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Magnetic Resonance Imaging of the Musculoskeletal System at 7T

Morphological Imaging and Beyond

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Abstract: In 2017, a whole-body 7T magnetic resonance imaging (MRI) device was given regulatory approval for clinical use in both the EU and United States for neuro and musculoskeletal applications. As 7 Tesla allows for higher signal-to-noise, which results in higher resolution images than those obtained on lower-field-strength scanners, it has attracted considerable attention from the musculoskeletal field, as evidenced by the increasing number of publications in the last decade. Besides morphological imaging, the quantitative MR methods, such as T₂, T₂*, T₁, mapping, sodium imaging, chemical-exchange saturation transfer, and spectroscopy, substantially benefit from ultrahigh field scanning. In this review, we provide technical considerations for the assessment of cartilage, tendon, meniscus, and muscle. The first part of the review is dedicated to morphological applications at 7T, and the second part describes the most recent developments in quantitative MRI at 7T.

Key Words: 7 Tesla, cartilage, meniscus, muscle, musculoskeletal, quantitative, tendon, ultrahigh-field

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A lthough 7T magnetic resonance (MR) scanners have been in operation in many large hospitals and research centers for more than a decade, only recently have the regulatory authorities given approval for clinical use. The first European regulatory approval, in July 2017, for a clinical 7 Tesla MRI was followed by the U.S. Food and Drug Administration (FDA), which cleared the first clinical 7 Tesla MRI scanner in North America at the Mayo Clinic in October 2017.^{1,2} Musculoskeletal clinical applications are widespread on standard field strengths, such as 1.5 and 3T MR scanners. Ultrahigh field MR scanners provide, however, significant improvements in signal gain, a shortened scan time, or increasing spatial resolution. In theory, as MR signal

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amplitude is linearly proportional to the net magnetic polarization of the (mostly commonly imaged) hydrogen nuclei, signal should increase by a factor of 2.3 between 3 and 7 Tesla. As the noise also increases slightly between the 2 field strengths (physiological noise increases with the square of the main field strength), the signal-to-noise (SNR) ratio increases approximately by a factor of 2.3 This fact can be exploited either by shortening the scan time for particular protocols or by increasing the spatial resolution while maintaining the other parameters, such as field-of-view, the same. This gain is, however, not straightforward, due to inherent obstacles of ultrahigh field MR scanners. Radiofrequency (RF) deposition, measured by specific absorption rate (SAR), is much higher, which results in greater heating of the tissue. B₁ inhomogeneity is also a considerable problem of ultrahigh field MR scanners, that is, the shortened wavelength (\sim 15cm) causes difficulties in penetrating the body and the scanner does not work in the near field regime anymore.³ In the early days of 7 Tesla, B₁ inhomogeneity and subsequent challenges in 7T coil construction resulted in the lack of dedicated coils for many tissues and organs of interest. However, recent developments, driven by the demand from the MR community, show a positive outlook. In addition to neuroimaging, spectroscopy, and Xnuclei applications, the musculoskeletal (MSK) system is one of the main targets of ultrahigh-field MR. One of the most frequently imaged MSK tissue is articular cartilage. Increased age of the population and the incidence of obesity have led to one of the most prevalent MSK conditions-osteoarthritis (OA). OA is a complex joint disease that, in addition to cartilage, also affects the menisci, the synovium, and other tissues of the joint. Conventional modalities, such as radiographs and morphological MRI, are reliable for the detection of the symptoms of later OA stages (joint space narrowing or cartilage tissue loss). There is, however, a strong demand for techniques that provide information about the early structural changes of cartilage, thereby offering an opportunity for early interventions. Quantitative MR techniques have a great potential to establish the imaging markers for glycosaminoglycan (GAG) loss and collagen fiber remodeling. Ultrahigh field scanners provide the basis for the improvement of GAG-specific techniques [chemical exchange saturation transfer (gagCEST) and sodium MRI)] and collagen-specific techniques (T₂ and T₂* mapping, magnetization transfer). Also, focal cartilage lesions after injuries and repair tissue after cartilage repair surgeries are of interest for imaging modalities to avoid the multiple biopsies required for follow-up of patients after treatment. Ultrahigh field MR is very powerful in imaging rapidly relaxing tissues, including tendons, menisci, ligaments, and bones. Higher SNR at 7T enables an accurate estimation of mono- and multicomponent relaxation rates in tendons, calcified cartilage, and bones. Degenerative disk disease (DDD) has received much literature attention in the last several years, as the incidence of intervertebral disk (IVD) disease has steadily increased. As for tendons, in addition to the acute complete tear, other abnormalities, such as tendinopathy, intratendinous necrosis, and partial tears, are relatively common. Skeletal muscle is another important target tissue for MSK imaging and spectroscopy, as many metabolic and degenerative diseases affect the muscle, and thus, dramatically decrease the quality of life for patients. This review summarizes the most recent developments and findings in both morphological and quantitative 7T MR imaging, which improve the diagnosis and treatment monitoring of MSK disorders.

MORPHOLOGICAL MSK IMAGING AT 7 TESLA

With the advent of 7T MR scanners, it was important to elucidate the benefits that could be obtained from 7T compared with 3T with respect to scan time reduction or increases in spatial resolution. Welsch et al⁴ compared several protocols and dedicated coils for musculoskeletal MR applications for 3T and 7 T. Two standard 2D sequences (PD-TSE and T₁-SE) and 3 isotropic 3D sequences (TRUFI, FLASH, and PD-TSE SPACE) were included for comparison. As for acquisition (scan) time (TA), 7T performed generally faster (by a mean factor of 2); TRUFI and FLASH stood out with more than a 3-fold decrease of TA. As for higher spatial resolution, the authors reported better resolution (by a mean factor of 2) with signal-intense sequences, such as the 3D-FLASH sequence, and an almost 4-fold increase in resolution. They concluded that the spatial resolution at 7T can be increased and acquisition time can be reduced, with superior quantitative and comparable qualitative results compared to 3T, but careful optimization must be performed and dedicated coils have to be used (Fig. 1). In addition to the knee joint, Juras et al systematically analyzed the ankle joint at 7T.⁵ They compared 3T and 7T MRI clinical sequences by quantifying SNR and CNR. Ten volunteers were tested using T1-weighted 3D GRE, as well as T1-weighted 2D SE and PD-weighted 2D FSE. SNR was calculated for various tissues (cartilage, bone, muscle, synovial fluid, Achilles tendon, and Kager fat-pad). CNR was calculated for cartilage/bone, cartilage/fluid, cartilage/muscle, and muscle/fat-pad. There was a significant increase of SNR at 7T compared with 3T on 3D GRE (60.9%) and 2D TSE sequences (86.7%); however, an SNR decrease was observed with 2D SE (25%). Furthermore, there was a CNR increase on 2D TSE and in the majority of 3D GRE images. These authors underlined the fact that protocol optimization is a must, and the use of dedicated coils is necessary to achieve the best possible results.

With respect to pathologies in joint imaging, Springer et al compared quantitative and semi-quantitative parameters [SNR and diagnostic confidence score (DCS)] between 3T and 7T on 40 patients who had knee pain.⁶ DSC was determined using 5 sequences (2D TSE T2 coronal, 2D TSE PD sagittal, 2D TSE PD FS sagittal + axial, 2D TSE STIR coronal, 2D TSE T1 sag) and SNR was determined by 3 morphological sequences (2D T2 TSE, 2D TSE PD, 3D DESS T2). In a quantitative analysis, all tested sequences provided voxel-volume adjusted SNR higher at 7T than at 3T. DCS was evaluated in 4 anatomically defined areas for potential pathological findings and revealed higher values at 7T than at 3T. In conclusion, 7T improved overall DCS in routine MRI, despite some degradation of images due to chemical shift artifacts in nonfat-saturated sequences. Fine structures in joints, as well as subtle lesions in bone, menisci, and cartilage, were more clearly depicted on 7T.

