Hepatitis E Virus in the Food of Animal Origin: A Review

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Abstract

Hepatitis E virus (HEV) is a cosmopolitan foodborne pathogen. The viral agent infects humans through the consumption of contaminated food (uncooked or undercooked). Most cases of infection are asymptomatic and for this reason, this pathology is considered underdiagnosed. Domestic and wild animals are considered natural reservoirs: that is, domestic pig, wild boar, sheep, goat, deer, rabbit, and so on. Therefore, various work categories are at risk: that is, veterinarians, farmers, hunters, slaughterhouse workers, and so on. In these last decades, researchers found a high percentage of positivity to the molecular viral detection in several food matrices included: ready-to-eat products, processed meat products, milk, and shellfish. This review aims to provide an international scenario regarding HEV ribonucleic acid (RNA) detection in several foodstuffs. From this investigative perspective, the study aims to highlight various gaps of the current knowledge about technologies treatments' impact on viral loads. The purpose was also to provide an innovative point of view "One Health"-based, pointing out the strategic role of environmental safety.

Keywords: hepatitis E virus, RNA, foodborne disease, viral detection, one health, public health

Introduction

OODBORNE ILLNESSES CAUSED by viruses are a significant global health problem. In the recent decades, knowledge regarding foodborne viral infections has increased. Furthermore, these pathogens reduce economic growth in many countries in the world (Shirazi et al., 2018).

Data regarding foodborne viral infections change every year. Foodborne viral agents are responsible for 12% of related disease outbreaks, as reported by the European Food Safety Authority (EFSA BIOHAZ Panel et al., 2017).

In the United States, norovirus (NoV) causes $\sim 58\%$ of foodborne illnesses. However, contaminated food ingestion can also vehicle another important pathogen: hepatitis E virus (HEV).

Indeed, in the last decade there were several outbreaks in the world (Hirneisen et al., 2012).

HEV is transmitted through orofecal route. In general, contaminated water or food of animal origin ingestion (raw or undercooked products) is the main source of infection (Colson et al., 2010).

Furthermore, travels to endemic countries (HEV) pose tourists' health at risk (Guerra et al., 2017; Jemeršić et al., 2019).

HEV belongs to the Hepeviridae family and it is classified as Orthohepevirus A (Smith et al., 2014). It is a nonenveloped single-strand ribonucleic acid (RNA) virus (Purdy et al., 2017). For this structural characteristic, Johne et al. (2016) discovered that virions maintain infectivity for up to 21 days at 37°C and 28 days at room temperature.

Three overlapping open reading frames (ORF) characterize the viral genome. ORF1, ORF2, and ORF3 encode for nonstructural, capsid, and small multifunctional proteins, respectively (Emerson and Purcell, 2003).

Orthohepevirus comprises seven genotypes (HEV1-HEV7). HEV1-HEV4 isolated in human hosts, HEV3 in pigs and wild boars (Prpić et al., 2015). HEV5 and HEV6 amplified from wild boars only (Doceul et al., 2016). Recently, an eighth genotype (HEV8) has been proposed as camel variants (Sridhar et al., 2017).

HEV1 and HEV2 are human specific (especially in Asiatic and African continents). These two genotypes

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infect humans through contaminated water ingestion. This condition is related to an inappropriate wastewaters' management (EFSA BIOHAZ Panel *et al.*, 2017).

HEV3 is widespread in the world (including industrialized countries: European and American continents). It has been derived from both humans and other animal species (domestic swine and wild boar). In general, HEV3 causes sporadic cases in human patients (Kamar *et al.*, 2014; Sayed *et al.*, 2015).

However, EFSA BIOHAZ Panel *et al.* (2017) reported an increasing number of cases of hepatitis E infection (HEV3) among Western European countries. For this reason, many authors suggested that the epidemiological scenario is dynamic.

Consequently, a significant proportion of people in industrialized countries are seropositive (high titers of anti-HEV antibodies Abs), but without typical symptoms of acute hepatitis (Sayed *et al.*, 2015).

HEV4 shows a zoonotic potential. It is considered an Asiatic genotype (Lu *et al.*, 2006; Wang *et al.*, 2021), but Colson *et al.* (2012) isolated this genotype in the European continent too.

In 2013, Yugo and Meng (2013) reported an expanding host range (new animal species found positive for HEV RNA) and demonstrated cross-species infections (Table 1).

There are multiple contact points between humans, wild and domestic animals, and HEV RNA (Nan *et al.*, 2017). This "viral cycle" begins from multiple primary productions: breeding farm/mariculture (especially in extensive realities), slaughterhouses (meat products), processing meat/seafood industries, and wastewaters' management. It concludes with consumers' and environment's health.

Therefore, HEV is a global public health issue that involves humans, animals, and environment. The aim of this article was to purpose a holistic and critical point of view, focusing on the strategic role of dietary transmission that permits HEV virions diffusion. It also explains the impact of food processing technologies on the viral loads and highlights numerous gaps of knowledge concerning HEV contamination in several alimentary industries.

 TABLE 1. ANIMAL SPECIES RESERVOIR OR RECEPTIVE

 TO THE VIRUS (HEPATITIS E VIRUS)

Species	References
Pig	Sasaki <i>et al.</i> (2018)
Wild boar	Fredriksson-Ahomaa (2019)
Cattle	Tritz et al. (2018)
Sheep	Wang and Ma (2010)
Goat	Tritz et al. (2018)
Deer	Prpić et al. (2015)
Chicken	Marek et al. (2010)
Rat	Lack et al. (2012)
Rabbit	Liu et al. (2017)
Mongoose	Nakamura et al. (2006)
Bat	Drexler et al. (2012)
Ferret	Raj et al. (2012)
Fish	Batts et al. (2011)
Moose	Meng et al. (2016)
Camels	Woo <i>et al.</i> (2016)
Marine mammals	Montalvo Villalba et al. (2017)

Categories at Risk

Veterinary food inspectors, slaughterhouse workers, farmers, and hunters

Veterinary food inspectors, swine farmers, hunters, pork retailers, meat processors, and so on, are routinely exposed to the risk of HEV infection (Montagnaro *et al.*, 2015).

Unsafe raw food, poor storage infrastructure, poor personal hygiene, improper handling methods, and crosscontamination of raw food are just some of the common causes of foodborne diseases that need to be prevented by new measures (Shirazi *et al.*, 2018).

Therefore, they are more likely to be infected with HEV than the general population by 50% (Huang *et al.*, 2018).

Molecular biology (real-time PCR [RT-PCR] assay) and seroprevalence screening assays (immunoglobulin G [IgG] and/or immunoglobulin M [IgM] detection) are goldstandard methods that are used by researchers to evaluate the epidemiological scenario (De Schryver *et al.*, 2015).

Direct contact between infected wild animals (i.e., wild boar) and humans, especially for the above-mentioned work categories, causes high prevalence of anti-HEV Abs.

Carpentier *et al.* (2012) investigated the risk of viral seroprevalence by using enzyme-linked immunosorbent assay. They evaluated seroprevalence in 593 forestry workers (including woodcutters, game or fishing keepers, rangers, and controls) and 421 wild boars. Results evidenced anti-HEV Abs in 31% of the forestry workers and 14% of the wild boars. This study is one of the first papers in which scientists demonstrated that these worker categories are more exposed to HEV virions.

