

Human Vδ2 versus non-Vδ2 γδ T cells in antitumor immunity

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The Vδ2 and non-Vδ2 (mainly Vδ1) subsets of human γδ T cells have distinct homing patterns and recognize different types of ligands, yet both exert potent antitumor effects. While the T-cell receptor of Vδ2 T cells primarily recognizes tumor cell-derived pyrophosphates, non-Vδ2 γδ T cells preferentially recognize stress-associated surface antigens. Here, we discuss the pros and cons of Vδ2 versus non-Vδ2 γδ T cells as tools for future immunotherapeutic interventions against cancer.

Introduction

γδ T cells are commonly considered to bridge innate and adaptive immunity as they share with cells belonging to the adaptive immune system the expression of clonally rearranged antigen receptors and with cells of the innate immune system the expression of natural killer receptors (such as Natural Killer Group 2 Member D, NKG2D) and pattern recognition receptors.^{1,2} Moreover, γδ T cells recognize antigens independently of MHC presentation/restriction. In fact, some γδ T-cell receptors (TCRs) such as human Vδ2Vγ9 act like pattern recognition receptors, hence detecting pyrophosphates derived from multiple microbes (and tumor cells) as ‘molecular patterns’.^{2,3} γδ T cells share with conventional αβ T cells many effector functions including cytotoxicity, cytokine production and regulatory activity.^{4,5} In addition, it appears that human γδ T-cell subsets can also compete with mature dendritic cells in their capacity to take up, process and present foreign antigens to CD4⁺ and CD8⁺ αβ T cells.⁶ The MHC-nonrestricted cytotoxicity of γδ T cells towards tumor cells of epithelial as well

as hematological origin has recently raised great interest.^{7–9} Human γδ T cells come in two major flavors: Vδ2 T cells account for the majority (50–95%) of circulating γδ T cells (in turn constituting only 5% of T cells in the peripheral blood), whereas γδ T cells expressing other Vδ elements (‘non-Vδ2’) are rare in the blood but appear at increased frequencies in mucosal tissues and in the skin.^{4,10,11} Although Vδ1 is the second most frequently used Vδ element, γδ T cells expressing one of the few other available Vδ gene segments have been identified. For the purpose of this article, these cells are collectively referred to as non-Vδ2 T cells.

Vδ2 T Cells: Everybody’s Darling

Vδ2 is almost exclusively paired with Vγ9 and Vδ2Vγ9 T cells recognize in a TCR-dependent fashion phosphorylated intermediates of the isoprenoid biosynthesis pathway involved in cholesterol synthesis.¹² Such molecules, collectively termed phosphoantigens, are produced by many microbes through the prokaryote-specific non-mevalonate pathway. Microbial phosphoantigens such as (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate (HMB-PP) operate as extremely potent and selective ligands for Vδ2 T cells, stimulating their activation at pico- to nanomolar concentrations.¹³ Structurally-related pyrophosphates (such as isopentenyl pyrophosphate, IPP) are also generated by mammalian cells via the mevalonate pathway. Like HMB-PP, IPP is recognized by the Vδ2 TCR, but micromolar concentrations are required for the activation of γδ T cells.¹⁴ Vδ2 T cells kill a wide variety of tumor cells including epithelial cancer cells of various origin, acute myeloid

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leukemia (AML) blasts, lymphoma cells as well as putative cancer stem cells.^{7,15–18} The stimulation of V δ 2 in vitro with natural or synthetic phosphoantigens (in the presence of interleukin-2, IL-2) triggers a rapid, massive and selective expansion of V δ 2 T cells. Due to the ease whereby V δ 2 T cells are expanded in vitro (even under good manufacturing practice conditions), the adoptive transfer of these cells to tumor patients has been performed in several studies (see below). In addition to phosphoantigens, aminobisphosphonates (N-BPs) can be used to selectively activate V δ 2 T cells. N-BPs such as zoledronic acid (ZOL) are licensed for the treatment of patients with osteoporosis as well as metastatic cancer patients, as they inhibit osteoclastic bone resorption. In addition, ZOL and related N-BPs interfere with the mevalonate pathway by inhibiting the enzymatic processing of the V δ 2-activating metabolite IPP, leading to its accumulation.¹⁹ Importantly, N-BPs enhance the susceptibility of multiple tumor cells to $\gamma\delta$ T cell-mediated lysis, following increased IPP production.^{15,16,20} As a consequence, the activation of V δ 2 T cells in vivo by therapeutic applications of N-BPs together with low doses of IL-2 has been explored as an alternative approach to the adoptive transfer of V δ 2 T cells expanded in vitro. The role of tumor- (or microbe-) derived phosphoantigens in the activation of human V δ 2 T cells is undisputed, yet also stress-induced molecules such as homologues of bacterial mismatch repair proteins (human MutS homologue 2, hMSH2) can be recognized by the V δ 2 TCR when ectopically expressed on cancer cells.^{21,22} Moreover, V δ 2 T cells can kill tumor cells independently of TCR-mediated recognition, for instance upon the activation of NKG2D by tumor cell-expressed NKG2D ligand MHC Class I-related chain A (MICA).²³ UL16-binding proteins (ULBPs) constitute a second group of human NKG2D ligands. ULBP1 has been shown to determine the susceptibility of leukemia and lymphoma cell lines to V δ 2 T cells.^{17,24} As demonstrated by antibody-mediated blocking experiments, the respective contribution of TCR- versus NKG2D-dependent activation to the killing of different tumor cell targets by V δ 2 T cells varies considerably.¹⁵ In addition to