Benerjee et al investigated GRAPPA-based parallel imaging for high-field MRI of osteoporosis and OA and to ascertain whether implementation of parallel imaging with 7T would require modifications in the calibration or reconstruction strategy.² Considering the limited commercial availability of array coils for MSK applications at 7T, the feasibility of a quadrature coil with 2 elements was evaluated. A modified 3D fs-SPGR, 3D FIESTA-c, and m-bSSFP were used for cartilage. The m-bSSFP sequence was used for the trabecular bone microarchitecture. These authors concluded that the feasibility of in vivo high-resolution MRI at 7T can be considerably increased using parallel imaging because of reduced RF power deposition and increased flexibility in protocol design. Krug et al have investigated the benefits of ultrahigh field MRI at 7T for the examination of knee cartilage and trabecular bone using a fully balanced, steady-state, free-precession (bSSFP) sequence.³ They showed that bSSFP and spin-echo imaging is possible at 7T within an SAR limit of 3.2 W/kg. Cartilage SNR of the bSSFP sequence was 2.4 times higher at 7T than at 3T. Trabecular bone SNR increased only minimally compared with 3T. In conclusion, they emphasized that cartilage and bone imaging benefits in SNR increased at 7T. Chang et al have pointed out the problem of trabecular bone microarchitecture with hip imaging at 7T.⁴ The advantages of higher SNR at 7 Tesla have translated into the visualization of individual trabeculae within the proximal femur, employing a T₁-weighted fast low



FIGURE 1. The coronal fat saturated (fs) 2D proton-density turbo-spin-echo (PD TSE) sequence is provided for 3 T (left), 7 T quick (middle), and 7 T high-resolution (right) measurements. The medial femoral condyle with the medial meniscus is enlarged for better visualization of the image quality. Reprinted with permission from *Eur Radiol* 2012; 22:1852–1859.



FIGURE 2. A, T₂ map acquired by CPMG, pseudo-color-coded, and overlaid on a morphological T₂-weighted image from a 32-year-old healthy volunteer (TE = 11.9 ms); B, T₂ map acquired by 3D-TESS, pseudo-color-coded, and overlaid on the second acquired MR image (F₀). The typical zonal stratification in cartilage is observable with both techniques. However, T₂ (CPMG) provides apparently higher dynamic contrast of T₂ values with better depiction of cartilaginous layers. Reprinted with permission from *Eur Radiol* 2016; 26:1905–1912.

angle shot 3D FLASH sequence for bone microarchitecture imaging. Furthermore, they devised a T_1 -weighted 3D FLASH with water excitation and volumetric interpolated breath-hold technique (VIBE) with saturation or inversion recovery based, fat suppression for cartilage imaging. Visualization of cartilage, labrum joint capsule, and tendons was achieved using a 2D intermediate-weighted fast spin-echo sequence, which can be readily applied for the clinical imaging of hip. It may allow insight into the pathogenesis of different hip disorders and to detect changes in the hip joint at its earliest stages.

QUANTITATIVE MRI

T₂ Mapping

Cartilage T₂ reflects the interaction of water and the extracellular matrix on a molecular level, particularly changes in collagen and water content and anisotropy of tissue.⁵ The orientation dependence of T₂ relaxation time observed in collagen-rich tissues originates in residual dipolar interactions of water protons due to their restricted motion in the collagen network. Another source of T₂ anisotropy was attributed to intermolecular dipolar interaction between water protons and those of biopolymers.⁶ In healthy articular cartilage, T₂ values differ from deep to superficial cartilage layers depending on the collagen orientation. To visualize cartilage zonal stratification in vivo, high in-plane resolution and short scan times are necessary. In general, T₂ is useful for evaluating cartilage hydration, progression in OA, or assessing damage or cartilage health after physical trauma to a joint, or for surgical procedures.⁷⁻¹⁰ Welsch et al¹¹ showed the feasibility of delayed gadolinium-enhanced MRI of cartilage (dGEMRIC), and T₂ and T₂* mapping to monitor patients after MACT. They observed a significant zonal stratification in healthy cartilage, although almost no stratification in MACT repair sites. In another study from the same group, the authors compared T_2 , T_2 *, and magnetization ratio between 3 and 7 Tesla.¹² In 17 knees of healthy volunteers, there were significantly lower global values at 7T than at 3T. The zonal stratification, however, was less pronounced at 7 Tesla and the coefficients of variation, indicating that reproducibility was slightly lower at 7T. As suggested in early works on quantitative MR of cartilage at 7 Tesla, the lower reproducibility was due to the lack of dedicated extremity coils. Chang et al compared a birdcage-transmit, 28-channel receive array with a quadrature volume coil for 7T for T₂ mapping and they observed 17% to 400% increased SNR/CNR when using a 28-channel knee coil that enabled more reliable morphological imaging and T₂ mapping.¹³ In thinner cartilage, such as in the hip or ankle joint, where the resolution is even more important, an ultrahigh field scanner may be of great interest. Domayer et al¹⁴ demonstrated, for the first time, the capability of 7T MRI T₂ mapping in talus cartilage, even detecting the bilayer zonal pattern of T₂ in the volunteers in these rather thin cartilage layers compared with the knee joint cartilage layers (Fig. 2). T₂ values found in the superficial and the deep layer were 39.3 \pm 5.9 and 21.1 \pm 3.1 ms, respectively. The field-of-view was $140 \text{ mm} \times 140 \text{ mm}$ and the matrix was 320×320 , resulting in an in-plane resolution of 0.4×0.4 mm with a slice thickness of 3 mm. Moreover, they compared T₂ values of 2 different surgical methods: in MFX, T₂ values in the superficial/ deep zone were 43.4 \pm 10.5 and 36.3 \pm 7.7 ms, and, in MACT, 39.0 ± 9.1 and 27.1 ± 6.6 ms.

The thin cartilage of the hip joint, together with the complex anatomy, is also a challenge in terms of T_2 mapping. Lazik et al¹⁵ showed the feasibility of T_2 , T_2* mapping, and dGEMRIC in the hips of 11 healthy volunteers. They used T_2 and T_{2*} mapping in the sagittal view with an FOV of $160 \times 160 \text{ mm}^2$, a matrix of 320×320 , and a slice thickness of 2.5 mm. Echo times used were 10.1, 20.2, 30.3, 40.4, and 50.5 ms for T_2 mapping (TR 1500 ms) and 3.06, 8.0, 12.94, 17.88, and 22.82 ms for T_{2*} mapping (TR 130 ms). They found a high correlation between T_2 and T_{2*} in acetabular cartilage (P = 0.009) and femoral cartilage (P = 0.0002) and described regional differences of T₂ and T₂*.¹⁵

The standard multiecho spin echo (MESE) sequence for T₂ mapping has many limitations when used at ultrahigh field MR. As it acquires signals from multiple time points along the T₂ decay for each k-space line during a single repetition time, it suffers from the contamination by stimulated echoes and easily reaches SAR limits due to multiple pulse utilization. A recently introduced triple-echo steady-state sequence (TESS) provides fast T1 and T2 quantification within a single scan, and, in particular, B1-insensitive T2 calculation.¹⁶ Juras et al¹⁷ used this sequence to estimate T₂ values in knee cartilage to compare the conventional T2 mapping MESE technique with TESS. Using similar sequence parameters for both techniques (slice thickness 3 mm, FOV 160×160 mm, matrix size 320×320 , slice spacing 3 mm), they set up a "standard" TESS protocol with a TA of 4:35 minutes and a "quick" protocol with a TA of 2:05. Although T₂ values calculated from TESS were generally lower (total mean of 31 ± 5 ms) than MESE (total mean of 46 ± 9 ms), the correlation of T₂ values between these 2 approaches was very high (ranging from 0.750 to 0.911 depending on the region) (Fig. 3). This study demonstrated the clinical utility of a TESS sequence and concluded that TESS provides results similar to those of a conventional MESE sequence with many benefits, such as shortening of total acquisition time, lowering SAR demands, and insensitivity to B₁ changes. A similar comparison was done by Kraff and Lazik¹⁸ in hip joint cartilage with the aim to investigate whether this sequence would be a good candidate for improving workflow in quantitative hip MRI protocols at 7 T. For TESS, these authors used a slice thickness of 2.5 mm, an FOV of 180 × 180 mm, a matrix size of 384×384 , 32 slices, and a TA of 6 minutes, and, for the MESE, a slice thickness of 2.5 mm, a FOV of 160×160 mm, a matrix size of 320×320 , 6 slices, and a TA of 4:53 minutes. Interestingly, they found much lower correlation coefficients between TESS T2 values and those acquired with MESE, ranging from 0.3 to 0.8. T₂ values were higher when using CPMG-this was attributed to the

contributions from stimulated echoes as a result of imperfect refocusing pulses. The authors reported that TESS was most advantageous in clinical studies where the measurement time is a critical factor and where the B₁ optimization is challenging. Recently, TESS has been used to assess axonal bundles (fascicles) in the median nerve and to determine the normative T₂ values.¹⁹ That study found different T₂ values between 2 subject groups, with significantly higher (P = 0.023) values in patients with idiopathic carpal tunnel syndrome (24.27 ± 0.97 ms) than in healthy volunteers (21.01 ± 0.65 ms). It was possible to image the relatively small median nerve due to the high resolution provided by a 7T scanner.

