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REVIEW

Natural products remodel cancer-associated fibroblasts in desmoplastic tumors



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KEY WORDS

Natural products; Desmoplastic tumors; Cancer-associated fibroblasts; Tumor microenvironment; Extracellular matrix; Traditional Chinese medicine; Cancer treatment **Abstract** Desmoplastic tumors have an abundance of stromal cells and the extracellular matrix which usually result in therapeutic resistance. Current treatment prescriptions for desmoplastic tumors are usually not sufficient to eliminate the malignancy. Recently, through modulating cancer-associated fibroblasts (CAFs) which are the most abundant cell type among all stromal cells, natural products have improved chemotherapies and the delivery of nanomedicines to the tumor cells, showing promising ability to improve treatment effects on desmoplastic tumors. In this review, we discussed the latest advances in inhibiting desmoplastic tumors by modeling CAFs using natural products, highlighting the potential therapeutic abilities of natural products in targeting CAFs for cancer treatment.

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1. Introduction

The tumor microenvironment consists of stromal cells (e.g., endothelial cells, immunological cells, stromal macrophages, pericytes and fibroblasts) and the acellular compartments including extracellular matrix (ECM) and cytokines, supporting and promoting tumor progression (Fig. 1)¹. Desmoplastic tumors, such as breast cancer², pancreatic ductal adenocarcinoma $(PDAC)^3$, prostate cancer $(PC)^4$ and cholangiocarcinoma⁵, that are aggressive with very poor prognosis, are usually characterized with high levels of stromal cell density and ECM concentrations. Among all the stromal cells, cancer-associated fibroblasts (CAFs) are the most abundant cell type. The rapidly proliferating cancer cells, CAFs and ECM fibers (such as collagen and hyaluronan) in desmoplastic tumors with the surrounding normal tissue⁶ form a highly complex and heterogeneous tumor microenvironment, causing intratumoral solid stresses gathering, vessel constricting and hypoperfusion⁷. The highly complex and heterogonous pathophysiology limits the effective delivery of drugs to tumors⁸.

There are both quiescent and activated fibroblasts in the tumors. Activated fibroblasts related to cancer have been termed CAFs or tumor-associated fibroblasts⁹. The α -smooth muscle actin (α -SMA) has been regarded as the most reliable marker of activated fibroblasts¹⁰. The CAFs can be developed from several cell types, including resident tissue fibroblasts, epitheliums *via* epithelial-to-mesenchymal transition (EMT), endothelial cells *via*



Figure 1 Desmoplastic tumor microenvironment. Desmoplastic tumor microenvironment is characterized with the whole noncancer components around tumor cells, including fibroblasts, macrophages, lymphocytes, extracellular matrix and intertwined blood vessels composed of endothelial cells and pericytes.

endothelial-to-mesenchymal transition, bone marrow-derived cells (BMDCs), adipose cells and stellate cells¹⁰. Through expressing a series of proteins such as α -SMA, tenascin and collagen, the CAFs enhance the complexity and heterogeneity of pathophysiology in tumor microenvironment, causing physiological abnormalities¹¹. In desmoplastic tumor microenvironment, solid stress, winding and leaky blood vessel networks, collapsed and nonfunctional lymphatic vessels and highly dense ECM accumulation are the major physiological abnormalities¹², compromising the main important process for efficient molecular transports: vascular transport, transvascular transport, interstitial transport, and cellular uptake $(Fig. 1)^{13}$. Due to these transport barriers, most anticancer drugs cannot reach the effective dose in the cancer site as they have poor distribution and penetration into desmoplastic tumors. Therefore, chemotherapy and nanotherapy many times fail to treat desmoplastic tumors even if they are potent enough to kill cancer cells in vitro.

Besides, CAFs are the dominant component of the tumor stroma, where they produce a wide range of mediators including transforming growth factor- β (TGF- β)¹⁴, interleukin 6 (IL-6), matrix metalloproteinases (MMPs)¹⁵ vascular endothelial growth factor (VEGF)¹⁶ hepatocyte growth factor (HGF)¹⁷, stromal cellderived factor 1 (SDF-1)¹⁸, hypoxia-inducible factor-1 (HIF-1)¹⁹ and tissue inhibitors of metalloproteinases (TIMPs)²⁰, etc. These secreted mediators are receptor ligands overexpressed by other cell types in the tumor microenvironment, which cause intercellular crosstalk, resulting in cancer progression and metastasis. What's more, desmoplasia results in high activity of CAFs and the accumulation of stromal components around tumors. CAFs further talk with different stromal components in the desmoplastic tumor microenvironment. By activating various pathways such as TGF- β^{14} , CAFs can affect the angiogenesis and the ECM of the desmoplastic tumor stroma, as well as affect the invasion, proliferation and migration of cancer cells. By remodeling the ECM, CAFs may be involved in the generation and maintenance of the desmoplastic tumor stem cell niche²¹. By affecting WNT signaling, they can also induce cancer cell drug resistance²². Therefore, CAFs play an important role in the initiation, development and metastasis of desmoplastic tumors.

Recently, the study of Ligorio et al.²³ also found that CAFs influenced tumor architecture by changing the intrinsic patterns of tumor glands in human PDAC, which further demonstrated the importance of tumor-stroma interaction in desmoplastic tumor treatment. More and more scientists realized the importance of CAFs in the treatment of desmoplastic tumors, and attempted to target them to improve desmoplastic tumor therapy 24 , such as directly depleting CAFs and normalizing CAFs. Ji et al.²⁵ designed an MMP-2 responsive peptide-hybrid liposome to down-regulate ECM levels through regulating the CAFs, and enhance penetration of GEM, which greatly enhanced the therapeutic efficacy on pancreatic cancer. Recently, Zhang et al.²⁴ also modulated CAFs to promote the accumulation of docetaxel in tumor with dexamethasone, which gave insights in overcoming the physiological abnormalities in desmoplastic tumor and provided a rational strategy to increase antitumor efficacy. Therefore, CAFs have emerged as the key target of drug delivery in antidesmoplastic tumor therapies.

In recent years, natural products have been widely studied and showed promising ability to improve treatment on desmoplastic tumors. They have been reported to regulate tumor stroma especially CAFs through multiple mechanisms (Fig. 2), thus normalizing the microenvironment of desmoplastic tumors to improve



Figure 2 Schematic diagram illustrates the CAF-remodeling effects of natural products in the desmoplastic tumor microenvironment. There are three general aspects that natural products target or modulate CAFs in the desmoplastic tumor microenvironment. First, natural products inhibit the expression of mediators which could initiate cell crosstalk between CAFs and tumor cells and result in cancer invasion, proliferation and migration. Second, natural products induce the ECM remodeling process and indirectly inhibit cancer cell invasion. Third, natural products can inhibit angiogenesis which is promoted by CAFs. CAV-1, caveolin 1; CXCR4, CXC chemokine receptor 4; M-CSF, macrophage colony-stimulating factor; MCP-1, monocyte chemotactic protein-1; ROS, reactive oxygen species; SDF-1, stromal cell-derived factor 1.

the delivery of chemotherapeutics and nanomedicines²⁶. Besides, natural products can inhibit the interaction between tumorassociated stroma and cancer cells, thus inhibiting cancer cell proliferation, invasion, migration and drug-resistance. The delivery of chemotherapeutics could be improved and the efficacy of chemical drugs could be enhanced through remodeling CAFs by natural products. In this review, we summarize recent studies on natural products which can inhibit desmoplastic tumor progression by remodeling CAFs.

2. Natural products affect the CAF-cancer cell crosstalk

CAFs produce a wide range of mediators, which are receptor ligands overexpressed by other cell types in the tumor microenvironment, initiating cell crosstalk, resulting in cancer invasion, proliferation and migration. Natural products can modulate the signal transduction involved in the interaction of CAFs with cancer cells, thereby inhibiting the invasion, proliferation and migration of desmoplastic tumors.

2.1. Curcumin

Curcumin, commonly known as turmeric, is a natural yellow polyphenolic compound derived from the rhizomes of the plant *Curcuma longa Linnaeus* and is considered as an effective drug to cure various diseases, including allergy, asthma, anorexia, bronchial hyperactivity, cough and sinusitis²⁷. Curcumin has a variety of biological functions, such as anti-oxidant, anti-bacterial, anti-inflammatory, anti-microbial, and anti-cancer effects²⁸.

Curcumin is capable of abrogating the invasion-promoting capacity of CAFs by increasing E-cadherin levels and decreasing vimentin levels. Moreover, through inhibiting monoamine oxidase A/mammalian target of rapamycin/HIF-1 α signaling pathway, curcumin inhibits the production of reactive oxygen species and the expression of CXC chemokine receptor 4 (CXCR4) and IL-6 receptor in PC cells, which supports the therapeutic effects of curcumin in PC²⁹.

Curcumin is able to inhibit CAF-induced migration of desmoplastic tumors through reducing the mesenchymal characteristic of CAFs³⁰ or up-regulating tumor suppressor proteins (P16, P21, and P53). Curcumin can up-regulate tumor suppressor proteins in CAFs while inactivate the JAK2/STAT3 pathway, leading to the decrease of α -SMA. Moreover, curcumin also inhibits the migration of tumor cells which is induced by CAFs *via* reducing the secretion of SDF-1, IL-6, MMP-2, MMP-9, and TGF- β^{31} . In addition, curcumin can repress CAF-induced migration and invasion of desmoplastic tumors through increasing the level of the P16 coding *CDKN2A* mRNA and miR-146b-5p which is an important tumor suppressor miRNA, leading to IL-6 production³².

