Research Article

SP110 Polymorphisms Are Genetic Markers for Vulnerability to Latent and Active Tuberculosis Infection in Taiwan

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Received 12 September 2018; Accepted 15 November 2018; Published 5 December 2018

Academic Editor: Frank Tacke

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One-fourth of the human population is estimated to have been exposed to Mycobacterium tuberculosis (Mtb) and carries the infection in its latent form. This latent infection presents a lifelong risk of developing active tuberculosis (TB) disease, and persons with latent TB infection (LTBI) are significant contributors to the pool of active TB cases. Genetic polymorphisms among hosts have been shown to contribute to the outcome of Mtb infection. The SP110 gene, which encodes an interferoninduced nuclear protein, has been shown to control host innate immunity to Mtb infection. In this study, we provide experimental data demonstrating the ability of the gene to control genetic susceptibility to latent and active TB infection. Genetic variants of the SP110 gene were investigated in the Taiwanese population (including 301 pulmonary TB patients, 68 LTBI individuals, and 278 healthy household contacts of the TB patients), and their association with susceptibility to latent and active TB infection was examined by performing an association analysis in a case-control study. We identified several SNPs (rs7580900, rs7580912, rs9061, rs11556887, and rs2241525) in the SP110 gene that are associated with susceptibility to LTBI and/or TB disease. Our studies further showed that the same SNPs may have opposite effects on the control of susceptibility to LTBI versus TB. In addition, our analyses demonstrated that the SP110 rs9061 SNP was associated with tumor necrosis factor- α $(TNF\alpha)$ levels in plasma in LTBI subjects. The results suggest that the polymorphisms within SP110 have a role in controlling genetic susceptibility to latent and active TB infection in humans. To the best of our knowledge, this is the first report showing that the SP110 variants are associated with susceptibility to LTBI. Our study also demonstrated that the identified SP110 SNPs displayed the potential to predict the risk of LTBI and subsequent TB progression in Taiwan.

1. Introduction

Tuberculosis (TB), caused by *Mycobacterium tuberculosis* (*Mtb*) infection, remains one of the top ten causes of death in the world [1]. Approximately one-fourth of the world population has been infected with *Mtb* [2], but only 10% of these infected persons develop progressive disease during their lifetimes [3]. The majority of infected individuals remain healthy and noninfectious but carry *Mtb* in a latent form. Latent TB infection (LTBI) is a state during which a

persistent host immune response to stimulation by *Mtb* antigens is sustained without evidence of clinically manifested active TB [4, 5]. As many as 10% of people with LTBI will go on to develop progressive disease in the near or remote future (a process named "TB reactivation"), and the risk is significantly higher in the presence of predisposing factors, such as coinfection with human immunodeficiency virus (HIV).

Persons with LTBI can be diagnosed by skin (tuberculin skin test (TST)) and/or blood (interferon-gamma (IFN γ)

release assay (IGRA)) tests [6]. A positive result from these assays indicates an immune response to stimulation by *Mtb* antigens in the LTBI population, despite the fact that these individuals have a negative bacteriological test. However, neither TST nor IGRA can distinguish between LTBI and active TB disease or predict who will progress to active TB [7]. Given that the LTBI individuals represent a potential reservoir for future active TB cases, preventive therapy for LTBI is as important a goal as timely anti-TB treatment to reduce the burden of TB [8]. Therefore, identifying and treating cases with LTBI will contribute to TB elimination. Fundamental research for the development of diagnostic assays with improved performance and predictive assessment for TB reactivation will have practical applications and offer a substantial benefit for LTBI management.

In both humans and experimental animal models, genetic polymorphisms among hosts have been shown to contribute to the outcome of *Mtb* infection [9–13]. In mice, the Ipr1 (intracellular pathogen resistance 1) gene is located within the sst1 (supersusceptibility to tuberculosis 1) locus on chromosome 1 (49-54 cM) [14] and has been identified as a genetic determinant conferring host innate immunity to Mtb infection [15]. The previous studies indicate that the *Ipr1* gene may function to integrate mechanisms on controlling cell death, innate immunity, and pathogenesis during intracellular pathogen infections [15, 16]. The gene orthologous to the mouse Ipr1 in humans is SP110, located on chromosome 2q37.1. Expression of both Ipr1 and SP110 genes is intensively regulated by IFNs, suggesting that the function of both genes is related to the IFN-mediated immune response [17].

Genetic defects in the SP110 gene have been found to be responsible for hepatic veno-occlusive disease and immunodeficiency [18, 19], indicating that the gene plays important roles in immunity [20]. The SP110 gene encodes the SP110 nuclear body protein, which has at least three isoforms, including the dominantly expressed SP110a, b, and c isoforms that are believed to be the result of alternative mRNA splicing. Our recent study demonstrated that SP110b, which is most similar to mouse Ipr1 and is expressed more abundantly than SP110a and SP110c, modulates nuclear factor- κB (NF- κB) activity resulting in the downregulation of tumor necrosis factor- α (TNF α) production and concomitant upregulation of NF- κ B-induced antiapoptotic gene expression, thereby suppressing IFNy-mediated monocyte/ macrophage cell death [21]. This indicates that the protein is crucial in the control of the activation of macrophages, the reservoir for *Mtb* persistence.

Although a number of genetic variants of the *SP110* gene have been reported to be associated with susceptibility to human TB, the results of studies regarding the relationship between *SP110* polymorphisms and TB susceptibility are inconsistent [22–29]. A family-based study in West Africa identified 3 *SP110* polymorphisms that are associated with TB susceptibility [22]; however, no significant associations between SP110 and disease susceptibility were identified by other, larger case-control studies conducted on various populations [24–26]. After screening Taiwanese populations for polymorphisms in *SP110*, we identified some singlenucleotide polymorphisms (SNPs) in *SP110* that are significantly associated with susceptibility to LTBI as well as TB disease. These results suggest that the *SP110* variants may provide novel predictive markers for TB infection status and disease outcome.

2. Materials and Methods

2.1. Human Subject Study. The study was conducted in accordance with the terms of the informed consent that was provided to, and received from, participants prior to inclusion in the study. This study was approved by the National Taiwan University Hospital Institutional Review Board (IRB No. 200612009M and IRB No. 201512169RINA). Human blood was ethically collected from patients with cultureconfirmed pulmonary TB and their household contacts as described previously [30]. Briefly, the participating contacts received chest radiography, and mycobacteriology studies were conducted for 3 sputum samples (including acid-fast smear and mycobacterial culture) to exclude the possibility of active TB disease. Because routine BCG vaccination for newborns in Taiwan could affect the accuracy of tuberculin skin test, all enrolled contacts were then tested for LTBI using a T-SPOT. TB assay (Oxford Immunotec Ltd., Abingdon, UK) or QuantiFERON-TB Gold In-Tube assay (QFT) (Qiagen, Hilden, Germany), and the assays were interpreted according to the manufacturers' criteria. Both patients and contacts were excluded if they were tested positive for HIV infection. In total, 301 pulmonary TB patients, 68 individuals with LTBI, and 278 healthy household contacts were included in the study, and genomic DNA was extracted from their peripheral blood (1-2 mL) using a kit from Qiagen according to the manufacturer's protocol. SP110 polymorphisms were identified from the National Center for Biotechnology Information dbSNP database (https://www.ncbi.nlm.nih. gov/snp). The SNPs were genotyped using the MassARRAY System (Sequenom, San Diego, CA, US), and the primer extension products were analyzed by MALDI-TOF mass spectrometry as previously described [31, 32]. Details of the primers that were used are listed in Table S1 in the Supplementary Materials.

Plasma samples were prepared from blood samples by centrifugation and then stored at -80° C until analysis. The TNF α levels in plasma samples were determined by a MAG-PIX[®] platform (Luminex Corp., Austin, TX. US) with a MILLIPLEX MAP Human Cytokine/Chemokine Magnetic Bead Panel I kit (Merck Millipore, Billerica, MA, US) according to the manufacturer's instructions.

