Review article

Korean J Pediatr 2014;57(12):505-513 http://dx.doi.org/10.3345/kjp.2014.57.12.505 pISSN 1738-1061 • eISSN 2092-7258





Necrotizing enterocolitis in newborns: update in pathophysiology and newly emerging therapeutic strategies

Young Youn Choi, MD, PhD

Department of Pediatrics, Chonnam National University Hospital, Chonnam National University Medical School, Gwangju, Korea

While the survival of extremely premature infants with respiratory distress syndrome has increased due to advanced respiratory care in recent years, necrotizing enterocolitis (NEC) remains the leading cause of neonatal mortality and morbidity. NEC is more prevalent in lower gestational age and lower birth weight groups. It is characterized by various degrees of mucosal or transmural necrosis of the intestine. Its exact pathogenesis remains unclear, but prematurity, enteral feeding, bacterial products, and intestinal ischemia have all been shown to cause activation of the inflammatory cascade, which is known as the final common pathway of intestinal injury. Awareness of the risk factors for NEC; practices to reduce the risk, including early trophic feeding with breast milk and following the established feeding guidelines; and administration of probiotics have been shown to reduce the incidence of NEC. Despite advancements in the knowledge and understanding of the pathophysiology of NEC, there is currently no universal prevention measure for this serious and often fatal disease. Therefore, new potential techniques to detect early biomarkers or factors specific to intestinal inflammation, as well as further strategies to prevent the activation of the inflammatory cascade, which is important for disease progression, should be investigated.

Key words: Necrotizing enterocolitis, Extremely premature infants, Pathophysiology, Risk factor, Prevention

Corresponding author: Young Youn Choi, MD,

Department of Pediatrics, Chonnam National University Hospital, Chonnam National University Medical School, 42 Jebong Ro, Dong-gu, Gwangju

501-757, Korea Tel: +82-62-220-6642 Fax: +82-62-222-6103 E-mail: yychoi@jnu.ac.kr

Received: 25 July, 2014 Accepted: 7 October, 2014

Introduction

Necrotizing enterocolitis (NEC) was first described by Mizrahi et al.¹⁾ in 1965, and was classified into three stages based on the severity of the clinical presentation and treatment strategies by Bell et al.²⁾ in 1978. Later, Walsh and Kliegman³⁾ proposed the modified Bell's criteria, which subdivided each stage into A and B according to the clinical and radiologic signs and treatment strategies. There are inverse correlations between gestational age or birth weight and the incidence of NEC^{4,5)}, with, as demonstrated in multicenter and large population-based studies, its prevalence being 7%–11% in very low birth weight infants who weigh less than 1,500 g^{6,7)}.

Several experimental therapies have been tested in animal models and human trials⁴. Despite this progress, there has been no method to identify infants who would be ideal candidates for therapy or those who will progress to more severe stages of disease. This paper summarizes the recent advances in pathophysiology, the newly developed diagnostic approaches, and the current emerging therapeutic strategies for NEC.

Copyright © 2014 by The Korean Pediatric Society

This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Pathogenesis

Various combinations of risk factors, which may be based on genetic predisposition, have been demonstrated to increase the risk of mucosal and epithelial injury⁴.

1. Prematurity with impaired host defense

The most consistent and important risk factor is immaturity itself⁴⁻⁷⁾. Immature motility, digestion, absorption, and circulatory regulation predispose the preterm infant to an increased risk of intestinal injury⁸⁾. Especially, in these patients, the gastrointestinal (GI) host defense (physical barrier) and immunologic host defense, including intraepithelial lymphocytes and secretory IgA, are markedly impaired; and many biochemical factors such as lactoferrin, epidermal growth factor (EGF), heparin binding (HB)-EGF, transforming growth factor, insulin-like growth factor, oligosaccharides, and polyunsaturated fatty acids (PUFA), are deficient or absent 4.8).

2. Enteral feeding

Enteral alimentation, especially formula feeding, is the second most important risk factor for NEC, with over 90% of NEC cases reportedly occurring after initiation of enteral feeding⁴⁾. Since early trophic feeding is introduced in the care of extremely low birth weight infants, the onset is generally delayed for several weeks. The exact relationship between enteral feeding and NEC remains unclear, but the volume and rate of feeding advancement (overdistention may compromise splanchnic circulation), osmolarity, and substrate fermentation are considered important factors^{9,10)}. Short chain fatty acids, derived from fermentation of undigested carbohydrates by intestinal bacterial fermentation, have important effects on epithelial functions and maturation. In one animal study, overproduction/accumulation of short chain fatty acids in the proximal colon and/or distal ileum was shown to play a potential role in the pathogenesis of NEC¹¹; however, no studies testing the effects of these molecules are available at the moment. Moreover, some studies have shown that deficiency of lactase or other brush border enzymes or accumulation of bile acid may also lead to mucosal injury^{4,12)}.

3. Bacterial colonization

In utero, the intestine is sterile, and no cases of NEC have been documented. The fact that NEC typically develops before extensive colonization of the GI tract by anaerobic organisms, and that probiotic therapy may prevent the disease, support the theory that microbes also play a key role in the initiation of intestinal injury⁴. Colonization of the intestines of a healthy breast-fed infant with anaerobic species such as *Bifidobacterium* and *Lactobacillus* occurs within 1 week of age¹³. However, a hospitalized, very low birth weight infant has less species

diversity and fewer or absent anaerobes. This imbalance promotes pathologic proliferation and binding and invasion of otherwise nonpathogenic intestinal bacteria, and reduces the anti-inflammatory effects and mucosal defense from probiotic organisms¹⁴⁾.

