

# Lung function tests to monitor respiratory disease in preschool children

*Valentina Fainardi<sup>1</sup>, Enrico Lombardi<sup>2</sup>*

<sup>1</sup> Department of Medicine and Surgery, University of Parma, Parma, Italy; <sup>2</sup> Paediatric Pulmonary Unit, "Anna Meyer" Paediatric University Hospital, Florence, Italy

**Summary.** Pulmonary function tests are routinely used in the diagnosis and follow-up of respiratory diseases. In preschool children assessment and evaluation of lung function has always been challenging but improved techniques that require only minimal collaboration allowed obtaining reliable and useful results even in this group of patients. In this review we will describe the different techniques used in clinical practice to measure lung function in preschool children. ([www.actabiomedica.it](http://www.actabiomedica.it))

**Key words:** preschool, lung function, interrupter, forced oscillations, airway resistance, multiple breath wash-out

## Introduction

Pulmonary function testing plays a key role in the diagnosis and follow-up of respiratory disease (1, 2). However, performing the tests and obtaining objective results in preschool children (i.e. 2-5 years) has always been very challenging due to the poor cooperation in this age range. Ongoing research in this field has allowed to improve the techniques and obtain reliable and useful results even in this group of patients.

In infants lung volumes can be measured by plethysmography (that can measure also airway resistance) or multiple-breath inert gas washout (MBW) with the infant sleeping in a supine position with or without sedation. Other tests are also used to assess forced expiratory flow-volume loops and respiratory mechanics in sedated infants. However, all these techniques are difficult to use in routine clinical practice and are performed in a few specialised centres.

For older children (>2-3 years), who can provide a minimal collaboration, lung volumes and forced expiratory flows can be assessed by means of spirometry using specific criteria of acceptability. Plethysmogra-

phy, interrupter technique (Rint) and forced oscillation technique (FOT) can be performed to measure respiratory resistance and reactance. Published international guidelines and reference values can now facilitate the clinical use of some of these tests.

This review aims to describe the different techniques that can be used in clinical practice to measure lung function in preschool children. Plethysmography in this age is less standardised and problematic to use routinely and it will not be included in this review.

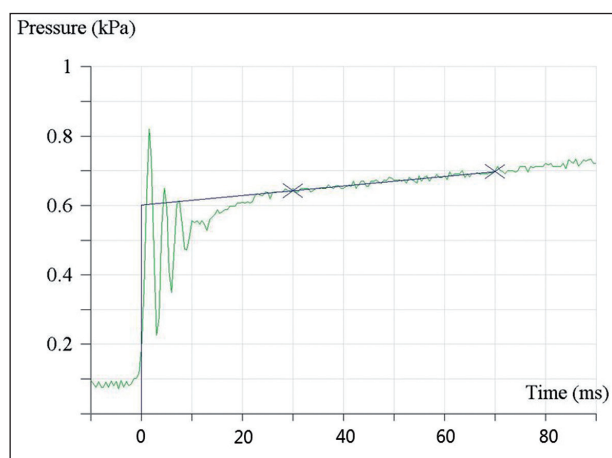
## Interrupter technique

The interrupter technique measures respiratory resistance (Rint) (including lung, airways and rib cage) during tidal breathing. Therefore, it is a quick and non-invasive test that can be used in preschoolers not enough collaborative to perform spirometry. The child will be sitting, wearing a nose clip, with the head in a neutral position and the cheeks supported by the hands of an operator. During tidal breathing through a mouthpiece and bacterial filter, a valve closing in less

than 10 ms will interrupt the flow in correspondence of the peak expiratory flow for about 100 ms. In this fashion the pressure at the mouth quickly equilibrates with pressure in the alveoli, giving an estimate of the pressure in the airways. Respiratory resistance is then calculated as the ratio between the change in mouth pressure and the flow measured immediately before (“classical” technique) or after (“opening” technique) the interruption.

Because of the viscoelastic properties of the respiratory system, when pressure is measured at the beginning of the interruption Rint will tend to measure pure airway resistance, when pressure is measured at the end of the interruption Rint will approach the resistance of the whole respiratory system including lung tissue and rib cage. Several methods of measuring mouth pressure have been proposed: in the “classical” technique, mouth pressure is back-extrapolated to 0 ms from 30 and 70 ms after the interruption (Fig. 1); in the “opening” technique, mouth pressure is measured at the end of the interruption. Usually, ten measurements are recorded to obtain at least five acceptable measurements of which the median is reported (3).

In preschool children the interrupter technique is highly feasible with up to 98% of subjects able to perform the test (4). Reference equations for the Italian population for the classic technique have been published (4) and international reference values are also available (5).



