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Multiscale Agent-Based and Hybrid Modeling of the Tumor Immune Microenvironment

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Abstract

Multiscale systems biology and systems pharmacology are powerful methodologies that are playing increasingly important roles in understanding the fundamental mechanisms of biological phenomena and in clinical applications. In this review, we summarize the state of the art in the applications of agent-based models (ABM) and hybrid modeling to the tumor immune microenvironment and cancer immune response, including immunotherapy. Heterogeneity is a hallmark of cancer; tumor heterogeneity at the molecular, cellular, and tissue scales is a major determinant of metastasis, drug resistance, and low response rate to molecular targeted therapies and immunotherapies. Agent-based modeling is an effective methodology to obtain and understand quantitative characteristics of these processes and to propose clinical solutions aimed at overcoming the current obstacles in cancer treatment. We review models focusing on intratumor heterogeneity, particularly on interactions between cancer cells and stromal cells, including immune cells, the role of tumor-associated vasculature in the immune response, immune-related tumor mechanobiology, and cancer immunotherapy. We discuss the role of digital pathology in parameterizing and validating spatial computational models and potential applications to therapeutics.

Keywords

multiscale systems biology; computational biology; quantitative systems pharmacology (QSP); immuno-oncology; immunotherapy; immune checkpoint inhibitor; mathematical modeling

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1. Introduction

In recent years it has become increasingly evident that studying the tumor microenvironment (TME), in addition to studying cancer cell transformation, is crucial to understanding tumor growth, progression and dissemination. TME is a complex and heterogeneous milieu where cancer cells and stromal cells (including immune cells and other cells resident in the tissue) interact with each other and with the extracellular matrix (ECM), Figure 1. One of the critical elements of the TME is the tumor's interaction with the host immune system. Hanahan and Weinberg described evasion of the immune system as one of the hallmarks of cancer [1]. The importance of the stromal microenvironment in tumor progression was also recognized in the classical paper by Paget [2]. It has become clear that the tumor stromal component, and specifically, the host immune system, contributes to tumor growth, and new therapeutics are now being aimed at altering the immune system as a cancer target (see reviews [3,4]).

The cellular components of the TME can vary in different regions of the tumor [5], as well as between patients and even between tumors in a single patient [6]. Each of these cellular components has its own behavior in terms of migration, proliferation, differentiation, apoptosis, adhesion, and response to treatment. Cancer is so difficult to treat partly because of this degree of complexity that results in a highly unpredictable tumor behavior, partially due to the complex microenvironment [7], emphasizing the pressing need for personalized treatment for individual patients. Mathematical and computational modeling techniques provide a powerful tool in understanding the TME and predicting cancer progression [8]. Below, we provide a brief overview of the role of the immune system in cancer and introduce computational approaches to study the tumor immune microenvironment (TIME).

2. Immune System Biology and Cancer

The immune system consists of two major parts: the innate immune system, and the adaptive immune system. The innate immune system is the body's immediate defense against foreign antigens. The immunity via the innate immune system is nonspecific and short-lived, whereas the adaptive immune response is a late-stage immune response that is highly specific and can provide long-lasting defense [9]. The innate immune cells have pattern recognition receptors that recognize entities that are non-self and then induce an inflammatory response. The innate immune response is immediate, although some studies suggest these cells also have the capability for memory [10], and may be followed by an adaptive immune response [11]. The adaptive immune response is more specific than the innate immune response and can be antibody-mediated or cell-mediated, with T- and B-cells as the key cell types driving this response [12]. T-cells are a type of lymphocyte that matures in the thymus, with several different subtypes that play a distinct function in immune response [13]. Cytotoxic T-cells kill cancer cells [13], T-helper cells assist other cell types during the immune response, and T-regulatory cells (Treg) play an important role in immunological tolerance. In the case of cancer, in addition to normal antigens (self), cancer cells express antigens unique to the tumor, which can result in an immune response. The computational models discussed in this review will largely focus on the adaptive immune system.

Although the immune system is well equipped to eliminate abnormal cells, cancer cells have several ways to evade the immune system. For example, cancer cells can become invisible to the immune system by downregulating major histocompatibility complex (MHC) class-I receptors on their cell surface, and in turn, not presenting the mutation associated antigen for detection by T-cell receptors. Cancer cells attract regulatory T-cells (Treg) [14] and myeloidderived suppressor cells (MDSC) [15] to the tumors; an abundance of these cell types results in an immunosuppressive environment [16]. Furthermore, receptors such as CTLA-4 on Treg can bind to CD80 and CD86 on T-cells and antigen presenting cells (APC), which inhibits co-stimulation of these cells, and results in T-cell suppression. Treg can further inhibit the adaptive immune response by interfering with B-cell function and releasing immunosuppressive cytokines such as IL-10. In fact, under certain conditions, the immune response can contribute to tumor growth instead of inhibiting it [17]. Another way that tumors escape detection is through chronic inflammation [18]. During chronic inflammation, T-cells eventually lose their effectiveness over the course of the infection, called T-cell exhaustion [19]. T-cell exhaustion is also frequently found in the tumor microenvironment through the PD-L1/PD-1 pathway [20]. PD-1 blockage enhances T-cell and NK (Natural Killer) cell activity in tumors [21,22]. Several immune checkpoint inhibitors are currently being used in treatments of patients with cancer [23,24].

Innate immune cells, such as macrophages, neutrophils, and eosinophils, have also been shown to decrease or enhance tumor growth depending on their polarization state. Macrophages have a great deal of plasticity but are usually classified in one of two types: an M1-type that is immuno-enhancing, and an M2-type which is immuno-suppressing; though there is a spectrum of states between M1 and M2 [25]. Studies have shown that high numbers of tumor-associated macrophages (TAM) can lead to a worse clinical outcome [26]. TAM have been shown to promote tumor growth by increasing vascularization, cancer cell migration, cancer cell survival, and immuno-suppression [27]. Macrophages are recruited to hypoxic areas of the tumor [28], and aid in tumor progression [26,29]. Cancer cells recruit macrophages through Colony Stimulating Factor 1 (CSF1), and high CSF1 concentrations are correlated with poor prognoses [30]. Macrophages can be converted to TAM within the tumor by secreted factors, such as c-c chemokine receptor type-2 (CCR2) [31], which causes them to exhibit an M2-like phenotype [32]; this conversion of macrophages leads to a distinct subpopulation [33]. TAM secretion of c-c chemokine ligand type-18 (CCL18) can enable the epithelial-to-mesenchymal transition of breast cancer cells [34,35]. These TAM have also been shown to be associated with invasion, extravasation and metastasis [36–38]. Thus, M2-type macrophages are currently being targeted with therapeutics for tumor treatment [39]. Neutrophils are less abundant in tumors, but they are becoming more recognized for their duel role in the immune response to cancer [40]. Eosinophils are also commonly found within tumors [41], and have been found to enhance T-cell infiltration [42]. These cells also have a duel role in the immune response, and can promote or suppress tumor growth [43]. There is a complex interaction between cancer cells and the immune system. Thus, it is important to understand the conditions in which tumors are eliminated or enhanced by the immune system.

As is evident from the complex mechanisms of immune response and immune evasion described above, modeling the immune system is a challenging task [44]. For the specific

case of cancer, immune cells can be found within the TME, the lymphatic system and the lymph nodes, resulting in spatial complexity. Molecular and cellular components themselves are complex and have patient specific features such as unique lymphocyte antigen receptors. In addition, different functions of the immune system occur at different time scales, ranging from minutes to years. For example, intracellular signaling occurs in minutes, whereas memory cells exist on the order of years. Revealing this complexity across different scales as discussed above is very challenging or impossible to achieve in an experimental setting. Thus, computational modeling platforms provide a powerful tool to complement experimental measurements for better understanding of the immune system in cancer. In the following section, we provide an overview of the current computational modeling approaches for the study of cancer.

3. Overview of Computational Modeling Methodologies including Agent-Based Modeling

Computational modeling has provided great insight into studying intra-tumor heterogeneity [45] and the interplay between the tumor and the microenvironment [46]. Modeling has the benefit of providing a quantitative time- and cost-effective means to study the physical and chemical interactions in tumor initiation and growth. Modeling efforts complement experimental platforms by providing an understanding of clonal dynamics and microenvironmental cues over time. There are several ways to classify mathematical/ computational models in general, and cancer models in particular. One is deterministic vs. stochastic; another is continuum vs. discrete models. Deterministic models have an end state that does not change as long as the initial conditions remain the same, whereas stochastic models have randomness included, resulting in differences in end states, even with the same initial conditions. Continuum models treat cells as concentrations of cell types, whereas discrete models (such as agent-based or particle models) consider discrete cells; the cell behaviors, including interactions between cells, can be described as deterministic or stochastic. A multiscale setting (called a hybrid model, illustrated in Figure 2) can include both approaches, i.e., the discrete modeling of cells and continuous modeling of molecular species, such as oxygen, growth factors, chemokines, microRNAs, and drugs, but appropriate linking and calibration of such hybrid models should be performed. For continuum-based models, temporal ordinary differential equations (ODE) and spatiotemporal partial differential equations (PDE) have been used to model the immune response in cancer, e.g., [47-53]; here, we focus on discrete agent-based models (ABM) and hybrid models.

From another standpoint, the models that describe the immune system can be broadly categorized into top-down and bottom-up, and previous reviews have focused on computational modeling of the immune system [44,54]. The top-down approach models populations of cells, not single entities, and uses the mean behavior at the macroscopic level. ODE and PDE models are examples of this type of modeling where individual interactions are not simulated. Stochastic differential equation models are also a part of this class. On the other hand, the bottom-up approach focuses on the microscopic level. The model tracks each agent (e.g., a cell) and its interactions with the surrounding environment, and emergent

behavior arises from all the entities and their local behavior. Features such as stochastic behavior, spatial distribution, and heterogeneity of entities are inherent to bottom up models, and thus, easier to capture with this approach. Drawbacks of these models are that they require more computational power because they track individual agents and their interactions over time and space; also, there are computational limitations on the number of agents that can be considered; it is thus impossible to consider an entire organ or patient. Therefore, both approaches will need to be combined to achieve both spatial cellular and sub-cellular resolutions and whole patient pharmacokinetics and pharmacodynamics.

Agent-based models are an example of a bottom-up approach with applications in immunology and immune related diseases such as cancer [55]. An agent-based model is a discrete mathematical and computational framework that is capable of capturing emergent behavior of its interacting agents, defined as large-scale spatio-temperal patterns resulting from local spatial interactions between agents. The behavior and function of these agents are driven by the information they sense in their local environment and the rules of the agentbased model. Some of the characteristics that separate agent-based models apart from other rule-based modeling systems (in which outcomes are based on a set of rules that govern decisions) are that (1) they are spatial, (2) they incorporate agents that interact with other agents and their environment, (3) they may incorporate stochasticity, (4) they are modular, and (5) they produce emergent behavior [56]. These models allow the individual agents to adapt to their local environment (i.e., agents are adaptive instead of reactive), and take part in local interactions with other agents [57]. These result in complex aggregate behavior stemming from simple rules and emergent properties from agent interactions. Agent-based models can be lattice-based or lattice-free, depending on whether agents reside and move on a (regular or irregular) spatially discretized lattice, or have their locations and velocities represented by continuous variables, usually governed by forces in the environment. For example, in lattice-based ABM, agents are placed on a lattice structure that defines the locations of cells and their neighbors for cellular interactions. There are several model types that, although they are not explicitly characterized as agent-based, are reviewed here for completeness; those include cellular automata, Potts models, and Petri net models.

