Research Article

Association Study of Anticitrullinated Peptide Antibody Status with Clinical Manifestations and SNPs in Patients Affected with Rheumatoid Arthritis: A Pilot Study

Argul Issilbayeva,^{1,2} Bayan Ainabekova,² Sanzhar Zhetkenev,¹ Assel Meiramova,^{1,2} Zhanar Akhmetova,^{1,2} Karlygash Karina,² Samat Kozhakhmetov,¹ Madiyar Nurgaziyev,¹,¹ Laura Chulenbayeva,¹,¹ Dimitri Poddighe,^{3,4} Jeannette Kunz,³ and Almagul Kushugulova,¹

¹Laboratory of Human Microbiome and Longevity, Center for Life Sciences, National Laboratory Astana, Nazarbayev University, Nur-Sultan, Kazakhstan

²NJSC Medical University Astana, Department of Internal Medicine with the Course of Gastroenterology,

Endocrinology and Pulmonology, Nur-Sultan, Kazakhstan

- ³Department of Medicine, Nazarbayev University School of Medicine (NUSOM), Nur-Sultan, Kazakhstan
- ⁴Department of Pediatrics, National Research Center for Mother and Child Health, University Medical Center, Nur-Sultan, Kazakhstan

Correspondence should be addressed to Argul Issilbayeva; isilbayeva.a@gmail.com

Received 24 March 2022; Accepted 25 April 2022; Published 11 May 2022

Academic Editor: Roberta Rizzo

Copyright © 2022 Argul Issilbayeva et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction. Rheumatoid arthritis (RA) is an autoimmune disease of unknown etiology that leads to disability due to articular and extra-articular damage. RA prevalence is variable. The disease is most common among females with a 3:1 ratio. The interaction of environmental and host factors contributes to RA development. Currently, the genome-wide association studies (GWAS) give the opportunity to uncover the RA genetic background. Anticitrullinated peptide antibody (ACPA) is a highly specific RA antibody, associated with poor prognosis and severe course of RA, and regulated by numerous genes. Our study is aimed at investigating whether there are any clinical and genetic aspects correlate with ACPA presence in Kazakhstani patients with RA. Indeed, the available studies on this subject are focused on Caucasian and East Asian populations (mainly Japanese and Chinese), and there are scarce data from Central Asia. Methods. Our study included 70 RA patients. Patients' blood samples were collected and genotyped for 14 SNPs by real-time polymerase chain reaction (RT-PCR). General examination, anamnestic, and clinical and laboratory data collection were carried out. Statistical analysis was performed using R statistics. Results and Conclusion. Our study revealed a significant association of ACPA positivity with Fc receptor-like 3 (FCRL3) and ACPA negativity with signal transducer and activator of transcription 4 (STAT4) genes, but not with T cell activation Rho GTPase activating protein (TAGAP). In addition, ACPA positivity was associated with radiographic progression, rheumatoid factor (RF), erythrocyte sedimentation rate (ESR), age of RA onset, the patient global assessment, body mass index (BMI), and Gamma globulin. Conclusion. Remained 11 earlier identified significantly associated in Caucasian and Asian population SNPs were not replicated in our cohort. Further studies on larger cohorts are needed to confirm our findings with higher confidence levels and stronger statistical power.

1. Introduction

Rheumatoid arthritis (RA) is an autoimmune disease of unknown etiology that leads to disability due to articular and extra-articular damage. RA prevalence is variable. [1]. The disease is most common among females, making a ratio of 3:1 [2]. The interaction of environmental and host factors contributes to RA development [3]. The genetic factor is leading one [4, 5]. Currently, the genome-wide association studies (GWAS) give the opportunity to uncover genetic background of RA. The majority of pathogenetically crucial in RA development HLA and non-HLA genes have been revealed [6-10]. HLA DRB genes certainly occupied the major niche in RA etiopathogenesis [11]. The large-scale study by Okada et al. has presented more than 100 non-HLA RA gene loci, confirming their pivotal role in the RA development [10]. Among these genes, the most key and widely studied ones can be distinguished; moreover, a specific role in the RA pathogenesis is assigned to each of them. Accordingly, PADI2 (peptidyl arginine deiminase 2) promotes citrullination of proteins resulting in the conversion of arginine into citrullines [12]. Several risk factors affect T cell activation of function. These include CD28 and CD40, which affect T cell proliferation and proinflammatory cytokine synthesis [13, 14]. The CTLA4 (cytotoxic Tlymphocyte associated protein 4) sends the inhibitory signal to T cells and provides its regulation [15], STAT4 (signal transducer and activator of transcription 4) provides phosphorylation in response to proinflammatory cytokines and activates the transcription process under the interleukin-12 control [16], TAGAP (T cell activation Rho GTPase activating protein) regulates T-cell activation effecting its cytoskeleton [17], COG6 (component of oligomeric Golgi complex 6) provides a Golgi apparatus functioning [18], TRAF1 (tumor necrosis factor receptor-associated factor-1) encoding the certain protein of TNF superfamily mediates the antiapoptotic signals coming from TNF receptors [19], ETS1 (proto-oncogene 1, transcription factor) encodes a protein responsible of a large number of gene expression leading to stem cells and tumor process development [20], FCRL3 (Fc receptor like-3) encodes a protein that is an Fc receptor-like glycoprotein capable of influencing the activation of immunoreceptor-tyrosine [21], LBH (limb bud and heart development) regulates WNT signaling pathway [22], and the roles of LINC01104 (long intergenic nonprotein coding RNA 1104) and other genes are being investigated. Further studies aimed at validating early identified SNPs in different populations demonstrated significant variabilities in genetic predisposition to RA in various ethnicities [8, 23-29].