Since curcumin can improve the sensitivity of tumor cells to chemotherapeutic drugs and radiotherapy through regulating CAFs, scientists studied the inhibitory abilities of curcumin and 5-fluorouracil (5-FU) on the survival of cancer stem cell (CSC) in a 3D co-culture model (Table 1)³³. Through NF- κ B pathways, curcumin can sensitize CSCs to 5-FU treatment *via* blocking the CAF-induced invasion of CSCs, emphasizing that curcumin is a promising modulator to coordinate the crosstalk in the tumor microenvironment³³.

| Remodeling effect | Drug | Cancer model | Cell line | Animal model | Mechanism | Combi- nation with | Ref. |
|---|------------------------------------|-------------------------------------|---|---|--|-----------------------|------|
| The CAF-cancer cell crosstalk | Curcumin | PC | PC3; Human prostate CAFs | _ | E-cadherin elevation; Vimentin inhibition | _ | 29 |
| | | Breast cancer | MDA-MB-231; MCF-10; human breast CAFs | MDA-MB-231 xenograft model; | Tumor suppressor proteins elevation; JAK2/STAT3 pathway inhibition; tumor suppressor miRNA elevation | - | 31 |
| | | Colon tumor | HCT116; MRC-5 | _ | NF- κ B pathways inhibition | 5-FU | 33 |
| The CAF-cancer cell crosstalk | Silibinin | PC | PC3; DU-145; LNCaP; 22Rv1 | PC3 xenograft model | TGF- β pathways inhibition; E-cadherin elevation; MCP-1inhition; fibronectin inhibition | _ | 34-3 |
| Angiogenesis | | | LNCaP; 22Rv1 | 22Rv1 xenograft model | HIF-1 signaling pathways inhibition | - | 37 |
| Angiogenesis | Fraxinellone | Human non-small cell lung cancer | A549; HeLa; Hep3B; HUVEC; HLF-a; HCT110 | A549 xenograft model | HIF-1 and STAT3 signaling pathways inhibition | _ | 38 |
| The CAF-cancer cell crosstalk | | Melanoma | BPD6 | BPD6 transplantable tumor model | TGF- β pathways inhibition | Peptide vaccine | 39 |
| | | Pancreatic tumor | Panc-1; NIH3T3 | Orthotropic Panc-1 xenograft model | | siRNA | 40 |
| The CAF-cancer cell crosstalk | Triptolide | Pancreatic tumor | KPC001; Panc-02 | Patient-derived xenograft model; KPC transgenic mouse model | TGF- β pathways inhibition | Doxorubicin | 41,4 |
| The CAF-cancer cell crosstalk | Triptonide | Gastric cancer | BGC-823; human GCAFs | - | Regulate microRNA expression | . — | 43 |
| The CAF-cancer cell crosstalk | ASV | Gastric cancer | BGC-823; Human GCAFs | - | Regulate microRNA expression | | 44 |
| The CAF-cancer cell crosstalk | Resveratrol | Cholangiocarcinoma | KKU-213; KKU-100; MMNK-1; Cholangiocarcinoma CAFs | - | Inhibiting IL-6 expression | - | 45 |
| | | Breast cancer | MCF-7; MDA-MB-231; human breast CAFs | - | Inhibiting the mRNA t ranscription | _ | 46 |
| | | PDAC | Capan1; AsPC1; PanO2; MP1070 | Orthotropic xenograft model | _ | 5-FU | 47 |
| The CAF-cancer cell crosstalk | EGCG | PC | - | - | Inhibiting HGF and VEGF production | _ | 48 |
| ECM | | | WPMY-1; HPS-19I | - | TGF- β pathways inhibition | Luteolin | 49 |
| The CAF-cancer cell crosstalk | Emodin | Breast cancer | BT20; human breast CAFs | - | EMT programming inhibition | - | 50 |
| The CAF-cancer cell crosstalk | Artesunate and dihydroartemisin | Breast cancer | 4T1; L-929 | Orthotropic xenograft model | TGF- β pathways inhibition | _ | 51 |
| ECM and The CAF-cancer cell crosstalk | СРА | Non-small cell lung cancer | A549; NCI-H520; CCL-206; primary human lung squamous CAFs | - | Hh pathway inhibition | _ | 52 |
| | | Cholangiocarcinoma | SNU-1196; SNU-246; SNU-308; SNU-1079; HuCCT-1; Lx-2 | HuCCT-1 xenograft model; co-implant xenograft with | | - | 53 |

| Remodeling effect | Drug | Cancer model | Cell line | Animal model | Mechanism | Combi- nation with | Ref. |
|---|-------------|------------------|--|--|--|-----------------------|------|
| | | Pancreatic tumor | Capan-2; Human umbilical vein endothelial cell line | Capan-2 xenograft model | | I | 54 |
| | | PDAC | Miapaca-2; L3.6pl; PANC-1; human pancreatic stellate cell line | 1 | | Cs-137 radiation | 55 |
| | | | MiaPaca-2-luc; human pancreatic stellate cell line | Orthotopic MiaPaca-2- luc pancreatic tumor xenografts; Patient-derived orthotopic model; KPC-Luc transgenic | | УТХ | 56 |
| ECM | Celastrol | Melanoma | BPD6; D4M; NIH-3T3 | BPD6 transplantable tumor model | Reducing the production of collagen | Mitoxantrone 57 | 57 |
| ECM and the CAF-cancer Quercetin cell crosstalk | r Quercetin | Bladder cancer | NIH3T3; UMUC3 | Co-implant xenograft with NIH3T3 and UMUC3 | WNT16 expression inhibition | Cisplatin | 58 |
| The CAF-cancer cell crosstalk | | Breast cancer | MCF7; MDA-MB 231; human skin fibroblasts | I | CAV-1 expression elevation | I | 59 |

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2.2. Silibinin

Silibinin is the main bio-active component of silymarin which is isolated from the seeds of *Silybum marianum* (L.) Gaertn (Family Asteraceae). It is widely used as a hepatoprotective agent and has been marketed as a dietary supplement⁶⁰. Silibinin has shown broad-spectrum anticancer activities, and is currently being studied clinically⁶¹. It has been reported that silibinin disrupts important signaling pathways which are necessary for PC cell proliferation, invasion, migration and metastasis, induces PC cell apoptosis and inhibits angiogenesis and EMT process⁶².

It was also reported that silibinin could inhibit PC cells' ability to activate CAFs, which highlighted the potential use of silibinin in PC prevention. Silibinin could suppress the TGF- β 2-induced CAF-like phenotype of naïve fibroblasts. Ting et al.³⁴ found that control conditioned medium (CCM) from human PC cells could induce a CAF-like phenotype in human prostate stromal cells, leading to increased α -SMA and vimentin expression. And silibinin could strongly inhibit the TGF- β 2-dependent increase in α -SMA and CAF-like phenotype (Table 1)³⁴.

Silibinin is also able to inhibit CAF-induced invasion of desmoplastic tumors through enhancing E-cadherin expression. Ting et al.³⁵ explored the function of CAFs to promote PC invasion, and the targeting efficacy of silibinin on this response. They treated CAFs with silibinin or vehicle, and collected the conditioned media from CAFs, labeling as silibinin-treatment conditioned media (SBCM) or CCM, respectively. Their results illustrated that SBCM could significantly increase E-cadherin expression and inhibit invasiveness of PCA cells compared with CCM. Further studies found that silibinin was able to suppress CAF-induced invasion of tumors through reducing the expression of monocyte chemotactic protein-1 (MCP-1) which was a key component of CCM. Silibinin strongly reduced the expression of MCP-1 by inhibiting the DNA-binding activity of MCP-1 transcriptional regulators-AP-1 and NF-kB. Besides, silibinin can inhibit the CAF-induced desmoplastic tumor invasiveness and proliferation by inhibiting the CAFs' ability to secret fibronectin³⁶.

2.3. Fraxinellone

As a member of limonoids, fraxinellone is a natural product isolated from the root bark of the Rutaceae plant, *Dictamnus dasycarpus*, exhibiting neuroprotective, anti-inflammatory, antifibrosis and anti-cancer activities³⁸. It has been reported that fraxinellone could suppress proliferation and angiogenesis of cancer cells *in vivo* through inhibiting programmed cell deathligand 1 expression *via* reducing HIF-1 and STAT3 signaling pathways³⁸. Besides, fraxinellone can cure liver fibrosis through decreasing the expression of CUG-binding protein 1 (CUGBP1) and then regulating TGF- β and interferon γ signaling⁶³.

Fraxinellone is reported to inhibit tumor cells' ability to activate CAFs through regulating the TGF- β signaling pathway. Hou et al.³⁹ designed a nanoemulsion (NE) formulation to deliver fraxinellone to CAFs in desmoplastic melanoma. To improve the anti-melanoma effect, they combined fraxinellone NEs with a tumor-specific peptide vaccine. They found that fraxinellone NEs could down-regulate the protein expression of α -SMA and CUGBP1 in the NIH-3T3 cell line (which was activated with TGF- β to mimic CAFs *in vitro*), and after treated with fraxinellone NEs in melanoma-bearing mice, the mRNA expressions of TGF- β were reduced, accompanying with a decreased expression of α -SMA and CUGBP1. Moreover, combining the tumor-specific

peptide vaccine with fraxinellone NEs could improve the tumorspecific T-cell infiltration, activate death receptors on the tumor cell surface, and induce the death of apoptotic tumor cells (Table 1)³⁹. Pei et al.⁴⁰ designed a fraxinellone-loaded CGKRK peptidemodified nanoparticle (Frax-NP-CGKRK) to regulate the stromal TME of pancreatic cancer, and found that Frax-NP-CGKRK could also reverse the CAFs to the quiescent state by suppressing TGF- β signaling pathway, decrease the collagen accumulation and increase tumor blood perfusion (Fig. 3). Furthermore, they applied Frax-NP-CGKRK to enhance the intratumor drug penetration of siRNA-loaded lipid-coated calcium phosphate (LCP) biomimetic high-density lipoprotein nanoparticles (siKras-LCP-ApoE3), which could damage pancreatic cancer cells, and evaluated the antitumor activity of the combination therapy. They found that Frax-NP-CGKRK can make ways for the delivery of siKras-LCP-ApoE3, and this combination therapy remarkably suppressed the tumor growth and prolonged the survival time of animals bearing pancreatic cancer.

2.4. Triptolide and triptonide

Triptolide (TPL) is a diterpenoid epoxy compound extracted from the *Tripterygium wilfordii* Hook F, which has dual effects of antiinflammatory and anti-tumor⁶⁴. TPL is used to treat bleomyciinduced pulmonary fibrosis⁶⁵, liver fibrosis⁶⁶ and renal fibrosis⁶⁷ in mice. Besides the effect on fibrosis, TPL has also been tested in researches as an anticancer compound since the 1990's. It has been reported to have great efficacy on moderating pancreatic tumor⁶⁸. However, the poor solubility of TPL in aqueous medium restricts its use in clinical and preclinical studies. Therefore, a water-soluble prodrug of TPL, minnelide, has been evaluated in a phase I clinical trial against gastrointestinal cancer. Besides weakening the primary tumor burden, minnelide also decreases metastasis in pancreatic, liver and ovarian cancers.