The lack of normal maternal-infant physical interaction during feeding interferes with normal intestinal colonization, which protects against the pathologic microflora in breast-fed premature infants⁴. Furthermore, contamination or colonization of the feeding tubes has been demonstrated to contribute to the development of NEC¹⁵. The most commonly isolated bacteria are *Clostridium* sp., *Escherichia coli*, *Klebsiella* sp., *Staphylococcus epidermidis*, and *Enterobacter*⁴. In particular, the use of infant formula contaminated with *Enterobacter sakazakii* has been associated with outbreaks of NEC¹⁶.

4. Intestinal ischemia/asphyxia

Intestinal ischemia due to hypoxia-ischemia was reported as an important risk factor for NEC in early studies, especially in term and late preterm infants, as the histologic hallmark of end-stage NEC is an "infarct" characterized by full thickness coagulation (ischemic) necrosis with scanty acute inflammatory cells (neutrophils) and a predominant lymphocytic infiltration¹⁷⁾. However, intestinal ischemia is now unlikely to be considered the major perinatal event that contributes to the development of NEC. Instead, it is believed that it disturbs the balance in the microvascular tone related to the production of vascular regulators such as nitric oxide and endothelin, which likely play downstream roles in the pathogenic cascade of NEC¹⁸). Further, patent ductus arteriosus, another risk factor for NEC, results in intestinal hypoperfusion by diastolic steal phenomenon; and, in patients treated with indomethacin (a nonselective inhibitor of cyclooxygenase-1 and -2), focal intestinal necrosis or NEC may occur¹⁹⁾.

5. Other factors

Although an association between the elective transfusion of packed red blood cells and NEC has been reported, the mechanism through which transfusion might be related to alteration in the intestinal blood flow or hypoxia-ischemia remains unclear²⁰⁾. H2 receptor antagonists, which are inhibitors of gastric acid, increase the gastric pH, which may enhance pathogenic bacterial growth and increase the risk of NEC²¹⁾. As a genetic predisposition, the loss of the HB-EGF-like growth factor gene (which is essential for preservation of gut barrier function) was proposed in an animal study²²⁾. Furthermore, early use of postnatal steroids may cause intestinal perforation, independent of enteral feeding²³⁾; the use of umbilical artery or venous catheters does not seem to increase the risk of NEC²⁴⁾.

Pathophysiology

Recent advances in the understanding of NEC suggest that the epithelial barrier and innate immunity, together with the inflammatory response, play important roles in very preterm infants⁴. Intestinal mucosal stress (resulting from enteral feeding, bacterial products, or intestinal ischemia) and an inadequate host defense and repair (due to prematurity) can result in activation of the proinflammatory cascade, which is the final common pathway of intestinal injury and NEC. This cascade involves a balance between pro- and anti-inflammatory endogenous mediators, receptors, signaling pathways, and second messengers, and produces a variety of downstream effects that ultimately result in end-organ damage (Table 1, Fig. 1).

1. Epithelial barrier

In neonates, the intestinal mucosa presents a persistent equilibrium state between injury and repair. Injury to the intestinal mucosa may depend on a variety of conditions characteristic of prematurity, including hypoxia, infection, and starvation. Furthermore, microcirculatory dysfunction contributes to epithelial damage. In physiological conditions, healing of the epithelium begins immediately after an injury with mature enterocyte migration to the wounded area from healthy adjacent areas²⁵⁾. Subsequently, the proliferation of new enterocytes within the crypts of Lieberkühn completes the process of repair²⁶⁾. It has been recently suggested that NEC is associated with a marked inhibition in both enterocyte migration and proliferation, making the host uniquely susceptible to further injury, and that the loss of epithelial barrier allows translocation of pathogens from the intestinal lumen to the mucosa²⁷⁾.

2. Innate immunity: the role of Toll-like receptors

Innate immunity has been demonstrated to regulate the epithelial barrier in both experimental models and human cases of NEC. The specific Toll-like receptors (TLRs), important components of the innate immune system located on the epithelial surface, play major roles in tissue repair²⁸⁾. Among the known human TLRs, type 4 seems to play a crucial role in NEC development²⁹⁾. TLR4 may be activated by bacterial (i.e., lipopolysaccharides) or by other innate immune components. The activation of TLR4 signaling in the intestinal epithelium has been shown to inhibit enterocyte migration and to lead to enterocyte apoptosis in a mouse model, whereas inhibition of TLR4 signaling has been found to prevent NEC development and attenuate the degree of enterocyte apoptosis in mouse models and cell cultures²⁷⁻³⁰⁾.

Developing fetuses express elevated TLR4 expression until the end of the gestation, owing to the fact that TLR4 regulates proliferation and differentiation of the intestinal epithelium. The persistently elevated expression of TLR4 during intrauterine life does not increase the risk of NEC for the fetus, probably because it lives in a sterile or quasi-sterile environment. At the end of gestation, the neonate shows low expression of TLR4, and the signal remains inactivated in the presence of a normal intestinal microflora. However, the expression of TRL4 in preterm babies is very high, and when the premature intestine is colonized by pathologic microflora, TLR4 signaling may become overactivated, leading to decreased ability of the body to repair the epithelium after injury^{28,29}.

