E, Cui Q, Chen X, et al. ApoEisoform-dependent SARS-CoV-2 neurotropism and cellular response. *Cell Stem Cell* 2021, 28, 331–342.e5. doi:10.1016/j. stem.2020.12.018.
- 113. Hsu JT, Tien C-F, Yu G-Y, Shen S, Lee Y-H, Hsu P-C, et al. The effects of abeta(1-42) binding to the SARS-CoV-2 spike protein S1 subunit and angiotensin-converting enzyme 2. *Int J Mol Sci* 2021, 22, 8226. doi:10.3390/ijms22158226.
- 114. Rudnicka-Drozak E, Drożak P, Mizerski G, Zaborowski T, Ślusarska B, Nowicki G, Drożak M. Links between COVID-19 and Alzheimer's disease-what do we already know?. *Int J Environ Res Public Health* 2023, 20, 2146. doi:10.3390/ijerph20032146.

- 115. Ding Q, Shults NV, Gychka SG, Harris BT, Suzuki YJ. Protein expression of angiotensin-converting enzyme 2 (ACE2) is upregulated in brains with Alzheimer's disease. *Int J Mol Sci* 2021, 22, 1687. doi:10.3390/ijms22041687.
- 116. Zou K, Liu J, Watanabe A, Hiraga S, Liu S, Tanabe C, et al. Abeta43 is the earliest-depositing Abeta species in APP transgenic mouse brain and is converted to Abeta41 by two active domains of ACE. *Am J Pathol* 2013, 182, 2322–31. doi:10.1016/j. ajpath.2013.01.053.
- 117. Bassendine MF, Taylor-Robinson SD, Fertleman M, Khan M, Neely D. Is Alzheimer's disease a liver disease of the brain?. J Alzheimers Dis 2020, 75, 1–14. doi:10.3233/JAD-190848.
- 118. Lin HC, Xirasagar S, Lee H-C, Huang C-C, Chen C-H. Association of Alzhemier's disease with hepatitis C among patients with bipolar disorder. *PLoS One* 2017, 12, e0179312. doi:10.1371/ journal.pone.0179312.
- 119. Nho K, Kueider-Paisley A, Ahmad S, MahmoudianDehkordi S, Arnold M, Risacher Shannon L, et al.; Alzheimer's Disease Neuroimaging Initiative and the Alzheimer Disease Metabolomics Consortium. Association of altered liver enzymes with Alzheimer disease diagnosis, cognition, neuroimaging measures, and cerebrospinal fluid biomarkers. JAMA Netw Open 2019, 2, e197978. doi:10.1001/jamanetworkopen.2019.7978.
- 120. Lee JY, Acosta EG, Stoeck IK, Long G, Hiet M-S, Mueller B, et al. Apolipoprotein E likely contributes to a maturation step of infectious hepatitis C virus particles and interacts with viral envelope glycoproteins. *J Virol* 2014, 88, 12422–37. doi:10.1128/ JVI.01660-14.
- 121. Chakradhar S. A tale of two diseases: aging HIV patients inspire a closer look at Alzheimer's disease. *Nat Med* 2018, 24, 376–7. doi:10.1038/nm0418-376.
- 122. Yu X, Raji MA, Giordano TP, Berenson AB, Baillargeon J, Kuo Y-F. Prevalence of Alzheimer's disease and related dementias among older US Medicare beneficiaries with and without HIV: a successive cross-sectional study. *Lancet Health Longevit* 2022, 3, S6. doi:10.1016/s2666-7568(22)00067-8.
- 123. Canet G, Dias C, Gabelle A, Simonin Y, Gosselet F, Marchi N, et al. HIV neuroinfection and Alzheimer's disease: similarities and potential links?. *Front Cell Neurosci* 2018, 12, 307. doi:10.3389/ fncel.2018.00307.
- 124. Kim J, Yoon JH, Kim YS. HIV-1 tat interacts with and regulates the localization and processing of amyloid precursor protein. *PLoS One* 2013, 8, e77972. doi:10.1371/journal.pone.0077972.
- 125. Chai Q, Jovasevic V, Malikov V, Sabo Y, Morham S, Walsh D, et al. HIV-1 counteracts an innate restriction by amyloid precursor protein resulting in neurodegeneration. *Nat Commun* 2017, 8, 1522. doi:10.1038/s41467-017-01795-8.
- 126. Clifford DB, Fagan AM, Holtzman DM, Morris J C, Teshome M, Shah AR, et al. CSF biomarkers of Alzheimer disease in HIV-associated neurologic disease. *Neurology* 2009, 73, 1982–7. doi:10.1212/WNL.0b013e3181c5b445.
- 127. Giunta B, Ehrhart J, Obregon DF, Lam L, Le L, Jin JJ, et al. Antiretroviral medications disrupt microglial phagocytosis of betaamyloid and increase its production by neurons: implications for HIV-associated neurocognitive disorders. *Mol Brain* 2011, 4, 23. doi:10.1186/1756-6606-4-23.
- 128. Green DA, Masliah E, Vinters HV, Beizai P, Moore DJ, Achim CL. Brain deposition of beta-amyloid is a common pathologic feature in HIV positive patients. *AIDS* 2005, 19, 407–11. doi:10.1097/01. aids.0000161770.06158.5c.
- 129. Achim CL, Adame A, Dumaop W, Everall IP, Masliah E; Neurobehavioral Research Center. Increased accumulation of intraneuronal amyloid beta in HIV-infected patients. J Neuroimmune Pharmacol 2009, 4, 190–9. doi:10.1007/s11481-009-9152-8.
- 130. Zhang Y, Kwon D, Esmaeili-Firidouni P, Pfefferbaum A, Sullivan EV, Javitz H, et al. Extracting patterns of morphometry distinguishing HIV associated neurodegeneration from mild cognitive impairment via group cardinality constrained classification. *Hum Brain Mapp* 2016, 37, 4523–38. doi:10.1002/hbm.23326.

