Original Article

Mutant *IDH1* Enhances Temozolomide Sensitivity via Regulation of the ATM/CHK2 Pathway in Glioma

Lin Lin, Jinquan Cai, Zixiao Tan, Xiangqi Meng, Ruiyan Li, Yang Li, Chuanlu Jiang

Department of Neurosurgery, Second Affiliated Hospital of Harbin Medical University, Harbin, China

Purpose Isocitrate dehydrogenase 1 (*IDH1*) mutations are the most common genetic abnormalities in low-grade gliomas and secondary glioblastomas. Glioma patients with these mutations had better clinical outcomes. However, the effect of *IDH1* mutation on drug sensitivity is still under debate.

Materials and Methods IDH1-R132H mutant cells were established by lentivirus. IDH1-R132H protein expression was confirmed by western blot. The expression of ataxia telangiectasia mutated (ATM) signaling pathway and apoptosis-related proteins were detected by immunofluorescence and western blot. Temozolomide (TMZ) induced cell apoptosis was detected by flow cytometry. Tumor cell proliferation was detected by Cell Counting Kit-8. *In vivo* nude mice were used to confirm the *in vitro* roles of *IDH1* mutation.

Results We established glioma cell lines that expressed IDH1-R132H mutation stably. We found that TMZ inhibited glioma cells proliferation more significantly in *IDH1* mutant cells compared to wild type. The IC₅₀ of TMZ in IDH1-R132H mutant group was less than half that of wild-type group (p < 0.01). TMZ significantly induced more DNA damage (quantification of γ H2AX expression in *IDH1* mutation vs. wild type, p < 0.05) and apoptosis (quantification of AnnexinV⁺propidium iodide⁻ cells in *IDH1* mutation versus wild type, p < 0.01) in *IDH1* mutant gliomas compared to wild-type gliomas. The ATM-associated DNA repair signal was impaired in *IDH1* mutant cells. Inhibiting the ATM/checkpoint kinase 2DNA repair pathway further sensitized *IDH1* mutant glioma cells to chemotherapy. We found that *IDH1* mutation significantly inhibited tumor growth *in vivo* (the tumor size was analyzed statistically, p < 0.05). Moreover, we confirmed that gliomas with *IDH1* mutation were more sensitive to TMZ *in vivo* compared to wild type significantly and the results were consistent with the *in vitro* experiment.

Conclusion These results provide evidence that combination of TMZ and ATM inhibitor enhances the antitumor effect in *IDH1* mutant gliomas.

Key words Isocitrate dehydrogenase 1, Temozolomide, Glioma, Ataxia telangiectasia mutated

Introduction

Glioma is the most common malignant tumor in brain and extremely fatal. The survival time of patients is short, and the prognosis is poor. According to the World Health Organization glioma grading classification criteria, gliomas are classified into low-grade glioma (I-II) and high-grade glioma (III-IV) [1]. At present, first-line therapeutic schedule in clinic is surgery resection combined with chemotherapy and radiotherapy [2]. Temozolomide (TMZ) is a conventional chemotherapy drug for glioma clinical therapy since it readily crosses the blood-brain barrier. However, despite its clinical effect, TMZ shows limited efficacy in prolonging the survivals of patients with glioma [3]. Drug resistance is one major factor that leads to the failure of chemotherapy. It is a critical issue for the cancer therapy to eradicate drug resistance and the underlying mechanisms still need to be investigated. How to enhance the drug sensitivity of glioma to TMZ is a very important direction in the treatment of tumor.

Isocitrate dehydrogenase (IDH) including IDH1 and IDH2 are key enzymes in the citric acid cycle. The role of the enzymes is to catalyze the tranformation of isocitrate to α -ketoglutaric acid (α -KG). Research indicates that mutant *IDH1* directly converts α -KG to (R)-2HG [4]. 2HG is an oncometabolite that affects chromatin methylation, cellular differentiation and homologous recombination [5]. *IDH1* mutations occurred frequently in low-grade gliomas and in most secondary glioblastomas. Clinically, *IDH1* mutations were associated with longer overall survival [6]. Research demonstrated that most low-grade gliomas without *IDH* mutation presented a malignant phenotype that molecularly like glioblastoma [7]. Further analysis indicated that patients with

