

[REVIEW ARTICLE]

Recent Advances in Research Regarding Autoantibodies in Connective Tissue Diseases and Related Disorders

Kosaku Murakami and Tsuneyo Mimori

Abstract:

Connective tissue diseases (CTDs), also known as systemic autoimmune diseases, involve a variety of autoantibodies against cellular components. An important factor regarding these autoantibodies is that each antibody is exclusively related to a certain clinical feature of the disease type, which may prove useful in clinical practice. Thus far, more than 100 types of autoantibodies have been found in CTDs, and most of their target antigens have been identified. Many of these autoantigens are enzymes or regulators involved in important cellular functions, such as gene replication, transcription, repair/recombination, RNA processing, and protein synthesis, as well as proteins that form complexes with RNA and DNA. This article reviews the autoantibodies for each CTD, along with an assessment of their clinical significance, and provides suggestions regarding their utilization for clinical practice.

Key words: autoantibody, connective tissue disease

(Intern Med 58: 5-14, 2019)

(DOI: 10.2169/internalmedicine.1423-18)

Introduction

Autoantibodies have been recognized as diagnostic markers for a variety of connective tissue diseases (CTDs), including rheumatoid arthritis (RA), systemic lupus erythematosus (SLE), polymyositis (PM)/dermatomyositis (DM), systemic sclerosis (SSc), and other systemic autoimmune diseases. In recent decades, it has been elucidated that a certain number of autoantibodies play pathogenic roles in the manifestation of the associated disease. In addition, some autoantigens are thought to be associated with immunological functions affecting the onset or expansion of the diseases.

This review article discusses recent advances in research regarding autoantibodies and autoantigens in CTDs, including related disorders.

Rheumatoid Arthritis

RA was one of the first autoimmune diseases in which autoantibodies were found to play important roles, not only as clinical treatment entities but also as fundamental parts of the disease pathogenesis. Rheumatoid factor (RF) comprises

IgM class antibodies against the Fc portion of autologous IgG. As early as 1948, Rose et al. found that sheep erythrocytes were agglutinated by the sera of RA patients (1); this paper is now recognized as the first report of the detection of RF. Although RF is not a specific marker for RA, it is used as a diagnostic marker in the current classification criteria (2, 3). In addition, a variety of autoantibodies have been described as disease-specific markers of RA.

Autoantigens as specific proteins recognized with autoantibodies

Calpastatin is an endogenous inhibitor for calcium-dependent neutral proteinase (calpain) and has been reported as an autoantigen in RA (4, 5). Synthetic calpain inhibition or calpastatin expression was reported to ameliorate experimental arthritis (6, 7).

Follistatin-related protein [FRP; or follistatin-like 1 (FSTL1)], a secreted glycoprotein, has also been detected as an autoantigen in RA (8). However, reports regarding the effects of FRP on the pathogenesis of RA have been controversial (9-11); at present, FRP is recognized as an inflammatory or osteoclastogenic cytokine, especially via CD14-toll-like receptor (TLR) 4 signal transduction (12-16). Im-

munoglobulin heavy chain-binding protein (BiP) is a heat shock protein that was also detected as an autoantigen in RA (17). BiP was highly expressed in the synovial cells of RA patients and is recognized as a major target of B and T lymphocytes (18). At present, self-reactivity to BiP in RA is associated with immune responses to mycobacterial heat shock protein 70 (19).

Myelin basic protein was reported as both a genetic risk factor and a specific autoantigen in RA (20), suggesting that the generation of autoantibodies is associated with genetic pathogenicity.

In contrast to the autoantigens described above, all of which have some relationships with immune responses, 60S ribosomal protein L23a (RPL23A) was detected as an initial responder of T cell receptor (TCR) signaling specific to RA through an analysis of the SKG arthritogenic mouse model (21). This has helped improve our understanding of the underlying drivers of autoimmunity.

Autoantigens with post-translational modification

Anti-citrullinated protein antibodies (ACPAs) are currently recognized as the most practically useful autoantibodies in RA treatment, largely because of their specificity in differential diagnoses. According to a meta-analysis, the pooled sensitivity and specificity for anti-cyclic citrullinated protein (CCP) antibody were 67% and 95%, respectively (22). ACPA is now recognized as an important item in the diagnosis of RA based on the American College of Rheumatology (ACR) criteria (2, 3) as well as a predictive factor for joint destruction (23). Since citrullination is a non-specific process of protein modification, multiple proteins are detected as targets of ACPA in a single patient (24, 25). In addition, there are a number of reports describing associations between ACPA and genetic risk factors, especially HLA-DRB1 shared epitope (SE). In Japanese patients, there is a strong association between amino acid position 74 of HLA-DRB1 (mainly conferred by alanine residue) and ACPA levels in seropositive RA (26). In contrast, ACPA-negative RA in Japanese patients seems to have a distinct genetic character, including specific HLA-DRB1 alleles other than SE (27-30).

Carbamylated proteins (CarP) are also recognized as autoantigens for RA, although there are some conflicting reports (31). As with ACPA, patients with anti-CarP antibodies tend to experience progressive joint damage (32). Specific carbamylated protein autoantigens remain unknown, but albumin has been identified as a target antigen of anti-CarP antibodies (33).

Another post-transcriptional change involves the misfolding of proteins to become autoantigens by a unique mechanism. A specific HLA class II molecule, associated with susceptibility to RA, complexes with intact IgG heavy chain following transportation to the cell surface, which leads to the production of RF (autoantibodies to the Fc portion of IgG) (34). In addition to RF, β 2-Glycoprotein I (autoantigen of anti-phospholipid antibodies) and myeloperoxidase [auto-

antigen of anti-neutrophil cytoplasmic antibody (ANCA)] are associated with this type of antigen presentation (35, 36).

Autoantibodies associated with the clinical response to biologic disease-modifying anti-rheumatic drugs (bDMARDs)

When bDMARDs became available for RA, several patients were found to exhibit an insufficient reaction despite a good initial response (known as secondary failure). In particular, patients treated by anti-tumor necrosis factor (TNF) reagents tend to experience this problem. These kinds of agents are not self-antigens; therefore, their immunogenicity is caused by anti-drug antibodies (ADAs) (37). However, not only ADAs arise against non-self antigens, antinuclear antibody (ANA) and anti-dsDNA antibody are often additionally detected in RA patients who have received anti-TNF bDMARDs (38). The reason for the lupus-like autoantibody production in association with bDMARDs remains unknown but may be associated with type I interferon production by anti-TNF therapy (39).

Systemic Lupus Erythematosus

SLE is a representative systemic autoimmune disease that affects almost all organs, including the brain, kidney, heart, and lung. Affected patients show anti-DNA antibody as well as a variety of other autoantibodies, such as anti-Sm, anti-ribosomal P, and anti-phospholipid antibodies; indeed, some of these are included in the representative classification criteria for SLE [i.e. the ACR criteria or Systemic Lupus International Collaborating Clinics (SLICC) classification criteria] (40-42).

Anti-double strand DNA (dsDNA) antibody is commonly detected in SLE patients. While its detection depends on the assay method used, such as the Farr assay (radiolabeled DNA antigen) or enzyme-linked immunosorbent assay (ELISA), 70-90% of SLE patients are anti-dsDNA-positive (43). These antibodies are examined as a measure of the disease activity in clinical practice, but the nature of the antigen-antibody interaction remains unclear, as these antibodies can bind a spectrum of DNA and non-DNA structures, like nucleosomes (44, 45).