T₂* Mapping

Some tissues with high organization of collagen fibers, such as tendons, ligaments, and menisci usually appear black on conventional MR images due to the fast transverse relaxation. To acquire signal from these tissues, a very short echo time must be used. In the past, many k-space trajectories that allowed for an ultrashort echo were developed, such as SPRITE,²⁰ radial UTE,²¹ 3D cones,²² or a stack of spirals (AWSOS).²³ The 7T scanner enables signal acquisition using ultrashort techniques with higher SNR than clinical scanners. This allows for more robust mono- and bi-exponential T₂* mapping of connective tissues. Juras et al²⁴ used 3D-UTE at 7T to demonstrate the regional variation of both mono- and bi-exponential T₂* in the Achilles tendon. They found significant differences between the tendon insertion, the mid portion, and the musculotendinous junction, which were further separated into anterior, posterior, medial, and lateral regions. Moreover, the short T2* component was found to be a reliable marker for tendinopathies, and thus, may provide additional information to standard clinical imaging in reasonable scan times. Some metabolic diseases, such as diabetes, alter the collagen content and organization in tendons, which can be detected by UTE sequences.²⁵ Han et al²⁶ compared the performance of the UTE sequence in depicting the tendon microstructure between 3 and 7T. Proper setup of sequence parameters, overcoming



FIGURE 3. Example of the ROI setting in a case after MACT in the repair site (A) and the reference (B). Both ROIs are set at the same orientation to the static magnetic field. The deep (1) and superficial (2) ROIs are indicated by the white boxes. This case shows T2 values in the repair tissue that are comparable to the articular cartilage both in the deep and the superficial layer. Reprinted with permission from *Osteoarthritis Cartilage* 2012; 20:829–836.

increased B₀ and B₁ inhomogeneities, and increased RF power deposition using dedicated coils at 7T was necessary to gain higher SNR at 7T versus 3T, as expected. Hager et al²⁷ used a micro-MRI insert at 7T to investigate the orientation dependence of T₂* in the knee meniscus. With a resolution approximately 100 times higher than that which is used clinically, they were able to describe the meniscal structures and the strong dependence of T₂* values on orientation toward B₀. They also reported no bi-exponential decay on a pixel-by-pixel basis. The previously published bi-component T₂* decay in the meniscus probably originated because of the low resolution of analyzed images, which resulted in multiple tissue types contributing to a given voxel.²⁸

T₁ρ Mapping

Previous works demonstrated the potential of $T_{1\rho}$ to serve as a proteoglycan-specific measure in articular cartilage.^{29–31} Unlike sodium MRI and dGEMRIC, there is no need for special hardware or contrast agent administration while using this technique. The cartilage repair community, however, has been disputing the actual sensitivity of $T_{1\rho}$ to proteoglycans and its clinical applicability for many years. Keenan et al³² found, for instance, that $T_{1\rho}$ relaxation time is inversely proportional to proteoglycan content only in cartilage regions with normal T₂ relaxation. 7T scanners, with higher SNR, and thus, higher resolution, may help investigate this issue further. Singh et al³³ demonstrated the feasibility of T_{10} mapping at ultrahigh field MR with high SNR (\geq 90), and an in-plane resolution of 0.2 mm², and compared it to 3T measurements. They found 15% lower $T_{1\rho}$ values at 7T than at 3T, with the SNR and resolution substantially in favor of 7T. Moreover, 7T MRI helped to overcome the issues with SAR constraints and B_0 and B_1 inhomogeneities. Wyatt et al³⁴ studied 20 OA symptomatic subjects to validate 7T T_2 and T_{10} mapping as markers for differentiating between OA patients and controls. At both field strengths, T_2 and T_{1p} were higher in the lateral femoral condyle and patella in patients with OA, but more regions were significant or approached significance at 7T.³⁴

CHEMICAL EXCHANGE SATURATION TRANSFER

Glycosaminoglycan-specific chemical exchange saturation transfer (gagCEST) is a novel technique, which has the potential to provide information about the amount of GAGs in various cartilaginous structures. It uses selective saturation of the exchange able hydroxyl protons of GAG,³⁵ which exchange with water protons. The performance of this method increases with the magnetic field due to the larger difference in resonance frequencies of the hydroxyl GAG protons and those of water. Along with the larger SNR ratio at 7 Tesla, the larger frequency difference at this field increases the selectivity of saturation and, consequently, the gagC-EST effect. The main drawbacks of the gagCEST methodology are the small magnitude of this effect and its sensitivity to subject motion.

The first gagCEST data on articular cartilage at 7T were reported by Schmitt et al³⁶ and Sigh et al,³⁷ followed by a clinical study that combined 7T gagCEST and sodium MRI in patients with autologous osteochondral transplantation.³⁸ Another clinical study of cartilage adjacent to the site of repair surgery showed significant differences in the gagCEST effect compared with reference cartilage.³⁹ The gagCEST effect at 7T in the tibio-talar joint was also compared with the T_{1p} data.⁴⁰ In a study by Lee et al,⁴¹ simultaneous saturation with 2 distinct frequencies enabled better isolation of the CEST effect in the knee cartilage. In another methodological study, dynamic correction of the frequency drift was suggested, which enabled the measurement of more realistic gagCEST values.⁴² Parameters for the gagCEST pulse sequence and protocol at 7T were optimized for the best clinical performance in several studies.^{43–45} It was shown that the 7T gagCEST effect in knee cartilage had a moderate-to-good reproducibility when averaged over larger regions of interest. However, a surprisingly large range of the gagCEST values was reported even in healthy cartilage of femoral condyles.⁴⁵ A reason for that might be the variation in the water T2 relaxation time, which can affect the measured gagCEST values in articular cartilage at both 3T and 7T.⁴⁶

SODIUM MRI

Technical Considerations

Previously published studies have demonstrated that sodium MR imaging can serve as a noninvasive method for direct determination of the GAG content in cartilage, which plays a crucial role in cartilage homeostasis. New technical developments in the last decade have helped to transfer this method from in vitro to pre-clinical in vivo studies. Sodium imaging has already been applied for the evaluation of cartilage and repair tissue in patients after various cartilage repair surgery techniques and in patients with OA. Sodium MRI, however, is extremely technically challenging. There are a number of reasons for this: first, the sensitivity of sodium MR is only 9.3% of that of proton MR; second, the sodium concentration in healthy cartilage is more than 300 times lower than the concentration of protons; and third, sodium has very short biexponential transverse relaxation times in biological tissues. The resultant SNR is thus more than 3000 times lower in sodium MR than proton one. Ultrahigh field MR scanners equipped with dedicated resonators may help to overcome these challenges; nevertheless, the resolution is much lower and total scan times much longer than those of conventional proton MR imaging. Besides the technical difficulties with sodium MRI, the post-processing of the sodium signal also represents an obstacle to be overcome. The precision of the sodium content measurement is strongly affected by partial volume effects that originate from surrounding tissues, such as bone or synovial fluid. To correctly calculate sodium concentration in articular cartilage, the sodium MR signal must be converted to a sodium concentration. For that, a series of phantoms with known sodium concentration needs to be scanned along with the subject and the calibration curve is calculated by linear fitting of the sodium signal.⁴⁷ As the relaxation times of sodium in healthy and damaged cartilage may vary, this also has to be considered during the evaluation. Chang et al 48 used a relaxation times (T1, T2*short, T2*long)-corrected method to validate the usefulness of inversion pulses to suppress the synovial fluid in the assessment of native and repair cartilage. In another study, the authors clearly demonstrated a linear increase of SNR between 3T and 7T using a 3D cone pulse sequence.⁴⁹

