



Dosimetric Analysis of a Phase I Study of PSMA-Targeting Radiopharmaceutical Therapy With [^{177}Lu]Ludotadipep in Patients With Metastatic Castration-Resistant Prostate Cancer

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Objective: ^{177}Lu Lutetium [Lu] Ludotadipep is a novel prostate-specific membrane antigen targeting therapeutic agent with an albumin motif added to increase uptake in the tumors. We assessed the biodistribution and dosimetry of [^{177}Lu]Ludotadipep in patients with metastatic castration-resistant prostate cancer (mCRPC).

Materials and Methods: Data from 25 patients (median age, 73 years; range, 60–90) with mCRPC from a phase I study with activity escalation design of single administration of [^{177}Lu]Ludotadipep (1.85, 2.78, 3.70, 4.63, and 5.55 GBq) were assessed. Activity in the salivary glands, lungs, liver, kidneys, and spleen was estimated from whole-body scan and abdominal SPECT/CT images acquired at 2, 24, 48, 72, and 168 h after administration of [^{177}Lu]Ludotadipep. Red marrow activity was calculated from blood samples obtained at 3, 10, 30, 60, and 180 min, and at 24, 48, and 72 h after administration. Organ- and tumor-based absorbed dose calculations were performed using IDAC-Dose 2.1.

Results: Absorbed dose coefficient (mean $\pm$ standard deviation) of normal organs was 1.17 ± 0.81 Gy/GBq for salivary glands, 0.05 ± 0.02 Gy/GBq for lungs, 0.14 ± 0.06 Gy/GBq for liver, 0.77 ± 0.28 Gy/GBq for kidneys, 0.12 ± 0.06 Gy/GBq for spleen, and 0.07 ± 0.02 Gy/GBq for red marrow. The absorbed dose coefficient of the tumors was 10.43 ± 7.77 Gy/GBq.

Conclusion: [^{177}Lu]Ludotadipep is expected to be safe at the dose of 3.7 GBq times 6 cycles planned for a phase II clinical trial with kidneys and bone marrow being the critical organs, and shows a high tumor absorbed dose.

Keywords: PSMA; mCRPC; Radiopharmaceutical therapy; Dosimetry; Albumin

INTRODUCTION

The prostate-specific membrane antigen (PSMA) is a transmembrane protein highly expressed in prostate cancer cells [1]. PSMA functions as a folate hydrolase and

glutamate carboxypeptidase, and although it is associated with more aggressive prostate cancer, its role in prostate cancer progression is not completely understood [2,3]. Numerous glutamate-ureido-lysine (GUL)-based compounds that bind with high affinity to the substrate recognition

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site of PSMA have been developed for imaging and therapy. Following clinical trials demonstrating improved survival outcomes, ^{177}Lu Lutetium [Lu]Lu-labeled PSMA-targeting radiopharmaceuticals are now a treatment option for patients with metastatic castration-resistant prostate cancer (mCRPC).

Dosimetry of the commercially available [^{177}Lu]Lu-PSMA-617 showed rapid clearance from the blood pool as could be expected from small molecules, with a mean total body time-integrated activity coefficient of 37.9 ± 14.6 hours (h) reported [4,5]. Up to 6 cycles were administered in clinical trials that demonstrated clinical benefit of [^{177}Lu]Lu-PSMA-617 (7.4 GBq in the VISION trial and 6.0–8.5 GBq in the TheraP trial) [6,7]. To overcome the limitations of small-molecule radiopharmaceutical compounds that wash out quickly, various albumin-binding PSMA targets have been developed to prolong circulation time [8–13]. [^{177}Lu]Ludotadipep is a novel PSMA inhibitor labeled with [^{177}Lu]Lu, and its 4-(4-iodophenyl) butanoly moiety functions as an albumin binder to extend the blood circulation time (Fig. 1). In the PC3-PIP animal model, tumor uptake increased over 72 h, and tumor growth was inhibited in mice treated with 4 or 6 MBq of [^{177}Lu]Ludotadipep [14].

This study aimed to assess the biodistribution and dosimetry of single administration of [^{177}Lu]Ludotadipep in patients with mCRPC.

