**GENOME SEQUENCES** 





## Complete Genome Sequence of *Rhodococcus* sp. Strain SGAir0479, Isolated from Indoor Air Collected in Singapore

Elaine L. Oliveira,<sup>a</sup> Daniela I. Drautz-Moses,<sup>a</sup> Akira Uchida,<sup>a</sup>
Rikky W. Purbojati,<sup>a</sup> Anthony Wong,<sup>a</sup> Kavita K. Kushwaha,<sup>a</sup> Alexander Putra,<sup>a</sup> Ngu War Aung,<sup>a</sup> Balakrishnan N. V. Premkrishnan,<sup>a</sup> Cassie E. Heinle,<sup>a</sup> Vineeth Kodengil Vettath,<sup>a</sup>
Ana Carolina M. Junqueira,<sup>b</sup> Stephan C. Schuster<sup>a</sup>

<sup>a</sup>Singapore Centre for Environmental Life Sciences Engineering, Nanyang Technological University, Singapore <sup>b</sup>Departamento de Genética, Instituto de Biologia, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

**ABSTRACT** The complete genome sequence of *Rhodococcus* sp. strain SGAir0479 is presented here. This organism was isolated from an air sample collected in an indoor location in Singapore. The consensus assembly generated one chromosome of 4.86 Mb (G+C content of 69.8%) and one plasmid of 104,493 bp.

Members of the genus *Rhodococcus* are naturally present in diverse temperate and extreme environments, and they can persist and grow in highly contaminated soils and waters (1). Some strains show pathogenicity for humans, animals, and plants (2). In addition, most of the described species are capable of metabolizing a wide variety of environmental pollutants, including trichloroethene, haloalkanes, and dibenzothiophene (3–6).

*Rhodococcus* sp. strain SGAir0479 was isolated from an air sample collected in an indoor area in Singapore (1°20'42.5"N 103°40'44.2"E) using an Andersen single-stage impactor (SKC BioStage, USA) operating at 28.3 liters/min for 3 min. The air was impacted onto Trypticase soy agar (TSA) (Becton, Dickinson, USA) plates, which were then incubated overnight at 30°C. CFU were manually isolated, and strain SGAir0479 was further cultured in lysogeny broth (LB; Becton, Dickinson, USA) overnight at 30°C, followed by genomic DNA extraction with the Wizard genomic DNA purification kit (Promega, USA). Preliminary taxon identification screening was performed with Sanger sequencing using 16S rRNA universal primers 27F and 1392R (7) and subsequent BLASTn search of the sequencing result. The PacBio library was prepared with the SMRTbell template prep kit version 1.0 (Pacific Biosciences, USA) and subjected to single-molecule real-time (SMRT) sequencing on the PacBio RS II platform, which generated a total of 93,748 subreads with a combined total of 1,199,875,648 bases.

The sequencing reads were then assembled with the Hierarchical Genome Assembly Process (HGAP) version 3 (8) implemented in the PacBio SMRT Analysis 2.3.0 package. Polishing of the assembly was performed with Quiver (8). The consensus assembly generated two contigs, one chromosome of 4.86 Mb (156.5-fold coverage) and one plasmid of 104,493 bp (68.2-fold coverage). The chromosomal contig showed a mean G+C content of 69.8%.

The NCBI Prokaryotic Genome Annotation Pipeline (PGAP) version 4.2 (9) was used for genome annotation. Unless specified, all software was run using default settings. A total of 4,616 genes were predicted with 4,461 protein-coding genes (PCGs), 12 rRNA genes (4 each of 5S, 16S, and 23S rRNAs), 53 tRNAs, 3 noncoding RNAs, and 87 pseudogenes. Based on Rapid Annotations using Subsystems Technology (RAST) (10) analysis (using the ClassicRAST annotation scheme and addition of the "fix frameshifts" option), the three subsystem categories with the highest feature counts were amino acids and derivatives (488), carbohydrate metabolism (400), and cofactors, vitamins, A, Purbojati RW, Wong A, Kushwaha KK, Putra A, Aung NW, Premkrishnan BNV, Heinle CE, Vettath VK, Junqueira ACM, Schuster SC. 2019. Complete genome sequence of *Rhodococcus* sp. strain SGAir0479, isolated from indoor air collected in Singapore. Microbiol Resour Announc 8:e00622-19. https://doi.org/10.1128/ MRA.00622-19.

Citation Oliveira EL, Drautz-Moses DI, Uchida

**Editor** Jason E. Stajich, University of California, Riverside

**Copyright** © 2019 Oliveira et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Stephan C. Schuster, SCSchuster@ntu.edu.sg.

E.L.O. and D.I.D.-M. contributed equally to this work.

Received 30 May 2019 Accepted 23 August 2019 Published 3 October 2019 prosthetic groups, and pigment formation (321). In this respect, at least 66 genes were found to be potentially involved in metabolism of aromatic compounds. Among these, 11 genes are part of the biphenyl degradation pathway (e.g., *bphC, bphD, bphI*, and *bphj2*) previously described to be involved in polychlorinated biphenyl (PCB) degradation (6, 11). The PCB degradation is a particular metabolic capability which may provide this microorganism with the ability to perform aerobic bioremediation (12).