Osteoarthritis

Osteoarthritis is a target disease where sodium MRI can play an important role in diagnosis and treatment monitoring in the future. Early-stage OA assessment, when disease progression might be reversible, is of major interest in cartilage imaging. At the beginning of OA, a repair phase accompanied by hypertrophy occurs, resulting in softening of the articular cartilage due to increased water content as a secondary effect of GAG loss.⁵⁰ To detect this loss with a noninvasive imaging method is extremely desirable. The GAGs contain a high concentration of negatively charged sulfate and carboxyl groups, which results in a negative fixed charge density (FCD) of the articular cartilage. As early as the late 1960s, it was shown that there is a correlation between FCD and sodium concentration as a result of the equilibrium between positively charged sodium ions and negatively charged GAG molecules.⁵¹ Sodium MRI has great potential to detect subtle GAG content changes due to its specificity to GAG concentration in articular cartilage. One of the



FIGURE 4. Sodium maps from 1 OA patient, reconstructed from data acquired with fluid suppression (sequence IRW) and without fluid suppression (sequence R3D), at baseline (scan 1) and at the 16-month follow-up (scan 2). A, Transverse slices showing patellar cartilage (PAT). B, Coronal slices showing femorotibial lateral (LAT) and medial (MED) cartilage. The apparent sodium concentrations (ASC) measured with R3D look very similar in both scans, while a higher difference of ASC can be detected visually with IRW in PAT, MED, and LAT. Reprinted with permission from *Radiology* 2018; 28:133–142.

first works to discover the link between FCD maps and proteoglycan loss in early-stage OA was published by Wheaton et al⁵² in 2004. Using a 4T MR scanner, they found a mean FCD of -182 mmol/L \pm 9 in healthy controls and a decreased FCD with mean values ranging from -108 to -144 mmol/L in symptomatic subjects, which was indicative of proteoglycan loss from the articular cartilage matrix. The first sodium application for articular cartilage on a 7T scanner was reported by Wang et al⁵³ in 2009. On 5 controls and on 5 OA patients, they demonstrated the feasibility of sodium MR using a quadrature knee coil and a 3D gradient echo pulse sequence with a radial acquisition. In OA patients, the sodium concentration was reduced significantly by -30% to 60%, depending on the degree of degeneration. Moreover, this was the first study that showed the possibility of reducing sodium MR scan time to a clinically acceptable time of ~15 minutes. Sodium MRI as an OA marker was validated for the first time in a study involving 19 healthy volunteers and 28 symptomatic patients, using fluid suppression and adiabatic pulses.⁵⁴ Later, the same investigators asked the same patients to undergo a follow-up examination and 12 of the original cohort agreed. The results of this subsequent study demonstrated the sensitivity of sodium MRI to detect the changes in GAG content induced by progressive OA over time.⁵⁵ Figure 4 depicts the comparison of the sodium examinations of a patient 16 months apart.

Cartilage Repair

Cartilage has a very limited capability for self-repair, and the subsequent degeneration often develops into OA. To repair cartilage damage, there are a number of surgical options available, including microfracturing (MFX),⁵⁶ osteochondral graft transplantation,⁵⁷ and autologous chondrocyte implantation (ACI).⁵⁸ The next generation of ACI techniques is often referred to as matrix-associated autologous chondrocyte transplantation (MACT) using hyaluronan, polylactides, or other materials as scaffolds for cell growth.⁵⁹ Maturation of cartilage repair requires multiple follow-ups to monitor the GAG content in the repair site. Sodium MRI was used as a marker for the maturation grading in 2010 by Trattnig et al.⁶⁰ Twelve patients were involved in the study, and all of them were scanned 56 ± 28 months

after they underwent a MACT procedure. A 3D GRE pulse sequence with a ²³Na transmit-receive knee coil was used. A significantly different normalized sodium signal was observed between the cartilage transplant (174 ± 53 a.u.) and healthy cartilage (267 ± 42 a.u.). Subsequently, the same group compared 2 surgical methods for cartilage repair (MFX and MACT) using ²³Na MRI at 7 Tesla.⁶¹ The study involved 9 MFX patients and 9 MACT patients, with postoperative intervals of 33.5 ± 25.3 and 33.2 ± 25.7 months for MFX and MACT, respectively. The results showed that the normalized sodium signal was significantly lower in BMS and MACT repair tissue, compared with reference cartilage. Moreover, the normalized sodium was significantly higher in MACT than in MFX repair tissue, suggesting better quality in MACT than in MFX. In another study by Krusche-Mandl et al,³⁸ the authors followed-up patients 8 years after autologous osteochondral transplantation using gagCEST, sodium MRI, and T₂ mapping. Interestingly, of the 3 biochemical MR methods, only T₂ correlated with clinical outcome. Technical developments in sodium MR allowed acquisition of the signal from joints with thinner cartilage, as well, such as the ankle joint. Zbyn et al⁶² demonstrated the feasibility of ²³Na imaging for monitoring patients after MACT and MFX in the ankle and subtalar joints.

Sodium MRI of Tendons and Spine

Sodium MRI has also found a broader application for other structures, such as tendons and IVDs. The normalized sodium MR signal may serve as a marker for patients with Achilles tendinopathy by detecting the increased proteosynthesis (which results in increased GAG content) of the affected tendon. Juras et al used 7T whole-body MRI with a 15-channel sodium knee coil to investigate 8 patients with tendinopathy and 20 controls. They found increased sodium SNR in patients (9.3 ± 2.3), indicating an elevated GAG content, compared with healthy controls (4.9 ± 2.1) (Fig. 5).⁶³ The same group investigated sodium MRI as a potential marker for biochemical changes induced by side effects in the Achilles tendon after ciprofloxacin intake.⁶⁴ It has been reported in the orthopedic community that the intake of ciprofloxacin may lead to tendon harm with the development of tendinopathies and even partial or complete



FIGURE 5. MR images in a 27-year-old healthy volunteer. A, Proton-density-weighted, 2-dimensional, turbo spin-echo image depicts the centers of each ROI (arrows). B, Corresponding color-coded, 3-dimensional, gradient-echo sodium image depicts Achilles tendon sodium signal (arrows), which is minimal. MR images in a 46-year-old patient with chronic Achilles tendinopathy. C, Proton-density-weighted, 2-dimensional, turbo spin-echo image depicts centers of each ROI (arrows). D, Corresponding color-coded, 3-dimensional, gradient-echo sodium image depicts centers of each ROI (arrows). D, Corresponding color-coded, 3-dimensional, gradient-echo sodium image depicts Achilles tendon sodium signal (arrows), which is obviously higher than that in the healthy volunteer. The sodium signal increase was observed in the whole tendon, and not only in regions with clinical findings (middle region in this case). All patients had focal thickening of the Achilles tendon without a fluid-like signal intensity increase on proton-density-weighted, 2-dimensional, turbo spin-echo images. The color scale represents sodium signal intensity values. Reprinted with permission from *Radiology* 2012; 262:199–205.

tendon tears. They scanned patients 10 days before the ciprofloxacin treatment, and 10 days and 5 months after treatment. The mean sodium signal decreased significantly, by almost 25%, in the whole tendon [from baseline to 10 days after ciprofloxacin intake, 130 arbitrary units (au) ± 8 to 98 au ± 5 , respectively; P = 0.023] and returned closer to baseline after 5 months (116 au \pm 10). A similar behavior of the sodium signal was also found in tendon insertion. This study demonstrated a ciprofloxacin-induced reversible decrease of the normalized sodium MR imaging signal in the Achilles tendon of healthy volunteers.