The effects of a TLR4-mediated response include gut barrier failure; bacterial translocation; intestinal inflammation; and, finally, a systemic inflammatory response through the activation of proinflammatory mediators such as IL-1, -6, -8, and -18; platelet-activating factor (PAF); leukotrienes; thromboxanes; free oxygen radicals; and nuclear factor kappa B (NFKB)⁴⁾. Accumulation of ileal bile acids causes significant injury in the small intestine and acts in concert with the TLR4 pathway¹²⁾. Intestinal integrity restitution requires intercellular connectivity, which is mediated through small channels (gap junctions), rich in connexin protein. Proinflammatory cytokines such as interferon gamma cause endocytosis of connexin (it is engulfed by the cell membrane and drawn in to the cell), thereby impairing intercellular connectivity and reducing the extent of intestinal restitution²⁸⁾.

Most premature infants do not develop NEC despite high TLR4 expression, which suggests that TLR4 signaling is weakened in the newborn intestinal epithelium by counter-regulatory

Table 1. Inflammatory mediators involved in pathophysiology of necrotizing enterocolitis

Inflammatory mediator
Proinflammatory mediators
IL, IL-1, -6, -8, -10
INFγ
PAF, ET-1
Leukotrienes
Thromboxanes
Free oxygen radicals
NF-κB
Anti-inflammatory mediators
IL-1 RA, IL-11, -12
Hsp
IkB
PAF-acetylhydrolase
Growth factors, e.g., EGF, EPO, and IGF

IL, interleukin, INF γ , interferon gamma; PAF, platelet activating factor; ET, endothelin; NF- κ B, nuclear factor-kappa B; RA, receptor antagonist; Hsp, heat shock protein; Ifactor-kappa B, inhibitory protein of NF- κ B; EGF, epidermal growth factor; EPO, erythropoietin; IGF, insulin-like growth factor.

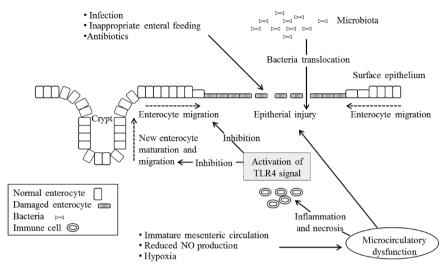


Fig. 1. Role of Toll-like receptor 4 (TLR4) in epithelial injury and repair mechanisms. Many factors related to prematurity such as infections, inappropriate enteral nutrition, antibiotics use, microcirculatory dysfunction, and hypoxia can induce epithelial injury. Hyperactivation of TLR4 enhances translocation of pathological bacteria across the epithelial barrier. Modified from Terrin et al. Biomed Res Int 2014;2014:543765²⁷).

mechanisms, including intra- and extracellular factors such as heat shock protein (Hsp), interleukin (IL) 1 receptor antagonist, IL-11 and -12, PAF acetylhydrolase, inhibitory protein of NF-κB, and EGF; thus, the development of NEC is limited. The intracellular factor Hsp70 (a major member of the Hsp family) is activated by stress, and plays a protective role in the intestine by limiting TLR4 signaling in enterocytes. TLR4 activation itself significantly increases Hsp70 expression in the enterocytes, which provides a mechanism of autoinhibition of the TLR4 signaling pathway. Reduced activity of Hsp70 or hyperactivation of TLR4 disrupts this balance and induces NEC, whereas upregulation of Hsp70 conversely leads to a reduction in TLR4 signaling³¹⁾.

Furthermore, the extracellular factor EGF may also affect TLR4 signaling. The fetus continuously swallows amniotic fluid (extremely rich in EGF), which limits the amplification of TLR4 signaling in the fetal intestinal mucosa and in cultured enterocytes exposed to bacterial products, thus markedly reducing the degree of proinflammatory cytokine release. This extracellular factor inhibits TLR4 signaling via the peroxisome proliferator-activated receptor gamma and NF-кB pathways³²⁾.

3. Gut microbiota

Although many studies have report an association between NEC and infection, and despite the fact that many pathogens may simulate features of NEC, no specific microbe has been identified as a determinant etiologic factor³³. Several findings³³⁻³⁵ such as the detection of abundant bacteria (including gram-negative pathogens) in the feces of NEC patients, a loss of gut microbial diversity and depletion of enterococcal populations in the feces before NEC development, and a correlation between the clinical

finding of pneumatosis intestinalis and the presence of clostridial species (*Clostridium butyricum* and *Clostridium paraputrificum*) suggest that NEC may not result from a single causative species but from a microbial imbalance (dysbiosis), which may favor TLR4 activation and translocation of pathogens across the epithelium.

Future approaches to prediction

Identification of the infants at increased risk of NEC may be possible through methods currently available at a few research centers. These methods use noninvasive indicators such as profiling of the fecal microbiota³⁶⁾ and identification of the expression of inflammatory proteins from the buccal epithelium³⁷⁾. In addition, oxidative stress determined by measuring concentrations of nonprotein bound iron, advanced oxidation protein products, and total hydroxides in the cord blood has been reported to be a useful predictor of NEC in high-risk infants³⁸⁾. However, additional validation is required. Recently, Ng et al.³⁹⁾ reported that the levels of three intestinal biomarkers, namely liver-fatty acid binding protein (L), intestinal-fatty acid binding protein (I), and trefoil factor 3 (T), as well as the combined value of the three (LIT score), were significantly higher in patients with NEC than in those with septicemia or control groups. Furthermore, the expression of these biomarkers, including the LIT score, was also found to be higher in NEC nonsurvivors than in the survivors.