Neuropsychiatric lupus (NPSLE) is a disease of interest in which autoantibodies have been found to be frequently associated with the disease pathogenesis. Anti-N-methyl-D-aspartate receptor subunit 2 (NR2) antibody, which shows cross-reaction with the anti-DNA antibody, is associated with cognitive dysfunction and an acute state of confusion in SLE (46-49). Anti-ribosomal P antibody is also associated with NPSLE (50, 51). Massardo et al. reported that anti-NR2 and anti-ribosomal P antibodies both independently contribute to cognitive dysfunction (52). There are several reports in which these two autoantibodies directly function to cause neuronal damage (53-56).

In addition, we reported that the anti-U1 ribonucleopro-

tein (RNP) antibody, especially antibodies to anti-70K protein of the U1-RNP molecule in cerebrospinal fluid, is a useful diagnostic marker for NPSLE (57). Serum anti-U1-70k antibody is associated with psychiatric syndromes in SLE but not with whole central nervous system (CNS) syndromes or neurologic syndromes. Anti-U1-70k antibody might be involved in the pathological mechanisms underlying the psychiatric syndromes of SLE (58).

While there have been few reports regarding specific autoantigens associated with lupus nephritis aside from historical studies of anti-DNA antibody, kidney-specific macrophages are thought to scavenge circulating immune complexes into the interstitium via trans-endothelial transport, thus triggering an Fc γ RIV-dependent inflammatory response involving the recruitment of monocytes and neutrophils (59). This suggests that the mechanism underlying the disease pathogenesis, mainly type III hypersensitivity reactions, differs depending on the organ affected in each case of SLE.

Polymyositis and Dermatomyositis

Polymyositis/Dermatomyositis (PM/DM) are systemic autoimmune disorders that involve skeletal muscle, skin, and internal organs, including the lung and heart. In recent decades, a number of novel autoantibodies have been detected in PM/DM patients, most of which are recognized as myositis-specific autoantibodies (MSAs) or myositis-associated autoantibodies (MAAs). Notably, each MSA/MAA is closely associated with characteristic symptoms, clinical subsets, complications, and prognoses. Therefore, PM/DM are recognized as diseases in which the detection of autoantibodies provides critical clinical implications (60).

Anti-aminoacyl-tRNA synthetase (ARS) antibodies, also known as anti-synthetase antibodies are the most frequent MSAs and are closely associated with interstitial lung disease (ILD) (60, 61). Among the 20 synthetases that correspond to the fundamental 20 amino acids, 8 specific ARSs have been recognized as autoantigens of MSAs: anti-Jo-1 (histidyl-tRNA synthetase) (62, 63), anti-PL-7 (threonyl) (64), anti-PL-12 (alanyl) (65), anti-EJ (glycyl) (66), anti-OJ (isoleucyl) (67), anti-KS (asparaginyl) (68), anti-Zo (phenylalanyl) (69), and anti-Ha (tyrosyl) antibodies (70). These antibodies can predict the clinical course of ILD, which responds well to initial treatment with high-dose glucocorticoids but frequently shows recurrence (71). Among these antibodies, Fujisawa et al. reported that anti-PL-7 antibody in particular predicts the long-term deterioration of ILD (72). Although each anti-ARS antibody has been detected by immunoprecipitation, an ELISA system was recently developed using a mixture of five major recombinant ARS antigens (Jo-1, PL-7, PL-12, EJ, and KS, but not OJ) and has been readily available in clinical practice in Japan since 2014 (73).

Anti-melanoma differentiation-associated gene 5 (MDA5) antibody is widely found in Asian patients with clinically amyopathic dermatomyositis (CADM) with typical skin

manifestations of DM and life-threatening ILD (74-77). Importantly, the frequency of anti-MDA5 antibody in American and European cohorts is lower than in Asian cohorts (78-80). ILD with anti-MDA5 antibody shows a characteristic disease course as well as distinct manifestations in blood tests and radiological findings. Serum levels of ferritin, interleukin (IL)-18, IFN- α , and IFN- β are increased in those patients compared with other ILD patients, although these cytokines are also increased in anti-ARS-positive patients (81-84). High-resolution computed tomography (HRCT) of ILD with anti-MDA5 tends to show intralobular reticular opacities and the predominance of consolidation or ground-glass attenuation (GGA) in the lower lung fields, as well as random GGA patterns (85). In Japan, an ELISA for anti-MDA5 has been established since 2016 and is recognized as a useful assay for detecting anti-MDA5 for the earlier initiation of intensive immunosuppression therapies (60, 86). We recently found that some anti-MDA5 antibody-positive patients had a novel antibody to splicing factor proline/glutamine-rich protein (SFPQ), which is known to play a role in innate immune responses (87).

Anti-Mi-2 autoantibody is detected in 10-20% of DM cases but is not found in PM cases (88). A major antigen recognized by anti-Mi-2 constitutes a component of the nucleosome remodeling-deacetylase (NuRD) complex, which is involved in transcriptional regulation (89). Anti-Mi-2-positive DM has a strong association with HLA-DR7 and shows a good prognosis (90).

Anti-transcriptional intermediary factor-1 γ (TIF-1 γ) antibody is commonly identified in 20-30% of DM cases. A number of reports have described a close association between anti-TIF-1 γ and internal malignancies (91, 92). Furthermore, cancer-associated myopathies (CAMs) have been revealed to be clinico-histopathologically heterogeneous disease entities, with anti-TIF-1 γ -positive CAMs showing a close temporal association with cancer detection, while CAMs with necrotizing autoimmune myopathy (NAM) comprise a subset of anti-TIF-1 γ -negative CAMs (93).

ELISAs for the detection of anti-TIF-1 γ and anti-Mi-2 antibodies were recently developed in Japan and have been available for clinical practice since 2016 (94). It should be noted that the ELISA tests of these two antibodies occasionally cross-react with each other.

Anti-signal recognition particle (SRP) autoantibody is recognized as an MSA typically found in 4-6% of patients with idiopathic inflammatory myopathies (IIMs) (95). Myopathy associated with anti-SRP has been well described in previous studies; it is characterized by severe and rapidly progressive symmetric proximal muscle weakness, extremely elevated levels of serum creatine kinase, and necrotizing myopathy with little evidence of inflammatory infiltrates on muscle biopsies (96-100). For the differential diagnosis, several autoantibodies have been reported as specific for inflammatory myopathies other than PM/DM. Anti-3-hydroxy-3-methylglutaryl-coenzyme A Reductase (HMGCR) antibody is recognized as a major target of antibodies in ne-

crotizing myopathy and is increased by statin use (101).

Inclusion body myositis (IBM) is characterized as a degenerative myopathy resistant to any immunosuppressive therapies with unique pathophysiological characteristics on a muscle biopsy analysis (inclusion bodies with an excess of cytochrome oxidase-deficient fibers) (102-104). Autoantibody against cytosolic 5'-nucleotidase 1A (NT5C1A) has recently been identified as a specific diagnostic marker for IBM (105). According to data from various European IBM registries, anti-NT5C1A autoantibodies are detected in 33% of IBM patients (106). Recently, anti-mitochondrial autoantibody (AMA), which was originally recognized as a marker of primary biliary cirrhosis, has been reported as a marker of IIMs (107). Myopathy with anti-AMA antibody exhibits a distinct phenotype, such as a lower degree of limb muscle weakness with frequent paravertebral muscle atrophy, in addition to cardiac muscle involvement (108-110).

Systemic Sclerosis (or Scleroderma)

SSc (or scleroderma) is characterized by cutaneous and visceral fibrosis with vascular abnormalities induced by an unknown mechanism, although most patients have certain autoantibodies in their sera. As in other CTDs, the clinical characteristics and subsets can be differentiated with specific autoantibodies (111-113).

Anti-DNA topoisomerase I (TOPO, initially known as Scl-70) antibody is most frequently detected in patients with diffuse SSc who have a high risk of diffuse cutaneous involvement and multiple organ lesions, such as those suffering from pulmonary fibrosis and cardiac lesions.

