# Association of Common Variants in TCF4 and PTPRG with Fuchs' Corneal Dystrophy: A Systematic Review and Meta-Analysis 

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#### Abstract

Topic: A meta-analysis of TCF4 and PTPRG gene variants in Fuchs' corneal dystrophy (FCD). Clinical relevance: To identify novel genetic markers in patients with FCD in different ethnic populations. Methods: MEDLINE and EMBASE were searched for eligible genetic studies on TCF4 and PTPRG in FCD. Odds ratios (OR) and $95 \%$ confidence intervals (CI) of each single-nucleotide polymorphism (SNP) in allelic, dominant and recessive models were estimated using fixed-effect model if $l^{2}<50 \%$ in the test for heterogeneity, otherwise the random effects model was used.

Results: Thirty-three records were obtained, with 8 being suitable for meta-analysis, which included five SNPs in TCF4 and two in PTPRG. There were 1610 FCD patients and 1565 controls tested for TCF4 rs613872. This SNP was strongly associated with FCD in Caucasians ( $\mathrm{P}=5.0 \times 10^{-106}$ ), with the risk allele G conferring an OR of 3.95 ( $95 \% \mathrm{Cl}$ : 3.49-4.46). A further 4 TCF4 SNPs (rs17595731, rs2286812, rs618869 and rs9954153) were also significantly associated with FCD in Caucasians ( $\mathrm{P}<10^{-8}$ ). However, we found no SNP associated with FCD in Chinese. In addition, there was no significant association between FCD and PTPRG.

Conclusion: TCF4 rs613872 is strongly associated with FCD in Caucasians but not in Chinese, which may suggest ethnic diversity in FCD SNP associations. SNPs in PTPRG were not associated with FCD in Caucasians or Chinese populations. Results of this meta-analysis indicate the need for larger-scale and multi-ethnic genetic studies on FCD to further explore the associated gene variants and their roles on the mechanism and genetic basis of FCD.


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## Introduction

Fuchs' corneal dystrophy (FCD) (MIM 136800) is a bilateral, asymmetric, progressive disorder affecting the corneal endothelium. It is characterized by the formation of guttata which are microscopic outgrowths on Descemet membrane. The resulting thickening of Descemet membrane causes a loss of corneal endothelial cells leading to corneal edema and eventual visual loss [1-4]. FCD affects $4 \%$ of the United States population aged 40 years and above and is a leading indication for corneal transplantation [1-4].

The etiology of FCD is not fully understood. Genetic factors have been suggested as a major risk factor [1,5]. Four FCD loci have been identified by genetic linkage analysis, namely FCD1, $F C D 2, F C D 3$ and $F C D 4$ on chromosome 13, 18, 5, and 9 respectively [6-11]. ZEB1 located on 10p11.2, SLC4A11 on

20p12 and LOXHD1 on 18q21.1 have been reported as causal genes [5]. ZEB1, encoding the Zinc finger E-box-binding homeobox 1 transcription factor, also known as TCF8, harbors the missense mutations p.Q840P, p.N78T, p.P649A, p.Q810P and p.A905T in patients with FCD [10,12]. There were 11 mutations in SLC4A11 that have been identified in both sporadic and familial cases [10,13]. Depletion of SLC4A11 in cultured human corneal endothelial cells resulted in the reduction of cellular proliferation with increased apoptosis [14]. LOXHD1 is located on chromosome 18 (FCD2) and mutations of LOXHD1 result in aggregates in the endothelium and increased thickness with protein abundance of the Descemet membrane which are both pathognomonic of FCD [15]. A nonsense mutation in $A G B L 1$ in the 15 q locus has been identified in patients with FCD recently and its truncated protein product AGBL1 has altered
protein-protein interaction with protein transcription factor 4 (TCF4) [16]. Another rare early-onset form of FCD has been linked to COL8A2 on chromosome $1 \mathrm{p} 34.3-\mathrm{p} 32.3$. This type of FCD shows gender differences but still has histopathological characteristics of the endothelial guttata from the classic late-onset form of FCD. It has therefore been regarded as a different form of the same disease [5].
In 2010 a genome-wide association study (GWAS) identified the single-nucleotide polymorphisms (SNPs) rs613872 in TCF4 and rs10490775 in the protein tyrosine phosphatase receptor type $G$ ( $P T P R G$ ) gene in FCD patients [17]. A strong association of rs 10490775 was found but did not reach genome-wide statistical significance in the combined analysis. Replication studies supported the association of rs613872 with FCD in Caucasians [18-22] but not in Chinese [23,24]. Other SNPs such as rs17089925 and rs17089887 were reported to be associated in Chinese [24]. The TCF4 gene is located on chromosomal region $18 q 21$. The upstream gene, CCDC68, is over 260 kbps away from TCF4, while the downstream hypothetical gene, FLJ45743, is over 600 kbps away (HapMap Data Release 28). Therefore, rs613872 is less likely to tag another gene in this region. The TCF4 gene encodes the E2-2 protein, which is expressed in the developing corneal epithelium [17]. TCF4 can induce epithelialmesenchymal transition (EMT) in epithelial cells and is vital in corneal damage repair [5,17]. TCF4 rs613872 is within binding site for two transcription factors; Ini1 (SMARCB1) and BRG1 (SMARCA4) which are components of the SWI/SNF chromatin remodeling complex involved in transcriptional regulation [25,26]. Although according to GWAS the TCF4 and PTPRG genes are significantly associated with FCD, these genes also show heterogeneity in the association profiles across populations. Thus the current meta-analysis will aim to give a comprehensive review of all the relevant studies to demonstrate clarity in association of these genes with FCD in different population groups. This is to establish the significance of TCF4 and PTPRG related to FCD, with the highlight of relevant SNPs. Included in this meta-analysis are all the reported associations of SNPs in or near TCF4 and PTPRG with FCD. The association credibility and the effects of the various genes are also evaluated.