**Figure 1.** Rint, classical technique: mouth pressure is back-extrapolated to 0 ms from 30 and 70 ms after the interruption

Several reports have been published on Rint in preschool children with wheezing. When used to compare children with different wheezing phenotypes, Rint was found to be higher in children with persistent wheeze compared to never wheezers and children with transient symptoms (6, 7). However, Rint measurements in young children failed to predict the development of asthma at school age (8, 9). As for spirometry, also for Rint assessing bronchodilator response (BDR) can be very useful in daily clinical practice. In one of the studies on BDR in preschool children, a decrease in Rint  $\geq 0.26$  kPa·L<sup>-1</sup>·s ( $\geq 1.25$  if expressed in Z-scores) after bronchodilation could discriminate children with respiratory symptoms at the time of the test with a sensitivity of 80% and a specificity of 82% (10). Hence, this cut-off could be appropriate for assessing BDR in preschool wheezing children. Other data from the literature show that BDR measured with Rint can distinguish children with wheeze from healthy children with a sensitivity that varies from 24% to 76% and a specificity between 70% and 92% (11).

In preschool children with cystic fibrosis (CF) Rint was assessed in a few studies. In a case-control study involving children aged 3–5 years Rint distinguished patients with asthma or CF from healthy controls showing higher resistance in the case population, but these values did not differ between the two diseases (12). Greater Rint values in preschool children with CF were also reported by Beydon et al. who demonstrated the highest resistance in the subjects exposed to passive smoke (13). When used in longitudinal studies, Rint could not reflect the progression of the disease and the worsening seen at the chest X rays did not correlate with changes of airway resistance over a 3 year period (14).

Only two studies used Rint in preschool children born preterm and compared lung function in children with and without bronchopulmonary dysplasia (BPD) (15, 16). Children diagnosed with BPD had stiffer airways as demonstrated by higher values of resistance (15, 16). In addition, prematurity alone was associated with higher airway resistance compared to reference values (16).

In summary, Rint can be a feasible and useful technique to assess airway resistance in preschool children with asthma, CF and BPD.

### Forced oscillation technique

Similarly to Rint, the forced oscillation technique (FOT) is a non-invasive technique performed during tidal breathing that requires only minimal cooperation from the patient. Small pressure oscillations at frequencies between 4 and 48 Hz are applied to the airways and the impedance of the respiratory system ( $Z_{rs}$ ) is calculated from the resulting changes of mouth pressure and flow (3).  $Z_{rs}$  comprises respiratory resistance ( $R_{rs}$ ) and respiratory reactance ( $X_{rs}$ ) (17).  $R_{rs}$  includes airway, lung tissue and chest wall resistance and represents the frictional pressure loss in the airway, while  $X_{rs}$  represents the balance of respiratory elastance (1/compliance) and inertance. Elastic forces give a negative  $X_{rs}$  and are predominant at low frequencies, while inertial forces give a positive  $X_{rs}$  and are predominant at high frequencies; the frequency at which elastic and inertial forces equal each other (resulting in 0  $X_{rs}$ ) is called resonant frequency ( $F_{res}$ ).  $AX$  is the total reactance (area under the curve) at all frequencies between 4-5 Hz and  $F_{res}$  and reflects  $X_{rs}$  at low frequencies and thus the elastance of the respiratory system.

The pressure oscillation applied to the mouth (forcing signal) can be a sinusoidal wave or a series of impulses (IOS), and both can be used at a single frequency or as multiple-frequency composite signals. Since low frequencies (4-10 Hz) can reach the peripheral lung while high frequencies (18-22 Hz) can be transmitted only into the central airways (18), the lowest frequencies represent an estimate of the whole respiratory system and the highest frequencies an estimate of the upper airways. The difference between  $R_{rs}$  at 5 Hz and  $R_{rs}$  at 20 Hz ( $R_{5-20}$ ) has been used to express the resistance of the peripheral airways (19). However, due to shunt and serial heterogeneity (20), the difference between  $R_{rs}$  at low and high frequencies can be an estimate of the resistance of any level of the airways. Also, the analysis can be performed in the frequency domain (spectral analysis), giving the mean  $Z_{rs}$  over the whole recorded breathing period, or in the time domain (within-breath analysis), giving the mean inspiratory and expiratory  $Z_{rs}$  for each breath (20) or even the end-inspiratory and end-expiratory  $Z_{rs}$  for each breath (21).

To obtain a valid manoeuvre the subject has to breath tidally into a mouthpiece and anti-bacterial fil-

ter for at least 8-16 seconds, wearing a nose clip and with the cheeks supported by the hands of an operator (3) (Fig. 2). At least 3 reproducible manoeuvres without artefacts due to coughing, swallowing, vocalization or breath holding have to be obtained to consider valid the test. The mean value of each index are then calculated (3, 11).

A certain number of clinical studies have established reference values for FOT in young children and, as for spirometry, the standing height appears to be the best and only independent variable for the regression equations (3, 11, 22-31).