Agent-based models are particularly suitable for capturing spatially-varying events and heterogeneities [58], and for understanding the immune system's function. With this aim in mind, several investigators have developed agent-based models of diseases with involvement of the immune system. Several ABM have simulated the immune system's involvement in maintaining homeostasis and disease conditions, such as bacterial infections [59], fungal infections [60], abnormal systemic inflammatory response [61], ulceration [62], allergens [63], ischemia [64], tuberculosis [65], sepsis [66], and wound healing [67]. For cancers, such models include tumor growth and invasion [68], as well as specific cancer types such as hepatocellular carcinoma [69], breast cancer [70], melanoma [71], colorectal [72], lung cancer [73,74], and metastasis [75]. Software packages have been developed based on the ABM framework to study the immune system; these include ImmSim [76–79], Immunogrid [80,81], Simmune [82], Cycell [83], and PhysiCell [84].

Now we discuss the latest agent-based and hybrid models that investigate the effects of the immune system on cancer progression and immunotherapy, see Table 1. In order to be

included in the review, the work needs to have an immune component, a tumor component, and include ABM. We limited our focus to papers that were published within the last ten years. However, we also included some studies on diseases other than cancer where we feel that the methodology is relevant and could be applied to cancer; we also refer to a few general software tools that can be readily adapted to cancer.

We break the review into the following sections, although there may be significant overlap:

- (1) Models focusing on immune-related tumor mechanobiology
- (2) Models focusing on tumor-associated vasculature in the immune response
- (3) Models focusing on tumor-associated lymphatics and lymph nodes
- (4) Models focusing on tumor immunotherapy
- (5) Models focusing on tumor-enhancing immune cells
- (6) Models focusing on intra-tumor heterogeneity

3.1. Models Focusing on Tumor Mechanobiology

Changes in the tumor extracellular matrix (ECM) have been known to contribute to tumor progression and metastasis [85], with several computational models focusing on investigating glioma invasion [86–88], but less is known about its contribution to immune response. Computational modeling has been used to shed light on the interactions between the ECM and the immune system in cancer dynamics. A hybrid agent-based model was used to investigate the role of cellular adhesion to the ECM in tumor and immune system dynamics [89]. Frascoli et al. found that the greater the motility of the cancer cells, the more likely they will escape from immunotherapy. They also found that intermediate levels of adhesion in general led to less successful outcomes, but these results were variable.

Kather et al. used ABM to investigate the combination of adoptive cell transfer and therapy that permeabilized the fibrotic stromal component in colorectal cancer [72]. Adoptive cell transfer is a therapeutic strategy that aims to increase the number of immune cells to strengthen immuno-surveillance and counter tumor development. Kather et al. simulated various conditions of immune surveillance. In their model, T-cell killing of tumor cells occurred in a purely stochastic manner, with killing probability representing the effect of tumor specificity, immunogenicity, stimulatory and inhibitory effects all in one parameter. An immune rich environment promoted immune escape, but tumor growth slowed in a lymphocyte deprived environment. Tumor control was observed in a subgroup of tumors with less stroma and a high numbers of immune cells. They found that high levels of fibrosis and low numbers of lymphocytes reduced overall survival. Their findings were validated with data from colorectal cancer patients, where low density stroma and high lymphocyte level correlated with better overall survival. In this study, Kather et al. simulated the effect of immunotherapy by boosting the number of immune cells by 2–8 fold. Therapy was intended to enhance fibrotic stromal permeability; this was implemented by modifying the corresponding parameter by a factor of 4% to 16%. The model predicted that optimal tumor eradication requires a combination of therapeutics aiming at both activating adaptive immune system and stromal depletion.

3.2. Models Focusing on Tumor-Associated Vasculature in the Immune Response

Tumor-associated vasculature is an important aspect of the tumor-immune complex because it not only provides oxygen and nutrients for the tumor to grow, but it is also the source of tumor dissemination via circulating tumor cells (CTC), and recruitment for many immune cells, such as monocytes/macrophages and T-cells. Studies have aimed to provide a better understanding of these processes [90,91]. An ABM of Early Metastasis (ABMEM) framework was used to model the interactions between tumor cells, platelets, neutrophils, and endothelial cells [92]. Receptor binding to Mac-1 (macrophage antigen-1) by endothelial cells, platelets, or tumor cells leads to reactive oxygen species (ROS) production by neutrophils. Uppal et al. examined two types of platelet inhibition: inhibition of thromboxane, inhibition of adenosine diphosphate (ADP) receptors and inhibition of both [92]. They found that thromboxane inhibition alone resulted in the best outcome.

Alfonso et al. developed an agent-based model of immune cell-epithelial cell interactions in breast lobular epithelium [93]. The model investigated the effect of menstrual cycle length and hormone status on inflammatory response to cell turnover in breast tissue. Blood vessels were homogeneously distributed in the intra- and interlobular stroma. The model accounts for myoepithelial and luminal cells. Cellular processes (i.e., epithelial cell proliferation, cell death via effector cells, programmed cell death, removal of dead cells, immune cell motility, and inhibition of effector cells by regulatory cells) are modeled as stochastic events. Effector CD8+ cells are the only cells responsible for killing of damaged epithelial cells. Regulatory CD4+ and CD8+ cells act by inducing inactivation of effector dependent response. Chemokines from damaged epithelial cells activate the immune cells. Immune cells become ineffective when such chemokines are absent, or due to the suppression via regulatory cells. The outcome of the model identified novel prognostic information for breast cancer, such as the number of immune clusters being associated with the degree of epithelial damage.

3.3. Models Focusing on Tumor-Associated Lymphatics and Lymph Nodes

Reddy developed the first mathematical model of the lymphatic system in 1977 [94]. In recent years, various computational modeling approaches have been used to study the lymphatic vessels [95–99] and lymph nodes [100–102] in general, and in the application to infectious disease [103,104], and simulating fluid and chemokine transport in the lymphatic system as it relates to health and disease conditions [105]. Agent-based models have more recently been used to simulate various processes that occur during an adaptive immune response in a lymph node. Meyer-Hermann developed an ABM of germinal centers of the lymph node [106,107]. The authors studied B-cell germinal center reactions and how they contribute to germinal center deregulation [106]. They expanded this model to study B-cell affinity maturation in the lymph node germinal centers [108]. They found that competition for T-cell rescue and increased refractory time leads to a more robust affinity maturation.

A series of studies on modeling of T-cell behavior in the lymph node have been conducted by Bogle and Dunbar [44,109–112]. They modeled T-cell trafficking, activation, and proliferation in the lymph node paracortex using an agent-based approach. The model included chemokine and cytokine gradients. Using this lattice-based approach, they were able to model the movement and behavior of T-cells in the lymph node paracortex [112]. In

the next step, they expanded the agent-based model of the lymph node paracortex in three dimensions to include T-cells and dendritic cells (DC). The model allows simulation of a large number of T-cells at physiologic densities. The virtual lymph node can shrink or swell, depending on the dynamics of cell trafficking. The model was able to simulate T-cell activation in agreement with in-vivo observations, and provide new understanding on T-cell-DC interactions. Not all the parameters of the model were experimentally measured; thus, the model can be refined by more accurate measurement of those parameters [110]. Next, the authors built on their previous models to simulate T-cell ingress and egress, as well as chemotaxis in the lymph node, by incorporating new numerical methods. The new model allows simulation of expansion and contraction of T-cells in the lymph node paracortex during an immune response. The ability to model chemotaxis could be useful in studying other biological processes involving chemotaxis [111].

Moreau et al. constructed a virtual lymph node using agent-based modeling to study T-cell activation by synapses (long-lasting contacts) and kinapses (transient interactions) [113]. The model incorporated T-cell migration and T-cell-DC interactions. Additionally, virtual fluorescence-activated cell sorting (FACs) profiles were obtained from modeling by visualizing T-cell proliferation. This virtual lymph node model provides new opportunities for understanding the mechanisms of T-cell regulation in infection or vaccine application [113].

The ABM developed for studies of the lymphatic system thus far mainly focus on the lymph node, and in particular, T-cell processes. Folcik et al. developed the basic immune simulator (BIS), which is an agent-based platform that includes parenchymal tissue, secondary lymphoid tissue and the lymphatic/humoral circulation [114]. Using agent-based and hybrid models, lymph node dynamics are studied in the context of infectious diseases and cancer. Kim and Lee used a hybrid model to study the efficacy of preventative cancer vaccines. The model comprised two compartments for interactions of tumor and immune cells at the tissue site and in the draining lymph nodes [115]. Jacob et al. developed a three-compartment ABM that includes lymph nodes, blood vessels, and organ/tissue. The model was used to study immune response against viruses in these compartments [116]. Marino et al. developed a hybrid model where the lymph node and blood compartment were simulated using ordinary differential equations and the lung compartment was simulated using agents. They focused on the formation of granulomas in the lung, which are organized structures of immune cells in the lung, and are a hallmark of infection. The model focused on the recruitment of APCs in the lymph node from the lung for Mycobacterium tuberculosis (Mtb, the causative bacterium of TB) infection [117]. In another study, they investigated the role of DC in Mtb infection [118]. The growth and dissemination of bacteria were highly affected by CD8+ and CD4+ T-cell proliferation rates and DC migration. Such multiscale models allow the study of tissue level dynamics during adaptive immune response [118], and although they focus on infectious disease, many of the components and processes involved in anti-cancer immunity and adaptive immunity against infection are shared. For example, Tcells specific to tumor antigens are primed and expanded in a similar fashion to that in which T-cells specific to foreign antigens are during their response to infection; the immune suppressive mechanisms that cancer cells hijack to evade immune surveillance are also deployed during an immune response against infection to prevent excessive tissue damage.

Since the body reacts similarly in response to an infection as it does in response to cancer (e.g., activation of similar signaling pathways), cancer models can heavily borrow from this literature.