Anti-RNA polymerase III (RNAP III) antibody is also associated with diffuse-type SSc and scleroderma renal crisis (114). In a recent report, anti-RNAP III was detected in 11% of SSc patients (115).

Anti-centromere antibody (CENP) is frequently detected in limited cutaneous SSc (lcSSc) patients, who tend to have a better prognosis than other SSc patients with regard to specific autoantibodies (114). However, SSc patients with anti-CENP antibodies have a high risk of pulmonary arterial hypertension (PAH). Recently, subspecificity of anti-centromeric protein A (CENP-A) antibody was found to have a strong association with PAH (116).

Anti-Th/To autoantibody was identified against RNase MRP (Th or 7-2 RNA) and RNase P (To or 8-2 RNA); the antibody recognizes a 40-kDa protein subunit common to both RNAs, known as Th40 (117). Anti-Th/To is a serological abnormality also found in localized scleroderma; the presence of anti-Th/To may be a serological indicator of a mild form of cutaneous involvement (118).

Anti-U3-RNP antibody directed against the 34-kDa protein (named fibrillar) can precipitate U3 RNA-containing particles (119, 120). The presence of anti-U3-RNP is considered to be relatively specific to SSc (121, 122), but anti-U3-RNP antibody has been described in some patients with SLE (123). Anti-U3-RNP-positive patients have more fre-

quent skeletal muscle involvement and PAH than Anti-U3-RNP negative patients, with PAH being the most common cause of death in these patients (124).

Although the prevalence is relatively low, other SSc-specific autoantibodies with potentially pathogenic roles in vascular damage and tissue fibrosis have been found. Antibodies against angiotensin II type 1 receptor [AT(1)R] and endothelin-1 type A receptor [ET(A)R] are detected in SSc patients. In an *in vitro* experiment, AT(1)R and ET(A)R autoantibodies increased the transforming growth factor β (TGF- β) gene expression in endothelial cells; this was able to be blocked with specific receptor antagonists (125). An autoantibody specifically inhibiting M3-muscarinic receptor that mediated enteric cholinergic neurotransmission was found, potentially providing a pathogenic mechanism for the gastrointestinal dysfunction seen in patients with scleroderma (126). An autoantibody against platelet-derived growth factor receptor (PDGF-R) was also found in SSc patients. Although there have been a few negative reports, anti-PDGF-R is thought to agonistically bind to PDGF-R and cause fibrosis via the upregulation of the reactive oxygen species (ROS) function and collagen production (127-134). Anti-U11/U12-RNP antibody, also known as anti-RNA-binding protein-containing 3 (RNPC-3) antibody, was specifically detected in approximately 3% of SSc patients (135). This autoantibody has recently been reported to be associated with an increased risk of cancer at the onset of scleroderma (136).

Mixed CTD (MCTD) and Overlap Syndrome

In contrast to the disease-specific autoantibodies mentioned above, other antibodies have been detected in patients who show characteristics of more than two CTDs; these patients are diagnosed with MCTD or overlapping syndrome. MCTD was initially proposed as a distinct disease in 1972 by Sharp et al. (137). While whether or not it is indeed a distinct disease entity remains controversial (138), the characteristics of MCTD are considered to be a combination of features similar to those of SLE, SSc, and PM. The disease pathogenesis of U1-RNP remains unclear, but the RNA-binding motifs of relevant autoantigens might underlie its susceptibility as a target of common CTDs (139).

Other autoantibodies have been detected in patients who do not exhibit "solo" CTD but instead suffer from double or triple diseases (so-called overlap syndrome). Anti-Ku antibody was first described in PM-SSc overlap cases (140). The Ku antigen, a heterodimer of 70-kDa (p70) and 80-kDa (p80) subunits, is a component of the DNA-dependent protein kinase (DNA-PK), which binds the free ends of double-stranded DNA (dsDNA) during DNA repair and recombination (141-144). Hoa et al. found that, among 2,140 SSc patients, 24 (1.1%) had anti-Ku autoantibody, and 13 (0.6%) exhibited single specificity. Subjects with single-specific anti-Ku antibody were likely to have ILD and increased creatine kinase levels than the others (145). Another article re-

Table. Autantibodies Detected in Connective Tissue Diseases.

Disease Category	Autoantigens	Prevalence (%)	Clinical Characteristics	Reference
RA	IgG (RF)	70–80	Structural progression	1-3
	Calpastatin	50–80	Inhibition of the function of calpastatin	4-7
	FRP	30	High disease activity of RA	8-16
	BiP	50–60	Citrullinated BiP is an autoantigen of ACPAs	17-19
	MBP	N.A.	Genetic risk factors	20
	RPL23A	N.A.	T cell responses to autoantigens confirmed	21
	Citrullinated proteins	60–80	Structural progression	22-30
	Carbamylated proteins	50–70	10–20 % positive of ACPA-negative RA patients	31-33
	ADA	N.A.	Detected when treated with bDMARDs	37-39
SLE	DNA	70-90	Correlated with disease activity	40-45
	Sm	15–25	Neuropsychiatric involvement	40
	Ribosomal P	10	Neuropsychiatric involvement	52
	Phospholipid	10–20	Thromboembolic events, pregnancy morbidity	40
	NR2	30	Neuropsychiatric involvement	46-49
PM / DM	ARS	30–40	ILD, mechanic's hand	60-73
	MDA5	50 (CADM)	Acute progressive ILD	74-86
	SFPQ	N.A.	53% positive in anti-MDA5-positive sera	87
	Mi-2	10–20 (DM)	DM, photosensitivity	88-90
	TIF-1 γ	20–30 (DM)	DM, association with cancer	91-94
	SRP	5–10	Necrotizing myopathy	95-100
	HMGCR	5–8	Statin-related myositis	101
	NT5C1A	N.A.	40–50 % positive in IBM	105-106
	Mitochondria	N.A.	PBC, Cardiac involvement	107-110
SSc	TOPO (Scl-70)	20–30	Diffuse type of scleroderma	111-113
	RNAP III	5–10	Diffuse type of scleroderma, renal crisis	114-115
	CENP	20–30	Limited type of scleroderma	116
	Th/To	2–4	Mild form of cutaneous involvement	117-118
	U3-RNP	3–8	Muscle involvement, PAH	119-124
	AT(1)R / ET (A)R	80	TGF-beta expression in endothelial cells (<i>in vitro</i>)	125
	M3 muscarinic receptor	60–80	Association with gastrointestinal dysfunction	126
	PDGF-R	90	Fibrosis with collagen production	127-134
	RNPC-3	3	Increased risk of cancer	135-136
Overlap	U1-RNP	100 (MCTD)	Raynaud's phenomenon, pulmonary arterial hypertension	57-58, 139
	Ku	30	SSc - PM overlap	140-146
	PM-Scl	10 (PM/DM)	Overlap with PM/DM	147-151

RA: rheumatoid arthritis, SLE: systemic lupus erythematosus, PM: polymyositis, DM: dermatomyositis, SSc: systemic sclerosis, FRP: foliastatin-related protein, BiP: immunoglobulin heavy chain binding protein, ACPA: anti-citrullinated protein antibody, MBP: myelin basic protein, RPL23A: 60S ribosomal protein L23a, IgG: immunoglobulin G, RF: rheumatoid factor, ADA: anti-drug antibodies, bDMARDs: biologic disease modifying anti-rheumatic drugs, NR2: N-methyl-D-aspartate receptor subunit 2, ARS: aminoacyl-tRNA synthetase, ILD: interstitial lung disease, MDA5: melanoma differentiation-associated gene 5, SFPQ: splicing factor proline/glutamine-rich protein, TIF-1 γ : transcriptional intermediary factor-1 γ , SRP: signal recognition particle, PBC: primary biliary cirrhosis, HMGCR: 3-hydroxy-3-methylglutaryl-coenzyme A reductase, NT5C1A: cytosolic 5'-nucleotidase 1A, IBM: inclusion body myositis, TOPO: DNA topoisomerase I, RNAP III: RNA polymerase III, CENP: centromere, RNP: RNA-containing particles, PAH: pulmonary arterial hypertension, AT(1)R: angiotensin II type 1 receptor, ET (A)R: endothelin-1 type A receptor, TGF-b: transforming growth factor-beta, PDGF-R: platelet-derived growth factor receptor, RNPC-3: RNA-binding protein-containing 3

ported that these antibodies appear more commonly among African American patients than among Caucasian patients with SLE, and that they were not present in samples obtained from patients with scleroderma (146). These observations suggest that ethnic differences influence the clinical manifestation in patients with anti-Ku antibody.

