

RESEARCH

Open Access



# Investigation of hub gene associated with the infection of *Staphylococcus aureus* via weighted gene co-expression network analysis

Jia-xin Li<sup>1,2</sup>, Xun-jie Cao<sup>1,3</sup>, Yuan-yi Huang<sup>2</sup>, Ya-ping Li<sup>4</sup>, Zi-yuan Yu<sup>1,3</sup>, Min Lin<sup>5</sup>, Qiu-ying Li<sup>1,3</sup>, Ji-chun Chen<sup>1,3</sup> and Xu-guang Guo<sup>1,6\*</sup>

## Abstract

**Introduction:** *Staphylococcus aureus* is a gram-positive bacterium that causes serious infection. With the increasing resistance of bacteria to current antibiotics, it is necessary to learn more about the molecular mechanism and cellular pathways involved in the *Staphylococcus aureus* infection.

**Methods:** We downloaded the GSE33341 dataset from the GEO database and applied the weighted gene co-expression network analysis (WGCNA), from which we obtained some critical modules. Kyoto Encyclopedia of Genes and Genomes (KEGG) and Gene Ontology (GO) were applied to illustrate the biological functions of genes in these modules. We constructed the protein-protein interaction (PPI) network by Cytoscape and selected five candidate hub genes. Five potential hub genes were validated in GSE30119 by GraphPad Prism 8.0. The diagnostic values of these genes were calculated and present in the ROC curve based on the GSE13670 dataset. Their gene functions were analyzed by Gene Set Enrichment Analysis (GSEA).

**Results:** A co-expression network was built with 5000 genes divided into 11 modules. The genes in green and turquoise modules demonstrated a high correlation. According to the KEGG and GO analyses, genes in the green module were closely related to ubiquitination and autophagy. Subsequently, we picked out the top five hub genes in the green module. And UBB was determined as the hub gene in the GSE30119 dataset. The expression level of UBB, ASB, and MKRN1 could significantly differentiate between *Staphylococcus aureus* infection and healthy controls based on the ROC curve. The GSEA analysis indicated that lower expression levels of UBB were associated with the P53 signal pathway.

**Conclusions:** We identified some hub genes and significant signal enrichment pathways in *Staphylococcus aureus* infection via bioinformatics analysis, which may facilitate the development of potential clinical therapeutic strategies.

**Keywords:** *Staphylococcus aureus*, Weighted gene co-expression network analysis, Mitophagy, Bioinformatic analysis

## Introduction

As a gram-positive bacterium, [1–3] *Staphylococcus aureus* can infect various tissue and organs in human beings, causing mild skin infection or even severe illnesses [4]. It causes conditions such as endocarditis, osteomyelitis, septicemia, and toxic shock syndrome

\*Correspondence: gysyngx@gmail.com

<sup>1</sup> Department of Clinical Laboratory Medicine, The Third Affiliated Hospital of Guangzhou Medical University, Guangzhou 510150, China  
Full list of author information is available at the end of the article



© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

toxin (TSST), [5–7] primarily through the production of a variety of toxic factors and toxins [5, 6]. Though most of the antibiotics used to treat *Staphylococcus aureus* work well, emerging evidence has shown that *Staphylococcus aureus*'s resistance becomes a major issue for clinicians [2, 6]. For instance, the methicillin-resistant *S. aureus* (MRSA), [8, 9] one of the most common resistant strains, leads to increased mortality and morbidity, which makes it difficult for clinicians to prescribe the suitable treatment therapy [1, 10]. By forming colonies in human nares and skin, *Staphylococcus aureus* causes mild symptoms while invading deeper tissues and exposed organs in a vulnerable environment, which causes severe diseases [2, 6]. Given the lack of effective treatment regimens and vaccines for drug-resistant *Staphylococcus aureus*, [11, 12] it is necessary to explore potential target genes associated with *Staphylococcus aureus*.

In this study, we construct a gene co-expression network using the WGCNA method and select some modules that pique our interest for further analysis. KEGG and GO analyses were applied to explore the biological functions of the modules. We obtained five hub genes through STRING and Cytoscape and explored the relationship between these genes and *Staphylococcus aureus* infection prognosis through the ROC curve. Moreover, we further addressed the biological roles of these genes during infection through GSEA. These results offer some critical genetic candidates for treating *Staphylococcus aureus* infection, which may facilitate vaccine development and treatment optimization for *Staphylococcus aureus* infection.

## Methods and materials

### Data inclusion and processing

Figure 1 displayed the workflow in our study. The data processing methods applied in our study were carried out in accordance with relevant guidelines and regulations. The Gene Expression Omnibus (GEO) is a comprehensive database of gene expression, collecting microarray and high-throughput resources. (<http://www.ncbi.nlm.nih.gov/geo/>). In this study, we downloaded the gene expression profile of GSE33341 [13] from the GEO database. The GSE33341 dataset was based on the GPL1261 platform of Affymetrix Mouse Genome 430 2.0 Array and GPL571 platform of Affymetrix Human Genome U133A 2.0 Array. We extracted the data that included 32 human samples infected with *Staphylococcus aureus* and 43 healthy counterparts. Then, we converted gene probes into gene symbols based on microarray annotation information on the GPL571 platform. For probes corresponding to multiple gene symbols, we randomly selected one gene symbol for matching the probes. Probes with no corresponding gene symbols were removed. For gene

symbols corresponding to multiple probes, we reserved the probe with the highest average value. Through the above steps, we ensure the one-to-one correspondence between probes and gene symbols. Furthermore, genes with negative values were deleted.

### Weighted gene co-expression network analysis

The top 5000 genes with high expression values in 32 infected samples were selected to construct co-expression modules through the R package “WGCNA”. (Fig. 2A) WGCNA is a systems biology approach applied to search for highly correlated gene modules or identify biomarkers for candidate diseases [14]. When the soft thresholding power  $\beta$  was 7, the scale-free  $R^2$  of the co-expression network was close to 0.9. (Fig. 2B). Therefore, network construction and module detection were continued based on the soft thresholding power  $\beta$ . To enhance the reliability of the results, we set the minimum number of genes in modules as 30. We further defined 0.4 as the threshold for cut height to merge modules with high correlation.