N-BPs, tumor cell killing by V δ 2 T cells is enhanced by Toll-like receptor (TLR) agonists²⁵ and by antibody-mediated cellular cytotoxicity (ADCC) in the presence of tumor-targeting monoclonal antibodies (mAbs). Therapeutically used mAbs such as trastuzumab (anti-HER2) and rituximab (anti-CD20) enhance the cytotoxicity of CD16-expressing $\gamma\delta$ T cells by inducing ADCC.^{26,27} Superior activity, both in terms of in vitro killing and clinical efficacy, can be expected from bispecific antibodies that cross-link tumor-cell surface antigens with signaling molecules on T cells.²⁸ While bispecific antibody constructs based on CD16 or CD3 do not specifically activate or recruit $\gamma\delta$ T cells, we are currently exploring constructs targeting V γ or V δ chains, which would selectively engage $\gamma\delta$ T cells. Pre-clinical studies have demonstrated the efficacy of adoptively transferred V δ 2 T cells against various hematological and solid tumors.^{27,29,30} Based on these results, and in view of the ease whereby V δ 2 T cells are activated in vivo and expanded in vitro to large cell numbers (by N-BP or phosphoantigen stimulation), several pilot trials have explored N-BPs plus low-dose IL-2 or the adoptive transfer to cancer patients of V δ 2 T cells expanded in vitro. Favorable responses including survival benefits were observed by Dieli et al. in a Phase I study involving a small cohort of patients affected by hormone-refractory prostate cancer and receiving ZOL plus IL-2, correlating with the activation of V δ 2 T cells in vivo.³¹ In contrast, no objective clinical responses were reported in a pilot study and a prospective Phase I/II trial involving renal cell carcinoma (RCC) and melanoma patients.^{32,33} Although V δ 2 T cells are transiently activated in vivo upon the administration of ZOL, $\gamma\delta$ T cells rapidly exhaust upon repeated application of N-BPs. In a recent study, we observed a dramatic decline of peripheral blood $\gamma\delta$ T cells in osteoporotic patients who were on i.v. or oral N-BP treatment.³⁴ Even though these patients did not receive IL-2 together with N-BPs, a similar reduction was also observed in cancer patients upon repeated administrations of ZOL together with IL-2.³² Therefore, the activation of potentially tumor-reactive V δ 2 T cells in vivo by repeated N-BP plus IL-2

administrations—in the absence of other strategies—does not hold promise as an effective anticancer therapy. Alternatively, the safety and efficacy of the adoptive transfer of V δ 2 T cells expanded in vitro has been assessed in several clinical trials involving patients affected by RCC, non-small cell lung cancer and other solid tumors (see refs. 7 and 9 for recent overviews). Generally, the infusion of $\gamma\delta$ T cells expanded in vitro appears to be well tolerated, and no major adverse effects have been observed. So far, however, only limited therapeutic benefits have been reported.^{7,9,35} Interestingly, objective responses were reported in a recent Phase I/II study enrolling 11 patients with advanced RCC. In this setting, the adoptive transfer of V δ 2 T cells was combined with the administration of ZOL, perhaps accounting for transient adverse reactions but also for beneficial effects.³⁶ Intriguingly, the retrospective analysis of intratumoral $\gamma\delta$ T cells and clinicopathological features (i.e., age, gender, tumor size, stage, grade and necrosis) in a large cohort of RCC patients did not reveal any correlation between the abundance of tumor-infiltrating $\gamma\delta$ T cells (which were in the 1% range in most cases) and disease outcome.³⁷ While such data might question the role of $\gamma\delta$ T cells in RCC, they do not preclude a potential therapeutic efficacy of adoptively transferred $\gamma\delta$ T cells.³⁶

Non-V δ 2 $\gamma\delta$ T Cells: V δ 1 and Beyond

In the absence of omnipotent ligands for the selective expansion of non-V δ 2 $\gamma\delta$ T cells (comparable to phosphoantigens for V δ 2 T cells), it is a demanding task to characterize the potential function of such cells in antitumor immunity. This notwithstanding, there are clear hints for an antitumor function of non-V δ 2 $\gamma\delta$ T cells. V δ 1 T cells, which usually constitute a minor proportion of circulating $\gamma\delta$ T cells, can exert potent cytotoxic effects against blasts from patients with acute lymphoblastic leukemia (ALL) or AML,³⁸ as well as against chronic lymphocytic leukemia cells^{39,40} and primary multiple myeloma cells.⁴¹ The reactivity of V δ 1 T cells towards hematological malignancies is not limited to cytotoxicity. In fact, a proliferative response associated

with IL-4 production was reported for V δ 1 T cells in low-grade non-Hodgkin lymphoma patients.⁴² The ligands potentially recognized on leukemia/lymphoma cells by the V δ 1 TCR have not been unambiguously identified. However, in addition to TCR-dependent pathways, signals delivered via activating receptors such as NKG2D, natural cytotoxicity receptors (NCR) like NKp30, and DNAX accessory molecule-1 (DNAM-1) play a prominent role in the recognition of tumor cells by these more ambivalent T cells.^{40–42} Intriguingly, MICA is recognized not only by NKG2D but also directly via the V δ 1 TCR, thereby possibly enabling a “super-stimulation” of V δ 1 T cells by TCR plus NKG2D.⁴³ In fact, MICA is frequently expressed on the surface of AML and ALL cells.⁴⁴ ULBPs, notably ULBP3, are also expressed on tumor cells of hematological origin and trigger cytotoxicity and/or cytokine production by V δ 1 T cells.^{42,45} Together with the observation that the inducible NKp30 as well as other NCRs enable V δ 1 T cells to kill cells that are resistant to phosphoantigen-activated V δ 2 T cells,⁴⁰ it is safe to conclude that V δ 1 have a substantial capacity to attack various leukemia and lymphoma cells, and thus might carry immunotherapeutic potential, provided that efficient large scale expansion would be achievable. Recently, experimental protocols based on the mitogenic stimulation with concanavalin A or immobilized anti-CD3 mAbs have been reported to allow for a robust expansion of V δ 1 T cells (in addition to V δ 2 T cells) when total $\gamma\delta$ T cells are used as a starting cell population.^{39,46,47} Therefore, it appears we are approaching the moment when V δ 1 T cells might also be amenable for adoptive cell transfer studies.

