Catechol-O-Methyltransferase Val158Met Polymorphism Modulates Gray Matter Volume and Functional Connectivity of the Default Mode Network

Tian Tian¹, Wen Qin¹, Bing Liu^{2,3}, Dawei Wang¹, Junping Wang¹, Tianzi Jiang^{2,3,4,5}, Chunshui Yu^{1*}

1 Department of Radiology and Tianjin Key Laboratory of Functional Imaging, Tianjin Medical University General Hospital, Tianjin, China, **2** Brainnetome Center, Institute of Automation, Chinese Academy of Sciences, Beijing, China, **3** National Laboratory of Pattern Recognition, Institute of Automation, Chinese Academy of Sciences, Beijing, China, **4** Key Laboratory for NeuroInformation of Ministry of Education, School of Life Science and Technology, University of Electronic Science and Technology of China, Chengdu, China, **5** The Queensland Brain Institute, the University of Queensland, Brisbane, Australia

Abstract

The effect of catechol-O-methyltransferase (COMT) Val158Met polymorphism on brain structure and function has been previously investigated separately and regionally; this prevents us from obtaining a full picture of the effect of this gene variant. Additionally, gender difference must not be overlooked because estrogen exerts an interfering effect on COMT activity. We examined 323 young healthy Chinese Han subjects and analyzed the gray matter volume (GMV) differences between Val/Val individuals and Met carriers in a voxel-wise manner throughout the whole brain. We were interested in genotype effects and genotype × gender interactions. We then extracted these brain regions with GMV differences as seeds to compute resting-state functional connectivity (rsFC) with the rest of the brain; we also tested the genotypic differences and gender interactions in the rsFCs. Val/Val individuals showed decreased GMV in the posterior cingulate cortex (PCC) compared with Met carriers; decreased GMV in the medial superior frontal gyrus (mSFG) was found only in male Val/Val subjects. The rsFC analysis revealed that both the PCC and mSFG were functionally correlated with brain regions of the default mode network (DMN). Both of these regions showed decreased rsFCs with different parts of the frontopolar cortex of the DMN in Val/Val individuals than Met carriers. Our findings suggest that the COMT Val158Met polymorphism modulates both the structure and functional connectivity within the DMN and that gender interactions should be considered in studies of the effect of this genetic variant, especially those involving prefrontal morphology.

Citation: Tian T, Qin W, Liu B, Wang D, Wang J, et al. (2013) Catechol-O-Methyltransferase Val158Met Polymorphism Modulates Gray Matter Volume and Functional Connectivity of the Default Mode Network. PLoS ONE 8(10): e78697. doi:10.1371/journal.pone.0078697

Editor: Hengyi Rao, University of Pennsylvania, United States of America

Received April 20, 2013; Accepted September 16, 2013; Published October 16, 2013

Copyright: © 2013 Tian et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by the National Basic Research Program of China (973 program, No. 2011CB707801), the Natural Science Foundation of China (Grant Nos. 81271551 and 81061120533), and the Natural Science Foundation of Tianjin (No. 11JCZDJC19300). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

* E-mail: chunshuiyu@tijmu.edu.cn

Introduction

Catechol-O-methyltransferase (COMT) catalyzes the degradation of synaptic dopamine (DA) in the brain, especially in the prefrontal cortex (PFC), where COMT may account for more than half of DA decline because of the lack of DA transporter in PFC synapses [1,2]. The COMT gene is located on chromosome 22q11 and contains a functional polymorphism (Val158Met) that results in a fourfold decrease in enzymatic activity at body temperature in Met-allele carriers [1]. Decreased enzymatic activity leads to increased synaptic DA concentrations, which may affect cognitive and emotional functions via modulation of brain structure and function, especially in the PFC [3-9]. However, dopaminergic modulation of phenotypes is complex and has been described as an

inverted U-shaped relationship [10,11] in which both the lowest and highest DA levels may impair behavioral performance [12-15].

Although Met allele carriers exhibit better performance in episodic memory and executive functions [16-26], Val carriers show increased activation of the prefrontal cortex during a variety of cognitive tasks [17,21,27,28]. In contrast, Val carriers exhibit better performance in emotion processing tasks [29,30]. These individuals show greater activation during emotional awareness [30] and regulation tasks [31]; however, they exhibit decreased limbic and prefrontal reactivity [32,33] and prefrontal-limbic connectivity [32,34] while processing unpleasant stimuli. These inverse effects have been ascribed to selective modulation of COMT Val158Met on prefrontal dopamine-associated processing and opposing effects of the Val158Met genotype on stable and flexible demands of cognition [35,36]. The effects of COMT Val158Met on brain function have also been investigated during non-task states. Resting-state electroencephalogram has revealed that Val/Val individuals exhibit lower baseline prefrontal activation [37] but greater connectivity strengths between frontal and temporal/ parietal areas [38]. Moreover, previous resting-state fMRI studies have reported that Val/Val individuals showed weaker prefrontal-related resting-state functional connectivity (rsFC) in the default mode network (DMN) [39] and greater prefrontalrelated rsFC in the executive control network than Met carriers [40]. Beyond the modulation of COMT Val158Met on functional brain characteristics, the Val158Met polymorphism has been shown to affect structural profiles of the brain. Most structural MRI studies have suggested that the COMT Val158Met was related to structural differences in healthy people [4,41-50]; however, one recent study reported no difference in grav matter volume (GMV) between genotypes in healthy young adults [51].

As mentioned above, most previous studies performed either structural or functional analysis, but to date, these methods have not been combined to provide a complete picture of the effect of COMT Val158Met polymorphism on the brain in healthy individuals. Only one study of healthy young men has combined GMV and rsFC analyses; these authors reported that the COMT Val158Met polymorphism affected the rsFC of the PFC, although this difference occurred in the absence of any alterations in gray matter [40]. Another study of healthy children has combined GMV and activation analyses; these authors reported increased GMV in the left hippocampal head and increased activation in the right parahippocampal gyrus during emotional processing in Met carriers [52]. Inspired by these multimodal imaging studies aiming at the structure-functional interactions [53], we will explore whether the rsFC differences between genotypes are driven by the corresponding structural differences. Additionally, most previous studies have followed a hypothesis-driven method and focused on regional changes, especially in the PFC and hippocampus. In the present study, we combined voxel-based structural (GMV) and rsFC analyses throughout the whole brain to characterize the structural and functional differences between Val158Met genotypes.

Estrogen can down-regulate COMT activity [54,55], resulting in decreased COMT activity in females compared with that in males. After Gogos and colleagues demonstrated the gender effects on DA levels and behaviors in COMT knockout mice [56], gender differences in the effects of COMT Val158Met on structural brain characteristics [50] and behavioral performance [23,57-62] have also been identified in healthy subjects. However, it should be noted that several previous studies have reported that there are no gender differences in brain structure and behavioral performance [51,63]. As a potential interacting factor, gender differences must not be ignored when considering the impact of COMT Val158Met on the human brain.

In the present study, we used a voxel-based morphometry (VBM) technique to investigate both the genotypic effects and gender interactions on GMV throughout the whole brain. We then extracted brain regions with GMV differences as seed

regions to compute rsFC with the rest of the brain; we also tested group differences and gender interactions in the rsFCs. Notably, we only recruited healthy young adults aged 18-31 years because previous studies have reported the interaction between COMT Val158Met genotype and age on brain structure [64] and functions [20,65,66].

Materials and Methods

Subjects

A total of 323 healthy young right-handed subjects (mean age: 22.7 ± 2.5 years; 157 males) participated in this study. Participants were carefully screened to ensure that there was no history of psychiatric or neurological illness, psychiatric treatment, or drug or alcohol abuse, and had no contraindications for MRI examinations. Only Chinese Han subjects were included to avoid artifacts of population stratification. All subjects were strongly right-handed according to the Chinese edition of the Edinburgh Handedness Inventory [67]. After a complete description of the study, all subjects provided written informed consent. The protocol was approved by the Ethics Committee of Tianjin Medical University. Subject intelligence quotient (IQ) scores were measured using the Chinese Revised Wechsler Adult Intelligence Scale (WAIS-RC) [68]. Depression scores were evaluated with the Beck Depression Inventory (BDI) [69]. Anxiety levels were obtained using the Self-Rating Anxiety Scale (SAS) [70]. The Tridimensional Personality Questionnaire (TPQ) was used to quantify each temperamental characteristic [71]. The above behavioral scales are known to reflect structural and functional characteristics of the brain [45,58,72-74] and were compared between genotypes. All data are stored according to the requirements of our hospital at the Tianjin Key Laboratory of Functional Imaging, where the experiments were done. These data are not publicly deposited.