The IVD containing GAG is also an interesting tissue for sodium MRI. Noebauer-Huhmann et al⁶⁵ performed a study in 10 asymptomatic volunteers (mean age, 30 years; range, 23 to 43 years) to validate the feasibility of sodium MR in IVD and to find the correlation between normalized sodium signal intensity and T₂ mapping/Pfirrmann score. The Pearson coefficient showed a correlation between sodium imaging and the Pfirrmann score, a moderate negative correlation between T₂ values and the Pfirrmann score (r = -0.62), and no correlation between sodium imaging and T₂ mapping (r = 0.06). This was the first study to demonstrate the feasibility of in vivo 7T MR sodium imaging of the IVD, and to determine the link between quantitative values and clinical score. This might be interesting for future in vivo studies of disk composition and the mechanisms of degenerative disk disease.

Future Outlook

Modern methods, such as machine-learning, may play a crucial role in sodium MRI evaluation and bring it closer to clinical use by overcoming the tedious manual assessment of low-resolution and low SNR sodium images. Madelin et al⁶⁶ at New York University recently suggested a machine-learning method for the assessment of the possible utility of machine-learning to classify subjects with and subjects without OA. In this study of 19 controls and 28 OA patients, they were able to isolate the best OA predictors with an accuracy as high as 74%.⁶⁶ In other work by the same group, compressed sensing was applied to under-sample a 3D radial pulse sequence for sodium MRI at 7 Tesla. With this technique, it is possible to decrease the scan time by a factor of 2 without losing accuracy in sodium concentration detection over different regions in order to detect early signs of OA. The best results were achieved by a combination of compressed sensing and nonuniform Fast Fourier Transform (NUFFT).

MRI and MRS of Muscle

Muscle MRI provides valuable information about many aspects of the structure and function of skeletal muscles, such as the presence of edema or fatty infiltration, or changes in mitochondrial metabolism or tissue perfusion. This makes muscle MRI a very powerful tool in the diagnosis and follow-up of patients with several muscle diseases, for example, muscular dystrophies,⁶⁷ and/or whole-body metabolic disorders.⁶⁸ The next few paragraphs will describe several muscle MRI techniques that benefit from the increased SNR of ultrahigh field systems.

Structural Imaging

Muscle structural MRI is primarily focused not only on detecting edema and fatty infiltration^{67,69} but can be also used to identify skeletal muscle denervation⁷⁰ or measure the muscle architecture.⁷¹ The fatty infiltration observed in muscle MRI is of particular interest in limb-girdle muscle dystrophies, as this has been shown to correlate with muscle strength and functional status.⁷² Fatty infiltration can be identified by T_1 -weighted imaging, the quantification of which can be challenging,⁷³ or, by the more quantitative Dixon technique.⁷⁴ The Dixon technique utilizes the in-phase/out-of-phase cycling of fat and water. The in-phase and out-of-phase images can be combined to create fat-only and water-only images and provide a proton density fat fraction.⁷⁵ Alternatively, proton magnetic resonance spectroscopic imaging (¹H-MRSI) can be used to define not only fatty infiltration of muscle tissue but also of muscle cells themselves,⁷⁶ which is particularly important in type 2 diabetes mellitus.⁷⁷ The separation of extra- and intramyocellular lipids grows larger at ultrahigh fields and the increase in SNR permits high spatial resolution acquisition in a reasonable time.⁷⁸

Edema and denervation are quite identifiable on T_2 -weighted images, and better still on short inversion time inversion-recovery (STIR) MR images.^{67,70} As the chemical shift separation between fat and water is pronounced at ultrahigh fields, the length of fat saturation pulses can be substantially shortened.⁷⁹ Note that simple qualitative or semi-quantitative comparisons could lead to data misinterpretation, and thus, proper quantitation methods have to be applied to provide information about disease severity.⁸⁰

Direct information about the spatial architecture of skeletal muscle tissue can be obtained by diffusion-weighted MRI, which exploits the restricted diffusion of water molecules along the muscle fibers. Acquiring several diffusion-weighted images allows full characterization of the diffusion tensor.⁸¹ The diffusion tensor imaging (DTI) approach yields a number of indices that closely define the anisotropic organization of skeletal muscle tissue,⁷¹ providing an interesting tool for the investigation of muscle injuries.⁸² It has been demonstrated that the reliability of estimating DTI parameters is substantially affected by the SNR and the percentage of fat composition (%fat).^{83,84} In particular, low SNR data may result in an overestimation of the eigenvalues and an underestimation of the mean diffusivity, making DTI a very good target for ultrahigh field MRI, which provides increased SNR.^{85,86}

Functional Imaging

In addition to structural information, muscle MRI also provides information about muscle energy metabolism through the utilization of phosphorus (³¹P) MR.⁸⁷ The high-energy phosphate bonds in adenosine-triphosphate (ATP) and phosphocreatine, both readily detectable by ³¹P-MRS, serve as an energy source for muscle cells and for the rapidly available energy reserve for ATP resynthesis, respectively. Furthermore, by exploiting the chemical shift difference between PCr and inorganic phosphate (Pi) resonances, intramyocellular pH can be calculated.⁸⁸ But, the true strength of ³¹P-MR of skeletal muscles lies in the fact that it allows noninvasive assessment of the rates of chemical reactions involved in energy metabolism.⁸⁹ Of particular interest is the possibility to investigate the oxidative energy production by muscle mitochondria during exercise and subsequent recovery.^{87,89} This has been repeatedly shown to be impaired in patients with muscle dystrophies,⁹⁰ peripheral arterial disease,^{91,92} heart failure,⁹³ and in systemic disease, such as obesity⁹⁴ and type 1 and 2 diabetes mellitus,^{95,96} and is also of great interest in sports medicine.⁶⁸

The inherently low SNR of ³¹P-MR on clinical scanners typically leads to the use of surface coils and limits the choice of sequences to nonlocalized acquisitions.⁸⁷ However, it has been recently shown that such rough localization is not sufficient for exercise-recovery studies, if several differently active muscle groups are detected simultaneously.^{97,98} The use of ultrahigh fields significantly improves the SNR of ³¹P-MR, allowing advanced multi-voxel localization schemes with sufficient temporal resolution to be used for dynamic studies.^{99–102} High spatial resolution can be achieved through spectrally selective MRI; however, it is typically limited to PCr dynamics measurement only,⁹⁹ and losing all other information present in the spectra. Although recent developments have allowed the information about intramyocellular pH to be extracted from ³¹P-MRI as well,^{100,103} still, more information can be achieved either by acquiring a rapidly sampled MRSI matrix¹⁰¹ or by interleaving 2 single-voxel excitations.¹⁰²

The ability of skeletal muscle mitochondria to synthesize ATP is highly reliant on oxygen supply. Therefore, the dynamics of muscle tissue perfusion, quantifiable by arterial spin labeling (ASL), are of great interest, particularly in patients with peripheral arterial disease.¹⁰⁴ Signal decay during inflow time due to T₁ relaxation, and



FIGURE 6. ³¹P real-value MR images acquired in a subject during the exercise-recovery experiment (averaged over 3 acquisitions). PCr (A) and Pi (B) at rest. The lower PCr signal during exercise (C) is accompanied by a detectable Pi signal (D) in the gastrocnemius muscles. E, A pH image obtained during exercise (averaged over 5 acquisitions) showing different pH values within the depicted ROIs of the examined muscle groups (gastrocnemius medialis, green; gastrocnemius lateralis, blue; and soleus, red), overlaid over a water image. Reprinted with permission from *Magn Reson Med* 2016; 75:2324–2331.

low SNR, often limit ASL investigations. The use of ultrahigh fields mitigates both these limitations¹⁰⁵ and allows the combination with T_2 *-weighted imaging that provides complementary information about blood oxygenation.¹⁰⁶

Muscular dystrophies, inherited myopathies, muscle injury, as well as metabolic diseases can be identified and their treatment monitored by skeletal muscle MRI. Both structural and functional changes can be quantified, and together, provide complex information about muscle health. Increased SNR and chemical shift separation of ultrahigh fields significantly increase the accuracy, specificity, and reproducibility of muscle MRI examinations.

