G3, 2021, 11(11), jkab269
DOI: 10.1093/g3journal/jkab269
Advance Access Publication Date: 30 July 2021
Investigation

Genome-wide association mapping of *Pyrenophora teres* f. *maculata* and *Pyrenophora teres* f. *teres* resistance loci utilizing natural Turkish wild and landrace barley populations

Shaun J. Clare [1,†] Arzu Çelik Oğuz,[2,†] Karl Effertz,[1] Roshan Sharma Poudel,[3] Deven See,[4,5] Aziz Karakaya,[2] and Robert S. Brueggeman [1,*]

[1]Department of Crop and Soil Sciences, Washington State University, Pullman, WA 99163, USA
[2]Department of Plant Protection, Faculty of Agriculture, Ankara University, Dışkapı, Ankara 06110, Turkey
[3]Sygenta Seed Inc., Durham, NC 27709, USA
[4]Wheat Health, Genetics and Quality Research Unit, Agricultural Research Service, U.S. Department of Agriculture, Pullman, WA 99163, USA, and
[5]Department of Plant Pathology, Washington State University, Pullman, WA 99163, USA

*Corresponding author: Department of Drop and Soil Science, Washington State University, Johnson Hall Room 215, Pullman, WA 99164-6420, USA. Email: bob.brueggeman@wsu.edu
†These authors have contributed equally to this work and share first authorship.

Abstract

Unimproved landraces and wild relatives of crops are sources of genetic diversity that were lost post domestication in modern breeding programs. To tap into this rich resource, genome-wide association studies in large plant genomes have enabled the rapid genetic characterization of desired traits from natural landrace and wild populations. Wild barley (*Hordeum spontaneum*), the progenitor of domesticated barley (*Hordeum vulgare*), is dispersed across Asia and North Africa, and has co-evolved with the ascomycetous fungal pathogens *Pyrenophora teres* f. *teres* and *P. teres* f. *maculata*, the causal agents of the diseases net form of net blotch and spot form of net blotch, respectively. Thus, these wild and local adapted barley landraces from the region of origin of both the host and pathogen represent a diverse gene pool to identify new sources of resistance, due to millions of years of co-evolution. The barley—*P. teres* pathosystem is governed by complex genetic interactions with dominant, recessive, and incomplete resistances and susceptibilities, with many isolate-specific interactions. Here, we provide the first genome-wide association study of wild and landrace barley from the Fertile Crescent for resistance to both forms of *P. teres*. A total of 14 loci, four against *P. teres* f. *maculata* and 10 against *P. teres* f. *teres*, were identified in both wild and landrace populations, showing that both are genetic reservoirs for novel sources of resistance. We also highlight the importance of using multiple algorithms to both identify and validate additional loci.

Keywords: Barley landrace; wild barley; *Hordeum spontaneum*; *Pyrenophora teres* f. *teres*; *Pyrenophora teres* f. *maculata*; resistance; susceptibility; GWAS

Introduction

Net blotch caused by the ascomycetous fungal pathogen *Pyrenophora teres* (anamorph: *Drechslera teres*) is an economically important disease of barley worldwide. *P. teres* f. *teres* (*Ptt*) incites the net form of net blotch (NFNB) and *P. teres* f. *maculata* (*Ptm*) incites the spot form of net blotch (SFNB) (Smedegård-Petersen 1971). Both forms are responsible for large crop losses that typically range between 10% and 40% when susceptible cultivars are grown, however, under conducive environmental conditions losses can reach 100% (Piening and Kaufmann 1969; Mathre 1982; Moya *et al.* 2018). At least one form of the disease has been reported from all barley growing regions and, in many regions, both forms are present with annual fluctuation in predominance. This presents challenges to breeders, as both *Ptt* and *Ptm* interact with host resistance/susceptibility genes differentially, thus are considered distinct and treated as different diseases when breeding for resistance. However, as further characterization of resistant/susceptibility loci continues, overlaps in host-pathogen genetic interactions in both pathosystems are becoming more prevalent.

Both *Ptt* and *Ptm* occur as genetically distinct populations and can be separated in the field based on lesion morphology. Although these two forms can be hybridized under laboratory conditions (Campbell and Crous 2003), hybridization under field conditions is extremely rare (Campbell *et al.* 2002; McLean *et al.* 2014; Akhavan *et al.* 2015; Çelik Oğuz *et al.* 2018; Poudel *et al.* 2019). However, both forms of *P. teres* undergo form specific sexual as well as asexual reproduction (Karakaya *et al.* 2004; Serenius et al. 2005; Akhavan *et al.* 2015; Çelik Oğuz *et al.* 2018, 2019b; Poudel *et al.* 2019). The complex nature of this reproduction system poses serious evolutionary risks for resistance

Received: June 04, 2021. **Accepted:** July 26, 2021

breeding as populations contain diverse effector repertoires, that in different combinations, can rapidly overcome deployed resistances (McDonald and Linde 2002). Yet, the use of resistant barley cultivars is the most environmentally friendly and economically feasible method of NFNB and SFNB control (Robinson and Jalli 1996; Afanasenko *et al.* 2009).

Wild barleys and barley landraces are important sources of resistance against diverse biotic and abiotic stresses (Allard and Bradshaw 1964; Ceccarelli 1996; Yitbarek *et al.* 1998; Ellis *et al.* 2000; Jakob *et al.* 2014; Karakaya *et al.* 2016, 2020). Wild barley (*Hordeum spontaneum*) is known as the progenitor of modern-day barley (*Hordeum vulgare*) and grows naturally in the Fertile Crescent, regions of south and southeastern Turkey, North Africa, and Southwest Asia (Harlan and Zohary 1966; Nevo 1992; Zohary and Hopf 2000). Barley landraces gave rise to modern barley varieties (Thomas *et al.* 1998) as they were subjected to natural and artificial selection for the last 10,000 years (Ceccarelli and Grando 2000). Within the barley center of origin, Turkey is located at the ancestral hub of barley diversification regions including the Mediterranean, Horn of Africa, and the Tibetan Plateau (Muñoz-Amatriaín *et al.* 2014; Poets *et al.* 2015), with diverse barley landraces still used by Turkish farmers (Helbaek 1969; Kün 1996; Pourkheirandish and Komatsuda 2007; Ergün *et al.* 2017). Turkey is also at the center of origin of the *P. teres* pathogens, *Ptt* and *Ptm*. Barley lines can show differential responses to either or both forms of net blotch due to distinct yet complex genetic interactions with each form. Many barley genotypes may be resistant to the majority of isolates of one form yet susceptible to the alternate form of most isolates (Bockelman *et al.* 1983; Grewal *et al.* 2012). Thus, when breeding for resistance, the two forms of net blotch are treated as separate diseases (Liu *et al.* 2011; Usta *et al.* 2014; Yazıcı *et al.* 2015).

In barley, genetic resistance to *P. teres* was first reported in Geschele (1928). Since both forms of net blotch were not described at that time, it was assumed that the net form was used. In the first studies to genetically characterize barley—*P. teres* interactions, Tifang was the resistant parent in the Tifang × Atlas cross (Schaller 1955). Mode and Schaller (1958) identified the resistance genes Pt_1, Pt_2, and Pt_3 segregating in this cross. Bockelman *et al.* (1977) revised the naming of Pt_1, Pt_2, and Pt_3 resistance genes and described the *Rpt1* (Pt_1 and Pt_2), *Rpt2* (novel), and *Rpt3* (Pt_3) loci on chromosomes 3H, 1H, and 2H, respectively. Further studies identified *Rpt4* on chromosome 7H (Williams *et al.* 1999, 2003), *Rpt5* on chromosome 6H, *Rpt6* on chromosome 5H (Manninen *et al.* 2006), *Rpt7* on chromosome 4HL, and *Rpt8* on chromosome 4HS (Franckowiak and Platz 2013), however, there are over 340 QTLs previously identified (Clare *et al.* 2020). Manninen *et al.* (2006) reclassified the locus Pt_a as *Rpt5* on chromosome 6H, and subsequently, three genes/alleles have been characterized at the locus (*Rpt5.f, Spt1.k, Spt1.r*) as dominant resistance or susceptibility genes (Franckowiak and Platz 2013; Richards *et al.* 2016). In multiple barley—*Ptt* genetic interaction studies, it has been shown that the *Rpt5* locus is the most important resistance/susceptibility locus in this system. This complex locus putatively contains multiple resistance as well as susceptibility genes that have been characterized in diverse barley—*P. teres* interactions from around the world (Clare *et al.* 2020). Because the *Rpt5* locus also shows dominant susceptibility in certain barley lines, additional alleles were designated *Susceptibility to P. teres 1* (*Spt1*) by Richards *et al.*, (2016). Furthermore, high-resolution genetic mapping and positional cloning efforts have identified *Rpt5* and *Spt1* candidate genes and functional validation are underway (Brueggeman *et al.* 2020).

Resistance to *Ptm* originally appeared to be less complicated when compared to *Ptt* due to the presence of three major loci. These three loci were identified as *Rpt4* on chromosome 7H (Williams *et al.* 1999, 2003), *Rpt6* on chromosome 5HS (Manninen *et al.* 2006), and *Rpt8* on chromosome 4HS (Friesen *et al.* 2006; Franckowiak and Platz 2013). To date, over 140 QTLs have been reported to be implicated in the *Ptm* reaction, which have been collapsed into 36 unique loci, five of which are specific to *Ptm* and the rest showing some degree of overlap with known *Ptt* loci (Clare *et al.* 2020). These five unique loci that are specific to the *Ptm* interaction are *SFNB-3H-78.53* on chromosome 2H (Burlakoti *et al.* 2017), *QRptm-4H-120-125* on 4H (Tamang *et al.* 2019), *QRptts-5H-106.00* on 5H (Adhikari *et al.* 2019), *QRptm7-3* on 7H (Wang *et al.* 2015), and *QRptm7-6/QRptm-7H-119-137* on 7H (Wang *et al.* 2015; Tamang *et al.* 2019). Considering that all currently designated resistance/susceptibility loci except for *Rpt2* (only implicated in the *Ptt* interaction) have now been implicated in both *Ptm* and *Ptt* interactions (Clare *et al.* 2020), it is with some caution that it can be concluded that host-pathogen genetic interactions with the two forms and barley should be considered distinct. Thus, for both forms, with the exception of *Rpt6*, numerous researchers have described synonyms of all loci (Clare *et al.* 2020).