Genotyping

We extracted genomic DNA from 3000 µl of whole blood using the EZgeneTM Blood gDNA Miniprep Kit (Biomiga Inc, San Diego, CA, USA). We then genotyped the COMT rs4680 in each subject using the PCR and ligation detection reaction (LDR) method [75,76] with technical support from the Shanghai Biowing Applied Biotechnology Company. The PCR primer sequences of COMT were as follows: forward: 5' GGGCCTACTGTGGCTACTCA 3'. and reverse: 5' CCCTTTTTCCAGGTCTGACA 3'. PCR was performed at a 20 μL volume containing 1 μL genomic DNA, 0.4 μL primer mixture, 2 µL dNTPs, 0.6 µL Mg2+, 2 µL buffer, 4 µL Q-Solution, and 0.3 µL Tag DNA polymerase. The amplification protocol consisted of an initial denaturation and enzyme activation phase at 95°C for 15 min, followed by 35 cycles of denaturation at 94°C for 30 sec, annealing at 59°C for COMT rs4680 for 1 min and 30 sec, extension at 72°C for 1 min, and then a final extension at 72°C for 7 min. PCR products were verified in 3% agarose gels that had been stained with ethidium bromide to regulate the amount of DNA added to the LDR.

Three probes were designed for the LDR reactions for each SNP, including one common probe (rs4680_modify: P-

Table 1. The number of samples collected, gender ratio and distributions of COMT Val158Met polymorphism.

Gender	N (%)	COMT gen	otypes(%)	Allele (%)		
		Val/Val	Val/Met	Met/Met	Val	Met
Male	144	64 (44 44)	62 (43.06)	18 (12.50)	190	98
	(47.68)	64 (44.44)			(65.97)	(34.03)
Female	158	77 (40 72)	69 (43.67)	12 (7.59)	223	93
	(52.32)	77 (48.73)			(70.57)	(29.43)
Total	302	141	131	20 (0 02)	413	191
	(100.00)	(46.69)	(43.38)	30 (9.93)	(68.38)	(31.62)

doi: 10.1371/journal.pone.0078697.t001

(rs4680 A: the SNP TTTTTTTTTTTTTTTTTCAGGCATGCACACCTTGTCCTT CAT: and rs4680 G: TTTTTTTTTTTTTTTTTTCAGGCATGCACACCTTGTCCT TCAC). These reactions were carried out in a 10 µL mixture containing 1 µL buffer, 1 µL probe mix, 0.05 µL Taq DNA ligase, 1 µL PCR product, and 6.95 µL deionized water. The reaction program consisted of initial heating at 95°C for 2 min, followed by 35 cycles of 30 sec at 94°C and 2 min at 50°C. Reactions were stopped by chilling the tubes in an ethanol-dry ice bath and adding 0.5 mL of 0.5 mM EDTA. Aliquots of 1 µL of the reaction products were mixed with 1 µL of loading buffer (83% formamide, 8.3 mM EDTA and 0.17% blue dextran) and 1 µL ABI GS-500 Rox-Fluorescent molecular weight marker, which were denatured at 95°C for 2 min and chilled rapidly on ice prior to being loaded on a 5 Murea-5% polyacrylamide gel and electrophoresed on an ABI 3100 DNA sequencer at 3000 V. Finally, fluorescent ligation products were analyzed and quantified using the ABI GeneMapper software.

Twenty-one of the 323 subjects were excluded from further analysis due to genotyping failure. The COMT rs4680 genotype distribution in the sample was in Hardy–Weinberg equilibrium (P > 0.05). The frequencies of the COMT genotypes are presented in Table 1. No genotype distribution differences were found between males and females. Subjects who were homozygous and heterozygous for the A-allele were merged into a group of A-allele carriers and compared with homozygotes for the G-allele; this method has been used previously to address skewed genotypic distributions [45,48,77,78].

Image acquisition

MR images were acquired using a Signa HDx 3.0 Tesla MR scanner (General Electric, Milwaukee, WI, USA). Tight but comfortable foam padding was used to minimize head motion and earplugs were used to reduce scanner noise. Resting-state functional MRI data were obtained using the Single-Shot Echo-Planar Imaging sequence (SS-EPI) with the following imaging parameters: repetition time (TR)/echo time (TE) = 2000/30 ms; field of view (FOV) = 240 mm × 240 mm; matrix = 64 × 64; flip angle (FA) = 90°, slice thickness = 4 mm; no gap; 40

transversal slices; 180 volumes. During fMRI scans, all subjects were instructed to keep their eyes closed, to stay as motionless as possible, to think of nothing in particular, and not to fall asleep. Sagittal 3D T1-weighted images were acquired by a brain volume (BRAVO) sequence (TR/TE = 8.1/3.1 ms; inversion time = 450 ms; FA = 13° ; FOV = 256 mm × 256 mm; matrix = 256×256 ; slice thickness = 1 mm, no gap; 176 sagittal slices).

Data preprocessing

All structural images were carefully checked slice by slice. Three of the remaining 302 subjects were excluded due to poor image quality or visible structural abnormality. Thus, a total of 299 subjects were included in the VBM analysis. The structural MR images were segmented into gray matter (GM), white matter and cerebrospinal fluid (CSF) using the new segmentation model in the Statistical Parametric Mapping software package (SPM8, http://www.fil.ion.ucl.ac.uk/spm). The new segmentation model is an extension of the "unified segmentation" algorithm [79], which includes additional tissue probability maps to better model CSF and other non-brain voxels, resulting in a more accurate segmentation. Following segmentation, GM population templates were generated from the entire image dataset using diffeomorphic anatomical registration through the exponentiated Lie algebra (DARTEL) technique [80]. After an initial affine registration of the GM DARTEL template to the tissue probability map in the Montreal Neurological Institute (MNI) space (http://www.mni.mcgill.ca/), non-linear warping of GM images was performed to the DARTEL GM template in the MNI space with a resolution of 1.5-mm³ (as recommended for the DARTEL procedure). The GMV of each voxel was obtained by multiplying the GM concentration map by the non-linear determinants derived during spatial normalization. Finally, to compensate for residual between-subjects anatomical differences, the GMV images were smoothed with a full width at half maximum (FWHM) kernel of 8 mm. In effect, the regional GMV represents normalized GMVs after removing the confounding effects of variance in individual brain sizes. After spatial pre-processing, normalized, modulated, and smoothed GMV maps were used for statistical analysis.

Each of the functional MR images from the 299 subjects was carefully checked slice by slice to exclude scans with obvious distortions, signal loss, or artifacts. Functional MRI data for 299 subjects were preprocessed using the Data Processing Assistant for Resting-State fMRI (DPARSF) [81]. The first 10 volumes of each subject were discarded for signal equilibrium and participants' adaptation to scanning noise. The remaining 170 volumes were first corrected for the acquisition time delay between slices. Head motion parameters were estimated and each volume was realigned to the mean whole volume map to correct for geometrical displacements using a six-parameter rigid-body transformation. Eleven participants were excluded from further analysis because their maximum displacement in any of the orthogonal directions (x, y, z) more than 2 mm, or there was a maximum rotation (x, y, z) greater than 2.0°. Thus, a total of 288 subjects were included in further rsFC preprocessing. We also calculated framewise displacement (FD), which indexes volume-to-volume changes in head position. These changes were obtained from derivatives of the rigid body realignment estimates that are used to realign blood level-dependent (BOLD) data during oxygen fMRI preprocessing [82,83]. There were no main effect of the genotype and interaction effect between genotype and gender on the FD (Table S1). Then all data were spatially normalized to the standard echo-planar imaging (EPI) template, and resampled to 3-mm³ voxels. The normalized data were smoothed with an 8-mm FWHM. Linear drifts were removed, and a temporal filter (0.01-0.08 Hz) was performed to reduce the effect of low-frequency drifts and high-frequency noise. Finally, a multiple regression method was performed to remove the possible influences of confounding factors, including six estimated motion parameters and average BOLD signals in the whole brain and CSF and white matter regions. The whole brain, CSF and white matter masks were defined using the tissue probabilistic maps in MNI space [81], i.e., the whole brain mask was thresholded at 50%, the white matter mask was thresholded at 90%, and the CSF mask was thresholded at 70%.

Statistical analysis

Statistical analyses for the demographic, cognitive and psychological data were performed using Statistical Package for the Social Sciences version 18.0 (SPSS, Chicago, IL, USA) for Windows. A two-way (genotype and gender) analysis of variance (ANOVA) was used to evaluate the main effects of genotype and gender and their interactions on age, years of education, IQ, depression scores, anxiety levels and personality traits.

