



# Metoclopramide-Induced Acute Dystonic Reactions May Be Associated With the CYP2D6 Poor Metabolizer Status and Pregnancy-Related Hormonal Changes

#### Eng Wee Chua<sup>1\*</sup>, Simon P. Harger<sup>2</sup> and Martin A. Kennedy<sup>3</sup>

<sup>1</sup> Faculty of Pharmacy, Universiti Kebangsaan Malaysia, Kuala Lumpur, Malaysia, <sup>2</sup> Hawke's Bay District Health Board, Hastings, New Zealand, <sup>3</sup> Carney Centre for Pharmacogenomics and Department of Pathology and Biomedical Science, University of Otago, Christchurch, Christchurch, New Zealand

**OPEN ACCESS** 

#### Edited by:

Elena Ramírez, University Hospital La Paz, Spain

#### Reviewed by:

Nathalie K. Zgheib, American University of Beirut, Lebanon Amit P. Bhavsar, University of Alberta, Canada

\*Correspondence:

Eng Wee Chua cew85911@ukm.edu.my

#### Specialty section:

This article was submitted to Translational Pharmacology, a section of the journal Frontiers in Pharmacology

**Received:** 20 May 2019 **Accepted:** 22 July 2019 **Published:** 22 August 2019

#### Citation:

Chua EW, Harger SP and Kennedy MA (2019) Metoclopramide-Induced Acute Dystonic Reactions May Be Associated With the CYP2D6 Poor Metabolizer Status and Pregnancy-Related Hormonal Changes. Front. Pharmacol. 10:931. doi: 10.3389/fphar.2019.00931 We report two cases of metoclopramide-induced acute dystonia in pregnant women and consider the role of genetic variation in the pathogenesis of the adverse effect. By whole-gene sequencing, we found that both women were CYP2D6 poor metabolizers. We theorize that CYP2D6 governs the risk of metoclopramide-related acute dystonia through its role in the synthesis of serotonin, which inhibits the dopamine tone. The effect of CYP2D6 poor metabolism is exaggerated by rises in the estrogen levels during pregnancy, as the hormone augments dopamine sensitivity. Together, the two factors may create a hyper-dopaminergic state that is easily upset by metoclopramide, resulting in acute dystonia.

Keywords: metoclopramide, acute dystonia, CYP2D6, poor metabolizer, pregnancy, estrogen, dopamine

# INTRODUCTION

Metoclopramide is a commonly used antiemetic agent. It improves gut movement partly through its antagonistic action on dopamine, which has an inhibitory effect on the gut. However, because it disrupts central dopaminergic signaling, metoclopramide may produce rare movement disorders, such as acute dystonic reactions (Albibi and McCallum, 1983; Bateman et al., 1985). The incidence rate of metoclopramide-induced acute dystonia and akathisia has been reported as 0.5% (Bateman et al., 1989). To date, there is a dearth of studies that examined the genetic basis of metoclopramide-induced acute dystonia. Only one case report detailed the presence of the deleterious *CYP2D6\*4* allele in two patients who were given metoclopramide for chemotherapy-induced nausea (van der Padt et al., 2006). The allele involves a guanine-to-adenine transition within the intron 3-exon 4 boundary of *CYP2D6*, shifting the position of an "AG" acceptor site, illustrated as follows: 5'-CCACCCCCAGGACGCCCCTT-3' to 5'-CCACCCCCAAGACGCCCCTT-3'. This causes erroneous splicing and deletion of one nucleotide from the fourth exon, thereby altering the ensuing reading frame. Consequently, a non-functional protein is produced (Hanioka et al., 1990).

It was thought that impaired CYP2D6 function reduced the clearance of metoclopramide and substantially increased the risk of acute dystonia caused by the drug (van der Padt et al., 2006). However, the assumption might have overlooked other factors that could also contribute to the development of the adverse effect. In the *CYP2D6*–metoclopramide case report, the chemotherapy

1

that the patients received could have been equally culpable of the dystonic reactions. Chemotherapeutic agents have been reported to cause dystonia, possibly by adversely affecting the movement-related brain circuits, i.e., those housed in the basal ganglia (Brashear and Siemers, 1997).

