

http://pubs.acs.org/journal/acsodf

Novel Stereoselective Syntheses of (+)-Streptol and (–)-1-epi-Streptol Starting from Naturally Abundant (–)-Shikimic Acid

Xing-Liang Zhu, Yong-Qiang Luo, Lei Wang, Yong-Kang Huang, Yun-Gang He, Wen-Jing Xie, Shi-Ling Liu,* and Xiao-Xin Shi*



1. INTRODUCTION

 C_7 -cyclitols are an important category of natural products possessing a broad spectrum of biological activities, so that a lot of natural C_7 -cyclitols and their derivatives have become the targets of organic synthesis due to these attractive biological properties.¹ (+)-Streptol [also known as (+)-valienol, **1** in Figure 1] and (-)-1-*epi*-streptol [or (-)-1-*epi*-valienol, **2** in

steps in 29 and 36% overall yields, respectively.





Figure 1] are two members of C_7 -cyclitols. (+)-Streptol 1 has been isolated from some microbial organisms² such as *Streptomyces* sp. no. 1409,^{2a} *Dacthylosporangium aurantiacum* SANK 61299,^{2b} and *Streptomyces lincolnensis* DSM 40355.^{2c} It has shown plant growth inhibitory activity^{2a,b,3} and antitumor activity.⁴ (-)-1-*epi*-Streptol 2 is an intermediate involved in the biosyntheses of acarbose and salbostatin (α -glucosidase inhibitors) in *Actinoplanes* and *Streptomyces*.⁵ Several total syntheses of (+)-streptol 1 and (-)-1-*epi*-streptol 2 have been reported.⁶ (+)-Streptol 1 has been synthesized from (*R*,*R*)tartaric acid,^{6a} D-glucose,^{6b,c} D-gluconolactone,^{6d,e} and some chiral building blocks.^{6f-h} (-)-1-*epi*-Streptol 2 has also been synthesized from D-glucose^{6b,i} and D-gluconolactone.^{6e} Despite the above-mentioned syntheses, novel practical and efficient syntheses of cyclitols 1 and 2 might be highly desirable to further investigate the biological activities of these two particular C_7 -cyclitols and their derivatives.

(-)-Shikimic acid (see Figure 1) has captured worldwide attention⁷ in recent decades due to its wide use in the syntheses of drugs or pharmaceutically valuable molecules (Tamiflu, valiolamine, valienamine, and so on)⁸ as well as some chiral building blocks.⁹ Many researchers have tried to improve the production of (-)-shikimic acid by means of extraction from plants,^{7a,10} fermentation based on microbial engineering,^{7c,e,g,11} and chemical syntheses,^{7a,12} (-)-Shikimic acid has been found in many plant species.¹³ It is noted to be in extremely high abundance in Chinese star anise (Illicium verum)¹⁴ and thus can be readily manufactured in a large quantity by extraction from the Chinese star anise due to the development of new methods for rapid and high-yielding extraction.¹⁵ Recently, we have been engaged in developing novel stereoselective syntheses of various pharmaceutically valuable molecules from (-)-shikimic acid.^{8a-j} Herein, we want to disclose highly stereoselective, efficient, and practical syntheses of (+)-streptol 1 and (-)-1-epi-streptol 2 by commercially available and inexpensive (-)-shikimic acid as the starting material.

2. RESULTS AND DISCUSSION

The novel total syntheses of (+)-streptol 1 and (-)-1-*epi*streptol 2 starting from (-)-shikimic acid are depicted in Scheme 1. The total syntheses can be briefly described as follows: esterification of (-)-shikimic acid first produced

 Received:
 May 12, 2021

 Accepted:
 June 14, 2021

 Published:
 June 23, 2021







methyl (-)-shikimate 3 in 97% yield.¹⁶ Next, when compound 3 was exposed to 5.0 equiv of SOCl₂ in anhydrous N.Ndimethylformamide (DMF) at room temperature (r.t.) for 18 h, compound 4 was thus obtained in 92% yield; the regioselectivity of this chlorination is very high just like a previous report.^{8e} Reaction of compound 4 with 2.0 equiv of K_2CO_3 in absolute methanol at room temperature for 9 h furnished epoxide 5 in 95% yield. Protection of the hydroxyl at the C-5 position with the *tert*-butyldiphenylsilyl (TBDPS) group could be achieved by treatment of compound 5 with 1.5 equiv of tert-butyldiphenylsilyl chloride and 3.0 equiv of imidazole in CH₂Cl₂ under reflux for 2 h; compound 6 was thus obtained in 93% yield. When compound 6 was treated with 5.0 equiv of AcOH and 0.5 equiv of CF₃COOH in CH₂Cl₂ under reflux for 6 h, high regioselective ring openning of epoxide took place smoothly to give compound 7 in 86% yield, and during this epoxide opening, the nucleophile (AcOH) favorably attacked the more reactive allylic C-3 position rather than the C-4 position. When compound 7 was treated with 1.5 equiv of Ac₂O, 2.0 equiv of Et₃N, and 0.1

equiv of *N*,*N*-dimethylaminopyridine (DMAP) in anhydrous ethyl acetate at 0 $^{\circ}$ C for 2 h, acylation of the hydroxyl at the C-4 position occurred rapidly to afford compound **8** in 96% yield.

Subsequently, RuCl₃-catalyzed highly stereoselective dihydroxylation¹⁷ of $\alpha_{,\beta}$ -unsaturated ester 8 produced the desired pinacol 9. When compound 8 was treated with 0.02 equiv of RuCl₃ and 1.5 equiv of NaIO₄ in a mixed solvent (CH₃CN/ EtOAc/H₂O = 3:3:1) at -5 °C for 1 h, compound 9 could be obtained in 89% yield. During the stereoselective dihydroxylation of compound 8, the ruthenium catalyst coordinated with the double bond in the opposite direction of the OAc (at C-3) and o-tert-butyldiphenylsilyl (OTBDPS) (at C-5) groups due to their high steric hindrance, so that two hydroxyls at C-1 and C-2 of compound 9 should have the desired downward orientation. Compound 9 was then exposed to 5.0 equiv of NaBH₄ at room temperature for 1 h in a mixed solvent (EtOAc/H₂O = 10:1), the ester group (CO₂Me) at C-1 was selectively reduced, and other two ester groups (two OAc at C-3 and C-4) remained intact during the reaction; compound 10 was thus obtained in 91% yield.

ACS Omega



Figure 2. ${}^{1}H-{}^{1}H$ COSY and ${}^{1}H-{}^{1}H$ NOESY spectra of 9.

Next, when compound **10** was treated with 3.0 equiv of benzoyl chloride, 4.0 equiv of Et_3N , and 0.1 equiv of p-dimethylaminopyridine (DMAP) in dichloromethane at 0 °C to room temperature for 5 h, selective benzoylation of primary and secondary hydroxyls occurred smoothly to afford compound **11** in 94% yield; in the meantime, the tertiary hydroxyl in compound **10** remained unchanged during the reaction probably due to its high steric hindrance. Exposure of compound **11** to 5.0 equiv of SOCl₂ and 3.0 equiv of pyridine in dichloromethane under reflux for 6 h led to the formation of olefinic compound **12** in 87% yield via regioselective β -

elimination. The silyl protecting group (TBDPS) was then removed by treatment of compound 12 with 4.5 equiv of Bu_4NF and 4.5 equiv of AcOH in tetrahydrofuran for 8 h at room temperature; compound 13 could be thus obtained in 92% yield.

Compound 13 is a common intermediate for syntheses of targeted molecules 1 and 2. When compound 13 was treated with a large excess of aqueous ammonia in methanol for 24 h at room temperature, all four acyl groups were removed in the one-pot reaction, and (-)-1-*epi*-streptol 2 was obtained in 90% yield. Next, when compound 13 was exposed to 2.0 equiv of

methanesulfonyl chloride (MsCl) and 1.5 equiv of Et₃N in ethyl acetate at 0 °C for 1 h, olefinic methanesulfonate I-A was formed and was immediately used for the following step without purification due to its unstability. When crude intermediate I-A was treated with 6.0 equiv of acetic acid (AcOH) and 3.0 equiv of 1,8-diazabicyclo[5,4,0]undec-7-ene (DBU) in toluene at 85 °C for 5 h according to our previous report,¹⁸ an S_N2-type substitution took place to furnish compound 14 in 80% yield over two steps. The (*R*) configuration of the chiral center at the C-1 position was inversed to an (*S*) configuration during the S_N2-type substitution. Finally, when compound 14 was treated with a large excess of aqueous ammonia in methanol for 24 h at room temperature, all five acyl groups were removed in the one-pot reaction, and (+)-streptol 1 was thus obtained in 90% yield.

In addition, some particular points in the above-described total syntheses (as shown in Scheme 1) are worthy to be further discussed in the following.