Many studies have measured lung function with FOT in young wheezy children with conflicting results: some studies reported abnormal lung function (32, 33), while others showed no difference from controls (34, 35). However, a novel method using the within-breath analysis detected airway obstruction with a sensitivity of 92% and a specificity of 89% (21). Cut-offs for BDR have also been suggested: -32% for  $R_{rs8}$ , +65% for  $X_{rs8}$  and -82% for  $AX$  (30). In young children with intermittent asthma IOS predicted the probability of acute exacerbations better than  $FEV_1$  and methacholine challenge (36), with  $R_5$  showing a sensitivity of 68% and a specificity of 83% and  $R_{5-20}$  a sensitivity of 90% and a specificity of 57%. Impaired  $R_{10}$  in children with asthma was reported in a cohort of 3-6 year-old Asian children (37) and lower z-scores of reactance  $X_{rs5}$  distinguished between intermittent and persistent asthma in 162 subjects aged 2-5 years



**Figure 2.** Forced oscillation technique in a preschool child (permission obtained by parents to reproduce this picture)

(38). In 157 asthmatic children R5 negatively correlated with pre and post-bronchodilator FEV<sub>1</sub>, FEV<sub>1</sub>/FVC and MEF50 and in a multivariate analysis increased Rrs5 was associated with decreased post-bronchodilator FEV<sub>1</sub> and FEV<sub>1</sub>/FVC with a specificity of 85% and 86% respectively (39). BDR could add important information in preschool children who wheezed and could distinguish asthmatic children from controls (35, 37, 40, 41). These findings suggest that wheezing can be associated with airways with higher resistance and lower reactance even in early age. It can be speculated that these impairments may persist in the subjects at risk of persistent asthma.

Although with a pulmonary function within the normal range, preschool children with CF showed higher Rrs and lower Xrs than reference values (42). These parameters worsened till abnormal if the child was symptomatic suggesting that measurements outside the normal range may reveal a clinical deterioration (42). However, the technique failed to distinguish CF subjects from controls (43) and multiple measurements over one year did not correlate with the worsening of the disease described as airway inflammation, pathogens in the BAL and structural changes at the CT (44). When IOS was used in a group of patients with CF including children aged >3 years, R, Fres and AX values increased during exacerbation and decreased after treatment, while X (10-15 Hz) values decreased during exacerbation and increased after recovery (45). However, when compared with spirometry, IOS was not as sensitive as spirometry to detect and follow in the long-term lung function deterioration (46).

Only a few studies measured Rrs and Xrs in young children born preterm, showing less compliant airways as demonstrated by abnormal values of resistance and reactance (16, 47, 48) that were particularly altered in those with BPD (16, 27). Furthermore, airway resistance correlated with oxygen therapy duration (47). Higher resistance R5 and R10, and also lower reactance X5 in those exposed to passive smoke, were found in preschool children born late preterm (49).

In summary, FOT seems to be more useful for showing differences between groups of patients rather than for following a disease (11). There is a large variability of Zrs in healthy preschool-age children and several reference equations have been published (3, 11).

## Preschool spirometry

Spirometry is the gold standard technique to measure lung function in children aged  $\geq 6$  years and adults. In preschool age the forced manoeuvre requires good collaboration and coordination to sustain the effort throughout the expiration and obtaining reliable results can be difficult and time-consuming. That said, several studies report successful spirometry measurements in preschool children (50-52). Criteria for acceptability in this age group are at least two good flow-volume curves with a rapid rise to peak flow and a smooth descending limb with no evidence of cough or glottic closure (3). In this age range a total expiration time of 0.5 (FEV<sub>0.5</sub>) or 0.75 (FEV<sub>0.75</sub>) seconds can be accepted (3), because only 41-75% of children younger than 4 years are able to produce a good FEV<sub>1</sub> (50, 53). Indications for FEV<sub>0.5</sub> value interpretation have been reported (52, 54). The GLI equations include reference equations for preschool children since 3 years of age and also predictive equations for FEV<sub>0.75</sub> (55).

In young children with wheeze spirometry impairment vary among the studies. In a big population study set in Seoul, children with recurrent wheeze had lower FEV<sub>1</sub>/FVC and FEF<sub>25-75</sub> than healthy controls [56]. In Argentinian children aged 3-5 years FVC and FEV<sub>0.75</sub> were significantly lower than those of healthy peers (57). In the U-Biopred cohort spirometry did not detect any difference in lung function between preschool children with severe and mild/moderate wheeze arguing that FEV<sub>1</sub> is a poor index of disease severity (58). When spirometry is used to assess response to bronchodilators there is little evidence that it can be used in wheezing preschool children (59), but the greatest BDR is usually seen in those who are at more risk for a diagnosis of asthma in later age (60). A change in FEV<sub>0.75</sub> of 11% has been suggested as cut-off to distinguish preschool children with asthma (57).