In this work, we consider two cases of metoclopramidetriggered acute dystonia and the role of *CYP2D6* variation in the pathogenesis of the adverse effect. These cases were referred to us for genetic analysis, as part of a long-term initiative (<u>Understanding Adverse Drug Reactions Using Genomic Sequencing</u>, or UDRUGS) to systematically accrue cases of unusual adverse drug reactions in a biobank (Maggo et al., 2017). Based on the participants' medical history and their *CYP2D6* and *SULT2A1* genotypes, we provide new insights into the relation between CYP2D6 poor metabolism, pregnancy-related hormonal changes, and acute dystonia caused by metoclopramide.

# TWO CASES OF METOCLOPRAMIDE-INDUCED ACUTE DYSTONIA

#### Case 1

Participant A, a 39-year-old female of European descent, was 12 weeks pregnant when the dystonic reaction occurred. She was taking oral metoclopramide 10 mg three times daily for pregnancyassociated hyperemesis, and not long thereafter (the exact interval was unclear), she started experiencing back spasms; however, she was not aware that metoclopramide was causing the adverse effect. After about 1 day of oral therapy, the vomiting persisted. She was given an additional 10-mg dose intravenously, and 3-4 h later, the back spasms developed into a full-blown dystonic reaction, resulting in torticollis and globally increased muscle tone. Also, she was unable to speak because her throat was "closing down." She received treatment at a local hospital, and the reaction resolved after she was given three 0.5-mg intravenous doses of benztropine, which is an anticholinergic agent. On arrival at the hospital, her vital signs were normal. She was afebrile, with a heart rate of 80/ min and an oxygen saturation level of 100%. Overall, we estimated that the gap between the first dose of metoclopramide and the occurrence of the acute dystonia was 1-2 days.

The participant consented to provide a blood sample for genetic analysis. We used a modified Pharmacist's Workup of Drug Therapy form (Strand et al., 1988) to document her medical history. She was otherwise healthy except for a history of atopy. The only additional medication that she was taking was oral folic acid, which has no known interactions with metoclopramide. Using the Naranjo algorithm (Naranjo et al., 1981), we rated the causality of the adverse reaction as *possible*.

#### Case 2

Participant B, a 32-year-old female of European descent, was 9–10 weeks pregnant when the dystonic reaction occurred. She was taking oral metoclopramide three times daily for pregnancy-associated nausea; the exact dosage was unknown. About 2 days after she started taking metoclopramide, she experienced the following symptoms: anxiety, jitteriness, involuntary upward

movement of her eyes, neck spasms, and back arching. The symptoms subsided after she was given two doses of an "anti-Parkinsonian" agent. Because we had no access to the participant's medical records, we did not have the details of her vital signs or other relevant physical examination findings for when she was treated for the adverse reaction.

The participant learned of UDRUGS by word of mouth, and she volunteered to take part in the project. We documented her medical history using the modified Pharmacist's Workup of Drug Therapy form (Strand et al., 1988). She was healthy and had no underlying illnesses. The only medication that she was taking alongside metoclopramide was prenatal vitamins. The participant also described to us that she had previously taken metoclopramide at a lower dosage for a month (once instead of thrice daily), but she did not develop a reaction. Interestingly, she was not pregnant at the time. Using the Naranjo algorithm (Naranjo et al., 1981), we rated the adverse reaction as being *possibly* linked to metoclopramide.

## **GENETIC ANALYSIS**

We obtained blood samples from the two participants, extracted DNA, and Sanger-sequenced the CYP2D6 and SULT2A1 genes. The CYP2D6 genotypes were determined using a previously reported method (Wright et al., 2010), whereby two duplex long PCRs were performed to detect CYP2D6 deletion and duplication alleles and to amplify the entire gene (6.6 kb; Supplementary Figure 1). The co-amplification of a 3.5-kb product would indicate whole-gene deletion (\*5) or duplication; but the product was absent from all the long PCRs performed for the participants. Therefore, we concluded that both participants did not harbor the duplication or the deletion allele. Using the diluted long amplicon as a template, we then performed touchdown PCRs to generate short, overlapping fragments that spanned CYP2D6 and a portion of its promoter (Supplementary Table 1). We then Sanger-sequenced the resultant PCR products. We used a similar approach for SULT2A1 sequencing and performed touchdown PCRs to generate products that covered the promoter region and all six exons of SULT2A1 (Supplementary Table 2). The details of all the PCRs can be found in Supplementary Methods.

By whole-gene sequencing, we found that both participants were homozygous for the loss-of-function *CYP2D6\*4* allele (**Supplementary Figure 2**). In other words, the participants were CYP2D6 poor metabolizers. We did not identify any deleterious variants in their *SULT2A1* gene. Therefore, they were likely to have normal SULT2A1 function.