© Copyright © 2021 by the Korean Cancer Association 367 This is an Open-Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Correspondence: Chuanlu Jiang

Department of Neurosurgery, Second Affiliated Hospital of Harbin Medical University, 246 Xuefu Road, Nangang District, Harbin 150086, China Tel: 86-13904634931 Fax: 86-605089 E-mail: jcl6688@163.com

Received May 26, 2020 Accepted October 12, 2020 Published Online October 13, 2020

Co-correspondence: Yang Li

Department of Neurosurgery, Second Affiliated Hospital of Harbin Medical University, 246 Xuefu Road, Nangang District, Harbin 150086, China Tel: 86-13895470833 Fax: 86-605089 E-mail: lee9mao@126.com

IDH1 mutation and O⁶-methylguanine-DNA methyltransferase (*MGMT*) promoter methylation were more sensitive to TMZ treatment in secondary glioblastoma versus *IDH1* wild type and MGMT promoter demethylation [8]. Although the relationship between *IDH1* mutation and chemosensitivity is increasingly recognized, the molecular mechanism determining the vulnerability of *IDH1* mutation remains to debate.

As clinical first-line drugs for glioma chemotherapy, TMZ is a DNA-methylating agent that indirectly induces DNA double-strand breaks (DSB) [9]. Ataxia telangiectasia mutated (ATM) serves as a key controller of cellular responses to DSB by activating multiple signal transduction pathways. Activated ATM phosphorylates several target proteins including the protein kinases checkpoint kinase 2 (CHK) 1, CHK2, and p53. These proteins are involved in cell cycle, DNA repair, and apoptosis [10]. DNA repair signal pathway can make tumor cells survive from DNA damage caused by chemotherapy. It is an integral part of cancer treatment on account of DNA damage in chromosomal DNA, including radiotherapy and cytotoxic chemotherapy [11]. The combination of inhibitors for specific DNA repair pathways and chemotherapy drugs for DNA damage deserves further investigation.

In the research, we established glioma cell lines that expressed the IDH1-R132H and wild-type control. We found that TMZ inhibited glioma cells proliferation more significantly in IDH1 mutant cells compared to wild type. A lower dose of TMZ was effective to induce cytotoxic damage in IDH1-R132H mutant cells. The expression of γ H2AX in IDH1-R132H mutant group was higher than wild-type group suggesting that TMZ induced more DNA damage in IDH1 mutant gliomas. To detect the annexin V/propidium iodide (PI) staining by flow cytometry we demonstrated that IDH1 mutation enhanced TMZ induced apoptosis. The TMZ induced activation of ATM and its substrates CHK2 and p53 were inhibited in IDH1-R132H mutant group. Furthermore, ATM inhibitor enhanced the cytotoxic effects of chemotherapy in IDH1-R132H mutant glioma cells. Taken together, our findings demonstrate that IDH1 mutations enhance TMZ sensitivity via regulating ATM/CHK2 pathway in glioma and suggest a combination therapeutic method in clinic.

Materials and Methods

1. Tumor cell lines

Human glioma cell lines U87 and Ln229 were purchased from the Chinese Academy of Sciences Cell Bank. We cultured the glioma cells in Dulbecco's modified Eagle's medium (Corning, Corning, NY) with 10% fetal bovine serum. We added penicillin (100 U/mL) and streptomycin (100 μ g/mL) into the culture medium. Cells were maintained at 37°C in a

humidified incubator with 5% CO₂.

2. Lentivirus production and stable cell line establishment

We created the glioma cell lines (Ln229 and U87) expressing IDH1-R132H mutation or wild-type control. IDH1-R132H mutation lentivirus and wild-type lentivirus were purchased from Shanghai GeneChem (Shanghai, China). The culture medium containing virus was added to cells. After infection for 48 hours, we discard the supernatant and change fresh culture medium. Treat the cells with puromycin to screen the cells. IDH1-R132H expression was confirmed by observation of green fluorescent protein fluorescent and western blot analysis.

3. Cell proliferation assay

Cell viability was detected by the Cell Counting Kit-8 (CCK-8) assay. Briefly, cells were harvested and counted. We seeded the specific number of tumor cells in 96-well plates. Treat the cells with different concentrations of TMZ or dimethyl sulfoxide control. After a certain time, we added the CCK-8 solution into the wells. Glioma cells were detected at 450 nm of absorbance in a microplate reader.