First, in the RuCl₃-catalyzed highly stereoselective dihydroxylation of α_{β} -unsaturated ester 8, the stereochemistry of product 9 has been unequivocally confirmed by the twodimensional (2D) NMR technique. ${}^{1}H-{}^{1}H$ correlation spectroscopy (COSY) and ¹H-¹H nuclear Overhauser effect (NOE) spectroscopy (NOESY) spectra of compound 9 are shown in Figure 2. As can be seen from the ${}^{1}H-{}^{1}H$ COSY spectrum of compound 9, the dd peak at 5.18 ppm could be assigned to proton H-4, which has correlation spots (a and b)with protons H-3 and H-5; the dd peak at 4.96 ppm could be assigned to proton H-3, which has correlation spots (c and a) with protons H-2 and H-4; the m peak at 4.19 ppm could be assigned to proton H-5, which has correlation spots (b and d)with protons H-4 and H-6; the d peak at 3.94 ppm could be assigned to proton H-2, which only has a correlation spot (c)with proton H-3; and the m peak at 1.86 ppm could be assigned to proton H-6, which only has a correlation spot (d)with proton H-5. As can be seen from the ¹H-¹H NOESY spectrum of compound 9, there are obvious NOE correction spots between H-3 and H-5, meaning that protons H-3 and H-5 have the cis relationship, there are obvious NOE correction spots between H-2 and H-4, and also there is no correction spot between H-2 and H-5, meaning that protons H-2 and H-4 have the cis relationship and that protons H-2 and H-5 have the trans relationship, so the chiral center at C-2 of compound **9** has an (S) configuration. According to the mechanism of Rucatalyzed dihydroxylation of olefins,^{17b} two hydroxyls at C-1 and C-2 positions in compound 9 should have the cis relationship, so the chiral center at C-1 of compound 9 has an (*R*) configuration.

Second, it is worth noting that chemoselectivity for the reduction of compound 9 with NaBH₄ was very high; a reasonable explanation for the high chemoselectivity of the reduction of compound 9 to compound 10 is proposed in Figure 3 according to the literature.¹⁹ The α -hydroxy group of CO₂Me in compound 9 first reacted with NaBH₄ to produce intermediate complex I-B, and then, it underwent intramolecular reduction of the methyl ester group (CO₂Me) to lead to the desired compound 10.

Third, as can be seen from Tables 1 and 2, ¹H and ¹³C NMR data of the samples of (+)-streptol 1 and (-)-1-*epi*-streptol 2 from present syntheses shown in Scheme 1 are consistent with the literature data for authentic samples.^{2a,6g} (Note that ¹H/¹³C NMR data of the authentic sample of compound 1 were published by Sakuda et al. in 1987^{2a} and ¹H/¹³C data of



Figure 3. Chemoselective reduction of 9 with NaBH₄.

Table 1. Comparison between ${}^{1}\text{H}/{}^{13}\text{C}$ NMR Data (δ ppm) of the Synthetic and Authentic Samples of (+)-Streptol 1^{*a*}

	synthetic sample		authentic sample	
positions	¹ H NMR	¹³ C NMR	¹ H NMR	¹³ C NMR
1	4.30	68.3	4.33	68.3
2	3.58	72.9	3.61	72.9
3	3.71	74.4	3.73	74.3
4	4.09	74.7	4.11	74.4
5		144.3		144.3
6	5.85	124.3	5.88	122.4
7	4.15 (7a)	63.4	4.17 (7a)	63.5
	4.24 (7b)		4.26 (7b)	
an o	1 .1 1			

^{*a*}D₂O was used as the solvent.

Table 2. Comparison between ${}^{1}\text{H}/{}^{13}\text{C}$ NMR Data (δ ppm) of the Synthetic and Authentic Samples of (-)-1-*epi*-Streptol 2^{*ab*}

	synthetic sample		authentic sample	
positions	¹ H NMR	¹³ C NMR	¹ H NMR	¹³ C NMR
1	4.10 (m)	71.3	4.17 (m)	73.6
2	3.36	71.9	3.43	74.2
3	3.42	75.1	3.49	77.4
4	4.10 (m)	75.5	4.17 (m)	77.8
5		138.3		140.6
6	5.51	124.9	5.58	127.2
7	3.99 (7a)	61.0	4.06 (7a)	63.3
	4.10 (m) (7b)		4.17 (m) (7b)	

^{*a*}D₂O was used as the solvent. ^{*b*}The same difference (0.07 ppm) of the δ value for each proton between ¹H NMR spectra of synthetic and authentic samples might result from a zero-point calibration error.

the authentic sample of compound 2 were published by Leermann et al. in 2010.^{6g})

3. CONCLUSIONS

In conclusion, we have performed stereoselective total syntheses of (+)-streptol [(+)-valienol] 1 and (-)-1-*epi*-streptol [(-)-1-*epi*-valienol] 2 starting from the naturally abundant (-)-shikimic acid. (+)-Streptol 1 has been synthesized starting from the naturally abundant (-)-shikimic acid in 14 steps in 29% overall yield; (-)-1-*epi*-streptol 2 has also been synthesized starting from (-)-shikimic acid in 12 steps in 36% overall yield. Moreover, the stereochemistry of key intermediate compound 9 has been unequivocally confirmed by analyses of its ${}^{1}\text{H}{-}{}^{1}\text{H}$ COSY and ${}^{1}\text{H}{-}{}^{1}\text{H}$ NOESY spectra (see Figure 2).

Compared with the known syntheses of (+)-streptol 1 and (-)-1-*epi*-streptol 2,⁶ present total syntheses, albeit with moderate overall yields, are more economic and practical because none of the expensive reagents such as lithium diisopropylamide (LDA),^{6e} diisobutylaluminum hydride

(DIBAL-H),^{6a,h} K-selectride^{6b,e} LiHMDS,⁶ⁱ TBSOTf,^{6f} and excessive Ag(OAc)₂^{6g} were used and also none of the drastic reaction conditions such as Ph₂O/230 °C,^{6f} LDA/-78 °C,^{6c,e} (COCl)₂/dimethyl sulfoxide (DMSO)/-78 °C,^{6d} trifluoro-acetic anhydride (TFAA)/DMSO/-78 °C,^{6c,i} and DIBAL-H/-78 °C^{6a,h} were used in every step.

4. EXPERIMENTAL SECTION

4.1. General Method. ¹H NMR and ¹³C NMR spectra were acquired on a Bruker AM-400 instrument; chemical shifts are given on the δ scale as parts per million (ppm) with tetramethylsilane (TMS) as the internal standard. IR spectra were recorded with a Nicolet Magna IR-550 instrument. Mass spectra were acquired with an HP1100 LC-MS spectrometer. Optical rotations of chiral compounds were measured on a PerkinElmer polarimeter at room temperature. Melting points were determined on a Mel-TEMP II apparatus. Column chromatography was performed on silica gel (Qingdao Ocean Chemical Corp.). Methyl shikimate **3** was prepared in 97% yield according to a known procedure.¹⁶

4.2. Methyl (3S,4S,5R)-3-Chloro-4,5-bis(formyloxy)cyclohex-1-ene-1-carboxylate 4. SOCl₂ (63.42 g, 533.1 mmol) was dropwise added to anhydrous DMF (120 mL) at room temperature for 15 min. The resulting solution was cooled to 5 °C by an ice bath, and then, the crushed methyl shikimate 3 (20.06 g, 106.6 mmol) was slowly added in portions. After the addition was finished, the ice bath was removed, and the solution was further stirred at room temperature for 18 h. The reaction solution was poured into stirred biphasic solvents of toluene (500 mL) and ice-water (600 mL). After the above well-stirred biphasic mixture was put into an ice bath, powered K_2CO_3 (148.0 g, 1.071 mol) was slowly added until the pH was adjusted to 8-9. The biphasic mixture was transferred into a separatory funnel, two phases were separated, and the aqueous phase was extracted again with toluene (200 mL). The extracts were combined and dried over anhydrous MgSO₄. Evaporation of toluene under vacuum gave an oily crude product that was purified by flash chromatography (eluent, EtOAc/hexane = 1:5) to furnish compound 4 (25.76 g, 98.08 mmol) as colorless oil in 92% yield. $[\alpha]_{D}^{25} = +28.3$ (c 1.0, CHCl₃). ¹H NMR (400 MHz, $CDCl_3$) δ 2.45–2.54 (m, 1H, H-6), 3.04 (dd, J_1 = 17.8 Hz, J_2 = 5.7 Hz, 1H, another H-6), 3.77 (s, 3H, OCH₃), 4.68 (dd, $J_1 =$ 2.5 Hz, $J_2 = 8.1$ Hz, 1H, H-3), 5.16–5.24 (m, 1H, H-5), 5.47 $(dd, J_1 = 9.5 Hz, J_2 = 8.1 Hz, 1H, H-4), 6.79 (d, J = 2.5 Hz, 1H, 1H)$ H-2), 8.03 (s, 1H, OCHO), 8.14 (s, 1H, another OCHO). ¹³C NMR (100 MHz, CDCl₃) δ 165.14, 159.65, 159.39, 135.17, 128.81, 73.25, 67.94, 55.53, 52.48, 29.13. High-resolution mass spectrometry (HRMS) (electrospray ionization (ESI)) calcd for C₁₀H₁₁O₆NaCl [M + Na]⁺: 285.0142, found: 285.0140. IR (neat) ν 2955, 1728, 1377, 1254, 1171, 1077, 752 cm⁻¹.