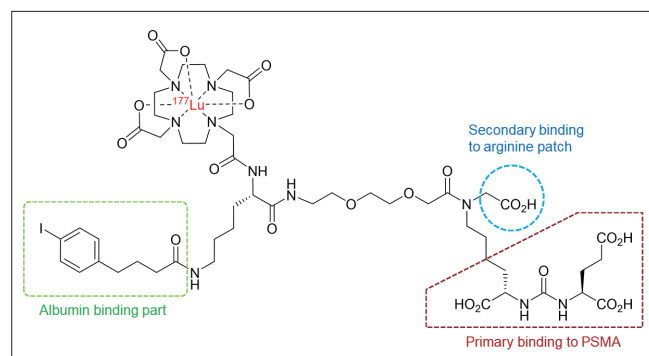


Fig. 1. [^{177}Lu]Ludotadipep structure. [^{177}Lu]Lutetium(III) 2,2',2''-(10-((4S,20S,24S)-20,24,26-tricarboxy-15-(carboxymethyl)-4-(4-(4-iodophenyl)butanamido)butyl)-2,5,14,22-tetraoxo-9,12-dioxo-3,6,15,21,23-pentaazahexacosyl)-1,4,7,10-tetraazacyclododecane-1,4,7-triyl)triacetate. The iodophenylbutanoly group (in green box) acts as an albumin binder, and increases circulation time and target uptake of the radiopharmaceutical. PSMA = prostate-specific membrane antigen

MATERIALS AND METHODS

Study Design

This study was approved by the hospital's Institutional Review Board (IRB No. KC20MDSF0483) and the Korean Ministry of Food and Drug Safety (KMFDS). Written informed consent was obtained from all participants and the study was conducted in compliance with the Declaration of Helsinki and local regulations (ClinicalTrials.gov Identifier: NCT04509557). In this phase I clinical trial, patients with mCRPC (blood testosterone level < 50 ng/dL) who progressed after standard treatment were enrolled [15]. [^{177}Lu]Ludotadipep doses of 1.85, 2.78, 3.70, 4.63, and 5.55 GBq were administered once in each subject and safety assessments were made at 2-, 3-, 4-, 6-, 8-, and 12-weeks post-injection (p.i.). In this activity escalation design, patient enrollment for the next dose began only if dose-limiting toxicity (grade 4 thrombocytopenia, grade 4 neutropenia, grade 3 febrile neutropenia, and other grade 3 or 4 non-hematological toxicities persisting for 5 days or more) was observed at 8 weeks in two or less of each group of six subjects.

Subjects were screened with [^{18}F]Florastamin PSMA PET/CT [16] for lesions with PSMA-reporting and data systems (RADS)-4 or 5 [17]. All screened patients had a history of disease progression following taxane-based chemotherapy and androgen receptor-targeted agents. A total of six subjects were included in each of the five dose groups, but one did not receive [^{177}Lu]Ludotadipep and four subjects did not have p.i. images for assessment ($n = 4$ due to COVID19 restrictions, and $n = 1$ for emergency surgery), and a total of 25 subjects were assessed for biodistribution and dosimetry.

Radiopharmaceuticals

[^{18}F]Florastamin was manufactured using an automatic synthesizer as previously described [16]. A good manufacturing practice-grade lutetium no-carrier-added product (Isotopia Molecular Imaging) approved by the Food and Drug Administration and European Medicines Agency was used for the radiolabeling of [^{177}Lu]Ludotadipep. [^{177}Lu]LuCl₃ solution was added to a reaction vial containing 1M NaOAc/HCl buffer (pH 4.88), L-ascorbic acid, and the Ludotadipep precursor solution. The reaction mixture was heated at 90°C for 5–10 min. The solid-phase purification was performed using a C-18 light cartridge. Tests were performed on the appearance and particle size, pH, identification (radioisotope, radiochemistry), purity

(radiochemical, residual solvent, chemical), bacterial endotoxin (limulus amoebocyte lysate test), and sterility to confirm the quality of [^{18}F]Florastamin and [^{177}Lu]Ludotadipep.

[^{18}F]Florastamin PSMA PET/CT

To evaluate adequate PSMA expression in tumors, [^{18}F]Florastamin, which has the same primary binding domain consisting of Glu-urea-Lys as that of [^{177}Lu]Ludotadipep, was used as a diagnostic radioligand. [^{18}F]Florastamin PSMA PET/CT was performed at screening, 4 weeks p.i., and 8 weeks p.i. using a dedicated PET/CT scanner (Discovery 710, GE Healthcare, Chicago, IL, USA). PET/CT images were acquired 90 min after the intravenous injection of 185 ± 19 MBq of [^{18}F]Florastamin. The static images were reconstructed using the ordered-subset expectation maximization and point-spread function (VFPX-S) method. The pre-treatment tumor maximum standardized uptake value (SUV_{max}) was measured using screening [^{18}F]Florastamin PSMA PET/CT. Additionally, the total PSMA-expression-positive tumor burden was defined and measured: total PSMA-tumor burden = (SUV mean) · (total tumor volume), applying fixed threshold of SUV 2.5 for segmentation of all visible tumor lesions on the [^{18}F]Florastamin PSMA PET/CT using LIFEx software v.7.0.16 [18]. [^{18}F]-Fluorodeoxyglucose PET/CT was not performed.