Approximately two-thirds of RA patients produce anticitrullinated protein antibodies (ACPAs) [30–32]. These autoantibodies may be detected years before the onset of clinical symptoms, suggesting a role of autoantibodies in early RA development [33, 34]. Anti-ACPA-positive and anti-ACPA-negative RA can be regarded as two disease entities with different predisposing factors, etiology, disease severity, prognosis, and presumably pathogenesis [35, 36]. Furthermore, the presence of ACPA significantly worsens the course of RA, as the presence of these antibodies is associated with the rapid development of deformities, visceral lesions, and comorbidities and a high risk of death [37–39]. The presence of these autoantibodies is also associated with an increase in disease activity measured by DAS28, X-ray progression, ultrasound of the joints, and a lower probability of achieving disease remission [40, 41]. Several studies have shown that the citrullination of proteins and the synthesis of autoantibodies, which underlies the origin of RA on the mucous membranes, are under genetic regulation [31-33]. Despite the large number of studies focusing on the genetic landscape of RA, most studies focused on Caucasian and East Asian (mainly Japanese and Chinese) populations. At present, studies on the Central Asian population are scarce. Nevertheless, the study of genetic markers associated with RA among various ethnicities opens up the opportunity to identify ethnic-specific RA risk genes, which may lead to the development of ethnic-specific diagnostic and therapeutic approaches. Moreover, while genetic predisposition has been investigated for RA in general, no studies focus on genetic markers linked to ACPA-positive RA.

2. Materials and Methods

2.1. Patients and Controls. 70 patients, all female, with the established RA diagnosis according to ACR/EULAR 2010 rheumatoid arthritis classification criteria [41], with the disease duration more than 1 year, were recruited in Nur-Sultan. The study was approved by the local ethics committee of National Laboratory Astana, Nazarbayev University, protocol No. 03-2019. Informed consent was obtained from all patients; the study was performed in accordance with the rules and principles of the Helsinki Declaration. All patients were examined by rheumatologists; all anamnestic and examination data were recorded in the individual cards. Examination of the bone and joint system was carried out according to generally accepted rules. The count of tender joint number (TJC) and swollen joint number (SJC) was carried out. The symptoms of transverse compression of the hands and feet and the strength of compression of the hands were evaluated. The deformities and disfigurations were recorded. Data on extra-articular manifestations and complication presence were recorded. The patient global assessment, indicated by marking a 10 cm line between very good and very bad state, was performed; the overall average score was analyzed. The assessment of disease activity was carried out according to the DAS-28 disease activity index [42]. The functional disability was assessed by the health assessment questionnaire (HAQ). The X-ray stage was set according to the X-ray images of patients over the past year.

All patients underwent laboratory examination. Blood sampling was carried out strictly after a 12–14-hour period of fasting. All patients underwent general clinical research methods with the determination of indicators of the general blood test: the content of hemoglobin, erythrocytes, platelets, leukocyte formula, and erythrocyte sedimentation rate (ESR) according to Westergren and the determination of indicators of general urinalysis. Biochemical blood tests determined the levels of alanine aminotransferase (ALT), aspartate aminotransferase (AST), total protein, protein fractions, creatinine, cholesterol, glucose, and C-reactive protein (CRP). Immunological parameters such as rheumatoid factor (RF) and antibodies to cyclic citrullinated peptide (ACPA) were also determined. The blood samples were collected from all participants in compliance with infection safety measures.

2.2. SNP Analysis. Genomic DNA was extracted from the collected whole blood of RA patients and healthy subjects, to the test tubes with EDTA-ACID. The Promega Wizard genomic DNA Purification Kit was used to DNA isolation, according to manufacturer's standard protocol. All DNA was stored at -20°C. The DNA concentration in isolated samples was determined using the NanoDrop 2000/2000c spectrophotometer (Thermo Fisher) and Qubit 2.0 using the dsDNA BR Qubit analysis kit (Thermo Fisher, catalog number 32853). The samples were genotyped for 14 SNPs (PADI2-rs761426, CD28-rs1980422, CD40-rs4810485, COG6-rs9603616, CTLA4-rs3087243, ETS1-rs73013527, FCRL3-rs2317230, LBH-rs10175798, LINC01104-RASGRP1-rs8032939, STAT4-rs11889341, rs9653442, SYNGR1-rs909685, TAGAP-rs2451258, TRAF1-rs3761847) and considered the best candidates in previous GWAS [10], by real-time polymerase chain reaction (RT-PCR) using TaqMan technology according to the manufacturer's instructions (Applied Biosystems 7500, Foster City, CA).