4.3. Methyl (3*R*,4*S*,5*R*)-3,4-Epoxy-5-hydroxy-cyclohex-1-ene-1-carboxylate 5. Compound 4 (10.05 g, 38.26 mmol) was dissolved in anhydrous methanol (100 mL), and powered K_2CO_3 (10.58 g, 76.55 mmol) was added. The reaction mixture was allowed to be stirred at room temperature for 9 h. When the reaction was finished (checked by thin-layer chromatography (TLC); eluent, EtOAc/hexane = 1:3), the potassium salt was filtered by suction, and the filtrate was concentrated under vacuum to give an oily residue, which was partitioned between ethyl acetate (200 mL) and water (30 mL). Two phases were separated, and the aqueous solution was extracted twice with ethyl acetate (50 × 2 mL). Organic extracts were combined and dried over anhydrous MgSO₄. Removal of the solvent by vacuum distillation gave an oily crude product that was purified by flash chromatography (eluent, EtOAc/hexane = 1:3) to furnish compound **5** (6.186 g, 36.35 mmol) as colorless oil in 95% yield. $[\alpha]_D^{25} = +227.8$ (*c* 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 2.20 (ddd, $J_1 = 17.5$ Hz, $J_2 = 5.2$ Hz, $J_3 = 3.3$ Hz, 1H, H-6), 2.68 (dd, $J_1 = 17.5$ Hz, $J_2 = 3.0$ Hz, 1H, another H-6), 3.04 (d, J = 5.6 Hz, 1H, OH), 3.45–3.37 (m, 1H, H-5), 3.48 (dd, $J_1 = 4.6$ Hz, $J_2 = 3.5$ Hz, 1H, H-4), 3.68 (s, 3H, OCH₃), 4.44–4.48 (m, 1H, H-3), 7.04 (dd, $J_1 = 3.7$ Hz, $J_2 = 3.3$ Hz, 1H, H-2). ¹³C NMR (100 MHz, CDCl₃) δ 166.83, 133.59, 130.66, 63.04, 56.01, 52.16, 46.35, 29.11. HRMS (ESI) calcd for C₈H₁₀O₄Na [M + Na]⁺: 193.0477, found: 193.0475. IR (neat) ν 3421, 2954, 1715, 1645, 1439, 1268, 1096, 1005 cm⁻¹.

4.4. Methyl (3R,4S,5R)-5-(tert-Butyldiphenylsilyloxy)-3,4-epoxy-cyclohex-1-ene-1-carboxylate 6. Compound 5 (5.013 g, 29.46 mmol) was dissolved in CH₂Cl₂ (50 mL), TBDPSCl (12.15 g, 44.20 mmol) and imidazole (6.017 g, 88.38 mmol) were then added. The resulting solution was heated to reflux (41 °C) and further stirred for 2 h. After the reaction was complete (ckecked by TLC; eluent, EtOAc/ hexane = 1:5), the solution was concentrated under vacuum to remove dichloromethane, and then ethyl acetate (60 mL) and an aqueous solution of potassium carbonate (15% w/w, 50 mL) were added. After the mixture was vigorously stirred for 15 min at room temperature, two phases were separated, and the aqueous solotion was extracted again with ethyl acetate (50 mL). Organic extracts were combined and dried over anhydrous MgSO₄. Removal of the solvent by vacuum distillation gave an oily crude product that was purified by flash chromatography (eluent, EtOAc/hexane = 1:7) to furnish compound 6 (11.20 g, 27.41 mmol) as colorless oil in 93% yield. $[\alpha]_D^{25} = +113.5$ (c 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.03 (s, 9H, t-Bu), 2.09 (ddd, J_1 = 17.2 Hz, J_2 = 4.9 Hz, J₃ = 3.2 Hz, 1H, H-6), 2.73 (dd, J₁ = 17.2 Hz, J₂ = 2.0 Hz, 1H, another H-6), 3.35 (ddd, J_1 = 4.9 Hz, J_2 = 2.7 Hz, J_3 = 2.0 Hz, 1H, H-5), 3.44 (dd, $J_1 = 4.5$ Hz, $J_2 = 2.7$ Hz, 1H, H-4), 3.75 (s, 3H, OCH₃), 4.52 (dd, J₁ = 4.5 Hz, J₂ = 2.4 Hz, 1H, H-3), 7.16 (dd, J_1 = 4.5 Hz, J_2 = 3.2 Hz, 1H, H-2), 7.35–7.47 (m, 6H, Ph-H), 7.61–7.69 (m, 4H, Ph-H). ¹³C NMR (100 MHz, $CDCl_3$) δ 166.59, 135.77 (2C), 135.72 (2C), 133.65, 133.39, 133.15, 131.30, 129.95, 129.93, 127.81 (2C), 127.76 (2C), 64.98, 56.15, 51.98, 46.79, 29.14, 26.87 (3C), 19.24. HRMS (ESI) calcd for $C_{24}H_{28}O_4SiNa [M + Na]^+$: 431.1655, found: 431.1652. IR (neat) v 3071, 2955, 2931, 2858, 1719, 1647, 1429, 1263, 1109, 1094, 1008, 938, 823, 705, 611, 508 cm⁻¹.

4.5. Methyl (3S,4R,5R)-5-(tert-Butyldiphenylsilyloxy)-3-acetoxy-4-hydroxyl-cyclohex-1-ene-1-carboxylate 7. Compound 6 (10.15 g, 24.84 mmol) was first dissolved in dichloromethane (100 mL), and AcOH (7.458 g, 124.2 mmol) and CF₃COOH (1.416 g, 12.42 mmol) were then added. The resulting solution was heated to reflux (41 °C), and further stirred under reflux for 6 h. After the reaction was complete (checked by TLC; eluent, EtOAc/hexane = 1:4), dichloromethane was removed by vacuum distillation. Ethyl acetate (120 mL) and an aqueous solution of potassium carbonate (20% w/w, 100 mL) were added. After the mixture was vigorously stirred for 15 min at room temperature, two phases were separated, and the aqueous solution was extracted twice with ethyl acetate (2×100 mL). Organic extracts were combined and dried over anhydrous MgSO₄. Removal of the solvent by vacuum distillation gave an oily crude product that

was purified by flash chromatography (eluent, EtOAc/hexane = 1:4) to give compound 7 (10.01 g, 21.36 mmol) as colorless oil in 86% yield. $[\alpha]_{D}^{25} = +28.2$ (c 1.0, CHCl₃). ¹H NMR (400 MHz, $CDCl_3$) δ 1.09 (s, 9H, t-Bu), 2.10 (s, 3H, CH₃ in Ac), 2.25-2.35 (m, 1H, H-6), 2.51-2.59 (m, 1H, another H-6), 3.67 (s, 3H, OCH₃), 3.80-3.89 (m, 2H, H-4 and H-5), 5.34 $(dd, J_1 = 2.8 Hz, J_2 = 4.2 Hz, 1H, H-3), 6.53 (dd, J_1 = 2.8 Hz, J_2 = 2.8 Hz)$ $J_2 = 2.6$ Hz, 1H, H-2), 7.36–7.48 (m, 6H, Ph-H), 7.66–7.73 (m, 4H, Ph-H). ¹³C NMR (100 MHz, CDCl₃) δ 170.67, 166.02, 135.93 (2C), 135.67 (2C), 134.48, 133.12, 133.04, 130.09, 129.98, 127.99, 127.86 (2C), 127.84 (2C), 74.57, 73.54, 71.63, 52.05, 32.59, 27.00 (3C), 21.02, 19.32. HRMS (ESI) calcd for $C_{26}H_{32}O_6SiNa [M + Na]^+$: 491.1866, found: 491.1860. IR (neat) v 3500, 3071, 2954, 2933, 2858, 1723, 1659, 1430, 1373, 1235, 1112, 1088, 970, 823, 705, 612, 506 cm^{-1} .

4.6. Methyl (3S,4R,5R)-5-(tert-Butyldiphenylsilyloxy)-3,4-diacetoxy-cyclohex-1-ene-1-carboxylate 8. Compound 7 (9.986 g, 21.31 mmol), Et₃N (4.313 g, 42.62 mmol), and DMAP (260.5 mg, 2.132 mmol) were dissolved in anhydrous ethyl acetate (100 mL), and the solution was then cooled to 0 °C by an ice bath. Ac₂O (3.265 g, 31.98 mmol) was then dropwise added for 10 min. The mixture was further stirred at 0 °C for 2 h. After the reaction was complete (checked by TLC; eluent, EtOAc/hexane = 1:5), an aqueous solution of HCl (2 N, 50 mL) was added to quench the reaction. After the mixture was vigorously stirred for 5 min, the two phases were separated by a separatory funnel. The aqueous solution was extracted again with ethyl acetate (50 mL). Organic extracts were combined, washed with an aqueous solution of potassium carbonate (15% w/w, 50 mL), and then dried over anhydrous MgSO4. Removal of the solvent by vacuum distillation gave an oily crude product that was purified by flash chromatography (eluent, EtOAc/hexane = 1:6) to furnish compound 8 (10.45 g, 20.46 mmol) as white crystals in 96% yield; mp 88–90 °C. $[\alpha]_D^{25} = +34.5$ (c 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.05 (s, 9H, t-Bu), 1.79 (s, 3H, CH₃ in Ac), 2.04 (s, 3H, CH₃ in another Ac), 2.34–2.45 (m, 1H, H-6), 2.56 (dd, J_1 = 17.8 Hz, J_2 = 5.8 Hz, 1H, another H-6), 3.69 (s, 3H, OCH₃), 4.03 (dd, $J_1 = 9.0$ Hz, $J_2 = 5.8$ Hz, $J_3 =$ 5.6 Hz, 1H, H-5), 5.27 (dd, $J_1 = 9.0$ Hz, $J_2 = 7.2$ Hz, 1H, H-4), 5.41 (dd, $J_1 = 7.2$ Hz, $J_2 = 5.4$ Hz, 1H, H-3), 6.55 (dd, $J_1 = 2.2$ Hz, $J_2 = 5.4$ Hz, 1H, H-2), 7.35–7.46 (m, 6H, Ph-H), 7.62– 7.71 (m, 4H, Ph-H). ¹³C NMR (100 MHz, CDCl₃) δ 170.39, 170.22, 165.88, 136.00 (2C), 135.72 (2C), 133.76, 133.07, 130.20, 130.00, 129.78, 127.78 (2C), 127.66 (2C), 74.27, 71.50, 68.46, 52.12, 32.64, 26.78 (3C), 20.92, 20.89, 19.21. HRMS (ESI) calcd for C₂₈H₃₄O₇SiNa [M + Na]⁺: 533.1971, found: 533.1973. IR (KBr film) v 3072, 2955, 2930, 2857, 1750, 1723, 1657, 1430, 1367, 1240, 1112, 1061, 974, 822, 705, 610, 506 $\rm cm^{-1}$.