Recent publications demonstrated that both of minnelide and TPL are able to inhibit tumor epithelial cells' ability to activate CAFs, and normalize CAFs through inhibiting the TGF- β signaling pathway⁴¹. Besides, minnelide can deplete the stroma by preventing the hyaluronan synthesis and collagen stabilization in pancreatic tumor. Treatment with minnelide reduces the viability of CAFs isolated from the pancreatic tumor⁴², leading to suppression of tumor invasion.

Triptonide is also a major active compound of *Tripterygium wilfordii* Hook F⁶⁴, which can also inhibit the pathological functions of CAFs, resulting in the inhibition of cancer cell migration. Wang et al.⁴³ found that triptonide treatment strongly suppressed the colony formation, migration and invasion-promoting abilities of gastric CAFs (GCAFs) by down-regulating miRNA-301a expression and up-regulating miRNA-149 expression in GCAFs. Furthermore, treatment with triptonide could inhibit EMT in gastric cancer cells induced by GCAFs (Table 1)⁴³. Their research results showed that triptonide inhibited the cancer-promoting ability of GCAFs. Therefore, triptonide is a promising therapeutic agent to affect the CAF-cancer cell crosstalk and to treat gastric cancer.

2.5. Astragaloside IV

Astragaloside IV (ASV) is the main active component of Astragali Radix which is a crucial Chinese herb prescribed to strengthen the body of patients and eliminate toxins from their bodies⁶⁹. ASV has multiple pharmacologic effects for its potent immunoregulatory, anti-asthma, anti-inflammatory and anti-fibrotic activities⁷⁰. It has been demonstrated that ASV protects against the progression of liver fibrosis⁷¹, renal fibrosis⁷², myocardial fibrosis⁷³, as well as systemic sclerosis⁷⁴, all without evident toxicity or side effects. And the antifibrotic effect of ASV is mediated by the MAPK pathway and TGF- β /Smad signaling pathway⁷⁵. Che et al.⁷⁶ examined the effect of ASV on the procedures associated with renal fibrosis in cultured TGF- β 1-activated mouse renal fibroblasts. Their results illustrated that ASV inhibited TGF- β 1induced fibroblast proliferation, transdifferentiation, and ECM production in a dose-dependent manner. Moreover, the inhibition effect of ASV on fibroblast differentiation and ECM formation was achieved by regulating the activity of MAPK and NF- κ B signaling pathways.

Besides the prominent antifibrotic effects, ASV is able to inhibit the proliferation- migration- and invasion-promoting abilities of GCAFs. Wang et al.⁴⁴ treated GCAFs with ASV, and then observed the effect of ASV on the malignancy-promoting capacity of GCAFs to explore the mechanism. They found that ASV could significantly inhibit the proliferation, migration and invasionpromoting effects of GCAFs through enhancing miRNA-214 and reducing miRNA-301a expression. By reestablishing the miRNA expression balance, ASV could effectively suppress macrophage colony-stimulating factor expression in CAFs which is an important role in promoting tumor proliferation, and elevate the tissue inhibitor of metalloproteinase 2 expression in CAFs which is a tumor suppressive factor. Thus, ASV can suppress the pathological functions of GCAFs, thereby inhibiting the gastric cancer cell progression, suggesting that ASV is a potent therapeutic agent for cancer by regulating CAFs.

2.6. Resveratrol

As a member of stilbenoids, resveratrol (*trans*-3,5,4'-trihydroxystilbene) was first isolated from the roots of white hellebore (*Veratrum grandiflorum O*. Loes) in 1940. It is also a component of grapes and a main constituent in red wine, the consumption of which has been closely related to the lower incidence of cardiac infarction in France than in other comparable countries⁷⁷. Resveratrol is a phytoalexin which has broad-spectrum effects including anti-infective, anti-oxidant, and cardioprotective functions⁷⁸.

Since Jang et al.⁷⁹ published the first article about the anticancer potential of resveratrol in 1997, a great interest from cancer scientists has focused on this molecule from then on. Resveratrol is able to inhibit the NF- κ B signaling pathway effectively and has been found to inhibit the growth of a wide variety of transplanted tumors in rodents including neuroblastoma, hepatoma, breast carcinoma, gastric carcinoma, colon carcinoma, and leukemia⁸⁰. Additionally, studies demonstrate that resveratrol modulates the interaction between desmoplastic tumor cells and the CAFs in desmoplastic tumor microenvironment, thereby inhibiting cancer cell migration and invasion.

Resveratrol is able to inhibit CAF-induced invasion of desmoplastic tumors by inhibiting the expression of IL-6. Thongchot et al.⁴⁵ found that resveratrol blocked the pro-invasive communication between CAFs and cholangiocarcinoma cells through abrogating the secretion of IL-6 by CAFs. In their study, the CCM could strongly induce IL-6-mediated motility of cholangiocarcinoma cells, while the medium of CAFs pretreated with resveratrol could completely inhibit the movement of cancer cells and strongly induce autophagy of cholangiocarcinoma cells.



Figure 3 Combination therapy modulated the tumor microenvironment, silenced the KRAS mutation and induced apoptosis at the orthotropic pancreatic tumor sites. (A) Masson staining figures and immunohistochemistry images of indicators CD31, α -SMA, fibronectin, and IL-6 in all experimental groups. Scale bars, 200 µm. (B) KRAS protein expression levels in tumor tissues of siKras-LCPApoE3 and Frax-NP-C + sLApoE3 (b) groups. Data represent mea \pm SD (n = 6). The significance of the differences (*P < 0.05, **P < 0.01, ***P < 0.001) was evaluated by two-tailed Student's *t*-test. (C) TUNEL analyses of *in vivo* apoptosis in pancreatic orthotropic tumor cells (red) on the day after the last administration. Nuclei (blue) were stained with Hoechst 33,258. Scale bar, 200 µm. (Used with permission from Ref. 40. Copyright © 2019 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim.)

What is more, resveratrol is also able to inhibit CAF-induced invasion and migration of tumor cells through influencing the mRNA transcription of various mediators. In MDA-MB-231 cells, Suh et al.⁴⁶ found that through suppressing CCM induced transcription of *C-MYC*, cyclin D1, MMP-2 and MMP-9 mRNA (Table 1)⁴⁶, resveratrol could decrease the expression of cyclin D1 and *C-MYC*, as well as regulate MMP-2 and MMP-9, so as to inhibit the migration and invasion of breast cancer cells induced by CCM.

Since resveratrol has a superior effect on remodeling the malignancy-promoting functions of CAFs, resveratrol is regarded as a promising chemo-protector which can be combined with chemotherapeutic agents for cancer treatment. Endostatin, as an angiogenesis inhibitor, can target proliferating endothelial cells. Cytosine deaminase (CD) linked to uracil phosphoribosyltransferase, converts the prodrug 5-fluocytosine (5-FC) to the chemotherapeutic drug 5-FU and has a greater effect to kill cancer cells than CD alone. 5-FC and the anti-angiogenic protein consisting of endostatin and CD (EndoCD) can both inhibit angiogenesis and chemo-therapeutically target tumors, and the targeting induce a bystander-killing effect on endothelial cells and tumor cells surrounding the vessels⁸¹. Chen et al.⁴⁷ investigated the efficacy of the EndoCD/5-FC/resveratrol combination on stroma in stroma-enriched PDAC models. They found that the EndoCD/5-FC/resveratrol combination reduced the amount of collagen, the number of CAFs, the density of tumor vessels, and the number of leukocytes, indicating that EndoCD/5-FC/resveratrol not only killed tumor cells, but also induced apoptosis of stroma cells, including CAFs, endothelial cells and immune cells. By using a noninvasive high-frequency ultrasound imaging technique, they found that resveratrol also increased the protein stability of EndoCD through suppressing chymotrypsin-like proteinase activity so that enhanced EndoCD-mediated 5-FC-induced cell killing. As a result, EndoCD/5-FC/resveratrol regimen suppressed the formation of pancreatic stroma in PDAC, leading to reduced tumor growth and extended survival. Their findings intimated that EndoCD/5-FC/resveratrol may be an ideal treatment strategy for PDAC and should be tested in clinical trials.

2.7. Epigallocatechin-3-gallate

Green tea polyphenols, showing potential in cancer therapy, have been attracting the attention of scientists for a long time⁸². Epigallocatechin-3-gallate (EGCG) isolated from green tea leaves, is reported with anti-oxidant, anti-fibrotic, anti-cancer and antiinflammatory activities. As a prominent antioxidant agent, EGCG is able to be used as an electron trap to scavenge free radicals, suppress the formation of ROS and reduce oxidative stress⁸³. The highly potent antioxidant capability makes EGCG to be a good candidate for reducing both oxidative stress and fibrogenesis in patients with scleroderma (SSc)⁸⁴, decreasing the production of ECM (collagen type I, fibronectin) and the markers of fibrosis (connective tissue growth factor) in scleroderma fibroblasts. Furthermore, it has been reported that the reduction of both oxidant stress and the fibrotic effects on SSc induced by EGCG is associated with the intracellular ROS, ERK1/2 kinase signalling and NF- κ B activity⁸⁵.

In addition to the prominent antifibrotic effects, EGCG can inhibit the proliferation and invasion of tumors through reducing the HGF in CAFs, and suppress the metastasis of tumors through reducing VEGF as well⁴⁸, which indicated a potential role for EGCG in the treatment of desmoplastic tumors. Furthermore, it was reported that EGCG combined with luteolin could inhibit the TGF- β -induced CAFs. Gray et al.⁴⁹ found that both of EGCG and luteolin inhibited fibronectin expression which was induced by TGF- β , and decreased RhoA activation which was found to be necessary for fibronectin expression induced by TGF- β . Besides, they found EGCG and luteolin could inhibit TGF- β -induced ECM contraction, thereby suppressing tumor cell invasion. Their study results imply that combining EGCG with luteolin in clinic can prevent cancer progression by targeting CAFs, besides the tumor cell itself.