Slaughterhouses have a key role in HEV RNA diffusion. In most of cases, infected swine is asymptomatic, or it presents a self-limiting hepatitis. Therefore, veterinarians cannot identify or suspect positive animals from *ante mortem* evaluation. False-negative animals can enter the slaughterhouse's chain as "healthy." Consequently, contaminated meat and liver handling diffuses HEV in the entire slaughter lines (Widén, 2016).

In 2019, Milojević *et al.* (2019) screened viral prevalence and respective genotypes in slaughtered pigs in different Serbian slaughterhouses. From the same slaughter line, they collected a total of 345 samples from liver and environmental swabs.

Environmental swabs were collected from different sites/ surfaces on the slaughter line having contact with offal and fresh meat (manipulation hooks hanging liver, inspection tables, knives, sharpener, and offal collection containers). The 53% of environmental and 34% of liver samples were positive for the presence of HEV RNA.

Therefore, cross-contaminations start from slaughterhouses and arrive as final products with repercussions on consumers' health.

Krog *et al.* (2019) purpose a new approach of preventive medicine. It is characterized by fecal swabs collected from pigs before their arrival to the slaughterhouses. The detection of HEV RNA (through RT-PCR assay) in feces could be related to HEV positivity in organs. This approach will be able to produce satisfactory results preventing and reducing HEV diffusion in these structures.

In addition, other animal species pose at-risk slaughterhouse workers' health, that is, goats, rabbits (Wang *et al.*, 2016), and sheep (Mesquita *et al.*, 2020). In rabbit slaughterhouses, Geng *et al.* (2019a) registered high titers of anti-HEV IgG (43.6% male and 47.2% female workers). Seroprevalence was 64.1% among persons with working years >2 and 14.3% among those with working years <0.5 (Geng *et al.*, 2019a). Furthermore, goat farmers, shepherd, and sheep milk cheesemaker workers presented analogous results (Li *et al.*, 2017; Mesquita *et al.*, 2020).

This condition is related to the direct contact with live animals, animal blood (viremia phase), liver, and meat (Wang *et al.*, 2016; Mesquita *et al.*, 2020). Therefore, more attention is required especially during evisceration phase and avoiding bile waste dispersion and cross-contamination regarding muscle masses (Milojević *et al.*, 2019). This evidence highlights one of the crucial HEV pathogenesis principle: the entero-hepato-biliary viral cycle.

This pathogenetic aspect is strictly related to the orofecal viral transmission. It is important to remember that HEV infection is characteristically related to contaminated food ingestion.

Control groups formed by administrative staff, chemists, teachers, and so on, who have no direct contact with biohazard materials, were checked for anti-HEV Abs.

These groups presented high titers ranging between 15% IgG and 18% IgM (Carpentier *et al.*, 2012; Chaussade *et al.*, 2013; Geng *et al.*, 2019a).

Consequently, people living in industrialized countries are generally exposed to the virus owing to the dietary factors (ingestion of contaminated foodstuffs: pork liver, and meat products) (Rivero-Juarez *et al.*, 2017; Delage *et al.*, 2019).

Veterinarians have an important educative role in the implementation of knowledge about zoonoses and their impact on the public health (Bahnson *et al.*, 2001; Poizat *et al.*, 2017). An efficient food safety service has to provide a riskbased surveillance and formative plans (improvement of slaughterhouse's personnel education), as reported by the European Law (EC No. 625/2017) (Huang *et al.*, 2018).

In the near future, strict compliance between private producers and public food safety agencies will improve foodstuffs quality and safety.

Immunocompromised patients

In immunocompetent patients, HEV is usually asymptomatic (it depends on the genotype) (Weigand *et al.*, 2018). Only a certain percentage of cases present icterus with nausea, fever, abdominal pain, vomiting, and hepatomegaly after an incubation period of 2 to 8 weeks (Park *et al.*, 2016).

Immunocompromised individuals such as organ transplant recipients (Kamar *et al.*, 2008), patients with chronic liver disease (EFSA BIOHAZ Panel *et al.*, 2011), patients affected by HIV infection (Dalton *et al.*, 2009), and those affected by lymphoblastic leukemia (Motte *et al.*, 2012) are some of the categories showing high risk of HEV infection.

These patients usually require the support of blood or blood product transfusions. These administrations can be responsible for interhuman HEV3 and HEV4 transmission (Matsubayashi *et al.*, 2008), because HEV RNA resists blood cell inactivation (Hauser *et al.*, 2014). Therefore, some industrialized countries (Ireland, United Kingdom, Japan, the Netherlands, and Germany) introduced nationwide HEV RNA screenings of blood donations (Domanović *et al.*, 2017).

Most of screened blood samples contained low viral loads and low antibody prevalence (United States 7.7%) (Ditah *et al.*, 2014). Data from United States and Canadian are lower than European countries, that is, IgG 8.7% and IgM 0.4%. Seroprevalence variations depend on the dietary factors (i.e., ingestion of contaminated foodstuffs) (Delage *et al.*, 2019).

High seroprevalence values are also influenced by alimentary and geographical traditions, that is, hunting activities and game meat product consumes. It is largely performed in small communities (anti-HEV immunoglobulin over 30% in Sardinia and Abruzzo regions) (Spada *et al.*, 2019).

Therefore, industrialized countries adopt a new and no alarmistic approach focused on "risk-based decision making." It purposes molecular screenings of blood, blood products, and foodstuff destined to such vulnerable categories of patients. This system can be used if there is a condition characterized by the low seroprevalence in blood donors (Delage *et al.*, 2019).

Meat Products

The first evidence of HEV transmission in humans was described in Japan from wild animal meat. These cases were associated with the consumption of uncooked or under-cooked pork meat and venison (Matsuda *et al.*, 2003).

In general, contamination comes from the primary production of fresh products, food handling, or water used for food production and preparation (EFSA BIOHAZ Panel *et al.*, 2011).

In contrast to many other foodborne viruses, viral contamination of meat products is not only restricted to the food surface. HEV can also localize in the internal parts of food products (Bouwknegt *et al.*, 2009). Viremia is responsible for HEV virion diffusion in several muscles. Diaphragm muscle may also contain small amounts of pig liver tissue owing to its contiguity. Therefore, sanitary authorities advised to stop using diaphragm muscle in unheated pork products (Bouwknegt *et al.*, 2017).

There is little information concerning HEV survival in food matrices such as ready-to-eat and raw meat products containing swine meat or liver. Consequently, it is necessary to obtain more data about the viral infectious dose from contaminated food consumption.

In general, thermal treatments eliminate the risk of HEV infection. Conversely, there are few studies about the impact of food-processing technologies on viral loads (Sarno *et al.*, 2017).

In North Europe, pork liver sausages are typically uncooked or cooked shortly during processing. Therefore, they still contain virions (Di Bartolo *et al.*, 2012) and vehicle HEV to the final consumers (Kubacki *et al.*, 2017; Giannini *et al.*, 2018) (Table 2).

Furthermore, traditional and homemade food processing is another crucial aspect. These are typical in Mediterranean European countries (Italy, Spain, France, and Greece) (EFSA BIOHAZ Panel *et al.*, 2011).

