Research Article

Novel Design Strategy for Checkpoint Kinase 2 Inhibitors Using Pharmacophore Modeling, Combinatorial Fusion, and Virtual Screening

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Received 29 November 2013; Accepted 19 February 2014; Published 23 April 2014

Academic Editor: Che-Lun Hung

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Checkpoint kinase 2 (Chk2) has a great effect on DNA-damage and plays an important role in response to DNA double-strand breaks and related lesions. In this study, we will concentrate on Chk2 and the purpose is to find the potential inhibitors by the pharmacophore hypotheses (PhModels), combinatorial fusion, and virtual screening techniques. Applying combinatorial fusion into PhModels and virtual screening techniques is a novel design strategy for drug design. We used combinatorial fusion to analyze the prediction results and then obtained the best correlation coefficient of the testing set (r_{test}) with the value 0.816 by combining the Best_{train}Best_{test} and Fast_{train}Fast_{test} prediction results. The potential inhibitors were selected from NCI database by screening according to Best_{train}Fast_{test} + Fast_{train}Fast_{test} prediction results and molecular docking with CDOCKER docking program. Finally, the selected compounds have high interaction energy between a ligand and a receptor. Through these approaches, 23 potential inhibitors for Chk2 are retrieved for further study.

1. Introduction

DNA-damage is induced by ionizing radiation, genotoxic chemicals, or collapsed replication forks, and when DNA was damaged or the responses of cells were failure, the mutation associated with the breast or ovarian cancer of genes may occur. To prevent and repair the DNA-damage, mammalian cells will control and stabilize the genome by cell cycle checkpoint. The checkpoint pathway consists of several kinases, such as ataxia telangiectasia mutated protein (ATM [1, 2]), ataxia telangiectasia and Rad3-related protein (ATR [1, 2]), checkpoint kinase 1 (Chk1 [3, 4]), and checkpoint kinase 2 (Chk2 [5-8]). ATM and ATR are upstream kinases passing messages to downstream kinases and phosphorylating several proteins that initiate the activation of the DNAdamage checkpoint. Moreover, ATM is a primarily pathway to activate p53 (protein 53 [9]) by Chk2, and ATR may influence the phosphorylation of Chk1. Both Chk1 and Chk2 are key components in DNA-damage; however, their cellular

activities are different. Chk1 is involved in S and G2 phases of the cell cycle with ATR pathway. By contrast, Chk2 is activated in all phases through ATM-dependent pathway and plays an important role in response to DNA double-strand breaks and related lesions. Furthermore, Chk1 is an unstable protein and lacks the forkhead-associated domain (FHA) which was involved in several processes that protect against cancer and can be found in Chk2. Therefore, we concentrate on Chk2 in this study.

Chk2 is a protein containing 543 amino acid residues and the structure of Chk2 consists of some functional elements, including the N-terminal SQ/TQ cluster domain (SCD), FHA, and the N-terminal serine/threonine kinase domain (KD) [5–8]. The SCD is known to be the preferred site with the residue Thr68 for phosphorylation to respond to DNA-damage by ATM/ATP kinases. The FHA domain is a phosphopeptide recognition domain found in many regulatory proteins and thought to bind to the phosphoThr68 segment of SCD [5–8, 10–14]. Hence it is a good candidate

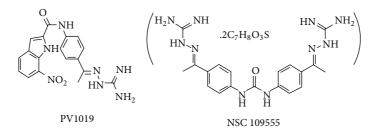


FIGURE 1: Two-dimensional chemical structures of known Chk2 inhibitors. The experimental IC_{50} of PV1019 and NSC 109555 were 138 nM and 240 nM, respectively.

for interactions of Chk2 with its upstream regulators or downstream targets in the cell-cycle-checkpoint signaling. The KD occupies almost the entire carboxy-terminal half of Chk2 and has been identified based on their homology with serine/threonine kinases. Some studies reported that when DNA was damaged, Chk2 is activated by ATM/ATR through the phosphorylation of residue Thr68. Moreover, Chk2 induces transautophosphorylation of residues Thr383 and Thr387 and then cis-phosphorylation of residue Ser516 [5-8, 10-14]. After that, Chk2 will phosphorylate several downstream substrates, such as BRCA1 (breast cancer 1, early onset [15, 16]), Cdc25A (cell division cycle 25 homolog A), Cdc25C, and p53 [7, 8, 10]. Several researches indicated that Chk2 phosphorylates Cdc25A which is considered an oncogene on the residue Ser123 in S phase of cell cycle, and it also phosphorylates Cdc25C on the residue Ser216 in G2 phase helping prevent mitotic entry in cells with damaged DNA [5]. Furthermore, BRCA1 and p53 are involved in DNA repair process in the breast or ovarian cancer. BRCA1 is a human caretaker gene and helps repair damaged DNA or destroys cells which cannot be repaired. The p53 is a tumor suppressor protein involved in preventing cancer in human and plays an important role in the G1 checkpoint in response to DNA damaging agents. We consider that the sites of the phosphorylations are important in the drug design for cell survival when DNA is damaged.

Recently, several studies identified the inhibitors of Chk2 [6-8, 10-14], and they also showed the crystal structures of Chk2 complex, such as PDB: 1GXC, 2W7X, and, and so forth. They are selective, reversible, and ATP-competitive Chk2 inhibitors demonstrating that they effectively restrain the radiation-induced phosphorylation of Chk2. In addition, several selective Chk2 inhibitors have been also identified (two examples were shown in Figure 1) and the researches indicated that they are potential and selective inhibitors of Chk2 with chemotherapeutic and radiosensitization potential. On structure-based drug design, several developments of Chk2 were published [17, 18]. Quantitative structureactivity relationship model (QSAR model) is a regression or classification model and is an important technique in the rational drug design. It is used to correlate the structure properties of compounds with their biological activities. The method to predict the quality by QSAR was improved by considering the three-dimensional structure QSAR (3D-QSAR) [19-24] of targeted inhibitor. Therefore, the compound structure can be directly optimized in the 3D space.

The comparative molecular field analyses (CoMFA) [18, 25– 30] and the comparative molecular similarity indices analyses (CoMSIA) [18, 27–32] for Chk2 inhibitors were performed by ligand-based and receptor-guided alignment. They used the cocrystal structure from protein data bank (PDB code: 2CN8) [7], and then they identified new plausible binding modes used as template for 3D-QSAR [18]. There is another research of Chk2 studied in QSAR/QSPR [17] providing structures that will improve reducing the side effects of Chk2 inhibitors.

Pharmacophore [20-24, 33-35] is a set of structural features responsible for the biological activity of a molecule. It allowed compounds with diverse structures to find the common chemical features by ligand pharmacophore mapping, and that is different from CoMFA and CoMSIA with the common structure constraint. Thus, pharmacophore can explain how diverse ligands bind to a receptor site by these features and visualize the feature of potential chemical interactions between ligands and receptors. Moreover, pharmacophore can easily and quickly identify candidate inhibitors for a target protein based on 3D query. Therefore, in this work, we first used 3D-QSAR study to build pharmacophore hypotheses (denoted as PhModels) for Chk2 inhibitors by HypoGen Best, Fast, and Caesar algorithms, respectively. Then we used the combinatorial fusion to select and combine prediction results for improving the predictive accuracy in biological activities of inhibitors. Virtual screening is a computational technique used in drug discovery research. There are two categories of screening techniques: ligandbased and structure-based. In this work, for ligand-based virtual screening, we used the selected PhModels as 3D structure query by pharmacophore hypothesis screening that each compound in National Cancer Institute (NCI) database will be mapped onto the pharmacophoric features of selected PhModels. When the chemical features of a compound fit the generated PhModels, it will be selected. All of feasible compounds in NCI database were selected in this work. Finally, the potential inhibitors were retrieved from selected compounds by using molecular docking program to predict the conformation and interaction energy between Chk2 and ligand. Applying combinatorial fusion into PhModels and virtual screening techniques is a novel design strategy for drug design and can help medicinal chemists to identify or design new Chk2 inhibitors. Besides, the potential inhibitors of Chk2 retrieved in this work can be estimated by biologists for further study.