## DISCUSSION

The acute dystonic reactions experienced by our participants were similar to those documented in other reports, namely, sustained contraction of the neck muscles, causing an arched back, and involuntary eye movements (Leus and van de Ven, 2015; Tianyi et al., 2017). Few studies have examined the genetic underpinnings of metoclopramide's adverse effects (van der Padt

et al., 2006; Parkman et al., 2012). To date, only CYP2D6 variation has been linked to the risk of metoclopramide-related acute dystonia (van der Padt et al., 2006). It was believed that deficient CYP2D6 function increased the exposure to metoclopramide, though this was not confirmed by measurement of the drug levels (van der Padt et al., 2006). However, considering the current understanding of metoclopramide's pharmacokinetics, we surmise that this is unlikely. Approximately 20% of metoclopramide is excreted unchanged, and 50% is renally eliminated in the conjugated form (Graffner et al., 1979). The major urinary metabolite is a sulfate conjugate, formed by SULT2A1. The de-ethylation and mono-oxygenation reactions, mediated largely by CYP2D6, are relatively minor and not believed to have a significant impact on plasma metoclopramide levels in vivo (Bateman et al., 1980; Desta et al., 2002; Senggunprai et al., 2009; Livezey et al., 2014). A previous study in children detected no relationship between metoclopramide levels and the occurrence of acute dystonia (Bateman et al., 1983).

Therefore, it seems more probable that the CYP2D6– metoclopramide relation is pharmacodynamic, not pharmacokinetic, in nature. CYP2D6 may regulate the central dopamine tone through its role in converting 5-methoxytryptamine to serotonin (Yu et al., 2003). Compared with the extensive metabolizers, the CYP2D6 poor metabolizers may have an inherently lower serotonin tone, allowing an unbridled increase in the dopamine tone. In a variety of studies, the interaction between the two neurotransmitters was thought to account for the association between CYP2D6 function and personality (Peñas-Lledó and Llerena, 2014). This suggests that variation in CYP2D6 activity is indeed accompanied by substantial biochemical changes in the brain.

However, impaired CYP2D6 function alone may not increase the dopamine tone sufficiently to trigger a major dystonic reaction in individuals taking metoclopramide. The influence of other physiological factors, such as sex hormone function, must also be present (Bateman et al., 1989). Otherwise, the incidence of the adverse effect would have been much higher, given the prevalence of the deleterious CYP2D6\*4 allele among Caucasians; the reported allele frequency was ~21% (Sachse et al., 1997). This hypothesis is supported by Case 2 participant's reported tolerance of metoclopramide when she was not pregnant and the known association between oral contraceptive use and acute extrapyramidal reactions (Bateman et al., 1985). Estrogen can augment dopamine sensitivity by upregulating the dopamine receptors (Hruska and Silbergeld, 1980). Moreover, pregnancy-related physiological changes may decrease the CYP2D6 activity further in the poor metabolizers (Wadelius et al., 1997). Hence, we theorize that the combined influence of CYP2D6 poor metabolism and pregnancyinduced rises in the estrogen levels may create a delicate, hyperdopaminergic state that is easily upset by metoclopramide (Askenasy et al., 1978), resulting in acute dystonia (Figure 1).

Nevertheless, it is worth noting that the presence of *CYP2D6\*4* in our participants could have been coincidental. In addition, the control of the number of dopamine receptors by estrogen



appears to be a dynamic process. Administration of estradiol was shown in a study involving ovariectomized monkeys to prevent antipsychotic agents from inducing a compensatory increase in the dopamine receptors and causing "dopamine supersensitivity" (Bédard and Boucher, 1986).

# CONCLUSIONS

The two cases provide further evidence that the CYP2D6 poor metabolizer status may be associated with metoclopramideinduced dystonia and that the role of other mediating factors, including hormonal changes, warrants further investigation. Prescribing an alternative antiemetic agent to the CYP2D6 poor metabolizers could help to prevent the distressing adverse reaction. Future work may focus on assembling a cohort for confirming the association between CYP2D6 variation and metoclopramide-induced dystonia. Measuring the blood concentrations of metoclopramide is necessary to ascertain the nature of the drug-CYP2D6 interaction. It may also be worthwhile to investigate other factors that could drive the pathogenesis of metoclopramide-induced dystonia, such as age, gender, DNA variation in non-metabolic genes (e.g., dopamine receptor gene polymorphisms), and co-administered drugs. Finally, the costeffectiveness of selectively prescribing metoclopramide based on CYP2D6 genotypes should be determined.