- 131. Ances BM, Vaida F, Yeh MJ, Liang CL, Buxton RB, Letendre S, et al. HIV infection and aging independently affect brain function as measured by functional magnetic resonance imaging. *J Infect Dis* 2010, 201, 336–40. doi:10.1086/649899.
- 132. Wiemken TL, Salas J, Morley JE, Hoft DF, Jacobs C, Scherrer JF. Comparison of rates of dementia among older adult recipients of two, one, or no vaccinations. J Am Geriatr Soc 2022, 70, 1157– 68. doi:10.1111/jgs.17606.
- 133. Eyting M, Xie M, Heß S, Heß S, Geldsetzer P. Causal evidence that herpes zoster vaccination prevents a proportion of dementia cases. *medRxiv* 2023. doi:10.1101/2023.05.23.23290253.
- 134. Dominy SS, Lynch C, Ermini F, Benedyk M, Marczyk A, Konradi A, et al. *Porphyromonas gingivalis* in Alzheimer's disease brains: evidence for disease causation and treatment with small-molecule inhibitors. *Sci Adv* 2019, 5, eaau3333. doi:10.1126/sciadv. aau3333.
- 135. Basak JM, Ferreiro A, Cohen LS, Sheehan PW, Nadarajah CJ, Kanan MF, et al. Bacterial sepsis increases hippocampal fibrillar amyloid plaque load and neuroinflammation in a mouse model of Alzheimer's disease. *Neurobiol Dis* 2021, 152, 105292. doi:10.1016/j.nbd.2021.105292.
- 136. Zhao J, Bi W, Xiao S, Lan X, Cheng X, Zhang J, et al. Neuroinflammation induced by lipopolysaccharide causes cognitive impairment in mice. *Sci Rep* 2019, 9, 5790. doi:10.1038/ s41598-019-42286-8.
- 137. Gerard HC, Dreses-Werringloer U, Wildt KS, Deka S, Oszust C, Balin BJ, et al. Chlamydophila (Chlamydia) pneumoniae in the Alzheimer's brain. *FEMS Immunol Med Microbiol* 2006, 48, 355– 66. doi:10.1111/j.1574-695X.2006.00154.x.
- 138. Piekut T, Hurła M, Banaszek N, Szejn P, Dorszewska J, Kozubski W, et al. Infectious agents and Alzheimer's disease. J Integr Neurosci 2022, 21, 73. doi:10.31083/j.jin2102073.
- 139. Lotz SK, Blackhurst BM, Reagin KL, Funk KE. Microbial infections are a risk factor for neurodegenerative diseases. *Front Cell Neurosci* 2021, 15, 691136. doi:10.3389/fncel.2021.691136.
- 140. Moir RD, Lathe R, Tanzi RE. The antimicrobial protection hypothesis of Alzheimer's disease. *Alzheimers Dement* 2018, 14, 1602–14. doi:10.1016/j.jalz.2018.06.3040.
- 141. Soscia SJ, Kirby JE, Washicosky KJ, Tucker SM, Ingelsson M, Hyman B, et al. The Alzheimer's disease-associated amyloid betaprotein is an antimicrobial peptide. *PLoS One* 2010, *5*, e9505. doi:10.1371/journal.pone.0009505.
- 142. Kumar DK, Choi SH, Washicosky KJ, Eimer WA, Tucker S, Ghofrani J, et al. Amyloid-beta peptide protects against microbial infection in mouse and worm models of Alzheimer's disease. *Sci Transl Med* 2016, 8, 340ra72. doi:10.1126/scitranslmed.aaf1059.
- 143. Eimer WA, Vijaya Kumar DK, Navalpur Shanmugam NK, Rodriguez AS, Mitchell T, Washicosky KJ, et al. Alzheimer's disease-associated beta-amyloid is rapidly seeded by herpesviridae to protect against brain infection. *Neuron* 2018, 100, 1527–32. doi:10.1016/j.neuron.2018.11.043.
- 144. Song C, Shi J, Zhang P, Zhang Y, Xu J, Zhao L, et al. Immunotherapy for Alzheimer's disease: targeting beta-amyloid and beyond. *Transl Neurodegener* 2022, 11, 18. doi:10.1186/s40035-022-00292-3.
- 145. Gilman S, Koller M, Black RS, Jenkins L, Griffith SG, Fox NC, et al.; AN1792(QS-21)-201 Study Team. Clinical effects of Abeta immunization (AN1792) in patients with AD in an interrupted trial. *Neurology* 2005, 64, 1553–62. doi:10.1212/01. WNL.0000159740.16984.3C.
- 146. Cao J, Hou J, Ping J, Cai D. Advances in developing novel therapeutic strategies for Alzheimer's disease. *Mol Neurodegener* 2018, 13, 64. doi:10.1186/s13024-018-0299-8.
- 147. Decourt B, Boumelhem F, Pope ED, Shi J, Mari Z, Sabbagh MN. Critical appraisal of amyloid lowering agents in AD. *Curr Neurol Neurosci Rep* 2021, 21, 39. doi:10.1007/s11910-021-01125-y.
- 148. Genentech. Genentech's Anti-Amyloid Beta Antibody Gantenerumab Granted FDA Breakthrough Therapy Designation in Alzheimer's Disease. 2021 https://www. gene.com/media/press-releases/14931/2021-10-08/