The voxel-based comparisons of GMV were performed using a factorial (genotype by gender) ANOVA (P < 0.001, cluster size > 200 voxels). Brain regions with significant genotypic effect and genotype × gender interaction in GMV were extracted and defined as seed regions for the rsFC analysis. For each subject, the correlation coefficient between the mean time series of each seed region and that of each voxel in the whole brain was computed and transformed into a z-value to improve normality. Subsequently, individuals' z-values were entered into a random effects one-sample t-test to identify brain regions exhibiting significant positive correlations with the seed region. Significant rsFC maps were corrected for multiple comparisons using the Family Wise Error (FWE, P < 0.05) method. A mask of brain areas with significant positive rsFCs with the seed region was generated and applied in a two-way (genotype and gender) ANOVA. Correction for multiple comparisons was performed using the Monte Carlo simulation with a corrected threshold of P < 0.05 and a cluster size of at least 11 voxels (AlphaSim program. Parameters: single voxel P = 0.001, FWHM = 8 mm, cluster connection radius r = 5 mm; with a GM mask and a resolution of 3-mm³).

To remove potential influence by age, years of education, IQ, depression scores, anxiety levels and personality traits on the GMV and rsFC analyses, we extracted brain regions with significant differences in GMV and rsFCs; and repeated the statistical analyses using these measures as covariates of no interest. To explore whether rsFC differences are driven by the

corresponding structural differences, we repeated the rsFC analyses using the GMVs of the regions of interest (ROIs) as covariates of no interest to remove potential variations in rsFCs related to GMV [53].

The main effects of genotype and gender, and the genotype × gender interaction were reported. If the interaction was significant, *post-hoc* comparisons were performed to determine genotypic differences in female and male subjects, respectively.

Results

Demographic and genetic characteristics

The VBM analysis included 299 subjects, which consisted of 139 Val/Val, 130 Val/Met, and 30 Met/Met individuals (Table 2). The rsFC analysis included 288 subjects, which consisted of 137 Val/Val, 124 Val/Met, and 27 Met/Met individuals (Table 2). Because of the relative low frequency of Met homozygotes (4-5 times lower than Val homozygotes), we merged the Met homozygotes and Met heterozygotes into a group of Met carriers. Detailed demographic, cognitive and psychological data are summarized in Table S2, Table S3 and Table S4. There was no significant main effect of genotype or genotype × gender interaction on any of the demographic, cognitive and psychological variables. However, significant (P < 0.05) main effect of gender was found in age, years of education, depression and anxiety scores, and harm avoidance of personality.

GMV differences

The main effects of the GMV differences between genotypes are shown in Figure 1. ANOVA revealed a significant main effect of genotype on GMV in the right posterior cingulate cortex (PCC) (BA 31; peak MNI coordinate: x = 15, y = -46.5, z = 37.5; 233 voxels; peak F = 14.99). Post-hoc testing showed that Val homozygotes exhibited significantly smaller GMV in the right PCC than Met allele carriers. The interaction between genotype and gender in GMV is shown in Figure 2. ANOVA demonstrated a significant interaction between genotype and gender in the GMV of the left medial superior frontal gyrus (mSFG) (BA 10; peak MNI coordinate: x = -16.5, y = 60, z =16.5; 231 voxels; peak F = 17.67). The post-hoc comparison showed that only male Val homozygotes exhibited significantly smaller GMV than male Met allele carriers. The main effects of the GMV differences between genders are shown in Figure S1. The gender differences in GMV were found in many brain regions, including the right PCC region that showed a significant main effect of genotype. Post-hoc testing showed that male subjects exhibited significantly smaller GMV in the right PCC than female subjects.

rsFC differences

For seed regions in the rsFC analysis, we extracted the right PCC because of the significant difference observed in GMV between genotypes, and the left mSFG was extracted because of the significant interaction identified between genotype and gender in GMV. A one-sample t-test (FWE, P < 0.05) revealed

Table 2. Demographic data for VBM analysis (n = 299) and rsFC analysis (n = 288).

			VBM analysis			rsFC analysis		
			n	Age (years)	Years of education	n	Age (years)	Years of education
COMT	Met carrie	r	160	22.7 (2.6)	15.4 (2.3)	151	22.7 (2.5)	15.5 (2.2)
	Val/Val		139	22.9 (2.4)	16.0 (2.0)	137	22.9 (2.4)	15.9 (2.0)
	F(P)		299	0.20 (0.65)	3.67 (0.06)	288	0.22 (0.64)	2.46 (0.12)
Gender	Male		141	22.3 (2.6)	15.1 (2.3)	134	22.2 (2.6)	15.1 (2.2)
	Female		158	23.3 (2.3)	16.1 (2.0)	154	23.3 (2.2)	16.2 (1.9)
	F(P)		299	12.00 (< 0.01)	17.18 (< 0.01)	288	14.59 (< 0.001)	18.22 (< 0.001)
COMT × gender	Male	Met carrier	79	22.2 (2.7)	14.9 (2.3)	74	22.1 (2.5)	15.0 (2.2)
		Val/Val	62	22.3 (2.6)	15.4 (2.2)	60	22.3 (2.6)	15.3 (2.2)
	Female	Met carrier	81	23.2 (2.4)	16.0 (2.1)	77	23.3 (2.3)	16.0 (2.1)
		Val/Val	77	23.3 (2.2)	16.4 (1.8)	77	23.3 (2.2)	16.4 (1.8)
		F(P)	299	0.11 (0.92)	0.01 (0.96)	288	0.11 (0.92)	0.01 (0.96)

The data are shown as the means (SD).

doi: 10.1371/journal.pone.0078697.t002



Figure 1. Significant GMV differences between genotypes. Val homozygotes exhibit significantly (P < 0.05, corrected) smaller GMV in the right PCC than Met allele carriers. GMV, gray matter volume; L, left; PCC, posterior cingulate cortex; R, right. doi: 10.1371/journal.pone.0078697.g001

that these seed regions showed similar rsFC patterns and that they were positively correlated with brain regions of the DMN, including the PCC, precuneus, medial prefrontal cortex, anterior cingulate cortex, lateral parietal cortex, anterior temporal lobe, and SFG, although these regions differed in rsFC strengths (Figure 3). This finding suggests that both the right PCC and the left mSFG are components of the DMN. Voxel-based comparisons of rsFCs were performed using a factorial ANOVA (genotype by gender). When the right PCC was treated as the seed region, there was a significant main effect of genotype on the rsFC between the right PCC and the left medial frontal pole (FP) (BA10; peak MNI coordinate: x = -6, y = 66, z = -6; 24 voxels; peak F = 14.20; P < 0.05, corrected) (Figure 4). *Post-hoc* testing showed that Val homozygotes exhibited decreased rsFC compared with Met allele carriers. The main effects of the rsFC differences of the right PCC between genders are shown in Figure S2. The gender differences in rsFCs of the right PCC were found in several brain regions, including the left FP that showed a significant main effect of genotype. *Post-hoc* testing showed that male subjects exhibited significantly weaker rsFC between the right PCC and the left FP than female subjects. Similarly,



Figure 2. The interaction between genotype and gender in GMV. A significant (P < 0.05, corrected) genotype × gender interaction effect is observed in the left mSFG. Only male Val homozygotes exhibit significantly smaller GMV than male Met allele carriers. GMV, gray matter volume; L, left; mSFG, medial superior frontal gyrus; R, right. doi: 10.1371/journal.pone.0078697.g002

when the left mSFG was treated as the seed region, there was also a significant main effect of genotype on the rsFC between the seed region and the left FP (BA10; peak MNI coordinate: x = -27, y = 57, z = 18; 23 voxels; peak F = 16.39; P < 0.05, corrected) (Figure 5). Post-hoc testing showed that Val homozygotes also exhibited decreased rsFC compared with Met allele carriers. However, no significant main effect of gender on the rsFC between the left mSFG and the left FP were found (Figure S3). No significant interactions were observed between genotype and gender in any of the rsFCs studied. After controlling for age, years of education, IQ, depression scores, anxiety levels and personality traits, we found the results of GMV and rsFC analyses were very similar with those without controlling for these factors (Table S5). To test the structural-functional relationship, we repeated the rsFC analysis while controlling for the GMV of the seed region; however, we did not find any significant changes between results with and without correcting for the GMV (Table S6).

According the genotype and gender, subjects were divided into four subgroups with different dopamine availability: Val/Val males, Val/Val females, male Met carriers, and female Met carriers. Met-allele carriers have lower COMT activity and higher dopamine availability than Val/Val individuals; and females have higher level of estrogen, lower COMT activity, and higher dopamine availability compared with males. Thus the dopamine signaling is theoretically the lowest in Val/Val males and the highest in female Met carriers. The left column of Figure 6 shows the mean GMV or rsFC of individual subgroup; whereas the right column of Figure 6 fits the mean GMV or rsFC of individual subgroup into the inverted U-shaped model of dopaminergic modulation [2,14,36,84].