2.2. Statistical Analyses. The associations of gene polymorphisms with LTBI and TB disease were analyzed by SAS 9.4 software (SAS Institute, Cary, NC, US) [33]. Linkage disequilibrium and haplotype analyses were performed using Haploview (https://www.broadinstitute.org/haploview) [34]. Chisquare tests were used to compare frequencies. Odds ratios (ORs) were calculated with 95% confidence intervals (CIs). Bonferroni correction or false discovery rate (FDR) correction was applied for multiple comparison adjustments as indicated. The difference of the TNF α levels in plasma between two genotype groups of samples was calculated using a two-tailed unpaired *t*-test with Prism software (GraphPad, San Diego, CA, US). *p* values less than 0.05 were considered to indicate statistical significance, and the number of asterisks represents the degree of significance with regard to the *p* values.

3. Results

3.1. Association of Polymorphisms in SP110 with LTBI and TB Susceptibility. To investigate the association between the SP110 gene and control of Mtb infection, we examined polymorphisms in the gene of members of the Taiwanese population for genetic association with TB disease status. In this study, 301 pulmonary TB patients (202 males and 99 females; mean age: 63.1 ± 19.9 years), 68 individuals with LTBI (35 males and 33 females; mean age: 46.9 ± 17.7 years), and 278 healthy household contacts of the patients (92 males and 186 females; mean age: 47.1 ± 17.3 years) were included (Table 1). In total, 20 SNPs in the SP110 gene were selected for analysis. Of these, 10 SNPs that were not polymorphic or had a minor allele frequency less than 1% were not further included in the analysis, and the remaining 10 SNPs were then analyzed. We found that 3 SNPs (rs7580912, p = 0.0426, OR: 1.52, 95% CI: 1.01–2.39; rs7580900, *p* = 0.008, OR: 1.68, 95% CI: 1.14–2.48; and rs9061, *p* = 0.0026, OR: 0.39, 95% CI: 0.21-0.73) showed differential allele frequency distributions in LTBI cases vs. healthy controls (Table 2). After Bonferroni correction, rs9061 remained significant in this analysis (p < 0.05). Although allele frequencies of none of the 10 SNPs differed significantly in TB patients vs. healthy controls (Table 2), 3 SNPs (rs7580900, *p* = 0.0319, OR: 0.66, 95% CI: 0.45-0.97; rs9061, p = 0.0116, OR: 2.21, 95% CI: 1.18-4.15; and rs2241525, *p* = 0.0309, OR: 0.6, 95% CI: 0.38–0.96) exhibited differential allele frequency distributions in TB cases vs. LTBI individuals (Table 2). All of these SNPs were in accordance with the Hardy-Weinberg equilibrium (HWE).

The associations between SP110 genotypes and susceptibility to LTBI and TB were then analyzed. In LTBI cases vs. healthy controls, we found that genotypes "GG" in rs7580912 (p = 0.025, OR: 2.451, 95% CI: 1.12–5.364) and "GG" in rs7580900 (p = 0.015, OR: 2.584, 95% CI: 1.208– 5.53) were associated with LTBI risk and that genotype "GA" in rs9061 exhibited a protective effect on LTBI (*p* = 0.044, OR: 0.494, 95% CI: 0.239–0.981) (Table 3). We also found that genotypes "GG" in rs7580912 (p = 0.02, OR: 0.392, 95% CI: 0.179-0.86) and "GG" in rs7580900 (p = 0.017, OR: 0.392, 95% CI: 0.182-0.848) in TB patients vs. LTBI cases (Table 3), as well as "CT" in rs11556887 (*p* = 0.039, OR: 0.626, 95% CI: 0.401–0.976) in TB patients vs. healthy controls had a protective effect on TB (Table 3). These results indicated that several SNPs (rs7580900, rs7580912, rs9061, and rs11556887) in SP110 were associated with susceptibility to LTBI and/or TB disease.

3.2. Association Analyses in Various Inheritance Models. The minor allele of each SNP was presumed as a risk factor

TABLE 1: Demographic characteristics of TB patients, LTBI cases, and healthy controls in this study.

Crown		Numbe	Age (years)		
Group	Total	Male (%)	Female (%)	Mean \pm SD	Range
Health	278	92 (33)	186 (67)	47.1 ± 17.3	15.2-93.9
LTBI	68	35 (51)	33 (49)	46.9 ± 17.7	19.5-86.5
TB	301	202 (67)	99 (33)	63.1 ± 19.9	19.4-98.7

compared to the major allele, and the associations between SNPs and susceptibility to LTBI and TB were analyzed in various inheritance models (Table 4). We found that in LTBI cases vs. healthy controls, rs9061 showed a protective effect on LTBI in both additive (*p* = 0.0059, OR: 0.41, 95% CI: 0.22– 0.78) and dominant (p = 0.0112, OR: 0.42, 95% CI: 0.21-0.82) models, while rs7580900 was associated with LTBI risk in both additive (*p* = 0.0147, OR: 1.62, 95% CI: 1.1–2.39) and recessive (*p* = 0.0195, OR: 2.13, 95% CI: 1.13–4.03) models. In addition to this finding, rs7580912 was also associated with LTBI risk in a recessive model (p = 0.0142, OR: 2.49, 95% CI: 1.2-5.18). In TB patients vs. LTBI cases, rs9061 was associated with TB risk in both additive (p = 0.015, OR: 2.17, 95% CI: 1.16–4.06) and dominant (p = 0.0273, OR: 2.14, 95% CI: 1.09-4.22) models, while rs7580900 exhibited a protective effect on TB in both additive (p = 0.0261, OR: 0.64, 95% CI: 0.43-0.95) and recessive (*p* = 0.0117, OR: 0.45, 95% CI: 0.24-0.84) models and rs2241525 was associated with protection from TB in both additive (p = 0.0407, OR: 0.62, 95% CI: 0.4–0.98) and dominant (*p* = 0.0389, OR: 0.55, 95% CI: 0.31-0.97) models. Additionally, rs7580912 had a protective effect on TB in a recessive model (p = 0.0043, OR: 0.35, 95% CI: 0.17-0.72). After false discovery rate (FDR) correction, rs7580900 and rs7580912 remained significant (p = 0.041 and 0.0301, respectively) in a recessive model in this analysis.

3.3. Linkage Disequilibrium and Haplotype Analyses. We then examined linkage disequilibrium (LD) with the SNP markers in the SP110 gene using a Haploview analysis. We found that the "ATATACGCGG" and "ATGTAC GCGA" haplotypes met statistical significance (p = 0.0193, OR: 2.11, 95% CI: 1.11–3.99 and p = 0.0225, OR: 2.76, 95% CI: 1.12-6.84, respectively) for association with LTBI risk in LTBI cases vs. healthy controls (see Figure S1 in the Supplementary Materials) and that the "ATGTAC GCGA" and "ATGAAAGCGA" haplotypes were statistically significant (*p* = 0.03, OR: 0.38, 95% CI: 0.16–0.95 and *p* = 0.0324, OR: 0.24, 95% CI: 0.06–0.99, respectively) with a protective effect on TB in TB cases vs. LTBI individuals (see Figure S2 in the Supplementary Materials). Interestingly, the "GCGTACGCGG" haplotype was associated with TB risk (p = 0.0169, OR: 3.81, 95% CI: 1.18– 12.31) in TB cases vs. healthy controls (see Figure S3 in the Supplementary Materials), although none of the SNPs studied show a significant difference in frequency distributions in this comparison (Table 2). Noteworthily, when we compared "ATGTACGCAA" (the most frequent haplotype) with "ATGTACGCGA" (the haplotype that was