2. Materials and Methods

2.1. Biological Data Collection. In order to construct the PhModels, at first, we collected the Chk2 inhibitors with two-dimensional structures and the biological activity values from the ChEMBL database [36]. Then, according to the structure variations and chemical differences in the kinase inhibitor activity, 158 known Chk2 inhibitors were selected and retrieved. The biological activity of 158 known Chk2 inhibitors was represented as IC_{50} (nanomolar, nM). There are 260,071 compounds from the NCI database (release version 3, http://cactus.nci.nih.gov/download/nci/) which were used in the database screening and molecular docking approach in this work.

2.2. Training and Testing Sets Selection. Before generating PhModels, we should divide the 158 Chk2 inhibitors into the training set and testing set, respectively. The rules used to select training set inhibitors are according to the following requirements as suggested by the Accelrys Discovery Studio. (1) All selected inhibitors should have clear and concise information including structure features and activity range. (2) At a minimum, 16 diverse inhibitors for training set were selected to ensure the statistical significance. (3) The training set should contain the most and the least active inhibitors. (4) The biological activities of the inhibitors spanned at least 4 orders of magnitude. Based on the above four rules, the 158 Chk2 inhibitors were divided, and the scatter diagram of training set and testing set inhibitors was shown in Figure 2. Figure 2 demonstrates the distribution of the inhibitors in the training set and testing set, and the representative points of the testing set are close to those of the training set. The training set with 25 inhibitors is used to construct PhModels, and the IC₅₀ values of these 25 inhibitors are ranged from 2.3 to 100,000 nM (Table 1). The testing set with remaining 133 inhibitors is used to test the predictive ability of generated PhModels, and the IC_{50} values of the 133 testing set inhibitors are ranged from 3.4 to 74,000 nM (Table 2). After selecting the training set and testing set inhibitors, we established PhModels at first, and then we used the correlation analysis to estimate the prediction abilities of PhModels.

2.3. Pharmacophore Generation. The workflow of PhModel generation for Chk2 inhibitors was shown in Figure 3. In this study, we used the HypoGen program [37] in Accelrys Discovery Studio 2.1 to generate PhModels. At the initial step, 3D conformations of the training set inhibitors were generated by using "3D-QSAR Pharmacophore Generation protocol" with the Best, Fast, and Caesar generating algorithms, respectively, based on the CHARMm-like force field. The conformational-space energy was constrained ≤20 kcal/mol which represented the maximum allowed energy above the global minimum energy. For each training set inhibitor, the number of the diverse 3D conformations was set to ≤255. All other parameters were set as default values. Following the above rules, the 3D conformations were generated, and then we can construct the PhModel by using "Ligand Pharmacophore Mapping protocol." Each of the ten PhModels using

HypoGen Best, Fast, and Caesar algorithms were generated in this study.

2.4. Combinatorial Fusion. In this study, we use a combinatorial fusion technique to facilitate prediction results selection and combination for improving predictive accuracy in biological activities of inhibitors. The combinatorial fusion we take is analogous to that used in information retrieval [38, 39], pattern recognition [40], molecular similarity searching and structure-based screening [41], and microarray gene expression analysis [42]. These works have demonstrated the following remark [43].

Remark 1. For a set of multiple scoring systems, each with a score function and a rank function, we have that (a) the combination of multiple scoring systems would improve the prediction accuracy only if (1) each of the systems has a relatively high performance, and (2) the individual systems are distinctive (or diversified), and (b) rank combination performs better than score combination under certain conditions.

Given an inhibitor and for each prediction result A, let s_A be a function as the predicted biological activity and it is represented as a real number. We view the function s_A as the score function. Since s_A only assigns a number not a set of numbers, in this work, no rank function would be used for an inhibitor. Therefore, the rank combination and the rule (b) in Remark 1 are not considered in the study. Suppose we have *m* prediction results (*m* scoring functions). There are combinatorially $2^m - 1$ combinations for all *m* individual prediction results $(\sum_{k=1}^m {m \choose k}) = 2^m - 1$ with score functions. The total number of combinations to be considered for predicting biological activity of an inhibitor is $2^m - 1$. This number of combinations can become huge when the number of prediction results m is large. Moreover, we have to evaluate the predictive power of each combination across all inhibitors. This study would start with combining only two prediction results which still retain fairly good prediction power.

Suppose *m* prediction results A_i , i = 1, 2, ..., m, are given with score function s_{Ai} ; there are several different ways of combination. Among others, there are score combination, voting, linear average combination, and weighted combination [38–42]. Voting is computationally simple and better than simple linear combinations when applied to the situation with large number of prediction results. However, a better alternative is to reduce the number of prediction results to a smaller number and then these prediction results are combined. In this paper, we reduce the set of prediction results to those which perform relatively well and then use the rank/score function to decide whether to combine by score. In this paper, we use the rules (a) (1) and (a) (2) stated in Remark 1 as our guiding principle to select prediction results and to decide on the method of combination. After generating each of the ten PhModels by using HypoGen Best, Fast, and Caesar algorithms for training set inhibitors, each of the best PhModel (denoted as Best_{train}, Fast_{train}, and Casear_{train}) was evaluated by its correlation coefficient of the

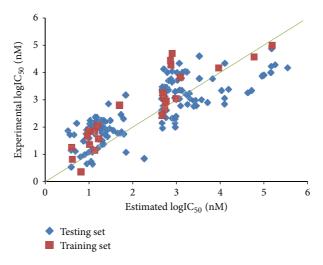


FIGURE 2: The scatter diagram of training set and testing set inhibitors.

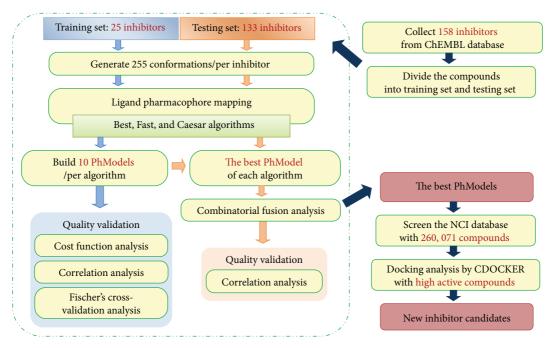


FIGURE 3: The workflow of PhModel generation for Chk2 inhibitors.

training set (r_{train}). Then these best PhModels were used to predict the biological activities of testing set inhibitors by using HypoGen Best, Fast, and Caesar algorithms. Therefore, there are nine prediction results (denoted as $Z_{train} \times Z_{test}$, $Z = \{Best, Fast, Caesar\}$, that is, $Best_{train}Best_{test}$) generated for testing set inhibitors. Using data fusion, results from various prediction results are combined to obtain predictions with larger accuracy rate. The diversity rank/score function is used to select the most suitable prediction results for combination. If these three best PhModels were selected, there are nine prediction results and then there are $2^9 - 1 = 511$ combinations. According to the rule (a) (1) in Remark 1, the r_{train} of Casear_{train} is far less than those of Best_{train} and Fast_{train} (Table 1); then, the Casear_{train} was not considered in the combinations. Therefore, there are six prediction results ($Z_{1\text{train}} \times Z_{2\text{test}}$, $Z_1 = \{\text{Best, Fast}\}$ and $Z_2 = \{\text{Best, Fast, Caesar}\}$) and $2^6 - 1 = 63$ combinations. A special diversity rank/score graph was used to choose the best discriminating prediction results for further combination.