While V δ 1 T cells seemingly have a particular affinity for leukemia and lymphoma cells, other non-V δ 2 $\gamma\delta$ T cells might be more prone to kill solid tumors. An exciting example extends the common theme of a shared role of $\gamma\delta$ T cells in infection and antitumor immunity, which has been first established for phosphoantigen-reactive V δ 2 T cells, to non-V δ 2 $\gamma\delta$ T cells. On the grounds of the previously described selective increase of non-V δ 2 $\gamma\delta$ T cells in the blood of renal allograft recipients who developed cytomegalovirus (CMV) infection after

Table 1. Activating ligands for human V δ 2 and non-V δ 2 $\gamma\delta$ T cells expressed by tumor cells: A simplified view

$\gamma\delta$ T cell subset	Ligands for		
	T-cell receptor	NKG2D	NKp30
V δ 2	IPP, hMSH2	MICA, ULBP1	
non-V δ 2:	V δ 1	unknown, MICA	ULBP3
	V δ 5	EPCR	n.d.

EPCR, endothelial protein C receptor; hMSH2, human MutS homologue 2; IPP, isopentenyl pyrophosphate; MICA, MHC Class I-related chain A; n.d., not determined; ULBP, UL16-binding protein.

transplantation,⁴⁸ Halary and coworkers discovered that these $\gamma\delta$ T cells recognize both CMV-infected cells and intestinal tumor cells.⁴⁹ Moreover, CMV-reactive V δ 2-negative $\gamma\delta$ T cells exhibited antitumor activity against colon carcinoma cells in a pre-clinical adoptive transfer model.⁵⁰ Interestingly, there is also clinical evidence for a role of V δ 2-negative $\gamma\delta$ T cells in immunosurveillance of kidney transplanted patients who are at an increased risk to develop cancer. Couzi et al. reported that an increase in V δ 2-negative $\gamma\delta$ T cells is significantly associated with a lower incidence of cancer development, but only in patients who experienced CMV infection.⁵¹ Recently, Déchanet-Merville’s group could identify the shared ligand of CMV-infected endothelial cells and epithelial tumor cells as the MHC-like endothelial protein C receptor (EPCR).⁵² EPCR is the newly minted stress-regulated molecule that is specifically recognized by V δ 5 T cells.⁵² A short summary of major activating ligands for V δ 2 and non-V δ 2 $\gamma\delta$ T cells expressed by tumor cells is provided in Table 1.

Potential of $\gamma\delta$ T Cells in Antitumor Immunity: Beyond Direct Cytotoxicity

Human $\gamma\delta$ T cells have additional capacities that are worth exploiting for immunotherapeutic purposes. As previously mentioned, activated V δ 2 T cells can take up and process antigens for subsequent (cross-)presentation to antigen-specific $\alpha\beta$ T cells.⁶ This property can also be extrapolated to tumor-associated antigens. In the tumor microenvironment, V δ 2 T cells might kill tumor cells and subsequently take up antigen by phagocytosis or trogocytosis, followed by presentation to tumor-reactive $\alpha\beta$ T cells.⁵³ The coating of tumor cells with antibodies (e.g., by therapeutic mAbs) could increase

the efficacy of this process and additionally drive the licensing of $\gamma\delta$ T cells for professional antigen presentation.⁵⁴ In view of the so far limited success of dendritic cell-based antitumor vaccination, it appears unrealistic to expect better results with V δ 2 antigen-presenting cells (APCs). Nevertheless, such an approach might be advantageous if combined to other antitumor strategies. Along these lines, recent data indicate that $\gamma\delta$ T cells play a pivotal role in determining the efficacy of anticancer chemotherapy. In several murine transplantable tumor models, anticancer drugs that induced immunogenic cell death (such as oxaliplatin or anthracyclines) triggered the local invasion of IL-17-producing $\gamma\delta$ T cells, which occurred before and was required for the subsequent invasion of tumor-reactive cytotoxic T lymphocytes.⁵⁵ Although it is presently unknown whether such a mechanism also applies to humans (and if so, which $\gamma\delta$ T-cell subset is involved), this is an important issue for the future development of combinatorial immunotherapies against cancer.⁹

Functional Plasticity of $\gamma\delta$ T Cells: Beware of the Suppressors

$\gamma\delta$ T cells enjoy a remarkable degree of functional plasticity.^{4,5} As discussed above, circulating V δ 1 T cells exert potent anti-leukemia/lymphoma effector activities. In contrast, V δ 1 T cells infiltrating breast tumors exhibit immunosuppressive functions and inhibit $\alpha\beta$ T-cell and dendritic-cell activation, thus supporting immune escape.⁵⁶ Under the influence of transforming growth factor β (TGF β), the regulatory activity associated with FOXP3 expression is also inducible in V δ 2 T cells.⁵⁷ As with CD4⁺ T cells, the local microenvironment impacts on the functional differentiation of $\gamma\delta$ T cells in the course of their activation.