4.7. (1*R*,2*S*,3*S*,4*R*,5*R*)-5-(*tert*-Butyldiphenylsilyloxy)-3,4-diacetoxy-1,2-dihydroxy-1-methoxycarbonyl-cyclohexane 9. Sodium periodate (4.412 g, 20.63 mmol), ruthenium trichloride (57.0 mg, 0.275 mmol), and water (10 mL) were added into a round-bottomed flask that was equipped with a stirrer bar. The mixture was stirred at room temperature for 15 min, and the color changed to bright yellow. Compound 8 (7.022 g, 13.75 mmol) was dissolved in a mixed solvent of ethyl acetate (30 mL) and acetonitrile (30 mL), and the resulting solution was cooled to -5 °C by a salt ice bath. The above bright yellow aqueous viscous solution was added, and the mixture was further stirred at -5 °C for 1 h. After the reaction was complete (checked by TLC; eluent, EtOAc/hexane = 1:3), ethyl acetate (100 mL) and a saturated aqueous solution of Na₂S₂O₃ (80 mL) were added. The mixture was vigorously stirred for 15 min, and then, the two phases were separated by a separatory funnel. The aqueous solution was extracted twice with ethyl acetate $(2 \times 80 \text{ mL})$. Organic extracts were combined, washed with brine (20 mL), and then dried over anhydrous MgSO4. The organic solution was concentrated under vacuum to give the crude product, which was then purified by flash chromatography (eluent, EtOAc/hexane = 1:2) to afford compound 9 (6.667 g, 12.24mmol) as white crystals in 89% yield; mp 133–134 °C. $[\alpha]_D^{25}$ = -24.9 (c 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.01 (s, 9H, t-Bu), 1.77 (s, 3H, CH₃ in Ac), 1.79–1.88 (m, 2H, two H-6), 2.02 (s, 3H, CH₃ in another Ac), 2.51 (s, 1H, OH), 3.36 (s, 1H, another OH), 3.76 (s, 3H, OCH₃), 3.90 (d, J = 9.7 Hz, 1H, H-2), 4.14–4.23 (m, 1H, H-5), 4.96 (dd, $J_1 = 9.9$ Hz, $J_2 =$ 9.8 Hz, 1H, H-4), 5.17 (dd, $J_1 = 9.8$ Hz, $J_2 = 9.7$ Hz, 1H, H-3), 7.34-7.46 (m, 6H, Ph-H), 7.58-7.67 (m, 4H, Ph-H). ¹³C NMR (100 MHz, CDCl₃) δ 173.73, 171.44, 170.20, 135.86 (2C), 135.71 (2C), 133.26, 133.21, 129.94, 129.81, 127.73 (2C), 127.65 (2C), 75.75, 75.38, 73.94, 73.78, 68.68, 53.55, 38.82, 26.75 (3C), 20.84, 20.79, 19.19. HRMS (ESI) calcd for $C_{28}H_{36}O_{9}SiNa [M + Na]^{+}$: 567.2026, found: 567.2025. IR (KBr film) v 3490, 3072, 2957, 2934, 2858, 1746, 1429, 1367, 1243, 1113, 1047, 826, 705, 608, 504 cm⁻¹

4.8. (1S,2S,3S,4R,5R)-3,4-Diacetoxy-1,2-dihydroxy-1hydroxymethyl-5-(tert-butyldiphenylsilyloxy)cyclohexane 10. Compound 9 (5.447 g, 10.00 mmol) was dissolved in ethyl acetate (60 mL), and water (6 mL) was added. While the mixture was well stirred, NaBH₄ (1.895 g, 50.09 mmol) was added in portions for 15 min at room temperature. After the addition was finished, the mixture was further stirred for 1 h. Ethyl acetate (60 mL) and water (50 $\,$ mL) were added, and the biphasic mixture was vigorously stirred for 5 min. Two phases were separated, and the aqueous phase was extracted again with ethyl acetate (60 mL). Organic extracts were combined and dried over anhydrous MgSO₄. The organic solution was concentrated under vacuum to give the crude product, which was then purified by flash chromatography (eluent, EtOAc/hexane = 1:2) to afford compound 10 (4.705 g, 9.107 mmol) as white crystals in 91% yield; mp 174-176 °C. $[\alpha]_D^{25} = -14.7$ (c 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.01 (s, 9H, t-Bu), 1.40 (dd, J_1 = 13.8 Hz, J_2 = 9.2 Hz, 1H, H-6), 1.72 (dd, $J_1 = 13.8$ Hz, $J_2 = 5.0$ Hz, 1H, another H-6), 1.77 (s, 3H, CH₃ in Ac), 1.99 (s, 3H, CH₃ in another Ac), 2.90 (br. s, 1H, OH), 3.04 (br. s, 1H, OH), 3.30 (dd, $J_1 =$ 11.2 Hz, $J_1 = 6.0$ Hz, 1H, CHHOH), 3.43 (dd, $J_1 = 11.2$ Hz, J_2 = 3.5 Hz, 1H, CHHOH), 3.52 (d, J = 6.0 Hz, 1H, OH), 3.62 $(dd, J_1 = 9.3 Hz, J_2 = 5.7 Hz, 1H, H-2), 4.18-4.26 (m, 1H, H-2)$ 5), 4.96 (dd, J_1 = 8.8 Hz, J_2 = 8.5 Hz, 1H, H-4), 5.04 (dd, J_1 = 9.3 Hz, J₂ = 8.5 Hz, 1H, H-3), 7.32-7.44 (m, 6H, Ph-H), 7.59–7.68 (m, 4H, Ph-H). ¹³C NMR (100 MHz, CDCl₃) δ 171.89, 170.44, 135.91 (2C), 135.74 (2C), 133.53, 133.48, 129.89, 129.74, 127.72 (2C), 127.63 (2C), 75.86, 74.47, 74.36, 72.19, 72.18, 68.48, 37.91, 26.79 (3C), 20.93, 20.82, 19.19. HRMS (ESI) calcd for $C_{27}H_{36}O_8SiNa [M + Na]^+$: 527.2077, found: 527.2071. IR (KBr film) v 3572, 3493, 3072, 2952, 2932, 2860, 1734, 1709, 1428, 1379, 1234, 1112, 1049, 823, 706, 608, 512 cm⁻¹.

4.9. (1*S*,2*S*,3*R*,4*R*,5*R*)-2-Benzoyloxy-1-benzoyloxymethyl-5-(*tert*-butyldiphenylsilyloxy)-3,4-diacetoxy-1hydroxy-cyclohexane 11. Compound 10 (4.000 g, 7.742

mmol) was dissolved in dichloromethane (50 mL), and the solution was cooled to 0 °C by an ice bath. Et₃N (3.134 g, 30.97 mmol), DMAP (95.0 mg, 0.778 mmol), and BzCl (3.265 g, 23.23 mmol) were added in turn. After the addition was finished, the ice bath was removed, and the mixture was further stirred at room temperature for 5 h. When the reaction was complete (checked by TLC; eluent, EtOAc/hexane = 1:3), dichloromethane was removed by vacuum distillation. Ethyl acetate (120 mL) and an aqueous solution of potassium carbonate (15% w/w, 50 mL) were added, and the biphasic mixture was vigorously stirred for 2 h. Two phases were separated, and aqueous phase was extracted again with ethyl acetate (60 mL). Organic extracts were combined and successively washed with an aqueous solution of HCl (2 N, 30 mL) and brine (15 mL). The organic solution was dried over anhydrous MgSO4 and then concentrated under vacuum to give the crude product, which was purified by flash chromatography (eluent, EtOAc/hexane = 1:4) to afford compound 11 (5.276 g, 7.278 mmol) as white crystals in 94% yield; mp 127–128 °C. $[\alpha]_D^{25} = -20.2$ (c 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.04 (s, 9H, t-Bu), 1.70–1.80 (m, 1H, H-6), 1.81 (s, 3H, CH₃ in Ac), 1.85 (s, 3H, CH₃ in another Ac), 2.00 (dd, J_1 = 13.9 Hz, J_2 = 5.0 Hz, 1H, another H-6), 2.45 (br. s, 1H, OH), 3.99 (d, J = 11.6 Hz, 1H, CHHOBz), 4.16 (d, J = 11.6 Hz, 1H, CHHOBz), 4.38 (ddd, J₁ = 10.0 Hz, J_2 = 5.2 Hz, J_3 = 5.0 Hz, 1H, H-5), 5.29 (dd, J_1 = 10.0 Hz, *J*₂ = 10.1 Hz, 1H, H-4), 5.43 (d, *J* = 9.9 Hz, 1H, H-2), 5.51 (dd, $J_1 = 10.1$ Hz, $J_2 = 9.9$ Hz, 1H, H-3), 7.28–7.46 (m, 10H, Ph-H), 7.52 (t, J = 7.8 Hz, 2H, para-H in Bz), 7.66 (t, J = 7.8 Hz, 4H), 7.81 (d, J = 7.8 Hz, 2H, meta-H in Bz), 7.89 (d, J = 7.8 Hz, 2H, ortho-H in Bz). ¹³C NMR (100 MHz, CDCl₃) δ 170.16, 170.15, 166.18, 165.25, 135.88 (2C), 135.76 (2C), 133.59 (2C), 133.44, 133.26, 129.93 (2C), 129.81 (2C), 129.66, 129.10, 128.73 (2C), 128.58 (2C), 128.35 (2C), 127.71 (2C), 127.66 (2C), 75.86, 73.97, 72.37, 71.21, 68.24, 68.08, 38.53, 26.78 (3C), 20.83, 20.54, 19.20. HRMS (ESI) calcd for C₄₁H₄₄O₁₀SiNa [M + Na]⁺: 747.2601, found: 747.2598. IR (KBr film) v 3474, 3071, 2960, 2932, 2857, 1724, 1601, 1428, 1365, 1269, 1111, 1050, 825, 708, 612, 514 cm^{-1} .