4. Western blotting

Glioma cells were treated with lysis buffer and centrifuged at 12,000 ×g. We separated the protein by gel electrophoresis. Then we transferred the protein from the gel to the PVDF membrane. Blocking the membrane in 5% nonfat milk at room temperature for one hour. The membranes were incubated with specific antibodies. All antibodies were diluted at specific concentrations given by manufacturer including anti-IDH1-R132H (Dianova DIA-H09, Hamburg, Germany), anti-ATM (Proteintech, Rosemont, IL), anti–p-ATM, anti-CHK2, anti–p-CHK2, anti-p53, anti–p-p53, anti-H2AX, anti-Bax, anti–Bcl-2 (Cell Signaling Technology, Beverly, MA) and anti-GAPDH (Proteintech). Incubating the membranes with secondary antibody. The membranes were treated with ECL and scanned by Gel Doc 2000 (Bio-Rad, Hercules, CA).

5. Immunofluorescence assay

Immunofluorescence assay was performed as previously described [12]. Briefly, glioma cells were fixed in 4% paraformaldehyde and infiltrated by 0.1% Triton. The cells were blocked in 1% bovine serum albumin. Anti- γ H2AX antibody (1:200, Cell Signaling Technology) and secondary antibody (1:1,000, Molecular Probes, Eugene, OR) were diluted and incubated with tumor cells. The slides were observed by fluorescence microscope (Nikon, Tokyo, Japan).

6. Flow cytometry apoptosis assay

We used the flow cytometry to check cell apoptosis accor-





VOLUME 53 NUMBER 2 APRiL 2021 369



Fig. 2. Temozolomide (TMZ) induced more DNA damage in isocitrate dehydrogenase 1 (*IDH1*) mutation cells. (A, B) Immunofluorescence staining of γ H2AX in glioma cells Ln229 and U87. Scale bars=100 μ m. (C, D) Quantification of γ H2AX is shown in A and B. *p < 0.05. (E, F) γ H2AX expression in glioma cells Ln229 and U87 were detected by western blotting. GAPDH, glyceraldehyde 3-phosphate dehydrogenase.

ding to the protocol. The glioma cells were collected and washed by phosphate buffered saline. Resuspend cells in binding buffer and transfer 100 μ L to 5 mL culture tube. Add phycoerythrin annexin V and 7-amino-actinomycin D. Incubate the cells for 15 minutes at room temperature in the dark. Add 400 μ L binding buffer and analyze by flow cytometry.

7. Immunohistochemistry staining

Tissue sections were fixed with formalin and embedded with paraffin. The slices were labeled with antibodies. Immunostaining intensity and reactivity were examined by Case Viewer after scanning using digital microscope. The expression of proteins was quantified by a 4-value score criterion that consisted with intensity score (0, 1, 2, 3 for none, weak, moderate and strong) and percentage score (0, < 10%; 1, 10-40%; 2, 40-70%; 3, > 70%). The final score was the intensity values plus percentage score.

8. Animal models and in vivo tumor formation

Four to 6-week-old female nude BALB/c mice were used for subcutaneous xenograft models. Glioma cells were subcutaneously transplanted into back flanks of BALB/c nude mice. After transplanted for 4 weeks 20 mg/kg TMZ was administered intraperitoneally into mice for consecutive 7



Fig. 3. Isocitrate dehydrogenase 1 (*IDH1*) mutant enhance temozolomide (TMZ) induced apoptosis. (A) Flow cytometry apoptosis analysis of *IDH1* mutant and wild-type cells treated TMZ at different time points. (B) Statistical quantification of annexin V⁺propidium iodide (PI)⁻ cells. (C) Statistical quantification of annexin V⁺PI⁺ cells. (D) Apoptosis-related protein Bax and Bcl-2 were detected by western blotting. GAPDH, glyceraldehyde 3-phosphate dehydrogenase. (E, F) Quantification of relative expression of Bax and Bcl-2. **p < 0.01.