^{177}Lu -Imaging and Blood Sampling

To evaluate organ level dosimetry, both whole body scan (WBS) and abdomen SPECT/CT (SymbiaT6, Siemens, Healthineers, Erlangen, Germany) were obtained at 2, 24, 48, 72, and 168 h p.i. WBS were acquired with medium energy collimator, 20% energy window centered on the energy peak of 208 keV (187.2–228.8 keV) and scan speed of 17 cm/min. In the subset of subjects administered with 5550 MBq, WBS was acquired with the following dual energy window (photopeak window 187.2–228.8 keV, scatter window 153.9–187.2 keV). A vial of the calibration source containing 3.7 MBq ^{177}Lu was placed between the knees at each time point to calibrate the images with attenuation and scatter correction. SPECT/CT images were acquired with a medium-energy collimator, 187.2–228.8 keV energy window, step and shoot for 60 frames, 15 s/frame, at a 180° angle, and reconstructed with CT-based attenuation correction and scatter correction according to the manufacturer's standard reconstruction method. To evaluate bone marrow dosimetry, 1–3 mL of peripheral venous blood samples was obtained from the arm opposite the radiopharmaceutical injection

site at 3, 10, 60, 180 min, 24, 48, and 72 h p.i.

Dose Calculations

The salivary glands, lungs, liver, kidneys, spleen, and red marrow were defined as accumulating organs. The activities of the liver, kidneys, and spleen at each time point were estimated from SPECT/CT images, whereas the activities of the salivary glands and lungs were estimated from the WBS. Red marrow activity was estimated from the blood samples. Lesions that were definite tumors based on all available data (previous anatomical images, [^{18}F]Florastamin PET/CT, and pathology results when available) and lesions with uptake that could reliably be accounted for by the tumor alone (i.e., lesions with uptake that could be clearly distinguished from the background activity on WBS and SPECT/CT) were selected to estimate the tumor absorbed dose. Lesions $> 2 \text{ cm}^3$ in volume were selected to minimize the partial volume effect. The SUV on [^{18}F]Florastamin PET/CT was not considered during selection.

QDOSE software v.1.1.18 (ABX-CRO, Dresden, Germany) was used to draw the region of interest (ROI) on the WBS and SPECT images, time-activity curve (TAC) fitting, and dose calculation. The ROI of the source tissues was drawn by sequential automatic segmentation using the fuzzy c-mean algorithm with the following settings: three clusters, 1.1 of weight exponent, and 3–4 included regions volume/activity, after co-registration of images for each time point. When necessary, an experienced nuclear medicine physician adjusted the ROI manually. The 2-dimensional activity calculations, including attenuation, scatter, and background corrections for the measurement of activity A_i for each source tissue ROI, were done referencing MIRD pamphlets 16 and 26 [19,20].

Bone marrow dosimetry was calculated using the venous blood samples. The intravenously administered [^{177}Lu]Ludotadipep was assumed to be uniformly distributed within the plasma and extracellular fluid spaces of the red marrow. A standardized solution of ^{177}Lu with 3.7 MBq/mL at the time of [^{177}Lu]Ludotadipep administration was diluted by $\times 3$, $\times 10$, $\times 100$ a week later and used to calibrate the gamma well counter. Red marrow-to-blood activity concentration ratio (RMBLR) of 1.0 was used [21,22]. Details of the dosimetry methods applied in this study are presented in the Supplementary Methods.

TAC Fitting and Absorbed Dose Analysis

The TAC of the source tissue was calculated using bi-

exponential fitting, except for the salivary glands. For the salivary glands and tumor lesions, the peaks were observed at later time points (24, 48, or 72 h p.i.), and the area under the curve was calculated by combining the trapezoidal method up to the peak and the mono-exponential function after the peak. The absorbed dose of the normal organs was calculated according to IDAC-Dose 2.1 implemented in the QDOSE software. The absorbed dose of tumor was calculated using a spherical model, assuming a density of 1 g/cm³ after measuring the volume from [¹⁸F]Florastamin PSMA PET by applying SUV threshold of 40% of maximum for tumor segmentation, which corresponded well to the anatomical margin seen on the CT images. The effective dose was calculated using the International Commission on Radiological Protection publication 103 weighting factors.

Statistical Analysis

Spearman's correlation test was performed to test the linear correlation between 1) pre-treatment tumor SUVmax and tumor absorbed dose coefficient and 2) total PSMA-tumor burden and absorbed doses of the salivary glands and kidneys. A partial correlation test was applied to control for the effect of tumor mass as a covariate, in addition to the correlation test between pre-treatment tumor SUVmax and tumor absorbed dose. A two-sided *P*-value less than 0.050 was considered statistically significant. Statistical analyses were performed using SPSS software version 24.0 (IBM Corp., Armonk, NY, USA).

