




# *Salmonella enterica* Newserovar Abeokuta Genome Sequence, Strain OG19FER4 Isolated from Poultry Feed in Nigeria

Idowu O. Fagbamila,<sup>a</sup> Alejandra Hernandez-Segura,<sup>b</sup>  Maaïke van den Beld,<sup>b</sup> Kirsten Mooijman,<sup>c</sup> Massimiliano Orsini,<sup>d</sup> Olawunmi T. Ajayi,<sup>a</sup> Sati Ngulukun,<sup>a</sup> Alexander Ray Jambalang,<sup>a</sup> Nancy Sati,<sup>a</sup> Paulinus Emennaa,<sup>a</sup> Paul I. Ankel,<sup>a</sup> Maryam Muhammad,<sup>a</sup> Lisa Barco<sup>d</sup>

<sup>a</sup>Bacterial Research Division, National Veterinary Research Institute, Vom, Plateau State, Nigeria

<sup>b</sup>Centre for Infectious Disease Control, National Institute for Public Health and the Environment (RIVM), Bilthoven, the Netherlands

<sup>c</sup>European Union Reference Laboratory (EURL) for Salmonella, National Institute for Public Health and the Environment (RIVM), Bilthoven, the Netherlands

<sup>d</sup>OIE and National Reference Laboratory for Salmonellosis, Istituto Zooprofilattico Sperimentale delle Venezie, Legnaro (PD), Italy

**ABSTRACT** This report announces the genome of a newly confirmed *Salmonella* serovar (*Salmonella enterica* serovar Abeokuta) that was isolated from a poultry feed sample collected on a farm in Abeokuta, capital of Ogun State in Nigeria. *Salmonella* Abeokuta has not been identified outside Nigeria, nor does it appear to be a cause for concern for animal and human health.

*Salmonella* is a common cause of gastroenteritis in humans (1) and even life-threatening disease (2). Moreover, *Salmonella* can lead to economic losses (3, 4).

*Salmonella* surveillance was conducted on poultry farms in Nigeria (2012 to 2015) (5). Here, we report the genome sequence of one inconclusive serovar that was isolated from a poultry feed sample collected on 13 February 2012 (latitude, 7.13N; longitude, 3.27E) in Abeokuta (Ogun State, Nigeria). The strain was isolated according to ISO 6579:2002 (6) and confirmed by ISO/TR 6579-3:2014 (7) as *Salmonella enterica* subsp. *enterica* 30:d:z<sub>6</sub>. This seroformula had never been published in the White-Kauffmann-Le Minor scheme (8), and the World Health Organization (WHO) Reference Center of *Salmonella* confirmed a new serovar by phenotypic and genotypic typing, now called *Salmonella enterica* subsp. *enterica* serovar Abeokuta (S. Abeokuta).

The isolate was cultured in tryptic soy broth (Sigma-Aldrich, Inc.) at 37°C overnight, and genomic DNA was extracted using the GenElute bacterial genomic DNA kit (Sigma-Aldrich, Inc.) according to the manufacturer's instructions. Whole-genome sequence was obtained using short-read and long-read sequencing approaches. Short-read libraries were prepared using the Illumina Nextera XT DNA library prep kit (Illumina, Inc.) and sequenced using Illumina NextSeq 500/550, resulting in 5,253,397 paired-end reads (150 bp), with a theoretical sequencing depth of about 150×. For long-read sequencing, the library was prepared with a rapid barcoding kit (SQK-RBK004) (no DNA shearing or size selection), loaded onto an R9.4.1 flow cell (FLO-MIN106), and sequenced in a GridION platform for 48 h. Live base calling was performed using GridION software (MinKNOW v21.05.25, MinKNOW GUI v4.0.20, and Ont-kingfisher-ui-gridion v4.3.28) obtaining 9,676 good-quality long reads with an average length of 8,591.66 bp, providing an average sequencing depth of 16×. Filtering was done using FastP (v0.20.1) (9) for short reads, and default filtering was performed with the minKNOW software followed by NanoFilt 2.8.0 (10) and Filtrong v0.2.1 (<https://github.com/rwrick/Filtrong>) for long reads. The quality control used FastQC (v0.11.9) (<https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>) for short reads and PycoQC 2.5.2 for long reads, resulting in an  $N_{50}$  read length of 20,549. The seroformula was confirmed using SeqSero2 (11). A hybrid assembly was performed using Unicycler (v0.4.8) (12), including a rotation step to ensure the *dnaA* gene was at the start of the forward strand. This resulted

**Editor** Vincent Bruno, University of Maryland School of Medicine

**Copyright** © 2022 Fagbamila et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Massimiliano Orsini, [morsini@izsvenezie.it](mailto:morsini@izsvenezie.it).

The authors declare no conflict of interest.

**Received** 20 May 2022

**Accepted** 14 September 2022

**Published** 28 September 2022

in one circular contig of 4,957,438 bp in length and 52.01% GC content. Analysis with CheckM (v1.1.3) (13) determined 99.54% genome completeness. The genome was annotated using PGAP (6.0) (14), which returned 4,788 genes: 4,564 protein-coding genes, 83 tRNAs, and 7 complete rRNA operons. ResFinder 4.0 software (15) predicted resistance to nalidixic acid, ciprofloxacin, amikacin, and tobramycin. The strain 10734/15 genome sequence was submitted to Enterobase (16) by the WHO Reference Center of *Salmonella*, resulting in 7-locus multilocus sequence type (MLST) type 8600, a unique ST. Further, using *Salmonella* cgMLST V2 plus HierCC V1 (17), the genome belongs to superlineage HC2000/219927, which includes *Salmonella enterica* serovar Kakikoka only.