Children with CF have been followed-up longitudinally using spirometry and most of the studies demonstrate that their lung function is already compromised at preschool age in up to 36% of subjects (61-64). Furthermore, the presence of a bacterial pathogen such as *Pseudomonas aeruginosa* or *Staphylococcus aureus* in the airways was associated with a reduction in FEV<sub>0.75</sub> ranging between 11.3% and 15.6%

(65). However, the abnormalities are often mild and sometimes not detectable, but when assessed longitudinally FEV<sub>1</sub> shows an inverse relationship with the more sensitive parameter lung clearance index (LCI) (66). Spirometry was also recently used in preschoolers with CF to assess the effect of a trial with hypertonic saline and showed improved FVC values after 16 weeks of treatment and decreased FEV<sub>1</sub> and FEF<sub>25-75</sub> in the group on normal saline. In the same study Rint did not detect any effect (67).

In preschool children born preterm, in whom the tracking of lung function might be very useful, the clinical application of spirometry is complicated by the possible cognitive impairment that sometimes is associated with prematurity. In a recent paper on children born extremely preterm including also children aged 4–5 years only 46% of the population was able to complete a full spirometry manoeuvre resulting in an acceptable FVC (48). Despite this, the results showed an impaired lung function in terms of FVC and FEV<sub>1</sub> compared to controls born at term.

Overall, the published studies on measurement of lung function with spirometry in preschoolers show that the test is safe, feasible and reproducible especially if performed by experienced personnel.

### Multiple breath washout

The multiple breath washout (MBW) describes the inhomogeneity of ventilation, particularly in the small airways, by measuring the clearance of a gas from the lungs. The test uses an open circuit and is performed at tidal breathing during which a marker gas, usually nitrogen, is washed out with 100% oxygen. The washout continues until gas concentration has reached levels lower than 1/40 of the initial concentration (3, 68).

Preschool children perform the test in a seated position and a video can be used for distraction and to promote a regular breathing pattern. Minimal cooperation and coordination are required; the test showed a feasibility of 91% in preschool children (85% under 4 years) (69). LCI, moment ratios and the conductive and acinar ventilation heterogeneity (Scond and Sacin) are some of the parameters used to measure ventilation

inhomogeneity. Functional residual capacity (FRC) and the dead space of the conducting airways can also be obtained. LCI is the principal measure considered in MBW and the value most used to interpret the test in clinical practice. LCI is calculated as the number of lung volume turnovers required to clear the lungs of the marker gas to 1/40th of the starting concentration (3, 68). A higher LCI value indicates greater ventilation inhomogeneity and therefore greater disease severity.

Increased LCI and Scond values were found in preschool children with multiple-trigger wheeze compared with episodic viral wheeze and healthy control subjects. In this cohort 39% and 68% of the subjects with multiple-trigger wheeze had abnormal values of LCI and Scond respectively (70). In a recent paper only Scond discriminated preschool children with asthma from healthy controls but the sample was smaller and in asthmatic subjects FeNO was normal maybe suggesting a less severe disease (69). Normal values of LCI were found also in a group of 32 children with asthma including subjects in preschool age but when compared to healthy peers they had slightly higher values (71). These results are concordant with those reported in adults where even in patients with mild asthma LCI is often normal, while the most consistent evidence of ventilation inhomogeneity is in the conducting airways (72). In an interesting paper by Sonnappa et al. MBW was used to evaluate lung function in preschool wheezers who previously had increased reticular basement membrane (RBM) thickness and increased airway eosinophils. The group showed significantly higher median LCI and Scond than healthy controls but these results did not correlate with past RBM thickness or mucosal eosinophilia (73). The only parameter that showed a significant BDR was Scond but just 16% of wheezy children showed a response larger than the determined threshold (73).

In preschool children with CF LCI is consistently elevated (74). LCI value has been shown to be more sensitive than FEV<sub>1</sub> for detecting alterations of peripheral airways (75) and data support its capacity in the recognition of early lung disease and in the prediction of lung function at school age (74, 76). When MBW was performed at different time points, LCI increased in preschool children with CF over 1 year

and worsened during pulmonary exacerbations suggesting that this parameter can track the progression of the disease and can be used to monitor these young patients (66, 77). Because of its sensitivity, MBW might be used to assess the effect of pharmacological treatment in patients with chronic lung disease. In one interventional study in young children with CF, LCI measured, at baseline and after a trial of 48 weeks with hypertonic saline 7% significantly improved (i.e. decreased) compared to the value measured in subjects on isotonic saline 0.9% (78). LCI has also been found to be sensitive in detecting the improvement in ventilation homogeneity 1 month after the antibiotic therapy taken during a pulmonary exacerbation (79). Furthermore, in the same study LCI values correlated with the magnetic resonance scores used to describe lung abnormalities (79).

To our knowledge there are no reports on MBW during preschool age in children born preterm.