RESULTS

Patient Characteristics

Data from 25 male (median age 73 years, range 60–90 years) were analyzed. No dose-limiting toxicities were observed. The average of total PSMA-tumor burden was 2620 cm³ [25.1–28000 cm³], and the 5.55 GBq dose group had the highest total PSMA-tumor burden of 8830 cm³ [1790–28000 cm³]. Patient characteristics are presented in Table 1. Images of a representative case are shown in Figure 2.

Biodistribution

Blood clearance of [¹⁷⁷Lu]Ludotadipep was relatively prolonged, falling to 70.7% ± 7.0% of the injected dose at 3 min p.i., 24.0% ± 9.2% at 1 h p.i., 7.5% ± 3.3% at 24 h p.i., 4.7% ± 1.7% at 48 h p.i., and 4.2% ± 1.0% at 72 h p.i. (Fig. 3). The half-times of the distribution phase and clearance phase were calculated as 27.0 ± 7.1 min

Table 1. Patient characteristics

	Group 1 of 1.85 GBq (n = 3)	Group 2 of 2.78 GBq (n = 6)	Group 3 of 3.70 GBq (n = 6)	Group 4 of 4.63 GBq (n = 6)	Group 5 of 5.55 GBq (n = 4)	Total (n = 25)
Administered activity, GBq	1.87 ± 0.02	2.85 ± 0.04	3.80 ± 0.04	4.83 ± 0.10	5.60 ± 0.04	-
Age, yrs	65 [63–78]	73.5 [63–83]	76.5 [64–90]	68 [60–86]	77.5 [73–83]	73 [60–90]
Gleason score	8 [8–10]	8 [7–9]	9 [7–9]	7.5 [7–9]	7.5 [7–10]	8 [7–10]
PSA, ng/mL	120 [7.9–125]	119 [18.7–545]	63.3 [5.3–149]	114 [58.7–3190]	790 [230–2990]	123 [5.3–2990]
Total PSMA tumor burden, g	418 [35.7–6280]	2890 [25.1–10000]	339 [49.2–3170]	4240 [618–5750]	8830 [1790–28000]	2620 [25.1–28000]

Data are mean ± standard deviation or median [range].

PSA = prostate-specific antigen, PSMA = prostate-specific membrane antigen

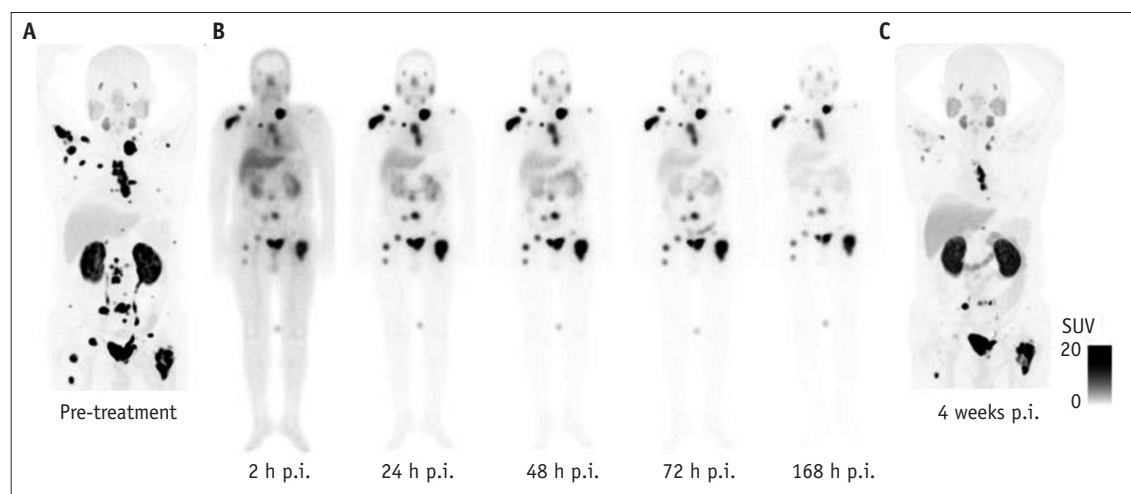


Fig. 2. A representative case. **A:** Pre-treatment [^{18}F]Florastamin PSMA PET/CT image of a patient with multiple bone and lymph node metastases. **B:** Serial whole body scans following administration of 4.63 GBq of [^{177}Lu]Ludotadipep demonstrate high uptakes in the sites corresponding to metastases seen on the PET/CT images. **C:** Post-radiopharmaceutical therapy [^{18}F]Florastamin PSMA PET/CT images show decreased intensity and extent of the tumors. PSMA = prostate-specific membrane antigen, p.i. = post-injection, SUV = standardized uptake value