Discussion

Considering the interactions between genotype and gender, we completed a stepwise investigation of COMT Val158Met modulation on GMV and rsFC using a voxel-based analysis of the whole brain in healthy young adults. The VBM analysis revealed decreased GMV in the right PCC of Val/Val individuals compared with that of Met carriers; however, decreased GMV of the left mSFG was observed only in male Val/Val subjects. When using these two brain areas as seed regions, the rsFC analysis revealed that both the right PCC and left mSFG were part of the DMN. Moreover, both seed regions showed decreased rsFCs with different parts of the FP of the DMN in Val/Val individuals than in Met carriers. No significant genotype × gender interaction was found in any of the rsFC studies.

To date, GMV differences between COMT genotypes have been explored in five studies of healthy subjects. Although a recent study failed to find any GMV differences between genotypes [51], most studies have reported GMV differences across genotypes in the temporal cortex [44,48], hippocampus [42,44,48,52] and frontal cortex [42]. Although Honea et al. [44] had found significant GMV differences in the left hippocampal region after multiple comparisons corrected at the whole-brain level, the other three studies found genotype differences in GMV using only a small volume correction [42,52] or a ROI analysis [48]. Even with these liberal statistical methods, several studies have reported no effects of COMT genotype on frontal [85,86] or hippocampal volume [87]. This discrepancy may be caused by differences in demographic characteristics, sample size, environmental background, and statistical



Figure 3. The rsFC maps of the right PCC and left mSFG. One-sample t-test (FWE, P < 0.05) reveals that the right PCC is positively correlated with brain regions of the DMN (A). The left mSFG shows a similar rsFC pattern (B), although the right PCC and left mSFG differed in rsFC strengths. DMN, default mode network; FWE, Family Wise Error; L, left; mSFG, medial superior frontal gyrus; PCC, posterior cingulate cortex; R, right; rsFC, resting-state functional connectivity. doi: 10.1371/journal.pone.0078697.g003



Figure 4. Genotypic differences in the rsFCs of the right PCC. There is only a significant (P < 0.05, corrected) main effect of genotype in the rsFC between the right PCC and the left medial FP. Val homozygotes exhibit decreased rsFC when compared with Met allele carriers. FP, frontal pole; L, left; PCC, posterior cingulate cortex; R, right; rsFC, resting-state functional connectivity. doi: 10.1371/journal.pone.0078697.g004

methods. In the present study, we recruited a large sample of 299 healthy young Chinese Han subjects, performed GMV analysis at the whole-brain level, and considered both the effect of the genotype, gender and their interactions.

We found that Val/Val males showed smaller GMV in the left mSFG than the other three groups, suggesting that COMT Val158Met effects on prefrontal morphology is genderdependent. As the key enzyme of DA degradation, COMT may



Figure 5. Genotypic differences in the rsFCs of the left mSFG. There is only a significant (P < 0.05, corrected) main effect of genotype in the rsFC between the left mSFG and the left FP. Val homozygotes exhibit decreased rsFC when compared with Met allele carriers. FP, frontal pole; L, left; mSFG, medial superior frontal gyrus; R, right; rsFC, resting-state functional connectivity. doi: 10.1371/journal.pone.0078697.g005

account for more than 60% of the DA degradation in the PFC; it thus plays a unique role in regulating DA levels of the PFC [88] because prefrontal DA transporters are scarce [1]. The Met variant results in a fourfold decrease in enzymatic activity at body temperature [1], resulting that Val/Val individuals have greater COMT activity and lower DA availability in the PFC compared with Met carriers. Additionally, estrogen may inhibit COMT activity; this effect is more prominent in the PFC [89]. Males who have lower level of estrogen may have greater COMT activity and lower DA availability relative to females. Consequently, both the Val/Val and male statuses may result in lowest DA availability in the PFC, which may contribute to the smallest GMV in the left mSFG in the Val/Val male group.

Using VBM analysis of the whole brain, we found significantly reduced GMV in the PCC of Val/Val individuals when compared with Met carriers. Although no studies have reported structural differences between COMT genotypes, functional differences between genotypes in the PCC have been frequently demonstrated [39,78,90-92]. GMV differences in the PCC between genotypes may also be explained by the different levels of COMT activity and DA availability between the two genotypic groups. Of course, this speculation must be validated in future studies because of the relatively low COMT expression in the PCC compared with that in the PFC. In the present study, we also found a main effect of gender on the GMV of the PCC. Although the underlying mechanism is unclear, the effect of estrogen on the COMT activity and DA availability may be one of the possible candidates.

In the rsFC analyses, we found significant main effects of genotypes on the rsFCs between the right PCC and the left medial FP and between the left mSFG and the left FP. These findings may be also explained by the different levels of COMT activity and DA availability between the two genotypic groups. However, the significant main effect of gender on the rsFC was only present in the rsFC between the right PCC and the left medial FP, suggesting gender plays a different role in these two rsFCs. Although the effect of estrogen on the COMT activity may partly explain for the gender difference in the rsFC between the right PCC and the left medial FP, this mechanism cannot explain for the lack of gender difference in the rsFC between the left mSFG and the left FP. Thus, other mechanisms may be implicated in gender differences in rsFCs and need to be further studied.

One intriguing finding in this study was that modulatory effects of COMT Val158Met were found in both structural and functional characteristics of the DMN. The DMN is an intrinsic brain system that exhibits higher metabolic activity during non-task states [93], plays an important role in human cognitive functions, and is impaired in several neuropsychiatric diseases [94]. Functional connectivity within the DMN has been recognized as being positively correlated with cognitive performance in task states as well as at rest [95]. Similar to our findings, a previous study revealed that Val homozygotes exhibited decreased PFC and PCC related rsFCs within the DMN [39]. Taken together, these findings highlight a possible neural pathway by which the COMT Val158Met polymorphism may affect cognitive functions via modulation of DMN structure



Cortical dopamine signaling

Figure 6. Illustration of the nonlinear curve model explaining the GMV and rsFC differences among different subgroups. Both COMT genotype and gender affect DA availability in the brain. Different DA levels may explain the GMV and rsFC differences that were observed among different subgroups. The left column shows the mean and 95% CI of imaging phenotypes of the four subgroups, whereas the right column shows the location of each imaging phenotype on the curve. Arrows represent significant group differences (P < 0.05, corrected). CI, confidence interval; COMT, catechol-O-methyltransferase; DA, dopamine; DMN, default mode network; FP, frontal pole; GMV, gray matter volume; mSFG, medial superior frontal gyrus; PCC, posterior cingulate cortex; rsFC, resting-state functional connectivity.

and function. For example, when compared with Met carriers, Val homozygotes exhibited smaller GMV and weaker rsFCs within the DMN; this results in reduced cognitive performance [16,21,22,96]. However, we did not find a direct relationship between the structural and functional changes in the DMN, which may suggest that the modulatory effects of COMT Val158Met on the structural and functional characteristics of the DMN are independent with each other. Future studies should be done to verify the results.

Nonlinear effects of COMT genotype on brain structure and function have been reported in previous studies [10-12,15,44,97]. Although the exact neural mechanisms by which COMT Val158Met affects GMVs of the mSFG and PCC are unclear, potential mechanism is DA level-dependent neurotrophic and neurotoxic effects [44]. The effect of DA levels on neuronal survival and growth has been described as an inverted U-shaped curve, in which the optimal level of extracellular DA can induce the expression of BDNF and facilitate neuronal growth [98]. However, both low and high levels of extracellular DA can impair neuronal integrity and survival [99]. For example, excessive extracellular DA levels in dopamine transporter knockout mice can reduce BDNF gene expression in the frontal cortex [100], whereas reduced DA signaling in D1 receptor mutant mice can impair the normal expression of DA-mediated behavioral responses by affecting the neurochemical architecture of the striatum [101]. Pharmacological studies in both animals [102] and humans [14,103-106] have indicated that relatively poor cognitive performance in individuals with lower DA levels tended to improve after dopamimetic agent stimulation, whereas performance of individuals with DA levels near or at the top of the inverted U-shaped curve showed no improvement or deterioration with these agents. Our findings may partly support the inverted U-shaped model in that Val/Val males who have the greatest COMT activity and the lowest DA availability exhibited the smallest GMV and the weakest rsFC among the four groups. However, no indication of the aversive effect of high DA availability on GMV or rsFC (the downside downslope of the inverted-U) (Figure 6) may weaken the inverted Ushaped hypothesis.

