

#### PLASTOME ANNOUNCEMENT

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# The complete chloroplast genome of ornamental and medicinal *Callerya dielsiana* (Fabaceae)

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#### **ABSTRACT**

Callerya dielsiana is a Chinese endemic tropical/subtropical liana. We sequenced the complete chloroplast genome with the Illumina Hiseq X-Ten platform. The genome is obtained with 132,301 bp in length, lacking an inverted repeat (IR) region, contains 4 rRNAs, 30 tRNAs genes, and 76 protein-coding genes. The overall GC content is 33.9%. Based on the whole chloroplast genomes of 14 legume species, a phylogenetic tree is constructed and indicated that *C. dielsiana* belongs to the well-supported tribe Wisterieae. The tribe is sister to *Glycyrrhiza* and nested within the IRLC (Inverted Repeat-Lacking Clade) group of the subfamily Papilionoideae (Fabaceae).

#### ARTICLE HISTORY

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#### **KEYWORDS**

Callerya dielsiana; chloroplast genome; tropical/subtropical liana

The tropical/subtropical leguminous species *Callerya dielsiana* (Harms ex Diels) P.K.Lôc ex Wei and Pedley (2010) is an ornamental woody liana species endemic to southern China (Compton et al. 2019; Duan et al. 2021), which can also be used as medicinal plant to promote blood circulation and to dissipate blood stasis (Song et al. 1992; Gong 2010). Few study focused on genome of *C. dielsiana*, a better genomics knowledge of this species would benefit the future works on population genetics, diversity and gardening.

The fresh leaves of C. dielsiana was collected in Tianjing Mt., Guangdong Province, China (24°41′24″E, 113°3′36″N), and the voucher specimen was deposited at South China Botanical Garden, Chinese Academy of Science (IBSC; http:// english.scbg.ac.cn/, Shi-Xiao Luo, luoshixiao@scbg.ac.cn) under the voucher number L.Duan 2016022. The total genomic DNA was extracted using CTAB approach (Doyle 1987), the cDNA library was prepared and sequenced with the Illumina Hiseg X-Ten platform (Illumina Inc., San Diego, CA). The sequences were filtered following the protocol of Yao et al. (2016), the resultant adaptor-free reads were then assembled with SPAdes 3.11 (Bankevich et al. 2012). The program of Dual Organellar GenoMe Annotator (DOGMA; Wyman et al. 2004) was used to annotate the assembly of complete chloroplast (cp) genome, which has been deposited in GenBank (accession number: MW007722). Raw reads were also deposited in the GenBank (SRA: SRR13871830; BioProject: PRJNA706880; Bio-Sample: SAMN18131232).

About 1.97 Gb raw reads of *C. dielsiana* were obtained. The cp genome, with 132,301 bp in length and lacking one inverted repeat (IR) region, contained 4 ribosomal RNA genes

(rRNA), 30 transfer RNA genes (tRNA), and 76 protein-coding genes (CDS). Within the cp genome, we found 15 one-intron genes (atpF, clpP, ndhA, ndhB, petB, petD, rpl2, rpl16, rpoC1, trnA-UGC, trnG-UCC, trnl-GAU, trnK-UUU, trnL-UAA, and trnV-UAC), and two two-intron genes (rps12 and ycf3). Overall GC content of the whole genome was 33.9%.

To infer the phylogenetic relationships among *C. dielsiana* and its related taxa, whole cp genomes of 13 Papilionoideae species were downloaded from GenBank, which were aligned with that of *C. dielsiana* by applying MAFFT v.7 (Katoh and Standley 2013). We constructed a maximum-likelihood (ML) tree taking TIM2 as model, based on the alignment using IQ-TREE v.1.6. The result (Figure 1) showed that *Callerya*, *Sarcodum*, *Wisteria* were members of the monophyletic tribe Wisterieae (as in Compton et al. 2019; Duan et al. 2021). Among them, *Callerya dielsiana* and *C. nitida* were sister to *Wisteria*, and together formed a monophytic clade with *Sarcodum*. Wisterieae was nested in the inverted repeat-lacking clade (IRLC), which in turn belonged to the Hologalegina group of the subfamily Papilionoideae (see Wojciechowski et al. 2004; Schrire 2005).