The clinical usefulness and the applicability of MBW in the daily care of the patient and in the decision of which treatment apply still need to be defined because there are gaps in the choice of the device and in the standardization of the technique across the different systems. However, LCI may be a valuable tool to investigate ongoing symptoms or as an outcome in clinical research studies (80). At present MBW is not routinely used in clinical practice but in centers where this technique is regularly performed an increase of 1 unit in the LCI value is considered a sign of pulmonary deterioration (82). As shown in the only published reference equations, LCI is dependent on body size and decreases as height increases, particularly in early childhood. Therefore, the upper limit of normal LCI is higher in infants and preschool children than in older subjects (81).

## Conclusions

In conclusion, Rint, FOT, spirometry and MBW are feasible and reproducible in preschoolers. They have a role in identifying changes in airway calibre and compliance and are potentially very useful in the clinical assessment and follow-up of a child with respiratory disease. To confirm their applicability and capability

in tracking lung function over time further studies on the short and long-term utility of these techniques are needed.

## References

1. Global Initiative for Asthma. Global Strategy for Asthma Management and Prevention, 2017. Available from: [www.ginasthma.org](http://www.ginasthma.org)
2. British Thoracic Society, Scottish Intercollegiate Guidelines Network. British guideline for the management of asthma; a national clinical guideline (SIGN 153). 2016 Sep; Available from: <https://www.brit-thoracic.org.uk/document-library/clinical-information/asthma/btssign-asthma-guide-line-2016/>
3. Beydon N, Davis SD, Lombardi E, et al. An Official American Thoracic Society/European Respiratory Society Statement: Pulmonary Function Testing in Preschool Children. *Am J Respir Crit Care Med* 2007; 175: 1304-45.
4. Lombardi E, Sly PD, Concutelli G, et al. Reference values of interrupter respiratory resistance in healthy preschool white children. *Thorax* 2001; 56: 691-95.
5. Merkus PJFM, Stocks J, Beydon N, et al. Reference ranges for interrupter resistance technique: the Asthma UK Initiative. *Eur Respir J* 2010; 36: 157-63.
6. Brussee JE, Smit HA, Koopman LP, et al. Interrupter resistance and wheezing phenotypes at 4 years of age. *Am J Respir Crit Care Med* 2004; 169: 209-13.
7. van de Kant KD, Koers K, Rijkers GT, et al. Can exhaled inflammatory markers predict a steroid response in wheezing preschool children? *Clin Exp Allergy* 2011; 41: 1076-83.
8. Klug B, Bisgaard H. Lung function and short-term outcome in young asthmatic children. *Eur Respir J* 1999; 14: 1185-89.
9. Caudri D, Wijga AH, Hoekstra MO, et al. Prediction of asthma in symptomatic preschool children using exhaled nitric oxide, Rint and specific IgE. *Thorax* 2010; 65: 801-07.
10. Mele L, Sly PD, Calogero C, et al. Assessment and validation of bronchodilation using the interrupter technique in preschool children. *Pediatr Pulmonol* 2010; 45: 633-38.
11. Rosenfeld M, Allen J, Arets BH, et al. An official American Thoracic Society workshop report: optimal lung function tests for monitoring cystic fibrosis, bronchopulmonary dysplasia, and recurrent wheezing in children less than 6 years of age. *Ann Am Thorac Soc* 2013; 10: S1-S11.
12. Vitaliti G, Leonardi S, La Rosa M. Opening interrupter technique in pre-school children with chronic respiratory diseases: a perspective case-control study in the diagnosis of airway hyperresponsiveness. *J Asthma* 2013; 50: 1045-48.
13. Beydon N, Amsallem F, Bellet M, et al. Pulmonary function tests in preschool children with cystic fibrosis. *Am J Respir Crit Care Med* 2002; 166: 1099-104.
14. Terheggen-Lagro SW, Arets HG, van der Laag J, van der Ent CK. Radiological and functional changes over 3 years