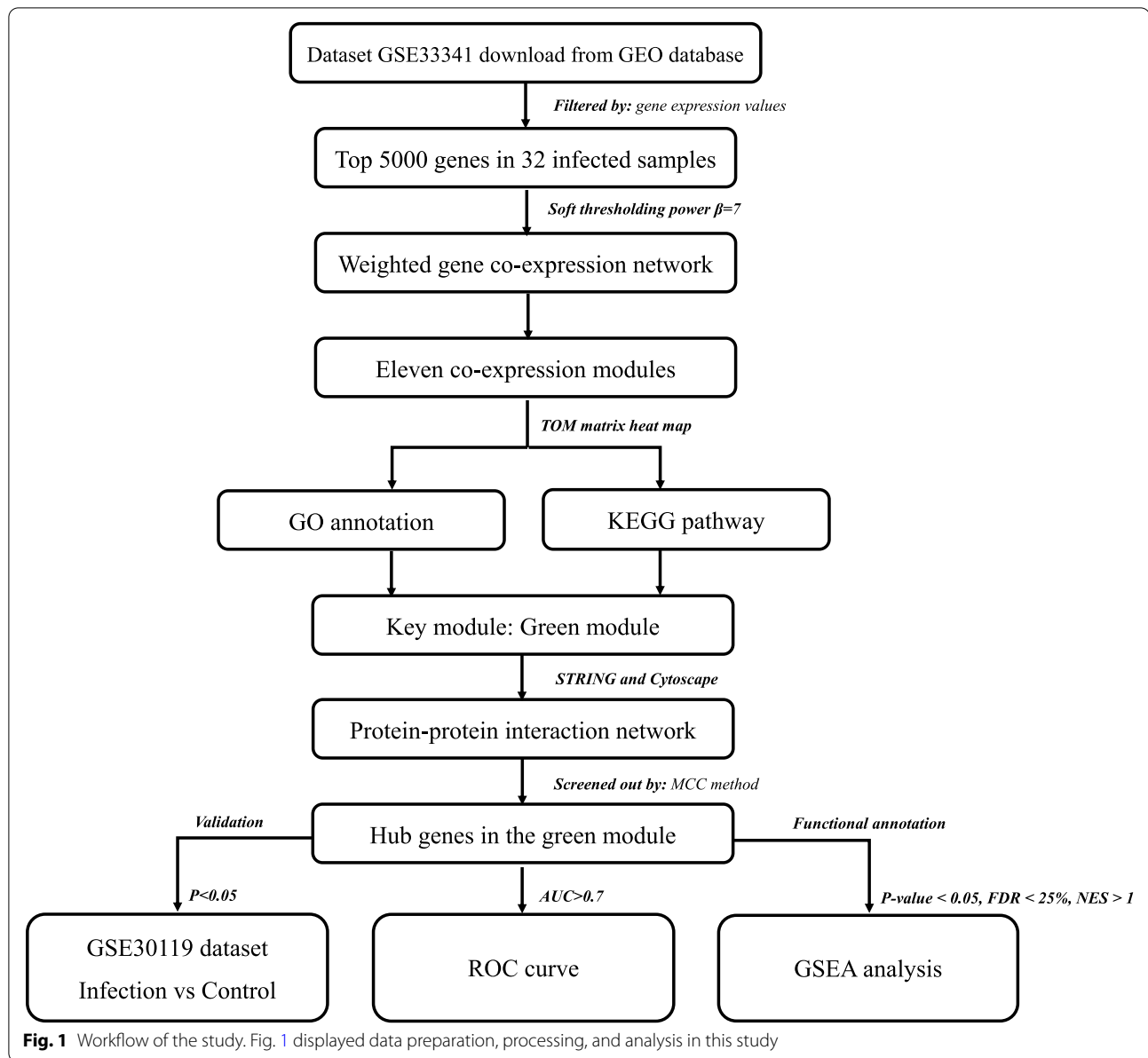
### Functional and pathway enrichment analyses of genes in the critical modules

WGCNA can divide genes with similar functions into the same module. Therefore, genes in the same co-expression modules possess a higher degree of connectivity, which means they may play similar roles. According to the TOM matrix heat map, we identified two key modules that were significantly associated with *Staphylococcus aureus* infection.

KEGG is an integrated database for analyzing genomes and biological data [15–17]. ([www.kegg.jp/kegg/kegg1.html](http://www.kegg.jp/kegg/kegg1.html)) Gene ontology (GO) is an international standardized classification system for gene function, covering biological process, molecular function, and cellular components [18]. To study the biological function of the genes in the important modules, we performed KEGG and GO pathway enrichment analyses by R language.  $P < 0.05$  was set as the cut-off value.

### Hub genes identification

PPI network is a useful tool to understand cell functions and disease machinery, which is crucial in predicting the function of interacting proteins [19]. We imported the genes previously obtained from the critical modules into the online database STRING (version 11.0; <https://string-db.org/>), which helps constructing the PPI network. The combined score was set as over 0.4. Subsequently, we searched the key modules and hub genes in the PPI network by the MCODE plugin and CytoHubba plugin in Cytoscape software. Finally, we defined the top five genes that displayed the highest degree of connectivity in key modules as hub genes.