genentechs-anti-amyloid-beta-antibody-ga (4 September 2023, date last accessed).

- 149. van Dyck CH, Swanson CJ, Aisen P, Bateman RJ, Chen C, Gee M, et al. Lecanemab in early Alzheimer's disease. N Engl J Med 2023, 388, 9–21. doi:10.1056/NEJMoa2212948.
- 150. ALZFORUM. Gantenerumab Mystery: How Did It Lose Potency in Phase 3? 2022. https://www.alzforum.org/news/conferencecoverage/gantenerumab-mystery-how-did-it-lose-potencyphase-3 (4 September 2023, date last accessed).
- 151. Lilly, E. Lilly's Donanemab Significantly Slowed Cognitive and Functional Decline in Phase 3 Study of Early Alzheimer's Disease. 2023. https://investor.lilly.com/news-releases/news-release-details/ lillys-donanemab-significantly-slowed-cognitive-and-functional (4 September 2023, date last accessed).
- 152. Bateman RJ, Cummings J, Schobel S, Salloway S, Vellas B, Boada M, et al. Gantenerumab: an anti-amyloid monoclonal antibody with potential disease-modifying effects in early Alzheimer's disease. *Alzheimers Res Ther* 2022, 14, 178. doi:10.1186/s13195-022-01110-8.
- 153. U.S. Food and Drug. Administration, FDA Grants Accelerated Approval for Alzheimer's Drug. FDA News Release, 2021.
- 154. Lancet T. Lecanemab for Alzheimer's disease: tempering hype and hope. *Lancet* 2022, 400, 1899.
- 155. Mahase E. Alzheimer's disease: Lecanemab gets full FDA approval and black box safety warning. *BMJ* 2023, 382, p1580. doi:10.1136/bmj.p1580.

- 156. Faridar A, Vasquez M, Thome AD, Yin Z, Xuan H, Wang JH, et al. Ex vivo expanded human regulatory T cells modify neuroinflammation in a preclinical model of Alzheimer's disease. *Acta Neuropathol Commun* 2022, 10, 144. doi:10.1186/s40478-022-01447-z.
- 157. Yang H, Park S-Y, Baek H, Lee C, Chung G, Liu X, et al. Adoptive therapy with amyloid-beta specific regulatory T cells alleviates Alzheimer's disease. *Theranostics* 2022, 12, 7668–80. doi:10.7150/thno.75965.
- 158. Olson KE, Mosley RL, Gendelman HE. The potential for tregenhancing therapies in nervous system pathologies. *Clin Exp Immunol* 2023, 211, 108–21. doi:10.1093/cei/uxac084.
- 159. Rubin DB, Danish HH, Ali AB, Li K, LaRose S, Monk AD, et al. Neurological toxicities associated with chimeric antigen receptor T-cell therapy. *Brain* 2019, 142, 1334–48. doi:10.1093/brain/ awz053.
- 160. Dhanwani R, Pham J, Premlal ALR, Frazier A, Kumar A, Pero ME, et al. Corrigendum: T cell responses to neural autoantigens are similar in Alzheimer's disease patients and age-matched healthy controls. *Front Neurosci* 2020, 14, 641809. doi:10.3389/ fnins.2020.641809.
- 161. Guen YL, Luo G, Ambati A, Damotte V, Jansen I, Yu E, et al. Protective association of HLA-DRB1*04 subtypes in neurodegenerative diseases implicates acetylated tau PHF6 sequences. *Alzheimers Dement* 2022, 18, e060159.