



Complete Genome Sequences of *Paenibacillus larvae* Phages BN12, Dragolir, Kiel007, Leyra, Likha, Pagassa, PBL1c, and Tadhana

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ABSTRACT We present here the complete genomes of eight phages that infect *Paenibacillus larvae*, the causative agent of American foulbrood in honeybees. Phage PBL1c was originally isolated in 1984 from a *P. larvae* lysogen, while the remaining phages were isolated in 2014 from bee debris, honeycomb, and lysogens from three states in the USA.

The Gram-positive bacterium *Paenibacillus larvae* is the causative agent of American foulbrood, currently the most destructive bacterial disease affecting the honeybee, *Apis mellifera* (1). With the rise of antibiotic-resistant strains of *P. larvae* (2), there is growing interest in phages that infect this pathogen. The first *P. larvae* phages were isolated in the 1950s (3), and the first complete *P. larvae* genome was published in 2013 (4). There are currently 18 complete *P. larvae* phage genomes in the literature (4–7). Here, we present eight complete *P. larvae* phage genomes obtained from samples across the United States. The phages' GenBank accession numbers, isolation sources, geographical provenance, and assembly results are shown in Table 1.

Phage PBL1c was isolated from a lysogen in 1984 by Dingman et al. (8) but was not sequenced until 2018 at Brigham Young University (BYU). The remaining seven phages were isolated over the period 2014 to 2016 from samples from the USA states of Utah, Idaho, and Wisconsin (Table 1) as part of the Phage Hunters course at BYU.

The phages were isolated from bee debris, honeycomb, and lysogens and amplified in *P. larvae* field isolates. Phage genomic DNA was isolated from high-titer lysates using Norgen phage DNA isolation kits (Norgen Biotek, Thorold, ON, Canada). Phage genomes were sequenced in the BYU DNA Sequencing Center using the Illumina HiSeq 2500 platform (Illumina, Hayward, CA, USA) and were assembled using Geneious 8 software (Biomatters Inc., Newark, NJ, USA).

All nine phages are members of the family *Siphoviridae* with linear double-stranded DNA genomes. The DNA packaging strategy was identified as “cohesive ends with 3' overhangs,” as explained in references 9 and 10. The overhangs were identified by sequence similarity with previously published phages (3–7). The overhangs are “CGACT-GCCC” for phages BN12, Kiel007, Leyra, Likha, Pagassa, PBL1c, and Tadhana, and “CGACGGACC” for phage Dragolir. The genomes were rearranged by setting the first base of the genome to be the base immediately after the 3' overhang.

Genome length is in the 37 kb to 42 kb range, and the G+C content was in the 41 to 44% range, consistent with 3' cohesive ends for *P. larvae* phages (11). Preliminary analysis shows that phages Pagassa and Tadhana are closely related to each other, with

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TABLE 1 *P. larvae* phages, GenBank accession numbers, and genome assembly results

Phage name	GenBank accession no.	Isolation source	Location	Genome length (bp)	GC content (%)
BN12	MG727695	Bee debris	Cedar City, Utah, USA	39,485	42.6
Dragolir	MG727697	Bee debris	Wisconsin, USA	41,131	44
Kiel007	MG727696	Bee debris	Salt Lake City, Utah, USA	37,985	41.8
Leyra	MG727701	Bee debris	Idaho, USA	42,276	41.4
Likha	MG727702	Honeycomb	American Fork, Utah, USA	39,778	41.3
Pagassa	MG727699	<i>P. larvae</i> lysogen	Provo, Utah, USA	40,035	42
PBL1c	MG727698	<i>P. larvae</i> lysogen	Iowa City, Iowa, USA	40,611	41.2
Tadhana	MG727700	<i>P. larvae</i> lysogen	Provo, Utah, USA	37,880	42.1

the other phages slightly more distant; phage Dragolir was shown to be an outlier. All eight phages encode a large terminase, a major tail protein, two tail assembly proteins, a tail tape measure protein, and an *N*-acetylmuramoyl-L-alanine amidase, among others. The tail assembly proteins appear to have a programmed translational frameshift similar to the G and G-T genes of phage lambda (12, 13), located in the 3' region of gp12 (the upstream tail assembly protein). We tentatively identified the heptanucleotide slippery sequence as "AAAAAAG" in phages BN12, Kiel007, Likha, Leyra, Pagassa, PBL1c, and Tadhana, and possibly "AAAAAAC" in phage Dragolir. Future studies will investigate this and other features of *P. larvae* phage genomes and also provide a detailed comparative genomic analysis of these and other *P. larvae* phages.