4.10. (1R,2R,3S,4R)-4-Benzoyloxy-5-benzoyloxymethyl-1-(tert-butyldiphenylsilyloxy)-2,3-diacetoxy-cyclohex-5-ene 12. Compound 11 (5.019 g, 6.924 mmol) was dissolved in dichloromethane (50 mL), and the solution was cooled to 0 °C by an ice bath. SOCl₂ (4.120 g, 34.63 mmol) and pyridine (1.643 g, 20.77 mmol) were slowly added. After the addition was finished, the ice bath was removed, and the mixture was heated to reflux (41 °C). The mixture was further stirred under reflux for 6 h. When the reaction was complete (checked by TLC; eluent, EtOAc/hexane = 1:4), dichloromethane was removed by vacuum distillation. Ethyl acetate (100 mL) and water (50 mL) were added, and the biphasic mixture was vigorously stirred for 10 min. Two phases were separated, and the aqueous phase was extracted again with ethyl acetate (60 mL). Organic extracts were combined and successively washed with an aqueous solution of potassium carbonate (15% w/w, 25 mL) and brine (15 mL). The organic solution was dried over anhydrous MgSO4 and then concentrated under vacuum to give the crude product, which was purified by flash chromatography (eluent, EtOAc/hexane = 1:5) to afford compound 12 (4.258 g, 6.024 mmol) as colorless oil in 87% yield. $[\alpha]_{D}^{25} = -94.5$ (c 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.06 (s, 9H, t-Bu), 1.87 (s, 3H,

CH₃ in Ac), 1.92 (s, 3H, CH₃ in another Ac), 4.64 (dd, $J_1 = 7.9$ Hz, $J_2 = 2.4$ Hz, 1H, H-1), 4.70 (s, 2H, CH₂OBz), 5.35 (dd, $J_1 = 11.0$ Hz, $J_2 = 7.9$ Hz, 1H, H-2), 5.51 (dd, $J_1 = 11.0$ Hz, $J_2 = 8.0$ Hz, 1H, H-3), 5.77 (d, J = 1.9 Hz, 1H, H-6), 6.18 (d, J = 1.9 Hz, 1H, H-4), 7.34–7.45 (m, 10H, Ph-H), 7.49–7.58 (m, 2H, Ph-H), 7.61–7.66 (m, 2H, Ph-H), 7.66–7.72 (m, 2H, Ph-H), 7.87–7.93 (m, 4H, Ph-H). ¹³C NMR (100 MHz, CDCl₃) δ 170.11, 169.80, 165.67, 165.66, 135.93 (2C), 135.79 (2C), 133.43, 133.10, 132.94, 132.87, 130.88, 130.34, 130.11, 130.00, 129.78, 129.69 (2C), 129.51 (2C), 129.02, 128.49 (2C), 128.35 (2C), 127.88 (2C), 127.82 (2C), 73.88, 72.12, 71.67, 70.98, 63.24, 26.76 (3C), 20.73, 20.59, 19.22. HRMS (ESI) calcd for C₄₁H₄₂O₉SiNa [M + Na]⁺: 729.2496, found: 729.2490. IR (neat) ν 3071, 2955, 2933, 2858, 1757, 1727, 1602, 1452, 1428, 1368, 1234, 1109, 823, 707, 610, 503 cm⁻¹.

4.11. (1R,2S,3S,4R)-4-Benzoyloxy-5-benzoyloxymethyl-2,3-diacetoxy-1-hydroxy-cyclohex-5-ene 13. Compound 12 (4.109 g, 5.813 mmol) was dissolved in tetrahydrofuran (35 mL). Bu₄NF (6.840 g, 26.16 mmol) and AcOH (1.571 g, 26.16 mmol) were added. The mixture was then stirred at room temperature for 8 h. When the reaction was complete (checked by TLC; eluent, EtOAc/hexane = 1:4), tetrahydrofuran was removed by vacuum distillation. Ethyl acetate (50 mL) and an aqueous solution of potassium carbonate (10% w/w, 30 mL) were added, and the biphasic mixture was vigorously stirred for 10 min. The two phases were separated, and the aqueous phase was extracted again with ethyl acetate (50 mL). Organic extracts were combined and dried over anhydrous MgSO4 and then concentrated under vacuum to give the crude product that was purified by flash chromatography (eluent, EtOAc/hexane = 1:4) to afford compound 13 (2.505 g, 5.347 mmol) as white crystals in 92% yield; mp 131–132 °C. $[\alpha]_D^{25} = -75.6$ (c 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.96 (s, 3H, CH₃ in Ac), 2.11 (s, 3H, CH₃ in another Ac), 3.04 (br. s, 1H, OH), 4.54 (dd, $J_1 =$ 7.9 Hz, $J_2 = 2.1$ Hz, 1H, H-1), 4.83 (s, 2H, CH₂OBz), 5.22 (dd, $J_1 = 10.2$ Hz, $J_2 = 7.9$ Hz, 1H, H-2), 5.53 (dd, $J_1 = 10.2$ Hz, $J_2 = 7.7$ Hz, 1H, H-3), 6.03 (d, J = 2.1 Hz, 1H, H-6), 6.18 (d, $J_1 = 7.7$ Hz, 1H, H-4), 7.35–7.44 (m, 4H, Ph-H), 7.50– 7.59 (m, 2H, Ph-H), 7.92-8.00 (m, 4H, Ph-H). ¹³C NMR (100 MHz, CDCl₃) δ 171.20, 170.04, 165.94, 165.75, 133.52, 133.21, 131.69, 130.30, 129.82 (2C), 129.70 (2C), 129.41, 128.97, 128.54 (2C), 128.37 (2C), 74.98, 71.64, 71.51, 70.12, 63.47, 20.83, 20.59. HRMS (ESI) calcd for C₂₅H₂₄O₉Na [M + Na⁺: 491.1318, found: 491.1315. IR (KBr film) ν 3424, 3071, 2951, 2926, 2873, 1752, 1724, 1601, 1451, 1377, 1261, 1120, 1068, 959, 712 cm⁻¹.

4.12. (1S,2S,3S,4R)-4-Benzoyloxy-5-benzoyloxymethyl-1,2,3-triacetoxy-cyclohex-5-ene 14. Compound 13 (1.450 g, 3.095 mmol) was dissolved in anhydrous ethyl acetate (15 mL), and the solution was cooled to 0 $^{\circ}$ C in an ice bath. Methanesulfonyl chloride (709.0 mg, 6.190 mmol) and Et₃N (470.0 mg, 4.645 mmol) were added, and the mixture was stirred at 0 °C for 1 h. When the reaction was complete (checked by TLC; eluent, EtOAc/hexane = 1:4), ethyl acetate (30 mL) and an aqueous solution of HCl (1 N, 20 mL) were added. After the biphasic mixture was vigorously stirred for 10 min, two phases were separated, and the aqueous phase was extracted again with ethyl acetate (30 mL). Organic extracts were combined, washed with an aqueous solution of potassium carbonate (10% w/w, 15 mL), and then dried over anhydrous MgSO₄. Removal of ethyl acetate under vacuum gave the crude unstable intermediate compound I-A, which was dissolved in toluene (5 mL). AcOH (1.115 g, 18.57 mmol) and DBU (1.414 g, 9.288 mmol) were added. The mixture was then heated to 85 °C and was stirred at this temperature for 5 h. When the reaction was complete (checked by TLC; eluent, EtOAc/hexane = 1:4), toluene was removed by vacuum distillation. Ethyl acetate (30 mL) and an aqueous solution of HCl (1 N, 15 mL) were added. After the biphasic mixture was vigorously stirred for 5 min, two phases were separated, and the aqueous phase was extracted twice with ethyl acetate $(2 \times$ 30 mL). Organic extracts were combined, washed with an aqueous solution of potassium carbonate (10% w/w, 15 mL), and then dried over anhydrous MgSO4. Evaporation of ethyl acetate under vacuum gave the crude product that was then purified by flash chromatography (eluent, EtOAc/hexane = 1:5) to furnish compound 14 (1.264 g, 2.476 mmol) as colorless viscous oil in 80% yield. $[\alpha]_D^{25} = +10.7$ (c 1.0, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 1.99 (s, 3H, CH₃ in Ac), 2.04 (s, 3H, CH₃ in Ac), 2.15 (s, 3H, CH₃ in Ac), 4.86 (s, 2H, CH₂OBz), 5.26 (dd, J_1 = 7.8 Hz, J_2 = 4.2 Hz, H-2), 5.68– 5.78 (m, 2H, H-6 and H-3), 6.03-6.13 (m, 2H, H-1 and H-4), 7.38-7.48 (m, 4H, Ph-H), 7.52-7.62 (m, 2H, Ph-H), 7.94-8.06 (m, 4H, Ph-H). ¹³C NMR (100 MHz, CDCl₃) δ 170.23, 169.94, 169.77, 165.73, 165.68, 137.62, 133.58, 133.28, 129.89 (2C), 129.71 (2C), 129.34, 128.91, 128.85, 128.55, 128.42 (2C), 123.61, 71.14, 69.51, 68.12, 65.40, 63.29, 20.93, 20.68, 20.62. HRMS (ESI) calcd for $C_{27}H_{26}O_{10}Na [M + Na]^+$: 533.1424, found: 533.1420. IR (neat) ν 3066, 2928, 2853, 1751, 1727, 1602, 1452, 1371, 1242, 1112, 1068, 940, 712 cm^{-1} .