SNP ID	Position ¹	Location ²	HWEp	Alleles	LTBI n (%)	Health n (%)	OR (95% CI)	<i>p</i> value
	220216600	Lature 2.2	0.1112	А	48 (40)	165 (30.4)	1.52 (1.01.2.20)	0.0426
rs/580912	230216690	Intron 2-3	0.1113	G	72 (60)	377 (69.6)	1.52 (1.01-2.39)	0.0426
ma7580000	220216660	Intuon 2.2	0 5446	А	67 (52.3)	214 (39.5)	1 60 (1 14 2 40)	0.000
rs/580900	230216669	Intron 2-3	0.5446	G	61 (47.7)	328 (60.5)	1.68 (1.14-2.48)	0.008
ma11556007	220212061	Europ 4	0.0502	С	113 (88.3)	476 (87.8)	0.06(0.52, 1.74)	0.0060
1811550887	230212901	EXOII 4	0.0595	Т	15 (11.7)	66 (12.2)	0.96 (0.55-1.74)	0.8862
wo0061	61 230212395 Exon 5	Evon 5	0 1272	G	118 (90.8)	427 (79.4)	0.20(0.21, 0.72)	0.0026*
189001		0.1272	А	12 (9.2)	111 (20.6)	0.39 (0.21-0.73)	0.0020	
	Introp 5 6	0.8257	С	93 (71.5)	366 (67.5)	0.83 (0.54, 1.26)	0 3774	
183820974	3820974 230211574 Intron 5-6	11111011 3-0	0.8257	А	37 (28.5)	176 (32.5)	0.83 (0.34-1.20)	0.3774
wo1265776	220207004	Evon 9	0 4241	А	111 (91)	479 (89.7)	0.96 (0.44, 1.70)	0 6709
181303770	230207994	EXOII 8	0.4341	G	11 (9)	55 (10.3)	0.80 (0.44-1.70)	0.0708
rc41300108	230201006	Introp 9 10	0.0333	Т	105 (80.8)	415 (76.6)	0.78 (0.48, 1.26)	0 3030
1841309108	230201000	1111011 9-10	0.0333	А	25 (19.2)	127 (23.4)	0.78 (0.46-1.20)	0.3039
rs22/1525	230178086	Introp 13-14	0.451	G	30 (24.6)	93 (17.4)	1 55 (0 97-2 47)	0.067
182241323	230178080	1111011 15-14	0.431	А	92 (75.4)	441 (82.6)	1.33 (0.97-2.47)	0.007
rc1135701	230177560	Evon 14	0 4826	Т	107 (89.2)	472 (87.4)	0.76(0.41 - 1.43)	0 5952
131133791	230177300	LX0II 14	0.4020	С	13 (10.8)	68 (12.6)	0.70 (0.41-1.45)	0.3932
rc10498244	230173117	Introp 14-15	0 5064	А	95 (89.6)	459 (86.6)	0.75 (0.38-1.47)	0 3972
rs10498244 230173117	2301/311/	/311/ Intron 14-15	0.5064	G	11 (10.4)	71 (13.4)	0./5 (0.38-1.4/)	0.3972

TABLE 2: Allele frequencies of polymorphisms in SP110 in TB patients, LTBI cases, and healthy controls.

(a) Allele frequencies in LTBI cases and healthy controls and odds ratio estimates for LTBI

¹NCBI Reference Sequence: NC_000002.12. ²Based on SP110c (NCBI Reference Sequence: NM_080424.2). HWE: Hardy-Weinberg equilibrium; OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic. *OR remains significant after Bonferroni correction.

SNP ID	Position ¹	Location ²	HWEp	Alleles	LTBI n (%)	Health n (%)	OR (95% CI)	<i>p</i> value
mo7590012	220216600	Introp 2.2	0.0012	А	201 (33.6)	165 (30.4)	1 16 (0.01 1 40)	0.2524
18/380912	230210090	Intron 2-3	0.9815	G	397 (66.4)	377 (69.6)	1.16 (0.91-1.49)	0.2524
ro7580000	220216660	Introp 2 3	0 8338	А	251 (42)	214 (39.5)	1 11 (0 88 1 41)	0 303
18/ 380 900	230210009	IIIti0II 2-3	0.8558	G	347 (58)	328 (60.5)	1.11 (0.88-1.41)	0.393
rc11556997	220212061	Evon 4	0 7206	С	543 (90.8)	476 (87.8)	0.73 (0.50, 1.07)	0 1028
1811330887	230212901	EXOII 4	0.7290	Т	55 (9.2)	66 (12.2)	0.75 (0.50-1.07)	0.1028
rc9061	220212205	Evon 5	0.0162	G	485 (81.6)	427 (79.4)	0.86 (0.64, 1.16)	0 3326
189001	230212393	Ex0II 5	0.0102	А	109 (18.4)	111 (20.6)	0.80 (0.04-1.10)	0.3320
re3820971	230211574	11574 Intron 5-6	0.9602	С	195 (32.6)	17 (32.5)	1.01 (0.79-1.29)	0.9609
183820974	230211374		0.9002	А	403 (67.4)	366 (67.5)	1.01 (0.79-1.29)	0.9009
rc1365776	230207004	Evon 8	0.0396	А	67 (11.2)	55 (10.3)	1 10 (0 75-1 59)	0.6242
131303770	230207994	LX0II 0	0.0570	G	531 (88.8)	479 (89.7)	1.10 (0.75-1.57)	0.0242
rc/1300108	230201006	Introp 9-10	0 0303	Т	463 (77.4)	415 (76.6)	0.95(0.72, 1.26)	0 7314
1341507100	230201000	mitron <i>y</i> -10	0.0595	А	135 (22.6)	127 (23.4)	0.95 (0.72-1.20)	0.7514
re22/11525	230178086	Introp 13-14	0 102	G	495 (83.6)	441 (82.6)	0.93 (0.68-1.27)	0.6447
182241323	230178080	1111011 13-14	0.102	А	97 (16.4)	93 (17.4)	0.95 (0.08-1.27)	0.0447
rc1135701	220177560	Evon 14	0 3517	Т	92 (15.4)	68 (12.6)	1 28 (0.02 1.80)	0.176
181133791	230177300	EXOII 14	0.3317	С	506 (84.6)	472 (87.4)	1.28 (0.92-1.80)	0.170
rc10198211	230173117	Introp 14-15	0 5808	А	88 (14.7)	71 (13.4)	1 13 (0 81-1 59)	0.525
1310470244	2301/311/	1111011 14-15	0.5000	G	510 (85.3)	459 (86.6)	1.15 (0.01-1.59)	0.525

(b) Allele frequencies in TB patients and healthy controls and odds ratio estimates for TB

¹NCBI Reference Sequence: NC_000002.12. ²Based on SP110c (NCBI Reference Sequence: NM_080424.2). HWE: Hardy-Weinberg equilibrium; OR: odds ratio; CI: confidence interval; ORs are adjusted for gender.

SNP ID	Position ¹	Location ²	HWEp	Alleles	LTBI n (%)	Health n (%)	OR (95% CI)	<i>p</i> value
	220216600	Lature 2.2	0.7251	А	397 (66.4)	72 (60)	0.76 (0.51, 1, 1, 4)	0 1707
rs/580912	230216690	Intron 2-3	0./351	G	201 (33.6)	48 (40)	0.76 (0.51-1.14)	0.1/9/
	22021////	Lature 2.2	0.0079	А	347 (58)	61 (47.7)	0 ((0 45 0 07)	0.0210
rs/580900	230216669	Intron 2-3	0.9968	G	251 (42)	67 (52.3)	0.66 (0.45-0.97)	0.0319
	220212071	Energy 4	0.0050	С	543 (90.8)	113 (88.3)	0.76 (0.42, 1.40)	0.3804
rs1155688/	230212961	Exon 4	0.8656	Т	55 (9.2)	15 (11.7)	0.76 (0.42-1.40)	
mc0061	220212205	Erron 5	0.0005	G	109 (18.4)	12 (9.2)	2.21 (1.18-4.15)	0.0116
rs9061	230212395	Exon 5	0.0993	А	485 (81.6)	118 (90.8)		0.0116
wa2920074	3820974 230211574 Intron 5-6	Introp E 6	0.8727	С	195 (32.6)	37 (28.5)	1 22 (0 80 1 85)	0.3577
183820974		0.8/2/	А	403 (67.4)	93 (71.5)	1.22 (0.00-1.05)	0.3377	
wo1265776	220207004	Evon 9	0.028	А	67 (11.2)	11 (9)	1 27 (0 65 2 40)	0 4796
181303770	230207994	EXOII 8	0.028	G	531 (88.8)	111 (91)	1.27 (0.03-2.49)	0.4780
wo41200109	220201006	Introp 0 10	0.2174	Т	135 (22.6)	25 (19.2)	1 22 (0 76 1 07)	0 4020
1841309108	230201000	11111011 9-10	0.2174	А	463 (77.4)	105 (80.8)	1.22 (0.76-1.97)	0.4039
wo2241525	220170006	Introp 12 14	0 1275	G	495 (83.6)	92 (75.4)	0 60 (0 28 0 06)	0.0200
182241323	2301/8080	11111011 13-14	0.1275	А	97 (16.4)	30 (24.6)	0.00 (0.38-0.90)	0.0309
re1135701	230177560	Evon 14	0 3733	Т	92 (15.4)	13 (10.8)	1.50(0.81-2.77)	0 1 9 7 9
181133791	230177300	EXOII 14	0.3733	С	506 (84.6)	107 (89.2)	1.30 (0.81-2.77)	0.1979
re10/082/1	230173117	Introp 14-15	0 8897	А	88 (14.7)	11 (10.4)	1 49 (0 77-2 89)	0 2363
1310470244	2301/311/	1111011 14-13	0.007/	G	510 (85.3)	95 (89.6)	1.47 (0.77-2.09)	0.2303