Anti-PM-Scl antibody, of which the autoantigen is a nuclear/nucleolar particle composed of several polypeptides, is

associated with PM/SSc overlap syndrome (147-149). Wodkowski et al. reported that, among 1,574 SSc patients, anti-PM-Scl antibodies were detected in <5% (48 subjects had antibody against PM75 antigen, and 18 had antibody against PM100 antigen) (150). As clinical manifestations, this antibody appears to be associated with lung and esophageal involvement; in addition, anti-PM-Scl may co-exist with malignancy in PM/DM patients (151).

Conclusion

The antibodies introduced in this review article are summarized in the included Table. As described above, in CTDs, various autoantibodies specific to each disease are produced; the types of targeted antigens vary, including the surface antigens of cells, antigens in the cytoplasm, and molecules in the nucleus. However, these autoantigens are poorly antigenic for healthy humans and animals, and it is generally difficult to obtain antibodies through artificial immunization in *in vivo* studies. No clear explanation has been agreed upon regarding why these autoantibodies are produced in patients. Furthermore, it has not been thoroughly clarified whether or not each of these antibodies is involved in the pathogenesis of the disease. Additional clues to clarify the mechanism underlying autoantibody production are anticipated in future studies.

Author's disclosure of potential Conflicts of Interest (COI).

Tsuneyo Mimori: Honoraria, Chugai Pharmaceutical, Bristol-Myers Squibb and Mitsubishi-Tanabe Pharma; Research funding, Chugai Pharmaceutical, Pfizer Japan, Eisai, Mitsubishi-Tanabe Pharma, Astellas Pharma, Daiichi Sankyo, AYUMI Pharmaceutical Corporation and Nippon Kayaku.

References

1. Rose HM, Ragan C, Pearce E, et al. Differential agglutination of normal and sensitized sheep erythrocytes by sera of patients with rheumatoid arthritis. *Proc Soc Exp Biol Med (New York, NY)* **68**: 1-6, 1948.
2. Aletaha D, Neogi T, Silman AJ, et al. 2010 Rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis Rheum* **62**: 2569-2581, 2010.
3. Aletaha D, Neogi T, Silman AJ, et al. 2010 rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Ann Rheum Dis* **69**: 1580-1588, 2010.
4. Mimori T, Suganuma K, Tanami Y, et al. Autoantibodies to calpastatin (an endogenous inhibitor for calcium-dependent neutral protease, calpain) in systemic rheumatic diseases. *Proc Natl Acad Sci USA* **92**: 7267-7271, 1995.
5. Despres N, Talbot G, Plouffe B, Boire G, Menard HA. Detection and expression of a cDNA clone that encodes a polypeptide containing two inhibitory domains of human calpastatin and its recognition by rheumatoid arthritis sera. *J Clin Invest* **95**: 1891-1896, 1995.
6. Yoshifuji H, Umehara H, Maruyama H, et al. Amelioration of experimental arthritis by a calpain-inhibitory compound: regulation of cytokine production by E-64-d *in vivo* and *in vitro*. *Int Immunol* **17**: 1327-1336, 2005.
7. Iguchi-Hashimoto M, Usui T, Yoshifuji H, et al. Overexpression of a minimal domain of calpastatin suppresses IL-6 production and Th17 development via reduced NF-kappaB and increased STAT5 signals. *PloS One* **6**: e27020, 2011.
8. Tanaka M, Ozaki S, Osakada F, Mori K, Okubo M, Nakao K. Cloning of follistatin-related protein as a novel autoantigen in systemic rheumatic diseases. *Int Immunol* **10**: 1305-1314, 1998.
9. Miyamae T, Marinov AD, Sowders D, et al. Follistatin-like protein-1 is a novel proinflammatory molecule. *J Immunol* **177**: 4758-4762, 2006.
10. Kawabata D, Tanaka M, Fujii T, et al. Ameliorative effects of follistatin-related protein/TSC-36/FSTL1 on joint inflammation in a mouse model of arthritis. *Arthritis Rheum* **50**: 660-668, 2004.
11. Clutter SD, Wilson DC, Marinov AD, Hirsch R. Follistatin-like protein 1 promotes arthritis by up-regulating IFN-gamma. *J Immunol* **182**: 234-239, 2009.
12. Miller M, Beppu A, Rosenthal P, et al. Fstl1 promotes asthmatic airway remodeling by inducing oncostatin M. *J Immunol* **195**: 3546-3556, 2015.
13. Liu Y, Wei J, Zhao Y, et al. Follistatin-like protein 1 promotes inflammatory reactions in nucleus pulposus cells by interacting with the MAPK and NFkappaB signaling pathways. *Oncotarget* **8**: 43023-43034, 2017.
14. Kim HJ, Kang WY, Seong SJ, Kim SY, Lim MS, Yoon YR. Follistatin-like 1 promotes osteoclast formation via RANKL-mediated NF-kappaB activation and M-CSF-induced precursor proliferation. *Cell Signal* **28**: 1137-1144, 2016.
15. Hayakawa S, Ohashi K, Shibata R, et al. Association of circulating follistatin-like 1 levels with inflammatory and oxidative stress markers in healthy men. *PloS One* **11**: e0153619, 2016.
16. Murakami K, Tanaka M, Usui T, et al. Follistatin-related protein/follistatin-like 1 evokes an innate immune response via CD14 and toll-like receptor 4. *FEBS Lett* **586**: 319-324, 2012.
17. Corrigan VM, Bodman-Smith MD, Fife MS, et al. The human endoplasmic reticulum molecular chaperone BiP is an autoantigen for rheumatoid arthritis and prevents the induction of experimental arthritis. *J Immunol* **166**: 1492-1498, 2001.
18. Blass S, Union A, Raymackers J, et al. The stress protein BiP is overexpressed and is a major B and T cell target in rheumatoid arthritis. *Arthritis Rheum* **44**: 761-771, 2001.
19. Shoda H, Hanata N, Sumitomo S, Okamura T, Fujio K, Yamamoto K. Immune responses to Mycobacterial heat shock protein 70 accompany self-reactivity to human BiP in rheumatoid arthritis. *Sci Rep* **6**: 22486, 2016.
20. Terao C, Ohmura K, Katayama M, et al. Myelin basic protein as a novel genetic risk factor in rheumatoid arthritis—a genome-wide study combined with immunological analyses. *PloS One* **6**: e20457, 2011.
21. Ito Y, Hashimoto M, Hirota K, et al. Detection of T cell responses to a ubiquitous cellular protein in autoimmune disease. *Science* **346**: 363-368, 2014.
22. Nishimura K, Sugiyama D, Kogata Y, et al. Meta-analysis: diagnostic accuracy of anti-cyclic citrullinated peptide antibody and rheumatoid factor for rheumatoid arthritis. *Ann Intern Med* **146**: 797-808, 2007.
23. Murakami K, Fujii T, Mimori T. [Rheumatoid arthritis: progress in diagnosis and treatment. Topics: II. Pathophysiology; 3. Autoantibodies in rheumatoid arthritis]. *Nihon Naika Gakkai Zasshi (Jpn Soc Intern Med)* **101**: 2844-2850, 2012 (in Japanese).
24. Ioan-Facsinay A, el-Bannoudi H, Scherer HU, et al. Anti-cyclic citrullinated peptide antibodies are a collection of anti-citrullinated protein antibodies and contain overlapping and non-overlapping reactivities. *Ann Rheum Dis* **70**: 188-193, 2011.
25. Snir O, Widhe M, Hermansson M, et al. Antibodies to several citrullinated antigens are enriched in the joints of rheumatoid arthritis patients. *Arthritis Rheum* **62**: 44-52, 2010.
26. Terao C, Suzuki A, Ikari K, et al. An association between amino acid position 74 of HLA-DRB1 and anti-citrullinated protein antibody levels in Japanese patients with anti-citrullinated protein antibody-positive rheumatoid arthritis. *Arthritis Rheumatol (Hoboken, NJ)* **67**: 2038-2045, 2015.
27. Ohmura K, Terao C, Maruya E, et al. Anti-citrullinated peptide antibody-negative RA is a genetically distinct subset: a definitive study using only bone-erosive ACPA-negative rheumatoid arthritis.