2.3. Statistical Analysis. The R ver 4.01 was used for statistical analysis: Chi square, t-test, and SNPAssoc package. Comparison of genotype distribution and allele frequencies between RA patients and healthy controls was evaluated by the Chi-square (χ 2) test and Fisher's exact test, using an odds ratio (ORs) and 95% confidence intervals (95% CIs). Correlation of the associated SNP with autoantibody status among RA cases was performed with χ^2 test. Case and control genotype frequencies did not deviate from Hardy-Weinberg equilibrium. The comparison of the clinical and laboratory parameters with the different genotypes was performed using *t*-test and χ^2 test with Yate's correction when necessary. A p value lower than 0.05 was considered as statistically significant. The adjusted significance level was considered using standard borderline significance p value of \leq 0.05 and Bonferroni correction.

3. Results

3.1. Demographic Description of the Study Sample and Bivariate Analysis. Sample consisted of 34 ACPA-negative and 36 ACPA-positive patients (Supplementary Table 1). The mean age of the responders is 45 (39.25-50.75) years. The highest portion of the patients presents with moderate and high activity of RA. The average age of start of the disease in patients with rheumatoid arthritis is 35.11 years. The bivariate statistical tests show the significant associations of ACPA with explanatory variables X-ray stage (p value = 0.017), RF status (p value=0.00185), erythrocyte sedimentation rate (p value=0.03218), age of RA onset



FIGURE 1: Allele frequency of significant SNPs.



FIGURE 2: Genotype frequency of significant SNPs.

(p value < 0.01), visual analogous scale (p value < 0.01), BMI (p value < 0.01), and gamma globulin (p value < 0.01).

Figure 1 shows the allele distribution of statistically significant single nucleotide polymorphisms. The major alleles are as follows: G (55.71%) in FCRL3 rs2317230, C (65.71%) in STAT4 rs11889341, and T (86.43%) in TAGAP rs2451258.

In Figure 2, the genotype distribution of statistically significant single nucleotide polymorphisms is illustrated. There is a more gradual distribution of sample in FCRL3 rs2317230 polymorphism than it is in STAT4 rs11889341 and TAGAP rs2451258 polymorphisms due to better recessive homozygous genotype distribution.

3.2. SNP Characteristics and Bivariate Analysis. Out of 14 SNPs of the patients tested, there was only one statistically significant result obtained using Hardy-Weinberg equilibrium (Supplementary Table 2). This can provide us with the assumption that samples are independent in 13 of the polymorphisms observed. COG6 gene SNP rs9603616 is found to violate the independence rule according to HWE and cannot be used in further analysis.

In Supplementary Table 2, the bivariate analysis between ACPA status and all the observed polymorphisms illustrates the statistically significant association in SNPs FCRL3 rs2317230, STAT4 rs11889341, and TAGAP rs2451258. Also, it is noticeable that polymorphism STAT4 rs11889341 is shown to have statistical significance with Bonferroni correction (Figure 3).

It is possible to observe the significance (p value = 0.023549316) of G/T genotype in FCRL3 rs2317230 polymorphism with an overdominant mode of inheritance with odds ratio of 3.1 (1.14-8.48) (Table 1).



FIGURE 3: Manhattan plot of SNPs by mode of inheritance.

Regarding STAT4, the C/T-T/T genotype in dominant mode $(0.29 \ (0.11-0.79))$ and C/T genotype in overdominant mode of inheritance $(0.21 \ (0.08-0.58))$ were significantly associated with the ACPA-negative RA form (Table 2).

Tests show significance in TAGAP rs2451258 codominant (*p* value = 0.046846672), dominant (*p* value = 0.014375636), overdominant (*p* value = 0.020765277), and log-additive (*p* value = 0.027666544) modes of inheritance (Table 3). Associations in SNP TAGAP rs2451258 are spurious due to relative small sample size and cannot be used to draw solid conclusions about its associations with ACPA, due to violation of Chi-square (χ 2) test assumptions.

4. Discussion

Studies focusing on the genetic landscape of RA have contributed significantly to identifying genetic risk factors involved in disease development. Likewise, these studies also carry promise for future exploration of ethnic and racial differences in epidemiologic trends in RA, which have revealed significant variations in terms of RA incidence and prevalence worldwide. To begin to provide insight into the epidemiology of RA in Central Asia, we conducted the first pilot study to identify clinical and genetic linkages associated with ACPA positivity status in RA patients.