In Italy, Istituto Zooprofilattico Sperimentale del Mezzogiorno and Istituto Superiore di Sanità evaluated the detection of HEV RNA in typical and regional homemade meat products. Results are given in Table 2. From these data evaluation, the Italian Public Health Minister purposed new guidelines for a safe domestic production (Montone *et al.*, 2019).

Spain is one of the principal European producers of pork from extensive farming (black Iberian pig) and bushmeat

Country	Meat products	Percentage of HEV RNA detection	References	
The Netherlands	255 raw pork sausages		Boxman <i>et al.</i> (2020)	
	Cervelaat	10.8%	· · · · · ·	
	Salami	18.5%		
	Metworst	26.1%		
	Snijworst	16.3%		
Switzerland	90 ready-to-eat products		Moor <i>et al.</i> (2018)	
	Local liver sausages	18.9%		
	Raw meat sausages	5.7%		
Germany	120 meat products		Szabo et al. (2015)	
2	Raw pig liver sausages	22%	× ,	
	Raw pig salami	20%		
	Raw wild boar sausages	10%		
Italy	99 traditional pork meat products	0.0%	Montone et al. (2019)	
2	63 wild boar homemade meat and liver sausages	6.3%		
France	394 samples		Pavio et al. (2014)	
	Typical pig liver sausages	30%		
	Dry salted liver	3%		
	Liver quenelles	25%		
China	107 retail samples		Hao et al. (2018)	
	Retail pork meats	33%		
	Retail pig livers	8.3%		
	Pig intestines	18.7%		
	Pig spleens	33.3%		
	Pig ureters	26.3%		
China	413 retail samples		Geng <i>et al.</i> (2019b)	
	Pig livers	6.1%	8	
	Pig kidneys	3.1%		
	Typical "blood curd"	1.2%		
The Netherlands	537 samples:		Boxman <i>et al.</i> (2019)	
	Liver	12.7%		
	Liver sausages ("Liverwurst")	70.7%		
	Liver pate	68.9%		
	Pork chops	0.0%		

TABLE 2. HEPATITIS E VIRUS RNA DETECTION IN MEAT PRODUCTS

HEV, hepatitis E virus; RNA, ribonucleic acid.

(deer and wild boar). Every year wild boar meat consumption level is ~ 3.5 kg per person and it represents an important risk factor for HEV infection, especially in Southern Spain (Pineda *et al.*, 2014).

A representative case was reported by Spanish authors at the Infectious Disease Unit of the Hospital in Cordoba (Rivero-Juarez *et al.*, 2017). A 32-year-old male patient (HIV infected) presented gastroenteric symptoms: malaise, diarrhea, jaundice, vomiting, and fever.

Molecular screenings confirmed HEV RNA detection. The patient's family represented an emblematic example because it comprised wild boar hunters and consumers of game meat (roasted or grilled wild boar meat).

Laboratories also screened two portions of wild boar meat (chap and loin). Both slices were positive with a viral load of 230–500 copies/mL and 155–300 copies/mL for chap and loin, respectively (Rivero-Juarez *et al.*, 2017).

In this case, cooking processes did not completely eliminate HEV, in contrast to the previous studies (Kamar *et al.*, 2014). However, a complete inactivation may also be dependent on the initial viral load and the exact composition of the food matrix (Cook and Van der Poel, 2015).

In the intercontinental scenario, China is the largest porkproducing and consuming country (Geng *et al.*, 2019b). It is common to consume hot pork (without being frozen). It means that pork products are delivered directly from the slaughterhouse to the retail markets. Pork viscera and blood (i.e., "blood curd" is a popular shortly cooked product) are usually consumed in Chinese popular tradition. These matrices pose at-risk consumers' health (Table 2).

There is a fragmentary knowledge about the resistance of HEV under food processing technologies, that is, fermentation, food aging, curing, drying (salami, sausages, etc.), smoking, and cooking processes.

Therefore, food product's label should be integrated with information about the potential risk of HEV infection. In this way, the public authorities have to prevent and control possible foodborne outbreaks (Geng *et al.*, 2019b), and protect vulnerable consumers (i.e., patients with preexisting chronic liver disease and transplant recipients) (Delage *et al.*, 2019).

A fundamental solution comes from innovative food technologies application. Emmoth *et al.* (2017) provided the possibility to apply new systems to inactivate viruses (using murine NoV MNV and feline calicivirus FCV as a model for HEV) in swine food matrices.

In particular, they evaluated the effects of high-pressure processing (HPP), lactic acid (LA), and intense light pulse (ILP).

Researchers applied three treatments (HPP, LA, and ILP) to fresh swine liver, dry-cured ham slices, and cold-smoked pork sausages (purchased from local grocery stores). HPP demonstrated efficacy (in the range 300–600 MPa) both on the food surfaces and on internalized viruses. LA and ILP obtained lower viral reduction. Therefore, physical parameters and properties in combination with biochemical ones will encourage new approaches to the food safety concept.

Milk

The European authorities suggest that milk is a possible source of HEV infection. Therefore, the consumption of unpasteurized and contaminated milk represents a risk for consumers (EFSA BIOHAZ Panel *et al.*, 2017).

The pasteurization process is an important phase in dairy industries. In general, it permits to prevent and control foodborne infection (Huang *et al.*, 2016).

However, in some cases, it is not able to inactivate all viral loads (HEV), and it is possible that consumers infect through pasteurized milk ingestion. Conversely, milk boiling process (100°C for 3 min) provides a complete sterilization (Huang *et al.*, 2016).

Cow's milk has been largely screened in numerous research projects by scientists. However, HEV has been also detected in other dairy species: goat (Long *et al.*, 2017), sheep, and donkey (Demirci *et al.*, 2019) (Table 3).

There are two different epidemiological realities: on one side industrialized countries, and on the other, developing nations.

In the European Union Member States, HEV RNA and anti-HEV antibodies were not detected (Baechlein and Becher, 2017; Vercouter *et al.*, 2018) (Table 3).

Conversely, in the African (Sayed *et al.*, 2020) and Asiatic continents (Huang *et al.*, 2016; Long *et al.*, 2017; Geng *et al.*, 2019c) it is possible to find contrasting data between rural and industrialized regions (Table 3).

In the rural regions, there are traditional small mixed farms, where different animal species (i.e., bovine, swine, ovine, caprine, and lagomorph) live in direct contact. This condition represents a particular epidemiological scenario that critically improve cross-species infections (Geng *et al.*, 2019c). It is also

strictly related to poor hygiene practices "from the stall to the consumers." Therefore, this factor contributes to increase seroprevalences of anti-HEV IgG Abs.

The role of mammary glands as source of HEV RNA excretion has been investigated in humans too. High titers of anti-HEV antibodies were registered in pregnant women (5.7%, 25/439 mainly owing to the dietary factors: contaminated water and food ingestion) (Alvarado-Esquivel *et al.*, 2014).

High steroid hormone (estrogens) levels (in pregnant women) promote viral multiplication (Singh *et al.*, 2019). Consequently, it is possible to isolate HEV RNA from human breast milk during acute phase of infection (Rivero-Juarez *et al.*, (2016).