- Rheumatology (Oxford) **49**: 2298-2304, 2010.
28. Terao C, Ikari K, Ohmura K, et al. Quantitative effect of HLA-DRB1 alleles to ACPA levels in Japanese rheumatoid arthritis: no strong genetic impact of shared epitope to ACPA levels after stratification of HLA-DRB1*09:01. *Ann Rheum Dis* **71**: 1095-1097, 2012.
 29. Terao C, Ohmura K, Ikari K, et al. ACPA-negative RA consists of two genetically distinct subsets based on RF positivity in Japanese. *PLoS One* **7**: e40067, 2012.
 30. Terao C, Ohmura K, Kochi Y, et al. A large-scale association study identified multiple HLA-DRB1 alleles associated with ACPA-negative rheumatoid arthritis in Japanese subjects. *Ann Rheum Dis* **70**: 2134-2139, 2011.
 31. Nakabo S, Yoshifuji H, Hashimoto M, et al. Anti-carbamylated protein antibodies are detectable in various connective tissue diseases. *J Rheumatol* 2017.
 32. Shi J, Knevel R, Suwannalai P, et al. Autoantibodies recognizing carbamylated proteins are present in sera of patients with rheumatoid arthritis and predict joint damage. *Proc Natl Acad Sci USA* **108**: 17372-17377, 2011.
 33. Nakabo S, Hashimoto M, Ito S, et al. Carbamylated albumin is one of the target antigens of anti-carbamylated protein antibodies. *Rheumatology (Oxford)* **56**: 1217-1226, 2017.
 34. Jin H, Arase N, Hirayasu K, et al. Autoantibodies to IgG/HLA class II complexes are associated with rheumatoid arthritis susceptibility. *Proc Natl Acad Sci USA* **111**: 3787-3792, 2014.
 35. Tanimura K, Jin H, Suenaga T, et al. β 2-glycoprotein I/HLA class II complexes are novel autoantigens in antiphospholipid syndrome. *Blood* **125**: 2835-2844, 2015.
 36. Hiwa R, Ohmura K, Arase N, et al. Myeloperoxidase/HLA class II complexes recognized by autoantibodies in microscopic polyangiitis. *Arthritis Rheumatol* **69**: 2069-2080, 2017.
 37. Prado MS, Bendtzen K, Andrade LEC. Biological anti-TNF drugs: immunogenicity underlying treatment failure and adverse events. *Expert Opin Drug Metab Toxicol* **13**: 985-995, 2017.
 38. Yukawa N, Fujii T, Kondo-Ishikawa S, et al. Correlation of antinuclear antibody and anti-double-stranded DNA antibody with clinical response to infliximab in patients with rheumatoid arthritis: a retrospective clinical study. *Arthritis Res Ther* **13**: R213, 2011.
 39. Ishikawa Y, Fujii T, Ishikawa SK, et al. Immunogenicity and lupus-like autoantibody production can be linked to each other along with type I interferon production in patients with rheumatoid arthritis treated with infliximab: a retrospective study of a single center cohort. *PLoS One* **11**: e0162896, 2016.
 40. Tan EM, Cohen AS, Fries JF, et al. The 1982 revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum* **25**: 1271-1277, 1982.
 41. Hochberg MC. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum* **40**: 1725, 1997.
 42. Petri M, Orbai AM, Alarcon GS, et al. Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum* **64**: 2677-2686, 2012.
 43. Tsokos GC. Systemic lupus erythematosus. *N Engl J Med* **365**: 2110-2121, 2011.
 44. Rekvig OP. The anti-DNA antibody: origin and impact, dogmas and controversies. *Nat Rev Rheumatol* **11**: 530-540, 2015.
 45. Pisetsky DS. Anti-DNA antibodies-quintessential biomarkers of SLE. *Nat Rev Rheumatol* **12**: 102-110, 2016.
 46. Hanly JG, Robichaud J, Fisk JD. Anti-NR2 glutamate receptor antibodies and cognitive function in systemic lupus erythematosus. *J Rheumatol* **33**: 1553-1558, 2006.
 47. Hirohata S, Arinuma Y, Yanagida T, Yoshio T. Blood-brain barrier damages and intrathecal synthesis of anti-N-methyl-D-aspartate receptor NR2 antibodies in diffuse psychiatric/neuropsychological syndromes in systemic lupus erythematosus. *Arthritis Res Ther* **16**: R77, 2014.
 48. Kowal C, Degiorgio LA, Lee JY, et al. Human lupus autoantibodies against NMDA receptors mediate cognitive impairment. *Proc Natl Acad Sci USA* **103**: 19854-19859, 2006.
 49. Lapteva L, Nowak M, Yarboro CH, et al. Anti-N-methyl-D-aspartate receptor antibodies, cognitive dysfunction, and depression in systemic lupus erythematosus. *Arthritis Rheum* **54**: 2505-2514, 2006.
 50. Hanly JG, Urowitz MB, Su L, et al. Autoantibodies as biomarkers for the prediction of neuropsychiatric events in systemic lupus erythematosus. *Ann Rheum Dis* **70**: 1726-1732, 2011.
 51. Shi ZR, Cao CX, Tan GZ, Wang L. The association of serum anti-ribosomal P antibody with clinical and serological disorders in systemic lupus erythematosus: a systematic review and meta-analysis. *Lupus* **24**: 588-596, 2015.
 52. Massardo L, Bravo-Zehnder M, Calderon J, et al. Anti-N-methyl-D-aspartate receptor and anti-ribosomal-P autoantibodies contribute to cognitive dysfunction in systemic lupus erythematosus. *Lupus* **24**: 558-568, 2015.
 53. Bravo-Zehnder M, Toledo EM, Segovia-Miranda F, et al. Anti-ribosomal P protein autoantibodies from patients with neuropsychiatric lupus impair memory in mice. *Arthritis Rheumatol (Hoboken, NJ)* **67**: 204-214, 2015.
 54. Kowal C, DeGiorgio LA, Nakaoka T, et al. Cognition and immunity; antibody impairs memory. *Immunity* **21**: 179-188, 2004.
 55. Matus S, Burgos PV, Bravo-Zehnder M, et al. Antiribosomal-P autoantibodies from psychiatric lupus target a novel neuronal surface protein causing calcium influx and apoptosis. *J Exp Med* **204**: 3221-3234, 2007.
 56. Nagai T, Arinuma Y, Yanagida T, Yamamoto K, Hirohata S. Anti-ribosomal P protein antibody in human systemic lupus erythematosus up-regulates the expression of proinflammatory cytokines by human peripheral blood monocytes. *Arthritis Rheum* **52**: 847-855, 2005.
 57. Sato T, Fujii T, Yokoyama T, et al. Anti-U1 RNP antibodies in cerebrospinal fluid are associated with central neuropsychiatric manifestations in systemic lupus erythematosus and mixed connective tissue disease. *Arthritis Rheum* **62**: 3730-3740, 2010.
 58. Yokoyama T, Fujii T, Kondo-Ishikawa S, et al. Association between anti-U1 ribonucleoprotein antibodies and inflammatory mediators in cerebrospinal fluid of patients with neuropsychiatric systemic lupus erythematosus. *Lupus* **23**: 635-642, 2014.
 59. Stamatiades EG, Tremblay ME, Bohm M, et al. Immune monitoring of trans-endothelial transport by kidney-resident macrophages. *Cell* **166**: 991-1003, 2016.
 60. Nakashima R, Hosono Y, Mimori T. Clinical significance and new detection system of autoantibodies in myositis with interstitial lung disease. *Lupus* **25**: 925-933, 2016.
 61. Ikeda N, Takahashi K, Yamaguchi Y, Inasaka M, Kuwana M, Ikezawa Z. Analysis of dermatomyositis-specific autoantibodies and clinical characteristics in Japanese patients. *J Dermatol* **38**: 973-979, 2011.
 62. Mathews MB, Bernstein RM. Myositis autoantibody inhibits histidyl-tRNA synthetase: a model for autoimmunity. *Nature* **304**: 177-179, 1983.
 63. Nishikai M, Reichlin M. Heterogeneity of precipitating antibodies in polymyositis and dermatomyositis. Characterization of the Jo-1 antibody system. *Arthritis Rheum* **23**: 881-888, 1980.
 64. Mathews MB, Reichlin M, Hughes GR, Bernstein RM. Anti-threonyl-tRNA synthetase, a second myositis-related autoantibody. *J Exp Med* **160**: 420-434, 1984.
 65. Bunn CC, Bernstein RM, Mathews MB. Autoantibodies against alanyl-tRNA synthetase and tRNAAla coexist and are associated with myositis. *J Exp Med* **163**: 1281-1291, 1986.
 66. Targoff IN, Trieu EP, Plotz PH, Miller FW. Antibodies to glycy-