Consistent with data reported in previous studies on the ACPA-positive RA form, our study identified significant correlations of ACPA presence with X-ray stage, RF, ESR, age of RA onset, VAS, BMI, and Gamma globulin level and is, thus, consistent with a more severe course of the disease in our cohort of RA patients [37, 43–45]. We further found a significant association of polymorphisms in two genes, FCRL3 rs2317230 and STAT4 rs11889341, with ACPA-positive and ACPA-negative forms of RA, respectively, in Central Asian patients.

Polymorphisms of FCRL3 were previously associated with autoantibody-positive RA (RF or ACPA) in several populations and reported to independently predict radiographic progression. These findings suggest that FCRL3 is involved in both disease susceptibility and progression. Our results are partially consistent with the study by Lin et al., which identified a significant association of the FCRL3 gene with an increased RF-positive and ACPA-positive RA form in the Chinese Han population [46]. A meta-analysis of 20 previously published studies by the same authors was in accordance with the study outcome [46]. The metaanalysis also suggested that SNPs in this gene correlate with a high risk of RA development in Asian populations [46]. Partially consistent with this data, our study revealed a strong association of FCRL3 rs2317230 with the ACPApositive form of RA, irrespective of RF status. However, researchers from Japan performed a meta-analysis of GWAS, which included 670 ACPA-negative RA patients and 16,891 controls and investigated a total of 1,948,138 markers. This study, which was followed by a replication study of the top 35 single nucleotide polymorphisms (SNPs)

Disease Markers

FCRL3 rs2317230	ACPA-	%	ACPA+	%	OR	Lower	Upper	p value	AIC
Codominant									
G/G	15	44.1	10	27.8	1			0.07679	97.9
G/T	9	26.5	19	52.8	3.17	1.03	9.77		
T/T	10	29.4	7	19.4	1.05	0.3	3.68		
Dominant									
G/G	15	44.1	10	27.8	1			0.15291	98.9
G/T-T/T	19	55.9	26	72.2	2.05	0.76	5.55		
Recessive									
G/G-G/T	24	70.6	29	80.6	1			0.33037	100
T/T	10	29.4	7	19.4	0.58	0.19	1.75		
Overdominant									
G/G-T/T	25	73.5	17	47.2	1			0.02355*	95.9
G/T	9	26.5	19	52.8	3.1	1.14	8.48		
Log-additive									
0,1,2	34	48.6	36	51.4	1.11	0.6	2.06	0.72789	100.9

TABLE 1: Statistically significant SNP FCRL3 rs2317230 characteristics and bivariate statistics.

* refers to statistically significant p value < 0.05 (no adjustment). ** refers to statistically significant p value < 0.01 (no adjustment).

STAT4 rs11889341	ACPA-	%	ACPA+	%	OR	Lower	Upper	p value	AIC
Codominant									
C/C	9	26.5	20	55.6	1			0.00716	93.1
C/T	23	67.6	11	30.6	0.22	0.07	0.62		
T/T	2	5.9	5	13.9	1.12	0.18	6.93		
Dominant									
C/C	9	26.5	20	55.6	1			0.01268*	94.8
C/T-T/T	25	73.5	16	44.4	0.29	0.11	0.79		
Recessive									
C/C-C/T	32	94.1	31	86.1	1			0.25659	99.7
T/T	2	5.9	5	13.9	2.58	0.47	14.31		
Overdominant									
C/C-T/T	11	32.4	25	69.4	1			0.00169*	91.1
C/T	23	67.6	11	30.6	0.21	0.08	0.58		
Log-additive									
0.1.2	34	48.6	36	51.4	0.6	0.28	1.26	0.16957	99.1

TABLE 2: Statistically significant SNP STAT4 rs11889341 characteristics and bivariate statistics.

* refers to statistically significant p value < 0.05 (no adjustment). ** refers to statistically significant p value < 0.01 (no adjustment).

using 916 RA cases and 3,764 healthy subjects, showed an association between FCRL3 rs17727339 and the ACPAnegative form of RA [43]. This result is distinct from our finding showing an association with ACPA-positive status,

Our data on STAT4 association with RA are in accordance with several published studies. Meta-analyses by Tong et al. [47], Jiang et al. [48], and Ebrahimiyan et al. [49] demonstrated the association of STAT4 polymorphisms with RA susceptibility across major ethnic groups. However, the observed association in these studies was irrespective of ACPA or RF status, whereas our study revealed an association with ACPA positivity. This outcome is similar to studies in other ethnicities that reported STAT4 association with ACPA and RF status. For example, Egyptian researchers reported the association of STAT4 rs7574865 polymorphism with RF and ACPA positivity [50]. In addition, the STAT4 rs7574865 TT genotype in Syrian RA patients showed a potential impact on ACPA positivity [51]. Furthermore, a study performed by Ciccacci et al. that included 192 RA patients and 278 healthy individuals revealed STAT-4 association with a severe disease phenotype in terms of ACPApositive status and radiographic damage in an Italian population [52].