Newly emerging therapeutic strategies

There are many evidence-based approaches to prevent NEC (Table 2)^{40,41)}. These include withholding enteral feedings; using enteral antibiotics; feeding the infant with the mother's expressed breast milk; administering probiotic agents, prebiotic agents, or both; and administering various growth factors, anticytokine agents, and glucocorticoids. Recent advances in understanding the pathophysiology of NEC suggest that future treatments may involve immunological approaches such as pharmacologic inhibition of TLR signaling and manipulation of the intestinal environment other than by administration of specific nutrients. Some options have already been tested in clinical trials, and many others are likely to be developed and verified in a clinical setting in the future²⁷⁾.

1. Nutritional support to preserve the epithelial barrier

Adequate feeding strategies and the use of specific molecules in enteral nutrition have been shown to have a protective effect on the risk of NEC. For example, human milk is able to protect against the development of NEC via induction of intestinal maturation and healing⁴². These positive effects of human milk have been attributed to several factors (i.e., macrophages, lymphocytes, secretory IgA, lysozymes, lactoferrin, oligosaccharides, nucleotides, cytokines, growth factors, PUFA, and enzymes) that influence host immunity, inflammation, and mucosal protection, but a specific component to transfer in preterm formula has not been definitively identified^{4,42}.

Although several growth factors such as recombinant EGF, HB-EGF, and granulocyte colony-stimulating factor may play distinct and critical roles in normal intestinal development and repair following GI mucosal injury, further studies are needed

Table 2. Evidence of preventive measures for necrotizing enterocolitis

Evidence of efficacy and safety

Breast-milk feeding

Nonaggressive enteral feeding

Evidence of efficacy but questionable safety

Enteral aminoglycosides

Probiotics

Glucocorticoids, arginine

Evidence of efficacy in animal models but not in humans

Anticytokines

Growth factors

Proposed efficacy but lacking evidence

Probiotics (derived from plants and breast milk)

Microbial components and Toll-like-receptor agonists

Glutamine, n-3 fatty acids

Adapted from Neu. Acta Paediatr Suppl 2005;94:100-5⁴⁰⁾ and Grave et al. Pediatr Res 2007:62:510-4⁴¹⁾.

to clarify the mechanisms of action of these growth factors ⁴³⁾. Additionally, erythropoietin, erythropoietin-like growth factor, and cytokines may also play roles in the growth and development of the GI tract. In a rat hypoxia NEC model, pretreatment with erythropoietin resulted in a decrease in the incidence of NEC⁴⁴⁾.

There have been some studies comparing the incidence of NEC between infants breast-fed with fortified bovine milk-based or human milk-based supplements and those breast-fed without fortification. The Cochrane review by Kuschel and Harding⁴⁵⁾ presents the result of 13 randomized controlled trials that examined the long and short-term outcomes of preterm infants fed on a diet of fortified expressed breast milk. They found that the use of a fortifier was associated with improved growth, with no significant increase in the adverse outcomes, including NEC, in this group. In contrast, Sullivan et al. 42) found a significantly higher incidence of NEC in infants receiving a bovine milk-based fortifier than in those receiving a human milk-based fortifier. However, the clinical significance of this result is questionable, as the incidence of NEC was small in both groups; moreover, it is possible that this finding may be the result of a dose-related association between increased expressed breast milk feeding and a reduced risk of NEC, rather than a true increase in risk due to the use of bovine milk fortifier⁴⁶⁾.

Endothelial nitric oxide, synthesized from the amino acid L-arginine, is an important regulator of vascular perfusion. Although the incidence of NEC has been found to be decreased with L-arginine supplementation in some studies, further human studies are needed⁴⁷⁾. Moreover, the use of zinc to prevent NEC is supported by evidences demonstrating the role of zinc in the maintenance of the epithelial barrier function and in the induction of adequate immune responses in experimental models of NEC; a recent clinical trial demonstrated that high-dose oral zinc supplementation decreased the incidence of NEC in preterm neonates⁴⁸⁾.

2. Modulation of inflammatory (TLR4) signal

The pharmacological upregulation of Hsp70 (inhibitor of TLR4 signal) within the intestinal mucosa by celastrol (a novel cell permeable triterpenoid antioxidant) has been tested in a mouse model of NEC⁴⁹. In this model, celastrol was found to reduce enterocyte apoptosis and attenuate the severity of NEC, but studies in humans are still lacking. Hence, further studies are required for the establishment of these novel therapeutic options for the prevention and treatment of human cases of NEC.

Lipopolysaccharide (LPS), one of the most powerful proinflammatory bacterial virulence factors, directly induces TLR signaling. LPS-binding protein (LBP) is secreted by enterocytes in response to inflammatory stimuli and has concentration-dependent effects. At basal concentrations, LBP stimulates the inflammatory response by presenting LPS to its receptor; however, at high concentrations, LBP is able to neutralize LPS and prevent an exaggerated inflammatory response. Hence, targeting LBP may represent a novel therapeutic strategy to facilitate wound healing after the acute phase of NEC and other forms of intestinal injury²⁶.