2.8. Emodin

Emodin (1,3,8-trihydroxy-6-methylanthraquinone), an predominant active component extracted from the rhizomes of rhubarb, aloes and other plants, has diverse biological activities, including laxative, immunosuppressive, anti-fibrosis, anti-inflammatory and anti-cancer effects. The anti-fibrosis effect of emodin is found in pulmonary⁸⁶, pancreatic⁸⁷ and liver fibrosis⁸⁸. Emodin is able to downregulate the activity of myeloperoxidase which is a marker for neutrophil influx into tissue, inhibit the expression of TGF- β 1 and collagen I, as well as suppress the cell proliferation in lung fibroblasts to weaken pulmonary fibrosis induced by bleomycin⁸⁶. Guan et al.⁸⁹ demonstrated that emodin was capacity of blocking pro-fibrotic signalings which were activated by TGF- β 1 in pulmonary fibroblasts, therefore inhibiting myofibroblast differentiation and ECM deposition. Their results also provide in vivo evidence for emodin to significantly inhibit bleomycin-induced lung inflammation and fibrosis.

Emodin has also been reported to possess the anti-cancer activity, such as inducing apoptosis in human lung adenocarcinoma cells⁹⁰. Studies have demonstrated that emodin inhibits migration and invasion of human breast cancer cells through downregulating MMP-2 and MMP-991. In addition, emodin represses cell migration and invasion by regulating EMT-related genes in head and neck squamous cell carcinoma⁹², colorectal cancer⁹³ and ovarian cancer⁹⁴. What is more, emodin is also able to inhibit the migration-promoting capacity of CAFs in tumors through blocking the EMT programming. Hsu et al.⁵⁰ tested the effects of emodin on EMT programming induced by CCM or the medium of interface zone fibroblasts (INFs-CM) in triple negative breast cancer. They analyzed the mesenchymal-marker expression, such as vimentin, β -catenin and MMP-2, and found that the increase of these mesenchymal markers stimulated by CCM or INFs-CM was reversed by emodin. The results showed that emodin inhibited INFs-CM or CCM-induced EMT programming in BT20 breast cancer cells, illustrating that emodin is a promising candidate for triple negative breast cancer prevention.

2.9. Derivatives of artemisinin

Artemisinin is a chemical isolated from the leaves of Artemisia annua Linn., a member of the Artemisia family which has a history of more than 2000 years in traditional Chinese medicine⁹⁵. Belonging to the family of sesquiterpene trioxane lactone, artemisinin has been used as a leading antimalarial drug since the end of 1990s. Besides, artemisinin possesses the highest efficacy among all the antimalarial drugs at present⁹⁶. Currently, a series of artemisinin derivatives, such as artemether (ARM), artesunate (ARS) and dihydroartemisinin (DHA), have been synthesized and shown improved bioactivity or solubility. Besides their antimalarial effects, it has been reported that artemisinin and its derivatives are efficacious in the treatment of infection and inflammatory diseases⁹⁷. Over the past two decades, studies have showed the anti-cancer effect of artemisinin and its derivatives, disclosing that artemisinin and its derivatives might also be effective therapeutic drugs to treat cancer 51 .

Researches have been also performed to study the ability of artemisinin and its derivatives to inactivate CAFs. ARS and DHA are found to be able to suppress the activation of CAFs through inhibiting the TGF- β pathway, and moreover, suppress the breast cancer growth and metastasis induced by CAFs *in vivo*. Yao et al.⁹⁸ explored the effects of artemisinin derivatives to inactivate breast cancer CAFs. They found that both ARS and DHA were capable of inhibiting TGF- β signaling, reverting CAFs from activated state to inactivated state, and suppressing the growth and metastasis of breast cancer induced by CAFs in the orthotopic model. Their study illustrated that artemisinin derivatives could be potential therapeutic drugs for breast cancer treatment.

3. Natural products normalize ECM

Besides the effects on the CAF–cancer cell crosstalk, natural products can also directly affect the ECM. By mechanical remodeling the ECM, CAFs contribute to invasion of desmoplastic tumor²¹. Natural products can effectively degrade the ECM and improve tumor perfusion⁹⁹ while maintaining the tumor-restraining function of ECM with minimal toxicity.

3.1. Cyclopamine

Cyclopamine (CPA) is an isosteroid alkaloid isolated from natural plants including Veratrum californicum, cornlily, V. grandiflorum and Fritillaria pallidiflora Schrenk. CPA is an inhibitor of smoothened, a G protein-coupled receptor presenting on CAFs that activates hedgehog (Hh) signaling¹⁰⁰. Thus, CPA is a potential treatment for patients with Hh-overexpressing tumors, such as cholangiocarcinoma, uveal melanoma, osteosarcoma, pancreatic, breast, and colon cancers. The paracrine Hh signaling between cancer cells and CAFs is a key regulator in promoting CAF growth and maintenance in desmoplastic cancers. Blocking the Hh pathway not only suppresses the proliferation of tumor cells but also disrupts tumor ECM and facilitates the delivery of other chemotherapy drugs to the tumor nest¹⁰¹. In non-small cell lung cancer, the increase of Hh expression by cancer cells improved CAF survival and proliferation, while CPA decreased it⁵². Kim et al.⁵³ focused on the interaction of cholangiocarcinoma cells with stromal cells to investigate the role of Hh signaling on the growth of cholangiocarcinoma cells. In animal experiments, they found that CPA scarcely suppresses cholangiocarcinoma cell proliferation at the concentration of lower than 10 µmol/L, while, in contrast, CPA could inhibit the growth of Lx-2 cells (stromal cells) at the same concentration. And they found CPA attenuated tumor growth in the co-implant xenograft group, but not in the single implant xenograft group (Table 1)⁵³. Their results indicated that stromal cells render cholangiocarcinoma cells susceptible to necrosis by CPA. As a result, CPA is a promising candidate to pharmacologically inhibit the interaction between tumorassociated stroma and cancer cells, thus inhibiting cancer cell proliferation, invasion, migration and drug resistance.

Because CPA is insoluble in water and has high systemic toxicity, it cannot be administrated directly to humans. To reduce toxicity of CPA and to enhance blood circulation, bioavailability and effects on tumor-microenvironment modulation of CPA, researchers developed various methods to deliver CPA to the tumor site. Jiang et al.⁵⁴ developed CPA-loaded membrane-camouflaged PLGA nanoparticles to effectively deliver CPA to the pancreatic tumor site, disrupt tumor ECM, increase functional vessels, and

improve tumor perfusion. To enhance the response of PDAC to ionizing radiation (IR), Zhao et al.⁵⁵ combined CPA-loaded corecrosslinked polymeric micelles (M-CPA) with Cs-137 radiation to enhance the radiation cvtotoxicity of Cs-137. In their results, M-CPA treatment can decrease the number of CAFs, intimating that M-CPA may disrupt the tumor-associated stroma in vivo, thus relieving the hypoxia condition within the tumor microenvironment. Later, in order to further enhance the effects on PDAC, Zhao et al.⁵⁶ co-delivered CPA, and paclitaxel (PTX) with a polymeric micelle formulation (M-CPA/PTX) to simultaneously regulate PDAC stroma and suppress tumor growth. The M-CPA could effectively modulate tumor stroma by increasing blood perfusion, improving tissue hypoxia, reducing matrix stiffness while sustaining the ECM tumor-compression function. The results that M-CPA/PTX apparently extended rodent survival suggested that it was a promising strategy for PDAC therapy to use multifunctional nanoparticles to target stromal and tumor cells concurrently.

3.2. Celastrol

Celastrol, also known as a tripterine, is a member of triterpenoids purified from traditional Chinese medicine named *Trypterygium wilfordii* Hook F. and has been used to treat autoimmune and neurodegenerative diseases, such as lupus erythematosus¹⁰². Celastrol possesses the activities of immunosuppressive, antiinflammatory, antioxidant and anti-fibrosis. It has been reported that celastrol could attenuate liver fibrosis through inhibiting inflammation by activating AMPK-SIRT3 signaling¹⁰³, and alleviate renal fibrosis through upregulating the expression of cannabinoid receptor 2 which is an anti-fibrotic factor through inhibiting the activation of *Smad3* signaling pathway¹⁰⁴.

Besides, celastrol has attracted great attention for its potent anticancer effects in breast cancer¹⁰⁵, PC¹⁰⁶, and osteosarcoma¹⁰⁷, etc. It has been reported that celastrol can promote tumor cells apoptosis through regulating mitochondria signal pathways¹⁰⁸. Recently, scientists found that celastrol could increase the sensitivity of CAFs to mitoxantrone. Liu et al.⁵⁷ combined celastrol with mitoxantrone to treat desmoplastic melanoma, and found that celastrol could decrease the IC₅₀ of mitoxantrone in CAF cell lines. Besides, co-delivery of mitoxantrone and celastrol in aminoethylanisamide-polymerdisulfide bond nanoparticles could reduce the amount of collagen in the tumor microenvironment and increase drug delivery to the tumor cells (Fig. 4).

3.3. Quercetin

Quercetin (3,3',4',5,7-penta-hydroxyflavone) is a natural flavonoid commonly found in fruits and vegetables. It is also the most abundant dietary flavonoid, which is widely used to prevent and treat cardiovascular diseases and cancers¹⁰⁹. Quercetin regulates multiple biological signaling pathways, inducing apoptosis of cancer cells as well as inhibiting proliferation of cancer cells¹¹⁰. Quercetin has also been proved to have anticancer effects on different cell lines in numerous *in vitro* studies¹¹¹. The anticancer effects of quercetin are primarily attributed to its anti-oxidant activity¹¹².