**Data availability.** Both long and short reads were submitted to the SRA database under the BioProject accession identifier (ID) [PRJNA816352](https://doi.org/10.6026/PRJNA816352), with the accession IDs [SRR18503845](https://doi.org/10.6026/SRR18503845) and [SRR18503846](https://doi.org/10.6026/SRR18503846), respectively. The annotated genome sequence is available in the NCBI RefSeq database (accession ID [NZ\\_CP093445.1](https://doi.org/10.6026/NZ_CP093445.1)).

## ACKNOWLEDGMENT

We thank F.X. Weill from the WHO Reference Center of *Salmonella* for submitting the genome of the *S. Abeokuta* strain to Enterobase.

## REFERENCES

- Scallan E, Hoekstra RM, Angulo FJ, Tauxe RV, Widdowson M-A, Roy SL, Jones JL, Griffin PM. 2011. Foodborne illness acquired in the United States—major pathogens. *Emerg Infect Dis* 17:7–15. <https://doi.org/10.3201/eid1701.P11101>.
- Ao TT, Feasey NA, Gordon MA, Keddy KH, Angulo FJ, Crump JA. 2015. Global burden of invasive nontyphoidal *Salmonella* disease, 2010(1). *Emerg Infect Dis* 21:941–949. <https://doi.org/10.3201/eid2106.140999>.
- Kagambega A, Lienemann T, Aulu L, Traoré AS, Barro N, Siitonen A, Haukka K. 2013. Prevalence and characterization of *Salmonella enterica* from the feces of cattle, poultry, swine and hedgehogs in Burkina Faso and their comparison to human *Salmonella* isolates. *BMC Microbiol* 13:253. <https://doi.org/10.1186/1471-2180-13-253>.
- Raufu I, Bortolaia V, Svendsen CA, Ameh JA, Ambali AG, Aarestrup FM, Hendriksen RS. 2013. The first attempt of an active integrated laboratory-based *Salmonella* surveillance programme in the north-eastern region of Nigeria. *J Appl Microbiol* 115:1059–1067. <https://doi.org/10.1111/jam.12304>.
- Fagbamila IO, Barco L, Mancin M, Kwaga J, Ngulukun SS, Zavagnin P, Lettini AA, Lorenzetto M, Abdu PA, Kabir J, Umoh J, Ricci A, Muhammad M. 2017. *Salmonella* serovars and their distribution in Nigerian commercial chicken layer farms. *PLoS One* 12:e0173097. <https://doi.org/10.1371/journal.pone.0173097>.
- International Organization for Standardization. 2002. ISO 6579:2002. Microbiology of food and animal feeding stuff —horizontal method for the detection of *Salmonella* spp. International Organization for Standardization, Geneva, Switzerland.
- International Organization for Standardization. 2014. ISO/TR 6579-3:2014. Microbiology of the food chain—horizontal method for the detection, enumeration and serotyping of *Salmonella*. Part 3: guidelines for serotyping of *Salmonella* spp. International Organization for Standardization, Geneva, Switzerland.
- Grimont P, Weill F-X. 2007. Antigenic formulae of the salmonella serovars, 9th ed. Institut Pasteur, Paris, France. [https://www.pasteur.fr/sites/default/files/veng\\_0.pdf](https://www.pasteur.fr/sites/default/files/veng_0.pdf).
- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ pre-processor. *Bioinformatics* 34:i884–i890. <https://doi.org/10.1093/bioinformatics/bty560>.
- De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. Nano-Pack: visualizing and processing long-read sequencing data. *Bioinformatics* 34:2666–2669. <https://doi.org/10.1093/bioinformatics/bty149>.
- Zhang S, den Bakker HC, Li S, Chen J, Dinsmore BA, Lane C, Lauer AC, Fields PI, Deng X. 2019. SeqSero2: rapid and improved *Salmonella* serotype determination using whole-genome sequencing data. *Appl Environ Microbiol* 85:e01746-19. <https://doi.org/10.1128/AEM.01746-19>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Florensa AF, Kaas RS, Clausen PTLC, Aytan-Aktug D, Aarestrup FM. 2022. ResFinder—an open online resource for identification of antimicrobial resistance genes in next-generation sequencing data and prediction of phenotypes from genotypes. *Microb Genom* 8:000748. <https://doi.org/10.1099/mgen.0.000748>.
- Zhou Z, Alikhan NF, Mohamed K, Fan Y, Agama Study Group, Achtman M. 2020. The Enterobase user's guide, with case studies on *Salmonella* transmissions, *Yersinia pestis* phylogeny and *Escherichia core* genomic diversity. *Genome Res* 30:138–152. <https://doi.org/10.1101/gr.251678.119>.
- Zhou Z, Charlesworth J, Achtman M. 2021. HierCC: a multi-level clustering scheme for population assignments based on core genome MLST. *Bioinformatics* 37:3645–3646. <https://doi.org/10.1093/bioinformatics/btab234>.