- transfer RNA synthetase in patients with myositis and interstitial lung disease. *Arthritis Rheum* **35**: 821-830, 1992.
67. Targoff IN, Trieu EP, Miller FW. Reaction of anti-OJ autoantibodies with components of the multi-enzyme complex of aminoacyl-tRNA synthetases in addition to isoleucyl-tRNA synthetase. *J Clin Invest* **91**: 2556-2564, 1993.
 68. Hirakata M, Suwa A, Nagai S, et al. Anti-KS: identification of autoantibodies to asparaginyl-transfer RNA synthetase associated with interstitial lung disease. *J Immunol* **162**: 2315-2320, 1999.
 69. Betteridge Z, Gunawardena H, North J, Slinn J, McHugh N. Anti-synthetase syndrome: a new autoantibody to phenylalanyl transfer RNA synthetase (anti-Zo) associated with polymyositis and interstitial pneumonia. *Rheumatology (Oxford)* **46**: 1005-1008, 2007.
 70. Satoh M, Ceribelli A, Chan EK. Common pathways of autoimmune inflammatory myopathies and genetic neuromuscular disorders. *Clin Rev Allergy Immunol* **42**: 16-25, 2012.
 71. Yoshifuji H, Fujii T, Kobayashi S, et al. Anti-aminoacyl-tRNA synthetase antibodies in clinical course prediction of interstitial lung disease complicated with idiopathic inflammatory myopathies. *Autoimmunity* **39**: 233-241, 2006.
 72. Fujisawa T, Hozumi H, Kono M, et al. Predictive factors for long-term outcome in polymyositis/dermatomyositis-associated interstitial lung diseases. *Respir Investig* **55**: 130-137, 2017.
 73. Nakashima R, Imura Y, Hosono Y, et al. The multicenter study of a new assay for simultaneous detection of multiple anti-aminoacyl-tRNA synthetases in myositis and interstitial pneumonia. *PLoS One* **9**: e85062, 2014.
 74. Sato S, Hirakata M, Kuwana M, et al. Autoantibodies to a 140-kd polypeptide, CADM-140, in Japanese patients with clinically amyopathic dermatomyositis. *Arthritis Rheum* **52**: 1571-1576, 2005.
 75. Sato S, Hoshino K, Satoh T, et al. RNA helicase encoded by melanoma differentiation-associated gene 5 is a major autoantigen in patients with clinically amyopathic dermatomyositis: association with rapidly progressive interstitial lung disease. *Arthritis Rheum* **60**: 2193-2200, 2009.
 76. Nakashima R, Imura Y, Kobayashi S, et al. The RIG-I-like receptor IFIH1/MDA5 is a dermatomyositis-specific autoantigen identified by the anti-CADM-140 antibody. *Rheumatology (Oxford)* **49**: 433-440, 2010.
 77. Chen F, Wang D, Shu X, Nakashima R, Wang G. Anti-MDA5 antibody is associated with A/SIP and decreased T cells in peripheral blood and predicts poor prognosis of ILD in Chinese patients with dermatomyositis. *Rheumatol Int* **32**: 3909-3915, 2012.
 78. Fiorentino D, Chung L, Zwerner J, Rosen A, Casciola-Rosen L. The mucocutaneous and systemic phenotype of dermatomyositis patients with antibodies to MDA5 (CADM-140): a retrospective study. *J Am Acad Dermatol* **65**: 25-34, 2011.
 79. Ceribelli A, Fredi M, Taraborelli M, et al. Prevalence and clinical significance of anti-MDA5 antibodies in European patients with polymyositis/dermatomyositis. *Clin Exp Rheumatol* **32**: 891-897, 2014.
 80. Labrador-Horrillo M, Martinez MA, Selva-O'Callaghan A, et al. Anti-MDA5 antibodies in a large Mediterranean population of adults with dermatomyositis. *J Immunol Res* **2014**: 290797, 2014.
 81. Gono T, Kawaguchi Y, Ozeki E, et al. Serum ferritin correlates with activity of anti-MDA5 antibody-associated acute interstitial lung disease as a complication of dermatomyositis. *Mod Rheumatol* **21**: 223-227, 2011.
 82. Gono T, Sato S, Kawaguchi Y, et al. Anti-MDA5 antibody, ferritin and IL-18 are useful for the evaluation of response to treatment in interstitial lung disease with anti-MDA5 antibody-positive dermatomyositis. *Rheumatology (Oxford)* **51**: 1563-1570, 2012.
 83. Gono T, Kaneko H, Kawaguchi Y, et al. Cytokine profiles in polymyositis and dermatomyositis complicated by rapidly progressive or chronic interstitial lung disease. *Rheumatology (Oxford)* **53**: 2196-2203, 2014.
 84. Horai Y, Koga T, Fujikawa K, et al. Serum interferon-alpha is a useful biomarker in patients with anti-melanoma differentiation-associated gene 5 (MDA5) antibody-positive dermatomyositis. *Mod Rheumatol* **25**: 85-89, 2015.
 85. Tanizawa K, Handa T, Nakashima R, et al. HRCT features of interstitial lung disease in dermatomyositis with anti-CADM-140 antibody. *Respir Med* **105**: 1380-1387, 2011.
 86. Abe T, Tsunoda S, Nishioka A, et al. Reliability and clinical utility of Enzyme-linked immunosorbent assay for detection of anti-aminoacyl-tRNA synthetase antibody. *Nihon Rinsho Men'eki Gakkaishi (Jpn J Clin Immunol)* **39**: 140-144, 2016 (in Japanese, Abstract in English).
 87. Hosono Y, Nakashima R, Serada S, et al. Splicing factor proline/glutamine-rich is a novel autoantigen of dermatomyositis and associated with anti-melanoma differentiation-associated gene 5 antibody. *J Autoimmun* **77**: 116-122, 2017.
 88. Targoff IN, Reichlin M. The association between Mi-2 antibodies and dermatomyositis. *Arthritis Rheum* **28**: 796-803, 1985.
 89. Wang HB, Zhang Y. Mi2, an auto-antigen for dermatomyositis, is an ATP-dependent nucleosome remodeling factor. *Nucleic Acids Res* **29**: 2517-2521, 2001.
 90. Mierau R, Dick T, Bartz-Bazzanella P, Keller E, Albert ED, Genth E. Strong association of dermatomyositis-specific Mi-2 autoantibodies with a tryptophan at position 9 of the HLA-DR beta chain. *Arthritis Rheum* **39**: 868-876, 1996.
 91. Hoshino K, Muro Y, Sugiura K, Tomita Y, Nakashima R, Mimori T. Anti-MDA5 and anti-TIF1-gamma antibodies have clinical significance for patients with dermatomyositis. *Rheumatology (Oxford)* **49**: 1726-1733, 2010.
 92. Fujimoto M, Hamaguchi Y, Kaji K, et al. Myositis-specific anti-155/140 autoantibodies target transcription intermediary factor 1 family proteins. *Arthritis Rheum* **64**: 513-522, 2012.
 93. Hida A, Yamashita T, Hosono Y, et al. Anti-TIF1-gamma antibody and cancer-associated myositis: a clinicohistopathologic study. *Neurology* **87**: 299-308, 2016.
 94. Fujimoto M, Murakami A, Kurei S, et al. Enzyme-linked immunosorbent assays for detection of anti-transcriptional intermediary factor-1 gamma and anti-Mi-2 autoantibodies in dermatomyositis. *J Dermatol Sci* **84**: 272-281, 2016.
 95. Dimitri D, Andre C, Roucoules J, Hosseini H, Humbel RL, Authier FJ. Myopathy associated with anti-signal recognition peptide antibodies: clinical heterogeneity contrasts with stereotyped histopathology. *Muscle Nerve* **35**: 389-395, 2007.
 96. Hengstman GJ, ter Laak HJ, Vree Egberts WT, et al. Anti-signal recognition particle autoantibodies: marker of a necrotising myopathy. *Ann Rheum Dis* **65**: 1635-1638, 2006.
 97. Kao AH, Lacomis D, Lucas M, Fertig N, Oddis CV. Anti-signal recognition particle autoantibody in patients with and patients without idiopathic inflammatory myopathy. *Arthritis Rheum* **50**: 209-215, 2004.
 98. Takada T, Hirakata M, Suwa A, et al. Clinical and histopathological features of myopathies in Japanese patients with anti-SRP autoantibodies. *Mod Rheumatol* **19**: 156-164, 2009.
 99. Targoff IN, Johnson AE, Miller FW. Antibody to signal recognition particle in polymyositis. *Arthritis Rheum* **33**: 1361-1370, 1990.
 100. Suzuki S, Nishikawa A, Kuwana M, et al. Inflammatory myopathy with anti-signal recognition particle antibodies: case series of 100 patients. *Orphanet J Rare Dis* **10**: 61, 2015.
 101. Mammen AL, Chung T, Christopher-Stine L, et al. Autoantibodies against 3-hydroxy-3-methylglutaryl-coenzyme A reductase in patients with statin-associated autoimmune myopathy. *Arthritis Rheum* **63**: 713-721, 2011.
 102. Badrising UA, Maat-Schieman M, van Duinen SG, et al. Epidemiology of inclusion body myositis in the Netherlands: a nation-