Quercetin is able to down-regulate the CAF-induced cancer drug resistance of desmoplastic tumors through suppressing the WNT16 expression, which is a key factor that can contribute to chemotherapy resistance in malignant tumors. However, quercetin is difficult to dissolve in water and its bioavailability is low, which limit its application as a pharmaceutical. Therefore,



Figure 4 Effective therapy significantly improved anti-tumor response and remodeled suppressive tumor microenvironment. (A) NP distribution in tumor-bearing mice. Mice were i.v. injected with Cy5-loaded (3 µg/kg) NPs and measured by IVIS imaging 24 h post-injection, n = 3; (B) Region-of-interest intensities of fluorescence signals among tumor and organs, n = 3; (C) Pharmaco-distribution of mitoxantrone and celastrol within tumor were measured by LC–MS, n = 5; (D) Tumor inhibition study and tumor weight comparison. Arrows indicate days of drug injection. Dosage: for M + C group: ~2 mg/kg of celastrol per dose; for M + C NP group: ~160 µg/kg of celastrol per dose. Tumors were surgically removed from the hosts at endpoint of study, weighted and compared between groups, n = 10-12; (E) Cell apoptosis were measured by TUNEL staining and collagen morphology changes were measured by Masson's trichrome staining, scale bar indicates 300 µm, n = 3. (F) Flow cytometry analysis of immune functioning cells within tumor microenvironment, n = 3. *P < 0.05, **P < 0.01, ***P < 0.001 (Used with permission from Ref. 57. Copyright © 2018 American Chemical Society).

Hu et al.⁵⁸ developed a quercetin prodrug by phosphorylating quercetin hydroxyl groups. In order to promote drug delivery, they prepared a targeted lipid/calcium/phosphate nanoparticle preparation composed of the quercetin phosphate. They found that quercetin could significantly remodel the CAFs and collagen

content within the bladder cancer through a significant downregulation of WNT16 expression. They also investigated the combination efficacy of the quercetin phosphate nanoparticles with cisplatin nanoparticles in a UMUC3 bladder cancer xenograft model, and found the antitumor efficacy of cisplatin



Figure 5 Nanoparticle distribution and tumor microenvironment remodeling *via* LCP-QP. (A) Effects of different treatments on the inhibition of fibroblast growth and Masson's trichrome stain for collagen and quantification results expressed as the percentage of total cell number. (B) Effect of LCP-QP on the penetration of DiI NPs and quantification of fluorescence signal (DiI labeled red) expressed as the percentage of cell number (DAPI signal) detected on frozen tumor sections. GFP positive fibroblasts (green), DAPI labeled nuclei (blue), and DiI labeled LCP-QP particles (red). **P < 0.01, *P < 0.05, n = 5. LCP, lipid calcium phosphate; QP, quercetin phosphate. (Used with permission from Ref. 58. Copyright © 2017 American Chemical Society.).

nanoparticles was improved by the quercetin phosphate nanoparticles (Fig. 5). Besides, in fibroblast-MCF7 co-cultures, studies found that treatment with quercetin can rescue Caveolin-1 expression which is related to early tumor recurrence, thus reversing the CAFs phenotypes⁵⁹.

3.4. Nab-PTX

Paclitaxel (PTX) is a taxane diterpenoid which was first isolated in 1971 from the Pacific yew and approved for medical usage in 1993. PTX has high anti-tumor activity and is widely used to treat ovarian cancer, breast cancer, non-small cell lung cancer, cervical cancer and brain cancer¹¹³.

Secreted protein acidic and rich in cysteine (SPARC) is an albumin-binding 42-kDa matricellular glycoprotein, overexpressed by CAFs in different types of tumor such as breast tumor, lung tumor, PDAC and melanoma¹¹⁴. SPARC has been shown to involve in proliferation, migration, and escape mechanisms of PDAC cells, and inversely correlate with survival in PDAC. Therefore, SPARC is gaining significant clinical interest as a potential biomarker. SPARC is expressed both in PDAC stroma and tumor cells, leading to the hypothesis that it may assist the delivery to the tumor of albumin-bound therapeutics. A drug formulation of PTX bound to the albumin (nab-PTX, Abraxane) is effective in depleting desmoplastic tumor stroma such as CAFs and ECM through binding albumins of nab-PTX to SPARC¹¹⁵ Study found that the combination of nab-PTX and GEM decreased CAF content and remodeled the ECM content, and further experiment confirmed that the stromal remodeling effects, such as alteration of collagen architecture and elimination of CAFs, are due to nab-PTX but not GEM¹¹⁶. In addition, the combination effect of GEM plus nab-PTX to remodel CAFs was greater than the combination of GEM plus tegafur¹¹⁷. These findings propose a potential role for nab-PTX in suppressing chemoresistance and metastasis by altering the tumor microenvironment.



Figure 6 Chemical structures of natural products mentioned in this review.

3.5. Epigallocatechin-3-gallate

Besides the effects of EGCG on HGF and VEGF and on CAFcancer crosstalk, Gray et al.⁴⁹ found that both of EGCG and luteolin inhibited fibronectin expression, and decreased RhoA activation which was disclosed to be essential for the expression of fibronectin induced by TGF- β . Furthermore, they found EGCG and luteolin could inhibit TGF- β -induced ECM contraction, thereby suppressing tumor cell invasion. Their study results imply that combining EGCG with luteolin in clinic can prevent or even reverse cancer progression by normalizing ECM through targeting CAFs.

4. Natural products inhibit the angiogenesis

Angiogenesis is a prerequisite for the growth and metastasis of tumors since tumors cannot maintain expansion without neovascularization to supply oxygen and nutrients. CAFs play a critical role in the construction of microenvironment to favor tumor angiogenesis through producing multiple regulatory molecules and ECM proteins, therefore promoting angiogenesis to meet the growth requirements of tumors.

Anti-angiogenesis therapy reduces blood supply and starves tumor cells of oxygen and nutrients. In clinic, VEGF-mediated signaling is one of the most promising anti-angiogenic therapeutic targets. While, anti-VEGF agents that are currently in use are mainly monoclonal antibodies, which have many serious adverse effects, such as bevacizumab. On the contrary, compared with the currently used synthetic medicines, natural products can suppress angiogenesis through multiple signal pathways in tumors with low systemic side effects. It has been reported that fraxinellone could suppress angiogenesis of tumors *in vivo* by inhibiting programmed cell death-ligand 1 expression *via* reducing HIF-1 and STAT3 signaling pathways³⁸. Silibinin and quercetin have been reported to inhibit the angiogenesis and proliferation of cancer cells in desmoplastic tumors through inhibiting VEGF expression^{37,118}.

Collectively, as an important complement to therapies targeted against cancer cells in desmoplastic tumors, various natural products that affect the CAF signals and effectors in the stroma have attracted scientists' attentions.

The pharmacological effects of natural products on desmoplastic tumors mainly reflect in three aspects: regulating ECM, inhibiting tumor angiogenesis, and influencing the interaction between tumor cells and CAFs. The current studies show that, through influencing the communication between CAFs and tumor cells, the proliferation, migration, invasion and drug resistance of tumor cells induced by CAFs can be significantly inhibited. On the one hand, CAFs can be depleted; on the other hand, CAFs can be reverted from the activated state into a quiescent state. However, direct CAF depletion could enhance hypoxia in the tumors and thus induce EMT in the desmoplastic tumor cells, which ultimately led to aggressive cancer progression¹¹⁹. Therefore, attempts to normalize CAFs could be a better way to provide new opportunities for the development of novel anti-desmoplastic cancer therapies. Natural products such as silibinin, DHA, artesunate, fraxinellone, and triptolide can effectively suppress the activation of CAFs induced by TGF- β , impel the normalization of CAFs, and then inhibit the development of desmoplastic tumors without the negative effects caused by the direct depletion of CAFs.

In addition, the current researches indicate that the effects of natural products are usually on multi-channels and multi-targets. They can affect the interaction between tumor cells and CAFs, while regulating the ECM or inhibiting tumor neovascularization, thus enhance the therapeutic effects for the desmoplastic tumor. For example, fraxinellone and silibinin can not only inhibit the activation of CAFs, but also inhibit tumor angiogenesis by inhibiting HIF-1 and VEGF, respectively; EGCG can suppress the CAF-induced proliferation and migration of tumor cells by downregulating the expressions of HGF and VEGF, and it can also remodel ECM through inhibiting the secretion of collagen induced by TGF- β ; triptolide can inhibit the activation of CAFs, while regulating ECM through inhibiting the secretion of collagen; quercetin can inhibit the activation of CAFs by promoting the expression of CAV-1, hinder tumor neovascularization by suppressing the secretion of VEGF, and inhibit CAF-induced drug resistance of tumor cells by down-regulating the expression of WNT16 (Fig. 2). From these natural products, we should be clearly aware that the pharmacological research of natural products should not be limited to a certain pathway or a target. Researchers should conduct extensive researches on the pharmacological mechanism of a natural product in multiple ways and directions.

A brief summary of the above mentioned natural products that potentially target CAFs is given in Fig. 6 and Table 1. Natural compounds that modify CAF signaling are waiting for further mechanistic and functional investigation.

5. Concluding remarks

The proliferation, migration and invasion characteristics of tumor cells must rely on their microenvironment. In the tumor microenvironment, fibroblasts as one important component of the dense desmoplastic stroma¹²⁰, contribute to the aggressiveness and chemotherapeutic resistance of desmoplastic tumor. As a consequence, it has been recognized that CAFs are attractive targets to reduce chemotherapy resistance for anticancer therapy and tumor recurrence over the past few years¹². Besides inhibiting the proliferation of tumor cells, many natural products also have the ability to target different types of stromal cells, by which exhibiting an indirect inhibitory effect on the invasion, proliferation and migration of desmoplastic tumors. These natural products usually modulate tumor microenvironment through various signal pathways, playing a comprehensive coordinating role in tumor treatment.

However, most of the natural products whose stability are poor are difficult to dissolve in water or general organic solvents, resulting in low bioavailability *in vivo*, which limits their pharmacological studies by researchers and their clinical application. Therefore, it is of great significance to develop rational drug delivery systems¹²¹ to enhance the bioavailability of natural products for their preclinical studies. In the study of anti-desmoplastic tumors, how to target natural products to tumor tissues to regulate CAFs should also be the focus of the researchers. Although there were a few studies which could deliver natural compounds to the tumor sites through using targeted drug delivery systems to load the natural product components, most studies of these formulations stayed at the pharmacodynamics level and the mechanisms were not widely explored. In addition, in the aspect of exploring the pharmacological mechanism of natural products for remodeling CAFs to inhibit tumor progression, the current researches mainly focused on TGF- β pathway. Therefore, the effect of natural products on other pathways such as WNT and Hh for CAF regulation should have potential values to be explored. Besides, there are still some important questions that should be further elucidated, such as when is the best time to deplete or remodel the CAFs by natural products in the tumor microenvironment? Are there other aspects of CAFs related to cancer progression which can be affected by chemicals from natural products? Are there any relations among the different remodeling aspects of natural products? And how can we better utilize natural products to assist current clinical treatments for desmoplastic tumors?