Average nucleotide identity (ANI) analysis (13) showed that strain SGAir0479 has only 84.0% sequence identity to the genome of the closest species, *Rhodococcus agglutinans*. These values are below the ANI criteria for accurate taxonomic classification of strain SGAir0479 on the species level (minimum of 95 to 96% identity required) (14). However, it is sufficient to show the relationship between strain SGAir0479 and the *Rhodococcus* genus. To further confirm or refute this taxonomic classification, Phyla-AMPHORA (15) was run using MarkerScanner.pl with the added DNA flag and using MarkerAlignTrim.pl with the options WithReference and OutputFormat phylip; 16S identification was also performed. The results showed 92.9% identity to *Rhodococcus* genus.

**Data availability.** The complete genome sequences of *Rhodococcus* sp. strain SGAir0479 and its plasmid have been deposited in DDBJ/EMBL/GenBank under the accession numbers CP039432 and CP039433, respectively, and in the SRA under accession number SRR9043824.

## ACKNOWLEDGMENTS

This work was supported by a Singapore Ministry of Education Academic Research Fund Tier 3 grant (MOE2013-T3-1-013).

We thank Anjali Bansal Gupta for providing advice on the manuscript review.

## REFERENCES

- Guevara G, Castillo Lopez M, Alonso S, Perera J, Navarro-Llorens JM. 2019. New insights into the genome of *Rhodococcus ruber* strain Chol-4. BMC Genomics 20:332. https://doi.org/10.1186/s12864-019-5677-2.
- Majidzadeh M, Fatahi-Bafghi M. 2018. Current taxonomy of *Rhodococcus* species and their role in infections. Eur J Clin Microbiol Infect Dis 37:2045–2062. https://doi.org/10.1007/s10096-018-3364-x.
- Chen BS, Médici R, van der Helm MP, van Zwet Y, Gjonaj L, van der Geest R, Otten LG, Hanefeld U. 2018. *Rhodococcus* strains as source for enereductase activity. Appl Microbiol Biotechnol 102:5545–5556. https://doi .org/10.1007/s00253-018-8984-7.
- Brooks SL, Van Hamme JD. 2012. Whole-genome shotgun sequence of *Rhodococcus* species strain JVH1. J Bacteriol 194:5492–5493. https://doi .org/10.1128/JB.01066-12.
- McLeod MP, Warren RL, Hsiao WWL, Araki N, Myhre M, Fernandes C, Miyazawa D, Wong W, Lillquist AL, Wang D, Dosanjh M, Hara H, Petrescu A, Morin RD, Yang G, Stott JM, Schein JE, Shin H, Smailus D, Siddiqui AS, Marra MA, Jones SJM, Holt R, Brinkman FSL, Miyauchi K, Fukuda M, Davies JE, Mohn WW, Eltis LD. 2006. The complete genome of *Rhodococcus* sp. RHA1 provides insights into a catabolic powerhouse. Proc Natl Acad Sci U S A 103:15582–15587. https://doi.org/10.1073/pnas .0607048103.
- Martínková L, Uhnáková B, Pátek M, Nesvera J, Kren V. 2009. Biodegradation potential of the genus *Rhodococcus*. Environ Int 35:162–177. https://doi.org/10.1016/j.envint.2008.07.018.
- Weisburg WG, Barns SM, Pelletier DA, Lane DJ. 1991. 165 ribosomal DNA amplification for phylogenetic study. J Bacteriol 173:697–703. https:// doi.org/10.1128/jb.173.2.697-703.1991.
- Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013.

Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563. https://doi.org/10.1038/nmeth .2474.

- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi .org/10.1093/nar/gkw569.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). Nucleic Acids Res 42:D206–D214. https://doi.org/10.1093/nar/qkt1226.
- Seto M, Kimbara K, Shimura M, Hatta T, Fukuda M, Yano K. 1995. A novel transformation of polychlorinated biphenyls by *Rhodococcus* sp. strain RHA1. Appl Environ Microbiol 61:3353–3358.
- Garrido-Sanz D, Manzano J, Martín M, Redondo-Nieto M, Rivilla R. 2018. Metagenomic analysis of a biphenyl-degrading soil bacterial consortium reveals the metabolic roles of specific populations. Front Microbiol 9:232. https://doi.org/10.3389/fmicb.2018.00232.
- Shen L, Liu Y, Xu B, Wang N, Zhao H, Liu X, Liu F. 2017. Comparative genomic analysis reveals the environmental impacts on two Arcticibacter strains including sixteen Sphingobacteriaceae species. Sci Rep 7:2055. https://doi.org/10.1038/s41598-017-02191-4.
- Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. Proc Natl Acad Sci U S A 106: 19126–19131. https://doi.org/10.1073/pnas.0906412106.
- Wang Z, Wu M. 2013. A phylum-level bacterial phylogenetic marker database. Mol Biol Evol 30:1258–1262. https://doi.org/10.1093/molbev/ mst059.