Fig. 4. Isocitrate dehydrogenase 1 (*IDH1*) mutation inhibits ataxia telangiectasia mutated (ATM)/checkpoint kinase 2 (CHK2) signaling triggered by temozolomide (TMZ). (A, B) The expression of ATM signaling including ATM, CHK2, p53, and their phosphorylated proteins in glioma cells treated with or without TMZ were detected by Western blotting. GAPDH, glyceraldehyde 3-phosphate dehydrogenase.

days. All nude mice were killed in the sixth week.

9. Statistical analysis

We conducted student t test to compare two groups. The difference between groups were calculated by one-way ANOVA analysis by SPSS software ver. 19.0 (SPSS Inc., Chicago, IL). Statistical results with mean \pm SEM in the experiments were calculated and conducted by GraphPad Prism software (ver. 7.0, GraphPad Software Inc., San Diego, CA). Results were performed at least three independent experiments and at least three technical replicates. p < 0.05 was statistical significance.

Results

1. *IDH1* mutation inhibited TMZ treated glioma cells proliferation

IDH1 mutant glioma cell lines were established that stably expressed IDH1-R132H in Ln229 and U87 cell lines. IDH1-R132H expression was confirmed by western blot (Fig. 1A). We treated glioma cells with or without TMZ and detected cell proliferation in different time points by CCK-8. The result demonstrated that *IDH1* mutant cells were more sensitive to TMZ versus wild type (Fig. 1B and C). Moreover, the proliferation rate of *IDH1* mutant cells was slower than that of wild-type cells especially in 72- and 96-hours' time points. The dose-response curves shifted significantly to the left in IDH1-R132H mutant group (for Ln229 cells, 50% inhibiting concentration [IC₅₀] IDH1^{WT}=399.4 μ M, IC₅₀ IDH1^{R132H}=149.2 μ M; for U87 cells, IC₅₀ IDH1^{WT}=543 μ M, IC₅₀ IDH1^{R132H}=259.7 μ M) when treated with TMZ in different concentrations (Fig. 1D and E). The IC₅₀ of TMZ in IDH1-R132H mutant group was significantly reduced than wild-type group suggesting that a lower dose of TMZ is effective inducing cytotoxic damage in IDH1-R132H mutant gliomas (Fig. 1F).

2. TMZ induced more DNA damage in IDH1 mutant glioma

We performed immunofluorescence to examine the DNA damage in glioma cells induced by TMZ. The result demonstrated that the expression of γ H2AX in IDH1-R132H mutant cells was stronger than that in the wild-type cells when treated with TMZ (Fig. 2A and B). Quantification of γ H2AX in wild-type cells and *IDH1* mutant cells treated with TMZ (Fig. 2C and D). In addition, the western blot analysis showed that the expression of γ H2AX in IDH1-R132H mutant group was more prominent than that in the wild-type group (Fig. 2E and F). Thus, TMZ induced more DNA damage in *IDH1* mutant cells compared with wild-type glioma cells.

3. IDH1 mutation enhanced TMZ induced apoptosis

We performed the annexin V/PI double staining to detect apoptosis. The result demonstrated that ATM inhibitor treatment enhanced the apoptosis induced by TMZ (Fig. 3A). Both annexinV⁺PI⁺ and annexinV⁺PI⁻ labeled cells were more abundant in IDH1-R132H mutant cells (Fig. 3B and C). We



Fig. 5. Ataxia telangiectasia mutated (ATM) inhibitor enhanced the effect of temozolomide (TMZ) on tumor apoptosis. (A) Isocitrate dehydrogenase 1 (*IDH1*) mutation and wild-type gliomas were divided into four groups treated with negative control, ATM inhibitor, TMZ and ATM inhibitor+TMZ, after 72 hours, cell apoptosis was detected by flow cytometry. (B) Statistical quantification of annexin V⁺propidium iodide (PI)⁻ cells. (C) *IDH1* mutation and wild-type gliomas were treated with negative control, ATM inhibitor, TMZ and ATM inhibitor+TMZ, respectively. Cell proliferation was detected by Cell Counting Kit-8. *p < 0.05, **p < 0.01, ***p < 0.001.

found that the expression of apoptosis-related protein Bax was higher in IDH1-R132H mutant cells when treated with TMZ. The expression of anti-apoptotic protein Bcl-2 was reduced in *IDH1* mutant cells compared to wild-type cells (Fig. 3D-F). The results demonstrated that TMZ induced more apoptosis in IDH1-R132H mutant glioma cells.