(c) Allele frequencies in TB patients and LTBI cases and odds ratio estimates for TB

¹NCBI Reference Sequence: NC_000002.12. ²Based on SP110c (NCBI Reference Sequence: NM_080424.2). HWE: Hardy-Weinberg equilibrium; OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic.

associated with LTBI risk in LTBI cases vs. healthy controls and had a protective effect on TB in TB cases vs. LTBI individuals), the latter was found to be significantly affected by the rs7580900 SNP in both comparisons (p = 0.0225, OR: 2.76, 95% CI: 1.12–6.84 for LTBI cases vs. healthy controls; p = 0.03, OR: 0.38, 95% CI: 0.16– 0.95 for TB cases vs. LTBI individuals) (see Figures S1 and S2 in the Supplementary Materials).

We analyzed a block that includes 3 SNPs (rs9061, rs7580900, and rs7580912) in two comparisons (LTBI cases vs. healthy controls and TB cases vs. LTBI individuals) (Figure 1). We found that the "GAA" (p = 0.0037, OR: 1.77, 95% CI: 1.20–2.60), "GGG" (*p* = 0.0009, OR: 2.06, 95% CI: 1.34-3.17), "GGA" (p = 0.0009, OR: 2.81, 95% CI: 1.49-5.27), and "GAG" (*p* = 0.0001, OR: 10, 95% CI: 2.36–42.32) haplotypes were associated with disease risk and that the "AGG" haplotype had a protective effect on LTBI (*p* = 0.0254, OR: 0.41, 95% CI: 0.18–0.92) for LTBI cases vs. healthy controls (Table 5). In addition, the "GAA" (p = 0.0111, OR: 0.61, 95% CI: 0.42-0.90),"GGG" (p = 0.0408, OR: 0.65, 95% CI: 0.43-0.98),"GGA" (p = 0.0001, OR: 0.3, 95% CI: 0.16-0.57), and "GAG"(p = 0.0001, OR: 0.1, 95% CI: 0.02-0.4) haplotypes showed a protective effect on TB disease for TB patients vs. LTBI individuals (Table 5). These genetic studies suggest that the SP110 gene plays a key role in modulating susceptibility to latent and active TB infection.

3.4. Association between the SP110 rs9061 SNP and the TNFa Production in LTBI Subjects. TNF α plays a crucial role in controlling Mtb infection and TB reactivation; however, overproduction of TNF α may cause pathology [35, 36]. Our previous studies demonstrated that the SP110b protein, which is encoded by the SP110 gene and whose expression is upregulated by IFNs, downregulates TNFa production in monocyte/macrophage cells activated by IFNy, thereby alleviating cell death [21]. This finding indicates that the protein functions as a regulator of proinflammatory cytokines of host immunity contributing to a reduction in tissue damage caused by excessive inflammation [21]. To further investigate the potential association between the SP110 SNPs and disease status, we next analyzed clinical parameters in the studied subjects. We measured TNFa levels in plasma from LTBI individuals who carry the different genotypes of the SP110 SNPs and demonstrated that the "GA" genotype of rs9061 in LTBI individuals was associated with lower TNFa levels in plasma compared to "GG" LTBI subjects (Figure 2). The TNF α levels of healthy controls with both genotypes were undetectable (not shown). These data are in agreement with the protective role of the "GA" genotype of rs9061 in LTBI (Table 3) and further support our recent finding showing that the SP110b protein prevents cell death and tissue damage by downregulating TNF α production.

TABLE 3: Association analyses of SP110 SNP genotypes with LTBI and TB susceptibility.

 a) Association between SP110 SN 	genotypes and LTBI risk in LTBI	cases vs. healthy controls
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SNP ID	Genotypes	Health no. (%)	LTBI no. (%)	OR (95% CI)	<i>p</i> value
	AA	134 (49)	26 (43)	Ref	_
rs7580912	GA	111 (41)	20 (33)	0.884 (0.464-1.686)	0.709
	GG	27 (10)	14 (23)	2.451 (1.12-5.364)	0.025
	AA	100 (37)	16 (25)	Ref	_
rs7580900	GA	128 (47)	29 (45)	1.403 (0.716-2.749)	0.324
	GG	43 (16)	19 (30)	2.584 (1.208-5.53)	0.015
	CC	206 (76)	49 (77)	Ref	_
rs11556887	СТ	64 (24)	15 (23)	0.982 (0.513-1.882)	0.957
	TT	1 (0.4)	0 (0)	<0.001 (<0.001-999.999)	0.967
	GG	177 (65)	54 (82)	Ref	_
rs9061	AA	16 (6)	0 (0)	<0.001 (<0.001-999.999)	0.901
	GA	79 (29)	12 (18)	0.494 (0.239-0.981)	0.044
	CC	125 (46)	34 (52)	Ref	_
rs3820974	AA	29 (11)	6 (9)	0.712 (0.269-1.886)	0.494
	CA	118 (43)	25 (38)	0.703 (0.391-1.264)	0.239
	AA	217 (81)	52 (83)	Ref	_
rs1365776	GA	47 (18)	10 (16)	0.757 (0.345-1.66)	0.487
	GG	4 (1)	1 (1)	1.212 (0.129-11.39)	0.867
	TT	153 (56)	41 (63)	Ref	_
rs41309108	AA	10 (4)	1 (1)	0.395 (0.048-3.248)	0.387
	TA	109 (40)	23 (35)	0.768 (0.433-1.362)	0.367
	GG	186 (69)	36 (58)	Ref	_
rs2241525	AA	10 (4)	4 (7)	1.984 (0.579-6.802)	0.276
	GA	75 (28)	22 (35)	1.509 (0.828-2.75)	0.179
	TT	206 (75)	48 (79)	Ref	_
rs1135791	CC	3 (1)	0 (0)	<0.001 (<0.001-999.999)	0.943
	СТ	64	13 (21)	0.86 (0.434-1.703)	0.667
	AA	199 (75)	44 (83)	Ref	_
rs10498244	AG	61 (23)	7 (13)	0.523 (0.223-1.223)	0.135
	GG	5 (2)	2 (4)	1.9 (0.354-10.203)	0.454

OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic.