3.4. Models Focusing on Tumor Immunotherapy

A variety of cancer immunotherapy strategies exist that range from boosting the overall immune response to specifically targeting cancer immunity. Some examples of immunotherapies are treatment vaccines, adoptive cell transfer, and immune checkpoint inhibitor treatments. Agent-based and hybrid models are developed to help understand these therapies when applied separately or in combination with other cancer treatments. One type of therapy that has been explored is cancer vaccines. Therapeutic cancer vaccines treat existing cancers by delivering immunogenic and tumor specific antigens to the patient to induce cellular and/or humoral anti-tumor immunity. Pennisi et al. have developed several hybrid models investigating the immune system effects on tumors. They developed a hybrid model to study the development of lung metastases from mammary carcinoma [75]. Pennisi et al. also developed a hybrid model MetastaSim to simulate the protection against lung metastases in mouse using Triplex cell vaccine [73]. In this simulation, macrophages could capture tumor-associated antigen and immunocomplexes, breaking them down and eliminating them from the system. This vaccine elicited a combination of three stimuli: the p185neu antigen expressed by the HER2/neu gene, allogeneic major histocompatibility complex (MHC) molecules, and IL-12 which enhances antigen presentation. Using this model, after calibration and validation, the authors were able to evaluate different protocols of vaccine administration. The simulation results suggested that in order to maximize protection while reducing the number of administrations, the vaccination strategy should include a significant dosage early on and a few recalls afterwards.

Dreau et al. developed an ABM model of solid tumor progression to understand the interplay between solid tumor growth, tumor vascular growth, and the host's immune system [119]. The model includes tumor and immune cells, vasculature, tumor cell proliferation, and immune system response. Their model supported immunotherapy as an effective cancer treatment in individuals with functioning immune systems. They concluded that a strong immune response limits tumor growth in a way that cannot be achieved under a weaker immune response. Another study focused on the role of T-cells in the effectiveness of response to immunotherapy in B-16 melanoma [120]. The model includes macrophages, DC, tumor vasculature, and interactions between these components. It was found that early entry of T-cells effectively eliminated the tumor and was dependent on CD137 (a costimulatory protein that helps in tumor rejection [121]) expression in tumor vasculature.

Oncolytic virus therapy is a strategy that utilizes viral infection to kill cancer cells, but not normal cells, with the potential of enhancing T-cell recruitment to the tumor and increasing their access to cancer cells. Several computational models have examined the conditions of success for this type of therapeutic in silico [122]. Walker et al. developed an agent-based model of pancreatic tumors to study the synergy between chimeric antigen receptor (CAR) T-cell therapy and oncolytic virus therapy [123]. CAR T-cell therapy is one type of adoptive cell transfer treatment involving genetically engineered T-cells specifically targeting cancer

cells, and has been the subject of several computational models [124]. The agent-based model recapitulates treatment mechanisms including cancer specific CAR T-cell recruitment to the tumor site via vasculature and the injection and spread of oncolytic virus. Rohrs et al. demonstrated the ability of the model to track the dynamics of cancer cells and stromal cells in space in the presence of the treatment combinations; optimization of the combination therapy requires more accurate calibration [124].

Immune checkpoint inhibitors are used in cancer immunotherapy that enhances anti-tumor immune response by targeting cancer immune evasion mechanisms. In many cancer types, tumor neoantigens are sufficiently immunogenic to promote the expansion of antitumor immune cells [125]; however, these immune cells are not functional due to the inhibitory signals from molecules adaptively induced during cancer development [24,126]. Among them, one of the most prominent mechanisms is PD-1/PD-L1 interaction, where T-cells are suppressed through PD-1 signaling upon contact with induced PD-L1 in the tumor microenvironment. Gong et al. developed an ABM of tumor-immune interaction in 3D to study the spatio-temporal dynamics of cancer cells and cytotoxic T-cells [127]. In this study, the inhibitor to the checkpoint molecules were modeled as a factor which modulates the parameter governing the suppression of tumor specific T-cells by PD-L1+ cancer cells. They found that patient responsiveness to such therapy could be associated with the level of mutational burden of the cancer and antigen strength among patients. They also found that tumor growth is insensitive to the vascular density of the tumor core. From these results, a scoring method was proposed to predict anti-PDL1 treatment efficacy in patients.

3.5. Models Focusing on Tumor-Enhancing Immune Cells

While the immune system has evolved to kill off tumor cells, there are many ways in which cancer cells can avoid immune detection. In addition, there is mounting evidence that immune cells can stimulate tumor growth under certain conditions. Several agent-based models have focused on understanding the tumor-enhancing contributions of the immune system. Enderling and colleagues explored the interactions between tumor cell death and the immune system using a cellular automata model focused on the interplay between cancer stem cells and the immune system [128]. They showed that immune system-induced tumor cell death led to stem cell selection, and thus, more aggressive tumors [128,129]. In this model, even though immune cells effectively killed off tumor cells, they also affected progenitor cells. This resulted in the creation of a space for cancer stem cells to proliferate and produce more cancer stem cells. This ultimately resulted in a larger stem cell population and a more aggressive tumor.

Several studies have specifically focused on the tumor-enhancing contribution of immune cells, such as tumor-associated macrophages (TAM). Macrophages are one of the most abundant immune cells found in tumors, but their population is heterogeneous [130]. M1-type macrophages have been shown to be tumor inhibiting, whereas M2-type macrophages have been shown to be tumor enhancing [26]. One model looked at the transition from the M1 to M2 macrophage phenotype on tumor growth and then predicted targeted therapies [131]. Knútsdóttir et al. used a hybrid model to investigate epidermal growth factor (EGF) and macrophage colony-stimulating factor 1 (CSF-1) signaling between macrophages and

cancer cells during macrophage aggregation [132]. They found that CSF-1/CSFR1 autocrine signaling affects the ratio of tumor cells to macrophages during tumor growth. In a further study, they found that the macrophage/tumor cell ratio was most sensitive to the strength of EGF signaling, but usually maintained a 1:3 ratio [133].

Another ABM of triple-negative breast cancer examined the tumor enhancing effects of macrophages [134]. Norton et al. investigated the interplay between tumor growth, blood vessel recruitment and macrophage recruitment through tumor vasculature. They observed that while macrophages increase tumor growth, excessive macrophage recruitment conversely leads to a decrease in tumor growth due to the inhibition of proliferation resulting from overcrowding.

3.6. Models Focusing on Intra-Tumor Heterogeneity

Intra-tumor heterogeneity and the characteristics of the tumor microenvironment are found to have important implications in the outcome of disease progression [135]. Patients often have varied responses to treatment because each patient is unique in their genome, microbiome, disease history, lifestyle, and environment. The case of tumors is especially complex, because this heterogeneity is observed not only between tumors, but also between subpopulations of cells from the same tumor, resulting in different response to drugs [136]. While capturing this degree of heterogeneity may be difficult in experiments and clinical trials, especially the temporal dynamics of spatial heterogeneity, computational models are especially suited to tackling this challenge. This section focuses on the models that have aimed to capture intra-tumor heterogeneity.

A 2D agent-based model was used to study the interactions between an avascular tumor and immune cells (NK cells and cytotoxic T-cells) [137]. They examined the effects of cancer cell proliferation on overall tumor growth under two conditions: the first, where cancer cells do not consider the microenvironment when deciding when to proliferate, and the second, where they proliferate based on the number of healthy cells surrounding them. Tumorimmune cell interaction can have three outcomes: tumor cell death, immune cell death, or no cell death based on the state of the tumor. The predicted growth of the tumor was then compared to a xenograft tumor growth. Spatial heterogeneity was also examined in a different model where cancer cells use glycolysis instead of oxidative phosphorylation to increase their energy production. In order to study how this increased energy production affects the surrounding stroma, a combination of computational modeling and in vitro/in vivo experiments was used [138]. They used agent-based modeling to understand tumor growth in a vascularized area of the tumor. They found that tumors develop spatial patterns where macrophages and tumor cells coexisted in areas with high levels of oxygen, but that only tumor cells survived in ischemic regions. They then used an in vitro tissue-mimetic system to create the directional gradients for oxygen and lactate, which also allowed for the co-culture of tumor cells and macrophages.

Figueredo et al. created a series of hybrid models to study the interplay between the immune system (including macrophages) and tumor cells [139,140]. An agent-based on-lattice model for tumors was created using Chaste (Cancer, Heart and Soft Tissue Environment), part of the Virtual Physiological Human (VPH) Toolkit; the model consists of three layers: a

diffusible layer, a cellular layer, and a subcellular layer [141]. The diffusible layer consists of diffusible species such as oxygen, the cellular layer consists of normal cells, tumor cells, and macrophages, and the subcellular layer governs apoptosis and cell-cycle in each cell. In this model, macrophages were M1-like, they supported the immune system, and aided immunotherapies. They investigated the growth of the tumor under oxygen-dependent proliferation. They found that the emergent behavior of agent-based models allowed for the generation of additional tumor architectures over other modeling methodologies [142].

4. Discussion and Emerging Applications

The immune system is made up of many interacting components that together drive a complex spatio-temporal behavior during immune response. Thus, agent-based modeling is particularly suitable for understanding the immune systems function in health and in disease conditions such as cancer. Here, we reviewed the latest agent-based and hybrid models that investigate the contributions of the immune system to cancer growth and the effect of immunotherapy. In this context, we focused on models of immune-related tumor mechanobiology, tumor-associated vasculature, tumor-associated lymphatics, tumor immunotherapies, tumor-enhancing immune cells, and finally, models focusing on intratumor heterogeneity. Overall, ABM can generate novel hypotheses to be validated and refined by future experiments. Development and refinement of multiscale agent-based models along with experiments through an iterative process can improve our understanding of biological processes in cancer and lead to the identification of novel prognostic and predictive biomarkers that can improve therapies and help design and interpret the results of clinical trials [143].

Models investigating the tumor-enhancing effects of the immune system can provide useful insights into managing tumor-immune interactions. Since the tumor microenvironment can be very heterogeneous, care must be taken to appropriately model cell-cell interactions between cancer, stromal, and immune cells, the extracellular matrix, and the secreted factors. Accurate data from in vitro and in vivo experiments must be used to understand the transition from tumor-inhibiting to tumor-enhancing immune cell types. In addition, since immune cells such as macrophages and T-cells are usually recruited to the tumor by secreted factors, an evolving tumor vasculature is necessary to accurately model these processes. Agent-based models of the tumor enhancing effects of the immune system can help us better understand how to prevent or revert the immune system back into a tumor-inhibiting phenotype. Thus, these models will help improve immunotherapies for cancer treatment.

One limitation of the immunotherapy studies mentioned above is that although the models are quantitative in many aspects, the modules governing drug delivery and response is relatively qualitative or semi-quantitative in nature. This could potentially be resolved by combining spatio-temporal agent-based models with traditional model types, such as Physiologically Based Pharmacokinetic (PBPK) models to track drug distribution in different physiological compartments, and pharmacodynamic (PD) models for individual cellular agents to represent effects of drugs on target cells [144]. Such hybrid quantitative systems pharmacology (QSP) models can be utilized not only as a platform for basic science research, but also as a potential complement to the clinical research and drug discovery

pipeline. A schematic of such a hybrid model based on the research in our own group is presented in Figure 3. Compared with continuous models, the discretely represented agents allow the flexibility to track intra-tumor heterogeneity, such as tumor neoantigen profile, T-cell clonality and local expression of immune checkpoint molecules with preferred levels of granularity. By running multiple simulations in parallel using different parameter values and initial conditions accounting for genetic background and environmental exposure, the models can represent cohorts of patients with desired population scale heterogeneity. These properties render agent-based and hybrid models a powerful platform for conducting virtual clinical trials.

