



Draft Genome Sequence of a New Delhi Metallo- β -Lactamase (NDM-1)-Producing *Providencia stuartii* Strain Isolated in Lima, Peru

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ABSTRACT *Providencia stuartii* is an opportunistic pathogen of the *Enterobacteriales* order. Here, we report the 4,594,658-bp draft genome sequence of a New Delhi metallo- β -lactamase (NDM-1)-producing *Providencia stuartii* strain that was isolated from an emergency patient in a private clinic in Lima, Peru.

*P*rovidencia stuartii is a motile Gram-negative bacterium of the *Morganellaceae* family that is commonly found in the human gut microbiota (1). It is an opportunistic pathogen in patients with urinary catheters and a cause of urinary tract infections (UTIs) (2). It displays intrinsic resistance to polymyxins, nitrofurans, sulfamethoxazole, tetracyclines, fluoroquinolones, and aminoglycosides (3). Outbreaks of carbapenem-resistant *P. stuartii* have been reported in intensive care units in Brazil (4), Greece (5), and Saudi Arabia (6). Here, we present the draft genome sequence of the first New Delhi metallo- β -lactamase (NDM-1)-positive *P. stuartii* isolate reported in Peru.

Isolate PS901 was obtained from a urine sample from a 76-year-old patient in the emergency service of a private clinic in Lima, Peru. The patient was diagnosed with a UTI, unspecified encephalopathy, and septicemia. Ethical approval (approval number 508-2019/DM-CCPJ) was obtained from the institutional review board at the Japanese-Peruvian Centennial Clinic (Lima, Peru). A standard urine culture was performed in blood agar and CHROMagar orientation medium (Becton, Dickinson, Heidelberg, Germany). The culture was positive after 24 h with >100,000 CFU/ml and identified as *Providencia stuartii* with the Vitek 2 Compact instrument (bioMérieux, France). Genomic DNA of the isolate was extracted using the column-based GeneJET genomic DNA purification kit (Thermo Fisher Scientific, Waltham, MA, USA) following the manufacturer's instructions. Sequencing libraries were prepared using the Nextera XT DNA library preparation kit (Illumina, Inc., San Diego, CA, USA) and sequenced on an Illumina MiSeq instrument using a v2 reagent kit, generating 1,205,030 paired-end 250-bp reads.

The European UseGalaxy server (<http://usegalaxy.eu>) (7) was used to conduct all data analyses. Raw sequencing reads were quality checked, trimmed, and *de novo* assembled using FastQC v0.72 (8), Trimmomatic v0.36 (9), and SPAdes v3.12.0 (10), respectively, resulting in a draft genome of 132 contigs larger than 200 bp (N_{50} , 193,853 bp), a total length of 4,608,029 bp with a mean coverage of 65-fold and a GC content of 41.68%. The NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v4.11 (11) predicted 4,170 protein-coding genes, 75 tRNA genes, and 36 rRNA genes. Default parameters were used for all software tools unless otherwise noted. ABRicate v0.9.8 (12)

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was used to identify antibiotic resistance and virulence genes using the NCBI Bacterial Antimicrobial Resistance Reference Gene Database (13) and the Virulence Factor Database (VFDB) (14). Fourteen antibiotic resistance genes were identified using a sequence similarity threshold of 90%, including genes producing resistance to aminoglycosides [*rmtG*, *aac(6')-Ib-cr*, and *aac(2')-Ia*], sulfonamides (*sul1* and *sul2*), fluoroquinolones (*qnrVC1*), trimethoprim (*dfrA6*), β -lactams (*bla_{OXA-1}* and *bla_{TEM-135}*), tetracyclines (*tetB*), phenicols (*catA2*, *catA3*, and *catB3*), and rifampin (*arr-3*). We identified the *cheY* (chemotaxis regulatory protein), *flhC* (flagellar biosynthesis transcription activator), *hcp-2* (type VI secretion system substrate), *kdsA* (2-dehydro-3-deoxyphosphooctonate aldolase), and *gmhA* (phosphoheptose isomerase, previously designated *lpcA*) virulence genes. The *bla_{NDM-1}* gene was identified in an 18,480-bp contig, followed by *ble_{MBL}* (a gene producing resistance to bleomycin), coinciding with previous reports (15, 16). A query against the PlasmidFinder database (17) identified plasmid replicon IncA/C2, which is commonly associated with *bla_{NDM-1}* (18).

This draft genome of *P. stuartii* reveals the dissemination of the *bla_{NDM}* gene in a nontraditional bacterial host and highlights the potential threat to public health of carbapenem resistance in Latin America.

Data availability. The draft genome assembly of *Providencia stuartii* PS901 has been submitted to NCBI under BioProject number [PRJNA627256](#), and the raw reads have been submitted to the SRA under accession number [SRR11611425](#).