(b) Association between SP110 SNP genotypes and TB risk in TB patients vs. LTBI cases

SNP ID	Genotypes	Health no. (%)	LTBI No. (%)	OR (95% CI)	<i>p</i> value
	AA	26 (43)	129 (43)	Ref	_
rs7580912	GA	20 (33)	139 (47)	1.377 (0.731-2.595)	0.322
	GG	14 (23)	31 (10)	0.392 (0.179-0.86)	0.02
	AA	16 (25)	98 (33)	Ref	_
rs7580900	GA	29 (45)	151 (50)	0.863 (0.444-1.878)	0.664
	GG	19 (30)	50 (17)	0.392 (0.182-0.848)	0.017
	CC	49 (77)	248 (83)	Ref	_
rs11556887	СТ	15 (23)	47 (16)	0.628 (0.324-1.219)	0.169
	TT	0 (0)	4 (1)	>999.999 (<0.001->999.999)	0.988
	GG	54 (82)	203 (68)	Ref	_
rs9061	AA	0 (0)	15 (5)	>999.999 (<0.001->999.999)	0.974
	GA	12 (18)	79 (27)	1.831 (0.924-3.627)	0.083

Disease Markers

SNP ID	Genotypes	Health no. (%)	LTBI No. (%)	OR (95% CI)	<i>p</i> value			
	CC	34 (52)	136 (45)	Ref	_			
rs3820974	AA	6 (9)	32 (11)	1.261 (0.483-3.293)	0.637			
	CA	25 (38)	131 (44)	1.31 (0.739-2.323)	0.356			
	AA	52 (83)	240 (80)	Ref	_			
rs1365776	GA	10 (16)	51 (17)	1.264 (0.58-2.753)	0.556			
	GG	1 (1)	8 (3)	2.028 (0.244-16.888)	0.513			
	TT	41 (63)	176 (59)	Ref	_			
rs41309108	AA	1 (1)	12 (4)	2.401 (0.3-19.219)	0.409			
	ТА	23 (35)	111 (37)	1.096 (0.621-1.932)	0.753			
	GG	36 (58)	211 (71)	Ref	_			
rs2241525	AA	4 (7)	12 (4)	0.47 (0.142-1.562)	0.218			
	GA	22 (35)	73 (25)	0.564 (0.31-1.026)	0.061			
	TT	48 (79)	212 (71)	Ref	_			
rs1135791	CC	0 (0)	5 (2)	>999.999 (<0.001->999.999)	0.986			
	СТ	13 (21)	82 (27)	1.33 (0.68-2.602)	0.404			
	AA	44 (83)	215 (72)	Ref	_			
rs10498244	AG	7 (13)	80 (27)	2.137 (0.917-4.981)	0.079			
	GG	2 (4)	4 (1)	0.441 (0.074-2.62)	0.368			

TABLE 3: Continued.

OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic.

(c) Association between SP110 SNP genotypes and TB risk in TB patients vs. healthy controls

SNP ID	Genotypes	Health no. (%)	LTBI no. (%)	OR (95% CI)	<i>p</i> value
	AA	134 (49)	129 (43)	Ref	_
rs7580912	GA	111 (41)	139 (47)	1.258 (0.869-1.822)	0.224
	GG	27 (10)	31 (10)	0.877 (0.474-1.622)	0.676
	AA	100 (37)	98 (33)	Ref	_
rs7580900	GA	128 (47)	151 (50)	1.258 (0.851-1.86)	0.249
	GG	43 (16)	50 (17)	1.023 (0.601-1.74)	0.934
	CC	206 (76)	248 (83)	Ref	_
rs11556887	СТ	64 (23.6)	47 (16)	0.626 (0.401-0.976)	0.039
	ΤT	1 (0.4)	4 (1)	2.387 (0.243-23.458)	0.456
	GG	176 (65)	203 (68)	Ref	_
rs9061	AA	17 (6)	15 (5)	0.792 (0.36-1.742)	0.561
	GA	79 (29)	79 (27)	0.886 (0.597-1.313)	0.546
	CC	125 (46)	136 (45)	Ref	_
rs3820974	AA	29 (11)	32 (11)	0.871 (0.473-1.603)	0.657
	CA	118 (43)	131 (44)	0.942 (0.651-1.363)	0.749
	AA	217 (81)	240 (80)	Ref	_
rs1365776	GA	47 (18)	51 (17)	0.931 (0.582-1.489)	0.767
	GG	4 (2)	8 (3)	2.291 (0.631-8.322)	0.208
	ТТ	153 (56)	176 (59)	Ref	_
rs41309108	AA	9 (3)	12 (4)	0.982 (0.382-2.527)	0.971
	ТА	109 (40)	111 (37)	0.852 (0.593-1.225)	0.387
	GG	186 (69)	211 (71)	Ref	_
rs2241525	AA	10 (3)	12 (4)	1.004 (0.402-2.505)	0.994
	GA	75 (28)	73 (25)	0.85 (0.569-1.268)	0.425
ma1125701	TT	206 (75)	212 (71)	Ref	_
181133/91	CC	3 (1)	5 (2)	1.681 (0.371-7.617)	0.5

TABLE 3: Continued.

SNP ID	Genotypes	Health no. (%)	LTBI no. (%)	OR (95% CI)	<i>p</i> value
	СТ	64 (24)	82 (27)	1.177 (0.788-1.76)	0.426
	AA	199 (75)	215 (72)	Ref	_
rs10498244	AG	61 (23)	80 (27)	1.185 (0.787-1.785)	0.417
	GG	5 (2)	4 (1)	0.774 (0.191-3.133)	0.719

OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic.

4. Discussion

SP110 is strongly regulated by IFNs [17], suggesting its possible role in microbial immunity. Although many groups have studied the associations between the gene and TB susceptibility in a variety of populations, these studies show inconclusive results [22-29]. In our study, we recruited healthy household contacts of TB patients as controls, as these contacts are at a high risk of exposure to Mtb. It has been reported that approximately 80-100% of the contacts may have Mtb infection, and on average, 20% of them may develop disease [37], indicating that household contacts are at high risk of LTBI and active TB disease. Therefore, examination of this group carries a considerable importance for prevention and control of TB disease. To the best of our knowledge, this is the first study to demonstrate an association between SP110 and LTBI, and it is the first study of the gene in the Taiwanese population. This work will help clarify the relationship between genetic variation in SP110 with latent and active TB infection in an Asian population.

Several SNPs (rs7580900, rs7580912, rs9061, rs11556887, and rs2241525) in SP110 showed an association with susceptibility to LTBI and/or TB disease in our study. In Table 3, we found that genotypes "GG" in rs7580912 (p = 0.025, OR: 2.451, 95% CI: 1.12-5.364) and "GG" in rs7580900 (*p* = 0.015, OR: 2.584, 95% CI: 1.208–5.53) were associated with LTBI susceptibility in LTBI cases vs. healthy controls, while the same SNP genotypes exhibited a protective effect on LTBI ("GG" in rs7580912, *p* = 0.02, OR: 0.392, 95% CI: 0.179–0.86; "GG" in rs7580900, *p* = 0.017, OR: 0.392, 95% CI: 0.182–0.848) in TB patients vs. LTBI individuals. In the Haploview analysis, we also found that the haplotype of multiple SNPs "ATGTACGCGA" in the SP110 gene was significantly associated with LTBI risk (p = 0.0225, OR: 2.76, 95% CI: 1.12-6.84) in LTBI cases vs. healthy controls, while the same haplotypes had a protective effect on TB disease (p = 0.03, OR: 0.38, 95% CI: 0.16-0.95) in LTBI individuals vs. TB patients (see Figures S1 and S2 in the Supplementary Materials). In addition, 3 SNPs (rs9061, rs7580900, and rs7580912) with the haplotype "GAA" (*p* = 0.0037, OR: 1.77, 95% CI: 1.20–2.60), "GGG" (*p* = 0.0009, OR: 2.06, 95% CI: 1.34–3.17), "GGA" (*p* = 0.0009, OR: 2.81, 95% CI: 1.49–5.27), and "GAG" (p = 0.0001, OR: 10, 95% CI: 2.36-42.32) were associated with disease risk in LTBI cases vs. healthy controls, while the same haplotypes showed a protective effect on TB disease for TB patients vs. LTBI individuals (p = 0.0111, OR: 0.61, 95% CI: 0.42–0.90 for "GAA"; p = 0.0408, OR: 0.65, 95% CI: 0.43–0.98 for "GGG"; p = 0.0001, OR: 0.3, 95% CI: 0.16–0.57 for "GGA"; and p = 0.0001, OR: 0.1, 95% CI: 0.02–0.4 for "GAG") (Table 5). These results revealed that the same SNP genotypes or haplotypes in the *SP110* gene had opposite effects on the control of susceptibility to LTBI and TB disease, suggesting that the gene may have differential roles in the control of susceptibility to LTBI and TB disease.