4. *IDH1* mutation inhibited ATM/CHK2 signaling triggered by TMZ

To further investigate the role of ATM signaling in TMZ

induced DNA damage, the activation of ATM and its substrates CHK2 and p53 were detected. The western blot analyses showed significant phosphorylation of ATM, CHK2, and -p53 in both glioma cells when treated with TMZ (Fig. 4A and B). We found that the expression of p-ATM was higher in wild-type cells compared with IDH1-R132H mutant groups. IDH1-R132H mutation also inhibited the activation of ATM downstream proteins including p-CHK2 and p-p53 (Fig. 4A and B). These data support that IDH1-R132H mutation inhibits ATM/CHK2/p53 signaling when treated with TMZ.



Fig. 6. The effect of isocitrate dehydrogenase 1 (*IDH1*) mutation on tumor *in vivo*. (A) The same number of *IDH1* mutant and wild-type glioma cells were injected into the subcutaneous of 4-6 weeks nude mice, and the mice were killed and photographed in the fifth week. (B) The tumor size was detected and analyzed statistically. (C) The tumor weight was detected and analyzed statistically. (D) The tumor tissue was paraffin-embedded and stained with immunohistochemistry to detect the expression of Ki-67. Scale bars=25 μ m. (E) The results of immunohistochemistry were scored and analyzed statistically. *p < 0.05, **p < 0.01, ***p < 0.001.

5. ATM inhibitor enhanced the antitumor effect of TMZ

Concerning IDH1-R132H mutation increased TMZ induced tumor damage via inhibiting ATM/CHK2/p53 signaling, we wondered if ATM inhibitor could strengthen the killing effect of TMZ. We added ATM inhibitor KU-55933 with or without TMZ to both wild-type and IDH1-R132H mutant groups. We found that ATM inhibitor combined with TMZ induced more apoptosis than TMZ alone group in IDH1-R132H mutant glioma cells. The antitumor effect of combination therapy was more significant than that of single-drug therapy in wild-type groups compared to IDH1 mutant groups. This may be *IDH1* mutations inhibited the activation of ATM more significant than wild type. Moreover, whether the ATM inhibitor was added or not, TMZ induced more apoptosis in IDH1-R132H mutant cells than wild-type cells (Fig. 5A and B). Moreover, combination of ATM inhibitor and TMZ inhibited cell proliferation more significantly in IDH1 mutant glioma cells (Fig. 5C). Taken together, ATM inhibition enhanced the antitumor effect of TMZ.

6. Effect of IDH1 mutation on tumor in vivo

IDH1 mutant cells and wild-type glioma cells were subcutaneously injected into the back flanks of mice, and the tumorigenesis and tumor size were recorded. In the 5th week after subcutaneous inoculation of tumor cells, the same number of mice were killed, and the tumor size and weight were recorded (Fig. 6A). The results showed that the size and weight of *IDH1* mutant gliomas were smaller than that of wild type (Fig. 6B and C). The results showed that *IDH1* mutation had a certain inhibitory effect on tumor growth *in vivo*. The expression of Ki67 in *IDH1* mutation group was lower than that in wild type (Fig. 6D). The results of immunohistochemistry showed that the score of wild-type Ki67 was significantly higher than that of *IDH1* mutant group (Fig. 6E). In conclusion, *IDH1* mutation can inhibit tumor growth *in vivo*.

7. The effect of TMZ on IDH1 mutant tumor in vivo

To further study the effect of TMZ on *IDH1* mutant glioma *in vivo*, we constructed subcutaneous tumor model in mice. Five weeks after tumor inoculation, TMZ was inject-



Fig. 7. The effect of temozolomide (TMZ) on isocitrate dehydrogenase 1 (*IDH1*) mutant glioma *in vivo*. (A) *IDH1* mutant and wild-type glioma cells were subcutaneously injected into 4-6 weeks BALB/c nude mice. After transplanted for 4 weeks 20 mg/kg TMZ was administered intraperitoneally into mice for consecutive 7 days. All nude mice were killed in the sixth week. (B) The tumor volume was monitored and analyzed. (C) The tumor weight was monitored and analyzed. (D) Primary glioma sections from mouse models were stained with Bax and Bcl-2 for immunohistochemistry (IHC) assay. Scale bars=25 μ m. (E, F) The results of IHC were scored and analyzed statistically. All results are expressed as the mean±standard deviation; n=5. *p < 0.05, **p < 0.01, ****p < 0.001, ****p < 0.0001.