It should be noted that different preprocessing methods had been used in previous rsFC studies. In the present study, we first used a temporal filter, and then remove the confounding factors (such as head motion) using a multiple regression according to several previous studies [82,107-109]. It has been criticized that this preprocessing method may reduce the effectiveness of removing the motion artifact using the regression method. To clarify the issue, we adopted an alternative preprocessing method, that is, we first used a multiple regression to remove motion parameters and other confounding factors, and then we used a temporal filter to reduce the effects of low-frequency drifts and high-frequency noise. We repeated the rsFC analysis on the preprocessed fMRI data with a different procedure and found that the new results (Figure S4 and Figure S5) were the same as the original results (Figure 4 and Figure 5) using the same statistical threshold (P < 0.05, corrected). These results suggest that the order of the preprocessing steps did not

significantly influence our findings. In the present study, we used two methods for multiple comparison correction. The FWE is a stronger correction method than the Monte Carlo simulation. The selection of different correction methods depends on the effective sizes. The one-sample t-test is to identify brain regions exhibiting significant positive correlations with the seed region. The effective size is relative large and then a stricter FWE correction was used in this situation. In contrast, the effective size of the genotypic differences between healthy adults is expected to be small. Therefore, we used a relatively loose threshold to avoid missing the subtle differences between groups. However, the intergroup differences cannot survive after the FWE correction for multiple comparisons. The lack of significant intergroup differences after a stricter FWE correction suggests that these findings should be validated in future studies.

In summary, this study used a relatively large sample size of healthy young adults and a whole brain analyzing method. We found that the COMT Val158Met polymorphism modulates anatomical morphology and related rsFCs within the DMN, indicating a potential neural pathway by which this polymorphism may affect cognitive function. Meanwhile, we found a genotype × gender interaction in the prefrontal GMV but not in the GMV of the PCC and the rsFCs within the DMN. The mechanisms of these findings need to be further investigated.

Supporting Information

Figure S1. Brain regions with significant GMV differences affected by genders (P < 0.05, corrected). The GMV of mSFG and PCC are effected by gender. GMV, gray matter volume; mSFG, medial superior frontal gyrus; L, left; PCC, posterior cingulate cortex; R, right. (DOC)

Figure S2. Brain regions with significant gender differences in rsFCs of the right PCC (P < 0.05, corrected). There is a significant main effect of gender on several brain regions, including the left FP that showed a significant main effect of genotype. FP, frontal pole; L, left; PCC, posterior cingulate cortex; R, right; rsFC, resting-state functional connectivity.

(DOC)

Figure S3. Brain regions with gender differences in the rsFCs of the left mSFG (P < 0.05, corrected). L, left; mSFG, medial superior frontal gyrus; R, right; rsFC, resting-state functional connectivity. (DOC)

Figure S4. Genotypic differences in the rsFCs of the right PCC. There is only a significant (P < 0.05, corrected) main effect of genotype in the rsFC between the right PCC and the left medial FP. Val homozygotes exhibit decreased rsFC when compared with Met allele carriers. FP, frontal pole; L, left; PCC, posterior cingulate cortex; R, right; rsFC, resting-state functional connectivity.

(DOC)

Figure S5. Genotypic differences in the rsFCs of the left mSFG. There is only a significant (P < 0.05, corrected) main effect of genotype in the rsFC between the left mSFG and the left FP. Val homozygotes exhibit decreased rsFC when compared with Met allele carriers. FP, frontal pole; L, left; mSFG, medial superior frontal gyrus; R, right; rsFC, resting-state functional connectivity. (DOC)

Table S1. Assessment of head motion in fMRI data using framewise displacement (FD) measure. (DOC)

Table S2. Demographic data and IQ scores of subjects (n = 297).

(DOC)

Table S3. Demographic data and depression scores and anxiety values of subjects (n = 279). (DOC)

References

- Männistö PT, Kaakkola S (1999) Catechol-O-methyltransferase (COMT): biochemistry, molecular biology, pharmacology, and clinical efficacy of the new selective COMT inhibitors. Pharmacol Rev 51: 593-628. PubMed: 10581325.
- Seamans JK, Yang CR (2004) The principal features and mechanisms of dopamine modulation in the prefrontal cortex. Prog Neurobiol 74: 1-58. doi:10.1016/j.pneurobio.2004.05.006. PubMed: 15381316.
- Barnett JH, Scoriels L, Munafò MR (2008) Meta-analysis of the cognitive effects of the catechol-O-methyltransferase gene Val158/108Met polymorphism. Biol Psychiatry 64: 137-144. doi: 10.1016/j.biopsych.2008.01.005. PubMed: 18339359.
- Gennatas ED, Cholfin JA, Zhou J, Crawford RK, Sasaki DA et al. (2012) COMT Val158Met genotype influences neurodegeneration within dopamine-innervated brain structures. Neurology 78: 1663-1669. doi:10.1212/WNL.0b013e3182574fa1. PubMed: 22573634.
- Goldberg TE, Weinberger DR (2004) Genes and the parsing of cognitive processes. Trends Cogn Sci 8: 325-335. doi:10.1016/j.tics. 2004.05.011. PubMed: 15242692.
- Mier D, Kirsch P, Meyer-Lindenberg A (2010) Neural substrates of pleiotropic action of genetic variation in COMT: a meta-analysis. Mol Psychiatry 15: 918-927. doi:10.1038/mp.2009.36. PubMed: 19417742.
- Sambataro F, Pennuto M, Wolf RC (2012) Catechol-O-Methyl Transferase Modulates Cognition in Late Life: Evidence and Implications for Cognitive Enhancement. CNS Neurol Disord Drug Targets 11: 195-208. doi:10.2174/187152712800672463. PubMed: 22483295.
- Wacker J, Mueller EM, Hennig J, Stemmler G (2012) How to consistently link extraversion and intelligence to the catechol-Omethyltransferase (COMT) gene: on defining and measuring psychological phenotypes in neurogenetic research. J Pers Soc Psychol 102: 427-444. doi:10.1037/a0026544. PubMed: 22180998.
- Favaro A, Clementi M, Manara R, Bosello R, Forzan M et al. (2012) Catechol-O-methyltransferase genotype modifies executive functioning and prefrontal functional connectivity in women with anorexia nervosa. J Psychiatry Neurosci 37: 120068. PubMed: 23046831.
- Goldman-Rakic PS (1998) The cortical dopamine system: role in memory and cognition. Adv Pharmacol 42: 707-711. PubMed: 9327997.
- Williams GV, Goldman-Rakic PS (1995) Modulation of memory fields by dopamine D1 receptors in prefrontal cortex. Nature 376: 572-575. doi:10.1038/376572a0. PubMed: 7637804.
- 12. Fallon SJ, Williams-Gray CH, Barker RA, Owen AM, Hampshire A (2012) Prefrontal Dopamine Levels Determine the Balance between

Table S4. Demographic data and personality traits of subjects (n = 292). (DOC)

Table S5. Statistical results before (out of the brackets) and after (in the brackets) removing demographic data and behavioral scales. (DOC)

Table S6. Statistical results before (out of the brackets) and after (in the brackets) removing the GMV of seed region.

(DOC)

Author Contributions

Conceived and designed the experiments: TT WQ CY. Performed the experiments: TT WQ DW JW BL. Analyzed the data: TT WQ DW JW BL. Contributed reagents/materials/ analysis tools: TT WQ DW JW BL. Wrote the manuscript: TT WQ CY. Preparation of Manuscript for publication: TT WQ CY TJ.

Cognitive Stability and Flexibility. Cereb Cortex 23: 361-369. PubMed: 22351648.

- Giakoumaki SG, Roussos P, Bitsios P (2008) Improvement of prepulse inhibition and executive function by the COMT inhibitor tolcapone depends on COMT Val158Met polymorphism. Neuropsychopharmacology 33: 3058-3068. doi:10.1038/npp.2008.82. PubMed: 18536698.
- Mattay VS, Goldberg TE, Fera F, Hariri AR, Tessitore A et al. (2003) Catechol O-methyltransferase val158-met genotype and individual variation in the brain response to amphetamine. Proc Natl Acad Sci U S A 100: 6186-6191. doi:10.1073/pnas.0931309100. PubMed: 12716966.
- 15. Qin S, Cousijn H, Rijpkema M, Luo J, Franke B et al. (2012) The effect of moderate acute psychological stress on working memory-related neural activity is modulated by a genetic variation in catecholaminergic function in humans. Front. J Integr Neurosci 6: 16.
- Barnett JH, Jones PB, Robbins TW, Müller U (2007) Effects of the catechol-O-methyltransferase Val158Met polymorphism on executive function: a meta-analysis of the Wisconsin Card Sort Test in schizophrenia and healthy controls. Mol Psychiatry 12: 502-509. PubMed: 17325717.
- Bertolino A, Rubino V, Sambataro F, Blasi G, Latorre V et al. (2006) Prefrontal-hippocampal coupling during memory processing is modulated by COMT val158met genotype. Biol Psychiatry 60: 1250-1258. doi:10.1016/j.biopsych.2006.03.078. PubMed: 16950222.
- Bruder GE, Keilp JG, Xu H, Shikhman M, Schori E et al. (2005) Catechol-O-methyltransferase (COMT) genotypes and working memory: associations with differing cognitive operations. Biol Psychiatry 58: 901-907. doi:10.1016/j.biopsych.2005.05.010. PubMed: 16043133.
- de Frias CM, Annerbrink K, Westberg L, Eriksson E, Adolfsson R et al. (2004) COMT gene polymorphism is associated with declarative memory in adulthood and old age. Behav Genet 34: 533-539. doi: 10.1023/B:BEGE.0000038491.06972.8c. PubMed: 15319576.
- de Frias CM, Annerbrink K, Westberg L, Eriksson E, Adolfsson R et al. (2005) Catechol O-methyltransferase Val158Met polymorphism is associated with cognitive performance in nondemented adults. J Cogn Neurosci 17: 1018-1025. doi:10.1162/0898929054475136. PubMed: 16102234.
- Egan MF, Goldberg TE, Kolachana BS, Callicott JH, Mazzanti CM et al. (2001) Effect of COMT Val108/158 Met genotype on frontal lobe function and risk for schizophrenia. Proc Natl Acad Sci U S A 98: 6917-6922. doi:10.1073/pnas.111134598. PubMed: 11381111.
- Goldberg TE, Egan MF, Gscheidle T, Coppola R, Weickert T et al. (2003) Executive subprocesses in working memory: relationship to

catechol-O-methyltransferase Val158Met genotype and schizophrenia. Arch Gen Psychiatry 60: 889-896. doi:10.1001/archpsyc.60.9.889. PubMed: 12963670.