3. Modification of gut microbiota composition

Probiotic colonization improves GI health and prevents proinflammatory signaling and disease progression by increasing mucosal barrier function and by up-regulating the immune system. Additionally, it also reduces mucosal colonization of potential pathogens, and alters the key components of intestinal inflammation⁴⁾. Several studies have demonstrated the efficacy and safety of prophylactic enteral probiotic administration in the prevention of NEC in infants with very low birth weight 50-56). In these studies, the administered probiotics were Lactobacillus alone or in combination with Bifidobacterium (Table 3). Many aspects may have influenced the different results obtained in each trial. Importantly, the effect of probiotics has been shown to be dramatically decreased in areas where the occurrence of NEC is low (<5% of very low birth weight infants), suggesting that the baseline NEC rate seems to be a major factor affecting the potential benefit of probiotic supplementation in a specific population⁵⁷⁾.

In a recent Cochrane Database Systematic Review and metaanalysis published⁵⁸⁾, 16 randomized or quasi-randomized controlled trials that enrolled a total of 2,842 preterm infants at <37 weeks gestational age and/or with a birth weight of <2,500 g were evaluated. The included studies showed highly variable enrollment criteria (birth weight and gestational age), probiotic species, times of initiation, doses, formulations, durations of therapy of probiotics, and feeding methods. However, nonetheless, the authors concluded that enteral supplementation of probiotics significantly reduced the incidence of severe NEC (stage II or more) and mortality in preterm infants weighing <1,500 g at birth. On the other hand, there are currently insufficient data with regard to the benefits and potential adverse effects in the most atrisk infants, namely those weighing <1,000 g at birth. U.S. Food and Drug Administration has not yet approved the use of probiotics in premature infants; however, this updated review of available evidence supports the routine use of probiotics in the neonatal intensive care unit⁵⁸. Because probiotic products have not been subjected to rigorous manufacturing quality control, and because the contents of such products may not be reproducible in compliance with drug or pharmaceutical standards⁴, more comparative trials among probiotic products rather than placebo-controlled trials are needed to assess the most optimal preparation, dose, and duration for extremely low birth weight infants. The main limitation to future trials is the large sample sizes required to demonstrate the benefit of probiotics in this setting^{4,57,58}.

Another proposed preventive strategy is to supplement with prebiotics, which enhance the growth of potentially beneficial intestinal microbes such as *Bifidobacterium*⁵⁹⁾. While the theoretical benefit has been reviewed, little information is currently available to support the benefit from their use in the prevention of NEC. Prebiotic agents include the oligosaccharides inulin, galactose, fructose, lactulose, and combinations of these nutrients.

4. Fecal transplantation

A novel perspective to manipulate intestinal microflora is represented by fecal transplantation. Fecal transplantation involves direct transfer of fecal material from a healthy donor to a recipient's upper or lower intestinal tract. This technique has proven effective in the treatment of refractory colitis by *Clostridium difficile* and in some cases of inflammatory bowel disease^{60]}. However, no data are currently available for neonates with NEC, and several important limitations should be considered before testing this therapeutic option in the newborn.

5. New surgical techniques

The most significant morbidity related to NEC is short bowel syndrome (SBS), which is an extensive, morbid condition with an increasing incidence. Currently, there are no functionally adequate surgical procedures for treating SBS. Bowel-lengthening

Table 3. Clinical trials for probiotic prophylaxis

Table 6. Official trials for problete propriyatio			
Reference	Report year	Species	Effect for NEC
Hoyos ⁵⁰⁾	1999	Lactobacillus acidophilus, Bifidobacterium infantis	Decreased NEC vs. historic controls
Dani et al.51)	2002	Lactobacillus GG	1.4% vs. 2.7%, not significant
Lin et al. ⁵²⁾	2005	Lactobacillus acidophilus, Bifidobacterium infantis	1.1% vs. 14%, P<0.05
Bin-Nun et al. 53)	2005	Lactobacillus bifidus, Streptococcus thermophillus, Bifidobacterium infantis	1% vs. 14%, <i>P</i> <0.05
Lin et al. 54)	2008	Bifidobacterium bifidum, Lactobacillus acidophilus	1.8% vs. 6.5%, <i>P</i> =0.02
Samanta et al. 55)	2008	Bifidobacteria infantis, Bifidobacteria bifidum, Bifidobacteria longum, and Lactobacillus acidophilus	5.5% vs. 15.8%, <i>P</i> =0.04
Sari et al.56)	2011	Lactobacillu sporogenes	5.5% vs. 8.1%, not significant

NEC, necrotizing enterocolitis.

procedures and parenteral and enteral nutrition have been shown to have modest effects. In a study from 1998, small bowel transplantation was reported to have a 5-year graft survival rate of 48%⁶¹, but this procedure is commonly accompanied by rejection and the need for lifelong immunosuppressant medications. Moreover, by serial transverse enteroplasty, a surgical method to increase the girth of the bowel in patients with SBS, the rate of independence from intravenous nutrition is only 58%⁶². Finally, tissue engineering techniques may represent a unique, novel strategy for intestinal failure after severe NEC in the future⁶³.