Interestingly, there was also no significant association of polymorphisms in the PADI2 gene with the ACPA-positive form of RA in our study, despite the known key role of

Гавle 3: Statisticall [,]	y significant	SNP	TAGAP	rs2451258	characteristics	and	bivariate	statistics
------------------------------------	---------------	-----	-------	-----------	-----------------	-----	-----------	------------

TAGAP rs2451258	ACPA-	%	ACPA+	%	OR	Lower	Upper	p value	AIC
Codominant									
T/T	22	64.7	32	88.9	1			0.04685	96.9
C/T	10	29.4	3	8.3	0.21	0.05	0.84		
C/C	2	5.9	1	2.8	0.34	0.03	4.03		
Dominant									
T/T	22	64.7	32	88.9	1			0.01437*	95
C/T-C/C	12	35.3	4	11.1	0.23	0.07	0.8		
Recessive									
T/T-C/T	32	94.1	35	97.2	1			0.51842	100.6
C/C	2	5.9	1	2.8	0.46	0.04	5.29		
Overdominant									
T/T-C/C	24	70.6	33	91.7	1			0.02076*	95.6
C/T	10	29.4	3	8.3	0.22	0.05	0.88		
Log-additive									
0,1,2	34	48.6	36	51.4	0.34	0.12	0.97	0.02766*	96.1

* refers to statistically significant p value < 0.05 (no adjustment). ** refers to statistically significant p value < 0.01 (no adjustment).

PADI2 and other PADI gene family members in the process of protein citrullination during RA development. Accordingly, a recent study by Guzmán-Guzmán et al. showed that PADI2 rs1005753 was associated with ACPA positivity and RF positivity in Mexican patients with RA [24]. Our study specifically investigating PADI2 rs761426 SNP association with ACPA-positive RA form and did not find any correlation. This result may thus indicate that various SNPs of one gene differ in their association with ACPA status in RA patients of diverse populations.

Recently, we published a study on HLA-DRB9, PADI4, and PTPN22 gene distribution in Kazakhstani RA patients, which revealed, among other data, an association of PADI4 with the ACPA-positive form of RA [29]. Similar to what was found in this study, PTPN22 did not show any association with RA, RF-positive, or ACPA-positive status [29].

The SNPs STAT4, CTLA4, CD40, LBH, ETS1, and TAGAP, identified earlier in Caucasian and Asian populations, as well as in African-Americans [8–10, 53], were not replicated in our study. This result is in part consistent with a study in a Latin American population [54].

5. Conclusion

Our study demonstrates a significant correlation of FCRL3 with the ACPA-positive form of RA, whereas STAT4 showed a significant association with the ACPA-negative form. In contrast, TAGAP SNP association with ACPA status was spurious due to violation of Chi-square (χ 2) test assumptions. Furthermore, 11 earlier identified SNPs as being significantly associated with ACPA status in Caucasian and East Asian populations were not replicated in our cohort. We recognize that the small sample size may be a major limitation of our study. A larger cohort of RA patients and an expanded number of SNPs will be needed to confirm

our findings with higher confidence levels and stronger statistical power.

Nevertheless, our current results do not fully correlate with published studies conducted on Asian and European populations, which shows the need for further research into the epidemiology of RA.

Data Availability

Data and material used during this study will be available from the corresponding author upon request.

Additional Points

Study Limitation. We recognize that the small sample size may be a major limitation of our study. A larger cohort of RA patients, control group, and an expanded number of SNPs will be needed to confirm our findings with higher confidence levels and stronger statistical power.

Conflicts of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Authors' Contributions

AK and BA designed the research and critically revised the paper. AI, AM, KK, and ZhA performed the patient recruitment and clinical and laboratory examination. AI, MN, and LCh performed the research. SZh and AI analyzed the data. ZhK, DP, and AK provided the key technical support and critically revised the paper. AI and SZh wrote the paper. JK edited the paper.

Acknowledgments

Financial support for this study was supported from the grant AP08052703 "Determination of microbiomic and genomic biomarkers of rheumatoid arthritis in Kazakhstan population" of the Ministry of Education and Science of the Republic of Kazakhstan.

Supplementary Materials

Supplementary 1. Supplementary Table 1: description of clinical characteristics and bivariate analysis.

Supplementary 2. Supplementary Table 2: SNP descriptive statistics and bivariate analysis.