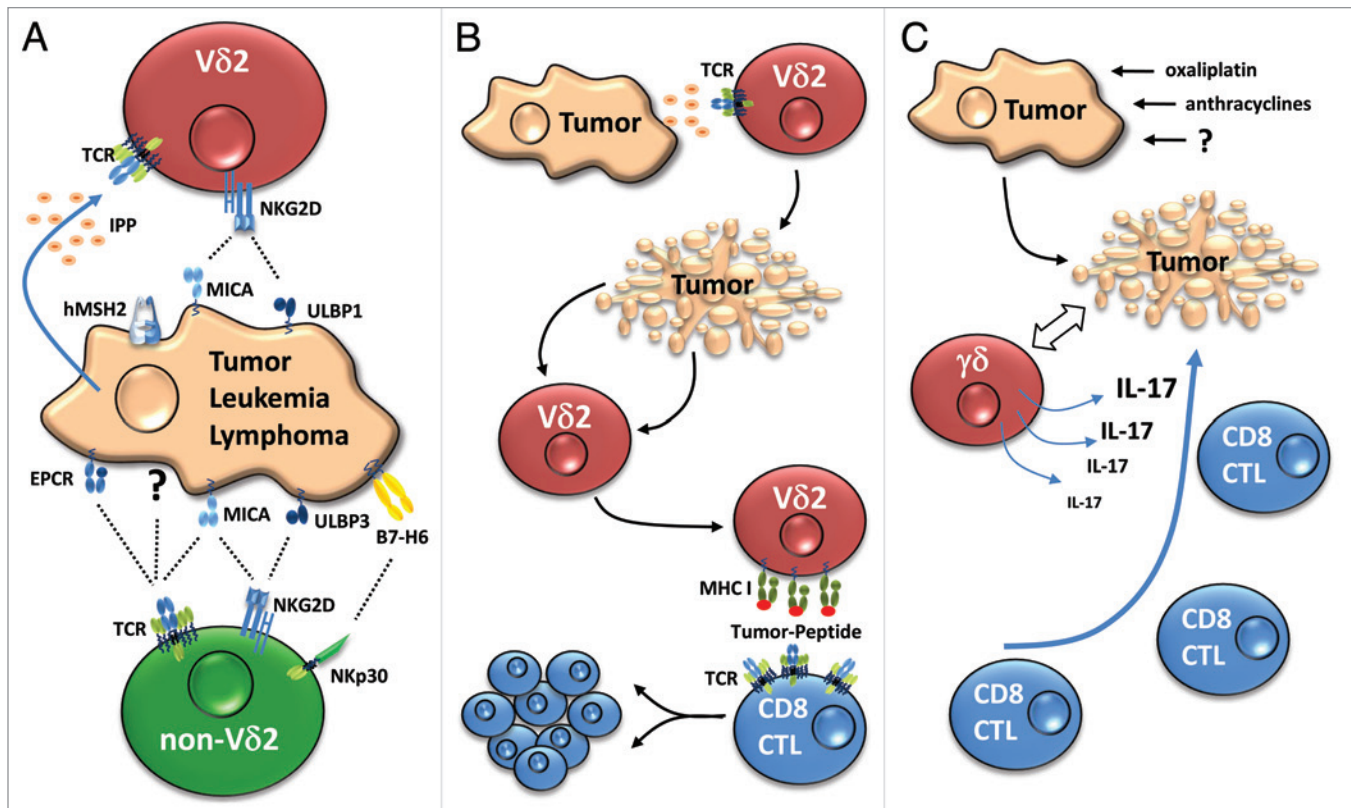


Figure 1. Possible roles of $\gamma\delta$ T cells in antitumor immunity. **(A)** Direct cytotoxic effector activity. The cytotoxic potential of V δ 2 T cells is activated following the T-cell receptor (TCR)-dependent recognition of tumor-associated phosphoantigens (e.g., isopentenyl pyrophosphate IPP) or ectopically expressed molecules, such as human MutS homologue 2 (hMSH2), as well as following the activation of NKG2D by MHC Class I-related chain A (MICA) or UL16-binding protein 1 (ULBP1). The specific ligands of non-V δ 2 TCRs have not been precisely identified, with the exception of MICA for V δ 1 and EPCR for V δ 5. NKG2D on V δ 1 $\gamma\delta$ T cells is preferentially activated by ULBP3, which is often expressed on the surface of leukemia and lymphoma cells. **(B)** Antigen-presenting function of V δ 2 T cells. Activated V δ 2 T cells kill tumor cells (top) and can engulf antigen by phagocytosis, endocytosis or trogocytosis (middle), process such antigens and subsequently present them to tumor-specific CD8⁺ cytotoxic T lymphocytes (CTLs) (bottom). **(C)** $\gamma\delta$ T cells contribute to effective chemotherapy. Certain chemotherapeutic agents induce immunogenic tumor cell death (top), activating interleukin-17 (IL-17)-secreting $\gamma\delta$ T cells (middle) that are required (at least in mice) for the subsequent recruitment and activation of tumor-specific CTLs (bottom).