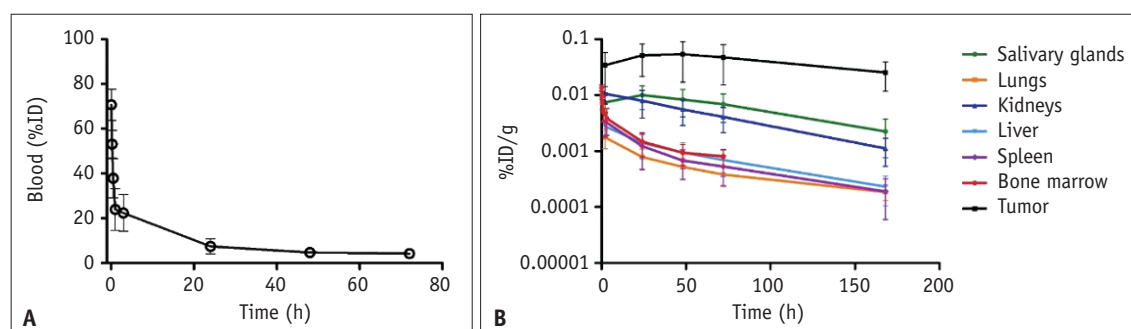


Fig. 3. Time-activity curves of [^{177}Lu]Ludotadipep (**A**) blood clearance and (**B**) normal organs and tumor.

Table 2. Absorbed dose coefficient of the source organs

Group (planned activity)	Absorbed dose coefficient (Gy/GBq)					
	Salivary glands	Lungs	Liver	Kidneys	Spleen	Bone marrow
Group 1 (1.85 GBq)	1.84 ± 1.52	0.04 ± 0.01	0.18 ± 0.09	0.86 ± 0.25	0.14 ± 0.05	NA [†]
Group 2 (2.78 GBq)	1.17 ± 0.43	0.06 ± 0.01	0.13 ± 0.05	0.78 ± 0.28	0.10 ± 0.02	NA [†]
Group 3 (3.70 GBq)	1.26 ± 0.69	0.06 ± 0.03	0.15 ± 0.06	0.76 ± 0.27	0.15 ± 0.06	0.09 [‡]
Group 4 (4.63 GBq)	0.89 ± 0.74	0.05 ± 0.01	0.13 ± 0.05	0.65 ± 0.31	0.11 ± 0.08	0.07 ± 0.01
Group 5 (5.55 GBq)	0.97 ± 0.27	NA*	0.12 ± 0.06	0.47 ± 0.25	0.07 ± 0.03	0.06 ± 0.02
All doses	1.17 ± 0.81	0.05 ± 0.02	0.14 ± 0.06	0.77 ± 0.28	0.12 ± 0.06	0.07 ± 0.02

Data are mean ± standard deviation.

*Disseminated rib metastases, [†]Insufficient blood sample data, [‡]Data from one subject.

NA = not assessable

and 60.3 ± 59.0 h, respectively. Among the normal organs, the highest activity level was $1.06 \pm 0.37 \cdot 10^{-2}\% \text{ID/g}$ in the kidneys at 2 h p.i. and $1.00 \pm 0.46 \cdot 10^{-2}\% \text{ID/g}$ in the salivary glands at 24 h p.i. The peaks were lower in the lungs ($0.18 \pm 0.07 \cdot 10^{-2}\% \text{ID/g}$), liver ($0.28 \pm 0.08 \cdot 10^{-2}\% \text{ID/g}$), and spleen ($0.34 \pm 0.16 \cdot 10^{-2}\% \text{ID/g}$), all observed at 2 h p.i. The

peak activity of tumor was $5.38 \pm 3.66 \cdot 10^{-2}\% \text{ID/g}$ at 48 h p.i.