Multiple genome-wide association mapping studies (GWAS) have investigated NFNB resistance in barley (Richards *et al.* 2017; Wonneberger *et al.* 2017b; Amezrou *et al.* 2018; Adhikari *et al.* 2019; Daba *et al.* 2019; Novakazi *et al.* 2019; Rozanova *et al.* 2019; Adhikari *et al.* 2020). A large proportion of the resistance markers associated with NFNB resistance have been localized to the centromeric region of barley chromosome 6H (Richards *et al.* 2017). In GWAS for SFNB resistance, 29 (Wang *et al.* 2015), 27 (Tamang *et al.* 2015), 11 (Burlakoti *et al.* 2017), and 1 (Daba *et al.* 2019) unique genomic loci were identified. Four important QTLs (*QRptm7-4*, *QRptm7-6*, *QRptm7-7*, and *QRptm7-8*) were mapped into a region covering the *Rpt4* locus on chromosome 7HS (Wang *et al.* 2015). Burlakoti *et al.* (2017) identified a new and important QTL on chromosome 2HS that was predominately found in 6-rowed barley lines as compared to 2-rowed. Daba *et al.* (2019) also defined a new QTL on chromosome 6H associated with *Ptm* susceptibility in a large number of genotypes, which is a common mechanism in inverse gene-for-gene interactions with this pathogen. Vatter *et al.* (2017) performed nested association mapping for NFNB and described further interactions at the important *Rpt5*/*Spt1* locus. In barley, the newest approach to identify marker trait associations (MTAs) with *Ptt* resistance is exome QTL-seq. This approach identified a large number of MTAs on chromosomes 3H and 6H when analyzing resistant and susceptible bulks (Hisano *et al.* 2017). The resistance status of wild barley genotypes and barley landraces to *P. teres* has been reported by several research groups worldwide (Jana and Bailey, 1995; Lakew *et al.*, 1995; Legge *et al.*, 1996; Sato and Takeda, 1997; Fetch *et al.*, 2003; Silvar *et al.*, 2010; Endresen *et al.*, 2011; Neupane *et al.*, 2015; Çelik Oğuz *et al.*, 2017, 2019a, 2019c). However, molecular mapping studies of *P. teres* resistance in wild and landrace barleys have been limited (Yun *et al.* 2005; Vatter *et al.* 2017; Adhikari *et al.* 2019; Gyawali *et al.* 2019, 2020). In this study, four novel loci representing resistance to NFNB were mapped in Turkish wild barley and landraces. This study highlights the importance of surveying wild and unimproved barley lines for sources of resistance that may have been lost during domestication and modern breeding. These studies, focused on diversity in the barley primary germplasm pool, will provide new sources of resistance and associated markers to aid in deploying robust resistances against NFNB and SFNB.

Materials and methods

Biological materials

A total of 295 barley accessions comprised of 193 landraces (*H. vulgare*) and 102 wild barley (*H. spontaneum*) genotypes which were collected from different growing regions of Turkey and maintained at the Field Crops Central Research Institute and Department of Plant Protection, Faculty of Agriculture, Ankara University located in Ankara, Turkey were utilized in the analyses (Çelik Oğuz *et al.* 2017, 2019a). Three virulent *Ptm* isolates (GPS263, 13–179, and 13–167) and three *Ptt* isolates (UHK77, GPS18, and 13–130) collected from different provinces of Turkey (Çelik Oğuz and Karakaya 2017) were used for the phenotypic assessment of the 295 barley accessions.

Pathogen assay phenotyping of barley lines

Phenotyping of the wild barley (*H. spontaneum*) genotypes and barley landraces (*H. vulgare*) were accomplished according to methods outlined in Çelik Oğuz *et al.* (2017) and Çelik Oğuz *et al.* (2019a). Briefly, a total of 5–10 seeds were planted in 7 cm diameter plastic pots containing sterile soil, sand, and organic matter mixtures (60, 20, 20; v/v/v, respectively), depending on the number of seeds of each wild and landrace barley. The pots were kept in greenhouse conditions at 18–23 ± 1°C night/day with 14 h/10 h light/dark regime before and after inoculation. Three virulent isolates of *Ptm* (GPS263, 13–179, and 13–167) and three virulent isolates of *Ptt* (GPS18, UHK77, and 13–130) were used in phenotyping studies. The inoculum was prepared from 10-day single spore cultures grown on potato dextrose agar kept at 16–23 ± 2°C night/day with a 10 h/14 h dark/light period. For the preparation of the inoculum, the mycelia were scraped from Petri dishes using a painting brush, washing with water, and filtered by cheesecloth. The inoculum concentration was adjusted to $15–20 \times 10^4$ mycelial fragments/ml. One drop of Tween® 20 was added to each 100 ml inoculum. Inoculation was carried out at the two to three-leaf stages by spraying inoculum over the barley lines with a hand spray until runoff. Following inoculation, plants were covered with nylon in transparent boxes with lids for 76 h. High humidity was maintained for a further 48 h with the nylon uncovered and ventilated. After 7 days, seedlings were evaluated for disease severity using the net and SFNB scales described by Tekauz (1985).

PCR-GBS library preparation and genotyping by sequencing

Two independent custom PCR-GBS SNP marker panels containing 365 (Panel 1) (Sharma Poudel *et al.* 2018; Tamang *et al.* 2019) and 1,272 (Panel 2) (Ruff *et al.* 2020) barley SNP markers were used to genotype 295 barley accessions. Marker primers were divided into six and three total primer pools for Panels 1 and 2 for PCR amplification, respectively. PCR amplification and barcoding reactions were performed as described by Sharma Poudel *et al.* (2018) and Ruff *et al.* (2020). Briefly, nine primary PCR amplification reactions were performed per sample. Following amplification, equal volumes of primary PCR products were pooled into 96-well plates with each well containing all amplified markers for a given DNA sample. Next, barcoding PCR reactions were performed with a universal barcoding reverse primer and unique forward barcoding primers for each sample. Following barcoding reactions, samples were pooled and purified. A final PCR reaction was performed using sequencing primers to ensure the barcoding reaction was successful. Samples taken before and after the final amplification were run on agarose gel to verify the appropriate product size and amplification. Quantification of the barcoded libraries were performed using the Qubit dsDNA HS assay kit (Life Technologies, Carlsbad, CA, USA). Enrichments were carried out using the Ion OneTouch™ 2 System (Panel 1) and the Ion PI™ Hi-Q Sequencing 200 kit on the Ion Chef (Panel 2). Finally, samples were sequenced on the Ion Torrent Personal Genome Machine™ (Panel 1) and the Ion Proton™ (Panel 2) Systems using two Ion 318™ chips and 3 Ion PI™ chips, respectively, following the manufacturer's standard protocols. The resulting marker panels were collapsed to eliminate duplicated markers from each panel. The Morex reference map (Beier *et al.* 2017; Mascher *et al.* 2017; Monat *et al.* 2019) and iSelect consensus map (Muñoz-Amatriaín *et al.* 2014) were downloaded from the Triticeae Toolbox (T3) barley database (https://triticeaetoolbox.org/barley/). The Morex reference map was used to determine the absolute marker position, whereas markers not included in the Morex reference map were estimated based on the genetic position of the iSelect consensus map relative to the flanking markers.

Imputation, filtering, and linkage disequilibrium

Due to the heterozygosity present in the natural population, heterozygous calls (5.23%) were included in the analysis. Accessions and markers with more than 30% missing data were removed from analysis, resulting in 282 barley accessions and 530 markers. Missing data were imputed using LinkImpute, which uses a linkage disequilibrium *k*-nearest neighbor imputation (LD-kNNi) method (Money *et al.* 2015) in Trait Analysis by aSSociation, Evolution, and Linkage (TASSEL) 5.2.60 (Bradbury *et al.* 2007). Markers with a minor allele frequency of <0.05 were included in the analysis but were treated with caution based on best practice from the Genomic Association and Prediction Integrated Tool (GAPIT) 3.0 user manual. Linkage disequilibrium was calculated in TASSEL using a window size of 50 markers and an R^2 threshold of 0.8 resulting in 522 markers.

Population structure, kinship matrices, and model algorithms

Population structure was accounted for using STRUCTURE analysis and principle component analysis. A total of 522 markers were used for analysis of population structure. The software STRUCTURE v2.3.4 (Pritchard *et al.* 2000) was used to estimate population structure of the barley panel to create a population structure matrix (Q) to be used as a covariate. To determine the optimal number of subpopulations, an admixture ancestry model was used with a burnin of 10,000, followed by 25,000 Monte Carlo Markov Chain (MCMC) replications for $k = 1$ to $k = 10$ with ten iterations. STRUCTURE HARVESTER (Earl and vonHoldt 2012) was used to identify the optimal number of subpopulations using the Δk method (Evanno *et al.* 2005). The optimal k value was subsequently used to run a new STRUCTURE analysis using a burn-in of 100,000 followed by 100,000 MCMC replications. An individual was deemed to be part of a population if the membership probability was >0.8 (Richards *et al.* 2017). Individuals that did not achieve a value of 0.8 were deemed to have admixture ancestry. The final Q matrix was used as a fixed covariate in association models. Principle component analysis was conducted in *R* 3.6.3 using GAPIT 3.0 (Wang and Zhang 2020) with default settings. Principle components explaining at least 25% (*PC1*) and 50% (*PC5*) were used for further analysis. A naïve model using only genotypic (Supplementary File S1 and S2) and phenotypic data (Supplementary File S3) and an additional three fixed-effect models accounting for population structure [Q (Supplementary File S4), *PC1*, and *PC5*] were all performed using the GLM method.

For initial discovery of the most appropriate method to account for random effect in the model, a kinship matrix (*K*) was constructed using the EMMA (Kang *et al.* 2008), Loiselle (Loiselle *et al.* 1995), and VanRaden (VanRaden 2008) algorithms in GAPIT 3.0 (Wang and Zhang 2020) with the MLM model (Yu *et al.* 2006). Based on these results, the EMMA derived *K* matrix (Supplementary File S5) was identified as the most powerful and used for subsequent analysis of mixed models for all isolates that included CMLM (Zhang *et al.* 2010), ECMLM (Li *et al.* 2014), and MLMM (Segura *et al.* 2012). Lastly, SUPER (Wang *et al.* 2014), FarmCPU (Liu *et al.* 2016), and BLINK (Huang *et al.* 2019) algorithms that reconstruct the kinship matrix were used for a total of nine random effect models. Due to the similarity of results, the MLM and MLMM methods were not used in further analysis. Mixed models included combinations to account for population structure (*Q*, *PC1*, and *PC5*), kinship (EMMA *K*), and algorithm methods (CMLM, ECMLM, SUPER, FarmCPU, and BLINK) for a total of 15 mixed models per isolate. The mean-squared deviation (MSD) was calculated for each model (Supplementary File S6, Mamidi *et al.* 2011), however visual inspection of QQ plots were performed to ensure the model was a good fit. This method was employed as models with the lowest MSD model often had highly correlated observed and expected -$\log_{10}(p)$ values yielding zero significant markers. A Bonferroni correction was calculated at an α level of 0.01 and 0.05 for a -log10(p) threshold of 4.72 and 4.02, respectively. Final Manhattan and QQ plots were generated with *R* 3.6.3 package *CMplot* 3.5.1 (https://github.com/YinLiLin/R-CMplot).

QTL identification

Absolute marker positions were extracted for significant MTAs from the Morex reference genome (Beier *et al.* 2017; Mascher *et al.* 2017; Monat *et al.* 2019) and compared to collapsed *P. teres* loci (Clare *et al.* 2020). Significant markers were declared distinct from previously identified loci, *i.e.*, novel, if the nearest neighboring marker that was closer to previously reported loci was not significant or if the gap to currently delimited locus exceeded 10 Mbp in physical distance when no closer marker was present.

Results

Phenotypic analysis

Wild barley was found to be statistically (Wilcoxon rank sum test) more resistant to *Ptm* isolate 13–179 and *Ptt* isolates GPS18 and UHK77, whereas landrace barley was shown to be statistically more resistant to *Ptm* isolate GPS263 and *Ptt* isolate 13–130 (Figure 1). There was no significant difference between landrace and wild barley to *Ptm* isolate 13–167. Despite UHK77 being statistically different between landrace and wild barley, no significant MTAs were found.

Marker panel analysis

Using a 30% missing data threshold, a total of 282 of the 295 barley accessions phenotyped were adequately genotyped. Using a similarity of individual matrix, all remaining individuals were unique. Similarly, 530 of the 598 collapsed markers were deemed to have sufficient coverage across barley accessions using a 30% missing data threshold and were used in the subsequent analysis of the six *P. teres* isolates. To eliminate markers in linkage disequilibrium, an LD R^2 threshold of 0.8 and sliding window of 50 markers was used, resulting in 522 markers for use in the final analysis.