# DATA AVAILABILITY

The raw data supporting the conclusions of this manuscript will be made available by the authors, without undue reservation, to any qualified researcher.

## REFERENCES

- Albibi, R., and McCallum, R. W. (1983). Metoclopramide: pharmacology and clinical application. Ann. Intern. Med. 98 (1), 86–95. doi: 10.7326/0003-4819-98-1-86
- Askenasy, J. J., Streifler, M., and Felner, S. (1978). The synaptic significance of metoclopramide induced dyskinetic–dystonic head and neck movements in pregnancy. J. Neural. Transm. 42 (1), 73–84. doi: 10.1007/BF01262731
- Bateman, D. N., Craft, A. W., Nicholson, E., and Pearson, A. D. (1983). Dystonic reactions and the pharmacokinetics of metoclopramide in children. Br. J. Clin. Pharmacol. 15 (5), 557–559. doi: 10.1111/j.1365-2125.1983.tb02090.x
- Bateman, D. N., Darling, W. M., Boys, R., and Rawlins, M. D. (1989). Extrapyramidal reactions to metoclopramide and prochlorperazine. QJM 71 (1), 307–311. doi: 10.1093/oxfordjournals.qjmed.a068325
- Bateman, D. N., Kahn, C., and Davies, D. S. (1980). The pharmacokinetics of metoclopramide in man with observations in the dog. Br. J. Clin. Pharmacol. 9 (4), 371–377. doi: 10.1111/j.1365-2125.1980.tb01064.x
- Bateman, D. N., Rawlins, M. D., and Simpson, J. M. (1985). Extrapyramidal reactions with metoclopramide. Br. Med. J. (Clin. Res. Ed.) 291 (6500), 930– 932. doi: 10.1136/bmj.291.6500.930
- Bédard, P. J., and Boucher, R. (1986). Estradiol can suppress haloperidol-induced supersensitivity in dyskinetic monkeys. *Neurosci. Lett.* 64 (2), 206–210. doi: 10.1016/0304-3940(86)90101-1
- Brashear, A., and Siemers, E. (1997). Focal dystonia after chemotherapy: a case series. J. Neurooncol. 34 (2), 163–167. doi: 10.1023/A:1005791623733
- Desta, Z., Wu, G. M., Morocho, A. M., and Flockhart, D. A. (2002). The gastroprokinetic and antiemetic drug metoclopramide is a substrate and

## **ETHICS STATEMENT**

The project was approved by the Southern Health and Disability Ethics Committee (New Zealand). Written consent to take part in this study was obtained from the participants by face-to-face interviews. They understood that the result of their genetic analysis, alongside their clinical data, might be published in a journal article; however, their identity would remain confidential.

# **AUTHOR CONTRIBUTIONS**

EWC and MK recruited the participants. EWC performed the experiments and wrote the manuscript. MK and SH reviewed the initial and final drafts of the manuscript.

## ACKNOLWEDGMENTS

The authors would like to thank Dr. Norazrina Azmi (Faculty of Pharmacy, Universiti Kebangsaan Malaysia) for her valuable inputs on the relation between estrogen, central dopaminergic signaling, and metoclopramide-related acute dystonic reactions.

# SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2019.00931/full#supplementary-material

inhibitor of cytochrome P450 2D6. Drug Metab. Dispos. 30 (3), 336–343. doi: 10.1124/dmd.30.3.336