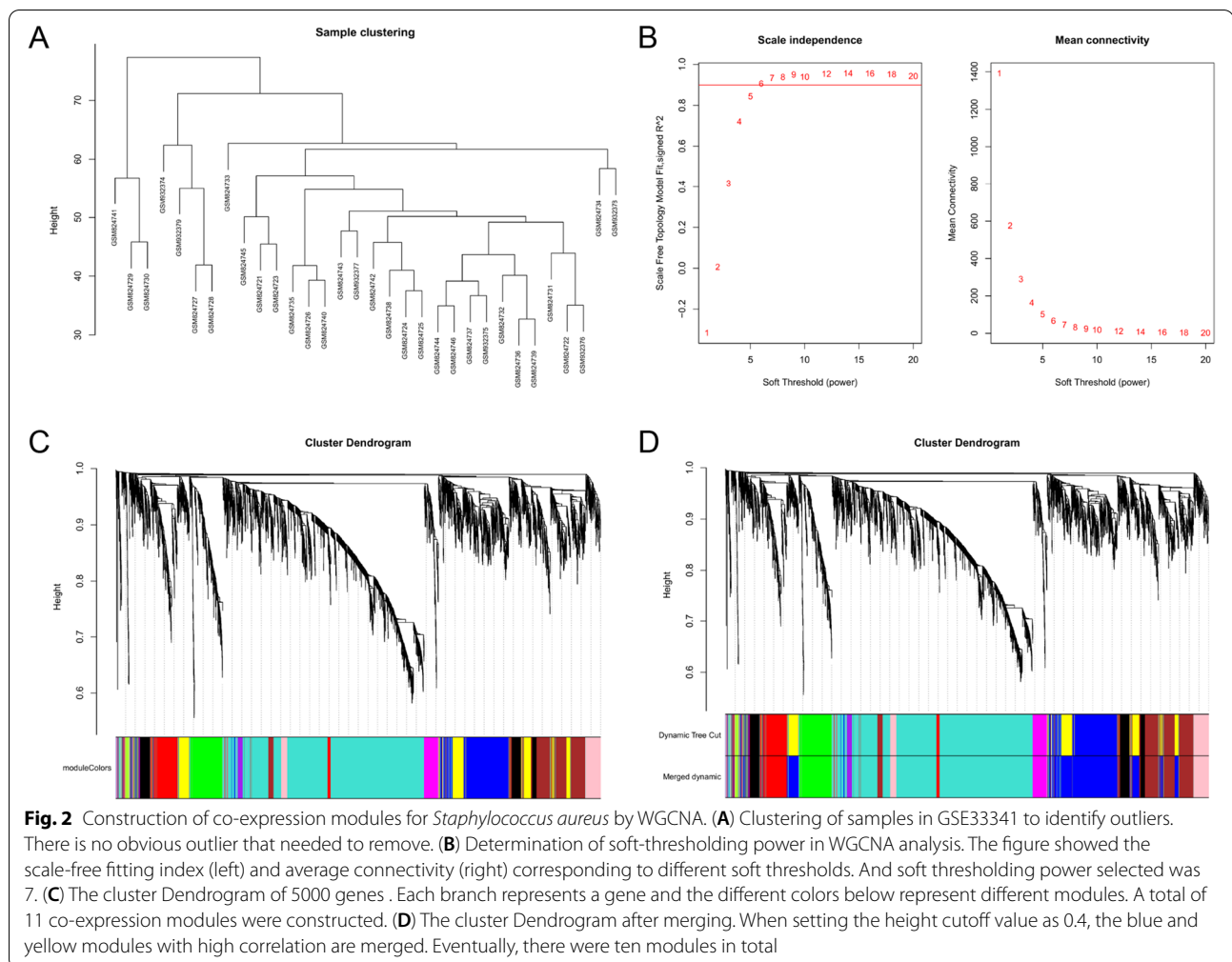
### Validation of hub genes in datasets

To ensure the result's rigor, we used two other datasets, GSE30119 [20] and GSE13670 [21], to verify the hub genes. In the GSE30119 dataset, the data of 99 infected samples and 44 healthy samples were extracted from whole blood and imported into GraphPad Prism (version 8.0.2) for t-tests and non-parametric tests. The hub genes with  $P < 0.05$  were considered significant. In the GSE13670 dataset, 15 infected blood samples and 15 healthy samples were utilized to plot ROC curves, from which we obtained their AUC through the "pROC" package. ROC curve is usually a helpful tool to evaluate the efficiency of gene diagnosis [22]. The hub

genes with  $AUC > 0.7$  were deemed useful for disease diagnosis.

### Gene set enrichment analysis

GSEA is an analytical method concentrating on groups of genes that share common biological functions and regulation [23]. To learn about the biological function, we performed the GSEA for the hub gene. Based on the median expression level of the hub gene, 32 infection samples were classified into the low-expression and the high-expression group. GSEA 4.1.0 was used to perform the analysis, and the c2.cp.kegg.v7.2.symbols.gmt in the Molecular Signatures Database (MSigDB) was selected as



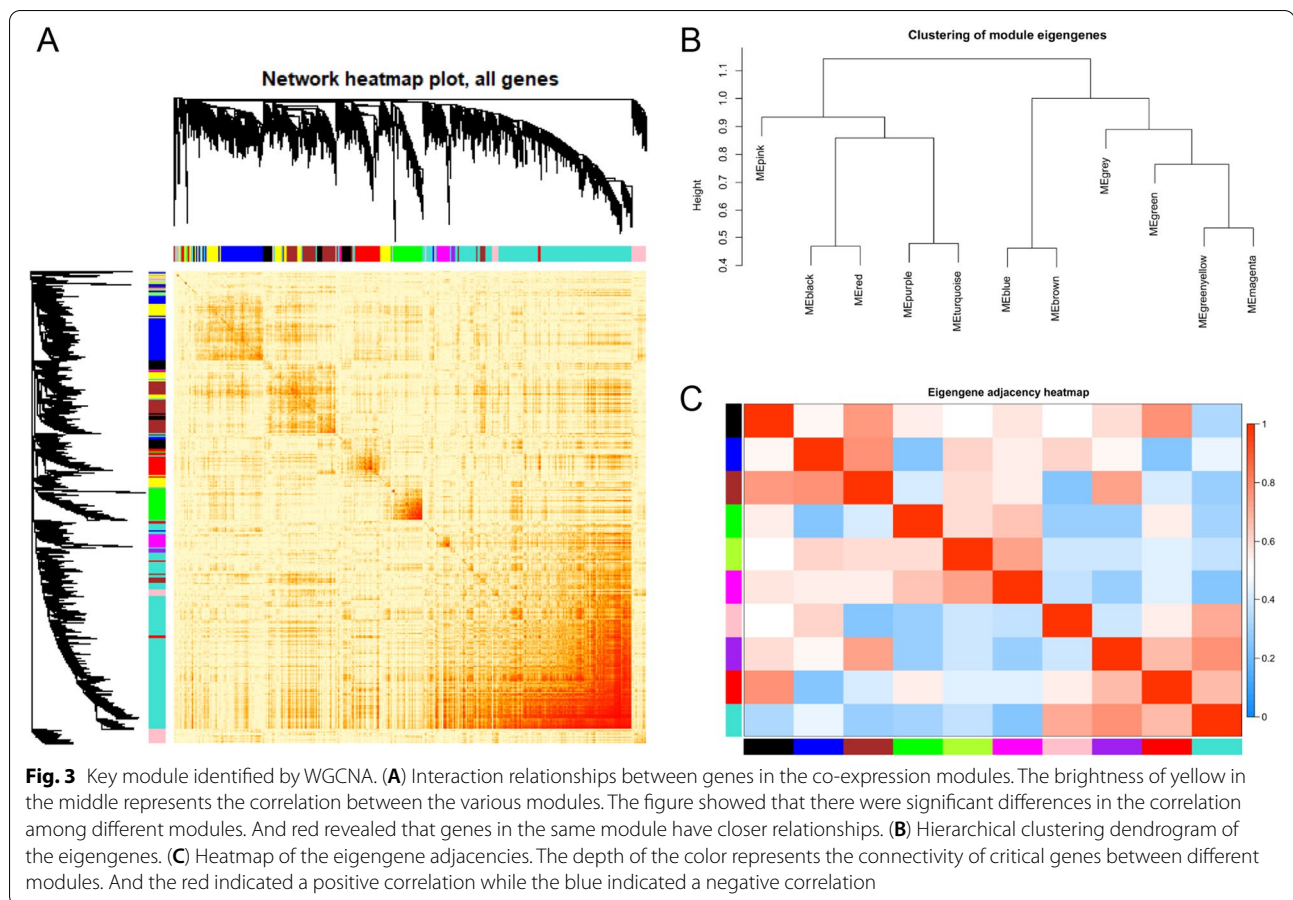
the reference gene set. The results met nominal  $p$ -value  $< 0.05$ , FDR  $< 25\%$  and normalized enrichment score  $> 1$  were considered statistically significant.