For an inhibitor p_i in the testing set $P = \{p_1, p_2, ..., p_t\}$ and the pair of prediction results *A* and *B*, the diversity score function $d_i(A, B)$ is defined as $d_i(A, B) = \sum |s_A - s_B|$. When there are *q* prediction results selected (in this study, q =6), there are $\binom{q}{2} = q(q-1)/2$ (in this study, the number is 15) diversity score functions. If we let *i* vary and fix the prediction result pair (*A*, *B*), then $d_i(A, B)$ is the diversity score function $s_{(A,B)}$ from $P = \{p_1, p_2, ..., p_t\}$. Sorting $s_{(A,B)}$ into descending order would lead to the diversity

CHEMBL ID	Experimental IC ₅₀ (nM)		Estimated IC ₅₀ (nM)				
CHEMBLID	Experimental IC_{50} (IIW)	Best _{train}	Fast _{train}	Caesar _{train}			
CHEMBL195041	2.3	15	9.9	1129			
CHEMBL193990	6.6	6.8	6.2	942			
CHEMBL248935	14	20	20	833			
CHEMBL195320	18	8.5	6.2	942			
CHEMBL176164	23	19	23	1151			
CHEMBL250765	37	30	22	950			
CHEMBL362677	47	23	23	1153			
CHEMBL249959	70	110	20	1000			
CHEMBL250992	72	47	6.9	72			
CHEMBL251155	110	220	23	756			
CHEMBL588536	270	670 790		78578			
CHEMBL400772	470	2200	268	231			
CHEMBL367390	640	2000	2237	1028			
CHEMBL608262	830	1200	1456	94262			
CHEMBL401105	900	1000	235	20			
CHEMBL176115	1100	970	1044	1449			
CHEMBL253542	1200	1100	189	3.8			
CHEMBL592490	1800	860	1275	93360			
CHEMBL589090	6700	1700	1419	3561			
CHEMBL199299	15000	22000	233	1745			
CHEMBL251629	19000	3600	615	411			
CHEMBL259084	28000	6800	31827	5300			
CHEMBL251628	37000	63000	1360	24786			
CHEMBL438485	MBL438485 50000		320	243			
CHEMBL589501	CHEMBL589501 100000		48276	96926			
Correlation coefficient (r_{train})	0.955	0.840	0.238				

TABLE 1: Experimental and estimated IC_{50} values of training set inhibitors.

rank function $r_{(A,B)}$. Consequently, the diversity rank/score function $f_{(A,B)}$ is defined as $f_{(A,B)} = (s_{(A,B)} \circ r_{(A,B)}^{-1})(j) =$ $s_{(A,B)}(r_{(A,B)}^{-1}(j))$, where j is in $T = \{1, 2, 3, \dots, t\}$. We note that the set T is different from the set P which is the testing set considered. The set T is used as the index set for the diversity rank function value and |T| = t is indeed the cardinality of P. The diversity rank/score function $f_{(A,B)}$ so defined exhibits the diversity trend of the prediction result pair (A, B)across the whole spectrum of input set of t inhibitors and is independent of the specific inhibitor under study. For two prediction results A and B, the graph of the diversity rank/score function $f_{(A,B)}(j)$ is called the diversity rank/score graph. This study aims to examine all the q(q-1)/2 diversity rank/score graphs to see which pair of prediction results would give the larger diversity measurement according to the rule (a) (2) in Remark 1.

2.5. Database Screen. After examining 15 diversity rank/score graphs, the PhModels *A* and *B* determined from the best prediction result pair were used to screen the NCI database for new Chk2 inhibitor candidates. Under the PhModel, pharmacophore hypothesis screening can be used to screen small molecule database to retrieve the compounds as potential inhibitors that fit the pharmacophoric features.

In this study, the "Search 3D Database protocol" with the Best/Fast/Casear Search option in Accelrys Discovery Studio 2.1 was employed to search the NCI database with 260,071 compounds. We could filter out and select the compounds in the NCI database based on the estimated activity and chemical features of PhModel.

2.6. Molecular Docking. After the database screening approach, the selected compounds can be further estimated according to the interaction energy between a receptor and a ligand through the molecular docking approach. In this study, selected compounds in the NCI database were docked into Chk2 active sites by CDOCKER docking program, and then their CDOCKER interaction energies were estimated. Finally, new potential candidates were retrieved from the NCI database with high interaction energy. The workflow of database screening and molecular docking approach was shown in Figure 4.

3. Results

3.1. PhModel Generation Results. Each of the ten PhModels using 25 training set inhibitors and HypoGen Best, Fast, and Caesar algorithms was generated by selecting hydrogen bond

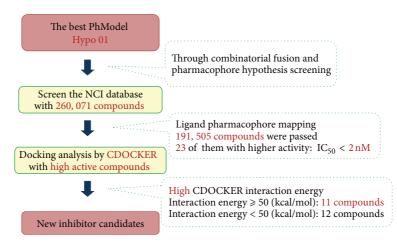


FIGURE 4: The workflow of database screening and molecular docking approach for new Chk2 inhibitor candidates.

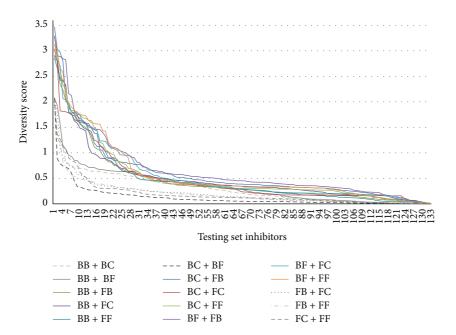


FIGURE 5: The diversity rank/score graphs for 15 combinations of prediction results.

acceptor (A), hydrogen bond donor (D), and hydrophobic (H) and hydrophobic aromatic (HYAR) features. Each of the best PhModels, $\text{Best}_{\text{train}}$, $\text{Fast}_{\text{train}}$, and $\text{Casear}_{\text{train}}$, was evaluated with the best r_{train} , and the predicted biological activities of training set inhibitors and r_{train} were listed in Table 1, respectively. From Table 1, the $\text{Best}_{\text{train}}$ obtained better r_{train} of value 0.955 than those by $\text{Fast}_{\text{train}}$ and $\text{Casear}_{\text{train}}$. Moreover, the r_{train} of $\text{Casear}_{\text{train}}$ is far less than those of $\text{Best}_{\text{train}}$ and $\text{Fast}_{\text{train}}$. Hence, HypoGen Best algorithm was used individually to generate the PhModels for most of target genes in the past. According to rule (a) (1) in Remark 1, the $\text{Casear}_{\text{train}}$ was not considered to be used for the prediction of testing set inhibitors.