Organ and Tumor Dosimetry

Table 2 summarizes the absorbed doses in normal organs. Salivary glands had the highest absorbed dose coefficient of 1.17 ± 0.81 Gy/GBq. The absorbed dose coefficient of kidney

Table 3. Tumor absorbed dose

Lesion number	Administered activity (GBq)	Tumor site	Tumor volume (cm ³)	Pre-treatment PSMA PET SUVmax	Pre-treatment PSMA PET SUVavg	Absorbed dose (Gy)	Absorbed dose coefficient (Gy/GBq)
1	1.86	Lumbar vertebra	7.5	41.6	9.1	19.7	10.6
2	1.86	Pelvic bone	8.1	58.7	15.0	66.7	35.8
3	1.85	Lumbar vertebra	3.8	6.8	3.7	1.8	0.9
4	1.89	Lumbar vertebra	16.7	9.3	3.9	6.7	3.5
5	2.83	Thoracic vertebra	18.3	23.5	8.8	30.4	10.7
6	2.90	Thoracic vertebra	12.0	26.6	11.0	19.9	6.9
7	2.81	Thoracic vertebra	3.0	9.9	4.7	11.0	3.9
8	3.75	Mesenteric lymph node	3.3	7.9	4.2	29.4	7.9
9	3.83	Thoracic vertebra	2.7	21.3	7.5	42.2	11.0
10	3.83	Lumbar vertebra	2.7	23.7	9.5	106.0	27.6
11	3.86	Thoracic vertebra	12.4	8.7	4.3	37.0	9.6
12	4.85	Thoracic vertebra	23.5	23.2	9.1	42.7	8.8
13	4.85	Lumbar vertebra	18.4	29.7	8.5	29.4	6.1
14	4.85	Lumbar vertebra	15.1	27.2	8.5	59.1	12.2
15	4.86	Rib	3.5	15.9	7.0	58.7	12.1
16	4.86	Thoracic vertebra	3.5	13.2	13.2	28.1	5.8
17	4.86	Lumbar vertebra	2.5	22.0	7.8	34.4	7.1
18	4.86	Lumbar vertebra	3.7	33.4	11.5	52.8	10.9
19	4.74	Aorticaval lymph node	4.1	12.5	7.9	25.4	5.4
20	4.66	Lumbar vertebra	50.2	15.1	6.5	39.3	8.9
21	4.66	Lumbar vertebra	39.5	17.2	6.0	41.6	8.9
22	5.59	Lumbar vertebra	14.8	17.2	5.7	38.2	6.8
23	5.63	Lumbar vertebra	5.8	52.1	14.1	39.7	7.0
24	5.63	Thoracic vertebra	2.0	51.3	11.1	123.9	22.0
Median [range]			6.7 [2.0–50.2]	21.7 [6.8–58.7]	8.2 [3.7–15.0]	37.6 [1.8–123.9]	8.9 [0.9–35.8]

PSMA = prostate-specific membrane antigen, SUVmax = maximum standardized uptake value, SUVavg = average standardized uptake value

and bone marrow was calculated as 0.77 ± 0.28 Gy/GBq and 0.07 ± 0.02 Gy/GBq, respectively. Table 3 describes the twenty-four tumor lesions selected for the tumor dosimetry analysis. The average volume was 11.55 ± 12.16 cm³, and the average absorbed dose coefficient of tumors was computed as 10.43 ± 7.77 Gy/GBq.

Pre-Treatment [¹⁸F]Florastamin PSMA PET vs. Absorbed Dose

The absorbed dose coefficient of the salivary glands showed a positive correlation with the SUVmax of the submandibular glands measured on pre-treatment [¹⁸F]Florastamin PSMA PET ($\rho = 0.78$, $P < 0.001$) and parotid glands ($\rho = 0.42$, $P = 0.035$). There was also a significant positive correlation between the absorbed dose coefficient of individual tumors and the pre-treatment tumor SUVmax ($\rho = 0.56$, $P = 0.005$) and average standardized uptake value (SUVavg) ($\rho = 0.46$, $P = 0.026$) (Fig. 4). There was

no significant difference in the ρ between the SUVmax and SUVavg data ($P = 0.66$).

Salivary gland doses showed a significant negative correlation with total PSMA-tumor burden ($\rho = -0.50$, $P = 0.011$) (Fig. 5). The absorbed dose coefficient of other source tissues did not significantly correlate with the total PSMA-tumor burden ($P > 0.050$).

DISCUSSION

In this first-in-human study of a novel PSMA-targeting radiopharmaceutical [¹⁷⁷Lu]Ludotadipep, data from 25 subjects were assessed for biodistribution and dosimetry. Blood clearance of [¹⁷⁷Lu]Ludotadipep was prolonged with half-times the distribution phase and clearance phase being 0.45 ± 0.12 h and 60.3 ± 59.0 h, respectively, compared to 0.16 ± 0.09 h and 10.8 ± 2.5 h reported for [¹⁷⁷Lu]Lu-PSMA-617 [23]. A peak in the kidneys was observed in