- in young children with cystic fibrosis. *Eur Respir J* 2007; 30: 279-85.
15. Kairamkonda VR, Richardson J, Subhedar N, et al. Lung function measurement in prematurely born preschool children with and without chronic lung disease. *J Perinatol* 2008; 28: 199-204.
  16. Vrijlandt EJ, Boezen HM, Gerritsen J, et al. Respiratory health in prematurely born preschool children with and without bronchopulmonary dysplasia. *J Pediatr* 2007; 150: 256-61.
  17. Oostveen E, MacLeod D, Lorino H, et al; ERS Task Force on Respiratory Impedance Measurements. The forced oscillation technique in clinical practice: methodology, recommendations and future developments. *Eur Respir J* 2003; 22: 1026-41.
  18. Goldman MD, Saadeh C, Ross D. Clinical applications of forced oscillation to assess peripheral airway function. *Respir Physiol Neurobiol* 2005; 148: 179-94.
  19. Shi Y, Aledia AS, Galant SP, George SC. Peripheral airway impairment measured by oscillometry predicts loss of asthma control in children. *J Allergy Clin Immunol* 2013; 131: 718-23.
  20. Dellacà RL, Pompilio PP, Walker PP, Duffy N, Pedotti A, Calverley PM. Effect of bronchodilation on expiratory flow limitation and resting lung mechanics in COPD. *Eur Respir J* 2009; 33: 1329-37.
  21. Czövek D, Shackleton C, Hantos Z, et al. Tidal changes in respiratory resistance are sensitive indicators of airway obstruction in children. *Thorax* 2016; 71: 907-15.
  22. Frei J, Jutla J, Kramer G, Hatzakis GE, Ducharme FM, Davis GM. Impulse oscillometry: reference values in children 100 to 150 cm in height and 3 to 10 years of age. *Chest* 2005; 128: 1266-73.
  23. Lai SH, Yao TC, Liao SL, et al. Reference value of impulse oscillometry in taiwanese preschool children. *Pediatr Neonatol* 2015; 56: 165-70.
  24. Park JH, Yoon JW, Shin YH, et al. Reference values for respiratory system impedance using impulse oscillometry in healthy preschool children. *Korean J Pediatr* 2011; 54: 64-68.
  25. Knihtilä H, Kotaniemi-Syrjänen A, Pelkonen AS, Kalliola S, Mäkelä MJ, Malmberg LP. Sensitivity of newly defined impulse oscillometry indices in preschool children. *Pediatr Pulmonol* 2017; 52: 598-605.
  26. De Assumpcao MS, da Silva Goncalves E, Oliveira MS, et al. Impulse Oscillometry System and Anthropometric Variables of Preschoolers, Children and Adolescents Systematic Review. *Curr Pediatr Rev* 2017; 13(2): 126-35.
  27. Dencker M, Malmberg LP, Valind S, et al. Reference values for respiratory system impedance by using impulse oscillometry in children aged 2-11 years. *Clin Physiol Funct Imaging* 2006; 26: 247-50.
  28. Gochicoa-Rangel L, Del Río-Hidalgo R, Hernández-Ruiz J, et al. Validating Reference Equations for Impulse Oscillometry in Healthy Mexican Children. *Respir Care* 2017; 62: 1156-65.
  29. Calogero C, Parri N, Baccini A, et al. Respiratory impedance and bronchodilator response in healthy Italian preschool children. *Pediatr Pulmonol* 2010; 45: 1086-94.
  30. Calogero C, Simpson SJ, Lombardi E, et al. Respiratory impedance and bronchodilator responsiveness in healthy children aged 2 to 13 years. *Pediatr Pulmonol* 2013; 48: 707-15.
  31. Shackleton C, Czovek D, Grimwood K, et al. Defining 'healthy' in preschool-aged children for forced oscillation technique reference equations. *Respirology* 2017 Oct 5 doi: 10.1111/resp.13186 [Epub ahead of print].
  32. Nielsen KG, Bisgaard H. Discriminative capacity of bronchodilator response measured with three different lung function techniques in asthmatic and healthy children aged 2 to 5 years. *Am J Respir Crit Care Med* 2001; 164: 554-59.
  33. Oostveen E, Dom S, Desager K, Hagendorens M, De Backer W, Weyler J. Lung function and bronchodilator response in 4-year-old children with different wheezing phenotypes. *Eur Respir J* 2010; 35: 865-72.
  34. Hamrin C, Gangell CL, Udomittipong K, et al. Assessment of bronchodilator responsiveness in preschool children using forced oscillations. *Thorax* 2007; 62: 814-19.
  35. Marotta A, Klinnert MD, Price MR, Larsen GL, Liu AH. Impulse oscillometry provides an effective measure of lung dysfunction in 4-year-old children at risk for persistent asthma. *J Allergy Clin Immunol* 2003; 112: 317-22.
  36. Schulze J, Biedebach S, Christmann M, Herrmann E, Voss S, Zielen S. Impulse Oscillometry as a Predictor of Asthma Exacerbations in Young Children. *Respiration* 2016; 91: 107-14.
  37. Song TW, Kim KW, Kim ES, Park JW, Sohn MH, Kim KE. Utility of impulse oscillometry in young children with asthma. *Pediatr Allergy Immunol* 2008; 19: 763-68.
  38. Shin YH, Yoon JW, Choi SH, et al. Use of impulse oscillometry system in assessment of asthma severity for preschool children. *J Asthma* 2013; 50: 198-203.
  39. Knihtilä H, Kotaniemi-Syrjänen A, Mäkelä MJ, Bondestam J, Pelkonen AS, Malmberg LP. Preschool oscillometry and lung function at adolescence in asthmatic children. *Pediatr Pulmonol* 2015; 50: 1205-13.
  40. Shin YH, Jang SJ, Yoon JW, et al. Oscillometric and spirometric bronchodilator response in preschool children with and without asthma. *Can Respir J* 2012; 19: 273-77.
  41. Komarow HD, Skinner J, Young M, et al. A study of the use of impulse oscillometry in the evaluation of children with asthma: analysis of lung parameters, order effect, and utility compared with spirometry. *Pediatr Pulmonol* 2012; 47: 18-26.
  42. Gangel CL, Horak F Jr, Patterson HJ, et al. Respiratory impedance in children with cystic fibrosis using forced oscillations in clinic. *Eur Respir J* 2007; 30: 892-97.
  43. Kerby G, Rosenfeld M, Ren CL, et al. Lung function distinguishes preschool children with CF from healthy controls in a multi-center setting. *Pediatr Pulmonol* 2012; 47: 597-605.
  44. Ramsey KA, Ranganathan SC, Gangell CL, et al; AREST CF. Impact of lung disease on respiratory impedance in young children with cystic fibrosis. *Eur Respir J* 2015; 46: 1672-79.