Population structure and linkage disequilibrium

STRUCTURE analysis identified an optimal k value = 2, with 89 and 118 individuals in subpopulations one and two (Supplementary Figure S1). Subpopulation one consisted of wild accessions, whereas subpopulation two consisted of landraces. A total of 75 barley accessions comprised of both landrace and wild barley had population membership probabilities of less than 0.8 and were deemed to have an admixture ancestry. The first five principle components accounted for 29.36, 9.4, 5.3, 4.0, and 3.7%, respectively, in the principle component analysis. Principle components were selected that accounted for at least 25% (*PC1*) and 50% (*PC5*) when eigenvalues were plotted on a cumulative scale.

Association mapping analysis

A total of 24 models were tested on each of the six *P. teres* isolates consisting of three *Ptm* and three *Ptt* isolates. The *Ptm* isolate 13-167 and *Ptt* isolate UHK77 contained no significant markers across all models tested. The *Ptm* isolates GPS263 and 13-179 contained one and three significant markers, respectively. The *Ptt* isolates GPS18 and 13-130 both contained five significant markers.

Ptm *isolate GPS263*

Only one significant MTA was identified with *Ptm* isolate GPS263 on chromosomes 5H based on the second version of the cv. Morex reference genome (Monat *et al.* 2019). The SNP marker 12_20350 located on chromosome 5H at physical position 446449782 was identified at the -log10(p) threshold = 4.72 in the K_{BLINK}. Marker 12_20350 is embedded within the collapsed *NBP_QRptt5-1* locus (Wonneberger *et al.* 2017a; Clare *et al.* 2020) and was not previously shown to be associated with *Ptm* interactions.

Ptm *isolate 13-179*

The three significant MTAs identified with *Ptm* isolate *13–179* were located on chromosomes 3H, 4H, and 5H based on the second version of the Morex reference genome (Monat *et al.* 2019). The marker 11_20866 located on chromosome 3H (physical position 153156749), embedded within the collapsed *QRptms3-2* locus (Wang *et al.* 2015; Burlakoti *et al.* 2017; Koladia *et al.* 2017; Vatter *et al.* 2017; Wonneberger *et al.* 2017a; Daba *et al.* 2019; Novakazi *et al.* 2019; Rozanova *et al.* 2019; Clare *et al.* 2020), was identified at the -log10(p) threshold = 4.02 in the $K_{FarmCPU}$ and PC1+$K_{FarmCPU}$ models. The marker 11_10510 is located on chromosome 4H at position 603258307 and was identified at the -log10(p) threshold = 4.72 in the K_{BLINK} and K_{SUPER} models and at the -log10(p) threshold = 4.02 in the $PC5_{GLM}$, Q+K_{SUPER}, and PC1+K_{SUPER} models. In addition, the 11_10510 marker almost met the significance threshold using the $K_{FarmCPU}$ model. The 11_10510 marker is embedded within the *Rpt8* locus (Friesen *et al.* 2006; Tamang *et al.* 2015; Richards *et al.* 2017; Vatter *et al.* 2017; Daba *et al.* 2019; Clare *et al.* 2020). Lastly, the marker SCRI_RS_160332 is located on chromosome 5H at position 474799503, ~3.0 Mbp distal to the *Qrptts-5HL.1* locus (Richards *et al.* 2017). SCRI_RS_160332 was identified with the $K_{FarmCPU}$ model at the -log10(p) threshold = 4.02.

Ptt *isolate GPS18*

The five significant MTAs identified using *Ptt* isolate GPS18 were distributed across chromosomes 1H, 6H, and 7H. The first marker 11_10176 located on chromosome 1H (position 397791042) was identified in the $K_{FarmCPU}$ model at the -log10(p) threshold = 4.02. The 11_10176 marker is located 13.6 Mbp distal to the collapsed

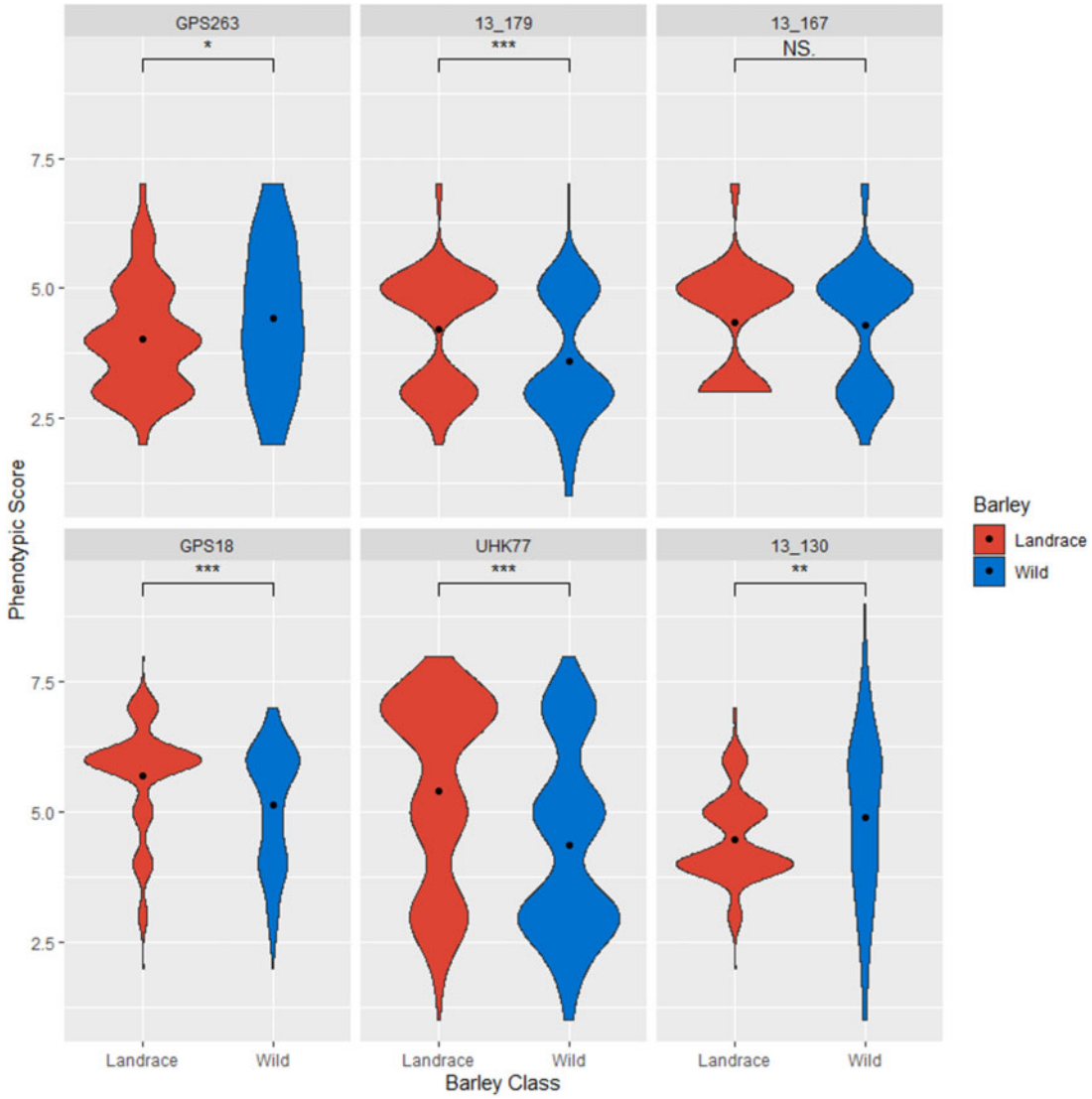

Figure 1 Violin plots of phenotypic distribution of landrace (red) and wild (blue) barley to each *Pyrenophora teres* f. *maculata* (top row) and *P. teres* f. *teres* (bottom row) isolate. Width of the violin indicates the number of accessions with that phenotypic score and the black dot represents the barley class mean. Wilcoxon test significance is indicated by asterisks above each plot.

NBP_QRptt1-1 (Wonneberger *et al.* 2017a) and 18.2 Mbp proximal to the QTL identified at 57.3-62.8 cM by Rozanova *et al.* (2019). The marker 11_20754 located on chromosome 1H (position 483805599) was identified in the K_{BLINK} and $K_{FarmCPU}$ models at the -log10(*p*) threshold = 4.72 and 4.02, respectively. The 11_20754 marker is embedded within the *QPt.1H-1* (Vatter *et al.* 2017; Clare *et al.* 2020) locus along with *QRptts-1H-92-93* (Amezrou *et al.* 2018) and a QTL from Tamang *et al.* (2015). The third marker 12_31282 located on chromosome 7H (position 617741299), is embedded within the collapsed QTL_{UHs}*-7H* locus (König *et al.* 2013; Tamang *et al.* 2015; Richards *et al.* 2017; Wonneberger *et al.* 2017a; Martin *et al.* 2018; Novakazi *et al.* 2019; Tamang *et al.* 2019; Clare *et al.* 2020). The 12_31282 MTA was identified using the $K_{FarmCPU}$ model at the -log10(*p*) threshold = 4.72. The fourth marker, 11_20972 on chromosome 6H (position 539551443) is embedded within the collapsed *AL_QRptt6-2* locus (Afanasenko *et al.* 2015; Vatter *et al.* 2017; Wonneberger *et al.* 2017b; Amezrou *et al.* 2018; Clare *et al.* 2020). Marker 11_20972 was identified using the K_{BLINK} model at the -log10(*p*) threshold = 4.02 and also nearly met the significant threshold using the $K_{FarmCPU}$ model. The last significant marker, 12_30545 located on chromosome 7H (54934072) is embedded within the collapsed *QNFNBAPR.Al/S-7Ha* locus (König *et al.* 2013; Tamang *et al.* 2015; Vatter *et al.* 2017; Wonneberger *et al.* 2017b; Amezrou *et al.* 2018; Daba *et al.* 2019; Novakazi *et al.* 2019; Clare *et al.* 2020). Marker 12_20545 was identified at the -log10(*p*) threshold = 4.02 using the $K_{FarmCPU}$ model. Two additional markers, 12_30250 and 12_111942 located on chromosome 3H and 4H, respectively, were almost significant at the -log10(*p*) threshold = 4.02 in the $K_{FarmCPU}$ model. The 12_30250 marker is embedded within the collapsed *QRpts3La* locus (Raman *et al.* 2003; Lehmensiek *et al.* 2007; Liu *et al.* 2015; Tamang *et al.* 2015; Burlakoti *et al.* 2017; Richards *et al.* 2017; Vatter *et al.* 2017; Daba *et al.* 2019; Tamang *et al.* 2019). The 12_11104 marker is located 1.8 Mbp distal from the *Rpt8* locus (Tamang *et al.* 2015; Vatter *et al.* 2017; Clare *et al.* 2020) and 4.4 Mbp proximal to the *QRptm-4H-120-125* locus (Tamang *et al.* 2015, 2019).

