



## Genome Sequence of Carbapenemase-Producing Enterobacter cloacae 0102-4P-1 Harboring the IncC-Type Plasmid with a Multidrug Resistance Site Encoding *bla*<sub>NDM-1</sub>, Isolated from Commercially Imported Shrimp

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**ABSTRACT** A carbapenem-resistant *Enterobacter cloacae* 0102-4P-1 strain was isolated from commercially imported shrimp in Japan. Here, we present a draft genome sequence. The complete plasmid sequence was also determined by hybrid assembly sequencing using Oxford Nanopore and Illumina methods. The assembled whole genome and plasmid were 5,164,033 bp and 162,852 bp long, respectively.

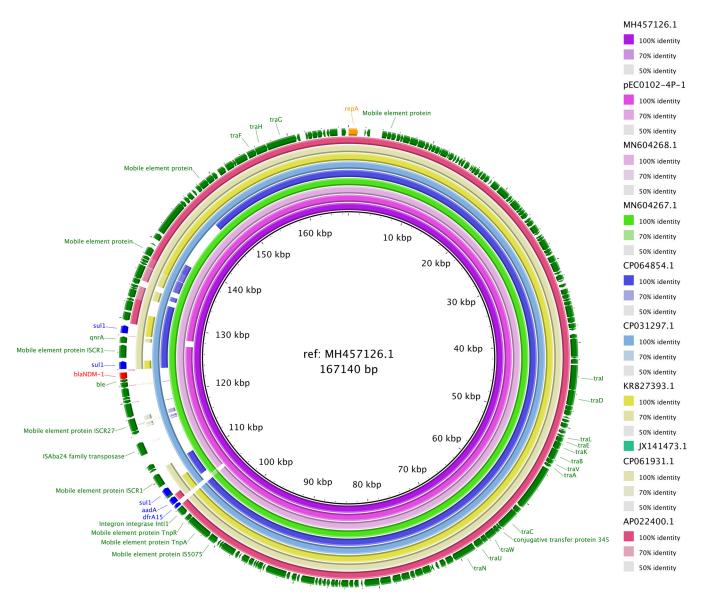
A ntibiotic-resistant bacteria that contaminate food can be transferred to humans (1). Research interest in the treatment of life-threatening infections caused by carbapenemresistant bacteria (1), which typically possess plasmid-mediated antibiotic resistance (2), is increasing. The spread of antibiotic resistance through plasmid-mediated horizontal gene transfer is an important aspect of this study.

We previously isolated carbapenem-resistant *Enterobacter cloacae* 0102-4P-1 from shrimp imported to Japan from Vietnam (3). Here, we describe a draft whole-genome sequence (WGS) and complete plasmid sequence (PS) of *E. cloacae* 0102-4P-1. In addition, a plasmid map harboring *bla*<sub>NDM-1</sub> was obtained and compared with maps of highly homologous plasmids.

TABLE 1 Genome information for	for <i>E. cloacae</i> 0102-4P-1 a	and pEC0102-4P-01
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	Data for:		
Parameter Sequencing type	<i>E. cloacae</i> 0102-4P-1 Whole genome	pEC0102-4P-01 Plasmid	
Total genome size (bp)	5,164,033	162,852	
No. of sequences	13	1	<b>Editor</b> Julie C. Dunning Hotopp, University of Maryland School of Medicine
GC content (%)	54.8	52.2	
No. of coding DNA sequences	4,920	198	Copyright © 2022 Nakayama et al. This is an
No. of rRNAs	21	0	open-access article distributed under the terms
No. of tRNAs	83	0	of the Creative Commons Attribution 4.0
Coding ratio (%)	88.8	87.8	International license.
Genotype <sup>a</sup>	Multilocus sequence typing, unknown; dnaA: 43, fusA:24, gyrB:66, leuS:247,	Plasmid Inc-type, IncC	Address correspondence to Tatsuya Nakayama t-nakayama@hiroshima-u.ac.jp.
Accession no. BPMY01000001	<i>pyrG</i> :3, <i>rpIB</i> :15, <i>rpoB</i> :3	AP024844	The authors declare no conflict of interest.
			Received 28 October 2021
Antibiotic resistance genes	dfrA16, aadA2b, sul1, bla <sub>NDM-1</sub> fosA, bla <sub>ACT-9</sub>		Accepted 31 March 2022 Published 20 April 2022

<sup>a</sup> Multi-locus sequence typing allele profile.