## Methods

## Literature search

A systematic literature search was conducted in the MEDLINE and EMBASE databases (accessed July 30, 2013) with the following free words and MeSH terms: "E2-2", "transcription factor 4", "TCF-4", "TCF4", "protein tyrosine phosphatase, receptor type, G", "HPTPG", "PTPG", "PTPRG" and "Corneal Dystrophies, Hereditary", and "Fuchs' Endothelial Dystrophy". We supplemented our search by screening the reference lists of all the relevant studies, including original articles, reviews, and metaanalyses. No language filter was applied.

## Inclusion and exclusion criteria

The following inclusion criteria were applied in the review process: (1) association studies from peer-reviewed journal evaluating the association between variants in selected genes and the disease; (2) genotype or allele counts and/or frequencies in case and control groups were presented, or such data could be calculated from enumerative data provided in the articles; (3) case and control groups were unrelated and drawn from the same temporally and geographically defined population; (4) control subjects are free of any form of corneal dystrophy. All animal studies, case reports, reviews, abstracts, conference proceedings,
editorials and reports with incomplete data were excluded. Although non-English articles were not deliberately excluded, the articles in the final analysis were all in English. We also searched some Chinese databases (CBM, CNKI, VIP) but did not find any additional relevant studies. For studies published by the same group on the same gene and markers on overlapping sample population only the most recent article or the article with the largest sample size was included for analysis. Data from nonoverlapped sample populations from same study are regarded as different sample collections.

## Literature review and data extraction

Two reviewers (LL and LM) separately extracted data from the retrieved records and confirmed the validity of the included articles. Any discrepancy was resolved by other reviewers (SSR and LJC ). The following variables were extracted: author, year of publication, ethnicity of study subjects, result of Hardy-Weinberg equilibrium (HWE) test in controls, the numbers of patients and controls, demographic information, and the allele and genotype counts or frequencies of each SNP. The allele or genotype counts were calculated from the frequencies, rounding to the closest integer, in those studies where the counts were not given [2022,24].

## Meta-analysis and test for potential bias

Polymorphisms reported in two or more studies were metaanalyzed. Odds ratios (ORs) and 95\% confidence interval (CIs) for the tested allele (or minor allele) were calculated with fixed effects model if $I^{2} \leq 50 \%$ or random effects model when $I^{2}>50 \%$ [27]. We also conducted subgroup analysis based on ethnicity where applicable. The results of individual studies were pooled using the software Review Manager (RevMan, version 5.2, The Cochrane Collaboration, Copenhagen, Denmark).

Inter-study heterogeneity was tested with the $I^{2}$ test. The $I^{2}$ value was interpreted as of no ( $0-25 \%$ ), low ( $25-50 \%$ ), moderate ( $50-75 \%$ ) and high heterogeneity ( $75-100 \%$ ) [28]. Statistical significance of the association between FCD and the polymorphism was evaluated with the $Z$-test. The P values were calculated using the Z scores. An association giving a pooled P value of $<0.05$ was considered statistically significant. The Funnel plots and Egger's test were used to evaluate potential biases [29,30]. When the Egger test reported $P<0.05$, publication bias was assumed to exist.