## Results

### Gene co-expression network construction and significant module identification

After preliminary screening, we screened out 13,345 genes from 32 infection samples to undergo WGCNA. The top 5000 genes with high expression values were screened out and used to construct the co-expression network. Through WGCNA analysis, 5000 genes were divided into 12 co-expression modules (Fig. 2C). Since yellow and blue modules had a high similarity, they were merged when MEDissThres was setting as 0.4. (Additional file 1) Therefore, there were a total of 11 modules, eventually (Fig. 2D). The turquoise module is the most extensive module and includes 1952 genes. The number of genes in the green-yellow module was the lowest, which only comprised 39 genes. A total of

92 genes that did not belong to any of the ten modules were classified as grey module. Next, we analyzed the interactions between genes in the co-expression modules. The results (Fig. 3A) revealed some noticeable differences in the correlation among other modules, especially the turquoise and green module, which aroused our attention and interest for further research. Moreover, we analyzed the connection between different co-expression modules by calculating the connectivity of eigengenes. As shown in Fig. 3B, 11 modules were divided into two clusters. One comprised pink, black, red, purple, and turquoise modules; the other included blue, brown, grey, green, green-yellow, and magenta modules. We observed that there were higher adjacencies between several pairs of modules from Fig. 3C, such as the brown and black, brown and blue, red and black, turquoise and purple modules.



### Functional enrichment analysis of genes in modules of interest

KEGG and GO were applied to obtain biological functions for the genes in the green and turquoise modules. According to the KEGG and GO enrichment analysis, the green module was defined as the critical module to undergo further analysis. The results of KEGG and GO analyses of the turquoise module were displayed in Additional file 2. Based on the KEGG analysis, mitophagy—animal and ubiquitin-mediated proteolysis were the most enriched terms in the green module (Fig. 4A). In terms of biological pathways, genes in the green module were enriched in the proteasomal protein catabolic process and myeloid cell differentiation catabolic process. (Fig. 4B) Concerning MF, genes were significantly enriched in ubiquitin-protein ligase binding (Fig. 4C). For CC, genes were involved in the vacuolar membrane. (Fig. 4D)

### PPI network analysis and hub genes identification

We built the PPI network of genes in the green module with the help of STRING. (Additional file 3) And

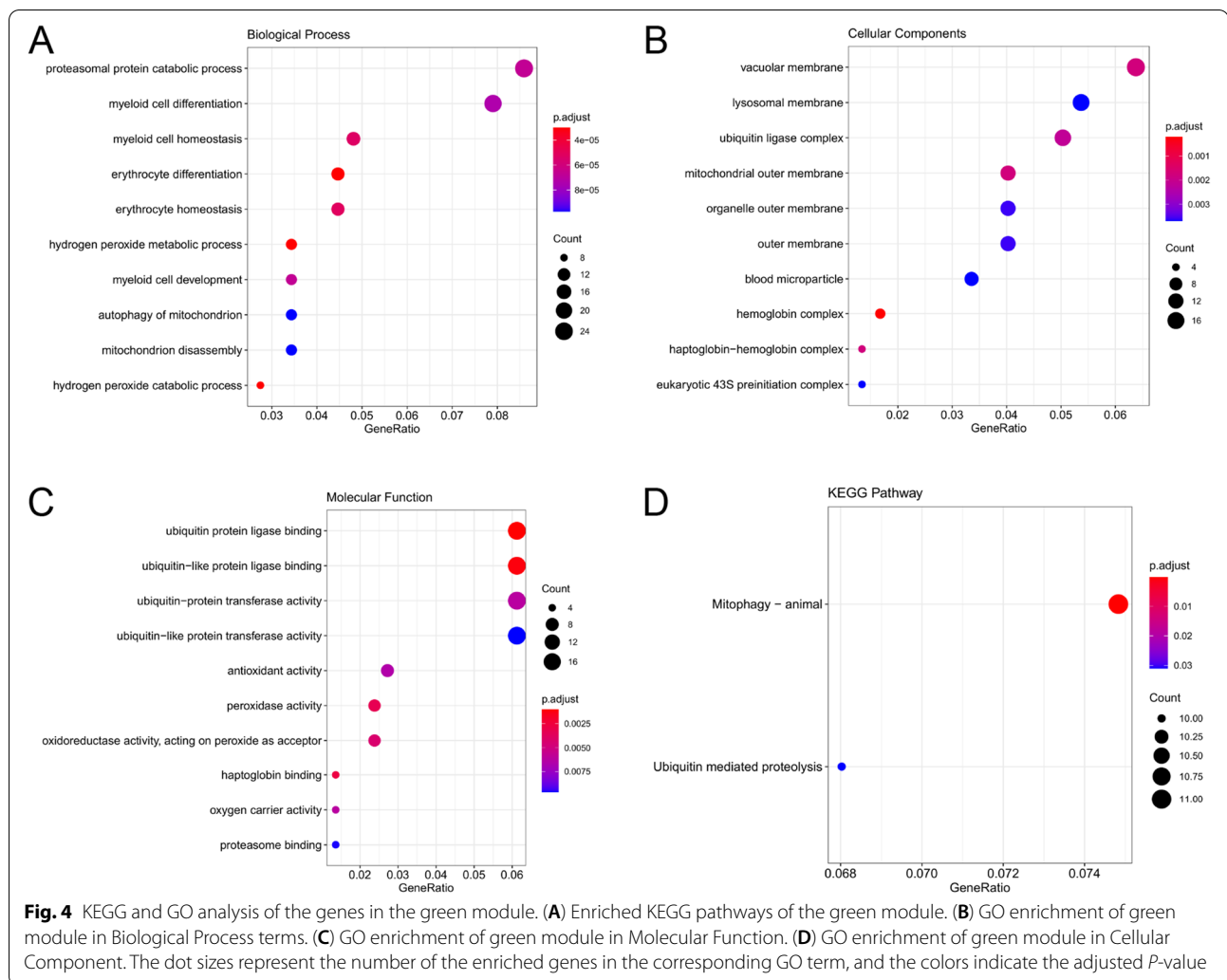
we identified the dense regions among the PPI network through the MCODE plugin. (Fig. 5A, B, C) Finally, we picked out the significant module with the highest score (score: 16), 16 nodes, and 120 edges from the green module (Fig. 5A). The genes with the top five MCC values in the final module were defined as hub genes (ASB1, CDC34, SKP1, MKRN1, and UBB) (Fig. 5D).