4.13. (1S,2S,3S,4R)-5-Hydroxymethyl-1,2,3,4-tetrahydr-oxy-cyclohex-5-ene [(+)-Streptol] 1. Compound 14 (1.020 g, 1.998 mmol) was dissolved in methanol (25 mL). Aqueous ammonia (25% w/w, 5 mL) was added, and the mixture was then stirred at room temperature for 24 h. The reaction solution was concentrated to dryness under vacumm. Ether (20 mL) and pure water (20 mL) were added, the biphasic mixture was vigorously stirred for 5 min, two phases were separated, and the organic phase was extracted again with pure water (20 mL). Aqueous extracts were combined and concentrated under vacumm to give the crude product that was then purified by chromatography on a column of Duolite-C20 resin (eluent, methanol/water =1:2) to afford pure (+)-streptol 1 (317.0 mg, 1.799 mmol) as colorless viscous oil in 90% yield. $[\alpha]_{D}^{25} = +95.2$ (c 0.5, CH₃OH) {lit.^{6c} $[\alpha]_{D}^{25} =$ +95.6 (c 0.45, CH₃OH)} ¹H NMR (400 MHz, D₂O) δ 3.58 (dd, $J_1 = 10.7$ Hz, $J_2 = 4.2$ Hz, 1H, H-2), 3.71 (dd, $J_1 = 10.7$ Hz, $J_2 = 7.8$ Hz, 1H, H-3), 4.09 (dd, $J_1 = 7.8$ Hz, $J_1 = 1.8$ Hz, 1H, H-4), 4.15 (d, J = 14.2 Hz, 1H, CHHO), 4.24 (d, J = 14.2 Hz, 1H, CHHO), 4.30 (dd, $J_1 = 4.3$ Hz, $J_2 = 4.2$ Hz, 1H, H-1), 5.85 (dd, 1H, J_1 = 4.3 Hz, J_2 = 1.8 Hz, H-6). ¹³C NMR (100 MHz, D_2O) δ 144.34, 124.31, 74.73, 74.43, 72.88, 68.31, 63.46. HRMS (ESI) calcd for $C_7H_{12}O_5Na [M + Na]^+$: 199.0582, found: 199.0580. IR (neat) v 3420, 2923, 1640, 1564, 1411, 1102, 1060, 998, 621 cm⁻¹.

4.14. (1*R*,2*S*,3*S*,4*R*)-5-Hydroxymethyl-1,2,3,4-tetrahydr-oxy-cyclohex-5-ene [(-)-1-*epi*-Streptol] **2.** Compound 13 (1.005 g, 2.145 mmol) was dissolved in methanol (25 mL). Aqueous ammonia (25% w/w, 5 mL) was added, and the mixture was then stirred at room temperature for 24 h. The reaction solution was concentrated to dryness under vacumm. Ether (20 mL) and pure water (20 mL) were added, the biphasic mixture was vigorously stirred for 5 min, two phases were separated, and the organic phase was extracted again with pure water (20 mL). Aqueous extracts were combined and concentrated under vacumm to give the crude product that was then purified by chromatography on a column of Duolite-C20 resin (eluent, methanol/water =1:2) to afford pure (-)-1-*epi*-streptol **2** (340.5 mg, 1.933 mmol) as a colorless viscous oil in 90% yield. $[\alpha]_D^{25} = -33.2$ (*c* 1.0, CH₃OH) {lit.⁶ⁱ $[\alpha]_D^{22} = -32.5$ (*c* 0.22, CH₃OH)}. ¹H NMR (400 MHz, D₂O) δ 3.36 (dd, $J_1 = 9.5$ Hz, $J_2 = 4.6$ Hz, 1H, H-2), 3.42 (dd, $J_1 = 9.5$ Hz, $J_2 = 7.8$ Hz, 1H, H-3), 3.99 (d, J = 13.5 Hz, 1H, CHHO), 4.10 (m, 3H, H-1, H-4 and CHHO), 5.51 (dd, $J_1 = 1.8$ Hz, $J_2 = 1.6$ Hz, 1H, H-6). ¹³C NMR (100 MHz, D₂O) δ 138.30, 124.91, 75.49, 75.10, 71.85, 71.33, 60.96. HRMS (ESI) calcd for $C_7H_{12}O_5$ Na [M + Na]⁺: 199.0582, found: 199.0582. IR (neat) ν 3360, 2975, 2897, 1658, 1564, 1422, 1091, 1049, 882, 651 cm⁻¹.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsomega.1c02502.

¹H and ¹³C NMR spectra of compounds 1, 2, and 4–14 (PDF)

AUTHOR INFORMATION

Corresponding Authors

- Shi-Ling Liu Zhejiang Arthur Pharmaceutical Co. Ltd., Jiaxing, Zhejiang 314000, P. R. China; Email: frank.liu@ arthurgroups.com
- Xiao-Xin Shi Engineering Research Center of Pharmaceutical Process Chemistry of the Ministry of Education, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, P. R. China; orcid.org/0000-0002-2751-5122; Email: xxshi@ecust.edu.cn

Authors

- Xing-Liang Zhu Engineering Research Center of Pharmaceutical Process Chemistry of the Ministry of Education, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, P. R. China
- Yong-Qiang Luo Engineering Research Center of Pharmaceutical Process Chemistry of the Ministry of Education, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, P. R. China
- Lei Wang Engineering Research Center of Pharmaceutical Process Chemistry of the Ministry of Education, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, P. R. China
- Yong-Kang Huang Engineering Research Center of Pharmaceutical Process Chemistry of the Ministry of Education, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, P. R. China
- Yun-Gang He Engineering Research Center of Pharmaceutical Process Chemistry of the Ministry of Education, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, P. R. China
- Wen-Jing Xie Engineering Research Center of Pharmaceutical Process Chemistry of the Ministry of Education, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, P. R. China

Complete contact information is available at: https://pubs.acs.org/10.1021/acsomega.1c02502

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

The authors are grateful to the National Natural Science Foundation of China (Nos. 20972048 and 20172015) for the financial support of this work.

REFERENCES

(1) (a) Shui, F.; Jia, J.; Yang, X.; Zhou, Q.; Jiang, Y.; Chen, X. Synthesis of (+)-epoxydon, (-)-phyllostine, (-)-RKTS 33, and (-)-parasitenone featuring selective sulfonylation and oxirane ring closure of Aldol cyclization products. Eur. J. Org. Chem. 2020, 3981-3988. (b) Osborn, A. R.; Kean, K. M.; Alseud, K. M.; Almabruk, K. H.; Asamiza, S.; Lee, J. A.; Karplus, P. A.; Mahmus, T. Evolution and distribution of C₇-cyclitol synthases in prokaryotes and eukaryotes. ACS Chem. Biol. 2017, 12, 979-988. (c) Mondal, S.; Sureshan, K. M. Carbasugar synthesis via vinylogous ketal: total syntheses of (+)-MK7607, (-)-MK7607, (-)-Gabosine A, (-)-Epoxydine B, (-)-Epoxydine C, epi-(+)-Gabosine E and epi-(+)-MK7607. J. Org. Chem. 2016, 81, 11635-11645. (d) Trapero, A.; Egido-Gabas, M.; Bujons, J.; Llebaria, A. Synthesis and evaluation of hydroxymethylaminocyclitols as glycosidase inhibitors. J. Org. Chem. 2015, 80, 3512-3529. (e) Vidyasagar, A.; Sureshan, K. M. Total syntheses of five uvacalols: structural validation of uvacalol A, uvacalol B and uvacalol C and disproval of the structures of uvacalol E and uvacalol G. Org. Biomol. Chem. 2015, 13, 3900-3910. (f) Usami, Y.; Mizuki, K. Stereostructure reassignment and determination of the absolute configuration of pericosine D_o by a synthetic approach. J. Nat. Prod. 2011, 74, 877-881. (g) Choi, W. S.; Xiumei, W. X.; Choeng, Y. H.; Mahmud, T.; Jeong, B. C.; Lee, S. H.; Chang, Y. K.; Kim, C-J.; Soon-Kwang Hong, S.-K. Genetic organization of the putative salbostatin biosynthetic gene cluster including the 2-epi-5-epi-valiolone synthase gene in Streptomyces albus ATCC 21838. Appl. Microbiol. Biotechnol. 2008, 80, 637-645. (h) Delgado, A. Recent advances in the chemistry of aminocyclitols. Eur. J. Org. Chem. 2008, 3893-3906. (i) Arjona, O.; Gómez, A. M.; López, J. C.; Plumet, J. Synthesis and conformational and biological aspects of carbasugars. Chem. Rev. 2007, 107, 1919-2036. (j) Mahmud, T. The C_7N aminocyclitol family of natural products. Nat. Prod. Rep. 2003, 20, 137-166. (k) Naganawa, H.; Hashizume, H.; Kubota, Y.; Sawa, R.; Takahashi, Y.; et al. Biosynthesis of the cyclitol moiety of pyralomicin 1a in Nonomuraea spiralis MI178-34F18. J. Antibiot. 2002, 55, 578-584. (1) Marco-Contelles, J. Cyclohexane epoxides -chemistry and biochemistry of (+)-cyclophellitol. Eur. J. Org. Chem. 2001, 1607-1618.