45. Sakarya A, Uyan ZS, Baydemir C, et al. Evaluation of children with cystic fibrosis by impulse oscillometry when stable and at exacerbation. *Pediatr Pulmonol* 2016; 51: 1151-58.
46. Moreau L, Crenesse D, Berthier F, Albertini M. Relationship between impulse oscillometry and spirometric indices in cystic fibrosis children. *Acta Paediatr* 2009; 98: 1019-23.
47. Udomittipong K, Sly PD, Patterson HJ, Gangell CL, Stick SM, Hall GL. Forced oscillations in the clinical setting in young children with neonatal lung disease. *Eur Respir J* 2008; 31: 1292-99.
48. Verheggen M, Wilson AC, Pillow JJ, Stick SM, Hall GL. Respiratory function and symptoms in young preterm children in the contemporary era. *Pediatr Pulmonol* 2016; 51: 1347-55.
49. Gunlemez A, Er İ, Baydemir C, Arisoy A. Effects of passive smoking on lung function tests in preschool children born late-preterm: a preventable health priority. *J Matern Fetal Neonatal Med* 2018; Feb 1: 1-6 [Epub ahead of print].
50. Aurora P, Stocks J, Oliver C, et al; London Cystic Fibrosis Collaboration. Quality control for spirometry in preschool children with and without lung disease. *Am J Respir Crit Care Med* 2004; 169: 1152-59.
51. Gaffin JM, Shotola NL, Martin TR, Phipatanakul W. Clinically useful spirometry in preschool-aged children: evaluation of the 2007 American Thoracic Society Guidelines. *J Asthma* 2010; 47: 762-67.
52. Kampschmidt JC, Brooks EG, Cherry DC, Guajardo JR, Wood PR. Feasibility of spirometry testing in preschool children. *Pediatr Pulmonol* 2016; 51: 258-66.
53. Crenesse D, Berlioz M, Bourrier T, Albertini M. Spirometry in children aged 3 to 5 years: reliability of forced expiratory maneuvers. *Pediatr Pulmonol* 2001; 32: 56-61.
54. Nystad W, Samuelsen SO, Nafstad P, Edvardsen E, Stensrud T, Jaakkola JJK. Feasibility of measuring lung function in preschool children. *Thorax* 2002; 57: 1021-27.
55. Quanjer PH, Stanojevic S, Cole TJ, et al; ERS Global Lung Function Initiative. Multi-ethnic reference values for spirometry for the 3-95-yr age range: the global lung function 2012 equations. *Eur Respir J* 2012; 40: 1324-43.
56. Soh JE, Kim K-M, Kwon J-W, et al. Recurrent wheeze and its relationship with lung function and airway inflammation in preschool children: a cross-sectional study in South Korea. *BMJ Open* 2017; 7: e018010.
57. Busi LE, Restuccia S, Tourres R, Sly PD. Assessing bronchodilator response in preschool children using spirometry. *Thorax* 2017; 72: 367-72.
58. Fleming L, Murray C, Bansal AT, et al; U-BIOPRED Study Group. The burden of severe asthma in childhood and adolescence: results from the paediatric U-BIOPRED cohorts. *Eur Respir J* 2015; 46: 1322-33.
59. Raywood E, Lum S, Aurora P, Pike K. The bronchodilator response in preschool children: A systematic review. *Pediatr Pulmonol* 2016; 51: 1242-50.
60. Vilozni D, Barak A, Efrati O, et al. The role of computer games in measuring spirometry in healthy and "asthmatic" preschool children. *Chest* 2005; 128: 1146-55.
61. Marostica PJ, Weist AD, Eigen H, et al. Spirometry in 3- to 6-year-old children with cystic fibrosis. *Am J Respir Crit Care Med* 2002; 166: 67-71.
62. Nielsen KG, Pressler T, Klug B, Koch C, Bisgaard H. Serial lung function and responsiveness in cystic fibrosis during early childhood. *Am J Respir Crit Care Med* 2004; 169: 1209-16.
63. Vilozni D, Bentur L, Efrati O, et al. Spirometry in early childhood in cystic fibrosis patients. *Chest* 2007; 131: 356-61.
64. Kerby GS, Rosenfeld M, Ren CL, et al. Lung function distinguishes preschool children with CF from healthy controls in a multi-center setting. *Pediatr Pulmonol* 2012; 47: 597-605.
65. Ramsey KA, Ranganathan S, Park J, et al; AREST CF. Early respiratory infection is associated with reduced spirometry in children with cystic fibrosis. *Am J Respir Crit Care Med* 2014; 190: 1111-16.
66. Stanojevic S, Davis SD, Retsch-Bogart G, et al. Progression of Lung Disease in Preschool Patients with Cystic Fibrosis. *Am J Respir Crit Care Med* 2017; 195: 1216-25.
67. Nenna R, Midulla F, Lambiase C, et al. Effects of inhaled hypertonic (7%) saline on lung function test in preschool children with cystic fibrosis: results of a crossover, randomized clinical trial. *Ital J Pediatr* 2017; 43: 60.
68. Robinson PD, Latzin P, Verbanck S, et al. Consensus statement for inert gas washout measurement using multiple- and single- breath tests. *Eur Respir J* 2013; 41: 507-22.
69. Vilmann L, Buchvald F, Green K, Nielsen KG. Fractional exhaled nitric oxide and multiple breath nitrogen washout in preschool healthy and asthmatic children. *Respir Med* 2017; 133: 42-47.
70. Sonnappa S, Bastardo CM, Wade A, et al. Symptom-pattern phenotype and pulmonary function in preschool wheezers. *J Allergy Clin Immunol* 2010; 126: 519-26.
71. Zwitterloot A, Fuchs SI, Müller C, Bisdorf K, Gappa M. Clinical application of inert gas Multiple Breath Washout in children and adolescents with asthma. *Respir Med* 2014; 108: 1254-59.
72. Verbanck S, Schuermans D, Paiva M, Vincken W. Nonreversible conductive airway ventilation heterogeneity in mild asthma. *J Appl Physiol* 2003; 94: 1380-86.
73. Sonnappa S, Bastardo CM, Wade A, Bush A, Stocks J, Aurora P. Repeatability and bronchodilator reversibility of lung function in young children. *Eur Respir J* 2013; 42: 116-24.
74. Belessis Y, Dixon B, Hawkins G, et al. Early cystic fibrosis lung disease detected by bronchoalveolar lavage and lung clearance index. *Am J Respir Crit Care Med* 2012; 185: 862-73.
75. Aurora P, Bush A, Gustafsson P, et al. Multiple-breath washout as a marker of lung disease in preschool children with cystic fibrosis. *Am J Respir Crit Care Med* 2005; 171: 249-56.
76. Aurora P, Stanojevic S, Wade A, et al. Lung clearance index at 4 years predicts subsequent lung function in children with cystic fibrosis. *Am J Respir Crit Care Med* 2011; 183: 752-58.