SNP rs9061 (G \rightarrow A) introduces an amino acid change from glutamic acid to lysine at codon position 207 of the SP110 protein. Transforming an acidic amino acid to a basic amino acid may alter the protein structure or posttranslational modification of the SP110 protein leading to better downregulation of TNF α production. It has been suggested that the A allele may cause changes in α -helices and β -sheets in the secondary structure of the SP110 protein compared with the G allele [38]. In addition, we analyzed the SP110 protein using the ELM database (http://elm.eu.org/) and found that this amino acid change generates a potential binding motif for the C-terminal ubiquitin-like domain (CTD) of ubiquitin specific protease 7 (USP7), one of the most abundant deubiquitinases [39]. USP7 plays important roles in various biological activities, including cell survival, proliferation, apoptosis, and tumorigenesis [40, 41]. As shown in Figure 2, the data demonstrated an association between the "GA" genotype of rs9061 with lower TNF α levels in plasma from LTBI individuals compared to "GG" LTBI subjects. One possible explanation for this result is that SP110 with the "GA" genotype at rs9061 may interact with USP7 and thus is more stable than the SP110 with the "GG" genotype, resulting in a more efficient downregulation of $TNF\alpha$ production. However, further functional studies are needed to elucidate the exact effects of SNP rs9061 on the SP110 gene and disease risk.

Individuals with LTBI, based on a positive result in the TST or the IGRA, usually show no disease symptoms and acquire an effective adaptive immunity. However, a proportion of people with LTBI might reactivate and develop clinical disease. The positive results from these assays indicate an immune response to stimulation by *Mtb* antigens in the LTBI population; however, neither TST nor IGRA can distinguish between LTBI and active TB. In addition, these assays also cannot predict which LTBI cases will progress to active TB [7]. To control and eliminate TB, worldwide TB eradication endeavors have been focused on identifying and treating cases with LTBI. Therefore,

TABLE 4: Association analyses of SP110 SNP genotypes with LTBI and TB susceptibility in various inheritance models.

SND ID	TB vs. hea	TB vs. health		BI	LTBI vs. health	
SNP ID	OR (95% CI)	<i>p</i> value	OR (95% CI)	<i>p</i> value	OR (95% CI)	<i>p</i> value
rs9061	0.89 (0.66,1.2)	0.4382	2.17 (1.16,4.06)	0.015	0.41 (0.22,0.78)	0.0059
rs7580900	1.07 (0.83,1.38)	0.5839	0.64 (0.43,0.95)	0.0261	1.62 (1.1,2.39)	0.0147
rs3820974	0.94 (0.72,1.23)	0.6661	1.2 (0.79,1.82)	0.4009	0.79 (0.51,1.21)	0.2761
rs41309108	0.9 (0.66,1.22)	0.4917	1.2 (0.73,1.97)	0.4824	0.73 (0.44,1.23)	0.2366
rs7580912	1.05 (0.8,1.37)	0.7307	0.74 (0.49,1.1)	0.1387	1.4 (0.94,2.08)	0.0992
rs1135791	1.2 (0.83,1.73)	0.3242	1.46 (0.77,2.77)	0.2444	0.81 (0.42,1.56)	0.525
rs1365776	1.1 (0.75,1.62)	0.6153	1.32 (0.7,2.5)	0.3933	0.84 (0.43,1.65)	0.6133
rs10498244	1.1 (0.77,1.59)	0.5919	1.4 (0.71,2.73)	0.3303	0.77 (0.4,1.5)	0.4457
rs2241525	0.91 (0.66,1.25)	0.5681	0.62 (0.4,0.98)	0.0407	1.46 (0.92,2.32)	0.1102
rs11556887	0.72 (0.48,1.08)	0.1155	0.75 (0.41,1.39)	0.362	0.96 (0.5,1.81)	0.8916

(a) Association analyses of SP110 SNP genotypes in an additive model

OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic.

(b) Association analyses of SP110 SNP genotypes in a dominant model

	TB vs. health		TB vs. LTBI		LTBI vs. health	
SNP ID	OR (95% CI)	<i>p</i> value	OR (95% CI)	<i>p</i> value	OR (95% CI)	<i>p</i> value
rs9061	0.87 (0.6,1.26)	0.4646	2.14 (1.09,4.22)	0.0273	0.42 (0.21,0.82)	0.0112
rs7580900	1.22 (0.84,1.76)	0.2929	0.68 (0.37,1.27)	0.2276	1.74 (0.93,3.24)	0.083
rs3820974	0.93 (0.65,1.32)	0.6879	1.3 (0.76,2.24)	0.3378	0.71 (0.41,1.23)	0.2193
rs41309108	0.86 (0.6,1.23)	0.4092	1.15 (0.66,2.01)	0.6191	0.74 (0.42,1.3)	0.2917
rs7580912	1.18 (0.83,1.68)	0.3571	0.98 (0.55,1.72)	0.9312	1.2 (0.68,2.12)	0.5388
rs1135791	1.2 (0.81,1.78)	0.3678	1.41 (0.72,2.75)	0.3146	0.83 (0.42,1.64)	0.5948
rs1365776	1.03 (0.66,1.6)	0.9033	1.34 (0.64,2.82)	0.4408	0.79 (0.37,1.67)	0.5353
rs10498244	1.15 (0.77,1.72)	0.4807	1.75 (0.81,3.78)	0.1536	0.63 (0.29,1.35)	0.2341
rs2241525	0.87 (0.59,1.27)	0.4667	0.55 (0.31,0.97)	0.0389	1.57 (0.88,2.78)	0.1239
rs11556887	0.66 (0.43,1.02)	0.0609	0.67 (0.35,1.3)	0.2363	0.97 (0.51,1.86)	0.9282

OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic.

(c)	Association	analyses	of SP110	SNP	genotypes	in a	recessive	model
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SNP ID	TB vs. health		TB vs. LTBI		LTBI vs. health	
	OR (95% CI)	<i>p</i> value	OR (95% CI)	<i>p</i> value	OR (95% CI)	<i>p</i> value
rs9061	0.82 (0.38,1.78)	0.6227	_		_	_
rs7580900	0.92 (0.57,1.48)	0.737	0.45 (0.24,0.84)	0.0117	2.13 (1.13,4.03)	0.0195*
rs3820974	0.92 (0.52,1.62)	0.7796	1.13 (0.45,2.83)	0.8018	0.84 (0.33,2.15)	0.7209
rs41309108	1.05 (0.41,2.66)	0.9241	2.35 (0.3,18.52)	0.4186	0.43 (0.05,3.55)	0.437
rs7580912	0.79 (0.44,1.42)	0.4367	0.35 (0.17,0.72)	0.0043	2.49 (1.2,5.18)	0.0142*
rs1135791	1.63 (0.36,7.51)	0.5284	_	_	_	_
rs1365776	2.25 (0.63,8.05)	0.2129	1.94 (0.23,16.11)	0.5375	1.24 (0.13,11.51)	0.8504
rs10498244	0.74 (0.18,3.05)	0.6804	0.38 (0.06,2.23)	0.2844	2.27 (0.42,12.19)	0.3385
rs2241525	1.05 (0.42,2.6)	0.9172	0.56 (0.17,1.83)	0.3374	1.73 (0.52,5.81)	0.3747
rs11556887	2.63 (0.27,25.7)	0.4064	_	_	_	_

OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic. *ORs remain significant after false discovery rate (FDR) correction.

our studies identified several SNPs in *SP110* that are significantly associated with controlling susceptibility to LTBI and TB disease and thus may provide novel predictive markers for latent and active TB infection. This study may also yield an improved strategy that can identify persons at increased risk of the disease.