(2) (a) Isogai, A.; Sakuda, S.; Nakayama, J.; Watanabe, S.; Suzuki, A. Isolation and structural elucidation of a new cyclitol derivative, streptol, as a plant growth regulator. *Agric. Biol. Chem.* **1987**, *51*, 2277–2279. (b) Kizuka, M.; Enokita, R.; Shibata, K.; Okamoto, Y.; Inoue, Y.; Okazaki, T. Studies on actinomycetes isolated from plant leaves. New plant growth inhibitors A-79197-2 and -3 from *Dacthylosporangium aurantiacum* SANK 61299. *Actinomycetologica* **2002**, *16*, 14–16. (c) Sedmera, P.; Halada, P.; Pospísil, S. New carbasugars from *Streptomyces lincolnensis. Magn. Reson. Chem.* **2009**, *47*, 519–522.

(3) (a) Hsiao, C.-C.; Sieber, S.; Georgiou, A.; Bailly, A.; Emmanouilidou, D.; Carlier, A.; Eberl, L.; Gademann, K. Synthesis and biological evaluation of the novel growth inhibitor streptol glucoside, isolated from an obligate plant symbiont. *Chem. – Eur. J.* **2019**, *25*, 1722–1726. (b) Iwu, M.; Okunji, C.; Tchimene, M.; Anele, N.; Chah, K.; Osonwa, U.; Akpa, P. A.; Onunkwo, G. C. Stability of cough linctus (streptol) formulated from named medicinal plant extracts. *Chem. Pharm. Bull.* **2009**, *57*, 229–232.

(4) Shing, T. K. M.; Wu, H. T.; Kwok, H. F.; Lau, C. B. S. Synthesis of chiral hydroxylated enones as potential anti-tumor agents. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 7562–7565.

(5) (a) Zhang, C.-S.; Stratmann, A.; Block, O.; Bruckner, R.; Podeschwa, M.; Altenbach, H.-J.; Wehmeier, U. F.; Piepersberg, W. Biosynthesis of the C₇-cyclitol moiety of acarbose in *Actinoplanes* species SE50/110. 7-O-Phosphorylation of the initial cyclitol precursor leads to proposal of a new biosynthetic pathway. *J. Biol. Chem.* **2002**, 277, 22853–22862. (b) Wehmeier, U. F. The biosynthesis and metabolism of acarbose in *Actinoplanes sp.* SE 50/110: A progress report. *Biocatal. Biotransform.* **2003**, 21, 279–284. (c) Zhang, C.-S.; Podeschwa, M.; Block, O.; Altenbach, H.-J.; Piepersberg, W.; Wehmeier, U. F. Identification of a 1-*epi*-valienol 7-kinase activity in the producer of acarbose, *Actinoplanes* sp. SE50/110. *FEBS Lett.* **2003**, 540, 53–57.

(6) (a) Krishna, P. R.; Kadiyala, R. R. Sequential Baylis-Hillman/ RCM protocol for the stereoselective synthesis of (+)-MK7607 and (+)-streptol. Tetrahedron Lett. 2010, 51, 2586-2588. (b) Shing, T. M.; Cheng, H. M. The structure and stereochemistry of gabosine K: syntheses of 7-O-acetyl-streptol and 7-O-acetyl-1-epi-streptol. Synlett 2010, 142-144. (c) Shing, T. K. M.; Chen, Y.; Ng, W-L. Carbocyclization of _D-glucose: syntheses of gabosine I and streptol. Tetrahedron 2011, 67, 6001-6005. (d) Yuan, P.; Liu, X. L.; Yang, X.; Zhang, Y. L.; Chen, X. C. Total syntheses of (+)-gabosine P, (+)-gabosine Q, (+)-gabosine E, (-)-gabosine G, (-)-gabosine I, (-)-gabosine K, (+)-streptol, and (-)-uvamalol A by a diversityoriented approach featuring tunable deprotection manipulation. J. Org. Chem. 2017, 82, 3692-3701. (e) Shing, T. K. M.; Chen, Y.; Ng, W. L. Short and efficient syntheses of gabosine I, streptol, 7-Oacetylstreptol, 1-epi-streptol, gabosine K, and carba- α -D-glucose from δ -D-gluconolactone. Synlett 2011, 1318–1320. (f) Mehta, G.; Pujar, S. R.; Ramesh, S. S.; Islam, K. Enantioselective total synthesis of polyoxygenated cyclohexanoids: (+)-streptol, ent-RKTS-33 and putative (+)-parasitenone. Identity of parasitenone with (+)-epoxydon. Tetrahedron Lett. 2005, 46, 3373-3376. (g) Leermann, T.; Block, B.; Podeschwa, M. A. L.; Pfuller, U.; Altenbach, H.-J. De novo synthesis and lectin binding studies of unsaturated carba-pyranoses. Org. Biomol. Chem. 2010, 8, 3965-3974. (h) Moschitto, M. J.; Vaccarello, D. N.; Lewis, C. A. Regiodivergent addition of phenols to allylic oxides: control of 1,2- and 1,4-additions for cyclitol synthesis. Angew. Chem., Int. Ed. 2015, 54, 2142-2145. (i) Mahmud, T.; Tornus, I.; Egelkrout, E.; Wolf, E.; Uy, C.; Floss, H. G.; Lee, S. Biosynthetic studies on the α -glucosidase inhibitor acarbose in Actinoplanes sp.: 2-epi-5-epi-valiolone is the direct precursor of the valienamine moiety. J. Am. Chem. Soc. 1999, 121, 6973-6983.

(7) (a) Candeias, N. R.; Assoah, B.; Simeonov, S. P. Production and synthetic modifications of shikimic acid. Chem. Rev. 2018, 118, 10458-10550. (b) Borah, J. C. Shikimic acid: a highly prospective molecule in pharmaceutical industry. Curr. Sci. 2015, 109, No. 1802. (c) Rawat, G.; Tripathi, P.; Saxena, R. K. Expanding horizons of shikimic acid. Recent progresses in production and its endless frontiers in application and market trends. Appl. Microbiol. Biotechnol. 2013, 97, 4277-4287. (d) Estevez, A. M.; Estevez, R. J. A short overview on the medicinal chemistry of (-)-shikimic acid. Mini-Rev. Med. Chem. 2012, 12, 1443-1454. (e) Ghosh, S.; Chisti, Y.; Banerjee, U. C. Production of shikimic acid. Biotechnol. Adv. 2012, 30, 1425-1431. (f) Bochkov, D. V.; Sysolyatin, S. V.; Kalashnikov, A. I.; Surmacheva, I. A. Shikimic acid: review of its analytical, isolation, and purification techniques from plant and microbial sources. J. Chem. Biol. 2012, 5, 5-17. (g) Krämer, M.; Bongaerts, J.; Bovenberg, R.; Kremer, S.; Muller, U.; Orf, S.; Wubbolts, M.; Raeven, L. Metabolic engineering for microbial production of shikimic acid. Metab. Eng. 2003, 5, 277-283.

(8) (a) Zhu, X.-L.; Wang, L.; Luo, Y.-Q.; He, Y.-G.; Li, F.-L.; Sun, M.-M.; Liu, S.-L.; Shi, X.-X. Efficient and highly stereoselective syntheses of (+)-*proto*-quercitol and (-)-*gala*-quercitol starting from the naturally abundant (-)-shikimic acid. ACS Omega 2020, 5, 1813–1821. (b) Li, F.-L.; Yu, J.-P.; Ding, W.; Sun, M.-M.; He, Y.-G.; Zhu, X.-L.; Liu, S.-L.; Shi, X.-X. Novel stereoselective syntheses of N-octyl-bvalienamine (NOV) and N-octyl-4-*epi*-bvalienamine (NOEV) from (-)-shikimic acid. RSC Adv. 2019, 9, 42077–42084. (c) Li, F.; Ding, W.; Quan, N.; Wu, J.; He, Y.; Zhu, X.; Shi, X.; Zhao, J. Improved stereoselective syntheses of (+)-valiolamine and (+)-valienamine starting from (-)-shikimic acid. Chin. J. Chem. 2017, 35, 457–464.