Ptt *isolate 13-130*

The five significant MTAs identified using *Ptt* isolate 13–130 were distributed across chromosomes 2H, 3H, 6H, and 7H. The first marker, 12_11452 located on chromosome 2H (position 34275254) was identified with the K_{BLINK} and $K_{FarmCPU}$ models at the -log10(*p*) threshold = 4.02. The marker 12_11452 has a minor allele frequency less than 5%, however the marker is embedded within the collapsed *SFNB-2H-8-10* locus (Tamang *et al.* 2015; Wonneberger *et al.* 2017a; Amezrou *et al.* 2018; Adhikari *et al.* 2019; Tamang *et al.* 2019; Clare *et al.* 2020). The marker 11_20968, located on chromosome 3H (position 19966889), was identified in the $K_{FarmCPU}$ model at the -log10(*p*) threshold = 4.72. The marker 11_20968 is located 7.6 Mbp distal to the boundary of the collapsed *QPt.3H-1* locus (Vatter *et al.* 2017; Daba *et al.* 2019; Rozanova *et al.* 2019) and 24 Mbp proximal to the boundary of the collapsed *Rpt-3H-4* locus (Tamang *et al.* 2015; Richards *et al.* 2017; Daba *et al.* 2019; Novakazi *et al.* 2019; Clare *et al.* 2020). The marker 12_10662, also located on chromosome 3H (position 553445025), was identified using the $K_{FarmCPU}$ and the $Q+K_{FarmCPU}$ models at the -log10(*p*) threshold = 4.72 and 4.02, respectively. The marker is located 4.6 Mbp distal to boundary of the collapsed *QRpts3La* locus (Liu *et al.* 2015; Tamang *et al.* 2015; Burlakoti *et al.* 2017; Richards *et al.* 2017; Vatter *et al.* 2017; Wonneberger *et al.* 2017a; Daba *et al.* 2019; Tamang *et al.* 2019) and 5.0 Mbp from the boundary of collapsed *Rpt1* locus (Tamang *et al.* 2015; Burlakoti *et al.* 2017; Martin *et al.* 2018; Adhikari *et al.* 2019; Novakazi *et al.* 2019; Clare *et al.* 2020). The last two markers, 11_20714 and 11_11243, were both identified using the $K_{FarmCPU}$ model at the -log10(*p*) threshold = 4.02 and are located on chromosomes 6H (position 489619101) and 7H (position 601974526), respectively. The marker 11_20714 is located 4.1 Mbp proximal to the *QPt.6H-3* locus (Vatter *et al.* 2017) and 11_11243 is embedded within the collapsed *QRptm7-6* locus (Tamang *et al.* 2015; Wang *et al.* 2015; Wonneberger *et al.* 2017a; Tamang *et al.* 2019; Clare *et al.* 2020).

Enrichment analysis

Enrichment of either resistance or susceptibility alleles at each MTA were calculated for landraces and wild barley (Figure 2, Supplementary Figure S2). For *Ptm* isolate GPS263 and *Ptt* isolate 13–130, the landraces that were more resistant than the wild barley showed enrichment for the majority of the resistance alleles or a depletion of susceptibility alleles. For *Ptm* isolate 13–179 and *Ptt* isolate GPS18, the opposite was observed as the wild barley showed more resistance and enrichment for the majority of the resistance alleles or depletion of susceptibility alleles.

Discussion

Both NFNB and SFNB are worldwide threats to barley production and recent evidence shows that both *Ptm* and *Ptt* have evolved to infect and threaten wheat production as well (Tóth *et al.* 2008; Mikhailova *et al.* 2010; Perelló *et al.* 2019). Wild barley and landraces from the origin of cereal domestication represent a rich reservoir of net blotch resistance that could be integrated into elite varieties to help mitigate the threat. Analysis of the phenotypic responses of Turkish wild barley and landraces to regional *Ptm* and *Ptt* isolates showed evidence of landraces under selective pressures by the pathogen during domestication compared with wild barley as seen by the more compact distribution of the phenotypic scores (Tables 1 and 2, Figure 1). This demonstrates that wild barley harbors additional diversity for net blotch resistance that is not present in the landraces and could be exploited for barley variety development. This is corroborated by enrichment analysis that shows enrichment of the resistant haplotype for marker 11_20866 and 11_20754 in the wild barley lines (Figure 2, Supplementary Figure S2). However, the opposite is true for other loci such as the 11_20968 haplotype near the *QRpt-3H.1* locus, that is located approximately 20 Mbp distal of the domestication gene non-brittle rachis 1, *btr1* on the chromosome 3HS (Komatsuda *et al.* 2004; Wang *et al.* 2019). None of the wild barley accessions analyzed contained the resistance haplotype but it is enriched within landrace accessions (Figure 2, Supplementary Figure S2). The complete lack of the "resistance" marker 11_20968 haplotype in the wild barley could be explained by removal of the "susceptible" haplotype through a selective sweep within close proximity to the *btr1* region during domestication. These results show the importance of surveying both landraces and wild barley accessions since important resistance and/or susceptibility loci that interact in the barley—*P. teres* pathosystem may have been lost or gained through domestication. We have found loci that are unique to wild barley or landraces indicating the importance of analyses of the entire primary barley germplasm pool to identify new sources of resistance for future breeding efforts.

To date, only a handful of studies have utilized wild or landrace barley to map resistance loci using biparental populations (Metcalfe *et al.* 1970; Bockelman *et al.* 1977; Manninen *et al.* 2000, 2006; Williams *et al.* 2003; Koladia *et al.* 2017) or association mapping (Tamang *et al.* 2015; Richards *et al.* 2017; Vatter *et al.* 2017; Wonneberger *et al.* 2017a; Amezrou *et al.* 2018; Adhikari *et al.* 2019; Daba *et al.* 2019; Novakazi *et al.* 2019). Only one study has incorporated both *Ptm* and *Ptt* (Daba *et al.* 2019) and zero have investigated the wild and landrace barley specifically present within the center of origin of the Fertile Crescent. Despite using a reduced marker set (Figure 3) which will reduce the amount of MTAs identified (Cui *et al.* 2020), a total of 14 unique MTAs have been identified using modern mapping algorithms (Figure 4). However, the two isolates for which no significant MTAs were identified may be due to the fact that low marker density was utilized in these analyses. Four of the MTAs were potentially novel and two that mapped to previously identified *Ptt* resistance loci that had not been reported to be involved in *Ptm* interactions. Additionally, while the remaining MTAs may not be novel, they may represent important alleles that could be incorporated into breeding programs. Thus, the association mapping identified an abundance of net blotch resistance/susceptibility loci within wild and landrace barley from the center of origin.

In this study, barley chromosomes 3H and 7H contained the most MTAs with three, followed by two MTAs on chromosomes 1H, 5H, and 6H and one MTA on chromosomes 2H and 4H (Figure 4, Table 3). Of the 14 MTAs identified, four were identified against the *Ptm* isolates GPS263 and 13-179. The remaining ten MTAs were identified against *Ptt* isolates. When selecting the appropriate GWAS algorithm, the BLINK algorithm identified five MTAs, whereas the FarmCPU algorithm identified eleven MTAs, of which only two MTAs overlapped in both BLINK and FarmCPU algorithms. The SUPER algorithm identified one MTA using *Ptm* isolate 13-179 but this was also identified by the BLINK algorithm. Investigating model selection, the kinship (*K*) model identified 14 MTA, whereas the mixed kinship and population structure ($Q+K$) model only identified one MTA, which was also identified by the *K* model and therefore no unique MTA. Although we would encourage higher marker saturation in future studies to confirm locus novelty, we would suggest continued best practice of testing multiple models (naïve, *K*, *Q*, $Q+K$), along with the addition of including all modern association mapping algorithms based on

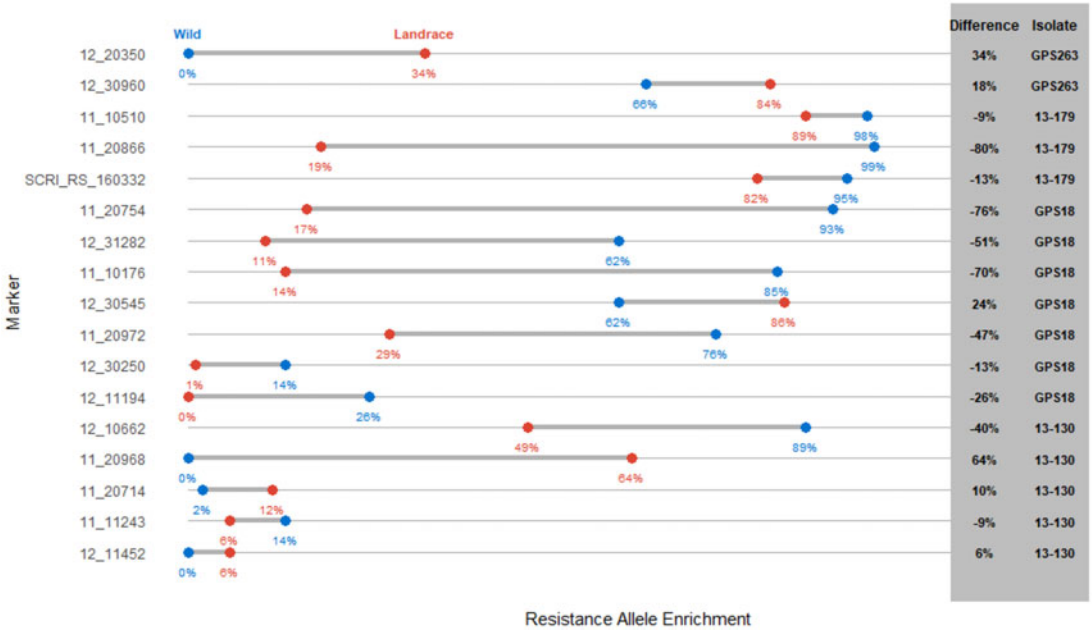

Figure 2 Enrichment dumbbell plot of enrichment of the resistance allele in landrace (red) and wild (blue) barley accession. Percentage is included below each data point and difference between the two barley classes as well as which isolate the loci was identified with.

Table 1 Phenotypic responses of wild barley and landraces to three *Pyrenophora teres* f. *maculata* isolates with absolute and percentage of accessions in each resistance class

Class	GPS263		13–179		13–167	
	Landrace	Wild	Landrace	Wild	Landrace	Wild
Resistant	6 (3%)	10 (10%)	0 (0%)	3 (3%)	6 (3%)	12 (12%)
Moderately resistant	42 (24%)	38 (39%)	65 (37%)	34 (35%)	66 (37%)	50 (51%)
Intermediate	42 (24%)	28 (29%)	105 (59%)	57 (58%)	100 (56%)	35 (36%)
Moderately susceptible	86 (49%)	21 (21%)	7 (4%)	4 (4%)	5 (3%)	1 (1%)
Susceptible	1 (1%)	1 (1%)	0 (0%)	0 (0%)	0 (0%)	0 (0%)
Mean score	5.4	4.4	4.3	4.3	4.2	3.6

The data are represented as the mean phenotypic score of each barley class to the respective isolate.

Table 2 Phenotypic responses of wild barley and landraces to three *Pyrenophora teres* f. *teres* isolates with absolute and percentage of accessions in each resistance class

Class	GPS18		UHK77		13-130	
	Landrace	Wild	Landrace	Wild	Landrace	Wild
Resistant	1 (1%)	1 (1%)	1 (1%)	7 (7%)	4 (2%)	8 (8%)
Moderately resistant	26 (15%)	32 (33%)	103 (58%)	32 (33%)	122 (69%)	44 (45%)
Intermediate	18 (10%)	14 (14%)	51 (29%)	18 (18%)	35 (20%)	21 (21%)
Moderately susceptible	131 (74%)	51 (52%)	22 (12%)	38 (39%)	16 (9%)	25 (26%)
Susceptible	1 (1%)	0 (0%)	0 (0%)	3 (3%)	0 (0%)	0 (0%)
Mean score	5.7	5.1	4.5	4.9	4.0	4.4

The data are represented as the mean phenotypic score of each barley class to the respective isolate.

the differing sets of MTAs identified using BLINK and FarmCPU algorithms.