Computer models provide large-scale predictive power by allowing us to simulate clinical trials with sufficient details to study response to various conditions. Using these models, it is possible to test and predict drug failures in simulations rather than in patients, which could result in improved drug design, reduced risks and side effects, and can dramatically decrease costs of drug development. Importantly, models can predict how the immune-tumor system evolves during the course of the treatment [136]. The challenge is that available data for individual patients is limited. To address this problem, machine learning approaches can be used to build statistical models based on available patient data, and these models can be employed to simulate virtual populations to predict the effects of therapies [145]. These approaches have already been expanded to identify biomarkers and find important mutations that affect response to treatment with drugs in cancer cell lines [146–148].

Mechanistic models are another suitable approach that provides large-scale predictive capabilities based on the available information on the interactions between various components of a biological system. These models can be discrete or continuous. A certain type of model is chosen based on the application and availability of the data [149,150]. Depending on the type of the model, predictions can be made for the behavior of the signaling system in a qualitative, semi-quantitative or quantitative manner. For example, for quantitative predictions of signaling and regulatory gene networks, continuous variables need to be modeled on continuous time scales using ODEs [151].

Such detailed modeling requires knowledge of important biological reactions at every step. This can only be achieved in several iterative steps that include the implementation of various components such as signaling events and defining values for related parameters and appropriate initial conditions. In recent years, numerous models have been developed that simulate individual signaling pathways [152–154]. The challenge is that these models often do not fully capture crosstalk mechanisms that are crucial in predicting patient response to treatment, as each drug perturbs multiple biological processes. ODE-based models can be combined with agent-based models to capture the dynamics of the system being modeled in a more complex fashion. Stochasticity is one of the main advantages of agent-based models, as it applies to biological processes [155]. In comparison to ODE models that make predictions of concentrations and other events over time, ABM allows the study of each agent, as it interacts with other agents in their proximity and the ways in which that affects the large-scale behavior. These models, however, are computationally more expensive. There are also challenges in validating results from ABM due to insufficient spatio-temporal data on tumor development [156].

One aspect that is typically missing from mechanistic knowledge-based models including QSP and ABM is an input from high-throughput data, genomic or proteomic; such data can inform the models and can supplement the data obtained at the cellular and tissue levels [157]. Examples include immune landscape information from different sources including patients' databases such as TCGA (The Cancer Genome Atlas) [33,158,159]. Another source of "Big Data" for parameterization and validation of the models, including ABM, will be the emerging methodologies of digital pathology, such as using multiplex immunofluorescence (mIF) of patients' biopsies and resected tumors, with subsequent analysis of cellular and molecular spatial patterns. Steps in this direction are already underway [160]. Another source of data is image-based, using microCT, confocal, multiphoton, and super resolution microscopy, both ex vivo and in vivo; examples include imaging entire tumor vasculature with subsequent computer simulation of blood flow and molecular transport [161,162].

In addition to modeling approaches used to simulate response to treatment, virtual patients are a key component of virtual clinical trials. A virtual population has the characteristics of the original patient population but also includes individual diversity, usually comprising parameter sets weighted by a clinical or response distribution [163]. This diversity allows testing a broad range of responses that can be missed in a clinical trial. In contrast to traditional clinical trials that can only be performed after costly and lengthy development, in silico trials could be performed at every stage of the drug development. In silico design of treatments can be conducted with data-driven or mechanistic-based (knowledge-based) approaches [164]. It should be noted that in silico clinical trials require integration of data at different scales via a multi-model approach using virtual patients. Similar to a traditional clinical trial, rigorous statistical approaches are needed at various steps of virtual clinical trials. The ability to test treatment via in silico clinical trials can significantly reduce the cost and increase the efficacy of drug development.

The main challenge in the way of predictive models for virtual clinical trials is the availability of input data for the model for each patient. Detailed knowledge about the situation at the start of the simulation can significantly affect the predictive power of that model. Such input information is being generated at a growing rate and a lower cost. Furthermore, proteomic data enable the modeling of interactions of different subgroups of cells from the same tumor with each other as well as immune cells and other stromal cells, allowing modeling tumors for individual patients. Computational models can make predictions for the optimal treatments, making it safer, faster, and cheaper to complement current clinical trials. These models will improve by continuous comparison of predicted and actual response to therapy. Additionally, as more detailed information on biological parameters and disease mechanisms become available, the accuracy of the models will increase.

Currently, only 1 in 10s of clinical trials results in drugs that make it to the market [165]. The process takes 10–12 years, costing billions of dollars, sometimes with low effectiveness when used by real patients [136]. Although virtual clinical trials with virtual patients and virtual cohorts cannot replace clinical trials, they can inform design of such trials to improve success rates and increase the efficacy of the process of drug development. Virtual trials can

also better address the need for personalized therapy [166]. Finally, combining agent-based models and data-driven artificial intelligence (AI) methods (e.g., machine learning, including deep learning), we can better understand the gap between preclinical findings and clinical outcomes.

In summary, in silico modeling and specifically agent-based modeling are powerful tools of cancer systems biology and cancer immune systems biology. Combined with novel measurement methodologies and increasing amounts and sophistication of data available from clinical trials, they should bring a better mechanistic understanding and predictive capabilities of therapeutic interventions in cancer, including immunotherapies. The field is ripe for conducting predictive virtual clinical trials as a prerequisite to clinical trials in patients.

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References

- 1. Hanahan D; Weinberg RA Hallmarks of cancer: The next generation. Cell 2011, 144, 646–674. [PubMed: 21376230]
- 2. Paget S The distribution of secondary growths in cancer of the breast. Lancet 1889, 133, 571–573.
- 3. Wei SC; Duffy CR; Allison JP Fundamental mechanisms of immune checkpoint blockade therapy. Cancer Discern. 2018, 8, 1069–1086.
- 4. Chen DS; Mellman I Elements of cancer immunity and the cancer-Immune set point. Nature 2017, 541, 321–330. [PubMed: 28102259]
- Quail DF; Joyce JA Microenvironmental regulation of tumor progression and metastasis. Nat. Med 2013, 19, 1423–1437. [PubMed: 24202395]
- Ansell SM; Vonderheide RH Cellular composition of the tumor microenvironment. Am. Soc. Clin. Oncol. Educ. B 2013, 33, e91–e97.
- Pitt JM; Marabelle A; Eggermont A; Soria JC; Kroemer G; Zitvogel L Targeting the tumor microenvironment: Removing obstruction to anticancer immune responses and immunotherapy. Ann. Oncol 2016, 27, 1482–1492. [PubMed: 27069014]
- 8. Crespo I; Coukos G; Doucey M; Xenarios I Modelling approaches to discovery in the tumor microenvironment. J. Cancer Immunol. Ther 2018, 1, 23–37.
- Netea MG; Joosten LAB; Latz E; Mills KHG; Natoli G; Stunnenberg HG; O'Neill LAJ; Xavier RJ Trained immunity: A program of innate immune memory in health and disease. Science 2016, 352, aaf1098. [PubMed: 27102489]
- 10. Netea MG; Quintin J; Van Der Meer JWM Trained immunity: A memory for innate host defense. Cell Host Microbe 2011, 9, 355–361. [PubMed: 21575907]
- 11. Kumar H; Kawai T; Akira S Pathogen recognition by the innate immune system. Int. Rev. Immunol 2011, 30, 16–34. [PubMed: 21235323]
- Gajewski TF; Schreiber H; Fu Y-X Innate and adaptive immune cells in the tumor microenvironment. Nat. Immunol 2013, 14, 1014–1022. [PubMed: 24048123]
- 13. Dykes SS; Hughes VS; Wiggins JM; Fasanya HO; Tanaka M; Siemann D Stromal cells in breast cancer as a potential therapeutic target. Oncotarget 2018, 9, 23761–23779. [PubMed: 29805773]
- 14. Chaudhary B; Elkord E Regulatory T cells in the tumor microenvironment and cancer progression: Role and therapeutic targeting. Vaccines 2016, 4, 28.

 Fleming V; Hu X; Weber R; Nagibin V; Groth C; Altevogt P; Utikal J; Umansky V Targeting myeloid-derived suppressor cells to bypass tumor-induced immunosuppression. Front. Immunol 2018, 9, 398. [PubMed: 29552012]

- Beyer M; Schultze JL Regulatory T cells in cancer. Blood 2006, 108, 804–811. [PubMed: 16861339]
- De Visser KE; Eichten A; Coussens LM Paradoxical roles of the immune system during cancer development. Nat. Rev. Cancer 2006, 6, 24–37. [PubMed: 16397525]
- 18. Palucka AK; Coussens LM The basis of oncoimmunology. Cell 2016, 164, 1233–1247. [PubMed: 26967289]
- Blank C; Mackensen A Contribution of the PD-L1/PD-1 pathway to T-cell exhaustion: An update on implications for chronic infections and tumor evasion. Cancer Immunol. Immunother 2007, 56, 739–745. [PubMed: 17195077]
- 20. Jiang Y; Li Y; Zhu B T-cell exhaustion in the tumor microenvironment. Cell Death Dis. 2015, 6, e1792. [PubMed: 26086965]
- Seifert AM; Zeng S; Zhang JQ; Kim TS; Cohen NA; Beckman MJ; Medina BD; Maltbaek JH; Loo JK; Crawley MH; et al. PD-1/PD-L1 blockade enhances T-cell activity and antitumor efficacy of Imatinib in gastrointestinal stromal tumors. Clin. Cancer Res 2017, 23, 454–465. [PubMed: 27470968]
- 22. Oyer JL; Gitto SB; Altomare DA; Copik AJ PD-L1 blockade enhances anti-tumor efficacy of NK cells. Oncoimmunology 2018, 7, e1509819. [PubMed: 30377572]
- Abdel-Wahab N; Shah M; Lopez-Olivo MA; Suarez-Almazor ME Use of immune checkpoint inhibitors in the treatment of patients with cancer and preexisting autoimmune disease: A systematic review. Ann. Intern. Med 2018, 168, 121–130. [PubMed: 29297009]
- 24. Sharma P; Allison JP Immune checkpoint targeting in cancer therapy: Toward combination strategies with curative potential. Cell 2015, 161, 205–214. [PubMed: 25860605]
- 25. Raggi F; Pelassa S; Pierobon D; Penco F; Gattorno M; Novelli F; Eva A; Varesio L; Giovarelli M; Bosco MC Regulation of human macrophage M1-M2 polarization balance by hypoxia and the triggering receptor expressed on myeloid cells-1. Front. Immunol 2017, 8, 1097. [PubMed: 28936211]
- 26. Choi J; Gyamfi J; Jang H; Koo JS The role of tumor-associated macrophage in breast cancer biology. Histol. Histopathol 2018, 33, 133–145. [PubMed: 28681373]
- 27. Seager RJ; Hajal C; Spill F; Kamm RD; Zaman MH Dynamic interplay between tumour, stroma and immune system can drive or prevent tumour progression. Converg. Sci. Phys. Oncol 2017, 3, 034002. [PubMed: 30079253]
- 28. Tripathi C; Tewari BN; Kanchan RK; Baghel KS; Nautiyal N; Shrivastava R; Kaur H; Bhatt MLB; Bhadauria S Macrophages are recruited to hypoxic tumor areas and acquire a pro-angiogenic M2-polarized phenotype via hypoxic cancer cell derived cytokines Oncostatin M and Eotaxin. Oncotarget 2014, 5, 5350–5368. [PubMed: 25051364]
- 29. Williams CB; Yeh ES; Soloff AC Tumor-associated macrophages: Unwitting accomplices in breast cancer malignancy. NPJ Breast Cancer 2016, 2, 15025. [PubMed: 26998515]
- 30. Luo H; Tu G; Liu Z; Liu M; Noy R; Pollard JWW Tumor-associated macrophages: From mechanisms to therapy. Immunity 2014, 41, 49–61. [PubMed: 25035953]
- 31. Hollmén M; Roudnicky F; Karaman S; Detmar M Characterization of macrophage—Cancer cell crosstalk in estrogen receptor positive and triple-negative breast cancer. Sci. Rep 2015, 5, 9188. [PubMed: 25776849]
- 32. Sousa S; Brion R; Lintunen M; Kronqvist P; Sandholm J; Mdnkkdnen J; Kellokumpu-Lehtinen P-L; Lauttia S; Tynninen O; Joensuu H; et al. Human breast cancer cells educate macrophages toward the M2 activation status. Breast Cancer Res. 2015, 17, 101. [PubMed: 26243145]
- 33. Gubin MM; Esaulova E; Ward JP; Malkova ON; Runci D; Wong P; Noguchi T; Arthur CD; Meng W; Alspach E; et al. High-dimensional analysis delineates myeloid and lymphoid compartment remodeling during successful immune-checkpoint cancer therapy. Cell 2018, 175, 1014–1030. [PubMed: 30343900]