Conclusions

NEC is one of the leading causes of mortality, and the most common reason for emergent GI surgery in newborns. However, NEC remains a major unsolved medical challenge, for which no specific therapy exists, and its pathogenesis remains controversial. A better understanding of the pathophysiology will offer new and innovative therapeutic approaches, and future studies should be focused on the roles of the epithelial barrier, innate immunity, and microbiota in this disorder. Bioinformatics modeling is a new emerging strategy aimed at understanding the dynamics of various inflammatory markers and their application in early diagnosis and treatment.

Conflict of interest

No potential conflict of interest relevant to this paper was reported.

References

- Mizrahi A, Barlow O, Berdon W, Blanc WA, Silverman WA. Necrotizing enterocolitis in premature infants. J Pediatr 1965;66: 697-705.
- 2. Bell MJ, Ternberg JL, Feigin RD, Keating JP, Marshall R, Barton L, et al. Neonatal necrotizing enterocolitis. Therapeutic decisions based upon clinical staging. Ann Surg 1978;187:1-7.
- 3. Walsh MC, Kliegman RM. Necrotizing enterocolitis: treatment based on staging criteria. Pediatr Clin North Am 1986;33:179-201.
- Caplan MS. Neonatal necrotizing enterocolitis: clinical observations, pathophysiology, and prevention. In: Martin RJ, Fanaroff AA, Walsh MC, editors. Fanaroff & Martin's neonatal-perinatal medicine. 9th ed. Missouri: Elsevier Mosby, 2010:1431-42.
- Maheshwari A, Carlo WA. Neonatal necrotizing enterocolitis. In: Kliegman RM, Stanton BF, St. Geme JW III, Schor NF, Behrman RE, editors. Nelson textbook of pediatrics. 19th ed. Philadelphia: Elsevier Saunders, 2011:601-3.
- 6. Fanaroff AA, Stoll BJ, Wright LL, Carlo WA, Ehrenkranz RA, Stark

- AR, et al. Trends in neonatal morbidity and mortality for very low birthweight infants. Am J Obstet Gynecol 2007;196:147.e1-8.
- Stoll BJ, Hansen NI, Bell EF, Shankaran S, Laptook AR, Walsh MC, et al. Neonatal outcomes of extremely preterm infants from the NICHD Neonatal Research Network. Pediatrics 2010;126:443-56.
- Martin CR, Walker WA. Intestinal immune defences and the inflammatory response in necrotising enterocolitis. Semin Fetal Neonatal Med 2006;11:369-77.
- 9. Kamitsuka MD, Horton MK, Williams MA. The incidence of necrotizing enterocolitis after introducing standardized feeding schedules for infants between 1250 and 2500 grams and less than 35 weeks of gestation. Pediatrics 2000;105:379-84.
- Tyson JE, Kennedy KA. Minimal enteral nutrition for promoting feeding tolerance and preventing morbidity in parenterally fed infants. Cochrane Database Syst Rev 2000;(2):CD000504.
- Lin J, Nafday SM, Chauvin SN, Magid MS, Pabbatireddy S, Holzman IR, et al. Variable effects of short chain fatty acids and lactic acid in inducing intestinal mucosal injury in newborn rats. J Pediatr Gastroenterol Nutr 2002;35:545-50.
- 12. Halpern MD, Holubec H, Saunders TA, Dvorak K, Clark JA, Doelle SM, et al. Bile acids induce ileal damage during experimental necrotizing enterocolitis. Gastroenterology 2006;130:359-72.
- Wold AE, Adlerberth I. Breast feeding and the intestinal microflora
 of the infant--implications for protection against infectious
 diseases. Adv Exp Med Biol 2000;478:77-93.
- 14. Walker WA. Mechanisms of action of probiotics. Clin Infect Dis 2008;46 Suppl 2:S87-91.
- Mehall JR, Kite CA, Saltzman DA, Wallett T, Jackson RJ, Smith SD. Prospective study of the incidence and complications of bacterial contamination of enteral feeding in neonates. J Pediatr Surg 2002;37:1177-82.
- van Acker J, de Smet F, Muyldermans G, Bougatef A, Naessens A, Lauwers S. Outbreak of necrotizing enterocolitis associated with Enterobacter sakazakii in powdered milk formula. J Clin Microbiol 2001;39:293-7.
- Ballance WA, Dahms BB, Shenker N, Kliegman RM. Pathology of neonatal necrotizing enterocolitis: a ten-year experience. J Pediatr 1990;117(1 Pt 2):S6-13.
- Nowicki PT, Nankervis CA. The role of the circulation in the pathogenesis of necrotizing enterocolitis. Clin Perinatol 1994;21: 219-34.
- Dollberg S, Lusky A, Reichman B. Patent ductus arteriosus, indomethacin and necrotizing enterocolitis in very low birth weight infants: a population-based study. J Pediatr Gastroenterol Nutr 2005;40:184-8.
- Mally P, Golombek SG, Mishra R, Nigam S, Mohandas K, Depalhma H, et al. Association of necrotizing enterocolitis with elective packed red blood cell transfusions in stable, growing, premature neonates. Am J Perinatol 2006;23:451-8.
- 21. More K, Athalye-Jape G, Rao S, Patole S. Association of inhibitors of gastric acid secretion and higher incidence of necrotizing enterocolitis in preterm very low-birth-weight infants. Am J Perinatol 2013;30:849-56.
- Radulescu A, Yu X, Orvets ND, Chen Y, Zhang HY, Besner GE.
 Deletion of the heparin-binding epidermal growth factorlike growth factor gene increases susceptibility to necrotizing
 enterocolitis. J Pediatr Surg 2010;45:729-34.
- 23. Stark AR, Carlo WA, Tyson JE, Papile LA, Wright LL, Shankaran S, et al. Adverse effects of early dexamethasone in extremely-low-birth-weight infants. National Institute of Child Health and Human Development Neonatal Research Network. N Engl J Med 2001;344:95-101.