CONCLUSION

The higher SNR, more reliable quantitative techniques, the improved contrast, and better X-nuclei MR capability at 7T translate into better spatiotemporal resolution and enhanced biochemical and metabolic tissue characterization. Thus, 7 Tesla MR imaging improves anatomic detail and supports a higher degree of diagnostic confidence when imaging cartilage and other joint tissues. Recent developments in quantitative MR techniques at 7T show the potential to include such techniques into routine clinical imaging, thus helping to merge morphological and compositional information. Clinically approved 7 Tesla MRI scanners will definitely propel the innovations in medical imaging in the future.

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REFERENCES

- medgadget.com. EU Gives First Approval for Ultra-High-Field MRI Scanner, The Siemens Magnetom Terra. 2017. Available at: https:// www.medgadget.com/2017/08/eu-gives-first-approval-ultra-high-field-mriscanner-siemens-magnetom-terra.html. Accessed December 6, 2018.
- medgadget.com. FDA Gives First Clearance to Siemens High-Field 7 Tesla MRI Scanner. 2017. Available at: https://www.medgadget.com/2017/10/fdagives-first-clearance-high-field-7-tesla-mri-scanner.html. Accessed December 6, 2018.
- Ugurbil K. Magnetic resonance imaging at ultrahigh fields. *IEEE Trans Biomed Eng.* 2014;61:1364–1379.
- Welsch GH, Juras V, Szomolanyi P, et al. Magnetic resonance imaging of the knee at 3 and 7 Tesla: a comparison using dedicated multi-channel coils and optimised 2D and 3D protocols. *Eur Radiol.* 2012;22:1852–1859.
- Mosher TJ, Dardzinski BJ. Cartilage MRI T2 relaxation time mapping: overview and applications. *Semin Musculoskelet Radiol.* 2004;8:355–368.
- Hanninen N, Rautiainen J, Rieppo L, et al. Orientation anisotropy of quantitative MRI relaxation parameters in ordered tissue. *Sci Rep.* 2017;7:9606.
- Keenan KE, Besier TF, Pauly JM, et al. Prediction of glycosaminoglycan content in human cartilage by age, T1 rho and T2 MRI. *Osteoarthritis Cartilage*. 2011;19:171–179.
- Nieminen MT, Rieppo J, Toyras J, et al. T-2 relaxation reveals spatial collagen architecture in articular cartilage: a comparative quantitative MRI and polarized light microscopic study. *Magn Reson Med.* 2001;46:487–493.
- Liess C, Lusse S, Karger N, et al. Detection of changes in cartilage water content using MRI T2-mapping in vivo. *Osteoarthritis Cartilage*. 2002;10:907–913.
- Mlynarik V, Trattnig S, Huber M, et al. The role of relaxation times in monitoring proteoglycan depletion in articular cartilage. *J Magn Reson Imaging*. 1999;10:497–502.

- Welsch GH, Mamisch TC, Hughes T, et al. In vivo biochemical 7.0 Tesla magnetic resonance: preliminary results of dGEMRIC, zonal T2, and T2* mapping of articular cartilage. *Invest Radiol.* 2008;43:619–626.
- Welsch GH, Apprich S, Zbyn S, et al. Biochemical (T2, T2* and magnetisation transfer ratio) MRI of knee cartilage: feasibility at ultra-high field (7T) compared with high field (3T) strength. *Eur Radiol.* 2011;21:1136–1143.
- Chang G, Wiggins GC, Xia D, et al. Comparison of a 28-channel receive array coil and quadrature volume coil for morphologic imaging and T2 mapping of knee cartilage at 7T. J Magn Reson Imaging. 2012;35:441–448.
- Domayer SE, Apprich S, Stelzeneder D, et al. Cartilage repair of the ankle: first results of T2 mapping at 7.0 T after microfracture and matrix associated autologous cartilage transplantation. *Osteoarthritis Cartilage*. 2012;20:829– 836.
- Lazik A, Theysohn JM, Geis C, et al. 7 Tesla quantitative hip MRI: T1, T2 and T2 star mapping of hip cartilage in healthy volunteers. *Eur Radiol.* 2016;26:1245–1253.
- Heule R, Ganter C, Bieri O. Triple echo steady-state (TESS) relaxometry. Magn Reson Med. 2014;71:230–237.
- Juras V, Bohndorf K, Heule R, et al. A comparison of multi-echo spin-echo and triple-echo steady-state T2 mapping for in vivo evaluation of articular cartilage. *Eur Radiol.* 2016;26:1905–1912.
- Kraff O, Lazik-Palm A, Heule R, et al. 7 Tesla quantitative hip MRI: a comparison between TESS and CPMG for T2 mapping. *MAGMA*. 2016;29:503–512.
- Riegler G, Drlicek G, Kronnerwetter C, et al. High-resolution axonal bundle (fascicle) assessment and triple-echo steady-state T2 Mapping of the Median Nerve at 7 T: Preliminary Experience. *Invest Radiol.* 2016;51:529–535.
- Szomolanyi P, Goodyear D, Balcom B, et al. SPIRAL-SPRITE: a rapid single point MRI technique for application to porous media. *Magn Reson Imaging*. 2001;19:423–428.
- Robson MD, Benjamin M, Gishen P, et al. Magnetic resonance imaging of the Achilles tendon using ultrashort TE (UTE) pulse sequences. *Clin Radiol.* 2004;59:727-735.
- Gurney PT, Hargreaves BA, Nishimura DG. Design and analysis of a practical 3D cones trajectory. *Magn Reson Med.* 2006;55:575–582.
- Qian YX, Boada FE. Acquisition-weighted stack of spirals for fast highresolution three- dimensional ultra-short echo time MR imaging. *Magn Reson Med.* 2008;60:135–145.
- Juras V, Zbyn S, Pressl C, et al. Regional variations of T-2* in healthy and pathologic Achilles tendon in vivo at 7 Tesla: preliminary results. *Magn Reson Med.* 2012;68:1607–1613.
- Oliveira RR, Medina de Mattos R, Magalhaes Rebelo L, et al. Experimental diabetes alters the morphology and nano-structure of the Achilles tendon. *PLoS One.* 2017;12:e0169513.
- Han M, Larson PE, Liu J, et al. Depiction of achilles tendon microstructure in vivo using high- resolution 3-dimensional ultrashort echo-time magnetic resonance imaging at 7 T. *Invest Radiol.* 2014;49:339–345.
- Hager B, Walzer SM, Deligianni X, et al. Orientation dependence and decay characteristics of T2 * relaxation in the human meniscus studied with 7 Tesla MR microscopy and compared to histology. *Magn Reson Med.* 2019;81:921–933.
- Juras V, Apprich S, Zbyn S, et al. Quantitative MRI analysis of Menisci using biexponential T- 2* fitting with a variable echo time sequence. *Magn Reson Med.* 2014;71:1015–1023.
- Borthakur A, Mellon E, Niyogi S, et al. Sodium and T1rho MRI for molecular and diagnostic imaging of articular cartilage. *NMR Biomed*. 2006;19:781–821.
- Burstein D, Gray M, Mosher T, et al. Measures of molecular composition and structure in osteoarthritis. *Radiol Clin North Am.* 2009;47:675–686.