The 14 MTAs identified in this study were compared to the collapsed loci of Clare *et al.* (2020) and a similar strategy of determining novel loci was dependent on the significance of the nearest neighbor marker and previous incorporation into a locus. Using this strategy, we identified four potentially novel loci (*QRpt-1H.1*, *QRpt-3H.1*, *QRpt-3H.3*, *QRpt-6H.1*) in the barley—*Ptt* interaction, and two novel loci in the barley—*Ptm* interaction (*QRpt-5H.1* and *QRpt-5H.2* corresponding to *NBP_QRptt5-1* and *Qrptts-5HL.1*, respectively) that had been previously described in the barley—*Ptt* interaction. The novel loci were detected on barley chromosomes 1H, 3H, and 6H. The *QRpt-1H.1* and *QRpt-6H.1* MTAs were associated in the interaction with *Ptt* isolate GPS18. *QRpt-1H.1* is

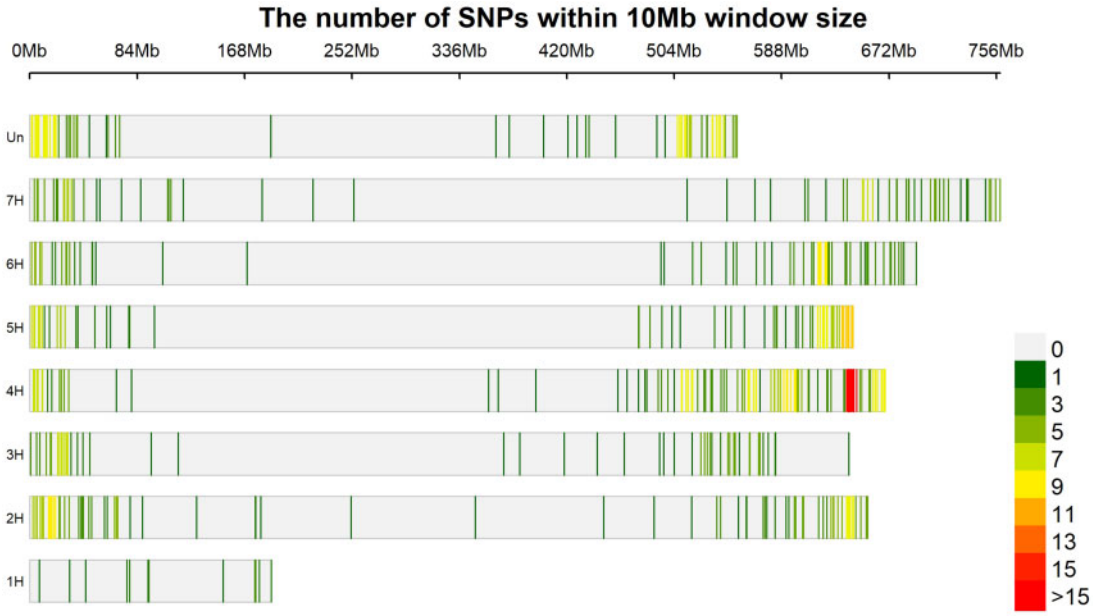

Figure 3 Marker density plot of all markers utilized within this study showing the distribution of markers across the barley genome with a window size of 10 Mb.

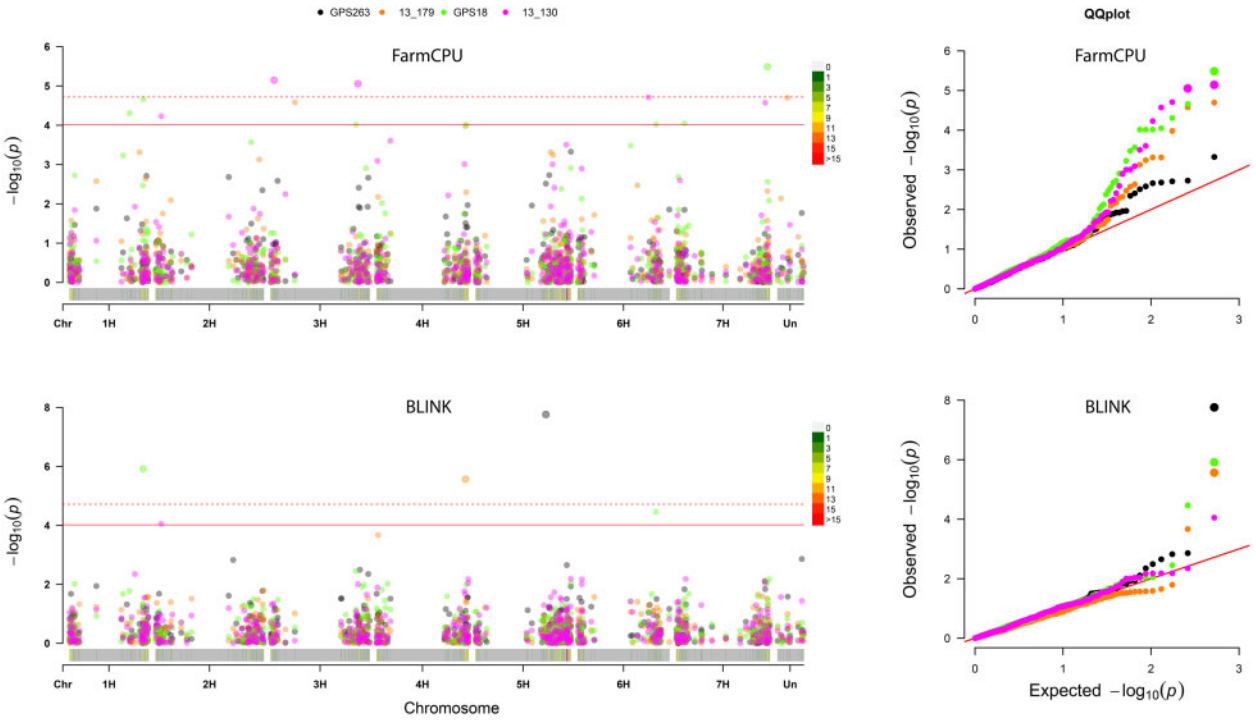

Figure 4 Manhattan and QQ plots for two models that show significant markers for more than one of the *Pyrenophora teres* f. *maculata* isolates GPS263 and 13–179, and *P. teres* f. *teres* isolates GPS18 and 13–130.

proximally flanked by the previously reported QTLs *NBP_QRptt1-1* (Wonneberger *et al.* 2017a) and a QTL identified at 57.3–62.8 cM (Rozanova *et al.* 2019) at distances of 13.6 Mbp and 18.2 Mbp, respectively, to the closest boundary of the delimited region of the loci (Clare *et al.* 2020). Markers located 18.2 Mbp proximal and 7.3 Mbp distal to *QRpt-1H.1* were not significant and therefore the MTA was deemed novel. Similarly, the closest locus to *QRpt-6H.1* is *Rpt5/Spt1* (Steffenson *et al.* 1996; Manninen *et al.* 2000; Raman *et al.* 2003; Read *et al.* 2003; Emebiri *et al.* 2005; Friesen *et al.* 2006; Manninen *et al.* 2006; Abu Qamar *et al.* 2008; Grewal *et al.* 2008; St. Pierre *et al.* 2010; Cakir *et al.* 2011; Gupta *et al.* 2011; Grewal *et al.* 2012; O'Boyle *et al.* 2014; Liu *et al.* 2015; Hisano *et al.* 2017; Islamovic *et al.* 2017; Koladia *et al.* 2017; Richards *et al.* 2017; Vatter *et al.* 2017; Wonneberger *et al.* 2017b; Amezrou *et al.* 2018; Martin *et al.* 2018; Adhikari *et al.* 2019; Daba *et al.* 2019; Novakazi *et al.* 2019; Rozanova *et al.* 2019; Adhikari *et al.* 2020) located 10.7 Mbp distal, however, a marker 28 Mbp distal to *QRpt-1H.1* and embedded within the *Rpt5/Spt1* locus was not significant, giving us reason to believe the locus is novel. The novel loci *QRpt-3H.1*, *QRpt-3H.3*, and *QRpt-6H.2* were all identified with *Ptt* isolate 13–

Table 3 Identified significant markers from genome wide association analysis order by isolate identified, followed by chromosome and base pair position

Loci	Marker	Chr*a*	Position*a*	Allele*b*	Isolate	Corresponding loci	Models identified	LOD score*c*
QRpt-5H.2	12_20350	5H	446449843	G/A	GPS263	*NBP_QRptt5-1*	K_{BLINK}	7.76
QRpt-3H.2	11_20866	3H	153156749	G/A	13_179	*QRptms3-2*	$K_{FarmCPU}$, PC1+$K_{FarmCPU}$	4.58
QRpt-4H.1	11_10510	4H	603258307	A/G		*Rpt8*	$K_{BLINK+SUPER}$, $PC5_{GLM}$, Q+K_{SUPER}, PC1+K_{SUPER}	5.57
QRpt-5H.1	SCRI_RS_160332	5H	474799503	A/G	*Qrptts-5HL.1*	$K_{FarmCPU}$	4.70	
QRpt-1H.1	11_10176	1H	397791042	G/C	GPS18	Novel	$K_{FarmCPU}$	4.31
QRpt-1H.2	11_20754	1H	483805599	C/G	*QPt.1H-1*	$K_{BLINK+FarmCPU}$	4.66–5.92	
QRpt-6H.2	11_20972	6H	539551443	T/A		*AL_QRptt6-2*	K_{BLINK}	4.46
QRpt-7H.1	12_30545	7H	54934072	A/G		*QNFNBAPR.Al/S-7Ha*	$K_{FarmCPU}$	4.05
QRpt-7H.3	12_31282	7H	617741299	C/T		QTL_{UHs}*-7H*	$K_{FarmCPU}$	5.49
QRpt-2H.1	12_11452	2H	34275254	G/A	13_130	*SFNB-2H-8-10*	$K_{BLINK+FarmCPU}$	4.05–4.23
QRpt-3H.1	11_20968	3H	19966889	A/G		Novel	$K_{FarmCPU}$	5.15
QRpt-3H.3	12_10662	3H	553445025	A/T		Novel	$K_{FarmCPU}$, Q+$K_{FarmCPU}$	4.22–5.06
QRpt-6H.1	11_20714	6H	489619101	G/A	Novel	$K_{FarmCPU}$	4.71	
QRpt-7H.2	11_11243	7H	601974526	A/G	*QRptm7-6*	$K_{FarmCPU}$	4.57	

Designation as well as predicted corresponding loci, models used to identify the marker and resistant/susceptibility alleles are also included.
[a] Location based on the second version of the Morex assembly (Monat *et al.*, 2019).
[b] Resistant/susceptible allele.
[c] LOD scores/ranges for BLINK and FarmCPU algorithms only.