References

- [1] J. S. Smolen, D. Aletaha, A. Barton et al., "Rheumatoid arthritis," *Nature Reviews Disease Primers*, vol. 4, p. 18001, 2018.
- [2] W. J. Kovacs and N. J. Olsen, "Sexual dimorphism of RA manifestations: genes, hormones and behavior," *Nature Publishing Group*, vol. 7, no. 5, pp. 307–310, 2011.
- [3] V. C. Romão and J. E. Fonseca, "Etiology and risk factors for rheumatoid arthritis: a state-of-the-art review," *Frontiers in Medicine*, vol. 8, pp. 1–20, 2021.
- [4] G. S. Firestein and I. B. McInnes, "Immunopathogenesis of rheumatoid arthritis," *Immunity*, vol. 46, no. 2, pp. 183–196, 2017.
- [5] D. van der Woude and M. A. H. M. van der Helm-van, "Update on the epidemiology, risk factors, and disease outcomes of rheumatoid arthritis," *Best Practice & Research Clinical Rheumatology*, vol. 32, no. 2, pp. 174–187, 2018.
- [6] A. Ferreiro-Iglesias, A. Montes, E. Perez-Pampin et al., "Evaluation of 12 GWAS-drawn SNPs as biomarkers of rheumatoid arthritis response to TNF inhibitors. A potential SNP association with response to etanercept," *PLoS One*, vol. 14, no. 2, pp. 1–14, 2019.
- [7] J. Kurkó, T. Besenyei, J. Laki, T. T. Glant, K. Mikecz, and Z. Szekanecz, "Genetics of rheumatoid arthritis - a comprehensive review," *Clinical Reviews in Allergy & Immunology*, vol. 45, no. 2, pp. 170–179, 2013.
- [8] R. X. Leng, D. S. Di, J. Ni et al., "Identification of new susceptibility loci associated with rheumatoid arthritis," vol. 79, no. 12, pp. 1565–1571, 2020.
- [9] S. Hayashi, T. Matsubara, K. Fukuda et al., "A genome-wide association study identifying the SNPs predictive of rapid joint destruction in patients with rheumatoid arthritis," *Biomedical Reports*, vol. 14, no. 3, pp. 1–7, 2021.
- [10] Y. Okada, D. Wu, G. Trynka et al., "Genetics of rheumatoid arthritis contributes to biology and drug discovery," *Nature*, vol. 506, no. 7488, pp. 376–381, 2014.
- [11] J. Karami, S. Aslani, A. Jamshidi, M. Garshasbi, and M. Mahmoudi, "Genetic implications in the pathogenesis of rheumatoid arthritis; an updated review," *Gene*, vol. 702, pp. 8–16, 2019.
- [12] H. Yu, C. H. Tung, K. Y. Huang, H. B. Huang, and M. C. Lu, "The essential role of peptidylarginine deiminases 2 for cytokines secretion, apoptosis, and cell adhesion in macrophage," *Apoptosis, and Cell Adhesion in Macrophage*, vol. 21, no. 16, p. 5720, 2020.