ed intraperitoneally for 7 days at a dose of 20 mg/kg. The tumor status was monitored, and the mice were killed at week seven. The size and weight of the subcutaneous tumor were recorded and analyzed statistically (Fig. 7A). We found that *IDH1* mutant group was more sensitive to TMZ, and the tumor size and weight were significantly inhibited in this group (Fig. 7B and C). The expression of Bax was higher in *IDH1* mutation group and Bcl-2 was lower in *IDH1* mutation group (Fig. 7D and E). It is suggested that *IDH1* mutation is more sensitive to TMZ, and chemotherapy might perform more antitumor effects.

Discussion

The standard therapeutic schedule of glioblastoma is radiotherapy with daily TMZ. Though TMZ treatment prolonged the median survival time for about 2 months (from 12.1 months to 14.6 months), less than one-third of the patients survived for more than 2 years [13,14]. Genomic analysis showed that mutations of IDH1 accounted for a large proportion of gliomas. Researches demonstrate that about 60%-80% low-grade gliomas and most secondary glioblastomas have a mutation and most of those *IDH1* mutations are replacing the arginine residue with histidine (IDH1-R132H) [15,16]. Tumors with IDH gene mutation have specific genetic and clinical characteristics, our previous studies identified IDH mutation with low ATRX expression were accompanied by longer progression time in glioma. To distinguish the IDH1-R132H mutation and ATRX expression status in primary and secondary gliomas is helpful for the choice of treatment strategy [17,18].

Studies of the role of IDH mutation on drug or radiation sensitivity were controversial. One kind of studies demonstrated that introduction of mutant *IDH1* reduced TMZ resistance. *IDH1* mutation impaired PARP1-mediated DNA repair signal in glioma [19]. Inoue et al. [20] showed that IDH1 mutation in acute myeloid leukemia impaired DNA repair and reduced hematopoietic stem cells. Patients with oligodendroglioma tumors that occurred IDH mutation benefit from alkylating-agent chemotherapy with radiotherapy [21]. However, some studies also demonstrated that mutant IDH1 expression increased TMZ resistance due to enhance the RAD51-mediated homologous recombination [9]. A clinical research also suggested that IDH1 mutation conferred resistance to TMZ. They indicated that there were no differences in overall survival between the TMZ plus radiotherapy or radiotherapy group [22]. Though the prognostic value for IDH mutation in clinic has been observed, the role of IDH1 mutation on drug sensitivity is still under debate. Therefore, the molecular characteristics of IDH1 mutant cells in response to TMZ need to be further investigated and the development of more effective treatment is still an important priority for patients with glioma.

As a DNA-methylating agent, TMZ has antitumor effect in the treatment of malignant glioma. TMZ induced multiple DNA adducts, of which the O6 methylguanine was more cytotoxic [23]. O6 methylguanine mispairs with thymine and is recognized by the mismatch repair. In the process of this error repair, secondary damage is formed, which blocks DNA replication in replication cycle, resulting in DNA DSBs [24]. DSBs finally trigger apoptosis, necrosis, and autophagy. DNA damage repair involves a protein network, which can make appropriate sense and response to DNA damage, and the interference of this network can promote the occurrence of tumor. Activated ATM recruits DNA repair mechanisms, activates cell cycle checkpoints, and induces apoptosis by phosphorylating CHK2 and tumor suppressor p53 [25,26]. It is suggested that IDH1 has a TET2 independent effect on DNA repair in acute myeloid leukemia cells, and its mechanism may be related to the down-regulation of ATM-mediated by histone methylation [20]. Whether TMZ mediated DNA damage affects ATM signal pathway in *IDH1* mutation of glioma was under debate.