130. The locus *QRpt-3H.1* is flanked by QTLs located at 12.1–17.4 cM (Rozanova *et al.* 2019) proximal and 53.42 cM (Tamang *et al.* 2019) distal on the Morex POPSEQ map (Mascher *et al.* 2013, 2017), equating to distances of 7.6 Mbp and 24 Mbp. The nearest neighbor markers are 5.2 Mbp proximal and 3.2 Mbp distal and are not significant indicating that this is potentially a novel locus. Similarly, the *QRpt-3H.3* locus is flanked by *QRpts3La* (Raman *et al.* 2003; Lehmensiek *et al.* 2007; Liu *et al.* 2015; Richards *et al.* 2017; Vatter *et al.* 2017; Daba *et al.* 2019) and *Rpt1* (Bockelman *et al.* 1977; Graner *et al.* 1996; Richter *et al.* 1998; Cakir *et al.* 2003; Raman *et al.* 2003; Manninen *et al.* 2006; Lehmensiek *et al.* 2007; Martin *et al.* 2018; Adhikari *et al.* 2019; Novakazi *et al.* 2019) approximately 4.6 Mbp proximal and 5 Mbp distal. However, the nearest neighbor markers are not significant at 6.2 Mbp proximal and 1.3 Mbp distal. The last novel locus, *QRpt-6H.2* is 4.1 Mbp proximal to *QPt.6H-3* (Daba *et al.* 2019); however, the nearest marker is 8.1 Mbp proximal and not significant.

Two loci that were previously implicated in *Ptt* resistance were also identified as novel MTAs for *Ptm* resistance in this study. The first locus, *QRpt-5H.1*, is located 3.0 Mbp distal to *Qrptts-5HL.1* (Richards *et al.* 2017) with marker SCRI_RS_160332. Additionally, since markers covering the *Qrptts-5HL.1* locus were not included in either panel, and due to the close proximity of *QRpt-5H.1* *Qrptts-5HL.1*, we believe that they are the same locus. The second locus, *QRpt-5H.2*, is embedded within the *NBP_QRptt5-1* locus (Wonneberger *et al.* 2017a). The remaining loci are all embedded within previously identified loci (Table 3).

Barley is predominately grown as a feed crop worldwide (IBGSC 2012), however in the United States where corn and soybeans are subsidized and used as feed crops, barley has been outcompeted and acreage has significantly dropped. This has pushed feed barley into less than optimal agricultural land due to its adaptability and hardiness (Brueggeman *et al.*, 2020). Quality malting barley demands premium prices because of its use in the multi-billion dollar added value brewing and distilling industry and is now the major class considered in breeding efforts in the US and Europe. However, in some regions of the world where traditional farming practices are still utilized barley is considered an important food crop (Helbaek 1969; Pourkheirandish and Komatsuda 2007; Geçit 2016; Ergün *et al.* 2017). Recent studies predicted that the effects of climate change will include higher temperatures, altered precipitation patterns, and higher disease pressure (Dawson *et al.* 2015), which could result in world malt barley shortages to supply the brewing and distilling industries (Xie *et al.* 2018). These predictions are beginning to be seen with the stagnating yields experienced in southern Europe (Dawson *et al.* 2015). Thus, barley breeding must maximize its potential in terms of quality and yield on the land it is currently afforded to sustain the demands for malting. One pillar of support for improving barley would be the introgression of predomestication resistance loci that are absent in current breeding programs to prevent substantial losses to net blotch, an important disease effecting barley production across the globe. Here, we report on the identification of novel loci from Turkish wild barley and landraces that could be introgressed into elite barley varieties.

Data availability

Isolates are available upon request. The authors ensure that all data necessary for confirming the conclusions of the article are present within the article, figures, and tables. Supplemental data has been submitted to figshare: https://doi.org/10.25387/g3.14725311. File SF1 contains genotyping data. File SF2 contains marker positions. File SF3 contains phenotyping data. Files SF4 and SF5 contain population structure and EMMA kindship matrix, respectively. File SF6 contains the mean-squared deviations of each association mapping model tested.

Author contributions

A.C.O., A.K., and R.S.B. conceived the study. A.C.O. and A.K. carried out phenotyping and DNA extractions. K.E. and D.S. carried out sequencing for genotyping data. S.C. and R.S.P. performed genotyping and analysis. S.C. wrote the manuscript with contributions from A.C.O., K.E., R.S.P. D.S., A.K., and R.B. All authors approved the final manuscript.

Acknowledgments

We thank the Western Regional Small Grains Genotyping Laboratory, specifically Travis Ruff and Marcus Hooker for the second Ion Torrent sequencing run, and to the staff and research personnel of the Central Research Institute for Field Crops (Ankara, Turkey) for their help.

Funding

The research presented in this manuscript was supported by the United States Department of Agriculture National Institute of Food and Agriculture (USDA-NIFA) award # 2018-67014-28491, the National Science Foundation grant award # 1759030, and the United States Department of Agriculture National Institute of Food and Agriculture Hatch project 1014919, Crop Improvement and Sustainable Production Systems (WSU reference 00011). Any opinions, findings, conclusions or recommendations expressed in this material are those of the author(s) and do not necessarily reflect the views of the United States Department of Agriculture National Institute of Food and Agriculture or the National Science Foundation.

Conflicts of interest

The authors declare that there is no conflict of interest.

Literature cited

Abu Qamar M, Liu ZH, Faris JD, Chao S, Edwards MC, *et al.* 2008. A region of barley chromosome 6H harbors multiple major genes associated with net type net blotch resistance. Theor Appl Genet. 117:1261–1270.

Adhikari A, Steffenson BJ, Smith MJ, Dill-Macky R. 2019. Genome-wide association mapping of seedling net form net blotch resistance in an Ethiopian and Eritrean barley collection. Crop Sci. 59:1625–1638.

Adhikari A, Steffenson BJJ, Smith KPP, Smith M, Dill-Macky R. 2020. Identification of quantitative trait loci for net form net blotch resistance in contemporary barley breeding germplasm from the USA using genome-wide association mapping. Theor Appl Genet. 133:1019–1037.

Afanasenko OS, Jalli M, Pinnschmidt HO, Filatova O, Platz GJ. 2009. Development of an international standard set of barley differential genotypes for *Pyrenophora teres* f. *teres*. Plant Pathol. 58: 665–676.

Afanasenko OS, Koziakov AV, Hedlay PE, Lashina NM, Anisimova AV, *et al.* 2015. Mapping of the loci controlling the resistance to *Pyrenophora teres* f. *teres* and *Cochliobolus sativus* in two double haploid barley populations. Russ J Genet Appl Res. 5:242–253.

Akhavan A, Turkington TK, Kebede B, Tekauz A, Kutcher HR, *et al.* 2015. Prevalence of mating type idiomorphs in *Pyrenophora teres* f. *teres* and *P. teres* f. *maculata* populations from the Canadian prairies. Can J Plant Pathol. 37:52–60.

Allard RW, Bradshaw AD. 1964. Implications of genotype-environmental interactions in applied plant breeding. Crop Sci. 4: 503–508.

Amezrou R, Verma RPS, Chao S, Brueggeman RS, Belqadi L, *et al.* 2018. Genome-wide association studies of net form of net blotch resistance at seedling and adult plant stages in spring barley collection. Mol Breed. 38:58.

Beier S, Himmelbach A, Colmsee C, Zhang X-Q, Barrero RA, *et al.* 2017. Construction of a map-based reference genome sequence for barley, *Hordeum vulgare* L. Sci Data. 4:170044.

Bockelman HE, Sharp EL, Bjarko ME. 1983. Isolates of *Pyrenophora teres* from Montana and the Mediterranean region that produce spot-type lesions on barley. Plant Dis. 67:696–697.

Bockelman HE, Sharp EL, Eslick RF. 1977. Trisomic analysis of genes for resistance to scald and net blotch in several barley cultivars. Can J Bot. 55:2142–2148.

Bradbury PJ, Zhang Z, Kroon DE, Casstevens TM, Ramdoss Y, *et al.* 2007. TASSEL: software for association mapping of complex traits in diverse samples. Bioinformatics. 23:2633–2635.

Brueggeman RS, Solanki S, Ameen G, Effertz K, Poudel RS, *et al.* 2020. Fungal diseases affecting barley. In: G, Fox C Li, editors. Achieving Sustainable Cultivation of Barley. Cambridge, UK: Burleigh Dodds Science Publishing Limited. p. 1–58.

Burlakoti RR, Gyawali S, Chao S, Smith KP, Horsley RD, *et al.* 2017. Genome-wide association study of spot form of net blotch resistance in the Upper Midwest barley breeding programs. Phytopathology. 107:100–108.

Cakir M, Gupta S, Li C, Hayden M, Mather DE, *et al.* 2011. Genetic mapping and QTL analysis of disease resistance traits in the barley population Baudin × AC Metcalfe. Crop Pasture Sci. 62: 152–161.

Cakir M, Gupta S, Platz GJ, Ablett GA, Loughman R, *et al.* 2003. Mapping and validation of the genes for resistance to *Pyrenophora teres* f. *teres* in barley (*Hordeum vulgare* L.). Aust J Agric Res. 54: 1369–1377.

Campbell GF, Crous PW. 2003. Genetic stability of net × spot hybrid progeny of the barley pathogen *Pyrenophora teres*. Austral Plant Pathol. 32:283–287.

Campbell GF, Lucas JA, Crous PW. 2002. Evidence of recombination between net- and spot-type populations of *Pyrenophora teres* as determined by RAPD analysis. Mycol. Res. 106:602–608.

Ceccarelli S. 1996. Adaptation to low/high input cultivation. Euphytica. 92:203–214.

Ceccarelli S, Grando S. 2000. Barley landraces from the Fertile Crescent. A lesson for plant breeders. In: SB Brush, editor. Genes in the Field, on –Farm Conservation of Crop Diversity. Boca Raton, London, New York, Washington, DC: IPGRI/IDRC/Lewis Publishers. p. 51–76.

Çelik Oğuz A, Karakaya A. 2017. Pathotypes of *Pyrenophora teres* on barley in Turkey. Phytopathol Mediterr. 56:224–234.

Çelik Oğuz A, Karakaya A, Duran RM, Özbek K. 2019a. Identification of *Hordeum spontaneum* genotypes resistant to net blotch disease. Tarım Bilim Derg. 25:115–122.

Çelik Oğuz A, Karakaya A, Ergün N, Sayim İ. 2017. Turkish barley landraces resistant to net and spot forms of *Pyrenophora teres*. Phytopathol Mediterr. 56:217–223.

Çelik Oğuz A, Ölmez F, Karakaya A. 2018. Mating type idiomorphs of *Pyrenophora teres* in Turkey. Zemdirbyste Agric. 105:271–278.

Çelik Oğuz A, Ölmez F, Karakaya A. 2019b. Genetic diversity of net blotch pathogens of barley in Turkey. Int J Agric Biol. 21: 1089–1096.

Çelik Oğuz A, Rahimi A, Karakaya A. 2019c. Seedling response of Iranian barley landraces to *Pyrenophora teres* f. *teres* and *Pyrenophora teres* f. *maculata*. Tarım Bilim Derg. 25:86–92.

Clare SJ, Wyatt NA, Brueggeman RS, Friesen TL. 2020. Research advances in the *Pyrenophora teres*–barley interaction. Mol Plant Pathol. 21:272–288.

Cui Z, Dong H, Zhang A, Ruan Y, Jiang S, *et al.* 2020. Denser markers and advanced statistical method identified more genetic loci associated with husk traits in maize. Sci Rep. 10:8165.

Daba SD, Horsley R, Brueggeman R, Chao S, Mohammadi M. 2019. Genome-wide association studies and candidate gene identification for leaf scald and net blotch in barley (Hordeum vulgare L.). Plant Dis. 103:880–889.