## Results

## Characteristics of the included studies

The workflow and results of the literature review are shown in Figure 1. A total of 33 records representing 16 independent studies were identified from the search. Twelve of these met our study criteria; however, 4 were excluded since they were review papers or abstracts (Appendix S2). Therefore, 8 articles involving 9 sample collections were included in the final meta-analysis (Appendix S1).

The characteristics of all the included articles are summarized in Table 1. The 8 eligible studies included a total of $1,707 \mathrm{FCD}$ cases and 2,184 controls [17-24]. Five SNPs (rs613872, rs17595731, rs2286812, rs618869 and rs9954153) in TCF4 and 2 SNPs (rs7640737 and rs 10490775) in PTPRG were tested in at least two sample collections and thus were eligible for meta-analysis.

## Meta-analysis of TCF4 and PTPRG polymorphisms

Five SNPs in the TCF4 gene, namely rs613872, rs17595731, rs2286812, rs618869 and rs9954153, were meta-analyzed. Nota-


Figure 1. Flow diagram (modified from The PRISMA Flow Diagram) and results of literature review. Flow diagram depicted the screening process of retrieved articles, including the number and reason of exclusion.
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bly rs613872 and rs17595731 were non-polymorphic in Chinese [23,24], thus the odds ratios could not be estimated for the Chinese population.

SNP rs613872 had been investigated in 9 study cohorts. Since it is not detected in Chinese, we only present data from Caucasians, consisting of 1610 FCD cases and 1565 controls. The allele G of rs613872 was more frequent in FCD patients than in controls in all Caucasian cohorts. It is strongly associated with FCD, conferring a pooled odds ratio of 3.95 ( $95 \%$ CI: 3.49-4.46, $\mathrm{Z}=21.87, \mathrm{P}=5.0 \times 10^{-106}, I^{2}=0 \%$; Figure 2a and Table 2). Rs 17595731 was tested in 3 study cohorts totaling 377 cases and 681 controls. The pooled OR for the C allele was 4.74 ( $95 \%$ CI: $3.10-7.25, \mathrm{Z}=7.20, \mathrm{P}=6.0 \times 10^{-13}, I^{2}=0 \%$; Figure 2 b and Table 2). SNP rs2286812 was investigated in four studies with

472 cases and 1293 controls, comprising of both Chinese and Caucasians. The association was significant after pooling the data ( $\mathrm{OR}=1.77,95 \% \mathrm{CI}: 1.19-2.63, \mathrm{P}=0.00051$, Figure 2c and Table 2); however, large heterogeneity $\left(I^{2}=67 \%\right)$ was detected, which could be explained by the differences in Caucasians and Chinese cohorts. In Chinese, this SNP was not significantly associated with FCD (pooled OR $=0.88,95 \% \mathrm{CI}: 0.45-1.72$, $\mathrm{Z}=0.36, \mathrm{P}=0.72, I^{2}=31 \%$ ). When the data from the two Chinese cohorts [23,24] were removed, the association became strongly significant in Caucasians (OR $=2.36,95 \%$ CI: $1.86-2.98$, $\left.\mathrm{Z}=7.16, \mathrm{P}=8.1 \times 10^{-13}, I^{2}=0 \%\right)$. SNP rs618869 was tested in two cohorts from the same study by Baratz et al. [17] The T allele was significantly associated with FCD, with a pooled OR of 2.94 $\left(95 \%\right.$ CI: $2.23-3.89, \mathrm{Z}=7.60, \mathrm{P}=3.0 \times 10^{-14}, I^{2}=0 \%$; Table 2).
Table 1. Characteristics of the studies included in the meta-analysis.


These two cohorts were separate sample collections [17] and should have low risk of overlapping subjects. SNP rs9954153 was tested in three cohorts from two studies and the G allele was significantly associated with FCD, with a pooled OR of 2.39 ( $95 \%$ CI: $1.93-2.96, \mathrm{Z}=8.03, \mathrm{P}=9.7 \times 10^{-16}, I^{2}=0 \%$; Figure 2 d and Table 2). In the dominant and recessive models, all SNPs showed a significant association with FCD except for TCF4 rs17595731 in the recessive model (Table 2). This is likely due to the small pooled-sample size.