Tumor-derived inhibitory cytokines such as TGF β and IL-10 are decisive factors for driving the development of regulatory $\gamma\delta$ T cells. Therefore, an important issue for the development of $\gamma\delta$ T cell-based immunotherapies, particularly adoptive cell transfer protocols, is to counteract the inhibitory differentiation pathway in $\gamma\delta$ T cells, for instance by co-stimulation with TLR agonists.⁵⁶

Concluding Remarks

$\gamma\delta$ T cells are attractive candidates for anticancer immunotherapy, mainly due to their MHC-non restricted antitumor activity. As discussed here, V δ 2 and non-V δ 2 $\gamma\delta$ T cells have a partially redundant antitumor profile. While the features and perspectives of these cell subsets have usually been investigated

independently from each other, it seems more than reasonable to exploit their combined activity, at least in certain types of cancer such as acute leukemia and multiple myeloma, two settings in which both V δ 1^{38,41} and V δ 2^{18,58} T cells have been implicated. Moreover, the APC capacity of V δ 2 T cells harbors interesting perspectives for antitumor vaccination. In this regard, it looks as if non-V δ 2 $\gamma\delta$ T cells might lose to V δ 2 T cells, but the potential APC function of non-V δ 2 $\gamma\delta$ T cells remains to be investigated. Extrapolating the fascinating results on the role of IL-17-producing $\gamma\delta$ T cells for successful chemotherapy in mice models to the human setting, it is presently unknown which one of the human $\gamma\delta$ T-cell subsets—if any—would mediate a similar functional outcome. Possible activities of human $\gamma\delta$ T-cell subsets that

can be targeted in immunotherapeutic approaches are summarized in Figure 1. Taken together, all the open questions need to be addressed when pursuing $\gamma\delta$ T cell immunotherapy, but there is no discernable reason to put V δ 2 T cells against non-V δ 2 $\gamma\delta$ T cells. Mutualism appears indeed to be an innate part of the multi-faceted nature of these cells.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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References