In conclusion, it is possible to consider that there is a multifocal sanitary perspective. It begins from healthy farms (through the improvement of zootechnic management during pre- and postdipping phases) and it continues in dairy industries through the introduction of new technologies (i.e., high hydrostatic pressures and ultrasounds) to provide safe food products.

Shellfish Products

Food handling, freshwaters used in aquaculture, wastewaters, and water used for vegetables and fruit irrigation can be vectors of enteric viruses (i.e., HEV) (EFSA BIOHAZ Panel *et al.*, 2011; Fusco *et al.*, 2019). These viral pathogens arrive to the lamellibranch mollusks' aquaculture farms. Mollusks accumulate and concentrate water microorganisms (bioaccumulation phenomenon) (Krog *et al.*, 2014; La Rosa *et al.*, 2018). Different studies demonstrate the persistence of HEV within shellfish for weeks (Lees, 2000; Loisy *et al.*, 2005; Benabbes *et al.*, 2012).

Therefore, raw edible mollusks' ingestion (i.e., raw oysters) (O'Hara *et al.*, 2018) is a risk for the final consumers' health (Said *et al.*, 2009; Diez-Valcarce *et al.*, 2012). On the other side, thermal treatments (cooking processes) always reduce viral loads (O'Hara *et al.*, 2018).

Mollusks health is strictly related to a correct wastewaters management (bioindicators) (Fiorito *et al.*, 2019). Consequently, environmental safety has repercussion on consumers (Suffredini *et al.*, 2014).

	TABLE 3.	INTERNATIONAL	Scenario	ABOUT	HEPATITIS	Е	VIRUS	RNA	DETECTION	IN	Milk	PRODUCT	гs
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Continent	Species	Samples	Percentage of HEV RNA detection	References
Europe (Belgium)	Bovine	504 milk samples (collected from 416 dairy farms)	0.0%	Vercouter et al. (2018)
Europe (Germany)	Bovine	400 milk samples	0.0%	Baechlein and Becher (2017)
Europe (Turkey)		231 raw milk		Demirci et al. (2019)
1 \)/	Bovine	48 cow milk	29.16% (positive for HEV RNA)	~ /
	Caprine	65 goat milk	18.46% (positive for HEV RNA)	
	Ovine	65 sheep milk	12.3% (positive for HEV RNA)	
	Donkey	53 donkey milk	24.5% (positive for HEV RNA)	
North Africa (Egypt)	Bovine	480 milk samples (collected from 12 farms)	0.2% (positive both for HEV RNA and anti-HEV IgG) 1.6% (positive for anti-HEV IgG)	Sayed et al. (2020)
Asia (China)	Bovine Bovine Caprine	140 milk samples 416 milk samples 54 milk samples	37.1% (positive for HEV RNA) 0.0% 74.07% (positive for anti-HEV IgG)	Huang <i>et al.</i> (2016) Geng <i>et al.</i> (2019c) Long <i>et al.</i> (2017)

IgG, immunoglobulin G.

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Screened species	Percentages of HEV RNA positive samples	References
81 Mytilus galloprovincialis	14.81%	Mesquita et al. (2016)
384 samples		La Rosa <i>et al.</i> (2018)
298 <i>Mytilus</i> spp.	2.08%	
41 Siliqua patula	0.52%	
33 Chamelea gallina	0.0%	
12 Donax spp.	0.0%	
39 seawater samples	12.8%	
168 samples:		
70 cultured mussels (<i>M. galloprovincialis</i>)	28.5%	Rivadulla et al. (2019)
35 wild mussels (<i>M. galloprovincialis</i>)	37.1%	
31 clams (<i>Ruditapes</i> spp.)	16.1%	
32 cockles (Cerastoderma edule)	9.3%	
108 mollusk samples	1%	Pupari <i>et al.</i> (2019)
23 water samples	4.3%	
63 Pacific oysters (<i>Crassostrea gigas</i>)	9.5%	Suffredini et al. (2020)
58 clams (<i>Meretrix lyrata</i>)	13.8%	· · · · · ·

TABLE 4. HEPATITIS E VIRUS RNA DETECTION IN SHELLFISH AND WATER SAMPLES

HEV particles are protagonists of a particular "water's cycle" (Fenaux *et al.*, 2019). It begins from virus elimination through human- and animal-infected feces that arrive to the waste treatment plants. Consequently, treated waters can be used in agriculture for fruits and crops irrigation, or they arrive to the sea in mariculture farms.

This cycle concludes with the final consumers, who are infected through contaminated waters, raw mollusks, raw or undercooked meat products, fruits and vegetables ingestion (Fenaux *et al.*, 2019).

Aquaculture farms are frequently localized near harvested areas, that is, urban areas, slaughterhouses and meat preparation industries (O'Hara *et al.*, 2018; Rivadulla *et al.*, 2019), or livestock areas (Barreira *et al.*, 2010). This localization explains the data reported in Table 4.

Therefore, waste treatment plants are a critical control point. New surveillance, control measures, and structural implementations could be useful to prevent viral diffusion and infections.

A parallel development of molecular diagnostic assays will provide epidemiological and ecological information regarding HEV circulation (Di Profio *et al.*, 2019).

Conclusions

HEV detection has been largely studied in meat production chains by numerous international research teams. Furthermore, it has been also detected in other food matrices that is, milk (cow, goat, sheep, and donkey) and mollusks.

Indeed, this review article reports the necessity of further investigations owing to the fragmentary knowledge about the efficacy of food processing technologies on viral loads.

For the future, the World Health Organization (2016) estimates and confirms that numerous foodborne cases of hepatitis E infection (caused by the following genotypes: HEV1, HEV2, and HEV4) will continue to affect endemic countries (African and Asiatic continents).

HEV3 will still be responsible for sporadic infections in Northern America and Europe. Therefore, public health authorities and private producers should enforce and improve monitoring activities in different food production chains. Regional and traditional realities will be crucial viral reservoirs. Therefore, adequate sanitary measures are required. For symptomatic human patients, there are two possible approaches to treat and prevent HEV infections.

On the one side, there is a pharmacological approach through ribavirin administration. The British Transplantation Society supports that this molecule provide satisfactory results in transplant recipients (McPherson *et al.*, 2018).

However, ribavirin is not recommended for pregnant women because of numerous adverse and teratogenic effects. Furthermore, emerging variants of HEV influence the efficacy and prognosis of therapies (Todt *et al.*, 2018).

On the other side, vaccination is a valid alternative.

It can be administrated in any epidemiological realities, where several outbreaks occur (endemic nations). In this way, it guarantees immunization for patients at risk.

Hecolin (Xiamen Innovax Biotech, China) is the first registered and licensed vaccine against HEV. It is a proteinbased vaccine, and Chinese scientists certified that it induces a valid immune response (Lee *et al.*, 2015).

It provides 100% protection against HEV1 and also crossprotects against HEV4. Therefore, it is strongly recommended for fragile categories.

However, no data are available about cross-protection against HEV3 (World Health Organization, 2016).

Authors underline that is necessary to discover other viral epitopes that can produce adequate immune responses. Furthermore, it is also important that patients' immunoglobulins will provide an efficient cross-protection against different HEV genotypes.

Therefore, this virus still remains an emerging and reemerging pathogen. It involves the relationships between HEV and multiple factors, that is, human and animal health, alimentary and environmental safety.