Two SNPs of the PTPRG gene were meta-analyzed. The rs7640737 has been studied in 4 sample collections totaling 416 FCD patients and 1175 control subjects. The minor allele T showed an opposite trend in the study of Kuot et al. as compared to the studies of Baratz et al. and Wang et al. The pooled OR was $1.56\left(95 \%\right.$ CI: $0.84-2.89, \mathrm{P}=0.16, I^{2}=83 \%$; Figure 3a and Table 2), but not statistically significant. Similarly, the A allele of rs10490775 also showed opposite effects between the studies of Kuot et al. and Baratz et al., and it was not significantly associated with FCD ( $\mathrm{OR}=1.49,95 \%$ CI: $0.67-3.27, \mathrm{P}=0.33, I^{2}=89 \%$; Figure 3b and Table 2). These Two SNPs were not significantly associated with FCD in either the dominant or recessive models (Table 2).

## Test for potential biases

A funnel plot revealed a symmetric inverted shape and no significant bias was detected.

## Discussion

In a recent genome-wide association study Baratz et al. identified the SNP rs613872 in the TCF4 gene to be significantly associated with FCD. They also found another 3 independently associated FCD SNPs (rs17595731, rs9954153 and rs2286812). Although the associations of FCD with SNPs in the PTPRG gene were indicated, the overall P value did not reach genome-wide significance [17]. These gene variants have subsequently been investigated in different ethnic populations however, the results were inconsistent and variable. In this systematic review and metaanalysis, a strong association of TCF4 rs613872 with FCD in Caucasians $\left(\mathrm{P}=5 \times 10^{-106}\right)$ was obtained and there was no heterogeneity found $\left(I^{2}=0 \%\right)$. The G allele increases the odds of FCD by nearly 4 folds. In contrast, rs613872 is not associated with FCD in ethnic Chinese [23,24]. In addition to rs613872, our data also revealed significant association of another 4 TCF4 SNPs (rs17595731, rs2286812, rs618869 and rs9954153) with FCD in Caucasians $\left(\mathrm{P}<10^{-8}\right)$. Rs2286812 has been studied in two Chinese cohorts but showed no significant association [23,24]. In contrast to TCF4, SNPs rs 10490775 and rs7640737 in PTPRG were not associated with FCD in Caucasians ( $\mathrm{P}>0.1$ ) and has distinct heterogeneity across study populations $\left(I^{2}>80 \%\right)$. Accordingly, TCF4 is the only susceptibility gene that has been confirmed so far for FCD.

This meta-analysis involves the largest sample size to date with a total of 1610 FCD cases and 1565 controls. The presence of TCF4 rs613872 in FCD among the Caucasian populations is confirmed and the G allele is a risk factor. However, TCF4 rs613872 and rs 17595731 has been investigated in a total of 91 FCD patients, 42 patients with non-Fuchs' corneal dystrophies, and 612 controls of Chinese origin $[23,24]$, but they were found to be nonpolymorphic. Instead, another TCF4 SNP rs17089887, near rs613872, was significantly associated with FCD in Singaporean Chinese, with the allele C conferring a 2.57 -fold of increased risk [24]. However, this SNP had not been included in other studies including a recent report in Chinese [23]. Its role in this





Figure 2. Forest plot of TCF4 allelic model: (a) TCF4 rs613872 (b) TCF4 rs17595731 (c) TCF4 rs2286812 (d) TCF4 rs9954153. Squares indicate the study-specific odds ratio (OR). The size of the box is proportional to the weight of the study. Horizontal lines indicate $95 \%$ confidence interval (CI). A diamond indicates the summary OR with its corresponding $95 \% \mathrm{Cl}$. doi:10.1371/journal.pone.0109142.g002
population has yet to be confirmed. Thalamuthu et al. and Baratz et al. respectively performed haplotype association tests between individual SNPs across the entire TCF4 gene and the associations were similar $[16,23]$. If rs 17089887 can be confirmed as a genuine FCD-associated SNP, then there could be another SNP in or near the TCF4 gene in linkage disequilibrium with both rs613872 and rs17089887 that may be responsible for the association signals detected on these two SNPs.

FCD has reportedly lower prevalence among Chinese than Caucasians, and it accounts for only $3 \%-4.5 \%$ of cases requiring penetrating keratoplasty [31-33]. The low occurrence may be due to the non-polymorphic TCF4 rs613872 in Chinese as shown in our meta-analysis. Furthermore, according to the database of the Human Genome Diversity Project [34], the risk allele G of TCF4 rs613872 is rare or nonexistent in populations from eastern Asia, Africa, and central and southern America but is more frequent in
Table 2. Pooled analyses on the relationship of gene polymorphisms with FCD.