3.2. Correlation Analysis of Testing Set Inhibitors. The testing set inhibitors were predicted by $\text{Best}_{\text{train}}$ and $\text{Fast}_{\text{train}}$ with HypoGen Best, Fast, and Caesar algorithms. Therefore, there

are six prediction results, $\text{Best}_{\text{train}}\text{Best}_{\text{test}}$ (denoted as BB), $\text{Best}_{\text{train}}\text{Fast}_{\text{test}}$ (denoted as BF), $\text{Best}_{\text{train}}\text{Casear}_{\text{test}}$ (denoted as BC), $\text{Fasr}_{\text{train}}\text{Best}_{\text{test}}$ (denoted as FB), $\text{Fast}_{\text{train}}\text{Fast}_{\text{test}}$ (denoted as FF), and $\text{Fast}_{\text{train}}\text{Casear}_{\text{test}}$ (denoted as FC), for testing set inhibitors. The predicted biological activities of testing set inhibitors and r_{test} by these six prediction results were listed in Table 2, respectively. From Table 2, for the $\text{Best}_{\text{train}}$, the best r_{test} of value 0.81 was achieved by the $\text{Best}_{\text{train}}$ Best $_{\text{test}}$; for the $\text{Fast}_{\text{train}}$, the best r_{test} of value 0.728 was achieved by the $\text{Fast}_{\text{train}}$ Fast $_{\text{test}}$. However, the $\text{Best}_{\text{train}}$ Best $_{\text{test}}$ obtained the best r_{test} in overall; moreover, the prediction results in the $\text{Best}_{\text{train}}$ all outperform those in the $\text{Fast}_{\text{train}}$.

3.3. Combinatorial Fusion Analysis. Under the six prediction results, the diversity score function $d_i(A, B)$ was calculated for each testing set inhibitor by a pair of prediction results (A, B). There are 15 diversity score functions $s_{(A,B)}$ that were

CHEMBL ID	Experimental IC ₅₀ (nM)	Estimated IC ₅₀ (nM)						
			Best _{train}			$Best_{train}Best_{test}$ +		
	10 ₅₀ (1111)	Best _{test}	Fast _{test}	Caesar _{test}	Best _{test}	Fast _{test}	Caesar _{test}	$Fast_{train}Fast_{test}$
CHEMBL195177	3.4	3.9	5.2	5.1	14.8	6.2	12.3	6.2
CHEMBL359881	4.4	12.0	46.1	42.3	17.3	22.5	29.4	22.5
CHEMBL179717	4.5	9.0	42.1	43.2	14.3	21.9	29.3	21.9
CHEMBL175553	5.5	11.2	57.8	43.1	17.6	20.5	29.1	20.5
CHEMBL192161	7	183.4	74.6	36.7	258.7	253.9	291.4	253.9
CHEMBL191969	8.2	6.4	15.3	14.3	10.5	9.4	13.5	9.4
CHEMBL175472	9.8	7.9	48.4	9.9	12.4	22.5	29.1	22.5
CHEMBL361378	12	9.9	48.1	43.7	14.7	21.7	29.3	21.7
CHEMBL362255	12	11.9	55.4	42.3	15.5	22.9	29.4	22.9
CHEMBL369254	12	70.2	51.3	45.6	38.0	23.7	31.5	23.7
CHEMBL364978	13	4.8	4.9	5.1	13.5	6.5	12.3	6.5
CHEMBL195846	14	3.9	5.0	5.1	12.8	6.2	12.3	6.2
CHEMBL179583	16	12.2	48.5	43.1	13.4	23.2	28.9	23.2
CHEMBL178972	17	15.9	52.5	43.1	18.0	23.3	29.1	23.3
CHEMBL250360	23	19.8	46.8	43.7	17.6	21.7	29.4	21.7
CHEMBL175879	24	21.6	57.0	42.3	24.0	23.2	29.2	23.2
CHEMBL179267	31	7.4	63.5	42.8	20.5	21.2	29.4	21.2
CHEMBL192022	32	20.3	47.5	42.6	23.9	22.4	29.4	22.4
CHEMBL250158	39	10.1	20.4	43.7	19.1	21.5	29.3	21.5
CHEMBL363339	41	10.1	59.8	42.2	18.1	24.0	29.5	24.0
CHEMBL250555	45	26.3	48.2	42.9	24.5	22.7	29.4	22.7
CHEMBL250359	52	3.7	5.2	3.5	20.6	7.0	28.8	7.0
CHEMBL251585	52	4.5	3.0	3.1	11.9	4.9	10.7	4.9
CHEMBL398529	53	23.0	48.4	43.5	21.4	22.4	29.4	22.4
CHEMBL178971	55	62.4	44.9	43.1	41.7	22.1	29.3	22.1
CHEMBL427879	55	13.7	45.4	42.4	16.9	19.9	29.5	19.9
CHEMBL250963	57	8.6	44.1	42.9	17.8	21.5	27.2	21.5
CHEMBL251170	60	51.3	45.3	43.1	26.3	21.5	29.4	21.5
CHEMBL250759	61	45.6	47.4	43.3	36.6	23.2	29.5	23.2
CHEMBL367263	61	19.7	50.3	9.6	17.3	23.9	29.1	23.9
CHEMBL250159	67	55.7	45.3	43.1	25.7	17.2	29.4	17.2
CHEMBL398467	70	11.4	46.5	42.7	21.0	20.4	29.4	20.4
CHEMBL250796	73	3.3	3.4	4.2	14.7	6.0	14.4	6.0
CHEMBL250957	74	36.1	44.9	43.7	30.5	20.4	29.5	20.4
CHEMBL206609	77	25.9	51.0	44.0	9.6	16.3	17.8	16.3
CHEMBL400755	78	8.8	26.6	43.0	12.9	22.5	29.4	22.5
CHEMBL249569	80	11.1	48.3	43.3	17.0	22.9	28.0	22.9
CHEMBL193397	81	9.9	43.8	42.8	14.5	21.2	28.4	21.2
CHEMBL438868	82	18.9	42.0	43.1	13.1	19.0	29.3	19.0
CHEMBL249566	86	17.6	48.8	43.1	23.5	22.8	29.6	22.8
CHEMBL249345	90	23.2	47.6	43.0	20.9	20.0	25.0	20.0
CHEMBL399146	90	29.8	47.9	42.7	28.4	22.1	29.3	22.1
CHEMBL602931	92	483.6	560.1	506.8	645.9	594.0	588.2	594.0
CHEMBL249347	95	10.1	46.3	43.1	27.2	20.1	29.2	20.1
CHEMBL193476	100	951.1	914.5	925.8	2435.2	1027.4	1981.9	1027.4

TABLE 2: Experimental and estimated $\mathrm{IC}_{\mathrm{50}}$ values of testing set inhibitors.

TABLE 2: Continued.