- Hayday AC. Gammadelta T cells and the lymphoid stress-surveillance response. *Immunity* 2009; 31:184-96; PMID:19699170; <http://dx.doi.org/10.1016/j.immuni.2009.08.006>.
- Bonneville M, O'Brien RL, Born WK. Gammadelta T cell effector functions: a blend of innate programming and acquired plasticity. *Nat Rev Immunol* 2010; 10:467-78; PMID:20539306; <http://dx.doi.org/10.1038/nri2781>.
- Morita CT, Mariuzza RA, Brenner MB. Antigen recognition by human $\gamma\delta$ T cells: pattern recognition by the adaptive immune system. *Springer Semin Immunopathol* 2000; 22:191-217; PMID:11116953; <http://dx.doi.org/10.1007/s002810000042>.
- Pang DJ, Neves JF, Sumaria N, Pennington DJ. Understanding the complexity of $\gamma\delta$ T-cell subsets in mouse and human. *Immunology* 2012; 136:283-90; PMID:22385416; <http://dx.doi.org/10.1111/j.1365-2567.2012.03582.x>.
- Kabelitz D, He W. The multifunctionality of human V γ 9V δ 2 $\gamma\delta$ T cells: clonal plasticity or distinct subsets? *Scand J Immunol* 2012; 76:213-22; PMID:22670577; <http://dx.doi.org/10.1111/j.1365-3083.2012.02727.x>.
- Meuter S, Eberl M, Moser B. Prolonged antigen survival and cytosolic export in cross-presenting human gammadelta T cells. *Proc Natl Acad Sci U S A* 2010; 107:8730-5; PMID:20413723; <http://dx.doi.org/10.1073/pnas.1002769107>.
- Braza MS, Klein B. Anti-tumour immunotherapy with V γ 9V δ 2 T lymphocytes: from the bench to the bedside. *Br J Haematol* 2013; 160:123-32; PMID:23061882; <http://dx.doi.org/10.1111/bjh.12090>.
- Siegers GM. Anti-leukemia activity of human $\gamma\delta$ T cells. *Oncoimmunology* 2012; 1:237-9; PMID:22720255; <http://dx.doi.org/10.4161/onci.1.2.18231>.
- Hannani D, Ma Y, Yamazaki T, Déchanet-Merville J, Kroemer G, Zitvogel L. Harnessing $\gamma\delta$ T cells in anticancer immunotherapy. *Trends Immunol* 2012; 33:199-206; PMID:22364810; <http://dx.doi.org/10.1016/j.it.2012.01.006>.
- Kalyan S, Kabelitz D. Defining the nature of human $\gamma\delta$ T cells: a biographical sketch of the highly empathetic. *Cell Mol Immunol* 2013; 10:21-9; PMID:23085947; <http://dx.doi.org/10.1038/cmi.2012.44>.
- Holtmeier W, Pfänder M, Hennemann A, Zollner TM, Kaufmann R, Caspary WF. The TCR- δ repertoire in normal human skin is restricted and distinct from the TCR- δ repertoire in the peripheral blood. *J Invest Dermatol* 2001; 116:275-80; PMID:11180004; <http://dx.doi.org/10.1046/j.1523-1747.2001.01250.x>.
- Wang H, Fang Z, Morita CT. Vgamma2Vdelta2 T Cell Receptor recognition of prenyl pyrophosphates is dependent on all CDRs. *J Immunol* 2010; 184:6209-22; PMID:20483784; <http://dx.doi.org/10.4049/jimmunol.1000231>.
- Hintz M, Reichenberg A, Altincicek B, Bahr U, Gschwind RM, Kollas AK, et al. Identification of (E)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate as a major activator for human gammadelta T cells in *Escherichia coli*. *FEBS Lett* 2001; 509:317-22; PMID:11741609; [http://dx.doi.org/10.1016/S0014-5793\(01\)03191-X](http://dx.doi.org/10.1016/S0014-5793(01)03191-X).
- Puan KJ, Jin C, Wang H, Sarikonda G, Raker AM, Lee HK, et al. Preferential recognition of a microbial metabolite by human Vgamma2Vdelta2 T cells. *Int Immunol* 2007; 19:657-73; PMID:17446209; <http://dx.doi.org/10.1093/intimm/dxm031>.
- Wrobel P, Shojaei H, Schitteck B, Gieseler F, Wollenberg B, Kalthoff H, et al. Lysis of a broad range of epithelial tumour cells by human $\gamma\delta$ T cells: involvement of NKG2D ligands and T-cell receptor- versus NKG2D-dependent recognition. *Scand J Immunol* 2007; 66:320-8; PMID:17635809; <http://dx.doi.org/10.1111/j.1365-3083.2007.01963.x>.
- Todaro M, D'Asaro M, Caccamo N, Iovino F, Francipane MG, Meraviglia S, et al. Efficient killing of human colon cancer stem cells by gammadelta T lymphocytes. *J Immunol* 2009; 182:7287-96; PMID:19454726; <http://dx.doi.org/10.4049/jimmunol.0804288>.
- Gomes AQ, Correia DV, Grosso AR, Lança T, Ferreira C, Lacerda JF, et al. Identification of a panel of ten cell surface protein antigens associated with immunotargeting of leukemias and lymphomas by peripheral blood gammadelta T cells. *Haematologica* 2010; 95:1397-404; PMID:20220060; <http://dx.doi.org/10.3324/haematol.2009.020602>.
- Gertner-Dardenne J, Castellano R, Mamessier E, Garbit S, Kochbati E, Etienne A, et al. Human V γ 9V δ 2 T cells specifically recognize and kill acute myeloid leukemic blasts. *J Immunol* 2012; 188:4701-8; PMID:22467661; <http://dx.doi.org/10.4049/jimmunol.1103710>.
- Gober HJ, Kistowska M, Angman L, Jenö P, Mori L, De Libero G. Human T cell receptor gammadelta cells recognize endogenous mevalonate metabolites in tumor cells. *J Exp Med* 2003; 197:163-8; PMID:12538656; <http://dx.doi.org/10.1084/jem.20021500>.
- Benzaïd I, Mönkkönen H, Stresing V, Bonnelye E, Green J, Mönkkönen J, et al. High phosphoantigen levels in bisphosphonate-treated human breast tumors promote Vgamma9Vdelta2 T-cell chemotaxis and cytotoxicity in vivo. *Cancer Res* 2011; 71:4562-72; PMID:21646473; <http://dx.doi.org/10.1158/0008-5472.CAN-10-3862>.