| Polymorphism | Ethnicity | Allele | Number of cohorts | Sample size (Case/Control) | Genetic model | OR (95\% CI) | Z score | P-value | $\mathrm{I}^{\mathbf{2}}$ (\%) | Reference |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| TCF4 rs613872 | Caucasian | G/T | 7 | 1610/1565 | G vs. T | 3.95 (3.49-4.46) | 21.87 | $5.0 \times 10^{-106}$ | 0 | [16-21] |
|  |  |  |  |  | GG+GT vs. $T 1$ | 6.05 (5.14-7.10) | 21.84 | $9.7 \times 10^{-106}$ | 12 |  |
|  |  |  |  |  | GG vs. TT+GT | 6.47 (4.55-9.20) | 10.41 | $2.2 \times 10^{-25}$ | 8 |  |
| TCF4 rs17595731 | Caucasian | C/G | 3 | 377/681 | C vs. G | 4.74 [3.10-7.25] | 7.20 | $6.0 \times 10^{-13}$ | 0 | [16,20] |
|  |  |  |  |  | CC+CG vs. GG | 5.12 (3.29-7.96) | 7.25 | $4.2 \times 10^{-13}$ | 0 |  |
|  |  |  |  |  | CC vs. GG+CG | 5.20 (0.81-33.42) | 1.74 | 0.082 | 0 |  |
| TCF4 rs2286812 | All ancestries | T/C | 5 | 472/1293 | T vs. C | 1.77 (1.19-2.63) | 2.80 | 0.0051 | 67 | [16,20,22,23] |
|  |  |  |  |  | T+TC vs. CC | 1.87 (1.15-3.04) | 2.53 | 0.011 | 70 |  |
|  |  |  |  |  | T v v. CC+TC | 2.58 (1.29-5.14) | 2.69 | 0.0071 | 0 |  |
| TCF4 rs618869 | Caucasian | T/C | 2 | 276/403 | T vs. C | 2.94 (2.23-3.89) | 7.60 | $3.0 \times 10^{-14}$ | 0 | [16] |
|  |  |  |  |  | T+TC vs. CC | 3.77 (2.70-5.26) | 7.81 | $5.7 \times 10^{-15}$ | 0 |  |
|  |  |  |  |  | TT vs. CC+TC | 4.54 (1.69-12.18) | 3.0 | 0.0027 | 44 |  |
| TCF4 rs9954153 | Caucasian | G/T | 3 | 376/676 | G vs. T | 2.39 (1.93-2.96) | 8.03 | $9.7 \times 10^{-16}$ | 0 | [16,20] |
|  |  |  |  |  | GG+GT vs. $T$ | 2.96 (2.27-3.86) | 7.99 | $1.3 \times 10^{-15}$ | 0 |  |
|  |  |  |  |  | GG vs. GT+TT | 3.59 (1.90-6.80) | 3.94 | $8.1 \times 10^{-5}$ | 0 |  |
| PTPRG rs7640737 | All ancestries | T/C | 4 | 416/1175 | T vs. C | 1.56 (0.84-2.89) | 1.42 | 0.16 | 83 | [16,20,22] |
|  |  |  |  |  | T+TC vs. CC | 1.54 (0.80-2.97) | 1.30 | 0.19 | 81 |  |
|  |  |  |  |  | TT vs. CC+TC | 3.19 (0.68-14.93) | 1.47 | 0.14 | 52 |  |
| PTPRG rs 10490775 | Caucasian | A/G | 3 | 380/683 | A vs. G | 1.49 (0.67-3.27) | 0.98 | 0.33 | 89 | [16,20] |
|  |  |  |  |  | AA+AG vs. GG | 1.45 (0.63-3.35) | 0.88 | 0.38 | 87 |  |
|  |  |  |  |  | AA vs. GG+AG | 3.05 (0.35-26.35) | 1.01 | 0.31 | 68 |  |



Figure 3. Forest plot of PTPRG allelic model: (a) PTPRG rs7640737 (b) PTPRG rs10490775. Squares indicate the study-specific odds ratio (OR). The size of the box is proportional to the weight of the study. Horizontal lines indicate $95 \%$ confidence interval (CI). A diamond indicates the summary OR with its corresponding $95 \% \mathrm{Cl}$.
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European, Middle Eastern and southern Asian populations. In order to determine the association of TCF4 rs613872 with FCD, further studies involving larger cohorts are thus needed to correlate the prevalence of FCD and the frequencies of TCF4 polymorphisms in other regions of China and Asia.