Furthermore, a combination of anti-cancer chemicals and natural product compounds is able to provide promising advantages in sensitizing monotherapy efficacy and overcoming druginduced resistance in desmoplastic tumor patients. The combination effects seen in these preclinical observations highlight the future application of such combinations for effective desmoplastic cancers treatments and reveal their clinical potential.

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Author contributions

Rujing Chen wrote the manuscript and made the figures; Kaili Hu and Leaf Huang supervised and edited the manuscript. All of the authors have read and approved the final manuscript.

Conflicts of interest

The authors have no conflicts of interest to declare.

References

- 1. Jain RK. Normalizing tumor microenvironment to treat cancer: bench to bedside to biomarkers. *J Clin Oncol* 2013;**31**:2205–18.
- Primac I, Maquoi E, Blacher S, Heljasvaara R, van Deun J, Smeland HYH, et al. Stromal integrin α11 regulates PDGFRβ signaling and promotes breast cancer progression. *J Clin Invest* 2019; 129:4609-28.
- Maloney E, Dufort CC, Provenzano PP, Farr N, Carlson MA, Vohra R, et al. Non-invasive monitoring of stromal biophysics with targeted depletion of hyaluronan in pancreatic ductal adenocarcinoma. *Cancers (Basel)* 2019;11:772.

- Loessner D, Holzapfel BM, Clements JA. Engineered microenvironments provide new insights into ovarian and prostate cancer progression and drug responses. *Adv Drug Deliv Rev* 2014;**79**: 193–213.
- Cadamuro M, Brivio S, Mertens J, Vismara M, Moncsek A, Milani C, et al. Platelet-derived growth factor-D enables liver myofibroblasts to promote tumor lymphangiogenesis in cholangiocarcinoma. *J Hepatol* 2019;**70**:700–9.
- Høgdall D, Lewinska M, Andersen JB. Desmoplastic tumor microenvironment and immunotherapy in cholangiocarcinoma. *Trends Cancer* 2018;4:239–55.
- 7. Stylianopoulos T. The solid mechanics of cancer and strategies for improved therapy. *J Biomech Eng* 2017;**139**:10.
- Yang S, Gao H. Nanoparticles for modulating tumor microenvironment to improve drug delivery and tumor therapy. *Pharmacol Res* 2017;**126**:97–108.
- 9. Chen X, Song E. Turning foes to friends: targeting cancer-associated fibroblasts. *Nat Rev Drug Discov* 2019;18:99–115.
- 10. Kalluri R. The biology and function of fibroblasts in cancer. *Nat Rev Cancer* 2016;**16**:582–98.
- 11. De Palma M, Biziato D, Petrova TV. Microenvironmental regulation of tumour angiogenesis. *Nat Rev Cancer* 2017;**17**:457–74.
- Affo S, Yu LX, Schwabe RF. The role of cancer-associated fibroblasts and fibrosisin liver cancer. *Annu Rev Pathol Mech Dis* 2017;12: 153–86.
- Jain RK. Delivery of molecular and cellular medicine to solid tumors. Adv Drug Deliv Rev 2012;64:353–65.
- 14. Sui H, Zhao J, Zhou L, Wen H, Deng W, Li C, et al. Tanshinone IIA inhibits β-catenin/VEGF-mediated angiogenesis by targeting TGFβ1 in normoxic and HIF-1α in hypoxic microenvironments in human colorectal cancer. *Cancer Lett* 2017;403:86–97.
- 15. Kesanakurti D, Chetty C, Dinh DH, Gujrati M, Rao JS. Role of MMP-2 in the regulation of IL-6/Stat3 survival signaling *via* interaction with $\alpha 5\beta 1$ integrin in glioma. *Oncogene* 2013;**32**:327–40.
- 16. Komiya E, Sato H, Watanabe N, Ise M, Higashi S, Miyagi Y, et al. Angiomodulin, a marker of cancer vasculature, is upregulated by vascular endothelial growth factor and increases vascular permeability as a ligand of integrin $\alpha\nu\beta$ 3. *Cancer Med* 2014;**3**:537–49.
- Wu X, Chen X, Zhou Q, Li P, Yu B, Li J, et al. Hepatocyte growth factor activates tumor stromal fibroblasts to promote tumorigenesis in gastric cancer. *Cancer Lett* 2013;335:128–35.
- 18. Kojima Y, Acar A, Eaton EN, Mellody KT, Scheel C, Ben-Porath I, et al. Autocrine TGF- β and stromal cell-derived factor-1 (SDF-1) signaling drives the evolution of tumor-promoting mammary stromal myofibroblasts. *Proc Natl Acad Sci U S A* 2010;**107**:20009–14.
- 19. De Francesco EM, Sims AH, Maggiolini M, Sotgia F, Lisanti MP, Clarke RB. GPER mediates the angiocrine actions induced by IGF1 through the HIF-1α/VEGF pathway in the breast tumor microenvironment. *Breast Cancer Res Treat* 2017;19:129.
- 20. Shimoda M, Principe S, Jackson HW, Luga V, Fang H, Molyneux SD, et al. Loss of the TIMP gene family is sufficient for the acquisition of the CAF-like cell state. *Nat Cell Biol* 2014;16: 889–901.
- 21. Insua-Rodríguez J, Oskarsson T. The extracellular matrix in breast cancer. *Adv Drug Deliv Rev* 2016;**97**:41–55.
- 22. Izumi D, Toden S, Ureta E, Ishimoto T, Baba H, Goel A. TIAM1 promotes chemoresistance and tumor invasiveness in colorectal cancer. *Cell Death Dis* 2019;**10**:267.
- 23. Ligorio M, Sil S, Malagon-Lopez J, Nieman LT, Misale S, Di Pilato M, et al. Stromal microenvironment shapes the intratumoral architecture of pancreatic cancer. *Cell* 2019;**178**:160–75.
- 24. Zhang L, Su H, Liu Y, Pang N, Li J, Qi XR. Enhancing solid tumor therapy with sequential delivery of dexamethasone and docetaxel engineered in a single carrier to overcome stromal resistance to drug delivery. *J Control Release* 2019;**294**:1–16.
- 25. Ji T, Lang J, Wang J, Cai R, Zhang Y, Qi F, et al. Designing liposomes to suppress extracellular matrix expression to enhance drug

penetration and pancreatic tumor therapy. ACS Nano 2017;11: 8668-78.

- Park SA, Surh YJ. Modulation of tumor microenvironment by chemopreventive natural products. Ann N Y Acad Sci 2017;1401:65–74.
- Li H, Sureda A, Devkota HP, Pittalà V, Barreca D, Silva AS, et al. Curcumin, the golden spice in treating cardiovascular diseases. *Biotechnol Adv* 2019;174:1325–48.
- Goel A, Aggarwal BB. Curcumin, the golden spice from Indian saffron, is a chemosensitizer and radiosensitizer for tumors and chemoprotector and radioprotector for normal organs. *Nutr Cancer* 2010;62:919–30.
- 29. Du Y, Long Q, Zhang L, Shi Y, Liu X, Li X, et al. Curcumin inhibits cancer-associated fibroblast-driven prostate cancer invasion through MAOA/mTOR/HIF-1α signaling. *Int J Oncol* 2015;47:2064–72.
- 30. Wang Q, Qu C, Xie F, Chen L, Liu L, Liang X, et al. Curcumin suppresses epithelial-to-mesenchymal transition and metastasis of pancreatic cancer cells by inhibiting cancer-associated fibroblasts. *Am J Cancer Res* 2017;7:125–33.
- Hendrayani SF, Al-Khalaf HH, Aboussekhra A. Curcumin triggers p16-dependent senescence in active breast cancer-associated fibroblasts and suppresses their paracrine procarcinogenic effects. *Neoplasia* 2013;15:631–40.
- Al-Ansari MM, Aboussekhra A. MiR-146b-5p mediates p16dependent repression of IL-6 and suppresses paracrine procarcinogenic effects of breast stromal fibroblasts. *Oncotarget* 2015;6: 30006–16.
- 33. Buhrmann C, Kraehe P, Lueders C, Shayan P, Goel A, Shakibaei M. Curcumin suppresses crosstalk between colon cancer stem cells and stromal fibroblasts in the tumor microenvironment: potential role of EMT. *PLoS One* 2014;9:e107514.
- 34. Ting HJ, Deep G, Jain AK, Cimic A, Sirintrapun J, Romero LM, et al. Silibinin prevents prostate cancer cell-mediated differentiation of naïve fibroblasts into cancer-associated fibroblast phenotype by targeting TGF β2. *Mol Carcinog* 2015;54:730–41.
- 35. Ting H, Deep G, Kumar S, Jain AK, Agarwal C, Agarwal R. Beneficial effects of the naturally occurring flavonoid silibinin on the prostate cancer microenvironment: role of monocyte chemotactic protein-1 and immune cell recruitment. *Carcinogenesis* 2016;37: 589–99.
- 36. Deep G, Kumar R, Jain AK, Agarwal C, Agarwal R. Silibinin inhibits fibronectin induced motility, invasiveness and survival in human prostate carcinoma PC3 cells *via* targeting integrin signaling. *Mutat Res* 2014;768:35–46.
- 37. Deep G, Kumar R, Nambiar DK, Jain AK, Ramteke AM, Serkova NJ, et al. Silibinin inhibits hypoxia-induced HIF-1α-mediated signaling, angiogenesis and lipogenesis in prostate cancer cells: *in vitro* evidence and *in vivo* functional imaging and metabolomics. *Mol Carcinog* 2017;**56**:833–48.
- 38. Xing Y, Mi C, Wang Z, Zhang ZH, Li MY, Zuo HX, et al. Fraxinellone has anticancer activity *in vivo* by inhibiting programmed cell death-ligand 1 expression by reducing hypoxia-inducible factor-1α and STAT3. *Pharmacol Res* 2018;135:166–80.
- 39. Hou L, Liu Q, Shen L, Liu Y, Zhang X, Chen F, et al. Nano-delivery of fraxinellone remodels tumor microenvironment and facilitates therapeutic vaccination in desmoplastic melanoma. *Theranostics* 2018;8:3781–96.
- 40. Pei Y, Chen L, Huang Y, Wang J, Feng J, Xu M, et al. Sequential targeting TGF-β signaling and KRAS mutation increases therapeutic efficacy in pancreatic cancer. *Small* 2019;**15**:e1900631.
- 41. Dauer P, Zhao X, Gupta VK, Sharma N, Kesh K, Gnamlin P, et al. Inactivation of cancer-associated-fibroblasts disrupts oncogenic signaling in pancreatic cancer cells and promotes its regression. *Cancer Res* 2018;78:1321–33.
- 42. Banerjee S, Modi S, McGinn O, Zhao X, Dudeja V, Ramakrishnan S, et al. Impaired synthesis of stromal components in response to minnelide improves vascular function, drug delivery, and survival in pancreatic cancer. *Clin Cancer Res* 2016;**22**:415–25.