Environmental health represents a "critical control point" in foodborne pathogen transmission. On this concept, the European Commission built the new concept of food safety control measures (EC No. 625/2017).

Authors suggest that public authorities should incentivize new system of collaboration between private industries and public research programs. In this way, it will be possible to fill the above-mentioned gaps to obtain a complete point of view regarding HEV. This article stimulates and inspires new approaches to elaborate data in an innovative way.

Authors' Contributions

All authors agreed to the publication of this article. The authors declare that they have no competing interests.

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References

- Alvarado-Esquivel C, Sánchez-Anguiano LF, Hernández-Tinoco J. Hepatitis E virus exposure in pregnant women in rural Durango, Mexico. Ann Hepatol 2014;13:510–517.
- Baechlein C, Becher P. No evidence for zoonotic HEV infection through dairy milk in Germany. Hepatology 2017;65:394– 395.
- Bahnson PB, Michalak MM, Miller GY. Pork producers' attitudes, knowledge, and production practices that relate to onfarm food safety. J Food Prot 2001;64:1967–1972.
- Barreira DM, Ferreira MS, Fumian TM, Checon R, de Sadovsky AD, Leite JP, Miagostovich MP, Spano LC. Viral load and genotypes of noroviruses in symptomatic and asymptomatic children in Southeastern Brazil. J Clin Virol 2010;47:60–64.
- Batts W, Yun S, Hedrick R, Winton J. A novel member of the family Hepeviridae from cutthroat trout (*Oncorhynchus clarkii*). Virus Res 2011;158:116–123.
- Benabbes L, Ollivier J, Schaeffer J, Parnaudeau S, Rhaissi H, Nourlil J, Le Guyader FS. Norovirus and other human enteric viruses in Moroccan shellfish. Food Environ Virol 2012;5: 35–40.
- Bouwknegt M, Rutjes SA, Reusken CB, Stockhofe-Zurwieden N, Frankena K, de Jong MC, de Roda Husman AM, Poel WH. The course of hepatitis E virus infection in pigs after contact-infection and intravenous inoculation. BMC Vet Res 2009;5:7.
- Bouwknegt M, van't Hooft BJ, Koppen K, Rietveld H, Straatsma G, Heres L. Ordinal QMRA to prioritize pork products that may contribute to foodborne hepatitis E virus transmission. Abstract T3-H.2, Presented Society for Risk Analysis Annual Meeting, December 13, 2017, SRA, Arlington, VA, USA.
- Boxman I, Jansen C, Zwartkruis-Nahuis A, Hägele G, Sosef NP, Dirks R. Detection and quantification of hepatitis E virus RNA in ready to eat raw pork sausages in the Netherlands. Int J Food Microbiol 2020;333:108791.
- Boxman ILA, Jansen CCC, Hagele G, Zwartkruis-Nahuis A, Tijsma ASL, Vennema H. Monitoring of pork liver and meat products on the Dutch market for the presence of HEV RNA. Int J Food Microbiol 2019;296:58–64.

- Carpentier A, Chaussade H, Rigaud E, Rodriguez J, Bernhault C, Boué F, Tognon M, Touzé A, Garcia-Bonnet N, Choutet P, Coursaget P. High Hepatitis E virus seroprevalence in forestry workers and wild boars in France. J Clin Microbiol 2012;50:2888–2893.
- Chaussade H, Rigaud E, Allix A, Carpentier A, Touzé A, Delzescaux D, Choutet P, Garcia-Bonnet N, Coursaget P. Hepatitis E virus seroprevalence and risk factors for individuals in working contact with animals. J Clin Virol 2013; 58:504–508.
- Colson P, Borentain P, Queyriaux B, Kaba M, Moal V, Gallian P, Heyries L, Raoult D, Gerolami R. Pig liver sausage as a source of hepatitis E virus transmission to humans. J Infect Dis 2010;202:825–834.
- Colson P, Romanet P, Moal V, Eono F, Renson P, Bourry O, Pavio N, Rose N. Autochthonous infections with hepatitis E virus genotype 4, France. Emerg Infect Dis 2012;18:1361– 1364.
- Cook N, Van der Poel W. Survival and elimination of hepatitis E virus: A review. Food Environ Virol 2015;7:189–194.
- Dalton HR, Bendall RP, Keane FE, Tedder RS, Ijaz S. Persistent carriage of hepatitis E virus in patients with HIV infection. N Engl J Med 2009;361:1025–1027.
- Delage G, Fearon M, Gregoire Y, Hogema BM, Custer B, Scalia V, Hawes G, Bernier F, Nguyen ML, Stramer SL. Hepatitis E virus infection in blood donors and risk to patients in the United States and Canada. Transfus Med Rev 2019;33: 139–145.
- Demirci M, Yiğin A, Ünlü Ö, Kılıç Altun S. Farklı hayvanlardan elde edilen çiğ sütlerde HEV RNA miktarının ve genotiplerinin tespiti [Detection of HEV RNA amounts and genotypes in raw milks obtained from different animals]. Mikrobiyol Bul 2019;53:43–52.
- De Schryver A, De Schrijver K, François G, Hambach R, van Sprundel M, Tabibi R, Colosio C. Hepatitis E virus infection: An emerging occupational risk? Occup Med (Lond.) 2015;65: 667–672.
- Di Bartolo I, Diez-Valcarce M, Vasickova P, Kralik P, Hernandez M, Angeloni G, Ostanello F, Bouwknegt M, Rodríguez-Lázaro D, Pavlik I, Ruggeri FM. Hepatitis E virus in pork production chain in Czech Republic, Italy and Spain. Emerg Infect Dis 2012;18:1282–1289.
- Diez-Valcarce M, Kokkinos P, Söderberg K, Bouwknegt M, Willems K, de Roda-Husman AM, von Bonsdorff CH, Bellou M, Hernández M, Maunula L, Vantarakis A, Rodríguez-Lázaro D. Occurrence of human enteric viruses in commercial mussels at retail level in three European countries. Food Environ Virol 2012;4:73–80.
- Di Profio F, Melegari I, Palombieri A, Sarchese V, Arbuatti A, Fruci P, Marsilio F, Martella V, Di Martino B. High prevalence of hepatitis E virus in raw sewage in Southern Italy. Virus Res 2019;272:197710.
- Ditah I, Ditah F, Devaki P, Ditah C, Kamath PS, Charlton M. Current epidemiology of hepatitis E virus infection in the United States: Low seroprevalence in the National Health and Nutrition Evaluation Survey. Hepatology 2014;60:815–822.
- Doceul V, Bagdassarian E, Demange A, Pavio N. Zoonotic hepatitis E virus: Classification, animal reservoirs and transmission routes. Viruses 2016;8:270.
- Domanović D, Tedder R, Blümel J, Zaaijer H, Gallian P, Niederhauser C, Sauleda Oliveras S, O'Riordan J, Boland F, Harritshøj L, Nascimento M, Ciccaglione AR, Politis C, Adlhoch C, Flan B, Oualikene-Gonin W, Rautmann G, Strengers P, Hewitt P. Hepatitis E and blood donation

safety in selected European countries: A shift to screening? Euro Surveill 2017;22:30514.