### Verification and efficacy evaluation of hub genes

In the GSE30119 dataset, with the standard of  $P < 0.05$ , UBB was defined as the hub gene (Fig. 5E). In the GSE13670 dataset, the area under the curve (AUC) of ASB1, UBB, and MKRN1 were both greater than 0.7, which suggested their potential diagnostic significance (Fig. 6A, B, C, and Additional file 4). In conclusion, we defined UBB as our hub gene to conduct further analysis in our study.

### Gene set enrichment analysis

After the GSEA for the final selected hub genes, we found that the KEGG P53 signaling pathway was associated with the samples with lower expression of UBB while KEGG porphyrin and chlorophyll metabolism was



associated with the samples with higher expression of UBB (Fig. 6D and E).

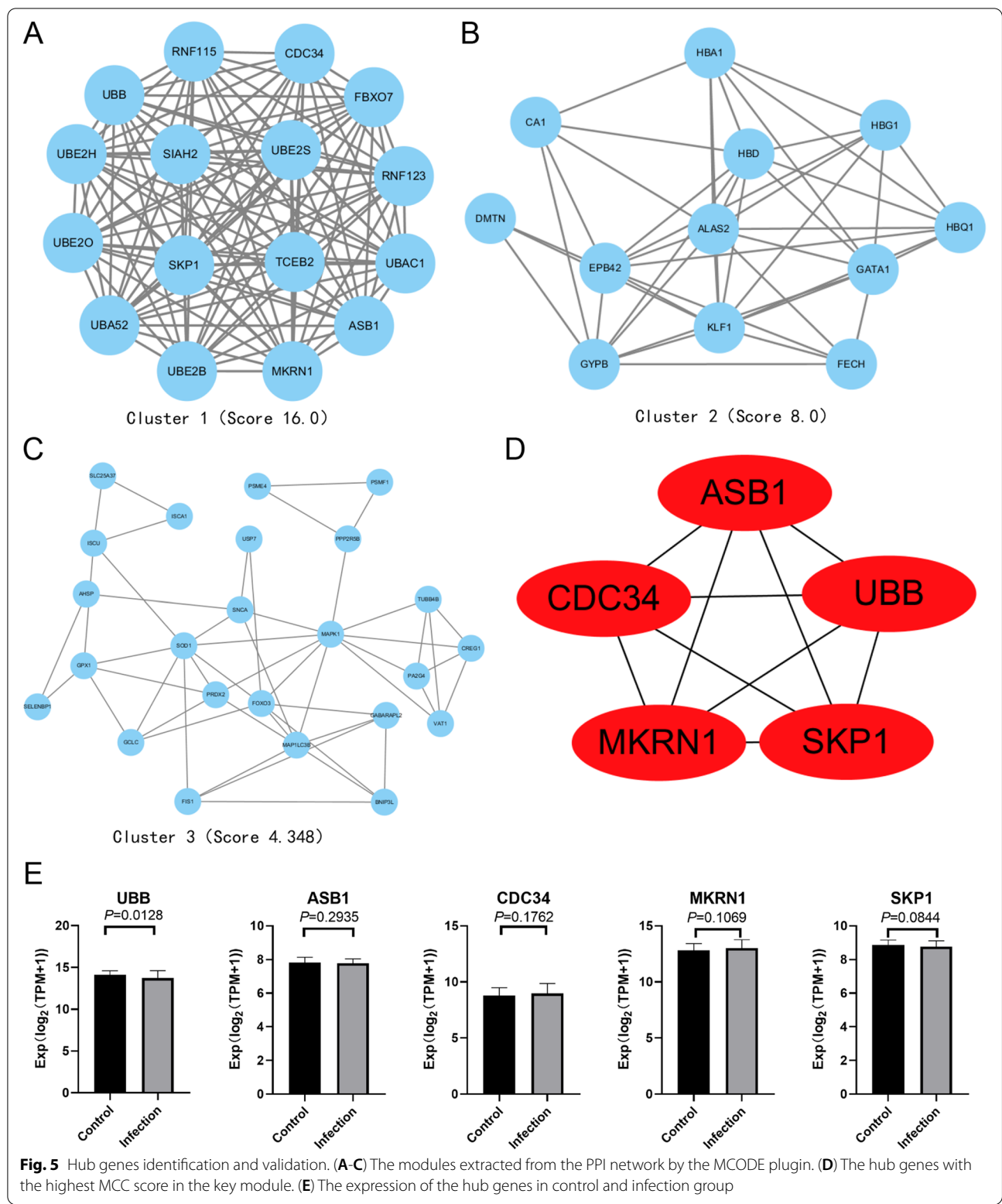
## Discussion

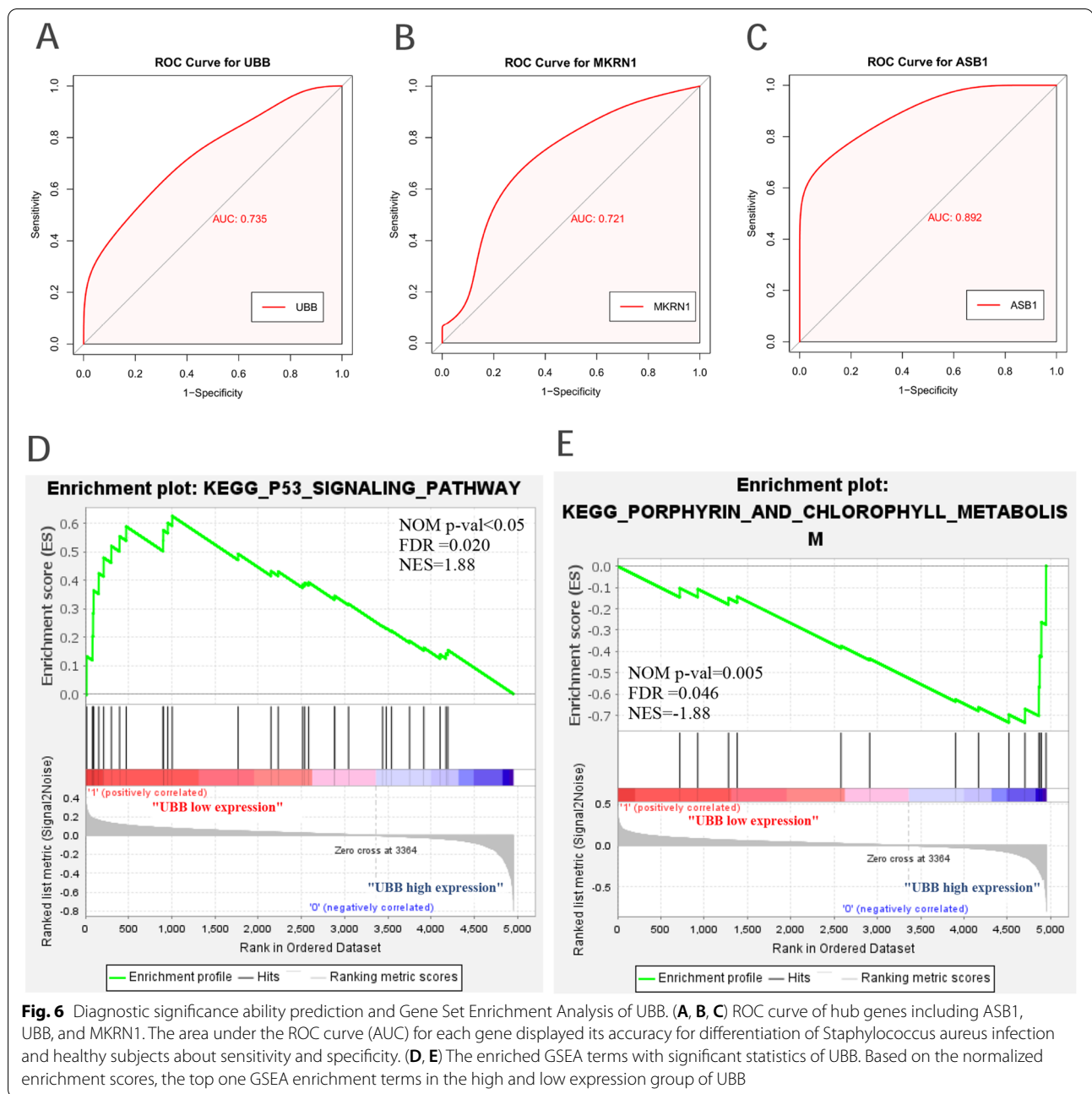
Multiple drug-resistant *Staphylococcus aureus* has become a public health issue, which renders the clinical treatment of the infection difficult [24]. We aimed to analyze the RNA transcriptional profiles and explore the highly correlated genes that participate in *Staphylococcus aureus* infection. In this study, a total of 5000 differentially expressed genes were constructed into 11 modules by WGCNA. Based on this, we applied bioinformatics methods to explore the characteristics of *Staphylococcus aureus* infection.