- Davey AM, Wagner CL, Cox C, Kendig JW. Feeding premature infants while low umbilical artery catheters are in place: a prospective, randomized trial. J Pediatr 1994;124(5 Pt 1):795-9.
- Neal MD, Sodhi CP, Dyer M, Craig BT, Good M, Jia H, et al. A critical role for TLR4 induction of autophagy in the regulation of enterocyte migration and the pathogenesis of necrotizing enterocolitis. J Immunol 2013;190:3541-51.
- Richter JM, Schanbacher BL, Huang H, Xue J, Bauer JA, Giannone PJ. LPS-binding protein enables intestinal epithelial restitution despite LPS exposure. J Pediatr Gastroenterol Nutr 2012;54:639-44.
- Terrin G, Scipione A, De Curtis M. Update in pathogenesis and prospective in treatment of necrotizing enterocolitis. Biomed Res Int 2014;2014:543765.
- 28. Leaphart CL, Cavallo J, Gribar SC, Cetin S, Li J, Branca MF, et al. A critical role for TLR4 in the pathogenesis of necrotizing enterocolitis by modulating intestinal injury and repair. J Immunol 2007;179:4808-20.
- 29. Sodhi CP, Shi XH, Richardson WM, Grant ZS, Shapiro RA, Prindle T Jr, et al. Toll-like receptor-4 inhibits enterocyte proliferation via impaired beta-catenin signaling in necrotizing enterocolitis. Gastroenterology 2010;138:185-96.
- 30. Huang W, Tang Y, Li L. HMGB1, a potent proinflammatory cytokine in sepsis. Cytokine 2010;51:119-26.
- 31. Afrazi A, Sodhi CP, Good M, Jia H, Siggers R, Yazji I, et al. Intracellular heat shock protein-70 negatively regulates TLR4 signaling in the newborn intestinal epithelium. J Immunol 2012;188:4543-57
- Good M, Siggers RH, Sodhi CP, Afrazi A, Alkhudari F, Egan CE, et al. Amniotic fluid inhibits Toll-like receptor 4 signaling in the fetal and neonatal intestinal epithelium. Proc Natl Acad Sci U S A 2012;109:11330-5.
- 33. Tran L, Ferris M, Norori J, Stark M, Craver R, Dowd S, et al. Necrotizing enterocolitis and cytomegalovirus infection in a premature infant. Pediatrics 2013;131:e318-22.
- 34. Stewart CJ, Marrs EC, Magorrian S, Nelson A, Lanyon C, Perry JD, et al. The preterm gut microbiota: changes associated with necrotizing enterocolitis and infection. Acta Paediatr 2012;101: 1121-7.
- 35. Smith B, Bode S, Petersen BL, Jensen TK, Pipper C, Kloppenborg J, et al. Community analysis of bacteria colonizing intestinal tissue of neonates with necrotizing enterocolitis. BMC Microbiol 2011;11:73.
- Mai V, Young CM, Ukhanova M, Wang X, Sun Y, Casella G, et al. Fecal microbiota in premature infants prior to necrotizing enterocolitis. PLoS One 2011;6:e20647.
- Warner BB, Ryan AL, Seeger K, Leonard AC, Erwin CR, Warner BW. Ontogeny of salivary epidermal growth factor and necrotizing enterocolitis. J Pediatr 2007;150:358-63.
- 38. Perrone S, Tataranno ML, Negro S, Cornacchione S, Longini M, Proietti F, et al. May oxidative stress biomarkers in cord blood predict the occurrence of necrotizing enterocolitis in preterm infants? J Matern Fetal Neonatal Med 2012;25 Suppl 1:128-31.
- Ng EW, Poon TC, Lam HS, Cheung HM, Ma TP, Chan KY, et al. Gutassociated biomarkers L-FABP, I-FABP, and TFF3 and LIT score for diagnosis of surgical necrotizing enterocolitis in preterm infants. Ann Surg 2013;258:1111-8.
- Neu J. Neonatal necrotizing enterocolitis: an update. Acta Paediatr Suppl 2005;94:100-5.
- 41. Grave GD, Nelson SA, Walker WA, Moss RL, Dvorak B, Hamilton FA, et al. New therapies and preventive approaches for necrotizing enterocolitis: report of a research planning workshop. Pediatr Res