The TCF4 gene is located on 18q21.1, encoding the E2-2 protein, a member of the ubiquitously expressed class I basic helix-loop-helix (bHLH) transcription factors which are necessary in cell growth and differentiation [35]. There is expression of E2-2 in the developing corneal epithelium [17]. TCF4 can induce epithelialmesenchymal transition (EMT) in epithelial cells, with a loss of the epithelial morphology, distinctive epithelial markers, and a gain or reorganization of mesenchymal markers, through an indirect Ecadherin repression and induction of downstream EMT regulators ZEB1 [36-38]. Defective EMT has been proposed as a potential pathway for FCD by impairing the mobilization of stem cells to repair corneal damage $[5,17,39]$. The TCF4 induced ZEB1 was also shown to be related to tumor invasiveness due to its effects in EMT [40]. ZEB1 was also shown to regulate type I collagen expression and is thus critical for the maintenance of an endothelial phenotype $[41,42]$. TCF4 rs613872 is within the chromatin immunoprecipitation sequence (ChIP-seq)-purported binding site for two transcription factors: Ini1 (SMARCB1) and BRG1 (SMARCA4) [43]. These two factors are components of the SWI/SNF chromatin remodeling complex involved in transcriptional regulation $[25,26]$. It has been postulated that variation over rs613872 would affect the spatiotemporal expression of TCF4 through Ini1 (SMARCB1) and BRG1 (SMARCA4) and hence its targets [19]. Though more evidence is needed to support this linkage, the association in TCF4 rs613872 played a significant role in identifying the potential causative genes and possibly future biomarkers for FCD.
This systematic review and meta-analysis is an overview of the published genetic studies on TCF4 and PTPRG in FCD. The
study has also revealed the limitations in the current FCD genetic studies. In particular, only a limited number of SNPs were found for this meta-analysis, and the number was even smaller in the Chinese populations making false negative errors likely. The small number of studies with small number of study participants in Chinese and Asian populations and non-polymorphism in the relevant SNPs had limited the generalizability of our results and introduced heterogeneity in the data. Further studies on TCF4 in FCD among different Chinese and Asian populations are needed to confirm the role of this gene. Besides, the homogeneity of the ethnical background in different Caucasian study cohorts may have played a role giving rise to subtle difference in the prevalence and severity of FCD which have not been accounted for in this study. It would be of higher impact and generalizability if more details and various ethnicities in relation to FCD were reported. Also, although a comprehensive search has been employed with different strategies to identify all published studies on TCF4 and $P R P R G$, relevant studies not meeting the search criteria can be missed. There may be useful results from association studies reported in abstracts, conferences and non-English journals. Thus, there can be selection and publication bias as a result of disproportionate exclusion of negative data, although we detected no significant bias by the Egger's test in the present study. Finally, important covariates like gender, age, severity of FCD, or potential gene-gene and gene-environment interactions could not be included in our analysis because raw allelic and genotypic data were not available. However, alleles are unlikely to display opposite effects based on different covariate. The association between FCD and variants in TCF4 and PTPRG as detected in this study should therefore be genuine.

In summary, results of this meta-analysis confirm 5 SNPs in TCF4, with rs613872 having the strongest effect, to be genetic susceptibility factors for FCD in Caucasians but not Chinese populations. To enhance our understanding of the mechanism of

FCD, re-sequencing studies and biological functional studies in multiple ethnic groups are required to ascertain FCD-associated gene variants and to determine their pathophysiological effects.