- Dai Y, Chen H, Mo C, Cui L, He W. Ectopically expressed human tumor biomarker MutS homologue 2 is a novel endogenous ligand that is recognized by human $\gamma\delta$ T cells to induce innate anti-tumor/virus immunity. *J Biol Chem* 2012; 287:16812-9; PMID:22433851; <http://dx.doi.org/10.1074/jbc.M111.327650>.
- Mo C, Dai Y, Kang N, Cui L, He W. Ectopic expression of human MutS homologue 2 on renal carcinoma cells is induced by oxidative stress with interleukin-18 promotion via p38 mitogen-activated protein kinase (MAPK) and c-Jun N-terminal kinase (JNK) signaling pathways. *J Biol Chem* 2012; 287:19242-54; PMID:22493490; <http://dx.doi.org/10.1074/jbc.M112.349936>.
- Rincon-Orozco B, Kunzmann V, Wrobel P, Kabelitz D, Steinle A, Herrmann T. Activation of V γ 9V δ 2 T cells by NKG2D. *J Immunol* 2005; 175:2144-51; PMID:16081780.
- Lança T, Correia DV, Moita CF, Raquel H, Neves-Costa A, Ferreira C, et al. The MHC class Ib protein ULBP1 is a nonredundant determinant of leukemia/lymphoma susceptibility to gammadelta T-cell cytotoxicity. *Blood* 2010; 115:2407-11; PMID:20101024; <http://dx.doi.org/10.1182/blood-2009-08-237123>.
- Shojaei H, Oberg HH, Juricke M, Marischen L, Kunz M, Mundhenke C, et al. Toll-like receptors 3 and 7 agonists enhance tumor cell lysis by human gammadelta T cells. *Cancer Res* 2009; 69:8710-7; PMID:19887600; <http://dx.doi.org/10.1158/0008-5472.CAN-09-1602>.
- Tokuyama H, Hagi T, Mattarollo SR, Morley J, Wang Q, So HF, et al. V γ 9V δ 2 T cell cytotoxicity against tumor cells is enhanced by monoclonal antibody drugs--rituximab and trastuzumab. *Int J Cancer* 2008; 122:2526-34; PMID:18307255; <http://dx.doi.org/10.1002/ijc.23365>.
- Capietto AH, Martinet L, Fournié JJ. Stimulated $\gamma\delta$ T cells increase the in vivo efficacy of trastuzumab in HER-2⁺ breast cancer. *J Immunol* 2011; 187:1031-8; PMID:21670311; <http://dx.doi.org/10.4049/jimmunol.1100681>.
- Bargou R, Leo E, Zugmaier G, Klinger M, Goebeler M, Knop S, et al. Tumor regression in cancer patients by very low doses of a T cell-engaging antibody. *Science* 2008; 321:974-7; PMID:18703743; <http://dx.doi.org/10.1126/science.1158545>.
- Kabelitz D, Wesch D, Pitters E, Zöllner M. Characterization of tumor reactivity of human V γ 9V δ 2 $\gamma\delta$ T cells in vitro and in SCID mice in vivo. *J Immunol* 2004; 173:6767-76; PMID:15557170.
- D'Asaro M, La Mendola C, Di Liberto D, Orlando V, Todaro M, Spina M, et al. V γ 9V δ 2 T lymphocytes efficiently recognize and kill zoledronate-sensitized, imatinib-sensitive, and imatinib-resistant chronic myelogenous leukemia cells. *J Immunol* 2010; 184:3260-8; PMID:20154204; <http://dx.doi.org/10.4049/jimmunol.0903454>.
- Dieli F, Vermijlen D, Fulfaro F, Caccamo N, Meraviglia S, Cicero G, et al. Targeting human $\gamma\delta$ T cells with zoledronate and interleukin-2 for immunotherapy of hormone-refractory prostate cancer. *Cancer Res* 2007; 67:7450-7; PMID:17671215; <http://dx.doi.org/10.1158/0008-5472.CAN-07-0199>.
- Lang JM, Kaikobad MR, Wallace M, Staab MJ, Horvath DL, Wilding G, et al. Pilot trial of interleukin-2 and zoledronic acid to augment $\gamma\delta$ T cells as treatment for patients with refractory renal cell carcinoma. *Cancer Immunol Immunother* 2011; 60:1447-60; PMID:21647691; <http://dx.doi.org/10.1007/s00262-011-1049-8>.
- Kunzmann V, Smetak M, Kimmel B, Weigang-Koehler K, Goebeler M, Birkmann J, et al. Tumor-promoting versus tumor-antagonizing roles of $\gamma\delta$ T cells in cancer immunotherapy: results from a prospective phase I/II trial. *J Immunother* 2012; 35:205-13; PMID:22306909; <http://dx.doi.org/10.1097/CJI.0b013e318245bb1e>.
- Kalyan S, Quabius ES, Wiltfang J, Mönig H, Kabelitz D. Can peripheral blood $\gamma\delta$ T cells predict osteonecrosis of the jaw? An immunological perspective on the adverse drug-effects of aminobisphosphonate therapy. *J Bone Miner Res* 2012; In Press; PMID:22991330; <http://dx.doi.org/10.1002/jbmr.1769>.
- Nicol AJ, Tokuyama H, Mattarollo SR, Hagi T, Suzuki K, Yokokawa K, et al. Clinical evaluation of autologous gamma delta T cell-based immunotherapy for metastatic solid tumours. *Br J Cancer* 2011; 105:778-86; PMID:21847128; <http://dx.doi.org/10.1038/bjc.2011.293>.
- Kobayashi H, Tanaka Y, Yagi J, Minato N, Tanabe K. Phase I/II study of adoptive transfer of $\gamma\delta$ T cells in combination with zoledronic acid and IL-2 to patients with advanced renal cell carcinoma. *Cancer Immunol Immunother* 2011; 60:1075-84; PMID:21519826; <http://dx.doi.org/10.1007/s00262-011-1021-7>.
- Inman BA, Frigola X, Harris KJ, Kuntz SM, Lohse CM, Leibovich BC, et al. Questionable relevance of $\gamma\delta$ T lymphocytes in renal cell carcinoma. *J Immunol* 2008; 180:3578-84; PMID:18292585.