- Drexler JF, Seelen A, Corman VM, Fumie Tateno A, Cottontail V, Melim Zerbinati R, Gloza-Rausch F, Klose SM, Adu-Sarkodie Y, Oppong SK, Kalko EK, Osterman A, Rasche A, Adam A, Müller MA, Ulrich RG, Leroy EM, Lukashev AN, Drosten C. Bats worldwide carry hepatitis E virus-related viruses that form a putative novel genus within the family Hepeviridae. J Virol 2012;86:9134–9147.
- EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), Andreoletti O, Budka H, Buncic S, Collins JD, Griffin J, Hald T, Havelaar AH, Hope J, Klein G, Koutsoumanis K, McLauchlin J, Messens W, Müller-Graf C, Nguyen-The C, Noerrung B, Peixe L, Maradona MP, Ricci A, Sofos J, Threlfall J, Vågsholm I, Vanopdenbosch E. Scientific opinion on an update on the present knowledge on the occurrence and control of foodborne viruses. EFSA J 2011;9:2190.
- EFSA BIOHAZ Panel (EFSA Panel on Biological Hazards), Ricci A, Allende A, Bolton D, Chemaly M, Davies R, Escamez PSF, Herman L, Koutsoumanis K, Lindqvist R, Nørrung B, Robertson L, Ru G, Sanaa M, Simmons M, Skandamis P, Snary E, Speybroeck N, Kuile BT, Threlfall J, Wahlström H, Di Bartolo I, Johne R, Pavio N, Rutjes S, van der Poel W, Vasickova P, Hempen M, Messens W, Rizzi V, Latronico F, Girones R. Public health risks associated with hepatitis E virus (HEV) as a foodborne pathogen. EFSA J 2017;15:e04886.
- Emerson SU, Purcell RH. Hepatitis E virus. Rev. Med Virol 2003;13:145–154.
- Emmoth E, Rovira J, Rajkovic A, Corcuera E, Wilches Pérez D, Dergel I, Ottoson JR, Widén F. Inactivation of viruses and bacteriophages as models for swine hepatitis E virus in food matrices. Food Environ Virol 2017;9:20–34.
- Fenaux H, Chassaing M, Berger S, Gantzer C, Bertrand I, Schvoerer E. Transmission of hepatitis E virus by water: An issue still pending in industrialized countries. Water Res 2019;151:144–157.
- Fiorito F, Amoroso MG, Lambiase S, Serpe FP, Bruno T, Scaramuzzo A, Maglio P, Fusco G, Esposito M. A relationship between environmental pollutants and enteric viruses in mussels (*Mytilus galloprovincialis*). Environ Res 2019;169: 156–162.
- Fredriksson-Ahomaa M. Wild boar: A reservoir of foodborne zoonoses. Foodborne Pathog Dis 2019;16:153–165.
- Fusco G, Anastasio A, Kingsley DH, Amoroso MG, Pepe T, Fratamico PM, Cioffi B, Rossi R, La Rosa G, Boccia F. Detection of hepatitis A virus and other enteric viruses in shellfish collected in the Gulf of Naples, Italy. Int J Environ Res Public Health 2019;16:2588.
- Geng Y, Zhao C, Geng K, Wang C, Wang X, Liu H, Wang Y. High seroprevalence of hepatitis E virus in rabbit slaughterhouse workers. Transbound Emerg Dis 2019a;66:1085–1089.
- Geng Y, Zhao C, Guo T, Xu Y, Wang X, Huang W, Liu H, Wang Y. Detection of hepatitis E virus in raw pork and pig viscera as food in Hebei Province of China. Foodborne Pathog Dis 2019b;16:325–330.
- Geng Y, Zhao C, Huang W, Wang X, Xu Y, Wu D, Du Y, Liu H, Wang Y. Hepatitis E virus was not detected in feces and milk of cows in Heibei province of China: No evidence for HEV prevalence in cows. Int J Food Microbiol 2019c;291:5–9.
- Giannini P, Jermini M, Leggeri L, Nuesch-Inderbinen M, Stephan R. Detection of hepatitis E virus RNA in raw cured sausages and raw cured sausages containing pig liver at retail stores in Switzerland. J Food Prot 2018;81:43–45.

- Guerra JAA, Kampa KC, Morsoletto DGB, Junior AP, Ivantes CAP. Hepatitis E: A literature review. J Clin Transl Hepatol 2017;5:376–383.
- Hao X, Cao W, Situ J, Zhao Y, Huang F, Yu W. Hepatitis E virus detected in pork products. Food Environ Virol 2018;10: 391–393.
- Hauser L, Roque-Afonso AM, Beylouné A, Simonet M, Deau Fischer B, Burin des Roziers N, Mallet V, Tiberghien P, Bierling P. Hepatitis E transmission by transfusion of Intercept blood system-treated plasma. Blood 2014;123:796–797.
- Hirneisen KA, Hoover DG, Hicks DT, Pivarnik LF, Kniel KE. Pressure inactivation of enteric viruses in a seafood salad-like product. J Aquatic Food Product Technol 2012;21:445–467.
- Huang F, Li Y, Yu W, Jing S, Wang J, Long F, He Z, Yang, C, Bi Y, Cao W, Liu C, Hua X, Pan Q. Excretion of infectious hepatitis E virus into milk in cows imposes high risks of zoonosis. Hepatology 2016;64:350–359.
- Huang X, Huang Y, Wagner AL, Chen X, Lu Y. Hepatitis E virus infection in swine workers: A meta-analysis. Zoonoses Public Health 2018;66:155–163.
- Jemeršić L, Prpić J, Brnić D, Keros T, Pandak N, Đaković Rode O. Genetic diversity of hepatitis E virus (HEV) strains derived from humans, swine and wild boars in Croatia from 2010 to 2017. BMC Infect Dis 2019;19:269.
- Johne R, Trojnar E, Filter M, Hofmann J. Thermal stability of hepatitis E virus as estimated by a cell culture method. Appl Environ Microbiol 2016;82:4225–4231.
- Kamar N, Dalton HR, Abravanel F, Izopet J. Hepatitis E virus infection. Clin Microbiol Rev 2014;27:116–138.
- Kamar N, Mansuy JM, Cointault O. Hepatitis E virus-related cirrhosis in kidney-pancreas-transplant recipients. Am J Transplant 2008;8:1744–1748.
- Krog JS, Larsen LE, Breum SO. Tracing hepatitis E virus in pigs from birth to slaughter. Front Vet Sci 2019;6:50.
- Krog JS, Larsen LE, Schultz AC. Enteric porcine viruses in farmed shellfish in Denmark. Int J Food Microbiol 2014;186: 105–109.
- Kubacki J, Fraefel C, Jermini M, Giannini P, Martinetti G, Ripellino P, Bernasconi E, Sidler X, Stephan R, Bachofen C. Complete genome sequences of two Swiss hepatitis E virus isolated from human stool and raw pork sausage. Genome Announc 2017;5:e00888-17.
- Lack JB, Volk K, Van Den Bussche RA. Hepatitis E virus genotype 3 in wild rats, United States. Emerg Infect Dis 2012;18:1268–1273.
- La Rosa G, Proroga YTR, De Medici D, Capuano F, Iaconelli M, Della Libera S, Suffredini E. First detection of hepatitis E virus in shellfish and in seawater from production areas in Southern Italy. Food Environ Virol 2018;10:127–131.
- Lee GY, Poovorawan K, Intharasongkroh D, Sa-Nguanmoo P, Vongpunsawad S, Chirathaworn C, Poovorawan Y. Hepatitis E virus infection: Epidemiology and treatment implications. World J Virol 2015;4:343–355.
- Lees D. Viruses and bivalve shellfish. Int J Food Microbiol 2000;59:81–116.
- Li S, Mingxia L, Cong J, Zhou Y, Miao Z. Detection and characterization of hepatitis E virus in goats at slaughterhouse in Tai'an Region, China. Biomed Res Int 2017;2017: 3723650.
- Liu B, Sun Y, Du T, Chen Y, Wang X, Huang B, Li H, Nan Y, Xiao S, Zhang G, Hiscox JA, Zhou EM, Zhao Q. Rabbit hepatitis E virus is an opportunistic pathogen in specificpathogen-free rabbits with the capability of cross-species transmission. Vet Microbiol 2017;201:72–77.