Dawson IK, Russell J, Powell W, Steffenson B, Thomas WTB, *et al.* 2015. Barley: a translational model for adaptation to climate change. New Phytol. 206:913–931.

Earl DA, vonHoldt BM. 2012. STRUCTURE HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. Conserv Genet Resour. 4:359–361.

Ellis RP, Forster BP, Robinson D, Handley LL, Gordon DC, *et al.* 2000. Wild barley: a source of genes for crop improvement in the 21st century? J Exp Bot. 51:9–17.

Emebiri LC, Platz CG, Moody DB. 2005. Disease resistance genes in a doubled haploid population of two-rowed barley segregating for malting quality attributes. Aust J Agric Res. 56:49–56.

Endresen DTF, Street K, Mackay M, Bari A, De Pauw E. 2011. Predictive association between biotic stress traits and eco-geographic data for wheat and barley landraces. Crop Sci. 51: 2036–2055.

Ergün N, S. Aydoğan İ, Sayim A, Karakaya AÇ., Oğuz 2017. Arpa (*Hordeum vulgare* L.) köy çeşitlerinde tane verimi ve bazı tarımsal özelliklerin incelenmesi. Tarla Bitk Merk Araştırma Enstitüsü Derg. 26:180–189.

Evanno G, Regnaut S, Goudet J. 2005. Detecting the number of clusters of individuals using the software structure: a simulation study. Mol Ecol. 14:2611–2620.

Fetch TG, Steffenson BJ, Nevo E. 2003. Diversity and sources of multiple disease resistance in *Hordeum spontaneum*. Plant Dis. 87: 1439–1448.

Franckowiak JD, Platz GJ. 2013. International database for barley genes and barley genetic stocks. Barley Genet Newsl. 43:48–223.

Friesen TL, Faris JD, Lai Z, Steffenson BJ. 2006. Identification and chromosomal location of major genes for resistance to *Pyrenophora teres* in a doubled-haploid barley population. Genome. 49:855–859.

Geçit HH. 2016. Serin İklim Tahılları (Buğday, Arpa, Yulaf, Triticale). Yayınları, Yayın No:1640, Ankara: Ankara Üniversitesi Ziraat Fakültesi.

Geschele EE. 1928. The response of barley to parasitic fungi *Helminthosporium teres* Sacc. Bull Appl Bot Genet Plant Breed. 19: 371–384 (in Rev. Appl. Mycol. 8, 165).

Graner A, Foroughi-Wehr B, Tekauz A. 1996. RFLP mapping of a gene in barley conferring resistance to net blotch (*Pyrenophora teres*). Euphytica. 91:229–234.

Grewal TS, Rossnagel BG, Pozniak CJ, Scoles GJ. 2008. Mapping quantitative trait loci associated with barley net blotch resistance. Theor Appl Genet. 116:529–539.

Grewal TS, Rossnagel BG, Scoles GJ. 2012. Mapping quantitative trait loci associated with spot blotch and net blotch resistance in a doubled-haploid barley population. Mol Breed. 30:267–279.

Gupta S, Li C, Loughman R, Cakir M, Westcott S, *et al.* 2011. Identifying genetic complexity of 6H locus in barley conferring resistance to *Pyrenophora teres* f. *teres*. Plant Breed. 130:423–429.

Gyawali S, Amezrou R, Verma RPS, Brueggeman R, Rehman S, *et al.* 2019. Seedling and adult stage resistance to spot form of net blotch (SFNB) in spring barley and stability of adult stage resistance to SFNB in Morocco. Eur J Plant Pathol. 153:475–487.

Gyawali S, Amezrou R, Verma RPS, Brueggeman R, Rehman S, *et al.* 2020. Seedling and adult stage resistance to net form of net blotch (NFNB) in spring barley and stability of adult stage resistance to NFNB in Morocco. J Phytopathol. 168:254–266.

Harlan JR, Zohary D. 1966. Distribution of wild wheats and barley. Science. 153:1074–1080.

Helbaek H. 1969. Plant-collecting, dry-farming and irrigation agriculture in prehistoric Deh Luran. In: F Hole, KV Flannery, JA Neely, editors. Prehistory and Human Ecology of the Deh Luran Plain. An Early Village Sequence from Khuzistan, Iran. University of Michigan. p. 383–426.

Hisano H, Sakamoto K, Takagi H, Terauchi R, Sato K. 2017. Exome QTL-seq maps monogenic locus and QTLs in barley. BMC Genomics. 18:125.

Huang M, Liu X, Zhou Y, Summers RM, Zhang Z. 2019. BLINK: a package for the next level of genome-wide association studies with both individuals and markers in the millions. Gigascience. 8: 1–12.

IBGSC. 2012. A physical, genetic and functional sequence assembly of the barley genome. Nature. 491:711–717.

Islamovic E, Bregitzer P, Friesen TL. 2017. Barley 4H QTL confers NFNB resistance to a global set of *P. teres* f. *teres* isolates. Mol. Breed. 37:29.

Jakob SS, Rödder D, Engler JO, Shaaf S, Ozkan H, *et al.* 2014. Evolutionary history of wild barley (*Hordeum vulgare* subsp. *spontaneum*) analyzed using multilocus sequence data and paleodistribution modeling. Genome Biol Evol. 6:685–702.

Jana S, Bailey KL. 1995. Responses of wild and cultivated barley from West Asia to net blotch and spot blotch. Crop Sci. 35:242–246.

Kang HM, Zaitlen NA, Wade CM, Kirby A, Heckerman D, *et al.* 2008. Efficient control of population structure in model organism association mapping. Genetics. 178:1709–1723.

Karakaya A, Çelik Oğuz A, Saraç Sivrikaya I. 2020. Diseases occurring on Hordeum bulbosum field populations at Bingöl province of Turkey. Work Fac Agric Food Sci Univ Sarajev LXV:75–81.

Karakaya A, Katırcıoğlu YZ, Aktaş H. 2004. Studies on the biology of *Drechslera teres* under Ankara conditions. Tarım Bilim Derg. 10: 133–135.

Karakaya A, Mert Z, Çelik Oğuz A, Ertaş MN, Karagöz A. 2016. Determination of the diseases occurring on naturally growing wild barley (*Hordeum spontaneum*) field populations. Work Fac Agric Univ Sarajev. 61:291–295.

Koladia VM, Faris JD, Richards JK, Brueggeman RS, Chao S, *et al.* 2017. Genetic analysis of net form net blotch resistance in barley lines CIho 5791 and Tifang against a global collection of *P. teres* f. *teres* isolates. Theor Appl Genet. 130:163–173.

Komatsuda T, Maxim P, Senthil N, Mano Y. 2004. High-density AFLP map of nonbrittle rachis 1 (*btr1*) and 2 (*btr2*) genes in barley (*Hordeum vulgare* L.). Theor Appl Genet. 109:986–995.

König J, Perovic D, Kopahnke D, Ordon F. 2013. Development of an efficient method for assessing resistance to the net type of net blotch (*Pyrenophora teres* f. *teres*) in winter barley and mapping of quantitative trait loci for resistance. Mol Breed. 32:641–650.

Kün E. 1996. Tahıllar-1 (Serin İklim Tahılları). Yayınları, Yayın No: 1451, Ankara: Ankara Üniversitesi Ziraat Fakültesi.

Lakew B, Semeane Y, Alemayehu F. 1995. Evaluation of Ethiopian barley landraces for disease and agronomic characters. Rachis. 14:21–25.

Legge WG, Metcalfe DR, Chiko AW, Martens JW, Tekauz A. 1996. Reaction of Turkish barley accessions to Canadian barley pathogens. Can J Plant Sci. 76:927–931.

Lehmensiek A, Platz GJ, Mace E, Poulsen D, Sutherland MW. 2007. Mapping of adult plant resistance to net form of net blotch in three Australian barley populations. Aust J Agric Res. 58:1191–1197.

Li M, Liu X, Bradbury P, Yu J, Zhang Y-M, *et al.* 2014. Enrichment of statistical power for genome-wide association studies. BMC Biol. 12:73.

Liu Z, Ellwood SR, Oliver RP, Friesen TL. 2011. *Pyrenophora teres*: profile of an increasingly damaging barley pathogen. Mol Plant Pathol. 12:1–19.

Liu Z, Holmes DJ, Faris JD, Chao S, Brueggeman RS, *et al.* 2015. Necrotrophic effector-triggered susceptibility (NETS) underlies the barley–*Pyrenophora teres* f. *teres* interaction specific to chromosome 6H. Mol Plant Pathol. 16:188–200.

Liu X, Huang M, Fan B, Buckler ES, Zhang Z. 2016. Iterative usage of fixed and random effect models for powerful and efficient genome-wide association studies. PLOS Genet. 12:e1005767.

Loiselle BA, Sork VL, Nason J, Graham C. 1995. Spatial genetic structure of a tropical understory shrub, *Psychotria officinalis* (Rubiacese). Am J Bot. 82:1420–1425.

Mamidi S, Chikara S, Goos RJ, Hyten DL, Annam D, *et al.* 2011. Genome-wide association analysis identifies candidate genes associated with iron deficiency chlorosis in soybean. Plant Genome. 4:154–164.

Manninen OM, Jalli M, Kalendar R, Schulman A, Afanasenko O, *et al.* 2006. Mapping of major spot-type and net-type net-blotch resistance genes in the Ethiopian barley line CI 9819. Genome. 49: 1564–1571.

Manninen O, Kalendar R, Robinson J, Schulman AH. 2000. Application of BARE-1 retrotransposon markers to the mapping of a major resistance gene for net blotch in barley. Mol Gen Genet. 264:325–334.

Martin A, Platz GJ, de Klerk D, Fowler RA, Smit F, *et al.* 2018. Identification and mapping of net form of net blotch resistance in South African barley. Mol Breed. 38:53.

Mascher M, Gundlach H, Himmelbach A, Beier S, Twardziok SO, *et al.* 2017. A chromosome conformation capture ordered sequence of the barley genome. Nature. 544:427–433.

Mascher M, Muehlbauer GJ, Rokhsar DS, Chapman J, Schmutz J, *et al.* 2013. Anchoring and ordering NGS contig assemblies by population sequencing (POPSEQ). Plant J. 76:718–727.

Mathre DE, editor. 1982. Compendium of Barley Diseases. St. Paul, MN: APS Press.

McDonald BA, Linde C. 2002. Pathogen population genetics, evolutioanry potential, and durable resistance. Annu Rev Phytopathol. 40:349–379.

McLean MS, Martin A, Gupta S, Sutherland MW, Hollaway GJ, *et al.* 2014. Validation of a new spot form of net blotch differential set and evidence for hybridisation between the spot and net forms of net blotch in Australia. Australasian Plant Pathol. 43:223–233.

Metcalfe DR, Buchannon KW, McDonald WC, Reinbergs E. 1970. Relationships between the 'Jet' and "Milton" genes for resistance to loose smut genes for reistance to other barley dieases. Can J Plant Sci. 50:423–427.

Mikhailova LA, Ternyuk IG, Mironenko NV. 2010. *Pyrenophora teres*, an agent causing wheat leaf spot. Microbiology. 79:561–565.

Mode CJ, Schaller CW. 1958. Two additional factors for host resistance to net blotch in barley. Agron J. 50:15–18.

Monat C, Padmarasu S, Lux T, Wicker T, Gundlach H, *et al.* 2019. TRITEX: chromosome-scale sequence assembly of Triticeae genomes with open-source tools. Genome Biol. 20:284.