- 43. Wang Z, Ma D, Wang C, Zhu Z, Yang Y, Zeng F, et al. Triptonide inhibits the pathological functions of gastric cancer-associated fibroblasts. *Biomed Pharmacother* 2017;96:757–67.
- 44. Wang ZF, Ma DG, Zhu Z, Mu YP, Yang YY, Feng L, et al. Astragaloside inhibits pathological functions of gastric cancer-associated fibroblasts. *World J Gastroenterol* 2017;**23**:8512–25.
- 45. Thongchot S, Ferraresi A, Vidoni C, Loilome W, Yongvanit P, Namwat N, et al. Resveratrol interrupts the pro-invasive communication between cancer associated fibroblasts and cholangiocarcinoma cells. *Cancer Lett* 2018;430:160–71.
- 46. Suh J, Kim DH, Surh YJ. Resveratrol suppresses migration, invasion and stemness of human breast cancer cells by interfering with tumorstromal cross-talk. Arch Biochem Biophys 2018;643:62–71.
- 47. Chen C Te, Chen YC, Du Y, Han Z, Ying H, Bouchard RR, et al. A tumor vessel-targeting fusion protein elicits a chemotherapeutic bystander effect in pancreatic ductal adenocarcinoma. *Am J Cancer Res* 2017;7:657–72.
- 48. Bigelow RLH, Cardelli JA. The green tea catechins, (-)-epigallocatechin-3-gallate (EGCG) and (-)-epicatechin-3-gallate (ECG), inhibit HGF/Met signaling in immortalized and tumorigenic breast epithelial cells. *Oncogene* 2006;25:1922–30.
- 49. Gray AL, Stephens CA, Bigelow RLH, Coleman DT, Cardelli JA. The polyphenols (–)-epigallocatechin-3-gallate and luteolin synergistically inhibit TGF-β-induced myofibroblast phenotypes through rhoa and ERK inhibition. *PLoS One* 2014;9:e109208.
- 50. Hsu HC, Liu LC, Wang HY, Hung CM, Lin YC, Ho CT, et al. Stromal fibroblasts from the interface zone of triple negative breast carcinomas induced epithelial-mesenchymal transition and its inhibition by emodin. *PLoS One* 2017;**12**:e0164661.
- Wang J, Zhang J, Shi Y, Xu C, Zhang C, Wong YK, et al. Mechanistic investigation of the specific anticancer property of artemisinin and its combination with aminolevulinic acid for enhanced anticolorectal cancer activity. ACS Cent Sci 2017;3:743–50.
- Bermudez O, Hennen E, Koch I, Lindner M, Eickelberg O. *Gli1* mediates lung cancer cell proliferation and sonic hedgehogdependent mesenchymal cell activation. *PLoS One* 2013;8:e63226.
- Kim Y, Kim MO, Shin JS, Park SH, Kim SB, Kim J, et al. Hedgehog signaling between cancer cells and hepatic stellate cells in promoting cholangiocarcinoma. *Ann Surg Oncol* 2014;21:2684–98.
- 54. Jiang T, Zhang B, Zhang L, Wu X, Li H, Shen S, et al. Biomimetic nanoparticles delivered hedgehog pathway inhibitor to modify tumour microenvironment and improved chemotherapy for pancreatic carcinoma. *Artif Cells Nanomedicine Biotechnol* 2018;46:1088–101.
- Zhao J, Wu C, Abbruzzese J, Hwang RF, Li C. Cyclopamine-loaded core-cross-linked polymeric micelles enhance radiation response in pancreatic cancer and pancreatic stellate cells. *Mol Pharm* 2015;12: 2093–100.
- 56. Zhao J, Wang H, Hsiao CH, Chow DSL, Koay EJ, Kang Y, et al. Simultaneous inhibition of hedgehog signaling and tumor proliferation remodels stroma and enhances pancreatic cancer therapy. *Biomaterials* 2018;159:215–28.
- Liu Q, Chen F, Hou L, Shen L, Zhang X, Wang D, et al. Nanocarriermediated chemo-immunotherapy arrested cancer progression and induced tumor dormancy in desmoplastic melanoma. ACS Nano 2018;12:7812–25.
- Hu K, Miao L, Goodwin TJ, Li J, Liu Q, Huang L. Quercetin remodels the tumor microenvironment to improve the permeation, retention, and antitumor effects of nanoparticles. *ACS Nano* 2017;11: 4916–25.
- 59. Martinez-Outschoorn UE, Balliet RM, Rivadeneira DB, Chiavarina B, Pavlides S, Wang C, et al. Oxidative stress in cancer associated fibroblasts drives tumor-stroma co-evolution: a new paradigm for understanding tumor metabolism, the field effect and genomic instability in cancer cells. *Cell Cycle* 2010;9:3256–76.
- Pradhan SC, Girish C. Hepatoprotective herbal drug, silymarin from experimental pharmacology to clinical medicine. *Indian J Med Res* 2013;137:491–504.

- 61. Singh RP, Raina K, Deep G, Chan D, Agarwal R. Silibinin suppresses growth of human prostate carcinoma PC-3 orthotopic xenograft via activation of extracellular signal-regulated kinase 1/2 and inhibition of signal transducers and activators of transcription signaling. *Clin Cancer Res* 2009;15:613–21.
- **62**. Deep G, Agarwal R. Targeting tumor microenvironment with silibinin: promise and potential for a translational cancer chemopreventive strategy. *Curr Cancer Drug Targets* 2013;**13**:486–99.
- Wu X, Wu X, Ma Y, Shao F, Tan Y, Tan T, et al. CUG-binding protein 1 regulates HSC activation and liver fibrogenesis. *Nat Commun* 2016; 7:13498.
- 64. An Y, Zhang W, Guo S, Gao T, Li C. 308 Triptolide, triptonide and celastrol alleviates the proliferation and IFN-γ-induced immune dysfunction of keratinocytes by inhibiting miR-17-92. *J Invest Dermatol* 2016;136:S213.
- **65.** Ren YX, Zhou R, Tang W, Wang WH, Li YC, Yang YF, et al. (5*R*)-5-Hydroxytriptolide (LLDT-8) protects against bleomycin-induced lung fibrosis in mice. *Acta Pharmacol Sin* 2007;**28**:518–25.
- 66. Chong LW, Hsu YC, Chiu YT, Yang KC, Huang YT. Antifibrotic effects of triptolide on hepatic stellate cells and dimethylnitrosamineintoxicated rats. *Phyther Res* 2011;25:990–9.
- **67.** Yuan XP, He XS, Wang CX, Liu LS, Fu Q. Triptolide attenuates renal interstitial fibrosis in rats with unilateral ureteral obstruction. *Nephrology* 2011;**16**:200–10.
- Noel P, Von Hoff DD, Saluja AK, Velagapudi M, Borazanci E, Han H. Triptolide and its derivatives as cancer therapies. *Trends Pharmacol Sci* 2019;40:327–41.
- **69**. Jung Y, Jerng U, Lee S. A systematic review of anticancer effects of Radix Astragali. *Chin J Integr Med* 2016;**22**:225–36.
- Li L, Hou X, Xu R, Liu C, Tu M. Research review on the pharmacological effects of astragaloside IV. *Fundam Clin Pharmacol* 2017; 31:17–36.
- 71. Li X, Wang X, Han C, Xing G, Zhou L, Li G, et al. Astragaloside IV suppresses collagen production of activated hepatic stellate cells *via* oxidative stress-mediated p38 MAPK pathway. *Free Radic Biol Med* 2013;60:168–76.
- 72. Zhou X, Sun X, Gong X, Yang Y, Chen C, Shan G, et al. Astragaloside IV from *Astragalus membranaceus* ameliorates renal interstitial fibrosis by inhibiting inflammation *via* TLR4/NF-κ*B in vivo* and *in vitro*. Int Immunopharm 2017;42:18–24.
- 73. Chen P, Xie Y, Shen E, Li GG, Yu Y, Zhang CB, et al. Astragaloside IV attenuates myocardial fibrosis by inhibiting TGF- β 1 signaling in coxsackievirus B3-induced cardiomyopathy. *Eur J Pharmacol* 2011; **658**:168–74.
- 74. Qian W, Cai X, Qian Q, Zhang W, Wang D. Astragaloside IV modulates TGF-β1-dependent epithelial-mesenchymal transition in bleomycin-induced pulmonary fibrosis. J Cell Mol Med 2018;22: 4354–65.
- 75. Xu W, Shao X, Tian L, Gu L, Zhang M, Wang Q, et al. Astragaloside IV ameliorates renal fibrosis *via* the inhibition of mitogen-activated protein kinases and antiapoptosis *in vivo* and *in vitro*. *J Pharmacol Exp Therapeut* 2014;**350**:552–62.
- 76. Che X, Wang Q, Xie Y, Xu W, Shao X, Mou S, et al. Astragaloside IV suppresses transforming growth factor-β1 induced fibrosis of cultured mouse renal fibroblasts *via* inhibition of the MAPK and NFκB signaling pathways. *Biochem Biophys Res Commun* 2015;464: 1260–6.
- Richard JL. Coronary risk factors. The French paradox. Arch Mal Coeur Vaiss 1987;80:17–21.
- Baur JA, Sinclair DA. Therapeutic potential of resveratrol: the *in vivo* evidence. *Nat Rev Drug Discov* 2006;5:493–506.
- 79. Jang M, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CWW, et al. Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science* 1997;275:218–20.
- Singh AP, Singh R, Verma SS, Rai V, Kaschula CH, Maiti P, et al. Health benefits of resveratrol: evidence from clinical studies. *Med Res Rev* 2019;39:1851–91.