- Graffner, C., Lagerström, P. O., Lundborg, P., and Rönn, O. (1979). Pharmacokinetics of metoclopramide intravenously and orally determined by liquid chromatography. *Br. J. Clin. Pharmacol.* 8 (5), 469–474. doi: 10.1111/j.1365-2125.1979.tb01028.x
- Hanioka, N., Kimura, S., Meyer, U. A., and Gonzalez, F. J. (1990). The human CYP2D locus associated with a common genetic defect in drug oxidation: A G1934 A base change in intron 3 of a mutant CYP2D6 allele results in an aberrant 3' splice recognition site. Am. J. Hum. Genet. 47 (6), 994–1001.
- Hruska, R. E., and Silbergeld, E. K. (1980). Increased dopamine receptor sensitivity after estrogen treatment using the rat rotation model. *Science* 208 (4451), 1466– 1468. doi: 10.1126/science.7189902
- Leus, M., and van de Ven, A. (2015). An acute dystonic reaction after treatment with metoclopramide. *N. Engl. J. Med.* 373 (14), e16. doi: 10.1056/NEJMicm1412207
- Livezey, M. R., Briggs, E. D., Bolles, A. K., Nagy, L. D., Fujiwara, R., and Furge, L. L. (2014). Metoclopramide is metabolized by CYP2D6 and is a reversible inhibitor, but not inactivator, of CYP2D6. *Xenobiotica* 44 (4), 309–319. doi: 10.3109/00498254.2013.835885
- Maggo, S. D., Chua, E. W., Chin, P., Cree, S., Pearson, J., and Doogue, M. (2017). A New Zealand platform to enable genetic investigation of adverse drug reactions. N. Z. Med. J. 130 (1466), 62–69.
- Naranjo, C. A., Busto, U., Sellers, E. M., Sandor, P., Ruiz, I., and Roberts, E. A. (1981). A method for estimating the probability of adverse drug reactions. *Clin. Pharmacol. Ther.* 30 (2), 239–245. doi: 10.1038/clpt.1981.154
- Parkman, H. P., Mishra, A., Jacobs, M., Pathikonda, M., Sachdeva, P., and Gaughan, J. (2012). Clinical response and side effects of metoclopramide:

associations with clinical, demographic, and pharmacogenetic parameters. *J. Clin. Gastroenterol.* 46 (6), 494–503. doi: 10.1097/MCG.0b013e3182522624

- Peñas-Lledó, E. M., and Llerena, A. (2014). CYP2D6 variation, behaviour and psychopathology: implications for pharmacogenomics-guided clinical trials. *Br. J. Clin. Pharmacol.* 77 (4), 673–683. doi: 10.1111/bcp.12227
- Sachse, C., Brockmöller, J., Bauer, S., and Roots, I. (1997). Cytochrome P450 2D6 variants in a Caucasian population: allele frequencies and phenotypic consequences. Am. J. Hum. Genet. 60 (2), 284–295.
- Senggunprai, L., Yoshinari, K., and Yamazoe, Y. (2009). Selective role of sulfotransferase 2A1 (SULT2A1) in the N-sulfoconjugation of quinolone drugs in humans. *Drug Metab. Dispos.* 37 (8), 1711–1717. doi: 10.1124/dmd.109.027441
- Strand, L. M., Cipolle, R. J., and Morley, P. C. (1988). Documenting the clinical pharmacist's activities: back to basics. *Drug Intell. Clin. Pharm.* 22 (1), 63–67. doi: 10.1177/106002808802200116
- Tianyi, F. L., Agbor, V. N., and Njim, T. (2017). Metoclopramide induced acute dystonic reaction: a case report. BMC Res. Notes 10 (1), 32. doi: 10.1186/s13104-016-2342-6
- van der Padt, A., van Schaik, R. H., and Sonneveld, P. (2006). Acute dystonic reaction to metoclopramide in patients carrying homozygous cytochrome P450 2D6 genetic polymorphisms. *Neth. J. Med.* 64 (5), 160–162.
- Wadelius, M., Darj, E., Frenne, G., and Rane, A. (1997). Induction of CYP2D6 in pregnancy. *Clin. Pharmacol. Ther.* 62 (4), 400–407. doi: 10.1016/ S0009-9236(97)90118-1
- Wright, G. E. B., Niehaus, D. J. H., Drögemöller, B. I., Koen, L., Gaedigk, A., and Warnich, L. (2010). Elucidation of CYP2D6 genetic diversity in a unique

African population: implications for the future application of pharmacogenetics in the Xhosa population. *Ann. Hum. Genet.* 74 (4), 340–350. doi: 10.1111/j. 1469-1809.2010.00585.x

Yu, A. M., Idle, J. R., Byrd, L. G., Krausz, K. W., Küpfer, A., and Gonzalez, F. J. (2003). Regeneration of serotonin from 5-methoxytryptamine by polymorphic human CYP2D6. *Pharmacogenetics* 13 (3), 173–181. doi: 10.1097/00008571-200303000-00007.

**Conflict of Interest Statement:** EC and MK received funding from the University of Otago, Christchurch, for the submitted work; there were no financial relationships with any organizations that might have an interest in the submitted work in the previous 3 years; and there were no other relationships or activities that could appear to have influenced the submitted work.

The remaining author declares that the research was conducted in the absence of any commercial or financial relationships that could be constructed as a potential conflict of interest.

Copyright © 2019 Chua, Harger and Kennedy. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.