Accession number(s). The genome sequences of the *P. larvae* phages reported here have been deposited in GenBank under the accession numbers listed in Table 1.

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REFERENCES

- de Graaf DC, Alippi AM, Antúnez K, Aronstein KA, Budge G, De Koker D, De Smet L, Dingman DW, Evans JD, Foster LJ, Fünfhaus A, Garcia-Gonzalez E, Gregore A, Human H, Murray KD, Nguyen BK, Poppinga L, Spivak M, van Engelsdorp D, Wilkins S, Genersch E. 2013. Standard methods for American foulbrood research. *J Apic Res* 52:1–28. <https://doi.org/10.3896/IBRA.1.52.1.11>.
- Miyagi T, Peng CYS, Chuang RY, Mussen EC, Spivak MS, Doi RH. 2000. Verification of oxytetracycline-resistant American foulbrood pathogen *Paenibacillus larvae* in the United States. *J Invertebr Pathol* 75:95–96. <https://doi.org/10.1006/jjpa.1999.4888>.
- Gochnauer TA. 1955. The isolation of a bacteriophage (bacterial virus) from *Bacillus larvae*. *Bee World* 36:101–103. <https://doi.org/10.1080/0005772X.1955.11094880>.
- Oliveira A, Melo LDR, Kropinski AM, Azeredo J. 2013. Complete genome sequence of the broad-host-range *Paenibacillus larvae* phage phi-IBB_PI23. *Genome Announc* 1(5):e00438-13. <https://doi.org/10.1128/genomeA.00438-13>.
- Carson S, Bruff E, DeFoor W, Dums J, Groth A, Hatfield T, Iyer A, Joshi K, McAdams S, Miles D, Miller D, Oufkir A, Raynor B, Riley S, Roland S, Rozier H, Talley S, Miller ES. 2015. Genome sequences of six *Paenibacillus larvae* Siphoviridae phages. *Genome Announc* 3(3):e00101-15. <https://doi.org/10.1128/genomeA.00101-15>.
- Beims H, Wittmann J, Bunk B, Spröer C, Rohde C, Günther G, Rohde M, von der Ohe W, Steinert M. 2015. *Paenibacillus larvae*-directed bacteriophage HB10c2 and its application in American foulbrood-affected honey bee larvae. *Appl Environ Microbiol* 81:5411–5419. <https://doi.org/10.1128/AEM.00804-15>.
- Tsourkas P, Yost D, Krohn A, LeBlanc L, Zhang A, Stamereilers C, Amy PS. 2015. Complete genome sequences of nine phages capable of infecting *Paenibacillus larvae*, the causative agent of American foulbrood disease of honeybees. *Genome Announc* 3(5):e01120-15. <https://doi.org/10.1128/genomeA.01120-15>.
- Dingman DW, Bakhiet N, Field CC, Stahly DP. 1984. Isolation of two bacteriophages from *Bacillus larvae*, PBL1 and PBL0.5, and partial characterization of PBL1. *J Gen Virol* 65:1101–1105. <https://doi.org/10.1099/0022-1317-65-6-1101>.
- Casjens SR, Gilcrease EB. 2009. Determining DNA packaging strategy by analysis of the termini of the chromosomes in tailed-bacteriophage virions, p. 91–111. In Clokie MRJ, Kropinski AM (ed), *Bacteriophages: methods and protocols, volume 2: molecular and applied aspects*. Humana Press, New York, NY. https://doi.org/10.1007/978-1-60327-565-1_7.
- Merrill BD, Ward AT, Grose JH, Hope S. 2016. Software-based analysis of bacteriophage genomes, physical ends, and packaging strategies. *BMC Genomics* 17:679. <https://doi.org/10.1186/s12864-016-3018-2>.
- Stamereilers C, LeBlanc L, Yost D, Amy PS, Tsourkas PK. 2016. Comparative genomics of 9 novel *Paenibacillus larvae* bacteriophages. *Bacteriophage* 6:e1220349. <https://doi.org/10.1080/21597081.2016.1220349>.
- Xu J, Hendrix RW, Duda RL. 2004. Conserved translational frameshift in dsDNA bacteriophage tail assembly genes. *Molecular Cell* 16:11–21. <https://doi.org/10.1016/j.molcel.2004.09.006>.
- Xu J, Hendrix RW, Duda RL. 2013. A balanced ratio of proteins from gene G and frameshift-extended gene GT is required for phage lambda tail assembly. *J Mol Biol* 425:3476–3487. <https://doi.org/10.1016/j.jmb.2013.07.002>.