34. Su S; Liu Q; Chen J; Chen J; Chen F; He C; Huang D; Wu W; Lin L; Huang W; et al. A Positive feedback loop between mesenchymal-like cancer cells and macrophages is essential to breast cancer metastasis. Cancer Cell 2014, 25, 605–620. [PubMed: 24823638]

- 35. Burger GA; Danen EHJ; Beltman JB Deciphering epithelial—Mesenchymal transition regulatory networks in cancer through computational approaches. Front. Oncol 2017, 7, 162. [PubMed: 28824874]
- 36. Mitchem JB; Brennan DJ; Knolhoff BL; Belt BA; Zhu Y; Sanford DE; Belaygorod L; Carpenter D; Collins L; Piwnica-Worms D; et al. Targeting tumor-infiltrating macrophages decreases tumor-initiating cells, relieves immunosuppression, and improves chemotherapeutic responses. Cancer Res. 2013, 73, 1128–1141. [PubMed: 23221383]
- 37. Qian B; Deng Y; Im JH; Muschel RJ; Zou Y; Li J; Lang RA; Pollard JW A distinct macrophage population mediates metastatic breast cancer cell extravasation, establishment and growth. PLoS ONE 2009, 4, e6562. [PubMed: 19668347]
- 38. Lohela M; Casbon A-J; Olow A; Bonham L; Branstetter D; Weng N; Smith J; Werb Z Intravital imaging reveals distinct responses of depleting dynamic tumor-associated macrophage and dendritic cell subpopulations. Proc. Natl. Acad. Sci. USA 2014, 111, E5086–E5095. [PubMed: 25385645]
- Ngambenjawong C; Cieslewicz M; Schellinger JG; Pun SH Synthesis and evaluation of multivalent M2pep peptides for targeting alternatively activated M2 macrophages. J. Control. Release 2016, 224, 103–111. [PubMed: 26772876]
- 40. Mishalian I; Granot Z; Fridlender ZG The diversity of circulating neutrophils in cancer. Immunobiology 222, 82–88. [PubMed: 26874580]
- 41. Davis BP; Rothenberg ME Eosinophils and cancer. Cancer Immunol. Res 2014, 2, 1–8. [PubMed: 24778159]
- 42. Hämmerling GJ; Carretero R; Beckhove P; Salgado OC; Sektioglu IM; Garbi N Eosinophils orchestrate cancer rejection by normalizing tumor vessels and enhancing infiltration of CD8+ T cells. Nat. Immunol 2015, 16, 609–617. [PubMed: 25915731]
- 43. Sakkal S; Miller S; Apostolopoulos V; Nurgali K Eosinophils in cancer: Favourable or unfavourable? Curr. Med. Chem 2016, 23, 650–666. [PubMed: 26785997]
- 44. Shinde SB; Kurhekar MP Review of the systems biology of the immune system using agent-based models. IET Syst. Biol 2018, 12, 83–92. [PubMed: 29745901]
- 45. Altrock PM; Liu LL; Michor F The mathematics of cancer: Integrating quantitative models. Nat. Rev. Cancer 2015, 15, 730–745. [PubMed: 26597528]
- 46. Rejniak KA; McCawley LJ Current trends in mathematical modeling of tumor-microenvironment interactions: A survey of tools and applications. Exp. Biol. Med 2010, 235, 411–423.
- 47. Eftimie R; Bramson JL; Earn DJD Interactions between the immune system and cancer: A brief review of non-spatial mathematical models. Bull. Math. Biol 2011, 73, 2–32. [PubMed: 20225137]
- 48. Alemani D; Pappalardo F; Pennisi M; Motta S; Brusic V Combining cellular automata and lattice Boltzmann method to model multiscale avascular tumor growth coupled with nutrient diffusion and immune competition. J. Immunol. Methods 2012, 376, 55–68. [PubMed: 22154892]
- 49. Bellomo N; Delitala M From the mathematical kinetic, and stochastic game theory to modelling mutations, onset, progression and immune competition of cancer cells. Phys. Life Rev 2008, 5, 183–206.
- 50. Eladdadi A; de Pillis L; Kim P Modelling tumour-immune dynamics, disease progression and treatment. Lett. Biomath 2018, 5, S1–S5.
- 51. Dritschel H; Waters SL; Roller A; Byrne HM A mathematical model of cytotoxic and helper T cell interactions in a tumour microenvironment. Lett. Biomath 2018, 5, S36–S68.
- 52. Nikolopoulou E; Johnson LR; Harris D; Nagy JD; Stites EC; Kuang Y Tumour-immune dynamics with an immune checkpoint inhibitor. Lett. Biomath 2018, 5, S137–S159.
- 53. Salgia R; Mambetsariev I; Hewelt B; Achuthan S; Li H; Poroyko V; Wang Y; Sattler M Modeling small cell lung cancer (SCLC) biology through deterministic and stochastic mathematical models. Oncotarget 2018, 9, 26226–26242. [PubMed: 29899855]

 Konstorum A; Vella AT; Adler AJ; Laubenbacher RC Addressing current challenges in cancer immunotherapy with mathematical and computational modelling. J. R. Soc. Interface 2017, 14, 20170150. [PubMed: 28659410]

- 55. Chiacchio F; Pennisi M; Russo G; Motta S; Pappalardo F Agent-based modeling of the immune system: NetLogo, a promising framework. BioMed Res. Int 2014, 907171. [PubMed: 24864263]
- 56. An G; Mi Q; Dutta-Moscato J; Vodovotz Y Agent-based models in translational systems biology. Wiley Interdiscip. Rev. Syst. Biol. Med 2009, 1, 159–171. [PubMed: 20835989]
- 57. Chavali AK; Gianchandani EP; Tung KS; Lawrence MB; Peirce SM; Papin JA Characterizing emergent properties of immunological systems with multi-cellular rule-based computational modeling. Trends Immunol. 2008, 29, 589–599. [PubMed: 18964301]
- 58. Shi ZZ; Wu C-H; Ben-Arieh D Agent-based model: A surging tool to simulate infectious diseases in the immune system. Open J. Model. Simul 2014, 02, 12–22.
- 59. Segovia-Juarez JL; Ganguli S; Kirschner D Identifying control mechanisms of granuloma formation during M. tuberculosis infection using an agent-based model. J. Theor. Biol 2004, 231, 357–376. [PubMed: 15501468]
- 60. Tokarski C; Hummert S; Mech F; Figge MT; Germerodt S; Schroeter A; Schuster S; Linde J; Hu G Agent-based modeling approach of immune defense against spores of opportunistic human pathogenic fungi. Front. Microbiol 2012, 3, 129. [PubMed: 22557995]
- Dong X; Foteinou PT; Calvano SE; Lowry SF; Androulakis IP Agent-based modeling of endotoxin-induced acute inflammatory response in human blood leukocytes. PLoS ONE 2010, 5, e9249. [PubMed: 20174629]
- 62. Solovyev A; Mi Q; Tzen YT; Brienza D; Vodovotz Y Hybrid equation/agent-based model of ischemia-induced hyperemia and pressure ulcer formation predicts greater propensity to ulcerate in subjects with spinal cord injury. PLoS Comput. Biol 2013, 9, e1003070. [PubMed: 23696726]
- 63. Santoni D; Pedicini M; Castiglione F Implementation of a regulatory gene network to simulate the TH1/2 differentiation in an agent-based model of hypersensitivity reactions. Bioinformatics 2008, 24, 1374–1380. [PubMed: 18413328]
- 64. Bailey AM; Lawrence MB; Shang H; Katz AJ; Peirce SM Agent-based model of therapeutic adipose-derived stromal cell trafficking during ischemia predicts ability to roll on p-selectin. PLoS Comput. Biol 2009, 5, e1000294. [PubMed: 19247427]
- 65. D'Souza RM; Lysenko M; Marino S; Kirschner D; Souza RMD; Arbor A Data-parallel algorithms for agent-based model simulation of tuberculosis on graphics processing units In Proceedings of the 2009 Spring Simulation Multiconference, San Diego, CA, USA, 22–27 3 2009.
- 66. Song SO; Hogg J; Peng ZY; Parker R; Kellum JA; Clermont G Ensemble models of neutrophil trafficking in severe sepsis. PLoS Comput. Biol 2012, 8, e1002422. [PubMed: 22412365]
- 67. Mi Q; Riviere B; Clermont G; Steed DL; Vodovotz Y Agent-based model of inflammation and wound healing: Insights into diabetic foot ulcer pathology and the role of transforming growth factor-β1 Wound Repair Regen. 2007, 15, 671–682. [PubMed: 17971013]
- 68. Alarcón T; Byrne HM; Maini PK A mathematical model of the effects of hypoxia on the cell-cycle of normal and cancer cells. J. Theor. Biol 2004, 229, 395–411. [PubMed: 15234206]
- 69. Hoehme S; Bertaux F; Weens W; Grasl-Kraupp B; Hengstler JG; Drasdo D Model prediction and validation of an order mechanism controlling the spatiotemporal phenotype of early hepatocellular carcinoma. Bull. Math. Biol 2018, 80, 1134–1171. [PubMed: 29568983]
- 70. Bianca C; Pennisi M The triplex vaccine effects in mammary carcinoma: A nonlinear model in tune with SimTriplex. Nonlinear Anal. Real World Appl 2012, 13, 1913–1940.
- 71. Wang J; Zhang L; Jing C; Ye G; Wu H; Miao H; Wu Y; Zhou X Multi-scale agent-based modeling on melanoma and its related angiogenesis analysis. Theor. Biol. Med. Model 2013, 10, 41. [PubMed: 23800293]
- 72. Kather JN; Poleszczuk J; Suarez-Carmona M; Krisam J; Charoentong P; Valous NA; Weis CA; Tavernar L; Leiss F; Herpel E; et al. In silico modeling of immunotherapy and stroma-targeting therapies in human colorectal cancer. Cancer Res. 2017, 77, 6442–6452. [PubMed: 28923860]
- 73. Pennisi M; Pappalardo F; Motta S Agent based modeling of lung metastasis-immune system competition In Lecture Notes in Computer Science (Including Subseries Lecture Notes in