- wide study. *Neurology* **55**: 1385-1387, 2000.
103. Brady S, Squier W, Hilton-Jones D. Clinical assessment determines the diagnosis of inclusion body myositis independently of pathological features. *J Neurol Neurosurg Psy* **84**: 1240-1246, 2013.
 104. Mastaglia FL, Needham M. Inclusion body myositis: a review of clinical and genetic aspects, diagnostic criteria and therapeutic approaches. *J Clin Neurosci* **22**: 6-13, 2015.
 105. Larman HB, Salajegheh M, Nazareno R, et al. Cytosolic 5'-nucleotidase 1A autoimmunity in sporadic inclusion body myositis. *Ann Neurol* **73**: 408-418, 2013.
 106. Lilleker JB, Rietveld A, Pye SR, et al. Cytosolic 5'-nucleotidase 1A autoantibody profile and clinical characteristics in inclusion body myositis. *Ann Rheum Dis* **76**: 862-868, 2017.
 107. Maeda MH, Tsuji S, Shimizu J. Inflammatory myopathies associated with anti-mitochondrial antibodies. *Brain* **135**: 1767-1777, 2012.
 108. Albayda J, Khan A, Casciola-Rosen L, Corse AM, Paik JJ, Christopher-Stine L. Inflammatory myopathy associated with anti-mitochondrial antibodies: a distinct phenotype with cardiac involvement. *Semin Arthritis Rheum* **47**: 552-556, 2018.
 109. Konishi H, Fukuzawa K, Mori S, et al. Anti-mitochondrial M2 antibodies enhance the risk of supraventricular arrhythmias in patients with elevated hepatobiliary enzyme levels. *Intern Med* **56**: 1771-1779, 2017.
 110. Uenaka T, Kowa H, Ohtsuka Y, et al. Less limb muscle involvement in myositis patients with anti-mitochondrial antibodies. *Eur Neurol* **78**: 290-295, 2017.
 111. Hamaguchi Y, Hasegawa M, Fujimoto M, et al. The clinical relevance of serum antinuclear antibodies in Japanese patients with systemic sclerosis. *Br J Dermatol* **158**: 487-495, 2008.
 112. Kayser C, Fritzler MJ. Autoantibodies in systemic sclerosis: unanswered questions. *Front Immunol* **6**: 167, 2015.
 113. Steen VD. Autoantibodies in systemic sclerosis. *Semin Arthritis Rheum* **35**: 35-42, 2005.
 114. Hamaguchi Y, Kodaera M, Matsushita T, et al. Clinical and immunologic predictors of scleroderma renal crisis in Japanese systemic sclerosis patients with anti-RNA polymerase III autoantibodies. *Arthritis Rheumatol* **67**: 1045-1052, 2015.
 115. Sobanski V, Dauchet L, Lefevre G, et al. Prevalence of anti-RNA polymerase III antibodies in systemic sclerosis: new data from a French cohort and a systematic review and meta-analysis. *Arthritis Rheumatol* **66**: 407-417, 2014.
 116. Perosa F, Favoino E, Favia IE, et al. Subspecificities of anticentromeric protein A antibodies identify systemic sclerosis patients at higher risk of pulmonary vascular disease. *Medicine* **95**: e3931, 2016.
 117. Yuan Y, Tan E, Reddy R. The 40-kilodalton to autoantigen associates with nucleotides 21 to 64 of human mitochondrial RNA processing/7-2 RNA *in vitro*. *Mol Cell Biol* **11**: 5266-5274, 1991.
 118. Yamane K, Ihn H, Kubo M, et al. Antibodies to Th/To ribonucleoprotein in patients with localized scleroderma. *Rheumatology (Oxford)* **40**: 683-686, 2001.
 119. Lischwe MA, Ochs RL, Reddy R, et al. Purification and partial characterization of a nucleolar scleroderma antigen (Mr = 34,000; pI, 8.5) rich in NG,NG-dimethylarginine. *J Biol Chem* **260**: 14304-14310, 1985.
 120. Reimer G, Steen VD, Penning CA, Medsger TA Jr, Tan EM. Correlates between autoantibodies to nucleolar antigens and clinical features in patients with systemic sclerosis (scleroderma). *Arthritis Rheum* **31**: 525-532, 1988.
 121. Okano Y, Steen VD, Medsger TA Jr. Autoantibody to U3 nucleolar ribonucleoprotein (fibrillarin) in patients with systemic sclerosis. *Arthritis Rheum* **35**: 95-100, 1992.
 122. Tormey VJ, Bunn CC, Denton CP, Black CM. Anti-fibrillarin antibodies in systemic sclerosis. *Rheumatology (Oxford)* **40**: 1157-1162, 2001.
 123. Mehra S, Walker J, Patterson K, Fritzler MJ. Autoantibodies in systemic sclerosis. *Autoimmun Rev* **12**: 340-354, 2013.
 124. Aggarwal R, Lucas M, Fertig N, Oddis CV, Medsger TA Jr. Anti-U3 RNP autoantibodies in systemic sclerosis. *Arthritis Rheum* **60**: 1112-1118, 2009.
 125. Riemekasten G, Philippe A, Nather M, et al. Involvement of functional autoantibodies against vascular receptors in systemic sclerosis. *Ann Rheum Dis* **70**: 530-536, 2011.
 126. Goldblatt F, Gordon TP, Waterman SA. Antibody-mediated gastrointestinal dysmotility in scleroderma. *Gastroenterology* **123**: 1144-1150, 2002.
 127. Balada E, Simeon-Aznar CP, Ordi-Ros J, et al. Anti-PDGFR-alpha antibodies measured by non-bioactivity assays are not specific for systemic sclerosis. *Ann Rheum Dis* **67**: 1027-1029, 2008.
 128. Baroni SS, Santillo M, Bevilacqua F, et al. Stimulatory autoantibodies to the PDGF receptor in systemic sclerosis. *New Engl J Med* **354**: 2667-2676, 2006.
 129. Classen JF, Henrohn D, Rorsman F, et al. Lack of evidence of stimulatory autoantibodies to platelet-derived growth factor receptor in patients with systemic sclerosis. *Arthritis Rheum* **60**: 1137-1144, 2009.
 130. Dragun D, Distler JH, Riemekasten G, Distler O. Stimulatory autoantibodies to platelet-derived growth factor receptors in systemic sclerosis: what functional autoimmunity could learn from receptor biology. *Arthritis Rheum* **60**: 907-911, 2009.
 131. Kaviani N, Servettaz A, Marut W, et al. Sunitinib inhibits the phosphorylation of platelet-derived growth factor receptor beta in the skin of mice with scleroderma-like features and prevents the development of the disease. *Arthritis Rheum* **64**: 1990-2000, 2012.
 132. Luchetti MM, Moroncini G, Jose Escamez M, et al. Induction of scleroderma fibrosis in skin-humanized mice by administration of anti-platelet-derived growth factor receptor agonistic autoantibodies. *Arthritis Rheumatol* **68**: 2263-2273, 2016.
 133. Makino K, Makino T, Stawski L, et al. Blockade of PDGF receptors by crenolanib has therapeutic effect in patient fibroblasts and in preclinical models of systemic sclerosis. *J Invest Dermatol* **137**: 1671-1681, 2017.
 134. Spies-Weissbart B, Schilling K, Bohmer F, Hochhaus A, Sayer HG, Scholl S. Lack of association of platelet-derived growth factor (PDGF) receptor autoantibodies and severity of chronic graft-versus-host disease (GvHD). *J Cancer Res Clin Oncol* **139**: 1397-1404, 2013.
 135. Fertig N, Domsic RT, Rodriguez-Reyna T, et al. Anti-U11/U12 RNP antibodies in systemic sclerosis: a new serologic marker associated with pulmonary fibrosis. *Arthritis Rheum* **61**: 958-965, 2009.
 136. Shah AA, Xu G, Rosen A, et al. Brief report: anti-RNPC-3 antibodies as a marker of cancer-associated scleroderma. *Arthritis Rheumatol* **69**: 1306-1312, 2017.
 137. Sharp GC, Irvin WS, Tan EM, Gould RG, Holman HR. Mixed connective tissue disease-an apparently distinct rheumatic disease syndrome associated with a specific antibody to an extractable nuclear antigen (ENA). *Am J Med* **52**: 148-159, 1972.
 138. Ciang NC, Pereira N, Isenberg DA. Mixed connective tissue disease-enigma variations? *Rheumatology (Oxford)* **56**: 326-333, 2017.
 139. Gunnarsson R, Hetlevik SO, Lilleby V, Molberg O. Mixed connective tissue disease. *Best Pract Res Clin Rheumatol* **30**: 95-111, 2016.
 140. Mimori T. Scleroderma-polymyositis overlap syndrome. Clinical and serologic aspects. *Int J Dermatol* **26**: 419-425, 1987.
 141. Francoeur AM, Peebles CL, Gompper PT, Tan EM. Identification of Ki (Ku, p70/p80) autoantigens and analysis of anti-Ki autoantibody reactivity. *J Immunol* **136**: 1648-1653, 1986.
 142. Mimori T, Hardin JA, Steitz JA. Characterization of the DNA-