Money D, Gardner K, Migicovsky Z, Schwaninger H, Zhong G-Y, *et al.* 2015. LinkImpute: Fast and accurate genotype imputation for nonmodel organisms. G3 (Bethesda)). 5:2383–2390.

Moya PA, Girotti JR, Toledo AV, Sisterna MN. 2018. Antifungal activity of Trichoderma VOCs against *Pyrenophora teres*, the causal agent of barley net blotch. J. Plant Prot. Res. 58:45–53.

Muñoz-Amatriaín M, Cuesta-Marcos A, Endelman JB, Comadran J, Bonman JM, *et al.* 2014. The USDA barley core collection: Genetic diversity, population structure, and potential for genome-wide association studies (L. Yan, Ed.). PLoS One. 9:e94688.

Neupane A, Tamang P, Brueggeman RS, Friesen TL. 2015. Evaluation of a barley core collection for spot form net blotch reaction reveals distinct genotype-specific pathogen virulence and host susceptibility. Phytopathology. 105:509–517.

Nevo E. 1992. Origin, evolution, population genetics and resources for breeding of wild barley, *Hordeum spontaneum*, in the Fertile Crescent. In: PR Shewry, editor. Barley: Genetics, Biochemistry, Molecular Biology and Biotechnology. C.A.B. International UK. p. 19–43.

Novakazi F, Afanasenko O, Anisimova A, Platz GJJ, Snowdon R, *et al.* 2019. Genetic analysis of a worldwide barley collection for resistance to net form of net blotch disease (*Pyrenophora teres* f. *teres*). Theor Appl Genet. 132:2633–2650.

O'Boyle PD, Brooks WS, Barnett MD, Berger GL, Steffenson BJ, *et al.* 2014. Mapping net blotch resistance in 'Nomini' and CIho 2291 barley. Crop Sci. 54:2596–2602.

Perelló AE, Couretot L, Curti A, Uranga JP, Consolo VF. 2019. First report of spot lesion of wheat caused by *Pyrenophora teres* f. sp *maculata* observed in Argentina. Crop Prot. 122:19–22.

Piening L, Kaufmann ML. 1969. Comparison of the effectors of net blotch and leaf removal on yield in barley. Can J Plant Sci. 49: 731–735.

Poets AM, Fang Z, Clegg MT, Morrell PL. 2015. Barley landraces are characterized by geographically heterogeneous genomic origins. Genome Biol. 16:173.

Poudel B, McLean MS, Platz GJ, McIlroy JA, Sutherland MW, *et al.* 2019. Investigating hybridisation between the forms of *Pyrenophora teres* based on Australian barley field experiments and cultural collections. Eur J Plant Pathol. 153:465–473.

Pourkheirandish M, Komatsuda T. 2007. The importance of barley genetics and domestication in a global perspective. Ann Bot. 100: 999–1008.

Pritchard JK, Stephens M, Donnelly P. 2000. Inference of population structure using multilocus genotype data. Genetics. 155:945–959.

Raman H, Platz GJ, Chalmers KJ, Raman R, Read BJ, *et al.* 2003. Mapping of genomic regions associated with net form of net blotch resistance in barley. Aust J Agric Res. 54:1359–1367.

Read BJ, Raman H, McMichael G, Chalmers KJ, Ablett GA, *et al.* 2003. Mapping and QTL analysis of the barley population Sloop × Halcyon. Aust J Agric Res. 54:1145–1153.

Richards J, Chao S, Friesen T, Brueggeman R. 2016. Fine mapping of the barley chromosome 6H net form net blotch susceptibility locus. G3 (Bethesda)). 6:1809–1818.

Richards JK, Friesen TL, Brueggeman RS. 2017. Association mapping utilizing diverse barley lines reveals net form net blotch seedling resistance/susceptibility loci. Theor Appl Genet. 130:915–927.

Richter K, Schondelmaier J, Jung C. 1998. Mapping of quantitative trait loci affecting *Drechslera teres* resistance in barley with molecular markers. Theor Appl Genet. 97:1225–1234.

Robinson J, Jalli M. 1996. Diversity among Finnish net blotch isolates and resistance in barley. Euphytica. 92:81–87.

Rozanova IV, Lashina NM, Mustafin ZS, Gorobets SA, Efimov VM, *et al.* 2019. SNPs associated with barley resistance to isolates of *Pyrenophora teres* f. *teres*. BMC Genomics. 20:292.

Ruff TM, Marston EJ, Eagle JD, Sthapit SR, Hooker MA, *et al.* 2020. Genotyping by multiplexed sequencing (GMS): A customizable platform for genomic selection. PLoS One. 15: e0229207.

Sato K, Takeda K. 1997. Net blotch resistance in wild species of *Hordeum*. Euphytica. 95:179–185.

Schaller CW. 1955. Inheritance of resistance to net blotch of barley. Phytopathology. 45:174–176.

Segura V, Vilhjálmsson BJ, Platt A, Korte A, Seren Ü, *et al.* 2012. An efficient multi-locus mixed-model approach for genome-wide association studies in structured populations. Nat Genet. 44: 825–830.

Serenius M, Mironenko N, Manninen O. 2005. Genetic variation, occurrence of mating types and different forms of *Pyrenophora teres* causing net blotch of barley in Finland. Mycol Res. 109:809–817.

Sharma Poudel R, Al-Hashel AF, Gross T, Gross P, Brueggeman R. 2018. Pyramiding *rpg4*- and *Rpg1*-mediated stem rust resistance in barley requires the *Rrr1* gene for both to function. Front Plant Sci. 9:1789.

Silvar C, Casas AM, Kopahnke D, Habekuß A, Schweizer G, *et al.* 2010. Screening the Spanish barley core collection for disease resistance. Plant Breed. 129:45–52.

Smedegård-Petersen V. 1971. *Pyrenophora teres* f. *maculata* f. nov. and *Pyrenophora teres* f. *teres* on barley in Denmark. In: 'Yearbook of the Royal Veterinary and Agricultural University (Copenhagen)'. pp. 124–144.

Steffenson BJ, Hayes PM, Kleinhofs A. 1996. Genetics of seedling and adult plant resistance to net blotch (*Pyrenophora teres* f. *teres*) and spot blotch (*Cochliobolus sativus*) in barley. Theor Appl Genet. 92: 552–558.

St. Pierre S, Gustus C, Steffenson B, Dill-Macky R, Smith KP. 2010. Mapping net form net blotch and Septoria speckled leaf blotch resistance loci in barley. Phytopathology. 100:80–84.

Tamang P, Neupane A, Mamidi S, Friesen T, Brueggeman R. 2015. Association mapping of seedling resistance to spot form net blotch in a worldwide collection of barley. Phytopathology. 105: 500–508.

Tamang P, Richards JK, Alhashal A, Sharma Poudel R, Horsley RD, *et al.* 2019. Mapping of barley susceptibility/resistance QTL against spot form net blotch caused by *Pyrenophora teres* f. maculata using RIL populations. Theor Appl Genet. 132:1953–1963.

Tekauz A. 1985. A numerical scale to classify reactions of barley to *Pyrenophora teres*. Can J Plant Pathol. 7:181–183.

Thomas WTB, Baird E, Fuller JD, Lawrence P, Young GR, *et al.* 1998. Identification of a QTL decreasing yield in barley linked to Mlo powdery mildew resistance. Mol Breed. 4:381–393.

Tóth B, Csősz M, Kopahnke D, Varga J. 2008. First report on *Pyrenophora teres* causing lesions of wheat leaves in Hungary. Plant Pathol. 57:385.

Usta P, Karakaya A, Çelik Oğuz A, Mert Z, Akan K, *et al.* 2014. Determination of the seedling reactions of twenty barley cultivars to six isolates of *Drechslera teres* f. *maculata*. Anadolu J Agr Sci. 29:20–25.

VanRaden PM. 2008. Efficient methods to compute genomic predictions. J Dairy Sci. 91:4414–4423.

Vatter T, Maurer A, Kopahnke D, Perovic D, Ordon F, *et al.* 2017. A nested association mapping population identifies multiple small effect QTL conferring resistance against net blotch (*Pyrenophora teres* f. *teres*) in wild barley. PLoS One. 12:e0186803.

Wang X, Mace ES, Platz GJ, Hunt CH, Hickey LT, *et al.* 2015. Spot form of net blotch resistance in barley is under complex genetic control. Theor Appl Genet. 128:489–499.

Wang Q, Tian F, Pan Y, Buckler ES, Zhang Z. 2014. A SUPER powerful method for genome wide association study. PLoS One. 9:e107684.

Wang YL, Ye H, Liu L, Wu JH, Ru WM, *et al.* 2019. Molecular insights on the domestication of barley (*Hordeum vulgare* L.). Crit Rev Plant Sci. 38:280–294.

Wang J, Zhang Z. 2020. GAPIT Version 3: boosting power and accuracy for genomic association and prediction. bioRxiv. doi: 10.1101/2020.11.29.403170.

Williams KJ, Lichon A, Gianquitto P, Kretschmer JM, Karakousis A, *et al.* 1999. Identification and mapping of a gene conferring resistance to the spot form of net blotch (*Pyrenophora teres* f. *maculata*) in barley. Theor Appl Genet. 99:323–327.

Williams KJ, Platz GJ, Barr AR, Cheong J, Willsmore K, *et al.* 2003. A comparison of the genetics of seedling and adult plant resistance to the spot form of net blotch (*Pyrenophora teres* f. *maculata*). Aust J Agric Res. 54:1387–1394.

Wonneberger R, Ficke A, Lillemo M. 2017a. Identification of quantitative trait loci associated with resistance to net form net blotch in a collection of Nordic barley germplasm. Theor Appl Genet. 130: 2025–2043.

Wonneberger R, Ficke A, Lillemo M. 2017b. Mapping of quantitative trait loci associated with resistance to net form net blotch (*Pyrenophora teres* f. *teres*) in a doubled haploid Norwegian barley population. PLoS One. 12:e0175773.

Xie W, Xiong W, Pan J, Ali T, Cui Q, *et al.* 2018. Decreases in global beer supply due to extreme drought and heat. Nat Plants. 4: 964–973.

Yazıcı B, Karakaya A, Oğuz AÇ, Mert Z. 2015. Determination of the seedling reactions of some barley cultivars to *Drechslera teres* f. *teres*. Bitki Koruma Bülteni. 55:239–245.

Yitbarek S, Berhane L, Fikadu A, Van Leur JAG, Grando S, *et al.* 1998. Variation in Ethiopian barley landrace populations for resistance to barley leaf scald and net blotch. Plant Breed. 117: 419–423.

Yu J, Pressoir G, Briggs WH, Vroh Bi I, Yamasaki M, *et al.* 2006. A unified mixed-model method for association mapping that accounts for multiple levels of relatedness. Nat Genet. 38: 203–208.

Yun SJ, Gyenis L, Hayes PM, Matus I, Smith KP, *et al.* 2005. Quantitative trait loci for multiple disease resistance in wild barley. Crop Sci. 45:2563–2572.

Zhang Z, Ersoz E, Lai C-Q, Todhunter RJ, Tiwari HK, *et al.* 2010. Mixed linear model approach adapted for genome-wide association studies. Nat Genet. 42:355–360.

Zohary D, Hopf M. 2000. Domestication of Plants in the Old World: The Origin and Spread of Cultivated Plants in West Asia, Europe, and the Nile Valley. New York: Oxford University Press.

Communicating editor: R. Wisser