There were some limitations in our study. First, this study dealt with relatively small sample sizes (301, 68,



FIGURE 1: Haplotype block maps for *SP110* with 3 SNPs (rs9061, rs7580900, and rs7580912). The haplotype blocks were analyzed in LTBI cases vs. healthy controls (a) and TB cases vs. LTBI individuals (b), respectively.

TABLE 5: Association of the haplotype frequencies of *SP110* SNPs with 3 SNPs (rs9061, rs7580900, and rs7580912) with LTBI and TB susceptibility.

Hanlatumaa	Frequencies		Chi aguana	OP(050/CI)	6 m.
napiotypes	LTBI	Health	Chi-square	OR (95% CI)	<i>p</i> value
GAA	0.417	0.558	8.417	1.77 (1.20-2.60)	0.0037
GGG	0.310	0.179	11.048	2.06 (1.34-3.17)	<i>9.00E – 04</i>
AGG	0.053	0.120	4.997	0.41 (0.18-0.92)	0.0254
GGA	0.134	0.052	10.99	2.81 (1.49-5.27)	9.00E - 04
AGA	0.021	0.043	1.429	0.47 (0.13-1.68)	0.232
AAA	0.019	0.042	1.579	0.44 (0.12-1.65)	0.209
GAG	0.047	0.005	14.5	10.00 (2.36-42.32)	1.00E - 04

(a) Association of haplotype frequencies with LTBI risk in LTBI cases and healthy controls

OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic.

(b) Association of haplotype frequencies with TB risk in TB cases vs. LTBI individuals

Hanlatumaa	Frequencies		Chianna		t
napiotypes	ТВ	LTBI	Chi-square	OR (95% CI)	<i>p</i> value
GAA	0.540	0.417	6.445	0.61 (0.42-0.90)	0.0111
GGG	0.228	0.313	4.184	0.65 (0.43-0.98)	0.0408
AGG	0.103	0.053	3.191	2.05 (0.91-4.62)	0.074
GGA	0.043	0.130	14.591	0.30 (0.16-0.57)	0.0001
AGA	0.045	0.021	1.564	2.16 (0.62-7.49)	0.211
AAA	0.035	0.018	0.993	1.95 (0.51-7.53)	0.3191
GAG	0.005	0.047	16.031	0.10 (0.02-0.40)	0.0001

OR: odds ratio; CI: confidence interval; ORs are adjusted for gender. The significant ORs are shown in italic.

and 278 for TB patients, LTBI cases, and healthy household contacts, respectively). Further large-scale studies are, therefore, needed to validate the predictive value of the *SP110* SNPs identified in this study. Second, the LTBI number of our study group is small, and besides, the results on plasma TNF α levels were analyzed from groups with even smaller sample sizes; therefore, the difference in TNF α levels in plasma of LTBI cases with two genotypes may be underestimated and should be verified in a different population. In addition, the analysis of TNF α levels in plasma of active TB patients with the different genotypes is in progress. The result may help support our finding in LTBI individuals and clarify the potential association between the *SP110* SNPs and disease status.

5. Conclusions

The results suggest that the *SP110* variants have a role in controlling genetic susceptibility to latent and active TB infection in humans. To the best of our knowledge,



FIGURE 2: Association between the *SP110* rs9061 SNP and the TNF levels in plasma of LTBI subjects. The TNF production was determined in plasma samples from LTBI cases carrying the different genotypes of the rs9061 SNP by a MAGPIX instrument with a MILLIPLEX MAP Human Cytokine/Chemokine Magnetic Bead Panel I kit. Sample numbers are 14 for "GG" and 8 for "GA" genotypes, respectively. Statistical significance was calculated using a two-tailed unpaired *t*-test. *p < 0.05.

this is the first report demonstrating associations of polymorphisms in *SP110* with LTBI susceptibility. Additionally, we provide evidence that the *SP110* rs9061 SNP is associated with plasma $\text{TNF}\alpha$ levels in LTBI individuals. These data suggest that the identified *SP110* SNPs may serve as a biomarker for latent and active TB infection in Taiwan.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare no conflict of interest.

Acknowledgments

This research was funded by the Ministry of Science and Technology, Taiwan (NSC 97-2320-B-010-006-MY3, NSC 99-2321-B-002-022-, NSC 100-2321-B-002-009-, NSC 100-2320-B-002-106-, NSC 101-2321-B-002-001-, NSC 101-2320-B-002-022-, NSC 103-2320-B-002-044-MY3, and 106-2320-B-002-020- to BSY; NSC 99-2320-B-002-078-MY3, NSC 103-2314-B-002-083-, and 104-2314-B-002-075- to MLC), and Institute for Biotechnology and Medicine Industry, Taiwan. We thank Hsiang-Ting Hsu and Bo-Wen Chen for technical assistance. We also thank the expert assistance provided by Microarray Core Facility of National Research Program for Genomic Medicine of National Science Council in Taiwan.

Supplementary Materials

Table S1: primers for genotype analysis for SP110. Figure S1: association of SP110 SNP haplotypes with LTBI risk in LTBI cases vs. healthy controls. (a) Haplotype block map for SP110 with 10 SNPs. (b) Association of haplotype frequencies with LTBI risk in LTBI cases and healthy controls. Figure S2: association of SP110 SNP haplotypes with TB risk in TB cases vs. LTBI individuals. (a) Haplotype block map for SP110 with 10 SNPs. (b) Association of haplotype frequencies with TB risk in TB cases vs. LTBI individuals. Figure S3: association of SP110 SNP haplotypes with TB risk in TB cases vs. healthy controls. (a) Haplotype block map for SP110 with 10 SNPs. (b) Association of haplotype frequencies with TB risk in TB cases and healthy controls. (*Supplementary Materials*)

References

- [1] WHO, Tuberculosis Fact Sheet N°104, WHO, 2018.
- [2] R. M. G. J. Houben and P. J. Dodd, "The global burden of latent tuberculosis infection: a re-estimation using mathematical modelling," *PLoS Medicine*, vol. 13, no. 10, 2016.
- [3] B. R. Bloom and C. J. L. Murray, "Tuberculosis-commentary on a reemergent killer," *Science*, vol. 257, no. 5073, pp. 1055– 1064, 1992.
- [4] C. Dye, S. Scheele, P. >Dolin, V. Pathania, M. C. Raviglione, and for the WHO Global Surveillance and Monitoring Project, "Global burden of tuberculosis - estimated incidence, prevalence, and mortality by country," *JAMA*, vol. 282, no. 7, pp. 677–686, 1999.
- [5] T. R. Frieden, T. R. Sterling, S. S. Munsiff, C. J. Watt, and C. Dye, "Tuberculosis," *Lancet*, vol. 362, no. 9387, pp. 887– 899, 2003.
- [6] M. Pai, L. W. Riley, and J. M. Colford Jr, "Interferon-γ assays in the immunodiagnosis of tuberculosis: a systematic review," *Lancet Infectious Diseases*, vol. 4, no. 12, pp. 761–776, 2004.
- [7] C. E. Barry, H. I. Boshoff, V. Dartois et al., "The spectrum of latent tuberculosis: rethinking the biology and intervention strategies," *Nature Reviews Microbiology*, vol. 7, no. 12, pp. 845–855, 2009.
- [8] D. N. Rose, "Benefits of screening for latent Mycobacterium tuberculosis infection," *Archives of Internal Medicine*, vol. 160, no. 10, pp. 1513–1521, 2000.
- [9] A. Apt and I. Kramnik, "Man and mouse TB: contradictions and solutions," *Tuberculosis*, vol. 89, no. 3, pp. 195–198, 2009.
- [10] J. L. Casanova and L. Abel, "Genetic dissection of immunity to mycobacteria: the human model," *Annual Review of Immunol*ogy, vol. 20, no. 1, pp. 581–620, 2002.
- [11] A. Fortin, L. Abel, J. L. Casanova, and P. Gros, "Host genetics of mycobacterial diseases in mice and men: forward genetic studies of BCG-osis and tuberculosis," *Annual Review of Genomics and Human Genetics*, vol. 8, no. 1, pp. 163–192, 2007.
- [12] A. V. S. Hill, "The genomics and genetics of human infectious disease susceptibility," *Annual Review of Genomics and Human Genetics*, vol. 2, no. 1, pp. 373–400, 2001.
- [13] C. J. Lynch, C. H. Pierce-Chase, and R. Dubos, "A genetic study of susceptibility to experimental tuberculosis in mice infected with mammalian tubercle bacilli," *The Journal of Experimental Medicine*, vol. 121, no. 6, pp. 1051–1070, 1965.
- [14] I. Kramnik, W. F. Dietrich, P. Demant, and B. R. Bloom, "Genetic control of resistance to experimental infection with

virulent Mycobacterium tuberculosis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 97, no. 15, pp. 8560–8565, 2000.