- Wheaton AJ, Casey FL, Gougoutas AJ, et al. Correlation of T1rho with fixed charge density in cartilage. J Magn Reson Imaging. 2004;20:519-525.
- Keenan KE, Besier TF, Pauly JM, et al. Prediction of glycosaminoglycan content in human cartilage by age, T1rho and T2 MRI. Osteoarthritis Cartilage. 2011;19:171–179.
- Singh A, Haris M, Cai K, et al. High resolution T1rho mapping of in vivo human knee cartilage at 7T. *PLoS One*. 2014;9:e97486.
- Wyatt C, Guha A, Venkatachari A, et al. Improved differentiation between knees with cartilage lesions and controls using 7T relaxation time mapping. J Orthop Trans. 2015;3:197–204.
- Ling W, Regatte RR, Navon G, et al. Assessment of glycosaminoglycan concentration in vivo by chemical exchange-dependent saturation transfer (gagCEST). *Proc Natl Acad Sci U S A*. 2008;105:2266–2270.
- Schmitt B, Zbyn S, Stelzeneder D, et al. Cartilage quality assessment by using glycosaminoglycan chemical exchange saturation transfer and Na-23 MR imaging at 7 T. *Radiology*. 2011;260:257–264.
- Singh A, Haris M, Cai KJ, et al. Chemical exchange saturation transfer magnetic resonance imaging of human knee cartilage at 3 T and 7 T. *Magn Reson Med.* 2012;68:588–594.
- Krusche-Mandl I, Schmitt B, Zak L, et al. Long-term results 8 years after autologous osteochondral transplantation: 7 T gagCEST and sodium magnetic resonance imaging with morphological and clinical correlation. *Osteoarthritis Cartilage*. 2012;20:357–363.
- Koller U, Apprich S, Schmitt B, et al. Evaluating the cartilage adjacent to the site of repair surgery with glycosaminoglycan-specific magnetic resonance imaging. *Int Orthop.* 2017;41:969–974.
- Kogan F, Hargreaves BA, Gold GE. Volumetric multislice GagCEST imaging of articular cartilage: optimization and comparison with T1rho. *Magn Reson Med.* 2017;77:1134–1141.
- Lee JS, Parasoglou P, Xia D, et al. Uniform magnetization transfer in chemical exchange saturation transfer magnetic resonance imaging (vol 3, 1707, 2013). Sci Rep. 2013;3:1–5.
- Windschuh J, Zaiss M, Ehses P, et al. Assessment of frequency drift on CEST MRI and dynamic correction: application to gagCEST at 7 T. *Magn Reson Med.* 2019;81:573–582.
- Schreiner MM, Zbyn S, Schmitt B, et al. Reproducibility and regional variations of an improved gagCEST protocol for the in vivo evaluation of knee cartilage at 7 T. MAGMA. 2016;29:513–521.
- 44. Krishnamoorthy G, Nanga RPR, Bagga P, et al. High quality threedimensional gagCEST imaging of in vivo human knee cartilage at 7 Tesla. *Magn Reson Med.* 2017;77:1866–1873.
- Brinkhof S, Nizak R, Khlebnikov V, et al. Detection of early cartilage damage: feasibility and potential of gagCEST imaging at 7T. *Eur Radiol.* 2018;28:2874–2881.
- Peterson P, Svensson OE. T2 relaxation time bias in gagCEST at 3T and 7T: comparison of saturation schemes. *Magn Reson Med.* 2019;81:1044–1051.
- Shapiro EM, Borthakur A, Dandora R, et al. Sodium visibility and quantitation in intact bovine articular cartilage using high field (23)Na MRI and MRS. *J Magn Reson*. 2000;142:24–31.
- Chang G, Madelin G, Sherman OH, et al. Improved assessment of cartilage repair tissue using fluid-suppressed (2)(3)Na inversion recovery MRI at 7 Tesla: preliminary results. *Eur Radiol.* 2012;22:1341–1349.
- Staroswiecki E, Bangerter NK, Gurney PT, et al. In vivo sodium imaging of human patellar cartilage with a 3D cones sequence at 3 T and 7 T. J Magn Reson Imaging. 2010;32:446–451.
- Favero M, Ramonda R, Goldring MB, et al. Early knee osteoarthritis. *RMD Open*. 2015;1(suppl 1):e000062.
- Maroudas A, Muir H, Wingham J. The correlation of fixed negative charge with glycosaminoglycan content of human articular cartilage. *Biochim Biophys Acta*. 1969;177:492–500.

- Wheaton AJ, Borthakur A, Shapiro EM, et al. Proteoglycan loss in human knee cartilage: quantitation with sodium MR imaging-feasibility study. *Radiology*. 2004;231:900–905.
- Wang LG, Wu Y, Chang G, et al. Rapid isotropic 3D-sodium MRI of the knee joint in vivo at 7T. J Magn Reson Imaging. 2009;30:606–614.
- Madelin G, Babb J, Xia D, et al. Articular cartilage: evaluation with fluidsuppressed 7.0-T sodium MR imaging in subjects with and subjects without osteoarthritis. *Radiology*. 2013;268:481–491.
- Madelin G, Xia D, Brown R, et al. Longitudinal study of sodium MRI of articular cartilage in patients with knee osteoarthritis: initial experience with 16-month follow-up. *Eur Radiol.* 2018;28:133–142.
- Mithoefer K, Williams RJ, Warren RF, et al. The microfracture technique for the treatment of articular cartilage lesions in the knee: a prospective cohort study. *Journal of Bone and Joint Surg Am.* 2005;87A:1911–1920.
- Hangody L, Vasarhelyi G, Hangody LR, et al. Autologous osteochondral grafting-technique and long-term results. *Injury*. 2008;39:S32–S39.
- Jones DG, Peterson L. Autologous chondrocyte implantation. J Bone Joint Surg Am. 2006;88A:2502–2520.
- Behrens P, Bitter T, Kurz B, et al. Matrix-associated autologous chondrocyte transplantation/implantation (MACT/MACI) - 5-year follow-up. *Knee*. 2006;13:194–202.
- Trattnig S, Welsch GH, Juras V, et al. Na-23 MR imaging at 7 T after knee matrix-associated autologous chondrocyte transplantation: preliminary results. *Radiology*. 2010;257:175–184.
- Zbyn S, Stelzeneder D, Welsch GH, et al. Evaluation of native hyaline cartilage and repair tissue after two cartilage repair surgery techniques with Na-23 MR imaging at 7 T: initial experience. *Osteoarthritis Cartilage*. 2012;20:837–845.
- Zbyn S, Brix MO, Juras V, et al. Sodium magnetic resonance imaging of ankle joint in cadaver specimens, volunteers, and patients after different cartilage repair techniques at 7 T initial results. *Invest Radiol.* 2015;50:246– 254.
- Juras V, Zbyn S, Pressl C, et al. Sodium MR imaging of Achilles tendinopathy at 7 T: preliminary results. *Radiology*. 2012;262:199–205.
- Juras V, Winhofer Y, Szomolanyi P, et al. Multiparametric MR imaging depicts glycosaminoglycan change in the Achilles tendon during ciprofloxacin administration in healthy men: initial observation. *Radiology*. 2015;275:763–771.
- Noebauer-Huhmann IM, Juras V, Pfirrmann CW, et al. Sodium MR imaging of the lumbar intervertebral disk at 7 T: correlation with T2 mapping and modified Pfirrmann score at 3 T: preliminary results. *Radiology*. 2012;265:555–564.
- Madelin G, Poidevin F, Makrymallis A, et al. Classification of sodium MRI data of cartilage using machine learning. *Magn Reson Med.* 2015;74:1435– 1448.
- Diaz-Manera J, Llauger J, Gallardo E, et al. Muscle MRI in muscular dystrophies. *Acta Myol.* 2015;34:95–108.
- 68. Valkovič L, Klepochová R, Krššák M. Multinuclear Magnetic Resonance Spectroscopy of Human Skeletal Muscle Metabolism in Training and Disease, Muscle Cell and Tissue – Current Status of Research Field, Kunihiro Sakuma, IntechOpen, (October 10, 2018). DOI: 10.5772/ intechopen.77107. Available from: https://www.intechopen.com/books/ muscle-cell-and-tissue-current-status-of-research-field/multinuclearmagnetic-resonance-spectroscopy-of-human-skeletal-muscle-metabolismin-training-and-dis
- Burakiewicz J, Sinclair CDJ, Fischer D, et al. Quantifying fat replacement of muscle by quantitative MRI in muscular dystrophy. *J Neurol*. 2017;264:2053–2067.
- Kim SJ, Hong SH, Jun WS, et al. MR imaging mapping of skeletal muscle denervation in entrapment and compressive neuropathies. *Radiographics*. 2011;31:319–332.