77. Ramsey KA, Foong RE, Grdasic J, et al; Australian Respiratory Early Surveillance Team for Cystic Fibrosis (AREST CF). Multiple-Breath Washout Outcomes Are Sensitive to Inflammation and Infection in Children with Cystic Fibrosis. *Ann Am Thorac Soc* 2017; 14: 1436-42.
78. Subbarao P, Stanojevic S, Brown M, et al. Lung clearance index as an outcome measure for clinical trials in young children with cystic fibrosis. A pilot study using inhaled hypertonic saline. *Am J Respir Crit Care Med* 2013; 188: 456-60.
79. Stahl M, Wielpütz MO, Graeber SY, et al. Comparison of Lung Clearance Index and Magnetic Resonance Imaging for Assessment of Lung Disease in Children with Cystic Fibrosis. *Am J Respir Crit Care Med* 2017; 195: 349-59.
80. Subbarao P, Milla C, Aurora P, et al. Multiple-Breath Washout as a Lung Function Test in Cystic Fibrosis. A Cystic Fibrosis Foundation Workshop Report. *Ann Am Thorac Soc* 2015; 12: 932-39.
81. Lum S, Stocks J, Stanojevic S, et al. Age and height dependence of lung clearance index and functional residual capacity. *Eur Respir J* 2013; 41: 1371-77.
82. Singer F, Kieninger E, Abbas C, et al. Practicability of nitrogen multiple-breath washout measurements in a pediatric cystic fibrosis outpatient setting. *Pediatr Pulmonol* 2013; 48: 739-46.

Received: 3 March 2018

Accepted: 5 March 2018

Correspondence:

Dr Enrico Lombardi

Paediatric Pulmonary Unit

“Anna Meyer” Paediatric University Hospital

Viale Pieraccini 24 - 50139 Florence, Italy

Tel. +39 055 566-2461

E-mail: enrico.lombardi@meyer.it