## Supporting Information

## Appendix S1 Lists of included studies. (DOC)

## References

1. Klintworth GK (2009) Corneal dystrophies. Orphanet J Rare Dis 4: 7
2. Krachmer JH, Purcell JJ, Young CW, Bucher KD (1978) Corneal endothelial dystrophy. A Study of 64 families. Arch Ophthalmol 96: 2036-2039.
3. Lorenzetti DWUM, Parikh N, Kaufman HE (1967) Central cornea guttata. Incidence in the general population. Am J Ophthalmol 64: 1155-1158.
4. Musch DC, Niziol LM, Stein JD, Kamyar RM, Sugar A (2011) Prevalence of corneal dystrophies in the United States: estimates from claims data. Invest Ophthalmol Vis Sci 52: 6959-6963.
5. Iliff BW, Riazuddin SA, Gottsch JD (2012) The genetics of Fuchs' corneal dystrophy. Expert Rev Ophthalmol 7: 363-375.
6. Gottsch JD, Sundin OH, Liu SH, Jun AS, Broman KW, et al. (2005) Inheritance of a novel COL8A2 mutation defines a distinct early-onset subtype of fuchs corneal dystrophy. Invest Ophthalmol Vis Sci 46: 1934-1939.
7. Gottsch JD, Zhang C, Sundin OH, Bell WR, Stark WJ, et al. (2005) Fuchs corneal dystrophy: aberrant collagen distribution in an L450W mutant of the COL8A2 gene. Invest Ophthalmol Vis Sci 46: 4504-4511.
8. Sundin OH, Broman KW, Chang HH, Vito EC, Stark WJ, et al. (2006) A common locus for late-onset Fuchs corneal dystrophy maps to 18q21.2-q21.32. Invest Ophthalmol Vis Sci 47: 3919-3926.
9. Sundin OH, Jun AS, Broman KW, Liu SH, Sheehan SE, et al. (2006) Linkage of late-onset Fuchs corneal dystrophy to a novel locus at 13pTel-13q12.13. Invest Ophthalmol Vis Sci 47: 140-145.
10. Riazuddin SA, Vithana EN, Seet LF, Liu Y, Al-Saif A, et al. (2010) Missense mutations in the sodium borate cotransporter SLC4A11 cause late-onset Fuchs corneal dystrophy. Hum Mutat 31: 1261-1268.
11. Riazuddin SA, Eghrari AO, Al-Saif A, Davey L, Meadows DN, et al. (2009) Linkage of a mild late-onset phenotype of Fuchs corneal dystrophy to a novel locus at 5q33.1-q35.2. Invest Ophthalmol Vis Sci 50: 5667-5671.
12. Mehta JS, Vithana EN, Tan DT, Yong VH, Yam GH, et al. (2008) Analysis of the posterior polymorphous corneal dystrophy 3 gene, TCF8, in late-onset Fuchs endothelial corneal dystrophy. Invest Ophthalmol Vis Sci 49: 184-188.
13. Vithana EN, Morgan PE, Ramprasad V, Tan DT, Yong VH, et al. (2008) SLC4A11 mutations in Fuchs endothelial corneal dystrophy. Hum Mol Genet 17: 656-666.
14. Liu J, Seet LF, Koh LW, Venkatraman A, Venkataraman D, et al. (2012) Depletion of SLC4A11 causes cell death by apoptosis in an immortalized human corneal endothelial cell line. Invest Ophthalmol Vis Sci 53: 3270-3279.
15. Riazuddin SA, Parker DS, McGlumphy EJ, Oh EC, Iliff BW, et al. (2012) Mutations in LOXHD 1, a recessive-deafness locus, cause dominant late-onset Fuchs corneal dystrophy. Am J Hum Genet 90: 533-539.
16. Riazuddin SA, Vasanth S, Katsanis N, Gottsch JD (2013) Mutations in AGBL1 cause dominant late-onset Fuchs corneal dystrophy and alter protein-protein interaction with TCF4. Am J Hum Genet 93: 758-764.
17. Baratz KH, Tosakulwong N, Ryu E, Brown WL, Branham K, et al. (2010) E2-2 protein and Fuchs's corneal dystrophy. N Engl J Med 363: 1016-1024.
18. Riazuddin SA, McGlumphy EJ, Yeo WS, Wang J, Katsanis N, et al. (2011) Replication of the TCF4 intronic variant in late-onset Fuchs corneal dystrophy and evidence of independence from the FCD2 locus. Invest Ophthalmol Vis Sci 52: 2825-2829.
19. Stamler JF, Roos BR, Wagoner MD, Goins KM, Kitzmann AS, et al. (2013) Confirmation of the association between the TCF4 risk allele and Fuchs endothelial corneal dystrophy in patients from the Midwestern United States. Ophthalmic Genet 34: 32-34.
20. Li YJ, Minear MA, Rimmler J, Zhao B, Balajonda E, et al. (2011) Replication of TCF4 through association and linkage studies in late-onset Fuchs endothelial corneal dystrophy. PLoS One 6: e18044.
21. Kuot A, Hewitt AW, Griggs K, Klebe S, Mills R, et al. (2012) Association of TCF4 and CLU polymorphisms with Fuchs' endothelial dystrophy and implication of CLU and TGFBI proteins in the disease process. Eur J Hum Genet 20: 632-638.