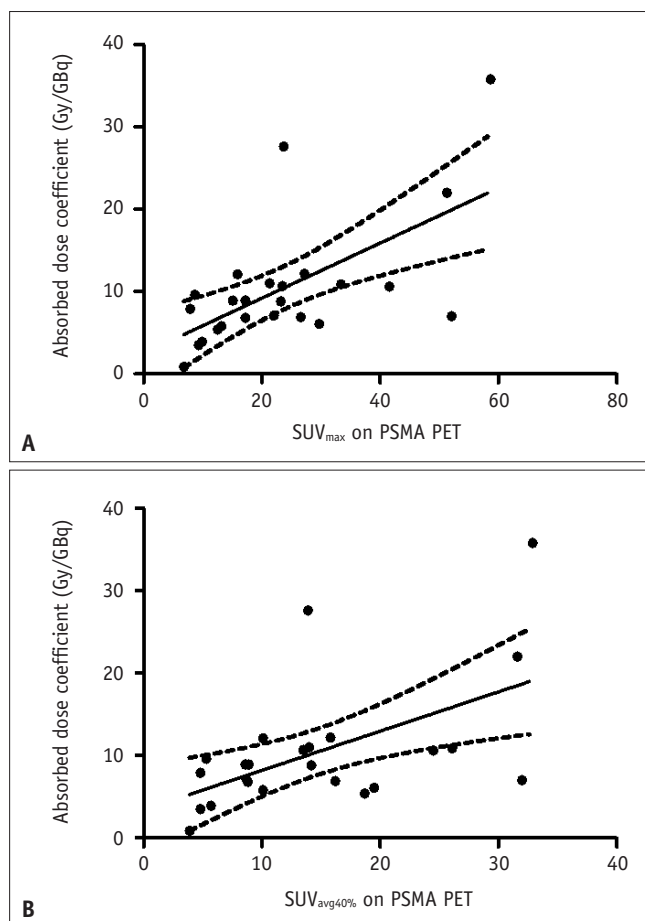


Fig. 4. Correlation of SUV from pre-treatment [^{18}F]Florastamin PSMA PET/CT with absorbed dose coefficient for tumor lesions. Correlation plots with SUV_{max} (**A**) and SUV_{avg} using 40% of SUV_{max} as a fixed threshold (**B**). SUV = standardized uptake value, PSMA = prostate-specific membrane antigen, SUV_{max} = maximum standardized uptake value, SUV_{avg} = average standardized uptake value

the first set of images obtained 2 h p.i. The mean tumor-absorbed dose coefficient was 13.6 times higher than the mean absorbed dose coefficient of the kidneys and 8.7 times higher than that of the salivary glands. As previously reported, a tumor sink effect was observed in the salivary glands of the study population [24]. Overall, the absorbed dose coefficients in various normal organs were comparable for the different activities of [^{177}Lu]Ludotadipep administered.

The red marrow was a dose limiting organ with absorbed dose coefficient of 0.07 Gy/GBq, and applying the RMBLR of 1.0 and limit of 2 Gy, the maximum injectable activity of [^{177}Lu]Ludotadipep was projected as 29.85 GBq per patient. The kidneys, another dose limiting organ with absorbed dose coefficient of 0.77 Gy/GBq and applying limit of 23 Gy derived from data of external beam radiation therapy (EBRT), allows similar maximum injectable activity of

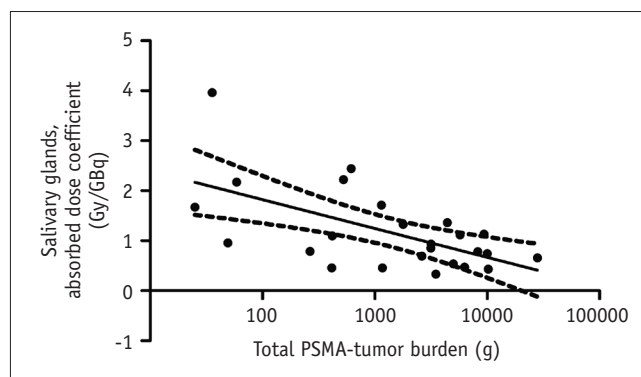


Fig. 5. Tumor sink effect observed as negative correlation between total PSMA-positive tumor burden and absorbed dose of the salivary glands. PSMA = prostate-specific membrane antigen

29.87 GBq per patient. A higher maximum injectable activity can be considered if a biologically effective dose of 40 Gy is applied as the renal dose limit in radioligand therapy, considering its low radiation dose rate compared with EBRT [25,26]. The mean absorbed dose coefficient in the tumors selected for assessment in this study was 10.43 ± 7.77 Gy/GBq, which may be high compared to the reported values for [^{177}Lu]Lu-PSMA-617, I&T, or J591 [27-29]. It should be noted that the pre-treatment PSMA SUV_{max} of the tumors selected for dosimetry was relatively low (< 10.0) in 5 out of 24 assessed lesions in our study. However, as only [^{177}Lu]Ludotadipep was used in this study, and considering the wide variability of inter- and intra-patient tumor PSMA expression, dosimetry profiles, and dosimetry computation methods [30-32] it would be injudicious to directly compare the reported tumor absorbed dose values of various PSMA-targeting compounds were directly compared. Full therapeutic efficacy can only be appreciated after long-term clinical outcome data are available.