Our data suggest that IDH1-R132H mutant gliomas are sensitive to chemotherapy which correspond to the clinical observation in the previous research. Compared with the wild-type group, the glioma cell proliferation in the *IDH1* mutant group was significantly inhibited by TMZ. A lower dose of TMZ was effective to induce cytotoxic damage in IDH1-R132H mutant cells. The IC₅₀ of TMZ was significantly decreased in IDH1-R132H mutant gliomas, for U87 cells, IC₅₀ of *IDH1* WT cells was 543 µmol/L while IC₅₀ of IDH1-R132H cells was 259.7 µmol/L. Consistent with previous findings [19], TMZ induced more DNA damage in IDH1-R132H mutant cells due to the high expression of γ H2AX. We confirmed that *IDH1* mutations elevated cell apoptosis induced by TMZ, as Annexin V–positive cells were increased in IDH1-R132H mutant group. Interestingly, the baseline apoptosis was also elevated in mutant group. The TMZ induced activation of ATM and its substrates CHK2 and p53 were inhibited in IDH1-R132H mutation group. Due to these molecular mechanisms, *IDH1* mutant gliomas are sensitive to TMZ therapy. Furthermore, ATM inhibitor enhanced the cytotoxic effects of chemotherapy in IDH1-R132H glioma cells.

Taken together, our findings demonstrate that *IDH1* mutations enhance TMZ sensitivity via regulating ATM/CHK2 pathway in glioma. We provided the preclinical evidence that combination of TMZ and ATM inhibitor enhanced the antitumor effect in *IDH1* mutant gliomas. TMZ treatment combined with DNA repair inhibitor might be served as a prospective therapeutic strategy for glioma.

Ethical Statement

All experiments were approved by the Ethics Committee of Harbin Medical University (HMU, Harbin, Heilongjiang, China) according to the Guidelines for the Care and Use of Laboratory Animals.

Author Contributions

Conceived and designed the analysis: Lin L, Li Y, Jiang C. Collected the data: Lin L, Tan Z, Meng X. Contributed data or analysis tools: Lin L, Li R. Performed the analysis: Lin L, Cai J. Wrote the paper: Lin L.

Conflicts of Interest

Conflicts of interest relevant to this article was not reported.

Acknowledgments

This study was supported by (1) China Postdoctoral Science Foundation (2018M640305, 2019M660074); (2) Chinese Society of Neurooncology, CACA Foundation (CSNO-2016-MSD12); (3) Hei-longjiang Postdoctoral Science Foundation (LBH-Z18103); (4) Heilongjiang Health and Family Planning Commission Foundation (2017-201, 2019-102); (5) Harbin Medical University Scientific Research Innovation Fund (2017LCZX46).

References

1. Louis DN, Perry A, Reifenberger G, von Deimling A, Figarella-Branger D, Cavenee WK, et al. The 2016 World Health Organization classification of tumors of the central nervous system: a summary. Acta Neuropathol. 2016;131:803-20.

- Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. N Engl J Med. 2005;352:987-96.
- 3. Stupp R, Gander M, Leyvraz S, Newlands E. Current and future developments in the use of temozolomide for the treatment of brain tumours. Lancet Oncol. 2001;2:552-60.
- 4. Dang L, White DW, Gross S, Bennett BD, Bittinger MA, Driggers EM, et al. Cancer-associated IDH1 mutations produce 2-hydroxyglutarate. Nature. 2009;462:739-44.
- 5. Sulkowski PL, Corso CD, Robinson ND, Scanlon SE, Purshouse KR, Bai H, et al. 2-Hydroxyglutarate produced by neomorphic IDH mutations suppresses homologous recombination and induces PARP inhibitor sensitivity. Sci Transl Med. 2017;9:eaal2463.
- 6. Parsons DW, Jones S, Zhang X, Lin JC, Leary RJ, Angenendt P, et al. An integrated genomic analysis of human glioblastoma multiforme. Science. 2008;321:1807-12.
- Cancer Genome Atlas Research Network; Brat DJ, Verhaak RG, Aldape KD, Yung WK, Salama SR, et al. Comprehensive, integrative genomic analysis of diffuse lower-grade gliomas. N Engl J Med. 2015;372:2481-98.
- SongTao Q, Lei Y, Si G, YanQing D, HuiXia H, XueLin Z, et al. IDH mutations predict longer survival and response to temozolomide in secondary glioblastoma. Cancer Sci. 2012;103:269-73.
- 9. Ohba S, Mukherjee J, See WL, Pieper RO. Mutant IDH1-driven cellular transformation increases RAD51-mediated homologous recombination and temozolomide resistance. Cancer Res. 2014;74:4836-44.
- 10. Matsuoka S, Rotman G, Ogawa A, Shiloh Y, Tamai K, Elledge SJ. Ataxia telangiectasia-mutated phosphorylates Chk2 in vivo and in vitro. Proc Natl Acad Sci U S A. 2000;97:10389-94.
- 11. Helleday T, Petermann E, Lundin C, Hodgson B, Sharma RA. DNA repair pathways as targets for cancer therapy. Nat Rev Cancer. 2008;8:193-204.
- 12. Lin L, Li Y, Liu M, Li Q, Liu Q, Li R. The Interleukin-33/ST2 axis promotes glioma mesenchymal transition, stemness and TMZ resistance via JNK activation. Aging (Albany NY). 2020;12:1685-703.
- 13. Stupp R, Hegi ME, Mason WP, van den Bent MJ, Taphoorn MJ, Janzer RC, et al. Effects of radiotherapy with concomitant and adjuvant temozolomide versus radiotherapy alone on survival in glioblastoma in a randomised phase III study: 5-year