	Г. : (1	Estimated IC ₅₀ (nM)						
CHEMBL ID	Experimental IC ₅₀ (nM)		Best _{train}			Fast _{train}		$Best_{train}Best_{test} +$
	10 ₅₀ (1101)	Best _{test}	Fast _{test}	Caesar _{test}	Best _{test}	Fast _{test}	Caesar _{test}	Fast _{train} Fast _{test}
CHEMBL250361	100	14.2	6.6	14.5	14.9	20.9	29.6	20.9
CHEMBL248934	109	20.0	52.5	43.3	14.0	22.4	29.4	22.4
CHEMBL249750	110	12.7	49.3	43.5	20.7	21.6	29.3	21.6
CHEMBL208463	133	10.4	46.0	41.0	968.6	2768.3	2670.7	2768.3
CHEMBL250566	140	5.2	17.3	26.3	18.4	20.9	29.6	20.9
CHEMBL251256	140	936.4	2453.5	2240.6	192.6	215.1	215.5	215.1
CHEMBL437331	142	471.8	521.6	450.6	64.2	222.1	60.7	222.1
CHEMBL249541	157	23.8	42.5	43.0	16.0	18.8	29.3	18.8
CHEMBL249776	158	13.9	47.2	43.7	16.7	20.9	29.4	20.9
CHEMBL249350	174	30.6	51.2	43.4	22.5	23.3	29.4	23.3
CHEMBL249546	176	10.6	47.3	42.8	17.9	21.0	29.4	21.0
CHEMBL251364	176	21.7	43.9	43.1	17.8	19.7	29.1	19.7
CHEMBL399933	180	14.8	45.0	43.5	20.8	18.9	29.4	18.9
CHEMBL400287	180	19.3	45.5	43.5	21.5	19.8	29.4	19.8
CHEMBL175780	200	61.7	47.5	42.3	36.0	21.9	29.5	21.9
CHEMBL176326	200	935.6	926.3	913.6	1896.0	986.2	1980.7	986.2
CHEMBL590335	210	791.5	721.8	807.4	616.3	639.0	644.7	639.0
CHEMBL398561	220	575.1	926.2	571.5	209.1	278.3	287.9	278.3
CHEMBL249777	231	15.0	45.7	42.5	25.5	21.7	29.4	21.7
CHEMBL442282	233	13.0	46.9	42.3	21.5	22.5	29.4	22.5
CHEMBL195599	250	981.5	908.0	925.8	2266.7	1057.6	1981.9	1057.6
CHEMBL176015	290	54.1	52.8	54.4	24.0	29.3	28.0	29.3
CHEMBL251284	310	484.8	429.7	533.7	189.6	196.8	217.4	196.8
CHEMBL600441	310	454.6	559.9	516.5	300.5	513.5	254.1	513.5
CHEMBL599581	410	594.7	496.6	509.1	506.1	252.9	299.2	252.9
CHEMBL592784	420	462.2	492.0	475.1	216.7	203.8	198.3	203.8
CHEMBL1197465	580	2492.8	8163.9	5925.9	995.1	896.8	488.9	896.8
CHEMBL590809	600	492.4	539.0	534.3	816.4	536.2	549.3	536.2
CHEMBL1197456	610	2871.8	7537.6	6733.2	615.7	4139.2	3514.6	4139.2
CHEMBL590637	610	1251.9	1786.7	1121.1	2379.0	1650.9	1262.3	1650.9
CHEMBL590057 CHEMBL591518	680	1797.9	1804.0	1516.5	6075.0	4714.4	2726.5	4714.4
CHEMBL598973	700	12585.6	151896.0	84151.4	1047.3	396.5	1318.1	396.5
CHEMBL251368	710	27.7	45.9	43.8	13.4	21.2	29.1	21.2
CHEMBL1197303	800	5153.6	45.9 36481.1	45.8 6594.4	1277.8	681.9	4077.9	681.9
CHEMBL1197303 CHEMBL1197320	890						4077.9	517.5
CHEMBL1197528		2537.8	7191.1	6002.7	1559.4	517.5		
	960	2752.7	7737.1	5925.9	765.7	654.9	559.0	654.9
CHEMBL215803	1000	3760.6	140257.0	74847.0	9672.7	50053.5	49263.9	50053.5
CHEMBL253324	1000	996.2	2416.5	603.1	264.7	553.1	272.4	553.1
CHEMBL589347	1100	458.3	560.2	482.3	209.7	299.1	196.7	299.1
CHEMBL604784	1100	1188.4	1365.6	1205.7	2305.3	1962.3	1307.9	1962.3
CHEMBL1197529	1120	2047.4	11678.5	8090.0	3368.8	3648.8	3659.9	3648.8
CHEMBL1197326	1130	12645.5	45428.8	7432.7	1138.2	465.0	416.0	465.0
CHEMBL176041	1200	925.0	906.8	913.6	1933.6	1040.0	1980.7	1040.0
CHEMBL590079	1350	548.9	548.5	550.6	1000.9	860.3	855.1	860.3
CHEMBL605083	1400	489.2	613.8	554.4	1224.9	1401.0	1345.6	1401.0
CHEMBL175481	1500	69.2	57.7	49.2	1853.9	1526.1	1852.1	1526.1

TABLE 2: Continued.

		Estimated IC ₅₀ (nM)						
CHEMBL ID	Experimental IC ₅₀ (nM)		Best _{train}			Fast _{train}		$Best_{train}Best_{test} +$
	1C ₅₀ (IIIVI)	Best _{test}	Fast _{test}	Caesar _{test}	Best _{test}	Fast _{test}	Caesar _{test}	$Fast_{train}Fast_{test}$
CHEMBL205906	1540	1696.1	1109.3	1153.0	980.6	17655.5	937.5	17655.5
CHEMBL590808	1600	1662.5	1799.1	1709.4	51882.2	48813.2	48302.3	48813.2
CHEMBL1170748	1700	1974.2	1940.4	1652.8	3251.4	2734.9	407.6	2734.9
CHEMBL253541	1800	43066.9	42831.1	79622.8	1255.9	1099.1	15566.3	1099.1
CHEMBL176554	1900	572.3	7226.3	9126.2	1096.1	3352.6	3926.3	3352.6
CHEMBL377597	2000	938.5	6719.7	5644.6	50.4	116.9	105.4	116.9
CHEMBL1170749	2200	10874.0	30873.8	3995.6	5835.1	25044.3	1090.5	25044.3
CHEMBL590336	2200	507.8	594.0	516.4	568.9	544.3	521.1	544.3
CHEMBL590807	2200	52944.6	70480.3	52639.4	44898.4	49196.7	36195.2	49196.7
CHEMBL600868	2200	1342.8	1603.8	1312.9	7373.6	5797.7	4864.3	5797.7
CHEMBL398759	2300	666.0	2847.2	1010.2	447.2	924.3	309.0	924.3
CHEMBL604459	2300	1176.9	2299.2	1369.3	614.2	877.8	682.5	877.8
CHEMBL179383	2400	15724.9	14621.7	14257.4	4508.5	4132.8	4023.9	4132.8
CHEMBL592489	2400	469.2	494.6	486.9	204.7	240.5	208.4	240.5
CHEMBL425904	2800	605.0	651.0	534.9	478.1	451.9	488.6	451.9
CHEMBL150894	3000	537.8	844.9	797.6	186.8	600.2	289.0	600.2
CHEMBL590793	3000	475.4	2480.4	1251.5	192.0	251.7	205.4	251.7
CHEMBL600865	4400	796.9	2072.3	1223.3	198.1	261.3	228.1	261.3
CHEMBL249253	5000	659.8	503.6	499.5	406.6	227.9	309.5	227.9
CHEMBL587506	5200	1050.8	1293.3	1083.2	2201.3	2162.7	1656.0	2162.7
CHEMBL204930	5800	755.5	1260.7	1105.5	47881.6	47912.3	47868.0	47912.3
CHEMBL554900	5900	6722.8	580526.0	807309.0	745.4	428.2	47897.1	428.2
CHEMBL176276	6000	3360.2	8955.9	8476.6	2075.6	2054.0	1970.7	2054.0
CHEMBL589091	6100	1319.5	1308.4	1172.0	579.2	603.1	600.9	603.1
CHEMBL559781	7400	91783.7	822917.0	1230010.0	414.5	6865.5	47911.9	6865.5
CHEMBL249252	8000	100671.0	38586.8	31129.8	590.5	491.6	467.9	491.6
CHEMBL589089	9800	1070.7	1321.5	1002.7	38589.3	48136.2	41143.2	48136.2
CHEMBL217090	10000	628.8	1010.9	1548.5	390.6	451.8	910.1	451.8
CHEMBL217092	10000	1030.2	1199.3	1908.8	379.9	625.6	707.2	625.6
CHEMBL382588	10000	1365.2	5880.3	5488.7	2560.7	3993.4	3649.4	3993.4
CHEMBL590581	10000	145206.0	149922.0	108067.0	50609.5	49348.5	48302.3	49348.5
CHEMBL242753	10300	1742.5	3393.1	1941.1	1202.5	2613.0	1297.3	2613.0
CHEMBL398758	11000	1582.8	187965.0	1573.6	237.0	4333.9	369.0	4333.9
CHEMBL399151	11000	812.1	1263.5	2069.9	334.5	1193.9	1271.6	1193.9
CHEMBL395080	13450	513.5	484.0	458.6	198.5	188.9	183.3	188.9
CHEMBL1171533	15000	349604.0	296515.0	159368.0	26784.5	26699.3	45506.4	26699.3
CHEMBL602729	17000	148756.0	149052.0	139465.0	224.1	324.7	229.4	324.7
CHEMBL249255	19000	182486.0	42615.8	41578.3	1698.2	616.1 210.7	683.9	616.1
CHEMBL202930	21730	12828.4	11945.6	12145.8	213.3	219.7	217.6	219.7
CHEMBL589986	22000	1167.1	1337.4	1143.5	53747.5	49155.6	49316.8	49155.6
CHEMBL251471 CHEMBL560056	40000 74000	3358.9 152006 0	1946.2 156723.0	2075.7 208466.0	512.2	423.7	3427.4	423.7
Correlation	/4000	152006.0	156723.0	200400.0	223.2	208.1	190.4	208.1
coefficient (r_{test})		0.810	0.771	0.783	0.710	0.728	0.714	0.816