- 2007:62:510-4.
- 42. Sullivan S, Schanler RJ, Kim JH, Patel AL, Trawoger R, Kiechl-Kohlendorfer U, et al. An exclusively human milk-based diet is associated with a lower rate of necrotizing enterocolitis than a diet of human milk and bovine milk-based products. J Pediatr 2010; 156:562-7.e1.
- Nair RR, Warner BB, Warner BW. Role of epidermal growth factor and other growth factors in the prevention of necrotizing enterocolitis. Semin Perinatol 2008;32:107-13.
- 44. Shiou SR, Yu Y, Chen S, Ciancio MJ, Petrof EO, Sun J, et al. Erythropoietin protects intestinal epithelial barrier function and lowers the incidence of experimental neonatal necrotizing enterocolitis. J Biol Chem 2011;286:12123-32.
- Kuschel CA, Harding JE. Multicomponent fortified human milk for promoting growth in preterm infants. Cochrane Database Syst Rev 2004;(1):CD000343.
- Meinzen-Derr J, Poindexter B, Wrage L, Morrow AL, Stoll B, Donovan EF. Role of human milk in extremely low birth weight infants' risk of necrotizing enterocolitis or death. J Perinatol 2009;29:57-62.
- 47. Polycarpou E, Zachaki S, Tsolia M, Papaevangelou V, Polycarpou N, Briana DD, et al. Enteral L-arginine supplementation for prevention of necrotizing enterocolitis in very low birth weight neonates: a double-blind randomized pilot study of efficacy and safety. JPEN J Parenter Enteral Nutr 2013;37:617-22.
- 48. Terrin G, Berni Canani R, Passariello A, Messina F, Conti MG, Caoci S, et al. Zinc supplementation reduces morbidity and mortality in very-low-birth-weight preterm neonates: a hospital-based randomized, placebo-controlled trial in an industrialized country. Am J Clin Nutr 2013;98:1468-74.
- Wallin RP, Lundqvist A, More SH, von Bonin A, Kiessling R, Ljunggren HG. Heat-shock proteins as activators of the innate immune system. Trends Immunol 2002;23:130-5.
- 50. Hoyos AB. Reduced incidence of necrotizing enterocolitis associated with enteral administration of Lactobacillus acidophilus and Bifidobacterium infantis to neonates in an intensive care unit. Int J Infect Dis 1999;3:197–202.
- 51. Dani C, Biadaioli R, Bertini G, Martelli E, Rubaltelli FF. Probiotics feeding in prevention of urinary tract infection, bacterial sepsis and necrotizing enterocolitis in preterm infants. A prospective double-blind study. Biol Neonate 2002;82:103-8.
- 52. Lin HC, Su BH, Chen AC, Lin TW, Tsai CH, Yeh TF, et al. Oral probiotics reduce the incidence and severity of necrotizing enterocolitis in very low birth weight infants. Pediatrics 2005;115:1-4.
- 53. Bin-Nun A, Bromiker R, Wilschanski M, Kaplan M, Rudensky B, Caplan M, et al. Oral probiotics prevent necrotizing enterocolitis in very low birth weight neonates. J Pediatr 2005;147:192-6.
- 54. Lin HC, Hsu CH, Chen HL, Chung MY, Hsu JF, Lien RI, et al. Oral probiotics prevent necrotizing enterocolitis in very low birth weight preterm infants: a multicenter, randomized, controlled trial. Pediatrics 2008;122:693-700.
- Samanta M, Sarkar M, Ghosh P, Ghosh Jk, Sinha Mk, Chatterjee S. Prophylactic probiotics for prevention of necrotizing enterocolitis in very low birth weight newborns. J Trop Pediatr 2009;55:128-31.
- 56. Sari FN, Dizdar EA, Oguz S, Erdeve O, Uras N, Dilmen U. Oral probiotics: lactobacillus sporogenes for prevention of necrotizing enterocolitis in very low-birth weight infants: a randomized, controlled trial. Eur J Clin Nutr 2011;65:434-9.
- 57. Mihatsch WA, Braegger CP, Decsi T, Kolacek S, Lanzinger H, Mayer B, et al. Critical systematic review of the level of evidence for routine use of probiotics for reduction of mortality and prevention of necrotizing enterocolitis and sepsis in preterm infants. Clin Nutr

- 2012;31:6-15.
- AlFaleh K, Anabrees J. Probiotics for prevention of necrotizing enterocolitis in preterm infants. Cochrane Database Syst Rev 2014;4:CD005496.
- 59. Sherman PM, Cabana M, Gibson GR, Koletzko BV, Neu J, Veereman-Wauters G, et al. Potential roles and clinical utility of prebiotics in newborns, infants, and children: proceedings from a global prebiotic summit meeting, New York City, June 27-28, 2008. J Pediatr 2009;155:S61-70.
- 60. Backhed F, Fraser CM, Ringel Y, Sanders ME, Sartor RB, Sherman PM, et al. Defining a healthy human gut microbiome: current concepts, future directions, and clinical applications. Cell Host

- Microbe 2012;12:611-22.
- Reyes J, Bueno J, Kocoshis S, Green M, Abu-Elmagd K, Furukawa H, et al. Current status of intestinal transplantation in children. J Pediatr Surg 1998;33:243-54.
- 62. Nucci A, Burns RC, Armah T, Lowery K, Yaworski JA, Strohm S, et al. Interdisciplinary management of pediatric intestinal failure: a 10-year review of rehabilitation and transplantation. J Gastrointest Surg 2008;12:429-35.
- 63. Guner YS, Chokshi N, Petrosyan M, Upperman JS, Ford HR, Grikscheit TC. Necrotizing enterocolitis: bench to bedside: novel and emerging strategies. Semin Pediatr Surg 2008;17:255-65.