analysis of the EORTC-NCIC trial. Lancet Oncol. 2009;10:459-66.

- 14. Lin L, Cai J, Jiang C. Recent advances in targeted therapy for glioma. Curr Med Chem. 2017;24:1365-81.
- 15. Yan H, Parsons DW, Jin G, McLendon R, Rasheed BA, Yuan W, et al. IDH1 and IDH2 mutations in gliomas. N Engl J Med. 2009;360:765-73.
- 16. Horbinski C. What do we know about IDH1/2 mutations so far, and how do we use it? Acta Neuropathol. 2013;125:621-36.
- Cai J, Zhu P, Zhang C, Li Q, Wang Z, Li G, et al. Detection of ATRX and IDH1-R132H immunohistochemistry in the progression of 211 paired gliomas. Oncotarget. 2016;7:16384-95.
- Cai J, Yang P, Zhang C, Zhang W, Liu Y, Bao Z, et al. ATRX mRNA expression combined with IDH1/2 mutational status and Ki-67 expression refines the molecular classification of astrocytic tumors: evidence from the whole transcriptome sequencing of 169 samples samples. Oncotarget. 2014;5:2551-61.
- Lu Y, Kwintkiewicz J, Liu Y, Tech K, Frady LN, Su YT, et al. Chemosensitivity of IDH1-mutated gliomas due to an impairment in PARP1-mediated DNA repair. Cancer Res. 2017;77:1709-18.
- Inoue S, Li WY, Tseng A, Beerman I, Elia AJ, Bendall SC, et al. Mutant IDH1 downregulates ATM and alters DNA repair and sensitivity to DNA damage independent of TET2. Cancer Cell. 2016;30:337-48.
- Cairncross JG, Wang M, Jenkins RB, Shaw EG, Giannini C, Brachman DG, et al. Benefit from procarbazine, lomustine, and vincristine in oligodendroglial tumors is associated with mutation of IDH. J Clin Oncol. 2014;32:783-90.
- 22. Yang P, Zhang W, Wang Y, Peng X, Chen B, Qiu X, et al. IDH mutation and MGMT promoter methylation in glioblastoma: results of a prospective registry. Oncotarget. 2015;6:40896-906.
- 23. Knizhnik AV, Roos WP, Nikolova T, Quiros S, Tomaszowski KH, Christmann M, et al. Survival and death strategies in glioma cells: autophagy, senescence and apoptosis triggered by a single type of temozolomide-induced DNA damage. PLoS One. 2013;8:e55665.
- 24. Quiros S, Roos WP, Kaina B. Processing of O6-methylguanine into DNA double-strand breaks requires two rounds of replication whereas apoptosis is also induced in subsequent cell cycles. Cell Cycle. 2010;9:168-78.
- 25. Burma S, Chen BP, Murphy M, Kurimasa A, Chen DJ. ATM phosphorylates histone H2AX in response to DNA double-strand breaks. J Biol Chem. 2001;276:42462-7.
- 26. Han B, Cai J, Gao W, Meng X, Gao F, Wu P, et al. Loss of ATRX suppresses ATM dependent DNA damage repair by modulating H3K9me3 to enhance temozolomide sensitivity in glioma. Cancer Lett. 2018;419:280-90.