- Loisy F, Atmar RL, Le Saux JC, Cohen J, Caprais MP, Pommepuy M, Le Guyader FS. Use of rotavirus virus-like particles as surrogates to evaluate virus persistence in shellfish. Appl Environ Microbiol 2005;71:6049–6053.
- Long F, Yu W, Yang C, Wang J, Li Y, Li Y, Huang F. High prevalence of hepatitis E virus infection in goats. J Med Virol 2017;89:1981–1987.
- Lu L, Li C, Hagedorn CH. Phylogenetic analysis of global hepatitis E virus sequences: Genetic diversity, subtypes and zoonosis. Rev Med Virol 2006;16:5–36.
- Marek A, Bilic I, Prokofieva I, Hess M. Phylogenetic analysis of avian hepatitis E virus samples from European and Australian chicken flocks supports the existence of a different genus within the Hepeviridae comprising at least three different genotypes. Vet Microbiol 2010;145:54–61.
- Matsubayashi K, Kang JH, Sakata H, Takahashi K, Shindo M, Kato M, Sato S, Kato T, Nishimori H, Tsuji K, Maguchi H, Yoshida J, Maekubo H, Mishiro S, Ikeda H. A case of transfusion-transmitted hepatitis E caused by blood from a donor infected with hepatitis E virus via zoonotic food-borne route. Transfusion 2008;48:1368–1375.
- Matsuda H, Okada K, Takahashi K, Mishiro S. Severe hepatitis E infection after ingestion of uncooked liver from a wild boar. J Infect Dis 2003;188:944.
- McPherson S, Elsharkawy AM, Ankcorn M, Ijaz S, Powell J, Rowe I, Tedder R, Andrews PA. Summary of the British Transplantation Society UK Guidelines for hepatitis E and solid organ transplantation. Transplantation 2018;102:15–20.
- Meng XJ. Expanding host range and cross-species infection of hepatitis E virus. PLoS Pathog 2016;12:e1005695.
- Mesquita JR, Oliveira D, Rivadulla E, Abreu-Silva J, Valera MF, Romalde JL, Nascimento MS. Hepatitis E virus genotype 3 in mussels (*Mytilus galloprovincialis*), Spain. Food Microbiol 2016;58:13–15.
- Mesquita JR, Santos-Ferreira N, Ferreira AS, Albuquerque C, Nóbrega C, Esteves F, Cruz R, Vala H, Nascimento M. Increased risk of hepatitis E virus infection in workers occupationally exposed to sheep. Transbound Emerg Dis 2020;67:1918–1921.
- Milojević L, Velebit B, Teodorović V, Kirbiš A, Petrović T, Karabasil N, Dimitrijević M. Screening and molecular characterization of hepatitis E virus in slaughter pigs in Serbia. Food Environ Virol 2019;11:410–419.
- Montagnaro S, De Martinis C, Sasso S, Ciarcia R, Damiano S, Auletta L, Iovane V, Zottola T, Pagnini U. Viral and antibody prevalence of hepatitis E in European Wild Boars (*Sus scrofa*) and Hunters at Zoonotic Risk in the Latium Region. J Comp Pathol 2015;153:1–8.
- Montalvo Villalba MC, Cruz Martínez D, Ahmad I, Rodriguez Lay LA, Bello Corredor M, Guevara March C, Martínez LS, Martínez-Campo LS, Jameel S. Hepatitis E virus in bottlenose dolphins Tursiops truncatus. Dis Aquat Organ 2017; 123:13–18.
- Montone AMI, De Sabato L, Suffredini E, Alise M, Zaccherini A, Volzone P, Di Maro O, Neola B, Capuano F, Di Bartolo I. Occurrence of HEV-RNA in Italian Regional Pork and Wild Boar Food Products. Food Environ Virol 2019;11:420–426.
- Moor D, Liniger M, Baumgartner A, Felleisen R. Screening of ready-to-eat products for hepatitis E virus in Switzerland. Food Environ Virol 2018;10:263–271.
- Motte A, Roquelaure B, Galambrun C, Bernard F, Zandotti C, Colson P. Hepatitis E in three immunocompromized children in southeastern France. J Clin Virol 2012;53:162–166.

- Nakamura M, Takahashi K, Taira K. Hepatitis E virus infection in wild mongooses of Okinawa, Japan: Demonstration of anti-HEV and a full-genome nucleotide sequence. Hepatol Res 2006;34:137–140.
- Nan Y, Wu C, Zhao Q, Zhou EM. Zoonotic hepatitis E virus: An ignored risk for public health. Front Microbiol 2017;8: 2396.
- O'Hara Z, Crossan C, Craft J, Scobie L. First report of the presence of hepatitis E virus in Scottish-harvested shellfish purchased at retail level. Food Environ Virol 2018;10:217–221.
- Park WJ, Park BJ, Ahn HS, Lee JB, Park SY, Song CS, Lee SW, Yoo HS, Choi IS. Hepatitis E virus as an emerging zoonotic pathogen. J Vet Sci 2016;17:1–11.
- Pavio N, Merbah T, Thébault A. Frequent hepatitis E virus contamination in food containing raw pork liver, France. Emerg Infect Dis 2014;20:1925–1927.
- Pineda JA, Cifuentes C, Parra M, Merchante N, Pérez-Navarro E, Rivero-Juárez A, Monje P, Rivero A, Macías J, Real LM. Incidence and natural history of hepatitis E virus coinfection among HIV-infected patients. AIDS 2014;28:1931– 1937.
- Poizat A, Bonnet-Beaugrand F, Rault A, Fourichon C, Bareille N. Antibiotic use by farmers to control mastitis as influenced by health advice and dairy farming systems. Prev Vet Med 2017;146:61–72.
- Prpić J, Černi S, Škorić D, Keros T, Brnić D, Cvetnić Ž, Jemeršić L. Distribution and molecular characterization of hepatitis E virus in Domestic Animals and Wildlife in Croatia. Food Environ Virol 2015;7:195–205.
- Pupari G, Macaluso G, Di Bella S, Gucciardi F, Mira F, Di Marco P, Lastra A, Petersen E, La Rosa G, Guercio A. Molecular characterization of human enteric viruses in food, water samples, and surface swabs in Sicily. Int J Infect Dis 2019;80:66–72.
- Purdy MA, Harrison TJ, Jameel S, Meng XJ, Okamoto H, Van der Poel W, Smith DB, ICTV Report Consortium. ICTV Virus Taxonomy Profile: *Hepeviridae*. J Gen Virol 2017;98: 2645–2646.
- Raj VS, Smits SL, Pas SD, Provacia LBV, Moorman-Roest H, Osterhaus ADME, Haagmans BL. Novel hepatitis E virus in ferrets, the Netherlands. Emerg Infect Dis 2012;18:1369– 1370.
- Rivadulla E, Valera MF, Mesquita JR, Nascimento MSJ, Romalde JL. Detection of hepatitis E virus in shellfish harvesting areas from Galicia (Northwestern Spain). Viruses 2019;11:618.
- Rivero-Juarez A, Frias M, Martinez-Peinado A, Risalde MA, Rodriguez-Cano D, Camacho A, García-Bocanegra I, Cuenca-López F, Gomez-Villamandos JC, Rivero A. Familial hepatitis E outbreak linked to wild boar meat consumption. Zoonoses Public Health 2017;64:561–565.
- Rivero-Juarez A, Frias M, Rodriguez-Cano D, Cuenca-López F, Rivero A. Isolation of hepatitis E virus from breast milk during acute infection. Clin Infect Dis 2016;62:1464.
- Said B, Ijaz S, Kafatos G, Booth L, Thomas HL, Walsh A, Ramsay M, Morgan D. Hepatitis E incident investigation team. Hepatitis E outbreak on cruise ship. Emerg Infect Dis 2009;15:1738–1744.
- Sarno E, Martin A, McFarland S, Johne R, Stephan R, Greiner M. Estimated exposure to hepatitis E virus through consumption of swine liver and liver sausages. Food Control 2017;73:821–828.