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REFERENCES

1. O'Hara CM, Brenner FW, Miller JM. 2000. Classification, identification, and clinical significance of *Proteus*, *Providencia*, and *Morganella*. *Clin Microbiol Rev* 13:534–546. <https://doi.org/10.1128/cmr.13.4.534-546.2000>.
2. Libertucci J, Bassis CM, Cassone M, Gibson K, Lansing B, Mody L, Young VB, Meddings J. 2019. Bacteria detected in both urine and open wounds in nursing home residents: a pilot study. *mSphere* 4:e00463-19. <https://doi.org/10.1128/mSphere.00463-19>.
3. Stock I, Wiedemann B. 1998. Natural antibiotic susceptibility of *Providencia stuartii*, *P. rettgeri*, *P. alcalifaciens* and *P. rustigianii* strains. *J Med Microbiol* 47:629–642. <https://doi.org/10.1099/00222615-47-7-629>.
4. Zavascki AP, Carvalhaes CG, da Silva GL, Tavares Soares SP, de Alcântara LR, Elias LS, Sandri AM, Gales AC. 2012. Outbreak of carbapenem-resistant *Providencia stuartii* in an intensive care unit. *Infect Control Hosp Epidemiol* 33:627–630. <https://doi.org/10.1086/665730>.
5. Oikonomou O, Liakopoulos A, Phee LM, Betts J, Mevius D, Wareham DW. 2016. *Providencia stuartii* isolates from Greece: co-carriage of cephalosporin (*bla_{SHV-5}*, *bla_{VEB-1}*), carbapenem (*bla_{VIM-1}*), and aminoglycoside (*rmtB*) resistance determinants by a multidrug-resistant outbreak clone. *Microb Drug Resist* 22:379–386. <https://doi.org/10.1089/mdr.2015.0215>.
6. Abdallah M, Alhababi R, Alqudah N, Aldiyat B, Alharthy A. 2018. First report of carbapenem-resistant *Providencia stuartii* in Saudi Arabia. *New Microbes New Infect* 26:107–109. <https://doi.org/10.1016/j.nmni.2018.09.007>.
7. Afgan E, Baker D, Batut B, van den Beek M, Bouvier D, Cech M, Chilton J, Clements D, Coraor N, Grüning BA, Guerler A, Hillman-Jackson J, Hiltemann S, Jalili V, Rasche H, Soranzo N, Goecks J, Taylor J, Nekrutenko A, Blankenberg D. 2018. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2018 update. *Nucleic Acids Res* 46:W537–W544. <https://doi.org/10.1093/nar/gky379>.
8. Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <https://www.bioinformatics.babraham.ac.uk/projects/fastqc>.
9. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
10. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
11. Tatusova T, Dicuccio M, Badretdin A, Chetvernin V, Nawrocki P, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
12. Seemann T. 2016. ABRicate: mass screening of contigs for antibiotic resistance genes. <https://github.com/tseemann/abricate>.
13. Feldgarden M, Brover V, Haft DH, Prasad AB, Slotta DJ, Tolstoy I, Tyson GH, Zhao S, Hsu C-H, McDermott PF, Tadesse DA, Morales C, Simmons M, Tillman G, Wasilenko J, Folster JP, Klimke W. 2019. Validating the AMR-Finder tool and resistance gene database by using antimicrobial resistance genotype-phenotype correlations in a collection of isolates. *Antimicrob Agents Chemother* 63:e00483-19. <https://doi.org/10.1128/AAC.00483-19>.
14. Chen L, Yang J, Yu J, Yao Z, Sun L, Shen Y, Jin Q. 2005. VFDB: a reference database for bacterial virulence factors. *Nucleic Acids Res* 33:D325–D328. <https://doi.org/10.1093/nar/gki008>.
15. Dorette L, Nordmann P, Poirel L. 2012. Association of the emerging carbapenemase NDM-1 with a bleomycin resistance protein in *Enterobacteriaceae* and *Acinetobacter baumannii*. *Antimicrob Agents Chemother* 56:1693–1697. <https://doi.org/10.1128/AAC.05583-11>.
16. Manageiro V, Sampaio DA, Pereira P, Rodrigues P, Vieira L, Palos C, Caniça M. 2015. Draft genome sequence of the first NDM-1-producing *Providencia stuartii* strain isolated in Portugal. *Genome Announc* 3:e01077-15. <https://doi.org/10.1128/genomeA.01077-15>.
17. Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. In silico detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. *Antimicrob Agents Chemother* 58:3895–3903. <https://doi.org/10.1128/AAC.02412-14>.
18. Carattoli A, Villa L, Poirel L, Bonnín RA, Nordmann P. 2012. Evolution of IncA/C *bla_{CMY-2}*-carrying plasmids by acquisition of the *bla_{NDM-1}* carbapenemase gene. *Antimicrob Agents Chemother* 56:783–786. <https://doi.org/10.1128/AAC.05116-11>.