- Damon BM, Ding Z, Anderson AW, et al. Validation of diffusion tensor MRIbased muscle fiber tracking. *Magn Reson Med.* 2002;48:97–104.
- Paradas C, Llauger J, Diaz-Manera J, et al. Redefining dysferlinopathy phenotypes based on clinical findings and muscle imaging studies. *Neurology*. 2010;75:316–323.
- Hollingsworth KG, Garrood P, Eagle M, et al. Magnetic resonance imaging in Duchenne muscular dystrophy: longitudinal assessment of natural history over 18 months. *Muscle Nerve*. 2013;48:586–588.
- Glover GH. Multipoint Dixon technique for water and fat proton and susceptibility imaging. J Magn Reson Imaging. 1991;1:521–530.
- Grimm A, Meyer H, Nickel MD, et al. Evaluation of 2-point, 3-point, and 6point Dixon magnetic resonance imaging with flexible echo timing for muscle fat quantification. *Eur J Radiol.* 2018;103:57–64.
- Schick F, Eismann B, Jung WI, et al. Comparison of localized proton NMR signals of skeletal muscle and fat tissue in vivo: two lipid compartments in muscle tissue. *Magn Reson Med.* 1993;29:158–167.
- Krssak M, Falk Petersen K, Dresner A, et al. Intramyocellular lipid concentrations are correlated with insulin sensitivity in humans: a 1H NMR spectroscopy study. *Diabetologia*. 1999;42:113–116.
- Just Kukurová I, Valkovič L, Bogner W, et al. Two-dimensional spectroscopic imaging with combined free induction decay and long-TE acquisition (FID echo spectroscopic imaging, FIDESI) for the detection of intramyocellular lipids in calf muscle at 7 T. NMR Biomed. 2014;27:980– 987.
- Regatte RR, Schweitzer ME. Ultra-high-field MRI of the musculoskeletal system at 7.0T. J Magn Reson Imaging. 2007;25:262–269.
- Deroide N, Bousson V, Mambre L, et al. Muscle MRI STIR signal intensity and atrophy are correlated to focal lower limb neuropathy severity. *Eur Radiol.* 2015;25:644–651.
- Mori S, van Zijl PC. Fiber tracking: principles and strategies: a technical review. NMR Biomed. 2002;15:468–480.
- Giraudo C, Motyka S, Weber M, et al. Normalized STEAM-based diffusion tensor imaging provides a robust assessment of muscle tears in football players: preliminary results of a new approach to evaluate muscle injuries. *Eur Radiol.* 2018;28:2882–2889.
- Damon BM. Effects of image noise in muscle diffusion tensor (DT)-MRI assessed using numerical simulations. *Magn Reson Med.* 2008;60:934–944.
- Williams SE, Heemskerk AM, Welch EB, et al. Quantitative effects of inclusion of fat on muscle diffusion tensor MRI measurements. *J Magn Reson Imaging*. 2013;38:1292–1297.
- Choi S, Cunningham DT, Aguila F, et al. DTI at 7 and 3 T: systematic comparison of SNR and its influence on quantitative metrics. *Magn Reson Imaging*, 2011;29:739–751.
- Giraudo C, Motyka S, Weber M, et al. Diffusion tensor imaging of healthy skeletal muscles: a comparison between 7 T and 3 T. *Invest Radiol.* 2019;54:48–54.
- Valkovič L, Chmelík M, Krššák M. In-vivo 31P-MRS of skeletal muscle and liver: a way for non-invasive assessment of their metabolism. *Anal Biochem.* 2017;529:193–215.
- Moon RB, Richards JH. Determination of intracellular pH by 31P magnetic resonance. J Biol Chem. 1973;248:7276–7278.
- Kemp GJ, Ahmad RE, Nicolay K, et al. Quantification of skeletal muscle mitochondrial function by 31P magnetic resonance spectroscopy techniques: a quantitative review. *Acta Physiol (Oxf)*. 2015;213:107–144.

- Griffiths RD, Cady EB, Edwards RH, et al. Muscle energy metabolism in Duchenne dystrophy studied by 31P-NMR: controlled trials show no effect of allopurinol or ribose. *Muscle Nerve*. 1985;8:760–767.
- Zatina MA, Berkowitz HD, Gross GM, et al. 31P nuclear magnetic resonance spectroscopy: noninvasive biochemical analysis of the ischemic extremity. *J Vasc Surg.* 1986;3:411–420.
- Sedivy P, Drobny M, Dezortova M, et al. 31P-MR spectroscopy in patients with mild and serious lower limb ischemia. *Int Angiol.* 2018;37:293–299.
- Melenovsky V, Hlavata K, Sedivy P, et al. Skeletal muscle abnormalities and iron deficiency in chronic heart failure: an exercise (31)P magnetic resonance spectroscopy study of calf muscle. *Circ Heart Fail*. 2018;11:e004800.
- 94. Valkovič L, Chmelík M, Ukropcová B, et al. Skeletal muscle alkaline Pi pool is decreased in overweight-to-obese sedentary subjects and relates to mitochondrial capacity and phosphodiester content. *Sci Rep.* 2016;6:20087.
- Cree-Green M, Newcomer BR, Coe G, et al. Peripheral insulin resistance in obese girls with hyperandrogenism is related to oxidative phosphorylation and elevated serum free fatty acids. *Am J Physiol Endocrinol Metab.* 2015;308:E726–E733.
- 96. Sedivy P, Dezortova M, Drobny M, et al. Differences in muscle metabolism in patients with type I diabetes: influence of gender and nephropathy studied by (31)P MR spectroscopy. *Physiol Res.* 2018;67:433–441.
- Meyerspeer M, Robinson S, Nabuurs CI, et al. Comparing localized and nonlocalized dynamic 31P magnetic resonance spectroscopy in exercising muscle at 7 T. Magn Reson Med. 2012;68:1713–1723.
- Valkovič L, Chmelík M, Just Kukurová I, et al. Depth-resolved surface coil MRS (DRESS)- localized dynamic 31P-MRS of the exercising human gastrocnemius muscle at 7 T. NMR Biomed. 2014;27:1346–1352.
- Parasoglou P, Xia D, Chang G, et al. Dynamic three-dimensional imaging of phosphocreatine recovery kinetics in the human lower leg muscles at 3T and 7T: a preliminary study. *NMR Biomed*. 2013;26:348–356.
- 100. Schmid AI, Meyerspeer M, Robinson SD, et al. Dynamic PCr and pH imaging of human calf muscles during exercise and recovery using (31) P gradient-Echo MRI at 7 Tesla. *Magn Reson Med.* 2016;75:2324–2331.
- Valkovič L, Chmelík M, Meyerspeer M, et al. Dynamic 31P-MRSI using spiral spectroscopic imaging can map mitochondrial capacity in muscles of the human calf during plantar flexion exercise at 7T. *NMR Biomed*. 2016;29:1825–1834.
- Niess F, Fiedler GB, Schmid AI, et al. Interleaved multivoxel (31) P MR spectroscopy. *Magn Reson Med.* 2017;77:921–927.
- 103. Khegai O, Madelin G, Brown R, et al. Dynamic phosphocreatine imaging with unlocalized pH assessment of the human lower leg muscle following exercise at 3T. *Magn Reson Med.* 2018;79:974–980.
- 104. Pollak AW, Meyer CH, Epstein FH, et al. Arterial spin labeling MR imaging reproducibly measures peak-exercise calf muscle perfusion: a study in patients with peripheral arterial disease and healthy volunteers. *JACC Cardiovasc Imaging*. 2012;5:1224–1230.
- 105. Schewzow K, Fiedler GB, Meyerspeer M, et al. Dynamic ASL and T2weighted MRI in exercising calf muscle at 7 T: a feasibility study. *Magn Reson Med.* 2015;73:1190–1195.
- 106. Schmid AI, Schewzow K, Fiedler GB, et al. Exercising calf muscle T2 * changes correlate with pH, PCr recovery and maximum oxidative phosphorylation. *NMR Biomed*. 2014;27:553–560.