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- Sasaki Y, Haruna M, Uema M, Noda M, Yamada Y. Prevalence and phylogenetic analysis of hepatitis E virus among Pigs in Japan. Jpn J Infect Dis 2018;71:75–78.
- Sayed IM, Elkhawaga AA, El-Mokhtar MA. Circulation of hepatitis E virus (HEV) and/or HEV-like agent in non-mixed dairy farms could represent a potential source of infection for Egyptian people. Int J Food Microbiol 2020;317:108479.
- Sayed IM, Vercouter A, Abdelwahab S, Vercautener K, Meuleman P. Is hepatitis E virus an emerging problem in industrialized countries? Hepatology 2015;62:1883–1892.
- Shirazi R, Pozzi P, Wax M, Bar-Or I, Asulin E, Lusting Y, Mendelson E, Ben-Ari Z, Schwartz E, Mor O. Hepatitis E in pigs in Israel: Seroprevalence, molecular characterization and potential impact on humans. Euro Surveill 2018;23:1800067.
- Singh S, Daga MK, Kumar A, Husain SA, Kar P. Role of oestrogen and its receptors in HEV-associated feto-maternal outcomes. Liver Int 2019;39:633–639.
- Smith DB, Simmonds P, Members of the International Committee on the Taxonomy of Viruses Hepeviridae Study Group, Jameel S, Emerson SU, Harrison TJ, Meng XJ, Okamoto H, Van der Poel W, Purdy MA. Consensus proposal for classification of the family *Hepeviridae*. J Gen Virol 2014;95(Pt 10):2223–2232.
- Spada E, Pupella S, Pisani G, Bruni R, Chionne P, Madonna E, Villano U, Simeoni M, Fabi S, Marano G, Marcantonio C, Pezzotti P, Ciccaglione AR, Liumbruno GM. A nationwide retrospective study on prevalence of hepatitis E virus infection in Italian blood donors. Blood Transfus 2018;16:413–421.
- Sridhar S, Teng JLL, Chiu TH, Lau SKP, Woo PCY. Hepatitis E virus genotypes and evolution: Emergence of camel hepatitis E variants. Int J Mol Sci 2017;18:869.
- Suffredini E, Lanni L, Arcangeli G, Pepe T, Mazzette R, Ciccaglioni G, Croci L. Qualitative and quantitative assessment of viral contamination in bivalve molluscs harvested in Italy. Int J Food Microbiol 2014;184:21–26.
- Suffredini E, Le QH, Di Pasquale S, Pham TD, Vicenza T, Losardo M, To KA, De Medici D. Occurrence and molecular characterization of enteric viruses in bivalve shellfish marketed in Vitnam. Food Control 2020;108:106828.
- Szabo K, Trojnar E, Anheyer-Behmenburg H, Binder A, Schotte U, Ellerbroek L, Klein G, Johne R. Detection of hepatitis E virus RNA in raw sausages and liver sausages from retail in Germany using an optimized method. Int J Food Microbiol 2015;215:149–156.
- Todt D, Meister TL, Steinmann E. Hepatitis E virus treatment and ribavirin therapy: Viral mechanisms of nonresponse. Curr Opin Virol 2018;32:80–87.

- Tritz SE, Khounvisith V, Pommasichan S, Ninnasopha K, Keosengthong A, Phoutana V, Camoin M, Hübschen JM, Black AP, Müller CP, Snoeck CJ, Pauly M. Evidence of increased Hepatitis E virus exposure in Lao villagers with contact to ruminants. Zoonoses Public Health 2018;65:690– 701.
- Vercouter AS, Sayed IM, Lipkens Z, De Bleecker K, De Vliegher S, Colman R, Koppelman M, Supré K, Meuleman P. Absence of zoonotic hepatitis E virus infection in Flemish dairy cows. Int J Food Microbiol 2018;281:54–59.
- Wang J, Li N, Zhang H, Li F, Fanning S, Jiang, T. Detection of hepatitis E virus in the pig livers and retail pork samples collected in selected cities in China. Foodborne Pathog Dis 2021;18:97–103.
- Wang L, Zhang Y, Gong W, Song WT, Wang L. Hepatitis E virus in 3 types of laboratory animals, China, 2012–2015. Emerg Infect Dis 2016;22:2157–2159.
- Wang Y, Ma X. Detection and sequences analysis of sheep hepatitis E virus RNA in Xinjiang Autonomous Region. Wei Sheng Wu Xue Bao Act Microbiol Sinica 2010;50:937–941.
- Weigand K, Weigand K, Schemmerer M, Müller M, Wenzel JJ. Hepatitis E seroprevalence and genotyping in a cohort of wild boars in Southern Germany and Eastern Alsace. Food Environ Virol 2018;10:167–175.
- Widén F. Hepatitis E as a Zoonosis. Adv Exp Med Biol 2016; 948:61–71.
- Woo PC, Lau SK, Teng JL, Cao KY, Wernery U, Schountz T, Chiu TH, Tsang AK, Wong PC, Wong EY, Yuen KY. New hepatitis E virus genotype in Bactrian Camels, Xinjiang, China, 2013. Emerg Infect Dis 2016;22:2219–2221.
- World Health Organization. Hepatitis E vaccine: WHO position paper, May 2015—Recommendations. Vaccine 2016;12;34: 304–305.
- Yugo DM, Meng XJ. Hepatitis E virus: Foodborne, waterborne and zoonotic transmission. Int J Environ Res Public Health 2013;10:4507–4533.

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