Appendix S2 Lists of excluded studies with reason. (DOC)

## Checklist S1 PRISMA checklist.

(DOC)

## Author Contributions

Conceived and designed the experiments: LCML LM LJC. Analyzed the data: LCML LM SSR. Wrote the paper: ALY VJ MEB CPP LJC.
22. Igo RP Jr, Kopplin LJ, Joseph P, Truitt B, Fondran J, et al. (2012) Differing roles for TCF4 and COL8A2 in central corneal thickness and fuchs endothelial corneal dystrophy. PLoS One 7: e46742.
23. Wang KJ, Jhanji V, Chen J, Law RW, Leung AT, et al. (2013) Association of Transcription Factor 4 (TCF4) and Protein Tyrosine Phosphatase, Receptor Type G (PTPRG) with Corneal Dystrophies in Southern Chinese. Ophthalmic Genet.
24. Thalamuthu A, Khor CC, Venkataraman D, Koh LW, Tan DT, et al. (2011) Association of TCF4 gene polymorphisms with Fuchs' corneal dystrophy in the Chinese. Invest Ophthalmol Vis Sci 52: 5573-5578.
25. Phelan ML, Sif S, Narlikar GJ, Kingston RE (1999) Reconstitution of a core chromatin remodeling complex from SWI/SNF subunits. Mol Cell 3: 247-253.
26. Kwon CS, Wagner D (2007) Unwinding chromatin for development and growth: a few genes at a time. Trends Genet 23: 403-412.
27. Lau J, Ioannidis JP, Schmid CH (1997) Quantitative synthesis in systematic reviews. Ann Intern Med 127: 820-826.
28. Higgins JP, Thompson SG, Deeks JJ, Altman DG (2003) Measuring inconsistency in meta-analyses. BMJ 327: 557-560.
29. Copas J, Shi JQ (2000) Meta-analysis, funnel plots and sensitivity analysis. Biostatistics 1: 247-262.
30. Harbord RM, Egger M, Sterne JA (2006) A modified test for small-study effects in meta-analyses of controlled trials with binary endpoints. Stat Med 25: 34433457.
31. Pan $\mathrm{Q}, \mathrm{LiX}, \mathrm{Gu} \mathrm{Y}$ (2012) Indications and outcomes of penetrating keratoplasty in a tertiary hospital in the developing world. Clin Experiment Ophthalmol 40: 232-238.
32. Chen WL, Hu FR, Wang IJ (2001) Changing indications for penetrating keratoplasty in Taiwan from 1987 to 1999. Cornea 20: 141-144.
33. Wang X, Wang W, Xu J, Wang Y (2012) Analysis of causes of bullous keratopathy in East China: a 10-year retrospective study. Graefes Arch Clin Exp Ophthalmol 250: 307-308.
34. Kent WJ, Sugnet CW, Furey TS, Roskin KM, Pringle TH, et al. (2002) The human genome browser at UCSC. Genome Res 12: 996-1006.
35. Murre C, Bain G, van Dijk MA, Engel I, Furnari BA, et al. (1994) Structure and function of helix-loop-helix proteins. Biochim Biophys Acta 1218: 129-135.
36. Sobrado VR, Moreno-Bueno G, Cubillo E, Holt LJ, Nieto MA, et al. (2009) The class I bHLH factors E2-2A and E2-2B regulate EMT. J Cell Sci 122: 1014 1024.
37. Sanchez-Tillo E, Lazaro A, Torrent R, Cuatrecasas M, Vaquero EC, et al. (2010) ZEB1 represses E-cadherin and induces an EMT by recruiting the SWI/ SNF chromatin-remodeling protein BRG1. Oncogene 29: 3490-3500.
38. Spaderna S, Schmalhofer O, Wahlbuhl M, Dimmler A, Bauer K, et al. (2008) The transcriptional repressor ZEB 1 promotes metastasis and loss of cell polarity in cancer. Cancer Res 68: 537-544.
39. Forrest MP, Hill MJ, Quantock AJ, Martin-Rendon E, Blake DJ (2014) The emerging roles of TCF4 in disease and development. Trends Mol Med 20: 322331.
40. Sanchez-Tillo E, de Barrios O, Siles L, Cuatrecasas M, Castells A, et al. (2011) beta-catenin/TCF4 complex induces the epithelial-to-mesenchymal transition (EMT)-activator ZEB1 to regulate tumor invasiveness. Proc Natl Acad Sci U S A 108: 19204-19209.
41. Terraz C, Toman D, Delauche M, Ronco P, Rossert J (2001) delta Efl binds to a far upstream sequence of the mouse pro-alpha $1(\mathrm{I})$ collagen gene and represses its expression in osteoblasts. J Biol Chem 276: 37011-37019.
42. Frisch SM (1994) Ela induces the expression of epithelial characteristics. J Cell Biol 127: 1085-1096.
43. Rosenbloom KR, Dreszer TR, Pheasant M, Barber GP, Meyer LR, et al. (2010) ENCODE whole-genome data in the UCSC Genome Browser. Nucleic Acids Res 38: D620-625.