- Holtzer R, Ozelius L, Xue X, Wang T, Lipton RB et al. (2010) Differential effects of COMT on gait and executive control in aging. Neurobiol Aging 31: 523-531. doi:10.1016/j.neurobiolaging. 2008.05.011. PubMed: 18547681.
- Joober R, Gauthier J, Lal S, Bloom D, Lalonde P et al. (2002) Catechol-O-methyltransferase Val-108/158-Met gene variants associated with performance on the Wisconsin Card Sorting Test. Arch Gen Psychiatry 59: 662-663. doi:10.1001/archpsyc.59.7.662. PubMed: 12090821.
- Malhotra AK, Kestler LJ, Mazzanti C, Bates JA, Goldberg T et al. (2002) A functional polymorphism in the COMT gene and performance on a test of prefrontal cognition. Am J Psychiatry 159: 652-654. doi: 10.1176/appi.ajp.159.4.652. PubMed: 11925305.
- Raz N, Rodrigue KM, Kennedy KM, Land S (2009) Genetic and vascular modifiers of age-sensitive cognitive skills: effects of COMT, BDNF, ApoE, and hypertension. Neuropsychology 23: 105-116. doi: 10.1037/a0013487. PubMed: 19210038.
- Bishop SJ, Fossella J, Croucher CJ, Duncan J (2008) COMT val158met genotype affects recruitment of neural mechanisms supporting fluid intelligence. Cereb Cortex 18: 2132-2140. doi:10.1093/ cercor/bhm240. PubMed: 18252743.
- Tan HY, Chen Q, Goldberg TE, Mattay VS, Meyer-Lindenberg A et al. (2007) Catechol-O-methyltransferase Val158Met modulation of prefrontal-parietal-striatal brain systems during arithmetic and temporal transformations in working memory. J Neurosci 27: 13393-13401. doi: 10.1523/JNEUROSCI.4041-07.2007. PubMed: 18057197.
- Montag C, Buckholtz JW, Hartmann P, Merz M, Burk C et al. (2008) COMT genetic variation affects fear processing: psychophysiological evidence. Behav Neurosci 122: 901-909. doi: 10.1037/0735-7044.122.4.901. PubMed: 18729643.
- Swart M, Bruggeman R, Larøi F, Alizadeh BZ, Kema I et al. (2011) COMT Val158Met polymorphism, verbalizing of emotion and activation of affective brain systems. NeuroImage 55: 338-344. doi:10.1016/ j.neuroimage.2010.12.017. PubMed: 21156209.
- Bishop SJ, Cohen JD, Fossella J, Casey BJ, Farah MJ (2006) COMT genotype influences prefrontal response to emotional distraction. Cogn Affect Behav Neurosci 6: 62-70. doi:10.3758/CABN.6.1.62. PubMed: 16869230.
- Drabant EM, Hariri AR, Meyer-Lindenberg A, Munoz KE, Mattay VS et al. (2006) Catechol O-methyltransferase val158met genotype and neural mechanisms related to affective arousal and regulation. Arch Gen Psychiatry 63: 1396-1406. doi:10.1001/archpsyc.63.12.1396. PubMed: 17146014.
- Smolka MN, Schumann G, Wrase J, Grüsser SM, Flor H et al. (2005) Catechol-O-methyltransferase val158met genotype affects processing of emotional stimuli in the amygdala and prefrontal cortex. J Neurosci 25: 836-842. doi:10.1523/JNEUROSCI.1792-04.2005. PubMed: 15673663.
- Rasch B, Spalek K, Buholzer S, Luechinger R, Boesiger P et al. (2010) Aversive stimuli lead to differential amygdala activation and connectivity patterns depending on catechol-O-methyltransferase Val158Met genotype. NeuroImage 52: 1712-1719. doi:10.1016/j.neuroimage. 2010.05.054. PubMed: 20510373.
- Bilder RM, Volavka J, Lachman HM, Grace AA (2004) The catechol-Omethyltransferase polymorphism: relations to the tonic-phasic dopamine hypothesis and neuropsychiatric phenotypes. Neuropsychopharmacology 29: 1943-1961. doi:10.1038/sj.npp. 1300542. PubMed: 15305167.
- Witte AV, Floel A (2011) Effects of COMT polymorphisms on brain function and behavior in health and disease. Brain. Res Bull 88: 418-428.
- Gianotti LR, Figner B, Ebstein RP, Knoch D (2012) Why Some People Discount More than Others: Baseline Activation in the Dorsal PFC Mediates the Link between COMT Genotype and Impatient Choice. Front Neurosci 6: 54. PubMed: 22586360.
- Lee TW, Yu YW, Hong CJ, Tsai SJ, Wu HC et al. (2011) The effects of catechol-O-methyl-transferase polymorphism Val158Met on functional connectivity in healthy young females: a resting EEG study. Brain Res 1377: 21-31. doi:10.1016/j.brainres.2010.12.073. PubMed: 21195697.
- Liu B, Song M, Li J, Liu Y, Li K et al. (2010) Prefrontal-related functional connectivities within the default network are modulated by COMT val158met in healthy young adults. J Neurosci 30: 64-69. doi:10.1523/ JNEUROSCI.3941-09.2010. PubMed: 20053888.
- Tunbridge EM, Farrell SM, Harrison PJ, Mackay CE (2013) Catechol-O-methyltransferase (COMT) influences the connectivity of the prefrontal cortex at rest. NeuroImage 68: 49-54. doi:10.1016/ j.neuroimage.2012.11.059. PubMed: 23228511.