- Artificial Intelligence and Lecture Notes in Bioinformatics); Springer: Berlin/Heidelberg, Germany, 2009; Volume 5666, pp. 1–3.
- 74. Jagiella N; Muller B; Muller M; Vignon-Clementel IE; Drasdo D Inferring growth control mechanisms in growing multi-cellular spheroids of NSCLC cells from spatial-temporal image data. PLoS Comput. Biol 2016, 12, e1004412. [PubMed: 26866479]
- 75. Pennisi M; Pappalardo F; Palladini A; Nicoletti G; Nanni P; Lollini P-L; Motta S Modeling the competition between lung metastases and the immune system using agents. BMC Bioinform. 2010, 11, S13.
- Bezzi M; Celada F; Ruffo S; Seiden PE The transition between immune and disease states in a cellular automaton model of clonal immune response. Phys. A Stat. Mech. Its Appl 1997, 245, 145–163.
- 77. Baldazzi V; Castiglione F; Bernaschi M An enhanced agent based model of the immune system response. Cell Immunol. 2006, 244, 77–79. [PubMed: 17416357]
- 78. Bernaschi M; Castiglione F Design and implementation of an immune system simulator. Comput. Biol. Med 2001, 31, 303–331. [PubMed: 11535199]
- 79. Celada F; Seiden PE A computer model of cellular interactions in the immune system. Immunol. Today 1992, 13, 56–62. [PubMed: 1575893]
- 80. Emerson A; Rossi E ImmunoGrid the virtual human immune system project. Stud. Heal. Technol. Inf 2007, 126, 87–92.
- 81. Halling-Brown M; Pappalardo F; Rapin N; Zhang P; Alemani D; Emerson A; Castiglione F; Duroux P; Pennisi M; Miotto O; et al. ImmunoGrid: Towards agent-based simulations of the human immune system at a natural scale. Philos. Trans. R. Soc. A Math. Phys. Eng. Sci 2010, 368, 2799.
- 82. Perelson AS; Ribeiro RM Modeling the within-host dynamics of HIV infection. BMC Biol. 2013, 11, 96. [PubMed: 24020860]
- 83. Warrender C; Forrest S; Koster F Modeling intercellular interactions in early Mycobacterium infection. Bull. Math. Biol 2006, 68, 2233–2261. [PubMed: 17086496]
- 84. Ghaffarizadeh A; Heiland R; Friedman SH; Mumenthaler SM; Macklin P PhysiCell: An open source physics-based cell simulator for 3-D multicellular systems. PLOS Comput. Biol 2018, 14, e1005991. [PubMed: 29474446]
- Gilkes DM; Semenza GL; Wirtz D Hypoxia and the extracellular matrix: Drivers of tumour metastasis. Nat. Rev. Cancer 2014, 14, 430–439. [PubMed: 24827502]
- 86. Alfonso JCL; Talkenberger K; Seifert M; Klink B; Hawkins-Daarud A; Swanson KR; Hatzikirou H; Deutsch A The biology and mathematical modelling of glioma invasion: A review. J. R. Soc. Interface 2017, 14, 20170490. [PubMed: 29118112]
- 87. Massey SC; Rockne RC; Hawkins-Daarud A; Gallaher J; Anderson ARA; Canoll P; Swanson KR Simulating PDGF-driven glioma growth and invasion in an anatomically accurate brain domain. Bull. Math. Biol 2018, 80, 1292–1309. [PubMed: 28842831]
- 88. Juliano J; Gil O; Hawkins-Daarud A; Noticewala S; Rockne RC; Gallaher J; Massey SC; Sims PA; Anderson ARA; Swanson KR; et al. Comparative dynamics of microglial and glioma cell motility at the infiltrative margin of brain tumours. J. R. Soc. Interface 2018, 15, 20170582. [PubMed: 29445035]
- 89. Frascoli F; Flood E; Kim PS A model of the effects of cancer cell motility and cellular adhesion properties on tumour-immune dynamics. Math. Med. Biol 2016, 34, dqw004.
- Noonan DM; De Lerma Barbaro A; Vannini N; Mortara L; Albini A Inflammation, inflammatory cells and angiogenesis: Decisions and indecisions. Cancer Metastasis Rev. 2008, 27, 31–40.
 [PubMed: 18087678]
- 91. Tian L; Goldstein A; Wang H; Ching Lo H; Sun Kim I; Welte T; Sheng K; Dobrolecki LE; Zhang X; Putluri N; et al. Mutual regulation of tumour vessel normalization and immunostimulatory reprogramming. Nature 2017, 544, 250–254. [PubMed: 28371798]
- 92. Uppal A; Wightman SC; Ganai S; Weichselbaum RR; An G Investigation of the essential role of platelet-tumor cell interactions in metastasis progression using an agent-based model. Theor. Biol. Med. Model 2014, 11, 17. [PubMed: 24725600]

93. Alfonso JCL; Schaadt NS; Schbnmeyer R; Brieu N; Forestier G; Wemmert C; Feuerhake F; Hatzikirou H In-silico insights on the prognostic potential of immune cell infiltration patterns in the breast lobular epithelium. Sci. Rep 2016, 6, 33322. [PubMed: 27659691]

- 94. Reddy NP; Krouskop TA; Newell PH A computer model of the lymphatic system. Comput. Biol. Med 1977, 7, 181–197. [PubMed: 891141]
- 95. Jamalian S; Jafarnejad M; Zawieja SD; Bertram CD; Gashev AA; Zawieja DC; Davis MJ; Moore JE Demonstration and analysis of the suction effect for pumping lymph from tissue beds at subatmospheric pressure. Sci. Rep 2017, 7, 12080. [PubMed: 28935890]
- 96. Jamalian S; Davis MJ; Zawieja DC; Moore JE Network scale modeling of lymph transport and its effective pumping parameters. PLoS ONE 2016, 11, e0148384. [PubMed: 26845031]
- 97. Jamalian S; Bertram CD; Richardson WJ; Moore JE Parameter sensitivity analysis of a lumped-parameter model of a chain of lymphangions in series. Am. J. Physiol. Circ. Physiol 2013, 305, H1709–H1717.
- 98. Wilson JT; van Loon R; Wang W; Zawieja DC; Moore JE Determining the combined effect of the lymphatic valve leaflets and sinus on resistance to forward flow. J. Biomech 2015, 48, 3584–3590. [PubMed: 26315921]
- 99. Roose T; Swartz MA Multiscale modeling of lymphatic drainage from tissues using homogenization theory. J. Biomech 2012, 45, 107–115. [PubMed: 22036032]
- 100. Jafarnejad M; Zawieja DC; Brook BS; Nibbs RJB; Moore JE A novel computational model predicts key regulators of chemokine gradient formation in lymph nodes and site-specific roles for CCL19 and ACKR4. J. Immunol 2017, 299, ji1700377.
- 101. Jafarnejad M; Woodruff MC; Zawieja DC; Carroll MC; Moore JE Modeling lymph flow and fluid exchange with blood vessels in lymph nodes. Lymphat. Res. Biol 2015, 13, 234–247. [PubMed: 26683026]
- 102. Cooper LJ; Heppell JP; Clough GF; Ganapathisubramani B; Roose T An image-based model of fluid flow through lymph nodes. Bull. Math. Biol 2016, 78, 52–71. [PubMed: 26690921]
- 103. Marino S; Gideon HP; Gong C; Mankad S; McCrone JT; Lin PL; Linderman JJ; Flynn JAL; Kirschner DE Computational and empirical studies predict mycobacterium tuberculosis-specific T cells as a biomarker for infection outcome. PLoS Comput. Biol 2016, 12, e1004804. [PubMed: 27065304]
- 104. Gong C; Linderman JJ; Kirschner D Harnessing the heterogeneity of T cell differentiation fate to fine-tune generation of effector and memory T cells. Front. Immunol 2014, 5, 57. [PubMed: 24600448]
- 105. Margaris KN; Black RA Modelling the lymphatic system: Challenges and opportunities. J. R. Soc. Interface 2012, 9, 601–612. [PubMed: 22237677]
- 106. Meyer-Hermann M A mathematical model for the germinal center morphology and affinity maturation. J. Theor. Biol 2002, 216, 273–300. [PubMed: 12183119]
- 107. Meyer-Hermann ME; Maini PK Cutting edge: Back to "one-way" germinal centers. J. Immunol 2005, 174, 2489–2493. [PubMed: 15728453]
- 108. Meyer-Hermann ME; Maini PK; Iber D An analysis of B cell selection mechanisms in germinal centers. Math. Med. Biol. A J. IMA 2006, 23, 255–277.
- 109. Bogle G; Dunbar PR T cell responses in lymph nodes. Wiley Interdiscip. Rev. Syst. Biol. Med 2010, 2, 107–116. [PubMed: 20836014]
- 110. Bogle G; Dunbar PR Agent-based simulation of T-cell activation and proliferation within a lymph node. Immunol. Cell Biol 2010, 88, 172–179. [PubMed: 19884904]
- 111. Bogle G; Dunbar PR On-lattice simulation of T cell motility, chemotaxis, and trafficking in the lymph node paracortex. PLoS ONE 2012, 7, e45258. [PubMed: 23028887]
- 112. Bogle G; Dunbar PR Simulating T-cell motility in the lymph node paracortex with a packed lattice geometry. Immunol. Cell Biol 2008, 86, 676–687. [PubMed: 18711399]
- 113. Moreau HD; Bogle G; Bousso P A virtual lymph node model to dissect the requirements for T-cell activation by synapses and kinapses. Immunol. Cell Biol 2016, 94, 680–688. [PubMed: 27089942]

114. Folcik VA; An GC; Orosz CG The Basic Immune Simulator: An agent-based model to study the interactions between innate and adaptive immunity. Theor. Biol. Med. Model 2007, 4, 39. [PubMed: 17900357]