- [13] M. A. Linterman, A. E. Denton, D. P. Divekar et al., "CD28 expression is required after T cell priming for helper T cell responses and protective immunity to infection," *Elife*, vol. 3, p. e03180, 2014.
- [14] R. Sender, S. Fuchs, and R. Milo, "Revised estimates for the number of human and bacteria cells in the body," *Science*, vol. 14, no. 8, 2016.
- [15] C. Le Coz, B. E. Nolan, M. Trofa et al., "Cytotoxic Tlymphocyte-associated protein 4 haploinsufficiencyassociated inflammation can occur independently of T-cell hyperproliferation," *Frontiers in Immunology*, vol. 9, p. 1715, 2018.
- [16] R. Nishikomori, T. Usui, C. Y. Wu, A. Morinobu, J. J. O'Shea, and W. Strober, "Activated STAT4 has an essential role in Th1 differentiation and proliferation that is independent of its role in the maintenance of IL-12R β 2," *The Journal of Immunology*, vol. 169, no. 8, pp. 4388–4398, 2022.
- [17] R. Chen, E. A. Stahl, F. A. S. Kurreeman et al., "Fine mapping the _TAGAP_ risk locus in rheumatoid arthritis," *Genes & Immunity*, vol. 12, no. 4, pp. 314–318, 2011.
- [18] A. Márquez, L. Vidal-Bralo, L. Rodríguez-Rodríguez et al., "A combined large-scale meta-analysis identi fi es COG6 as a novel shared risk locus for rheumatoid arthritis and systemic lupus erythematosus," vol. 76, no. 1, pp. 286–294, 2017.
- [19] M. I. Edilova, A. A. Abdul-Sater, and T. H. Watts, "TRAF1 signaling in human health and disease," *Frontiers in Immunology*, vol. 9, pp. 1–12, 2018.
- [20] J. Dittmer, "The biology of the Ets1 proto-oncogene," *Molecular Cancer*, vol. 2, no. 1, pp. 1–21, 2003.
- [21] Y. Kochi, R. Yamada, A. Suzuki et al., "A functional variant in FCRL3, encoding Fc receptor-like 3, is associated with rheumatoid arthritis and several autoimmunities," *Nature Genetics*, vol. 37, no. 5, pp. 478–485, 2005.
- [22] A. H. Ekwall, J. W. Whitaker, D. Hammaker, W. D. Bugbee, W. Wang, and G. S. Firestein, "The rheumatoid arthritis risk gene LBH regulates growth in fibroblast-like synoviocytes," *Arthritis & Rheumatology*, vol. 67, no. 5, pp. 1193–1202, 2015.
- [23] https://www.snpedia.com/index.php/Rs11889341.
- [24] I. P. Guzma, E. Navarro-zarza, and I. A. Gutie, "PADI2 Polymorphisms Are Significantly Associated With Rheumatoid Arthritis, Autoantibodies Serologic Status and Joint Damage in Women from Southern Mexico," *Frontiers in Immunology*, vol. 12, pp. 1–11, 2021.
- [25] Y. C. Kwon, J. Lim, S. Y. Bang et al., "Genome-wide association study in a Korean population identifies six novel susceptibility loci for rheumatoid arthritis," *Annals of the Rheumatic Dis*eases, vol. 79, no. 11, pp. 1438–1445, 2020.
- [26] C. J. Baños-Hernández, J. E. Navarro-Zarza, I. Parra-Rojas et al., "PADI4 polymorphisms and the functional haplotype are associated with increased rheumatoid arthritis susceptibility: A replication study in a Southern Mexican population," *Human Immunology*, vol. 78, no. 9, pp. 553–558, 2017.
- [27] F. Li, X. Ma, L. du et al., "Identification of susceptibility SNPs in CTLA-4 and PTPN22 for scleritis in Han Chinese," *Clinical & Experimental Immunology*, vol. 197, no. 2, pp. 230–236, 2019.
- [28] M. M. Aslam, P. John, K. H. Fan et al., "Investigating the GWAS-implicated loci for rheumatoid arthritis in the Pakistani population," *Disease Markers*, vol. 2020, 9 pages, 2020.
- [29] A. Issilbayeva, A. Kushugulova, A. Meiramova et al., "Epidemiological trends of rheumatoid arthritis and PADI4,

PTPN22, and HLA-DRB9 genes distribution in the Kazakhstan Population," *Open Access Macedonian Journal of Medical Sciences*, vol. 9, no. B, pp. 747–757, 2020.

- [30] A. H. Hensvold, T. Frisell, P. K. E. Magnusson, R. Holmdahl, J. Askling, and A. I. Catrina, "How well do ACPA discriminate and predict RA in the general population: a study based on 12 590 population-representative Swedish twins," *Annals of the Rheumatic Diseases*, vol. 76, no. 1, pp. 119–125, 2017.
- [31] Y. Ishikawa and C. Terao, "The impact of cigarette smoking on risk of rheumatoid arthritis: a narrative review," *Review*, vol. 9, no. 2, p. 475, 2020.
- [32] H. Ulrich, T. Häupl, and G. R. Burmester, "The etiology of rheumatoid arthritis," *Journal of Aautoimmunity*, vol. 110, p. 102400, 2020.
- [33] D. A. Fox, "Citrullination: a specific target for the autoimmune response in rheumatoid arthritis," vol. 195, no. 1, pp. 5–7, 2015.
- [34] K. Salma, A. Nessrine, E. Krystel et al., "Rheumatoid arthritis: seropositivity versus seronegativity; a comparative crosssectional study arising from Moroccan context," *Current Rheumatology Reviews*, vol. 16, no. 2, pp. 143–148, 2020.
- [35] M. M. J. Nielen, D. van Schaardenburg, H. W. Reesink et al., "Specific autoantibodies precede the symptoms of rheumatoid arthritis: a study of serial measurements in blood donors," *Arthritis and Rheumatism*, vol. 50, no. 2, pp. 380–386, 2004.
- [36] A. Tracy, C. D. Buckley, and K. Raza, "Pre-symptomatic autoimmunity in rheumatoid arthritis: when does the disease start?," *Seminars in Immunopathology*, vol. 39, no. 4, pp. 423–435, 2017.
- [37] A. Issilbayeva, A. Meiramova, A. R. Kushugulova et al., "The clinical course of rheumatoid arthritis in Kazakhstani patients," *Open Access Macedonian Journal of Medical Sciences*, vol. 9, no. B, pp. 1352–1358, 2020.
- [38] V. F. A. M. Derksen, T. W. J. Huizinga, and D. Woude, "The role of autoantibodies in the pathophysiology of rheumatoid arthritis," *Seminars in Immunopathology*, vol. 39, no. 4, pp. 437–446, 2017.
- [39] O. Gadeholt, K. Hausotter, H. Eberle, T. Klink, and A. Pfeil, "Differing X-ray patterns in seronegative and seropositive rheumatoid arthritis," *Clinical Rheumatology*, vol. 38, no. 9, pp. 2403–2410, 2019.
- [40] J. Grosse, E. Allado, C. Roux et al., "ACPA-positive versus ACPA-negative rheumatoid arthritis: two distinct erosive disease entities on radiography and ultrasonography," *Rheumatology International*, vol. 40, no. 4, pp. 615–624, 2020.
- [41] J. Kay and K. S. Upchurch, "ACR/EULAR 2010 rheumatoid arthritis classification criteria. Rheumatology," *Rheumatology*, vol. 51, no. 6, pp. 5–9, 2012.
- [42] M. L. L. Prevoo, M. Van'T Hof, H. H. Kuper, M. A. Van Leeuwen, L. B. A. Van De Putte, and P. L. C. M. Van Riel, "Modified disease activity scores that include twenty-eight-joint counts. Development and validation in a prospective longitudinal study of patients with rheumatoid arthritis," *Arthritis & Rheumatism: Official Journal of the American College of Rheumatology*, vol. 38, no. 1, pp. 44–48, 1995.
- [43] C. Terao, K. Ohmura, Y. Kochi et al., "Anti-citrullinated peptide/protein antibody (ACPA) -negative RA shares a large proportion of susceptibility loci with ACPA-positive RA : a metaanalysis of genome-wide association study in a Japanese population," *Arthritis Research & Therapy*, vol. 17, no. 1, pp. 1–11, 2015.