- Cerasa A, Cherubini A, Quattrone A, Gioia MC, Tarantino P et al. (2010) Met158 variant of the catechol-O-methyltransferase genotype is associated with thicker cortex in adult brain. Neuroscience 167: 809-814. doi:10.1016/j.neuroscience.2010.02.040. PubMed: 20219642.
- Cerasa A, Gioia MC, Labate A, Liguori M, Lanza P et al. (2008) Impact of catechol-O-methyltransferase Val(108/158) Met genotype on hippocampal and prefrontal gray matter volume. Neuroreport 19: 405-408. doi:10.1097/WNR.0b013e3282f5f784. PubMed: 18287936.
- 43. Ehrlich S, Morrow EM, Roffman JL, Wallace SR, Naylor M et al. (2010) The COMT Val108/158Met polymorphism and medial temporal lobe volumetry in patients with schizophrenia and healthy adults. Neuroimage 53: 992-1000. doi:10.1016/j.neuroimage.2009.12.046. PubMed: 20026221.
- Honea R, Verchinski BA, Pezawas L, Kolachana BS, Callicott JH et al. (2009) Impact of interacting functional variants in COMT on regional gray matter volume in human brain. Neuroimage 45: 44-51. doi: 10.1016/j.neuroimage.2008.10.064. PubMed: 19071221.
- 45. Li J, Yu C, Li Y, Liu B, Liu Y et al. (2009) COMT val158met modulates association between brain white matter architecture and IQ. Am J Med Genet B Neuropsychiatr Genet 150B: 375-380. doi:10.1002/ajmg.b. 30825. PubMed: 18615479.
- 46. Raznahan A, Greenstein D, Lee Y, Long R, Clasen L et al. (2011) Catechol-o-methyl transferase (COMT) val158met polymorphism and adolescent cortical development in patients with childhood-onset schizophrenia, their non-psychotic siblings, and healthy controls. NeuroImage 57: 1517-1523. doi:10.1016/j.neuroimage.2011.05.032. PubMed: 21620981.
- Shaw P, Wallace GL, Addington A, Evans A, Rapoport J et al. (2009) Effects of the Val158Met catechol-O-methyltransferase polymorphism on cortical structure in children and adolescents. Mol Psychiatry 14: 348-349. doi:10.1038/mp.2008.121. PubMed: 19308019.
- Taylor WD, Züchner S, Payne ME, Messer DF, Doty TJ et al. (2007) The COMT Val158Met polymorphism and temporal lobe morphometry in healthy adults. Psychiatry Res 155: 173-177. doi:10.1016/ j.pscychresns.2007.01.005. PubMed: 17521892.
- Thomason ME, Dougherty RF, Colich NL, Perry LM, Rykhlevskaia El et al. (2010) COMT genotype affects prefrontal white matter pathways in children and adolescents. Neuroimage 53: 926-934. doi:10.1016/ j.neuroimage.2010.01.033. PubMed: 20083203.
- Zinkstok J, Schmitz N, van Amelsvoort T, de Win M, van den Brink W et al. (2006) The COMT val158met polymorphism and brain morphometry in healthy young adults. Neurosci Lett 405: 34-39. doi: 10.1016/j.neulet.2006.06.034. PubMed: 16857316.
- Barnes A, Isohanni M, Barnett JH, Pietiläinen O, Veijola J et al. (2012) No Association of COMT (Val158Met) Genotype with Brain Structure Differences between Men and Women. PLOS ONE 7: e33964. doi: 10.1371/journal.pone.0033964. PubMed: 22479488.
- Mechelli A, Tognin S, McGuire PK, Prata D, Sartori G et al. (2009) Genetic vulnerability to affective psychopathology in childhood: a combined voxel-based morphometry and functional magnetic resonance imaging study. Biol Psychiatry 66: 231-237. doi:10.1016/ j.biopsych.2009.01.033. PubMed: 19278671.
- Voets NL, Beckmann CF, Cole DM, Hong S, Bernasconi A et al. (2012) Structural substrates for resting network disruption in temporal lobe epilepsy. Brain 135: 2350-2357. doi:10.1093/brain/aws137. PubMed: 22669081.
- Jiang H, Xie T, Ramsden DB, Ho SL (2003) Human catechol-Omethyltransferase down-regulation by estradiol. Neuropharmacology 45: 1011-1018. doi:10.1016/S0028-3908(03)00286-7. PubMed: 14573393.
- Xie T, Ho SL, Ramsden D (1999) Characterization and implications of estrogenic down-regulation of human catechol-O-methyltransferase gene transcription. Mol Pharmacol 56: 31-38. PubMed: 10385681.
- Gogos JA, Morgan M, Luine V, Santha M, Ogawa S et al. (1998) Catechol-O-methyltransferase-deficient mice exhibit sexually dimorphic changes in catecholamine levels and behavior. Proc Natl Acad Sci U S A 95: 9991-9996. doi:10.1073/pnas.95.17.9991. PubMed: 9707588.
- Barnett JH, Heron J, Ring SM, Golding J, Goldman D et al. (2007) Gender-specific effects of the catechol-O-methyltransferase Val108/158Met polymorphism on cognitive function in children. Am J Psychiatry 164: 142-149. doi:10.1176/appi.ajp.164.1.142. PubMed: 17202556.
- Chen C, Moyzis R, Dong Q, He Q, Zhu B et al. (2011) Sex modulates the associations between the COMT gene and personality traits. Neuropsychopharmacology 36: 1593-1598. doi:10.1038/npp.2011.39. PubMed: 21471954.
- Kempton MJ, Haldane M, Jogia J, Christodoulou T, Powell J et al. (2009) The effects of gender and COMT Val158Met polymorphism on fearful facial affect recognition: a fMRI study. Int J

Neuropsychopharmacol 12: 371-381. doi:10.1017/ S1461145708009395. PubMed: 18796186.

- Lang UE, Bajbouj M, Sander T, Gallinat J (2007) Gender-dependent association of the functional catechol-O-methyltransferase Val158Met genotype with sensation seeking personality trait. Neuropsychopharmacology 32: 1950-1955. doi:10.1038/sj.npp. 1301335. PubMed: 17299513.
- O'Hara R, Miller E, Liao CP, Way N, Lin X et al. (2006) COMT genotype, gender and cognition in community-dwelling, older adults. Neurosci Lett 409: 205-209. doi:10.1016/j.neulet.2006.09.047. PubMed: 17029783.
- 62. Zhang K, Zheng Z, Gao X, Li J, Zhang F (2007) Possible relationship between the COMT gene ValMet polymorphism and psychometric IQ in girls of the Qinba region in China. Neuropsychobiology 56: 98-103. doi: 10.1159/000112950. PubMed: 18182829.
- Hoth KF, Paul RH, Williams LM, Dobson-Stone C, Todd E et al. (2006) Associations between the COMT Val/Met polymorphism, early life stress, and personality among healthy adults. Neuropsychiatr Dis Treat 2: 219-225. doi:10.2147/nedt.2006.2.2.219. PubMed: 19412467.
- Rowe JB, Hughes L, Williams-Gray CH, Bishop S, Fallon S et al. (2010) The val158met COMT polymorphism's effect on atrophy in healthy aging and Parkinson's disease. Neurobiol Aging 31: 1064-1068. doi:10.1016/j.neurobiolaging.2008.07.009. PubMed: 18755526.
- Dumontheil I, Roggeman C, Ziermans T, Peyrard-Janvid M, Matsson H et al. (2011) Influence of the COMT genotype on working memory and brain activity changes during development. Biol Psychiatry 70: 222-229. doi:10.1016/j.biopsych.2011.02.027. PubMed: 21514925.
- Sambataro F, Reed JD, Murty VP, Das S, Tan HY et al. (2009) Catechol-O-methyltransferase valine(158)methionine polymorphism modulates brain networks underlying working memory across adulthood. Biol Psychiatry 66: 540-548. doi:10.1016/j.biopsych. 2009.04.014. PubMed: 19539269.
- Oldfield RC (1971) The assessment and analysis of handedness: the Edinburgh inventory. Neuropsychologia 9: 97-113. doi: 10.1016/0028-3932(71)90067-4. PubMed: 5146491.
- Gong YX (1982) Manual of modified Wechsler Adult Intelligence Scale (WAIS-RC) (in Chinese). Changsha, (China): Human Med College.
- Beck AT, Steer RA (1993) Manual for the Beck Depression Inventory. San Antonio: Psychological Corporation.
- Zung WWK (1971) A rating instrument for anxiety disorders. Psychosomatics 12: 371-379. doi:10.1016/S0033-3182(71)71479-0. PubMed: 5172928.
- Cloninger CR, Svrakic DM, Przybeck TR (1993) A psychobiological model of temperament and character. Arch Gen Psychiatry 50: 975-990. doi:10.1001/archpsyc.1993.01820240059008. PubMed: 8250684.
- Enoch MA, Xu K, Ferro E, Harris CR, Goldman D (2003) Genetic origins of anxiety in women: a role for a functional catechol-Omethyltransferase polymorphism. Psychiatr Genet 13: 33-41. doi: 10.1097/00041444-200303000-00006. PubMed: 12605099.
- Green AE, Kraemer DJ, Deyoung CG, Fossella JA, Gray JR (2012) A Gene-Brain-Cognition Pathway: Prefrontal Activity Mediates the Effect of COMT on Cognitive Control and IQ. Cereb Cortex 23: 552-559. PubMed: 22368081.
- Reuter M, Hennig J (2005) Association of the functional catechol-Omethyltransferase VAL158MET polymorphism with the personality trait of extraversion. Neuroreport 16: 1135-1138. doi: 10.1097/00001756-200507130-00020. PubMed: 15973162.
- 75. Thomas G, Sinville R, Sutton S, Farquar H, Hammer RP et al. (2004) Capillary and microelectrophoretic separations of ligase detection reaction products produced from low-abundant point mutations in genomic DNA. Electrophoresis 25: 1668-1677. doi:10.1002/elps. 200405886. PubMed: 15188256.
- 76. Yi P, Chen Z, Zhao Y, Guo J, Fu H et al. (2009) PCR/LDR/capillary electrophoresis for detection of single-nucleotide differences between fetal and maternal DNA in maternal plasma. Prenat Diagn 29: 217-222. doi:10.1002/pd.2072. PubMed: 19177453.
- 77. Aguilera M, Barrantes-Vidal N, Arias B, Moya J, Villa H et al. (2008) Putative role of the COMT gene polymorphism (Val158Met) on verbal working memory functioning in a healthy population. Am J Med Genet B Neuropsychiatr Genet 147B: 898-902. doi:10.1002/ajmg.b.30705. PubMed: 18213617.
- Ettinger U, Kumari V, Collier DA, Powell J, Luzi S et al. (2008) Catechol-O-methyltransferase (COMT) val158met genotype is associated with BOLD response as a function of task characteristic. Neuropsychopharmacology 33: 3046-3057. doi:10.1038/sj.npp. 1301658. PubMed: 18235427.