38. Meeh PF, King M, O'Brien RL, Muga S, Buckhalts P, Neuberger R, et al. Characterization of the gammadelta T cell response to acute leukemia. *Cancer Immunol Immunother* 2006; 55:1072-80; PMID:16328383; <http://dx.doi.org/10.1007/s00262-005-0094-6>.
39. Siegers GM, Dhamko H, Wang XH, Mathieson AM, Kosaka Y, Felizardo TC, et al. Human Vδ1 γδ T cells expanded from peripheral blood exhibit specific cytotoxicity against B-cell chronic lymphocytic leukemia-derived cells. *Cytotherapy* 2011; 13:753-64; PMID:21314241; <http://dx.doi.org/10.3109/14653249.2011.553595>.
40. Correia DV, Fogli M, Hudspeth K, da Silva MG, Mavilio D, Silva-Santos B. Differentiation of human peripheral blood Vδ1⁺ T cells expressing the natural cytotoxicity receptor NKP30 for recognition of lymphoid leukemia cells. *Blood* 2011; 118:992-1001; PMID:21633088; <http://dx.doi.org/10.1182/blood-2011-02-339135>.
41. Knight A, Mackinnon S, Lowdell MW. Human Vdelta1 gamma-delta T cells exert potent specific cytotoxicity against primary multiple myeloma cells. *Cytotherapy* 2012; 14:1110-8; PMID:22800570; <http://dx.doi.org/10.3109/14653249.2012.700766>.
42. Catellani S, Poggi A, Bruzzone A, Dadati P, Ravetti JL, Gobbi M, et al. Expansion of Vdelta1 T lymphocytes producing IL-4 in low-grade non-Hodgkin lymphomas expressing UL-16-binding proteins. *Blood* 2007; 109:2078-85; PMID:16973957; <http://dx.doi.org/10.1182/blood-2006-06-028985>.
43. Xu B, Pizarro JC, Holmes MA, McBeth C, Groh V, Spies T, et al. Crystal structure of a gammadelta T-cell receptor specific for the human MHC class I homolog MICA. *Proc Natl Acad Sci U S A* 2011; 108:2414-9; PMID:21262824; <http://dx.doi.org/10.1073/pnas.1015433108>.
44. Salih HR, Antropius H, Gieseke F, Lutz SZ, Kanz L, Rammensee HG, et al. Functional expression and release of ligands for the activating immunoreceptor NKG2D in leukemia. *Blood* 2003; 102:1389-96; PMID:12714493; <http://dx.doi.org/10.1182/blood-2003-01-0019>.
45. Poggi A, Venturino C, Catellani S, Clavio M, Miglino M, Gobbi M, et al. Vdelta1 T lymphocytes from B-CLL patients recognize ULBP3 expressed on leukemic B cells and up-regulated by trans-retinoic acid. *Cancer Res* 2004; 64:9172-9; PMID:15604289; <http://dx.doi.org/10.1158/0008-5472.CAN-04-2417>.
46. Siegers GM, Ribot EJ, Keating A, Foster PJ. Extensive expansion of primary human gamma delta T cells generates cytotoxic effector memory cells that can be labeled with Feraheme for cellular MRI. *Cancer Immunol Immunother* 2012; In Press; PMID:23100099; <http://dx.doi.org/10.1007/s00262-012-1353-y>.
47. Dokouhaki P, Han M, Joe B, Li M, Johnston MR, Tsao MS, et al. Adoptive immunotherapy of cancer using ex vivo expanded human gammadelta T cells: A new approach. *Cancer Lett* 2010; 297:126-36; PMID:20537791; <http://dx.doi.org/10.1016/j.canlet.2010.05.005>.
48. Déchanet J, Merville P, Lim A, Retière C, Pitard V, Lafarge X, et al. Implication of gammadelta T cells in the human immune response to cytomegalovirus. *J Clin Invest* 1999; 103:1437-49; PMID:10330426; <http://dx.doi.org/10.1172/JCI5409>.
49. Halary F, Pitard V, Dlubek D, Krzysiek R, de la Salle H, Merville P, et al. Shared reactivity of Vδ2^(ns) γδ T cells against cytomegalovirus-infected cells and tumor intestinal epithelial cells. *J Exp Med* 2005; 201:1567-78; PMID:15897274; <http://dx.doi.org/10.1084/jem.20041851>.
50. Devaud C, Bilhere E, Loizon S, Pitard V, Behr C, Moreau JF, et al. Antitumor activity of gammadelta T cells reactive against cytomegalovirus-infected cells in a mouse xenograft tumor model. *Cancer Res* 2009; 69:3971-8; PMID:19383918; <http://dx.doi.org/10.1158/0008-5472.CAN-08-3037>.
51. Couzi L, Levaillant Y, Jamai A, Pitard V, Lassalle R, Martin K, et al. Cytomegalovirus-induced gammadelta T cells associate with reduced cancer risk after kidney transplantation. *J Am Soc Nephrol* 2010; 21:181-8; PMID:19713314; <http://dx.doi.org/10.1681/ASN.2008101072>.
52. Willcox CR, Pitard V, Netzer S, Couzi L, Salim M, Silberzahn T, et al. Cytomegalovirus and tumor stress surveillance by binding of a human γδ T cell antigen receptor to endothelial protein C receptor. *Nat Immunol* 2012; 13:872-9; PMID:22885985; <http://dx.doi.org/10.1038/ni.2394>.
53. Moser B, Eberl M. γδ T-APCs: a novel tool for immunotherapy? *Cell Mol Life Sci* 2011; 68:2443-52; PMID:21573785; <http://dx.doi.org/10.1007/s00018-011-0706-6>.
54. Himoudi N, Morgenstern DA, Yan M, Vernay B, Saraiva L, Wu Y, et al. Human γδ T lymphocytes are licensed for professional antigen presentation by interaction with opsonized target cells. *J Immunol* 2012; 188:1708-16; PMID:22250090; <http://dx.doi.org/10.4049/jimmunol.1102654>.
55. Ma Y, Aymeric L, Locher C, Mattarollo SR, Delahaye NF, Pereira P, et al. Contribution of IL-17-producing γ δ T cells to the efficacy of anticancer chemotherapy. *J Exp Med* 2011; 208:491-503; PMID:21383056; <http://dx.doi.org/10.1084/jem.20100269>.
56. Peng G, Wang HY, Peng W, Kiniwa Y, Seo KH, Wang RF. Tumor-infiltrating gammadelta T cells suppress T and dendritic cell function via mechanisms controlled by a unique toll-like receptor signaling pathway. *Immunity* 2007; 27:334-48; PMID:17656116; <http://dx.doi.org/10.1016/j.immuni.2007.05.020>.
57. Casetti R, Agrati C, Wallace M, Sacchi A, Martini F, Martino A, et al. Cutting edge: TGF-β1 and IL-15 Induce FOXP3+ gammadelta regulatory T cells in the presence of antigen stimulation. *J Immunol* 2009; 183:3574-7; PMID:19710458; <http://dx.doi.org/10.4049/jimmunol.0901334>.
58. Burjanadze M, Condomines M, Reme T, Quittet P, Latry P, Lugagne C, et al. *In vitro* expansion of gamma delta T cells with anti-myeloma cell activity by Phosphostim and IL-2 in patients with multiple myeloma. *Br J Haematol* 2007; 139:206-16; PMID:17897296; <http://dx.doi.org/10.1111/j.1365-2141.2007.06754.x>.
59. Li Y, Wang Q, Mariuzza RA. Structure of the human activating natural cytotoxicity receptor NKP30 bound to its tumor cell ligand B7-H6. *J Exp Med* 2011; 208:703-14; PMID:21422170; <http://dx.doi.org/10.1084/jem.20102548>.