Through WGCNA analysis, we defined the green module as the key module and the genes in the green module were extracted for analysis. According to KEGG analysis, mitophagy and ubiquitin-mediated proteolysis were the most significant enrichment

results of the genes in the green module, both of which were closely associated with cell defense. Mitophagy is a metabolic process that participates in the removal of excessive or damaged mitochondria in eukaryotic cells, thus maintaining the stability of the intracellular microenvironment [25]. The previous study [26, 27] has proven that systemic sepsis or pneumonia caused by *Staphylococcus aureus* is associated with mitochondrial lung damage. Mitophagy is a strategy adopted by cells to eliminate damaged mitochondria and prevent their harmful byproducts, such as reactive oxygen species (ROS), from interfering with normal cell components like DNA and lipids [26, 28].

According to the GO analysis, the proteasomal protein catabolic process was one of the significant enrichment items in biological process. Proteasomal degradation was a crucial catabolic pathway in cells. And the up-regulation of this process contributes to activating the adaptive immune system and eliminating intracellular





pathogens [29]. In terms of molecular function, genes in the green module were also significantly enriched in ubiquitin–protein ligase binding, ubiquitin–like protein ligase binding, ubiquitin–protein transferase activity, and ubiquitin-like protein transferase activity. These all had a close connection with the ubiquitination process. Ubiquitination is a sequential process in which ubiquitin binds with target proteins via enzymatic cascade and degrades them through proteasome or lysosome pathways [30, 31]. It plays an essential role in the regulation

of immune responses and the defense against pathogens [32]. For example, Neumann, Yvonne, and Sakowski Erik reported that intracellular *Staphylococcus aureus* was ubiquitinated by the host cell shortly after the invasion and confused with lysosomes [33, 34]. In addition, ubiquitination is crucial in mitophagy. Damaged mitochondria were eliminated by binding to the ubiquitin ligase parkin [35].

Therefore, combining with KEGG, BP, and MF analysis, we concluded that the up-regulation of the protein



ubiquitination pathway and mitophagy might help defend against *Staphylococcus aureus* infection.

Concerning cellular components, the vacuolar membrane is the most notable enrichment result. Some studies reported that *Staphylococcus aureus* can survive phagocytosis by macrophages and neutrophils and replicate in the cell as a vacuolar pathogen [36, 37]. In addition, the vacuolar pathogen can escape from vacuolar and replicate in the cytosol by modifying the vacuolar membranes, leading to a vicious cycle of host phagocytosis, host cell death, and bacterial release [36]. Therefore, we inferred that block enzymes that modify vacuolar membranes, such as phospholipases, can help prevent bacterial escape.

Protein-protein interactions are crucial for knowing about biological processes in living cells [38]. We selected the top-five critical genes from the PPI network to investigate their biological function in *Staphylococcus aureus* infection. According to the ROC curve, ASB1, UBB, and MKRN1 can effectively act as potential diagnostic markers for distinguishing samples of *Staphylococcus aureus* infection from healthy counterparts. ASB1, a member of the ASB family, acted as a positive regulator of NF- $\kappa$ B- and MAPK-mediated inflammatory signaling pathways [39]. Appropriate inflammatory response helps immune cells fight microbial infections while excessive inflammatory response can damage tissues and cells [40]. The study pointed that ASB1 deficiency protected mice from LPS- or bacteria-induced death by inhibiting inflammation [39]. Therefore, we speculated that inhibition and down-regulation of ASB1 in the late stage of infection can against the inflammatory injury induced by *Staphylococcus aureus*. Makorin ring finger protein 1 (MKRN1) is a ubiquitin ligase, which is vital against viral pathogens [41]. Some studies displayed that the high expression of MKRN1 can help host cells to defend against infection by inducing ubiquitination of ppar $\gamma$  that is beneficial to *M. tuberculosis* growth [41]. However, there are few studies about the role of MKRN1 in the *Staphylococcus aureus*. Through the verification of the dataset GSE30119, UBB was also identified as a hub gene. UBB was known to be a vital gene that encodes ubiquitin, one of the most conserved proteins that play a significant role in targeting cellular proteins for degradation by the 26S proteasome [42]. GSEA results revealed that the group with a low expression of UBB was closely associated with the p53 signaling pathway in *Staphylococcus aureus* infection. P53 is well known for its anti-cancer function and regulation of autophagy [43–45]. Abundant evidence revealed that cytosolic p53 could also disturb the process of mitophagy through inhibitory interaction with Parkin, a ubiquitin E3 ligase [43, 44] Zhang, Fei pointed

out that the upregulation of p53 inhibited mitochondrial translocation of Parkin and activation of Parkin's E3 ubiquitin ligase, which eventually stopped cells from effectively removing damaged mitochondria [46]. Therefore, we speculate that this was a strategy for bacteria to avoid death.