(d) Ding, W.; Yu, J.-P.; Shi, X.-X.; Nie, L.-D.; Quan, N.; Li, F.-L. Stereoselective synthesis of (+)-valienamine starting from the naturally abundant (-)-shikimic acid. Tetrahedron Asymmetry 2015, 26, 1037-1042. (e) Quan, N.; Nie, L.-D.; Zhu, R.-H.; Shi, X.-X.; Ding, W.; Lu, X. Total syntheses of (+)-valiolamine and (-)-1-epivaliolamine from naturally abundant (-)-shikimic acid. Eur. J. Org. Chem. 2013, 6389-6396. (f) Nie, L.-D.; Wang, F.-F.; Ding, W.; Shi, X.-X.; Lu, X. A novel azide-free asymmetric synthesis of oseltamivir phosphate (Tamiflu) starting from Roche's epoxide. Tetrahedron Asymmetry 2013, 24, 638-642. (g) Nie, L.-D.; Ding, W.; Shi, X.-X.; Quan, N.; Lu, X. A novel and high-yielding asymmetric synthesis of oseltamivir phosphate (Tamiflu) starting from (-)-shikimic acid. Tetrahedron Asymmetry 2012, 23, 742-747. (h) Nie, L.-D.; Shi, X.-X.; Quan, N.; Wang, F.-F.; Lu, X. Novel asymmetric synthesis of oseltamivir phosphate (Tamiflu) from (-)-shikimic acid via cyclic sulfite intermediates. Tetrahedron Asymmetry 2011, 22, 1692-1699. (i) Nie, L.-D.; Shi, X.-X.; Ko, K. H.; Lu, W.-D. A short and practical synthesis of oseltamivir phosphate (Tamiflu) from (-)-shikimic acid. J. Org. Chem. 2009, 74, 3970-3973. (j) Nie, L.-D.; Shi, X.-X. A novel asymmetric synthesis of oseltamivir phosphate (Tamiflu) from (-)-shikimic acid. Tetrahedron Asymmetry 2009, 20, 124-129. (k) Kim, H.-K.; Park, K.-J. J. A new efficient synthesis of oseltamivir phosphate (Tamiflu) from (-)-shikimic acid. Tetrahedron Lett. 2012, 53, 1561-1563. (1) Karpf, M.; Trussardi, R. Efficient Access to Oseltamivir phosphate (Tamiflu) via the O-trimesylate of shikimic acid ethyl ester. Angew. Chem., Int. Ed. 2009, 48, 5760-5762. (m) Harrington, P. J.; Brown, J. D.; Foderaro, T.; Hughes, R. C. Research and development of a second-generation process for oseltamivir phosphate, prodrug for a neuraminidase inhibitor. Org. Process Res. Dev. 2004, 8, 86-91. (n) Karpf, M.; Trussardi, R. New, azide-free transformation of epoxides into 1,2-diamino compounds: synthesis of the anti-influenza neuraminidase inhibitor oseltamivir phosphate (Tamiflu). J. Org. Chem. 2001, 66, 2044-2051. (o) Kim, C. U.; Lew, W.; Williams, M. A.; Liu, H.; Zhang, L.; Swaminathan, S.; Bischofberger, N.; Chen, M. S.; Mendel, D. B.; Tai, C. Y.; Laver, W. G.; Stevens, R. C. Influenza neuraminidase inhibitors possessing a novel hydrophobic interaction in the enzyme active site: design, synthesis, and structural analysis of carbocyclic sialic acid analogues with potent anti-influenza activity. J. Am. Chem. Soc. 1997, 119, 681-690.

(9) (a) Quan, N.; Nie, L.; Shi, X.; Zhu, R.; Lu, X. Novel and efficient syntheses of four useful shikimate-derived epoxy chiral building blocks via cyclic sulfite intermediates. *Chin. J. Chem.* **2012**, *30*, 2759–2766. (b) Zhang, W.; Zhu, X.-L.; Ding, W.; Shi, X.-X. A novel stereoselective synthesis of (–)-quinic acid starting from the naturally abundant (–)-shikimic acid. *Tetrahedron Asymmetry* **2015**, *26*, 1375–1381. (c) Zhou, S.; Zhu, R.; Hu, J.; Zhang, L.; Lu, Q.; Yu, X. Economical process for preparation of the 19-nor A ring of paricalcitol from (–)-shikimic acid. *Org. Process Res. Dev.* **2019**, *23*, 1887–1891. (d) Gonda, J.; Psotka, M.; Martinkova, M.; Fazekasova, S.; Mitrikova, T.; Vilkova, M. A short synthesis of protected 3-deoxy-D-arabino-2-heptulosonates (DAH) from shikimic acid based on silyl group migration. *Tetrahedron Lett.* **2018**, *59*, 4620–4621.

(10) (a) Qadar, M. M.; Kumar, N. S.; Jayasinghe, L.; Fujimoto, Y. Shikimic acid production by *Fusarium decencellulare*, an endophytic fungus isolated from *Flacourtia inermis* fruits. *J. Biol. Act. Prod. Nat.* **2018**, *8*, 43–50. (b) Zhang, X.; Meng, X.; Wu, J.; Huang, L.; Chen, S. Global ecological regionalization of 15 *Illicium* species: nature sources of shikimic acid. *Chin. Med.* **2018**, *13*, No. 31. (c) Xu, S.; Hossain, M. M.; Lau, B. B. Y.; To, T. Q.; Rawal, A.; Aldous, L. Total quantification and extraction of shikimic acid from star anise (*llicium verum*) using solid-state NMR and cellulose-dissolving aqueous hydroxide solutions. *Sustainable Chem. Pharm.* **2017**, *5*, 115–121.

(11) (a) Kuriya, Y.; Araki, M. Dynamic flux balance analysis to evaluate the strain production performance on shikimic acid production in *Escherichia coli. Metabolites* 2020, 10, No. 198.
(b) Sato, N.; Kishida, M.; Nakano, M.; Hirata, Y.; Tanaka, T. Metabolic engineering of shikimic acid-producing *Corynebacterium glutamicum* from glucose and cellobiose retaining its phosphotransfer-

ase system function and pyruvate kinase activities. *Front. Bioeng. Biotechnol.* **2020**, *8*, No. 569406. (c) Lee, M.-Y.; Hung, W.-P.; Tsai, S.-H. Improvement of shikimic acid production in *Escherichia coli* with growth phase-dependent regulation in the biosynthetic pathway from glycerol. *World J. Microbiol. Biotechnol.* **2017**, *33*, No. 25.

(12) (a) Jiang, S. D.; Singh, G. Chemical synthesis of shikimic acid and its analogues. *Tetrahedron* **1998**, *54*, 4697–4753. (b) Bohm, B. A. Shikimic acid (3,4,5-trihydroxy-1-cyclohexene-1-carboxylic acid). *Chem. Rev.* **1965**, *65*, 435–466.

(13) (a) Kshirsagr, P. R.; Pai, S. R.; Hegde, H. V. Comparison of shikimic acid content: a precursor for drug against H5N1 from various plant species of western Ghats, India. *Natl. Acad. Sci. Lett.* **2018**, *41*, 399–402. (b) Aniya; Nomura, Y.; Fuerdeng; Appiah, K. S.; Fujii, Y. Evaluation of allelopathic activity of Chinese medicinal plants and identification of shikimic acid as an allelochemical from *Illicium verum* Hook. f. *Plants* **2020**, *9*, No. 684. (c) Avula, B.; Wang, Y.-H.; Smillie, T. J.; Khan, I. A. Determination of shikimic acid in fruits of illicium species and various other plant samples by LC–UV and LC–ESI–MS. *Chromatographia* **2009**, *69*, 307–314.

(14) (a) Shi, Y.; Chen, G.; Chen, K.; Chen, X.; Hong, Q.; Kan, J. Assessment of fresh star anise (*Illicium verum* Hook.f.) drying methods for influencing drying characteristics, color, flavor, volatile oil and shikimic acid. *Food Chem.* **2021**, *342*, No. 128359. (b) Niu, F.-X.; He, X.; Huang, Y.-B.; Liu, J.-Z. Biosensor-guided atmospheric and room-temperature plasma mutagenesis and shuffling for high-level production of shikimic acid from sucrose in *Escherichia coli. J. Agric. Food Chem.* **2020**, *68*, 11765–11773.

(15) (a) Just, J.; Deans, B. J.; Olivier, W. J.; Paull, B.; Bissember, A. C.; Smith, J. A. New method for the rapid extraction of natural products: efficient isolation of shikimic acid from star anise. *Org. Lett.* **2015**, *17*, 2428–2430. (b) Cai, M.; Luo, Y.; Chen, J.; Liang, H.; Sun, P. Optimization and comparison of ultrasound-assisted extraction and microwave-assisted extraction of shikimic acid from Chinese star anise. *Sep. Purif. Technol.* **2014**, *133*, 375–379. (c) Zirbs, R.; Strassl, K.; Gaertner, P.; Schroder, C.; Bica, K. Exploring ionic liquid–biomass interactions: towards the improved isolation of shikimic acid from star anise pods. *RSC Adv.* **2013**, *3*, 26010–26016. (d) Ohira, H.; Torii, N.; Aida, T. M.; Watanabe, M.; Smith, R. L., Jr. Rapid separation of shikimic acid from Chinese star anise (*Illicium verum* Hook. f.) with hot water extraction. *Sep. Purif. Technol.* **2009**, *69*, 102–108. (e) Payne, R.; Edmonds, M. Isolation of shikimic acid from star aniseed. J. Chem. Educ. **2005**, *82*, 599–600.

(16) Liu, A.; Liu, Z. Z.; Zou, Z. M.; Chen, S. Z.; Xu, L. Z.; Yang, S. L. Synthesis of (+)-zeylenone from shikimic acid. *Tetrahedron* **2004**, 60, 3689–3694.

(17) (a) Shing, T. K. M.; Tai, V. W.-F.; Tam, E. K. W. Practical and rapid vicinal hydroxylation of alkenes by catalytic ruthenium tetraoxide. *Angew. Chem., Int. Ed.* **1994**, *33*, 2312–2313. (b) Plietker, B.; Niggemann, M. An improved protocol for the RuO₄-catalyzed dihydroxylation of olefins. *Org. Lett.* **2003**, *5*, 3353–3356.

(18) Shi, X.-X.; Shen, C.-L.; Yao, J.-Z.; Nie, L.-D.; Quan, N. Inversion of secondary chiral alcohols in toluene with the tunable complex of $R_3NR'COOH$. *Tetrahedron Asymmetry* **2010**, *21*, 277–284.

(19) (a) Dalla, V.; Catteau, J. P.; Pale, P. Mechanistic rationale for the NaBH₄ reduction of α -keto esters. *Tetrahedron Lett.* **1999**, 40, 5193–5196. (b) Dalla, V.; Cotelle, P.; Catteau, J. P. Chemocontrolled reduction of aromatic α -ketoesters by NaBH₄: selective Synthesis of α -hydroxy esters or 1,2-diols. *Tetrahedron Lett.* **1997**, 38, 1577–1580.