- [44] D. M. Boeters, L. E. Burgers, E. H. Sasso, T. W. J. Huizinga, and A. H. M. van der Helm – van Mil, "ACPA-negative RA consists of subgroups : patients with high likelihood of achieving sustained DMARD-free remission can be identified by serological markers at disease presentation," *Arthritis Research & Therapy*, vol. 21, no. 1, pp. 1–9, 2019.
- [45] K. Bogdan, P. Dominika, and P. Rafał, "Ce pt cr//biomarkers," *Taylor & Francis*, vol. 2, pp. 1–9, 2020.
- [46] X. Lin, Y. Zhang, and Q. Chen, "FCRL3 gene polymorphisms as risk factors for rheumatoid arthritis," *Human Immunology*, vol. 77, no. 2, pp. 223–229, 2016.
- [47] G. Tong, X. Zhang, W. Tong, and Y. Liu, "Association between polymorphism in STAT4 gene and risk of rheumatoid arthritis: a meta-analysis," *Human Immunology*, vol. 74, no. 5, pp. 586–592, 2013.
- [48] X. Jiang, Z. Zhou, Y. Zhang, H. Yang, and K. Ren, "An updated meta-analysis of the signal transducer and activator of transcription 4 (STAT4) rs7574865 G/T polymorphism and rheumatoid arthritis risk in an Asian population," *Scandinavian Journal of Rheumatology*, vol. 43, no. 6, pp. 477–480, 2014.
- [49] H. Ebrahimiyan, S. Mostafaei, S. Aslani, A. Jamshidi, and M. Mahmoudi, "Studying the association between STAT4 gene polymorphism and susceptibility to rheumatoid arthritis Disease: an updated Meta-Analysis," *Iranian Journal of Immunology*, vol. 16, no. 1, pp. 71–83, 2019.
- [50] D. El-Lebedy, H. Raslan, A. Ibrahim, I. Ashmawy, S. A. El-Aziz, and A. M. Mohammed, "Association of STAT4 rs7574865 and PTPN22 rs2476601 polymorphisms with rheumatoid arthritis and non-systemically reacting antibodies in Egyptian patients," *Clinical Rheumatology*, vol. 36, no. 9, pp. 1981–1987, 2017.
- [51] I. Tarakji, W. Habbal, and F. Monem, "Association between STAT4 rs7574865 polymorphism and rheumatoid arthritis: debate unresolved," *Debate Unresolved*, vol. 12, no. 1, pp. 172–178, 2018.
- [52] C. Ciccacci, P. Conigliaro, C. Perricone et al., "Polymorphisms in STAT-4, IL-10, PSORS1C1, PTPN2 and MIR146A Genes are associated differently with prognostic factors in Italian patients affected by rheumatoid arthritis," *Clin Exp Immunol*, vol. 186, no. 2, pp. 157–163, 2016.
- [53] V. A. Laufer, H. K. Tiwari, R. J. Reynolds et al., "Genetic influences on susceptibility to rheumatoid arthritis in African-Americans," *Human Molecular Genetics*, vol. 28, no. 5, pp. 858–874, 2019.
- [54] R. A. Verdugo, M. A. Gutiérrez, and J. Suazo, "Association analysis in a Latin American population revealed ethnic differences in rheumatoid arthritis-associated SNPs in Caucasian and Asian populations," *Scientific Reports*, vol. 10, no. 1, pp. 1–7, 2020.