FIGURE 6: The r_{test} for all of 63 combinations from six prediction results.

Name

NSC 136954

NSC 70804

NSC 158029

NSC 603427

NSC 57782

NSC 16739

NSC 720227

NSC 618702

NSC 195178

NSC 653142

NSC 653143

NSC 32200

NSC 342015

NSC 343685

NSC 205750

NSC 96538

NSC 210455

NSC 314654

NSC 179894

NSC 370907

NSC 91710

performed at first and then these diversity score functions were sorted to become the diversity rank function $r_{(A,B)}$, respectively. Finally, 15 diversity rank/score functions $f_{(A,B)}$ were represented as diversity rank/score graphs shown in Figure 5. Among 15 diversity rank/score graphs, there are several combinations (gray color) that have less diversity scores than those by others, such as BB + BC, BB + BF, and FB + FB, shown in Figure 5. It means that these combinations may have less r_{test} than those by others according to rule (a) (2) in Remark 1. In other words, several combinations, such as BB + FC (purple color), BB + FF (blue color), and BF + FF (orange color), may have larger r_{test} than those by others due to larger diversity scores. For the six prediction results, all of the 63 combinations were preformed and evaluated by its r_{test} , respectively, as shown in Figure 6. In Figure 6, for 15 pairs of two prediction results, the combinations BB + FB, BB + FC, and BB + FF have larger r_{test} than those by others. Moreover, the combination BB + FF has best r_{test} of value 0.816 among 15 combinations, even for 63 combinations. Besides, the average r_{test} by the combinations is larger than the individual prediction results. It means that the predictive accuracy for Chk2 inhibitors may be improved by considering the Best_{train} and Fast_{train} concurrently.

3.4. Database Screen Results. The best PhModels, $Best_{train}$ and Fast_{train}, were used to screen the NCI database with 260,071 compounds for new Chk2 inhibitor candidates by using HypoGen Best and Fast algorithms, respectively. The $Best_{train}Best_{test}$ and $Fast_{train}Fast_{test}$ prediction results for NCI database were combined in order to filter out possible false positive candidates. Of the 260,071 compounds, 191,505 passed the screening and best fitted to the chemical features in 3D space. 23 drug-like compounds that had an estimated IC₅₀

TABLE 3: The 21 drug-like compounds with their estimated IC_{50} values and CDOCKER interaction energy greater than 37.786 (kal/mol).

Interaction energy (kal/mol)

61.239

58.967

57.944

56.963

56.54

56.342

55.839

55.196

51.351

51.19

50.055

49.439

47.327

46.436

45.542

44.344

42.258

42.082

41.707

40.533

40.502

Estimated IC₅₀ (nM)

1.989

1.682

1.885

1.87

1.6855

1.5385

1.914

1.862

1.7015

1.557

1.577

1.901

1.6515

1.7615

1.875

1.705

1.7935

1.947

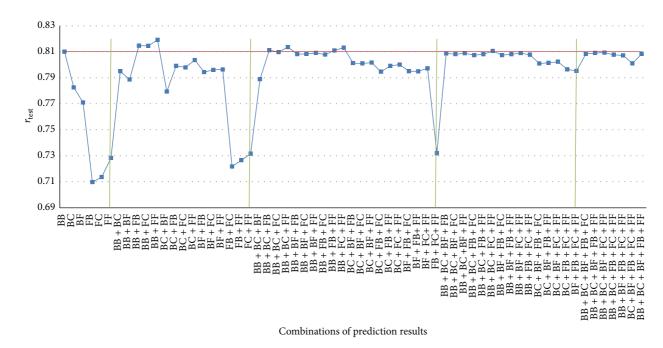
1.6135

1.701

1.8785

value of less th	han 2 nM	were s	studied	in a	molecular	docking
study (Figure -	4).					-

3.5. *Molecular Docking Results.* 23 drug-like compounds along with the training set compounds were docked into the



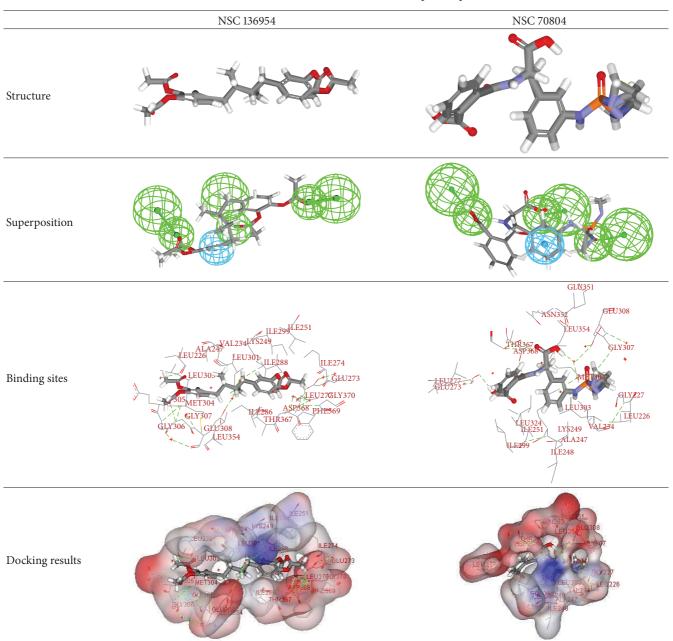


TABLE 4: The structures and characteristics of the top 2 compounds.

active sites that were defined based on the bound inhibitor, PV1019, in a crystal structure of Chk2 (PDB: 2W7X). We used CDOCKER program to confirm that inhibitor candidates bind to the receptor. CDOCKER uses molecular dynamics (MD) in conjunction with the CHARMm force field to individually dock the compounds into the binding sites. The coordinates of Chk2 from the Chk2/PV1019 crystal structure were used after removing PV1019 and solvent molecules and adding protein hydrogen atoms. After docking each screened compound, its interaction energy value was calculated. The PV1019 was redocked into the Chk2 binding site by the CDOCKER program. Its-CDOCKER interaction energy was calculated by CDOCKER and determined to be 37.786 (kal/mol). The 23 drug-like compounds were docked into the Chk2 binding sites. Finally, there are 21 drug-like compounds with CDOCKER interaction energies greater than 37.786 (kal/mol). In addition, 11 drug-like compounds had high interaction value greater than 50 (kal/mol) (Figure 4) and the top 2 are NSC136954 with 61.239 (kal/mol) and NSC70804 with 58.967 (kal/mol), respectively, kept for future characterization as inhibitors. The 21 drug-like compounds with their estimated IC₅₀ values and CDOCKER interaction energy greater than 37.786 (kal/mol) were shown in Table 3.