- Ashburner J, Friston KJ (2005) Unified segmentation. NeuroImage 26: 839-851. doi:10.1016/j.neuroimage.2005.02.018. PubMed: 15955494.
- Ashburner J (2007) A fast diffeomorphic image registration algorithm. NeuroImage 38: 95-113. doi:10.1016/j.neuroimage.2007.07.007. PubMed: 17761438.
- Chao-Gan Y, Yu-Feng Z (2010) DPARSF: A MATLAB Toolbox for "Pipeline" Data Analysis of Resting-State fMRI. Front Syst Neurosci 4: 13. PubMed: 20577591.
- Power JD, Barnes KA, Snyder AZ, Schlaggar BL, Petersen SE (2012) Steps toward optimizing motion artifact removal in functional connectivity MRI; a reply to Carp. NeuroImage 76: 439-441. PubMed: 22440651.
- Power JD, Barnes KA, Snyder AZ, Schlaggar BL, Petersen SE (2012) Spurious but systematic correlations in functional connectivity MRI networks arise from subject motion. NeuroImage 59: 2142-2154. doi: 10.1016/j.neuroimage.2011.10.018. PubMed: 22019881.
- Dickinson D, Elvevag B (2009) Genes, cognition and brain through a COMT lens. Neuroscience 164: 72-87. doi:10.1016/j.neuroscience. 2009.05.014. PubMed: 19446012.
- Ho BC, Wassink TH, O'Leary DS, Sheffield VC, Andreasen NC (2005) Catechol-O-methyl transferase Val158Met gene polymorphism in schizophrenia: working memory, frontal lobe MRI morphology and frontal cerebral blood flow. Mol Psychiatry 10: 229: 287-298. PubMed: 15668720.
- Ohnishi T, Hashimoto R, Mori T, Nemoto K, Moriguchi Y et al. (2006) The association between the Val158Met polymorphism of the catechol-O-methyl transferase gene and morphological abnormalities of the brain in chronic schizophrenia. Brain 129: 399-410. PubMed: 16330500.
- Dutt A, McDonald C, Dempster E, Prata D, Shaikh M et al. (2009) The effect of COMT, BDNF, 5-HTT, NRG1 and DTNBP1 genes on hippocampal and lateral ventricular volume in psychosis. Psychol Med 39: 1783-1797. doi:10.1017/S0033291709990316. PubMed: 19573260.
- 88. Karoum F, Chrapusta SJ, Egan MF (1994) 3-Methoxytyramine is the major metabolite of released dopamine in the rat frontal cortex: reassessment of the effects of antipsychotics on the dynamics of dopamine release and metabolism in the frontal cortex, nucleus accumbens, and striatum by a simple two pool model. J Neurochem 63: 972-979. PubMed: 7914228.
- Bixo M, Bäckström T, Winblad B, Andersson A (1995) Estradiol and testosterone in specific regions of the human female brain in different endocrine states. J Steroid Biochem Mol Biol 55: 297-303. doi: 10.1016/0960-0760(95)00179-4. PubMed: 8541226.
- McIntosh AM, Baig BJ, Hall J, Job D, Whalley HC et al. (2007) Relationship of catechol-O-methyltransferase variants to brain structure and function in a population at high risk of psychosis. Biol Psychiatry 61: 1127-1134. doi:10.1016/j.biopsych.2006.05.020. PubMed: 17014827.
- Schmahl C, Ludäscher P, Greffrath W, Kraus A, Valerius G et al. (2012) COMT val158met polymorphism and neural pain processing. PLOS ONE 7: e23658. doi:10.1371/journal.pone.0023658. PubMed: 22247753.
- Stokes PR, Rhodes RA, Grasby PM, Mehta MA (2011) The effects of the COMT Val108/158Met polymorphism on BOLD activation during working memory, planning, and response inhibition: a role for the posterior cingulate cortex? Neuropsychopharmacology 36: 763-771. doi:10.1038/npp.2010.210. PubMed: 21150912.
- Gusnard DA, Akbudak E, Shulman GL, Raichle ME (2001) Medial prefrontal cortex and self-referential mental activity: relation to a default mode of brain function. Proc Natl Acad Sci U S A 98: 4259-4264. doi: 10.1073/pnas.071043098. PubMed: 11259662.
- Broyd SJ, Demanuele C, Debener S, Helps SK, James CJ et al. (2009) Default-mode brain dysfunction in mental disorders: a systematic review. Neurosci Biobehav Rev 33: 279-296. doi:10.1016/j.neubiorev. 2008.09.002. PubMed: 18824195.
- Hampson M, Driesen NR, Skudlarski P, Gore JC, Constable RT (2006) Brain connectivity related to working memory performance. J Neurosci 26: 13338-13343. doi:10.1523/JNEUROSCI.3408-06.2006. PubMed: 17182784.
- Blasi G, Mattay VS, Bertolino A, Elvevåg B, Callicott JH et al. (2005) Effect of catechol-O-methyltransferase val158met genotype on attentional control. J Neurosci 25: 5038-5045. doi:10.1523/ JNEUROSCI.0476-05.2005. PubMed: 15901785.
- Meyer-Lindenberg A, Nichols T, Callicott JH, Ding J, Kolachana B et al. (2006) Impact of complex genetic variation in COMT on human brain function. Mol Psychiatry 11: 797-877, 10.1038/sj.mp.4001860. PubMed: 16786032.
- Küppers E, Beyer C (2001) Dopamine regulates brain-derived neurotrophic factor (BDNF) expression in cultured embryonic mouse

striatal cells. Neuroreport 12: 1175-1179. doi: 10.1097/00001756-200105080-00025. PubMed: 11338187.

- Santiago M, Matarredona ER, Granero L, Cano J, Machado A (2000) Neurotoxic relationship between dopamine and iron in the striatal dopaminergic nerve terminals. Brain Res 858: 26-32. doi:10.1016/ S0006-8993(99)02485-3. PubMed: 10700592.
- 100. Fumagalli F, Racagni G, Colombo E, Riva MA (2003) BDNF gene expression is reduced in the frontal cortex of dopamine transporter knockout mice. Mol Psychiatry 8: 898-899. doi:10.1038/sj.mp.4001370. PubMed: 14593425.
- 101. Xu M, Moratalla R, Gold LH, Hiroi N, Koob GF et al. (1994) Dopamine D1 receptor mutant mice are deficient in striatal expression of dynorphin and in dopamine-mediated behavioral responses. Cell 79: 729-742. doi:10.1016/0092-8674(94)90557-6. PubMed: 7954836.
- 102. Granon S, Passetti F, Thomas KL, Dalley JW, Everitt BJ et al. (2000) Enhanced and impaired attentional performance after infusion of D1 dopaminergic receptor agents into rat prefrontal cortex. J Neurosci 20: 1208-1215. PubMed: 10648725.
- Apud JA, Mattay V, Chen J, Kolachana BS, Callicott JH et al. (2007) Tolcapone improves cognition and cortical information processing in normal human subjects. Neuropsychopharmacology 32: 1011-1020. doi:10.1038/sj.npp.1301227. PubMed: 17063156.
- Kimberg DY, D'Esposito M, Farah MJ (1997) Effects of bromocriptine on human subjects depend on working memory capacity. Neuroreport

8: 3581-3585. doi:10.1097/00001756-199711100-00032. PubMed: 9427330.

- Mattay VS, Callicott JH, Bertolino A, Heaton I, Frank JA et al. (2000) Effects of dextroamphetamine on cognitive performance and cortical activation. NeuroImage 12: 268-275. doi:10.1006/nimg.2000.0610. PubMed: 10944409.
- 106. Mehta MA, Owen AM, Sahakian BJ, Mavaddat N, Pickard JD et al. (2000) Methylphenidate enhances working memory by modulating discrete frontal and parietal lobe regions in the human brain. J Neurosci 20: RC65–RC65. PubMed: 10704519.
- 107. Brier MR, Thomas JB, Snyder AZ, Benzinger TL, Zhang D et al. (2012) Loss of intranetwork and internetwork resting state functional connections with Alzheimer's disease progression. J Neurosci 32: 8890-8899. doi:10.1523/JNEUROSCI.5698-11.2012. PubMed: 22745490.
- Fox MD, Corbetta M, Snyder AZ, Vincent JL, Raichle ME (2006) Spontaneous neuronal activity distinguishes human dorsal and ventral attention systems. Proc Natl Acad Sci U S A 103: 10046-10051. doi: 10.1073/pnas.0604187103. PubMed: 16788060.
- 109. Pizoli CE, Shah MN, Snyder AZ, Shimony JS, Limbrick DD et al. (2011) Resting-state activity in development and maintenance of normal brain function. Proc Natl Acad Sci U S A 108: 11638-11643. doi:10.1073/ pnas.1109144108. PubMed: 21709227.