**FIG 1** Complete plasmid map of pEC0102-4P-01 from *E. cloacae* 0102-4P-1. The complete genomes of nine strains (GenBank accession no. MH457126, MN604268, MN604267, CP031297, CP064854, KR827393, JX141473, CP061931, and AP022400) with high homology (99.97% to 99.99%) were selected. The reference strain was MH457126. Plasmid maps were designed using BLAST Ring Image Generator v0.95. The plasmid map showed that four plasmids (MH457126, MN604268, MN604267, and CP031297) had approximately 20- to 30-kbp multidrug regions encoding antibiotic-resistance genes (ARGs), including *bla*<sub>NDM-1</sub>, mobile element proteins, and IS*Aba24* family transposase.

*E. cloacae* 0102-4P-1 was incubated at 37°C for 18 h in tryptic soy agar (Becton, Dickinson, Franklin Lakes, NJ). DNA was extracted using the DNeasy blood and tissue kit (Qiagen, Hilden, Germany) for short reads and Genomic-tip 100/G (Qiagen) for long reads. Short- and long-read sequencing libraries were prepared using Nextera DNA flex library kit (Illumina, San Diego, CA) and the rapid barcoding kit (SQK-RBK004; Oxford Nanopore Technologies, Oxford, UK), respectively. Short- and long-read sequencing were performed using the Illumina iSeq100 system with a 2 × 150-bp paired-end protocol and Oxford Nanopore MinION device with flow cell R9.4.1 (FLO-MIN106D) and MinKNOW software v20.10.3. The obtained fast5 reads were based on Guppy v4.2.3. Short-read sequences were trimmed and quality checks performed using fastp v0.20.0 (4). Short- and long-read quality checks were performed using fastqc v0.11.9 and NanoPlot v1.38.0 (5). A hybrid assembly of Illumina (total reads, 17,136,820; mean length after filtering, 2 × 143-bp reads [paired end]; total bases, 246.5 Mb; coverage, 51×) and MinION (total reads, 48,182;  $N_{50}$  value, 5,335 bp; total bases, 171.9 Mb)

sequencing data was performed using Unicycler v0.4.8 (6). The Unicycler bridge assembly graph determined that the contig 3 was completely cyclic (pEC0102-4P-1). Default parameters were used except where otherwise noted. Annotation was performed using DFAST software. The assembled WGS was analyzed for the *E. cloacae* complex using SpeciesFinder2.0, MLST2.0, and ResFinder4.1 (Table 1).

S1 nuclease pulsed-field gel electrophoresis and Southern hybridization were performed to confirm the localization of  $bla_{NDM-1}$  (7). WGS analysis showed that contig 3 encoding  $bla_{NDM}$  was the same size as the plasmid encoding  $bla_{NDM-1}$ . Hence, contig 3 was analyzed using PlasmidFinder2.1. BLAST was used to determine the PS similarity of pEC0102-4P-1. The *E. cloacae* WGS is 5,164,033 bp (Table 1). PS contig 3 (pEC0102-4P-1) is 162,852 bp. Plasmid typing was performed using an Inc-type plasmid (Table 1). To clarify the plasmid encoding  $bla_{NDM-1}$ , a complete map was designed, and the localization of  $bla_{NDM-1}$  was compared with that of a high homology plasmid (Fig. 1). Based on the composition of these genes, it is conceivable that the multidrug resistance site could spread horizontally to other plasmid harboring  $bla_{NDM-1}$  in seafood. Data from plasmid sequencing studies, such as ours, may reveal more information regarding plasmids encoding carbapenemase-related genes in seafood.

**Data availability.** The draft WGS of *E. cloacae* 0102-4P-1 and PS of pEC0102-4P-1 from *E. cloacae* 0102-4P-1 were deposited in DDBJ/GenBank (accession numbers BPMY01000001 and AP024844). The raw reads were deposited under accession numbers DRX313430 and DRX313429 with BioProject number PRJDB11927.

## ACKNOWLEDGMENT

This work was supported by the Japan Society for the Promotion of Science (JSPS) KAKENHI 18KK0168.

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