- 115. Kim PS; Levy D; Lee PP Modeling and simulation of the immune system as a self-regulating network. Methods Enzymol. 2009, 467, 79–109. [PubMed: 19897090]
- 116. Jacob C; Sarpe V; Gingras C; Feyt RP Swarm-based simulations for immunobiology: What can agent-based models teach us about the immune system? In Intelligent Systems Reference Library; Springer: Berlin/Heidelberg, Germany, 2011; Volume 11, pp. 29–64.
- 117. Marino S; El-Kebir M; Kirschner D A hybrid multi-compartment model of granuloma formation and T cell priming in Tuberculosis. J. Theor. Biol 2011, 280, 50–62. [PubMed: 21443879]
- 118. Marino S; Kirschner D A multi-compartment hybrid computational model predicts key roles for dendritic cells in Tuberculosis infection. Computation 2016, 4, 39. [PubMed: 28989808]
- 119. Dréau D; Stanimirov D; Carmichael T; Hadzikadic M An agent-based model of solid tumor progression In Lecture Notes in Computer Science (Including Subseries Lecture Notes in Artificial Intelligence and Lecture Notes in Bioinformatics); Springer: Berlin/Heidelberg, Germany, 2009; Volume 5462, pp. 187–198.
- 120. Pappalardo F; Forero IM; Pennisi M; Palazon A; Melero I; Motta S Simb16: Modeling induced immune system response against B16-melanoma. PLoS ONE 2011, 6, e26523. [PubMed: 22028894]
- 121. Sanchez-Paulete AR; Labiano S; Rodriguez-Ruiz ME; Azpilikueta A; Etxeberria I; Bolaños E; Lang V; Rodriguez M; Aznar MA; Jure-Kunkel M; et al. Deciphering CD137 (4–1BB) signaling in T-cell costimulation for translation into successful cancer immunotherapy. Eur. J. Immunol 2016, 46, 513–522. [PubMed: 26773716]
- 122. Santiago DN; Heidbuechel JPW; Kandell WM; Walker R; Djeu J; Engeland CE; Abate-Daga D; Enderling H Fighting cancer with mathematics and viruses. Viruses 2017, 9, 239.
- 123. Walker R; Navas PE; Friedman SH; Galliani S; Karolak A; MacFarlane F; Noble R; Poleszczuk J; Russell S; Rejniak KA; et al. Enhancing synergy of CAR T cell therapy and oncolytic virus therapy for pancreatic cancer. bioRxiv 2016, 055988.
- 124. Rohrs JA; Zheng D; Graham NA; Wang P; Finley SD Computational model of chimeric antigen receptors explains site-specific phosphorylation kinetics. Biophys. J 2018, 115, 1116–1129. [PubMed: 30197180]
- 125. Schumacher TN; Schreiber RD Neoantigens in cancer immunotherapy. Science 2015, 348, 69–74. [PubMed: 25838375]
- 126. Sharma P; Allison JP The future of immune checkpoint therapy. Science 2015, 348, 56–61. [PubMed: 25838373]
- 127. Gong C; Milberg O; Wang B; Vicini P; Narwal R; Roskos L; Popel AS A computational multiscale agent-based model for simulating spatio-temporal tumour immune response to PD1 and PDL1 inhibition. J. R. Soc. Interface 2017, 14, 20170320. [PubMed: 28931635]
- 128. Hillen T; Enderling H; Hahnfeldt P The tumor growth paradox and immune system-mediated selection for cancer stem cells. Bull. Math. Biol 2013, 75, 161–184. [PubMed: 23196354]
- 129. Enderling H; Hlatky L; Hahnfeldt P Immunoediting: Evidence of the multifaceted role of the immune system in self-metastatic tumor growth. Theor. Biol. Med. Model 2012, 9, 31. [PubMed: 22838395]
- 130. Dehne N; Mora J; Namgaladze D; Weigert A; Brüne B Cancer cell and macrophage cross-talk in the tumor microenvironment. Curr. Opin. Pharmacol 2017, 35, 12–19. [PubMed: 28538141]
- 131. Wells DK; Chuang Y; Knapp LM; Brockmann D; Kath WL; Leonard JN Spatial and functional heterogeneities shape collective behavior of tumor-immune networks. PLoS Comput. Biol 2015, 11, e1004181. [PubMed: 25905470]
- 132. Knútsdóttir H; Pálsson E; Edelstein-Keshet L Mathematical model of macrophage-facilitated breast cancer cells invasion. J. Theor. Biol 2014, 357, 184–199. [PubMed: 24810842]
- 133. Knutsdottir H; Condeelis JS; Palsson E 3-D individual cell based computational modeling of tumor cell-macrophage paracrine signaling mediated by EGF and CSF-1 gradients. Integr. Biol 2016, 8, 104–119.

134. Norton KA; Jin K; Popel AS Modeling triple-negative breast cancer heterogeneity: Effects of stromal macrophages, fibroblasts and tumor vasculature. J. Theor. Biol 2018, 452, 56–68. [PubMed: 29750999]

- 135. Marusyk A; Almendro V; Polyak K Intra-tumour heterogeneity: A looking glass for cancer? Nat. Rev. Cancer 2012, 12, 323–334. [PubMed: 22513401]
- 136. Lehrach H Virtual clinical trials, an essential step in increasing the effectiveness of the drug development process. Public Health Genom. 2015, 18, 366–371.
- 137. Pourhasanzade F; Sabzpoushan S; Alizadeh AM; Esmati E An agent-based model of avascular tumor growth: Immune response tendency to prevent cancer development. Simulation 2017, 93, 641–657.
- 138. Carmona-Fontaine C; Bucci V; Akkari L; Deforet M; Joyce JA; Xavier JB Emergence of spatial structure in the tumor microenvironment due to the Warburg effect. Proc. Natl. Acad. Sci. USA 2013, 110, 19402–19407. [PubMed: 24218566]
- 139. Figueredo GP; Aickelin U; Siebers PO Systems dynamics or agent-based modelling for immune simulation? In Proceedings of the Lecture Notes in Computer Science (Including Subseries Lecture Notes in Artificial Intelligence and Lecture Notes in Bioinformatics); Springer: Berlin/Heidelberg, Germany, 2011; Volume 6825, pp. 81–94.
- 140. Figueredo GP; Siebers P-O; Aickelin U Investigating mathematical models of immunointeractions with early-stage cancer under an agent-based modelling perspective. BMC Bioinform. 2013, 14, S6.
- 141. Figueredo GP; Joshi TV; Osborne JM; Byrne HM; Owen MR On-lattice agent-based simulation of populations of cells within the open-source Chaste framework. Interface Focus 2013, 3, 20120081. [PubMed: 24427527]
- 142. Figueredo GP; Siebers PO; Owen MR; Reps J; Aickelin U Comparing stochastic differential equations and agent-based modelling and simulation for early-stage cancer. PLoS ONE 2014, 9, e95150. [PubMed: 24752131]
- 143. Yankeelov TE; An G; Saut O; Luebeck EG; Popel AS; Ribba B; Vicini P; Zhou X; Weis JA; Ye K; et al. Multi-scale modeling in clinical oncology: Opportunities and barriers to success. Ann. Biomed. Eng 2016, 44, 2626–2641. [PubMed: 27384942]
- 144. Cosgrove J; Butler J; Alden K; Read M; Kumar V; Cucurull-Sanchez L; Timmis J; Coles M Agent-based modeling in systems pharmacology. CPT Pharmacometrics Syst. Pharmacol 2015, 4, 615–629. [PubMed: 26783498]
- 145. Rieger TR; Allen RJ; Bystricky L; Chen Y; Colopy GW; Cui Y; Gonzalez A; Liu Y; White RD; Everett RA; et al. Improving the generation and selection of virtual populations in quantitative systems pharmacology models. Prog. Biophys. Mol. Biol 2018, 139, 15–22. [PubMed: 29902482]
- 146. Barretina J; Caponigro G; Stransky N; Venkatesan K; Margolin AA; Kim S; Wilson CJ; Lehár J; Kryukov GV; Sonkin D; et al. The Cancer Cell Line Encyclopedia enables predictive modelling of anticancer drug sensitivity. Nature 2012, 483, 603–607. [PubMed: 22460905]
- 147. Garnett MJ; Edelman EJ; Heidorn SJ; Greenman CD; Dastur A; Lau KW; Greninger P; Thompson IR; Luo X; Soares J; et al. Systematic identification of genomic markers of drug sensitivity in cancer cells. Nature 2012, 483, 570–575. [PubMed: 22460902]
- 148. Rubio-Perez C; Tamborero D; Schroeder MP; Antolin AA; Deu-Pons J; Perez-Llamas C; Mestres J; Gonzalez-Perez A; Lopez-Bigas N In silico prescription of anticancer drugs to cohorts of 28 tumor types reveals targeting opportunities. Cancer Cell 2015, 27, 382–396. [PubMed: 25759023]
- 149. Anderson ARA; Maini PK Mathematical oncology. Bull. Math. Biol 2018, 80, 945–953. [PubMed: 29679280]
- 150. Sun X; Hu B Mathematical modeling and computational prediction of cancer drug resistance. Brief. Bioinform 2017, 19, 1382–1399.
- Arney K Improving brain-cancer therapies through mathematical modelling. Nature 2018, 561, S52–S53. [PubMed: 30258162]
- 152. Barua D; Hlavacek WS Modeling the effect of APC truncation on destruction complex function in colorectal cancer cells. PLoS Comput. Biol 2013, 9, e1003217. [PubMed: 24086117]

153. Klinger B; Sieber A; Fritsche-Guenther R; Witzel F; Berry L; Schumacher D; Yan Y; Durek P; Merchant M; Schafer R; et al. Network quantification of EGFR signaling unveils potential for targeted combination therapy. Mol. Syst. Biol 2014, 9, 673.