## **Ethical approval**

This study includes no human, animal, or endangered plant samples, and the sample was legally collected in accordance with guidelines provided by the authors' institution and national or international regulations. Field studies were complied with local legislation. No ethical approval/permission is required in this study.

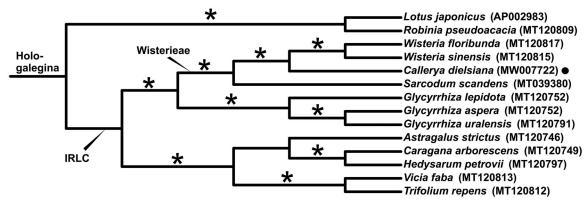


Figure 1. Maximum-likelihood (ML) phylogenetic tree based on 14 chloroplast genomes of Fabaceae. The position of Callerya dielsiana is indicated with black dot. The bootstrap values of 100% are shown on branches with asterisks.

### **Author contributions**

Qi-Fei Yi and Hong-Feng Chen involved in the conception, design and financial support; Ming-Chen Lin and Lin Fu collected the sample; Yong Tan and Lei Duan analyzed the data; Lei Duan drafted the paper, Li-Na Han revised the manuscript, and Qi-Fei Yi final approved the version to be published. All authors agree to be accountable for all aspects of the work.

#### Disclosure statement

No potential conflict of interest was reported by the author(s).

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## Data availability statement

The genome sequence data that support the findings of this study are openly available in GenBank of NCBI at [https://www.ncbi.nlm.nih.gov] (https://www.ncbi.nlm.nih.gov/) under the accession no. MW007722. The associated BioProject, SRA, and Bio-Sample numbers are PRJNA706880, SRR13871830, and SAMN18131232, respectively.

#### References

Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, et al. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol. 19(5):455-477.

Compton JA, Schrire BD, Könyves K, Forest F, Malakasi P, Mattapha S, Sirichamorn Y. 2019. The Callerya Group redefined and Tribe Wisterieae (Fabaceae) emended based on morphology and data from nuclear and chloroplast DNA sequences. PhytoKeys. 125:1-112.

Doyle JJ. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. Phytochem Bull. 19:11-15.

Duan L, Li SJ, Su C, Sirichamorn Y, Han LN, Ye W, Loc PK, Wen J, Compton JA, Schrire B, et al. 2021. Phylogenomic framework of the IRLC legumes (Leguminosae subfamily Papilionoideae) and intercontinental biogeography of tribe Wisterieae. Mol Phylogenet Evol. 163:

Gong T. 2010. Studies on the chemical constituents and bioactivities of Mucuna birdwoodiana Tutch and Millettia dielsiana Harms [PhD dissertation]. Beijing (China): Chinese Academy of Medical Sciences & Peking Union Medical College.

Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol. 30(4):772-780.

Schrire BD. 2005. Millettieae. In: Lewis G, Schrire B, Mackinder B, Lock M, editors. Legumes of the world. Richmond: Royal Botanic Gardens, Kew; p. 367-387.

Song JX, Hu WY, Luo SD. 1992. Studies on the chemical constituents of the stem of Millettia dielsiana Harms ex Diels. J Southwest Forest Coll. 12(1):40-43.

Wei Z, Pedley L. 2010. Callerya. In Wu ZY, Raven PH, Hong DY, editors. Flora of China. Vol. 10. Beijing (China): Science Press; p. 181-187.

Wojciechowski MF, Lavin M, Sanderson MJ. 2004. A phylogeny of legumes (Leguminosae) based on analysis of the plastid matK gene resolves many well-supported subclades within the family. Am J Bot. 91(11):1846-1862.

Wyman SK, Jansen RK, Boore JL, 2004. Automatic annotation of organellar genomes with DOGMA. Bioinformatics. 20(17):3252-3255.

Yao X, Tan YH, Liu YY, Song Y, Yang JB, Corlett RT. 2016. Chloroplast genome structure in *llex* (Aquifoliaceae). Sci Rep. 6:28559.