The structures and characteristics of the top 2 compounds are given in Table 4, and we can find that some active site residues were identified from the Chk2 complex. The interaction sites of NSC136954 were Leu226, Val234, Ala247, Lys249, Ile251, Glu273, Ile274, Leu277, Ile286, Ile288, Ile299, Leu301, Leu303, Met304, Glu305, Gly306, Gly307, Glu308, Leu354, Thr367, Asp368, Phe369, and Gly370. On the other hand, the interaction sites of NSC70804 were Leu226, Leu227, Val234, Ala247, Ile248, Lys249, Ile251, Glu273, Leu277, Ile299, Leu301, Leu303, Met304, Gly307, Glu308, Glu351, Asn352, Leu354, Thr367, and Asp368. Several studies indicated that they are involved in hydrophobic interactions with Val234, Ile251, Leu354, Ile299, and the aliphatic portions of the side chains of Lys249, Thr367, and Asp368, in addition to several interactions of van der Waals or hydrophobic with Leu226, Val234, Leu303, Gly307, Leu354, and the aliphatic portions the side chains of Met304 and Glu308 [10, 11]. Furthermore, the quinazoline was sandwiched between the lipophilic side chains of Val234 and Leu354, with the side chains of Ala247, Leu301, and Leu303 also contributing to a hydrophobic surface surrounding the core and an interaction between the pyrazole and Lys249 is likely to account for the increase in Chk2 potency [12]. And residue Thr367 of Chk2 is a serine in Chk1. Portions of the glycine-rich P-loop in Chk2, which is located directly above the inhibitor, are disordered (residues 229-231), whereas this loop is well defined in the structure of Chkl, and Leu301 in Chk2 corresponds to the "gatekeeper" residue in many kinases, which has been found to form contacts with bound inhibitors and is poorly conserved [44].

4. Conclusions

In this study, a novel design strategy for drug design was proposed to apply combinatorial fusion into PhModels and virtual screening techniques. 158 Chk2 inhibitors were divided into the training set and testing set, respectively. For 25 training set inhibitors, three best PhModels, Best_{train}, Fast_{train}, and Casear_{train}, were generated at first, and then six prediction results for 133 testing set inhibitors were used for calculating 15 diversity rank/score functions. Finally, the combination $Best_{train}Best_{test}$ and $Fast_{train}Fast_{test}$ prediction results achieved the best r_{test} of value 0.816 among 63 combinations. Through these approaches, 23 potential Chk2 inhibitors with IC_{50} value less than $2\,nM$ and interaction energy value larger than 37.786 (kal/mol) are retrieved from NCI database. This study can help medicinal chemists to identify or design new Chk2 inhibitors. Besides, the potential inhibitors of Chk2 retrieved in this work can be estimated by biologists for further study.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This work was supported in part by the National Science Council of Taiwan (under Grants NSC100-2221-E-182-057-MY3) and by Chang Gung Memorial Hospital (Grant CMRPD260033). The authors thank the National Center for High-Performance Computing for computer time and use of its facilities.

References

- K. M. Culligan, C. E. Robertson, J. Foreman, P. Doerner, and A. B. Britt, "ATR and ATM play both distinct and additive roles in response to ionizing radiation," *Plant Journal*, vol. 48, no. 6, pp. 947–961, 2006.
- [2] J. Yang, Z.-P. Xu, Y. Huang, H. E. Hamrick, P. J. Duerksen-Hughes, and Y.-N. Yu, "ATM and ATR: sensing DNA damage," *World Journal of Gastroenterology*, vol. 10, no. 2, pp. 155–160, 2004.
- [3] Q. Liu, S. Guntuku, X.-S. Cui et al., "Chkl is an essential kinase that is regulated by Atr and required for the G2/M DNA damage checkpoint," *Genes and Development*, vol. 14, no. 12, pp. 1448– 1459, 2000.
- [4] C. Tapia-Alveal, T. M. Calonge, and M. J. O'Connell, "Regulation of Chkl," *Cell Division*, vol. 4, article 8, 2009.
- [5] J. Bartek, J. Falck, and J. Lukas, "Chk2 kinase—a busy messenger," *Nature Reviews Molecular Cell Biology*, vol. 2, no. 12, pp. 877–886, 2001.
- [6] J. Li, B. L. Williams, L. F. Haire et al., "Structural and functional versatility of the FHA domain in DNA-damage signaling by the tumor suppressor kinase Chk2," *Molecular Cell*, vol. 9, no. 5, pp. 1045–1054, 2002.
- [7] A. W. Oliver, A. Paul, K. J. Boxall et al., "Trans-activation of the DNA-damage signalling protein kinase Chk2 by T-loop exchange," *EMBO Journal*, vol. 25, no. 13, pp. 3179–3190, 2006.
- [8] Z. Cai, N. H. Chehab, and N. P. Pavletich, "Structure and activation mechanism of the CHK2 DNA damage checkpoint kinase," *Molecular Cell*, vol. 35, no. 6, pp. 818–829, 2009.
- [9] Z. A. Stewart and J. A. Pietenpol, "p53 signaling and cell cycle checkpoints," *Chemical Research in Toxicology*, vol. 14, no. 3, pp. 243–263, 2001.
- [10] G. T. Lountos, A. G. Jobson, J. E. Tropea et al., "Structural characterization of inhibitor complexes with checkpoint kinase 2 (Chk2), a drug target for cancer therapy," *Journal of Structural Biology*, vol. 176, no. 3, pp. 292–301, 2011.
- [11] A. G. Jobson, G. T. Lountos, P. L. Lorenzi et al., "Cellular inhibition of checkpoint kinase 2 (Chk2) and potentiation of camptothecins and radiation by the novel Chk2 inhibitor PV1019 [7-nitro-1H-indole-2-carboxylic acid 4-[1-(guanidinohydrazone)-ethyl]-phenyl-amide]," *Journal of Pharmacology and Experimental Therapeutics*, vol. 331, no. 3, pp. 816–826, 2009.
- [12] J. J. Caldwell, E. J. Welsh, C. Matijssen et al., "Structure-based design of potent and selective 2-(quinazolin-2-yl)phenol inhibitors of checkpoint kinase 2," *Journal of Medicinal Chemistry*, vol. 54, no. 2, pp. 580–590, 2011.
- [13] G. T. Lountos, A. G. Jobson, J. E. Tropea et al., "X-ray structures of checkpoint kinase 2 in complex with inhibitors that target its gatekeeper-dependent hydrophobic pocket," *FEBS Letters*, vol. 585, no. 20, pp. 3245–3249, 2011.
- [14] S. Hilton, S. Naud, J. J. Caldwell et al., "Corrigendum to 'Identification and characterisation of 2-aminopyridine inhibitors of checkpoint kinase 2," *Bioorganic and Medicinal Chemistry*, vol. 18, no. 12, p. 4591, 2010.
- [15] E. M. Rosen, S. Fan, R. G. Pestell, and I. D. Goldberg, "BRCA1 gene in breast cancer," *Journal of Cellular Physiology*, vol. 196, no. 1, pp. 19–41, 2003.