In this study, we found that ubiquitination and mitophagy play important roles in defending against *Staphylococcus aureus* infection. In addition, we also identified some hub genes which act as regulators in *Staphylococcus aureus* infection. However, more molecular biological experiments will be needed to confirm the function of the identified genes.

### Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12866-021-02392-y>.

**Additional file 1.**

**Additional file 2.**

**Additional file 3.**

**Additional file 4.**

### Acknowledgments

We sincerely acknowledge the GEO database for offering their platform and their contributions for uploading their valuable dataset.

### Authors' contributions

Xu-guang Guo conceived and designed the experiment. Jia-xin Li, Xun-jie Cao, and Yuan-yi Huang conducted the data analysis and made the figures. Other people participated in the writing and revising of the manuscript.

### Availability of data and materials

Microarray datasets (GSE33341, GSE13670 and GSE30119) for this study are openly available in Gene Expression Omnibus database at <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE33341>, <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE13670>, and <https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE30119>, respectively.

### Declarations

#### Ethics approval and consent to participate

Not applicable for that section.

#### Consent for publication

Not applicable for that section.

#### Competing interests

The authors reports no conflict of interest.

#### Author details

<sup>1</sup>Department of Clinical Laboratory Medicine, The Third Affiliated Hospital of Guangzhou Medical University, Guangzhou 510150, China. <sup>2</sup>Department of Clinical Medicine, The First Clinical School of Guangzhou Medical University, Guangzhou 511436, China. <sup>3</sup>Department of Clinical Medicine, The Third Clinical School of Guangzhou Medical University, Guangzhou 511436, China. <sup>4</sup>Department of Clinical Medicine, The Second Clinical School of Guangzhou Medical University, Guangzhou 511436, China. <sup>5</sup>Department of Traditional Chinese and Western Clinical Medicine, The Traditional Chinese and Western Clinical School of Guangzhou Medical University, Guangzhou 511436, China. <sup>6</sup>Key Laboratory for Major Obstetric Diseases of Guangdong Province, The

Third Affiliated Hospital of Guangzhou Medical University, Guangzhou 510150, China.

Received: 14 August 2021 Accepted: 10 November 2021  
Published online: 01 December 2021