- 154. Kirouac DC; Du JY; Lahdenranta J; Overland R; Yarar D; Paragas V; Pace E; McDonagh CF; Nielsen UB; Onsum MD Computational modeling of ERBB2-amplified breast cancer identifies combined ErbB2/3 blockade as superior to the combination of MEK and AKT inhibitors (Science Signaling 6:288 (ra68)). Sci. Signal 2014, 7, er5.
- 155. Carbo A; Bassaganya-Riera J; Pedragosa M; Viladomiu M; Marathe M; Eubank S; Wendelsdorf K; Bisset K; Hoops S; Deng X; et al. Predictive computational modeling of the mucosal immune responses during Helicobacter pylori infection. PLoS ONE 2013, 8, e73365. [PubMed: 24039925]
- 156. Zand R; Abedi V; Hontecillas R; Lu P; Noorbakhsh-Sabet N; Verma M; Leber A; Tubau-Juni N; Bassaganya-Riera J Development of synthetic patient populations and in silico clinical trials In Accelerated Path to Cures; Springer International Publishing AG: Basel, Switzerland, 2018; pp. 57–77.
- 157. Edelman LB; Eddy JA; Price ND In silico models of cancer. Wiley Interdiscip. Rev. Syst. Biol. Med 2010, 2, 438–459. [PubMed: 20836040]
- 158. Jerby-Arnon L; Shah P; Cuoco MS; Rodman C; Su M-J; Melms JC; Leeson R; Kanodia A; Mei S; Lin J-R; et al. A cancer cell program promotes T cell exclusion and resistance to checkpoint blockade. Cell 2018, 175, 984–997.e24. [PubMed: 30388455]
- 159. Thorsson V; Gibbs DL; Brown SD; Wolf D; Bortone DS; Ou Yang T-H; Porta-Pardo E; Gao GF; Plaisier CL; Eddy JA; et al. The immune landscape of cancer. Immunity 2018, 48, 812–830.e14. [PubMed: 29628290]
- 160. Gong C; Anders RA; Zhu Q; Taube JM; Green B; Cheng W; Bartelink IH; Vicini P; Wang B; Popel AS Quantitative Characterization of CD8+ T Cell Clustering and Spatial Heterogeneity in Solid Tumors. Front Oncol. 2018, 8, 649. [PubMed: 30666298]
- 161. D'Esposito A; Sweeney PW; Ali M; Saleh M; Ramasawmy R; Roberts TA; Agliardi G; Desjardins A; Lythgoe MF; Pedley RB; et al. Computational fluid dynamics with imaging of cleared tissue and of in vivo perfusion predicts drug uptake and treatment responses in tumours. Nat. Biomed. Eng 2018, 2, 773–787.
- 162. Stamatelos SK; Kim E; Pathak AP; Popel AS A bioimage informatics based reconstruction of breast tumor microvasculature with computational blood flow predictions. Microvasc. Res 2014, 91, 8–21. [PubMed: 24342178]
- 163. Kirouac DC How do we "validate" a QSP model? CPT Pharmacometrics Syst. Pharmacol 2018, 7, 547–548. [PubMed: 29761661]
- 164. Gadkar K; Kirouac D; Parrott N; Ramanujan S Quantitative systems pharmacology: A promising approach for translational pharmacology. Drug Discov. Today Technol 2016, 21–22, 57–65.
- 165. Workman P; Draetta GF; Schellens JHM; Bernards R How much longer will we put up with 100,000 cancer drugs? Cell 2017, 168, 579–583. [PubMed: 28187281]
- 166. Zhang X; Li Y; Pan X; Xiaoqiang L; Mohan R; Komaki R; Cox JD; Chang JY Intensity-modulated proton therapy reduces the dose to normal tissue compared with intensity-modulated radiation therapy or passive scattering proton therapy and enables individualized radical radiotherapy for extensive stage IIIB non-small-cell lung canc. Int. J. Radiat. Oncol. Biol. Phys 2010, 77, 357–366. [PubMed: 19660879]

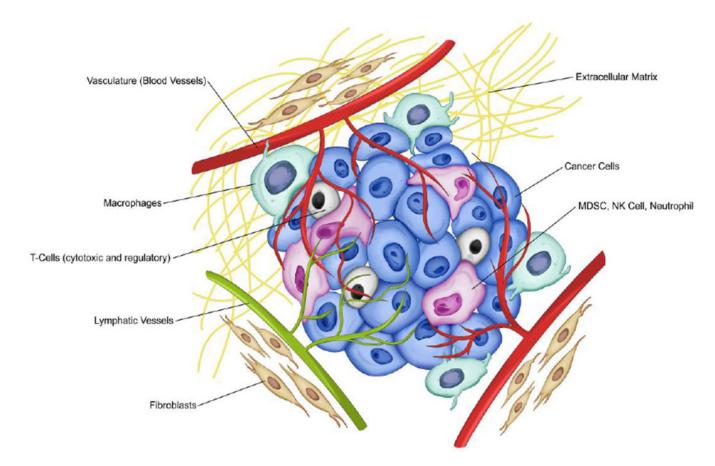


Figure 1. The Tumor Microenvironment (TME). The tumor microenvironment consists of different types of cells (cancer and stromal including immune cells), the extracellular matrix (ECM), and the myriad molecules such as chemokines, cytokines, microRNAs, and growth factors. Cancer cells (including cancer stem cells and progenitor cells), the tumor vessels (blood vessels and lymphatic vessels), immune cells (including tumor-associated macrophages (TAM) and T-cells (cytotoxic and regulatory), myeloid-derived suppressor cells (MDSC), natural killer cells (NK cell), neutrophils and other stromal components (including the extracellular matrix and cancer-associated fibroblasts (CAF)) are shown.

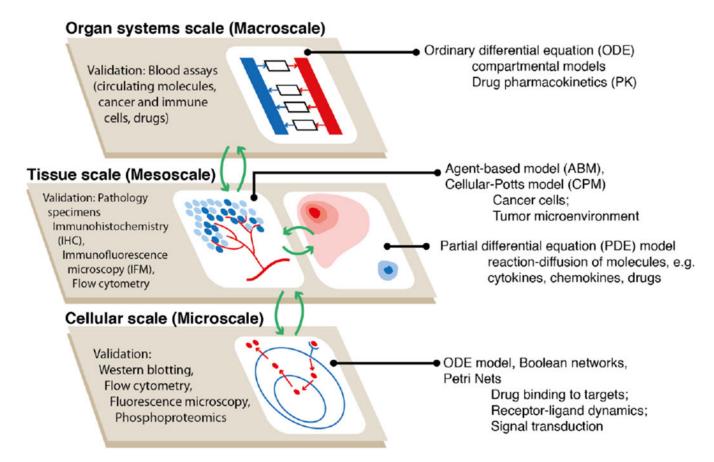


Figure 2.

Using hybrid models to study immuno-oncology. While agent-based models are ideal tools to recapitulate the spatio-temporal dynamics of cancer cells and the tumor microenvironment at the tissue scale, the mechanisms at other biological scales can be efficiently embodied using other types of mathematical representations; however, agent-based models (ABM) can also be used at any scale. Such multi-scale hybrid models increase the flexibility in model construction, improve computational performance, and enhance model credibility by allowing comparison between model output and a wide range of experimental and clinical observations

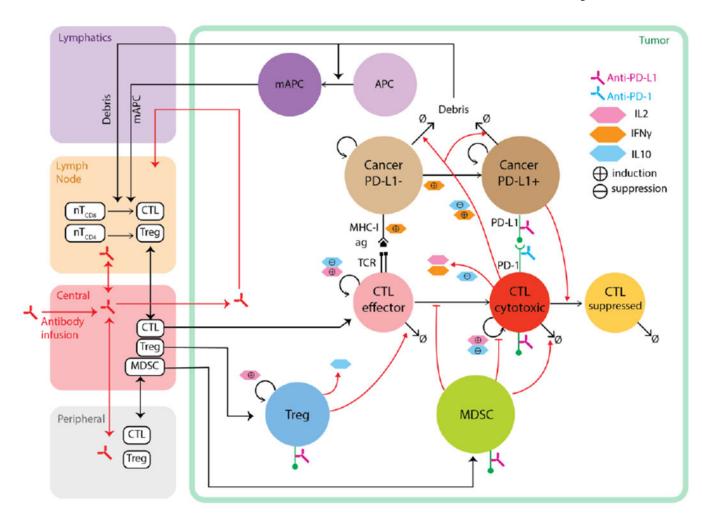


Figure 3.

Diagram of a multi-compartment hybrid model capturing tumor development and anti-tumor immune response. Dynamics of cells and pharmacokinetics of drug (e.g., antibody) in the lymphatics, tumor-draining lymph node, central (blood) and peripheral compartments are modeled using ordinary differential equation systems. Spatial dynamics of cells and molecules in the tumor compartment are captured using agent-based model and partial differential equations. Death of cancer cells produces antigens which drive maturation of APC and their migration to the tumor draining LN, where CD8+ and CD4+ T-cells go through priming and proliferation before they enter blood circulation and extravasate to the tumor microenvironment. Effector CD8+ T-cells can be further activated and expanded when they encounter tumor antigens. These cytotoxic cells kill cancer cells and also release various cytokines, including IL2 which drives further proliferation of T-cells, and IFNy which is proinflammatory and induces PD-L1 expression on cancer cells. PD-L1 can then bind to PD-1 molecules on cytotoxic T-cells, resulting in T-cell exhaustion. Both PD-L1 and PD-1 molecules are potential targets for immune checkpoint blockade antibodies. Regulatory cell types in the ABM include Treg and MDSC, which can inhibit cytotoxic T lymphocytes (CTL) through different mechanisms.

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Table 1.

Summary of Section 3: ABM and hybrid models discussed in each section.

3.6 Models Focusing on Intra- Tumor Heterogeneity	ly Ref	, NK (Pourhasanzade ic T- et al. 2017) 11 [137]	t of (Carmona- a on (Carmona- or Fontaine et al. al 2013) [138]	nne (Figueredo hage, 2011, 2013) cell [139,140] ions	ors er (Figueredo en- 2013, 2014) dent [141,142] ation			
	Study	Tumor, NK cell, cytotoxic T- cell interactions	Effect of stroma on tumor spatial patterns	Immune cell, macrophage, tumor cell interactions	Tumors under oxygen- dependent proliferation			
3.5 Models Focusing on Tumor- Enhancing Immune Cells	Ref	(Hillen 2013) (Enderling 2012) [128,129]	(Wells 2015) [131]	(Knútsdóttir et al. 2014, 2016) [132,133]	(Norton et al. 2018) [134]			
	Study	Cancer stem cell-immune cell interaction	Effect of M1 and M2 macrophages on tumor growth	Signaling between macrophages and cancer cells	Effect of macrophages on TNBC tumor growth			
3.4 Models Focusing on Tumor Immunotherapy	Ref	(Pennisi et al. 2009) [73]	(Pennisi et al. 2010) [75]	(Dréau et al. 2009) [119]	(Pappalardo et al. 2011) [120]	(Walker et al. 2016) [123]	(Gong et al. 2017) [127]	
	Study	Lung met in mammary carcinoma	Effect of vaccine on lung metastasis	Immunotherapy in solid tumors	Role of T-cells in response to immunotherapy	Effect of different therapies on pancreatic tumors	Spatio- temporal dynamics of tumor-immune cell interactions	
3,3 Models Focusing on Tumor- Associated Lymphatics	Ref	(Meyer-Hermann et al. 2002, 2005) [106– 108]	(Bogle et al. 2010, 2012, 2008) [109– 113]	(Moreau, 2016) [113]	(Kim et al. 2009) [100]	(Jacob et al. 2011) [116]	(Marino et al. 2011) [117]	(Marino et al. 2016) [118]
	Study	Germinal centers of LN	T-cell behavior in LN	T-cell activation in virtual LN	Model of LN to study cancer vaccines	Immune response against viruses	Recruitment of APCs in the LN from lung	T-cell trafficking and proliferation
3.2 Models Focusing on Tumor-Associated Vasculature in the Immune Response	Ref	(Uppal et al. 2017) [92]	(Alfonso et al. 2016) [93]					
	Study	Early metastasis	Immune- epithelial cell interactions in breast epithelium					
3.1. Models Focusing on Immune-Related Tumor Mechanobiology	Ref	(Frascoli et al. 2016) [89]	(Kather et al. 2017) [72]					
	Study	Cellular adhesion to ECM	Adoptive cell transfer in colorectal cancer					