- [16] E. S. Yang and F. Xia, "BRCA1 16 years later: DNA damageinduced BRCA1 shuttling," *FEBS Journal*, vol. 277, no. 15, pp. 3079–3085, 2010.
- [17] M. Gupta, S. Gupta, H. Dureja, and A. K. Madan, "Superaugmented eccentric distance sum connectivity indices: Novel highly discriminating topological descriptors for QSAR/QSPR," *Chemical Biology and Drug Design*, vol. 79, no. 1, pp. 38–52, 2012.
- [18] F. A. Pasha, M. Muddassar, and S. Joo Cho, "Molecular docking and 3D QSAR studies of Chk2 inhibitors," *Chemical Biology and Drug Design*, vol. 73, no. 3, pp. 292–300, 2009.
- [19] H. Kubinyi, G. Folkers, and Y. C. Martin, *3D QSAR in Drug Design*, Springer, 2002.
- [20] Y.-K. Jiang, "Molecular docking and 3D-QSAR studies on βphenylalanine derivatives as dipeptidyl peptidase IV inhibitors," *Journal of Molecular Modeling*, vol. 16, no. 7, pp. 1239–1249, 2010.
- [21] R. R. S. Pissurlenkar, M. S. Shaikh, and E. C. Coutinho, "3D-QSAR studies of Dipeptidyl peptidase IV inhibitors using a docking based alignment," *Journal of Molecular Modeling*, vol. 13, no. 10, pp. 1047–1071, 2007.
- [22] W. Sippl, 3D-QSAR—Applications, Recent Advances, and Limitations. Recent Advances in QSAR Studies, Springer, 2010.
- [23] A. Lauria, M. Ippolito, M. Fazzari et al., "IKK-β inhibitors: an analysis of drug-receptor interaction by using Molecular Docking and Pharmacophore 3D-QSAR approaches," *Journal* of Molecular Graphics and Modelling, vol. 29, no. 1, pp. 72–81, 2010.
- [24] S. John, S. Thangapandian, M. Arooj, J. C. Hong, K. D. Kim, and K. W. Lee, "Development, evaluation and application of 3D QSAR Pharmacophore model in the discovery of potential human renin inhibitors," *BMC Bioinformatics*, vol. 12, pp. 1–14, 2011.
- [25] H. Kubinyi and Comparative Molecular Field Analysis (CoMFA), Handbook of Chemoinformatics: From Data To Knowledge in 4 Volumes, Wiley, 2008.
- [26] K. W. Lee and J. M. Briggs, "Comparative molecular field analysis (CoMFA) study of epothilones-tubulin depolymerization inhibitors: pharmacophore development using 3D QSAR methods," *Journal of Computer-Aided Molecular Design*, vol. 15, no. 1, pp. 41–55, 2001.
- [27] S. Durdagi, T. Mavromoustakos, and M. G. Papadopoulos, "3D QSAR CoMFA/CoMSIA, molecular docking and molecular dynamics studies of fullerene-based HIV-1 PR inhibitors," *Bioorganic and Medicinal Chemistry Letters*, vol. 18, no. 23, pp. 6283–6289, 2008.
- [28] M.-E. Suh, S.-Y. Park, and H.-J. Lee, "Comparison of QSAR methods (CoMFA, CoMSIA, HQSAR) of anticancer 1-Nsubstituted imidazoquinoline-4,9-dione derivatives," *Bulletin of the Korean Chemical Society*, vol. 23, no. 3, pp. 417–422, 2002.
- [29] S. J. Bang and S. J. Cho, "Comparative molecular field analysis (CoMFA) and comparative molecular similarity index analysis (CoMSIA) study of mutagen X," *Bulletin of the Korean Chemical Society*, vol. 25, no. 10, pp. 1525–1530, 2004.
- [30] L. Ghemtio, Y. Zhang, and H. Xhaard, Virtual Screening, InTech, 2012.
- [31] G. Klebe, 3D QSAR in Drug Design, vol. 3, Springer, 2002.
- [32] G. Klebe and U. Abraham, "Comparative Molecular Similarity Index Analysis (CoMSIA) to study hydrogen-bonding properties and to score combinatorial libraries," *Journal of Computer-Aided Molecular Design*, vol. 13, no. 1, pp. 1–10, 1999.
- [33] I. Mitra, A. Saha, and K. Roy, "Pharmacophore mapping of arylamino-substituted benzo[b]thiophenes as free radical

scavengers," Journal of Molecular Modeling, vol. 16, no. 10, pp. 1585–1596, 2010.

- [34] K. Boppana, P. K. Dubey, S. A. R. P. Jagarlapudi, S. Vadivelan, and G. Rambabu, "Knowledge based identification of MAO-B selective inhibitors using pharmacophore and structure based virtual screening models," *European Journal of Medicinal Chemistry*, vol. 44, no. 9, pp. 3584–3590, 2009.
- [35] M. Chopra, R. Gupta, S. Gupta, and D. Saluja, "Molecular modeling study on chemically diverse series of cyclooxygenase-2 selective inhibitors: generation of predictive pharmacophore model using Catalyst," *Journal of Molecular Modeling*, vol. 14, no. 11, pp. 1087–1099, 2008.
- [36] A. Gaulton, L. J. Bellis, A. P. Bento et al., "ChEMBL: a large-scale bioactivity database for drug discovery," *Nucleic Acids Research*, vol. 40, pp. D1100–D1107, 2012.
- [37] K. C. Shih, C. W. Shiau, T. S. Chen et al., "Automated chemical hypothesis generation and database searching with Catalyst," *Perspectives in Drug Discovery and Design*, vol. 3, pp. 1–20, 1995.
- [38] D. F. Hsu and I. Taksa, "Comparing rank and score combination methods for data fusion in information retrieval," *Information Retrieval*, vol. 8, no. 3, pp. 449–480, 2005.
- [39] C. C. Vogt and G. W. Cotrell, "Fusion via a linear combination of scores," *Information Retrieval*, vol. 1, no. 3, pp. 151–172, 1999.
- [40] L. Xu, A. Krzyzak, and C. Y. Suen, "Methods of combining multiple classifiers and their applications to handwriting recognition," *IEEE Transactions on Systems, Man and Cybernetics*, vol. 22, no. 3, pp. 418–435, 1992.
- [41] J.-M. Yang, Y.-F. Chen, T.-W. Shen, B. S. Kristal, and D. F. Hsu, "Consensus scoring criteria for improving enrichment in virtual screening," *Journal of Chemical Information and Modeling*, vol. 45, no. 4, pp. 1134–1146, 2005.
- [42] M. A. Kuriakose, W. T. Chen, Z. M. He et al., "Selection and validation of differentially expressed genes in head and neck cancer," *Cellular and Molecular Life Sciences*, vol. 61, no. 11, pp. 1372–1383, 2004.
- [43] K.-L. Lin, C.-Y. Lin, C.-D. Huang et al., "Feature selection and combination criteria for improving accuracy in protein structure prediction," *IEEE Transactions on Nanobioscience*, vol. 6, no. 2, pp. 186–196, 2007.
- [44] G. T. Lountos, J. E. Tropea, D. Zhang et al., "Crystal structure of checkpoint kinase 2 in complex with NSC 109555, a potent and selective inhibitor," *Protein Science*, vol. 18, no. 1, pp. 92–100, 2009.