- Chen C Te, Yamaguchi H, Lee HJ, Du Y, Lee HH, Xia W, et al. Dual targeting of tumor angiogenesis and chemotherapy by endostatincytosine deaminase-uracil phosphoribosyltransferase. *Mol Cancer Ther* 2011;10:1327–36.
- Surh YJ. Cancer chemoprevention with dietary phytochemicals. *Nat Rev Cancer* 2003;3:768–80.
- 83. Lv Y, Chen L, Wu H, Xu X, Zhou G, Zhu B, et al. (–)-Epigallocatechin-3-gallate-mediated formation of myofibrillar protein emulsion gels under malondialdehyde-induced oxidative stress. *Food Chem* 2019;285:139–46.
- Rice-Evans C. Implications of the mechanisms of action of tea polyphenols as antioxidants *in vitro* for chemoprevention in humans. *Proc Soc Exp Biol Med* 1999;220:262–6.
- 85. Dooley A, Shi-Wen X, Aden N, Tranah T, Desai N, Denton CP, et al. Modulation of collagen type I, fibronectin and dermal fibroblast function and activity, in systemic sclerosis by the antioxidant epigallocatechin-3-gallate. *Rheumatology* 2010;49:2024–36.
- Chen XH, Sun RS, Hu JM, Mo ZY, Yang ZF, Jin GY, et al. Inhibitory effect of emodin on bleomycin-induced pulmonary fibrosis in mice. *Clin Exp Pharmacol Physiol* 2009;36:146–53.
- Wang CH, Gao ZQ, Ye B, Cai JT, Xie CG, Qian K Da, et al. Effect of emodin on pancreatic fibrosis in rats. *World J Gastroenterol* 2007;13: 378–82.
- Zhan Y, Wei H, Wang Z, Huang X, Xu Q, Li D, et al. Effects of emodin on hepatic fibrosis in rats. *Chin J Hepatol* 2001;9:235–6.
- 89. Guan R, Zhao X, Wang X, Song N, Guo Y, Yan X, et al. Emodin alleviates bleomycin-induced pulmonary fibrosis in rats. *Toxicol Lett* 2016;262:161–72.
- 90. Su YT, Chang HL, Shyue SK, Hsu SL. Emodin induces apoptosis in human lung adenocarcinoma cells through a reactive oxygen speciesdependent mitochondrial signaling pathway. *Biochem Pharmacol* 2005;70:229–41.
- 91. Sun Y, Wang X, Zhou Q, Lu Y, Zhang H, Chen Q, et al. Inhibitory effect of emodin on migration, invasion and metastasis of human breast cancer MDA-MB-231 cells *in vitro* and *in vivo*. Oncol Rep 2015;33:338–46.
- 92. Way TD, Huang JT, Chou CH, Huang CH, Yang MH, Ho CT. Emodin represses TWIST1-induced epithelial-mesenchymal transitions in head and neck squamous cell carcinoma cells by inhibiting the β -catenin and Akt pathways. *Eur J Cancer* 2014;**50**:366–78.
- 93. Zou J, Luo H, Zeng Q, Dong Z, Wu D, Liu L. Protein kinase CK2α is overexpressed in colorectal cancer and modulates cell proliferation and invasion via regulating EMT-related genes. J Transl Med 2011;9:97.
- 94. Hu C, Dong T, Li R, Lu J, Wei X, Liu P. Emodin inhibits epithelial to mesenchymal transition in epithelial ovarian cancer cells by regulation of GSK-3/-catenin/ZEB1 signaling pathway. *Oncol Rep* 2016; 35:2027–34.
- Tu Y. Artemisinin—a gift from traditional Chinese medicine to the world (Nobel Lecture). Angew Chem Int Ed Engl 2016;55:10210–26.
- Miller LH, Su X. Artemisinin: discovery from the Chinese herbal garden. *Cell* 2011;146:855–8.
- An J, Minie M, Sasaki T, Woodward JJ, Elkon KB. Antimalarial drugs as immune modulators: new mechanisms for old drugs. *Annu Rev Med* 2017;68:317–30.
- 98. Yao Y, Guo Q, Cao Y, Qiu Y, Tan R, Yu Z, et al. Artemisinin derivatives inactivate cancer-associated fibroblasts through suppressing TGF-β signaling in breast cancer. J Exp Clin Cancer Res 2018;37:1–14.
- 99. Zhang B, Wang H, Jiang T, Jin K, Luo Z, Shi W, et al. Cyclopamine treatment disrupts extracellular matrix and alleviates solid stress to improve nanomedicine delivery for pancreatic cancer. *J Drug Target* 2018;26:913–9.
- 100. Chen JK. I only have eye for Ewe: the discovery of cyclopamine and development of hedgehog pathway-targeting drugs. *Nat Prod Rep* 2016;33:595–601.
- 101. Zhang B, Jiang T, Shen S, She X, Tuo Y, Hu Y, et al. Cyclopamine disrupts tumor extracellular matrix and improves the distribution and efficacy of nanotherapeutics in pancreatic cancer. *Biomaterials* 2016; 103:12–21.

- 102. Li J, Hao J. Treatment of neurodegenerative diseases with bioactive components of *tripterygium wilfordii*. Am J Chin Med 2019;47: 769–85.
- 103. Wang Y, Li C, Gu J, Chen C, Duanmu J, Miao J, et al. Celastrol exerts anti-inflammatory effect in liver fibrosis *via* activation of AMPK-SIRT3 signaling. *J Cell Mol Med* 2020;24:941–53.
- 104. Tang M, Cao X, Zhang K, Li Y, Zheng QY, Li GQ, et al. Celastrol alleviates renal fibrosis by upregulating cannabinoid receptor 2 expression article. *Cell Death Dis* 2018;9:601.
- 105. Zhao Y, Tan Y, Meng T, Liu X, Zhu Y, Hong Y, et al. Simultaneous targeting therapy for lung metastasis and breast tumor by blocking the NF-κB signaling pathway using celastrol-loaded micelles. *Drug Deliv* 2018;25:341–52.
- 106. Guo J, Huang X, Wang H, Yang H. Celastrol induces autophagy by targeting AR/miR-101 in prostate cancer cells. *PLoS One* 2015;10: e0140745.
- 107. Li HY, Zhang J, Sun LL, Li BH, Gao HL, Xie T, et al. Celastrol induces apoptosis and autophagy via the ROS/JNK signaling pathway in human osteosarcoma cells: an *in vitro* and *in vivo* study. *Cell Death Dis* 2015;6:e1604.
- 108. Tan Y, Zhu Y, Zhao Y, Wen L, Meng T, Liu X, et al. Mitochondrial alkaline pH-responsive drug release mediated by celastrol loaded glycolipid-like micelles for cancer therapy. *Biomaterials* 2018;154: 169–81.
- 109. Russo M, Spagnuolo C, Tedesco I, Bilotto S, Russo GL. The flavonoid quercetin in disease prevention and therapy: facts and fancies. *Biochem Pharmacol* 2012;83:6–15.
- Reyes-Farias M, Carrasco-Pozo C. The anti-cancer effect of quercetin: molecular implications in cancer metabolism. *Int J Mol Sci* 2019;20:3177.
- Dajas F. Life or death: neuroprotective and anticancer effects of quercetin. J Ethnopharmacol 2012;143:383–96.
- 112. Cossarizza A, Gibellini L, Pinti M, Nasi M, Montagna JP, De Biasi S, et al. Quercetin and cancer chemoprevention. *Evid Based Complement Altern Med* 2011;2011:591356.
- 113. Du X, Khan AR, Fu M, Ji J, Yu A, Zhai G. Current development in the formulations of non-injection administration of paclitaxel. *Int J Pharm* 2018;542:242–52.
- 114. Giordano G, Pancione M, Olivieri N, Parcesepe P, Velocci M, Di Raimo T, et al. Nano albumin bound-paclitaxel in pancreatic cancer: current evidences and future directions. *World J Gastroenterol* 2017; 23:5875–86.
- 115. Neesse A, Frese KK, Chan DS, Bapiro TE, Howat WJ, Richards FM, et al. SPARC independent drug delivery and antitumour effects of nab-paclitaxel in genetically engineered mice. *Gut* 2014;63:974–83.
- 116. Rajeshkumar NV, Yabuuchi S, Pai SG, Tong Z, Hou S, Bateman S, et al. Superior therapeutic efficacy of nab-paclitaxel over cremophorbased paclitaxel in locally advanced and metastatic models of human pancreatic cancer. *Br J Cancer* 2016;115:442–53.
- 117. Miyashita T, Tajima H, Makino I, Okazaki M, Yamaguchi T, Ohbatake Y, et al. Neoadjuvant chemotherapy with gemcitabine plus nab-paclitaxel reduces the number of cancer-associated fibroblasts through depletion of pancreatic stroma. *Anticancer Res* 2018;**38**: 337–43.
- 118. Balakrishnan S, Bhat FA, Raja Singh P, Mukherjee S, Elumalai P, Das S, et al. Gold nanoparticle—conjugated quercetin inhibits epithelial—mesenchymal transition, angiogenesis and invasiveness via EGFR/VEGFR-2-mediated pathway in breast cancer. *Cell Prolif* 2016;49:678–97.
- 119. Özdemir BC, Pentcheva-Hoang T, Carstens JL, Zheng X, Wu CC, Simpson TR, et al. Depletion of carcinoma-associated fibroblasts and fibrosis induces immunosuppression and accelerates pancreas cancer with reduced survival. *Cancer Cell* 2014;25:719–34.
- 120. Hamada S, Masamune A, Shimosegawa T. Novel therapeutic strategies targeting tumor-stromal interactions in pancreatic cancer. *Front Physiol* 2013;4:331.
- 121. Li C, Wang J, Wang Y, Gao H, Wei G, Huang Y, et al. Recent progress in drug delivery. *Acta Pharm Sin B* 2019;9:1145–62.