- binding protein antigen Ku recognized by autoantibodies from patients with rheumatic disorders. *J Biol Chem* **261**: 2274-2278, 1986.
- 143.** Reeves WH. Use of monoclonal antibodies for the characterization of novel DNA-binding proteins recognized by human autoimmune sera. *J Exp Med* **161**: 18-39, 1985.
- 144.** Yaneva M, Busch H. A 10S particle released from deoxyribonuclease-sensitive regions of HeLa cell nuclei contains the 86-kilodalton-70-kilodalton protein complex. *Biochemistry* **25**: 5057-5063, 1986.
- 145.** Hoa S, Hudson M, Troyanov Y, et al. Single-specificity anti-Ku antibodies in an international cohort of 2140 systemic sclerosis subjects: clinical associations. *Medicine* **95**: e4713, 2016.
- 146.** Wang J, Satoh M, Kabir F, et al. Increased prevalence of autoantibodies to ku antigen in African American versus white patients with systemic lupus erythematosus. *Arthritis Rheum* **44**: 2367-2370, 2001.
- 147.** Alderuccio F, Chan EK, Tan EM. Molecular characterization of an autoantigen of PM-Scl in the polymyositis/scleroderma overlap syndrome: a unique and complete human cDNA encoding an apparent 75-kD acidic protein of the nucleolar complex. *J Exp Med* **173**: 941-952, 1991.
- 148.** Gelpi C, Alguero A, Angeles Martinez M, Vidal S, Juarez C, Rodriguez-Sanchez JL. Identification of protein components reactive with anti-PM/Scl autoantibodies. *Clin Exp Immunol* **81**: 59-64, 1990.
- 149.** Reimer G, Scheer U, Peters JM, Tan EM. Immunolocalization and partial characterization of a nucleolar autoantigen (PM-Scl) associated with polymyositis/scleroderma overlap syndromes. *J Immunol* **137**: 3802-3808, 1986.
- 150.** Wodkowski M, Hudson M, Proudman S, et al. Clinical correlates of monospecific anti-PM75 and anti-PM100 antibodies in a tri-nation cohort of 1574 systemic sclerosis subjects. *Autoimmunity* **48**: 542-551, 2015.
- 151.** Marie I, Lahaxe L, Tiev K, et al. [Idiopathic inflammatory myopathies with anti-PM-Scl antibodies: case series and literature review]. *Rev Med Interne* **31**: 540-544, 2010 (in French, Abstract in English).

The Internal Medicine is an Open Access journal distributed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License. To view the details of this license, please visit (<https://creativecommons.org/licenses/by-nc-nd/4.0/>).