- [15] H. Pan, B. S. Yan, M. Rojas et al., "Ipr1 gene mediates innate immunity to tuberculosis," *Nature*, vol. 434, no. 7034, pp. 767–772, 2005.
- [16] I. Kramnik, "Genetic dissection of host resistance to Mycobacterium tuberculosis: the sst1 locus and the Ipr1 gene," *Current Topics in Microbiology and Immunology*, vol. 321, pp. 123– 148, 2008.
- [17] S. Kadereit, D. R. Gewert, J. Galabru, A. G. Hovanessian, and E. F. Meurs, "Molecular cloning of two new interferoninduced, highly related nuclear phosphoproteins," *Journal of Biological Chemistry*, vol. 268, no. 32, pp. 24432–24441, 1993.
- [18] S. T. Cliffe, M. Wong, P. J. Taylor et al., "The first prenatal diagnosis for veno-occlusive disease and immunodeficiency syndrome, an autosomal recessive condition associated with mutations in SP110," *Prenatal Diagnosis*, vol. 27, no. 7, pp. 674–676, 2007.
- [19] T. Roscioli, S. T. Cliffe, D. B. Bloch et al., "Mutations in the gene encoding the PML nuclear body protein Sp110 are associated with immunodeficiency and hepatic veno-occlusive disease," *Nature Genetics*, vol. 38, no. 6, pp. 620–622, 2006.
- [20] S. T. Cliffe, D. B. Bloch, S. Suryani et al., "Clinical, molecular, and cellular immunologic findings in patients with SP110associated veno-occlusive disease with immunodeficiency syndrome," *Journal of Allergy and Clinical Immunology*, vol. 130, no. 3, pp. 735–742.e6, 2012.
- [21] J. S. Leu, M. L. Chen, S. Y. Chang et al., "SP110b controls host immunity and susceptibility to tuberculosis," *American Journal of Respiratory and Critical Care Medicine*, vol. 195, no. 3, pp. 369–382, 2017.
- [22] K. Tosh, S. J. Campbell, K. Fielding et al., "Variants in the SP110 gene are associated with genetic susceptibility to tuberculosis in West Africa," *Proceedings of the National Academy* of Sciences of the United States of America, vol. 103, no. 27, pp. 10364–10368, 2006.
- [23] G. J. Fox, D. N. Sy, N. V. Nhung et al., "Polymorphisms of SP110 are associated with both pulmonary and extrapulmonary tuberculosis among the Vietnamese," *PLoS One*, vol. 9, no. 7, article e99496, 2014.
- [24] T. Thye, E. N. Browne, M. A. Chinbuah et al., "No associations of human pulmonary tuberculosis with Sp110 variants," *Journal of Medical Genetics*, vol. 43, no. 7, p. e32, 2006.
- [25] C. Babb, E. H. Keet, P. D. van Helden, and E. G. Hoal, "SP110 polymorphisms are not associated with pulmonary tuberculosis in a South African population," *Human Genetics*, vol. 121, no. 3-4, pp. 521-522, 2007.
- [26] J. S. Szeszko, B. Healy, H. Stevens et al., "Resequencing and association analysis of the SP110 gene in adult pulmonary tuberculosis," *Human Genetics*, vol. 121, no. 2, pp. 155–160, 2007.
- [27] M. Moller and E. G. Hoal, "Current findings, challenges and novel approaches in human genetic susceptibility to tuberculosis," *Tuberculosis*, vol. 90, no. 2, pp. 71–83, 2010.
- [28] E. Png, B. Alisjahbana, E. Sahiratmadja et al., "Polymorphisms in SP110 are not associated with pulmonary tuberculosis in Indonesians," *Infection, Genetics and Evolution*, vol. 12, no. 6, pp. 1319–1323, 2012.
- [29] X. Lei, H. Zhu, L. Zha, and Y. Wang, "SP110 gene polymorphisms and tuberculosis susceptibility: a systematic review

and meta-analysis based on 10 624 subjects," *Infection, Genetics and Evolution*, vol. 12, no. 7, pp. 1473–1480, 2012.

- [30] J. Y. Wang, C. C. Shu, C. H. Lee, C. J. Yu, L. N. Lee, and P. C. Yang, "Interferon-gamma release assay and rifampicin therapy for household contacts of tuberculosis," *The Journal of Infection*, vol. 64, no. 3, pp. 291–298, 2012.
- [31] C. Jurinke, D. van den Boom, C. R. Cantor, and H. Köster, "The use of MassARRAY technology for high throughput genotyping," *Advances in Biochemical Engineering/Biotechnol*ogy, vol. 77, pp. 57–74, 2002.
- [32] C. Jurinke, D. van den Boom, C. R. Cantor, and H. Köster, "Automated genotyping using the DNA MassArray technology," *Methods in Molecular Biology*, vol. 187, pp. 179–192, 2002.
- [33] C. C. Tung, J. M. Wong, W. C. Lee et al., "Combining TNFSF15 and ASCA IgA can be used as a predictor for the stenosis/perforating phenotype of Crohn's disease," *Journal of Gastroenterology and Hepatology*, vol. 29, no. 4, pp. 723–729, 2014.
- [34] J. C. Barrett, B. Fry, J. Maller, and M. J. Daly, "Haploview: analysis and visualization of LD and haplotype maps," *Bioinformatics*, vol. 21, no. 2, pp. 263–265, 2005.
- [35] L. G. Bekker, A. L. Moreira, A. Bergtold, S. Freeman, B. Ryffel, and G. Kaplan, "Immunopathologic effects of tumor necrosis factor alpha in murine mycobacterial infection are dose dependent," *Infection and Immunity*, vol. 68, no. 12, pp. 6954–6961, 2000.
- [36] S. Ehlers, J. Benini, H. D. Held, C. Roeck, G. Alber, and S. Uhlig, " $\alpha\beta$ T cell receptor-positive cells and interferongamma, but not inducible nitric oxide synthase, are critical for granuloma necrosis in a mouse model of mycobacteriainduced pulmonary immunopathology," *The Journal of Experimental Medicine*, vol. 194, no. 12, pp. 1847–1859, 2001.
- [37] "Contact investigations for Tuberculosis," in Self-Study Modules on Tuberculosis, U.S. Department of Health and Human Services, CDC, Atlanta, GA, USA, 1999.
- [38] S. Zhang, X. B. Wang, Y. D. Han, C. Wang, Y. Zhou, and F. Zheng, "Certain polymorphisms in SP110 gene confer susceptibility to tuberculosis: a comprehensive review and updated meta-analysis," *Yonsei Medical Journal*, vol. 58, no. 1, pp. 165–173, 2017.
- [39] R. D. Everett, M. Meredith, A. Orr, A. Cross, M. Kathoria, and J. Parkinson, "A novel ubiquitin-specific protease is dynamically associated with the PML nuclear domain and binds to a herpesvirus regulatory protein," *EMBO Journal*, vol. 16, no. 7, pp. 1519–1530, 1997.
- [40] R. Q. Kim and T. K. Sixma, "Regulation of USP7: a high incidence of E3 complexes," *Journal of Molecular Biology*, vol. 429, no. 22, pp. 3395–3408, 2017.
- [41] B. Nicholson and K. G. Suresh Kumar, "The multifaceted roles of USP7: new therapeutic opportunities," *Cell Biochemistry* and Biophysics, vol. 60, no. 1-2, pp. 61–68, 2011.