## References

- Redi D, Raffaelli CS, Rossetti B, De Luca A, Montagnani F. Staphylococcus aureus vaccine preclinical and clinical development: current state of the art. *New Microbiol.* 2018;41(3):208–13.
- Chang J, Lee RE, Lee W. A pursuit of Staphylococcus aureus continues: a role of persister cells. *Arch Pharm Res.* 2020;43(6):630–8.
- Melehan JH, Duncan JA. Inflammasome activation can mediate tissue-specific pathogenesis or protection in Staphylococcus aureus infection. *Curr Top Microbiol Immunol.* 2016;397:257–82.
- Narita K, Asano K, Nakane A. IL-17A plays an important role in protection induced by vaccination with fibronectin-binding domain of fibronectin-binding protein a against Staphylococcus aureus infection. *Med Microbiol Immunol.* 2017;206(3):225–34.
- Stokes W, Parkins MD, Parfitt E, Ruiz JC, Mugford G, Gregson DB. Incidence and outcomes of Staphylococcus aureus bacteriuria: a population-based study. *Clinical infectious diseases: an official publication of the Infectious Diseases Society of America.* 2019;69(6):963–9.
- Sethi S, Chakraborty T. Role of TLR- / NLR-signaling and the associated cytokines involved in recruitment of neutrophils in murine models of Staphylococcus aureus infection. *Virulence.* 2011;2(4):316–28.
- Nair N, Biswas R, Götz F, Biswas L. Impact of Staphylococcus aureus on pathogenesis in polymicrobial infections. *Infect Immun.* 2014;82(6):2162–9.
- Cheung G, Bae JS, Otto M. Pathogenicity and virulence of Staphylococcus aureus. *Virulence.* 2021;12(1):547–69.
- Brady RA, Mocca CP, Plaut RD, Takeda K, Burns DL. Comparison of the immune response during acute and chronic Staphylococcus aureus infection. *PLoS One.* 2018;13(3):e0195342.
- Reilly J, Jacobs MA, Friedman B, Cleveland KO, Lombardi DA, Castaneda-Ruiz B. Clinical experience with Telavancin for the treatment of patients with bacteremia and endocarditis: real-world results from the Telavancin observational use registry (TOUR™). *Drugs - Real World Outcomes.* 2020;7(3):179–89.
- Xu C, Zhang BZ, Lin Q, Deng J, Yu B, Arya S, et al. Live attenuated salmonella typhimurium vaccines delivering SaEsxA and SaEsxB via type III secretion system confer protection against Staphylococcus aureus infection. *BMC Infect Dis.* 2018;18(1):195.
- Zeng H, Yang F, Feng Q, Zhang J, Gu J, Jing H, et al. Rapid and broad immune efficacy of a recombinant five-antigen vaccine against staphylococcus aureus infection in animal models. *Vaccines.* 2020;8(1):134.
- Hn SH, Tsalik EL, Cyr DD, Zhang Y, van Velkinburgh JC, Langley RJ, et al. Gene expression-based classifiers identify Staphylococcus aureus infection in mice and humans. *PLoS One.* 2013;8(1):e48979.
- Xie X, Zhao J, Xie L, Wang H, Xiao Y, She Y, et al. Identification of differentially expressed proteins in the injured lung from zinc chloride smoke inhalation based on proteomics analysis. *Respir Res.* 2019;20(1):36.
- Kanehisa M, Goto S. KEGG: Kyoto encyclopedia of genes and genomes. *Nucleic Acids Res.* 2000;28(1):27–30.
- Kanehisa M. Toward understanding the origin and evolution of cellular organisms. *Protein Sci Publ Protein Soc.* 2019;28(11):1947–51.
- Kanehisa M, Furumichi M, Sato Y, Ishiguro-Watanabe M, Tanabe M. KEGG: integrating viruses and cellular organisms. *Nucleic Acids Res.* 2021;49(D1):D545–51.
- Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, et al. Gene ontology: tool for the unification of biology. *Gene Ontol Consort Nature Gen.* 2000;25(1):25–9.
- Vella D, Marini S, Vitali F, Di Silvestre D, Mauri G, Bellazzi R. MTGO: PPI network analysis via topological and functional module identification. *Sci Rep.* 2018;8(1):5499.
- Banchereau R, Jordan-Villegas A, Ardura M, Mejias A, Baldwin N, Xu H, et al. Host immune transcriptional profiles reflect the variability in clinical disease manifestations in patients with Staphylococcus aureus infections. *PLoS One.* 2012;7(4):e34390.
- Koziel J, Maciag-Gudowska A, Mikolajczyk T, Bzowska M, Sturdevant DE, Whitney AR, et al. Phagocytosis of Staphylococcus aureus by macrophages exerts cytoprotective effects manifested by the upregulation of antiapoptotic factors. *PLoS One.* 2009;4(4):e5210.
- Zhou C, Yu J, Wang M, Yang J, Xiong H, Huang H, et al. Identification of glycerol-3-phosphate dehydrogenase 1 as a tumour suppressor in human breast cancer. *Oncotarget.* 2017;8(60):101309–24.
- Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A.* 2005;102(43):15545–50.
- Liu, X., Mao, Y., Kang, Y., He, L., Zhu, B., Zhang, W., Lu, Y., Wu, Q., Xu, D., & Shi, L. (2020). MicroRNA-127 promotes anti-microbial host defense through restricting A20-mediated De-ubiquitination of STAT3. *iScience*, 23(1), 100763.
- Zhao Y, Huang S, Liu J, Wu X, Zhou S, Dai K, et al. Mitophagy contributes to the pathogenesis of inflammatory diseases. *Inflammation.* 2018;41(5):1590–600.
- Chang AL, Ulrich A, Suliman HB, Piantadosi CA. Redox regulation of mitophagy in the lung during murine Staphylococcus aureus sepsis. *Free Radic Biol Med.* 2015;78:179–89.
- Suliman HB, Kraft B, Bartz R, Chen L, Welty-Wolf KE, Piantadosi CA. Mitochondrial quality control in alveolar epithelial cells damaged by S aureus pneumonia in mice. *Am J Physiol Lung Cell Mol Physiol.* 2017;313(4):L699–709.
- Ashrafi G, Schwarz TL. The pathways of mitophagy for quality control and clearance of mitochondria. *Cell Death Differ.* 2013;20(1):31–42.
- Gannagé M, Münz C. Macroautophagy in immunity and tolerance. *Traffic (Copenhagen, Denmark).* 2009;10(6):615–20.
- Lawrence DW, Willard PA, Cochran AM, Matchett EC, Kornbluth J. Natural killer lytic-associated molecule (NKLAM): an E3 ubiquitin ligase with an integral role in innate immunity. *Front Physiol.* 2020;11:573372.
- Claessen J, Witte MD, Yoder NC, Zhu AY, Spooner E, Ploegh HL. Catch-and-release probes applied to semi-intact cells reveal ubiquitin-specific protease expression in *Chlamydia trachomatis* infection. *Chembiochem: Eur J Chem Biol.* 2013;14(3):343–52.
- Genini S, Badaoui B, Sclep G, Bishop SC, Waddington D, Pinard van der Laan MH, et al. Strengthening insights into host responses to mastitis infection in ruminants by combining heterogeneous microarray data sources. *BMC Genomics.* 2011;12(1):225.
- Neumann Y, Bruns SA, Rohde M, Prajsnar TK, Foster SJ, Schmitz I. Intracellular Staphylococcus aureus eludes selective autophagy by activating a host cell kinase. *Autophagy.* 2016;12(11):2069–84.
- Sakowski ET, Koster S, Portal Celhay C, Park HS, Shrestha E, Hetzencker SE, et al. Ubiquitin 1 promotes IFN-γ-induced Xenophagy of mycobacterium tuberculosis. *PLoS Pathog.* 2015;11(7):e1005076.
- Manzanillo PS, Ayres JS, Watson RO, Collins AC, Souza G, Rae CS, et al. The ubiquitin ligase parkin mediates resistance to intracellular pathogens. *Nature.* 2013;501(7468):512–6.
- Bianchi F, van den Bogaart G. Vacuolar escape of foodborne bacterial pathogens. *J Cell Sci.* 2020;134(5):jcs247221.
- Tranchemontagne ZR, Camire RB, O'Donnell VJ, Baugh J, Burkholder KM. Staphylococcus aureus strain USA300 perturbs Acquisition of Lysosomal Enzymes and Requires Phagosomal Acidification for survival inside macrophages. *Infect Immun.* 2015;84(1):241–53.
- Lin, J. S., & Lai, E. M. (2017). Protein-protein interactions: co-immunoprecipitation. *Methods in molecular biology (Clifton, N.J.)*, 1615, 211–219.
- Hou P, Jia P, Yang K, Li Z, Tian T, Lin Y, et al. An unconventional role of an ASB family protein in NF-κB activation and inflammatory response during microbial infection and colitis. *Proc Natl Acad Sci U S A.* 2021;118(3):e2015416118.
- Wang H, Bi C, Wang Y, Sun J, Meng X, Li J. Selenium ameliorates Staphylococcus aureus-induced inflammation in bovine mammary epithelial cells by inhibiting activation of TLR2, NF-κB and MAPK signaling pathways. *BMC Vet Res.* 2018;14(1):197.
- Subrahmanian M, Marimuthu J, Sairam T, Sankaran R. In vitro ubiquitination of mycobacterium tuberculosis by E3 ubiquitin ligase, MKRN1. *Biotechnol Lett.* 2020;42(8):1527–34.
- Tian Y, Ding W, Wang Y, Ji T, Sun S, Mo Q, et al. Ubiquitin B in cervical cancer: critical for the maintenance of cancer stem-like cell characters. *PLoS One.* 2013;8(12):e84457.

43. Zheng R, Yao Q, Xie G, Du S, Ren C, Wang Y, et al. TAT-ODD-p53 enhances the radiosensitivity of hypoxic breast cancer cells by inhibiting Parkin-mediated mitophagy. *Oncotarget*. 2015;6(19):17417–29.
44. Song YM, Lee WK, Lee YH, Kang ES, Cha BS, Lee BW. Metformin restores Parkin-mediated Mitophagy, suppressed by cytosolic p53. *Int J Mol Sci*. 2016;17(1):122.
45. Nakamizo A, Amano T, Zhang W, Zhang XQ, Ramdas L, Liu TJ, et al. Phosphorylation of Thr18 and Ser20 of p53 in ad-p53-induced apoptosis. *Neuro-oncology*. 2008;10(3):275–91.
46. Zhang F, Peng W, Zhang J, Dong W, Wu J, Wang T, et al. P53 and Parkin co-regulate mitophagy in bone marrow mesenchymal stem cells to promote the repair of early steroid-induced osteonecrosis of the femoral head. *Cell Death Dis*. 2020;11(1):42.

### Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Ready to submit your research? Choose BMC and benefit from:**

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

**At BMC, research is always in progress.**

Learn more [biomedcentral.com/submissions](https://biomedcentral.com/submissions)