[^{177}Lu]Ludotadipep has an iodophenylbutanoly group incorporated for affinity to serum albumin [14]. A couple of other PSMA-targeting radiopharmaceuticals that were modified to increase tumor uptake by increasing albumin binding have published dosimetry data obtained from patients with mCRPC. Evans blue (EB), the dye that binds to serum albumin with moderate affinity [33], and [^{177}Lu]Lu-PSMA-617 with a truncated EB molecule, [^{177}Lu]Lu-EB-PSMA-617, showed a longer circulation time and improved tumor uptake than [^{177}Lu]Lu-PSMA-617 [10]. [^{177}Lu]Lu-PSMA-ALB-56 contains a PSMA ligand with a p-tolyl-entity that shows lower background activity in mice [34]. In this study, the computed radiation to the kidneys with [^{177}Lu]Ludotadipep was 0.77 ± 0.28 Gy/GBq, which may be low

when compared to the published data for [^{177}Lu]Lu-PSMA-ALB-56 (2.54 ± 0.94 Gy/GBq) and [^{177}Lu]Lu-EB-PSMA-617 (2.38 ± 0.69 Gy/GBq) [8,10]. The favorable renal dosimetry of [^{177}Lu]Ludotadipep could be attributed to the earlier peak renal activity at 2 h p.i. compared to that of [^{177}Lu]Lu-PSMA-ALB-56 (24 h p.i.) and [^{177}Lu]Lu-EB-PSMA-617 (72 h p.i.). The computed dose to the red marrow for [^{177}Lu]Ludotadipep (0.07 ± 0.02 Gy/GBq) was higher than [^{177}Lu]Lu-EB-PSMA-617 (0.05 ± 0.01 Gy/GBq) and lower than [^{177}Lu]Lu-ALB-56 (0.29 ± 0.07 Gy/GBq), while the salivary gland findings were reversed ([^{177}Lu]Lu-ALB-56 0.86 ± 0.42 Gy/GBq, vs. [^{177}Lu]Ludotadipep 1.17 ± 0.81 Gy/GBq, vs. [^{177}Lu]Lu-EB-PSMA-617 6.41 ± 1.40 Gy/GBq) [8,10]. The absorbed dose to the kidneys observed in this study was about twice the preclinical dose that was estimated based on biodistribution in mice, which had been calculated as 0.03 Gy/GBq, reflecting the difference between the species and highlighting the difficulty of estimating human dose and variability that comes from different masses. Newer radiopharmaceuticals with albumin binders, such as [^{177}Lu]HTK03121 reported a 2.9 fold improvement in the tumor-to-kidney absorbed dose ratio compared to [^{177}Lu]Lu-PSMA-617 in animal LNCaP tumor-bearing mice [35], and it remains to be seen if this advantage is retained in patients.

In our study, up to 5.6 GBq of [^{177}Lu]Ludotadipep was administered once to patients without immediate complications or an incidence of grade 3 or 4 toxicity. In a study with patients in three escalating activities (1.2, 2.1, and 3.5 GBq) of up to 3 cycles of [^{177}Lu]Lu-EB-PSMA-617 radiopharmaceutical therapy at 8-week intervals, 2.1 GBq was shown to be safe and adequate in tumor treatment [36]. Data are currently being collected for a more comprehensive assessment of the clinical safety profile and the efficacy of [^{177}Lu]Ludotadipep. Based on the available data so far, the KMFDS has approved phase II clinical trial with up to 3.7 GBq $\times$ 6 cycles of [^{177}Lu]Ludotadipep. In a multicycle clinical trial, positively charged amino acid infusion was added to the protocol for kidney protection. We used a more conservative RMBL (1.0) for the estimation of the bone marrow dose, as recommended for both [^{177}Lu]Lu-SSRT and [^{177}Lu]Lu-PSMA [22], but a lower RMBL could be more appropriate for [^{177}Lu]Ludotadipep considering its albumin-binding character. Additionally, 3 Gy as the marrow dose limit instead of 2 Gy should be cautiously tested in the future to better understand normal tissue tolerance to radiopharmaceutical therapy [37,38].

This pilot dosimetry study of [^{177}Lu]Ludotadipep had

several limitations. Whole-body scans were acquired in different energy windows, and blood sampling was performed in only half of the patients. Instead of the total tumor burden, a selected minority of the tumor lesions was assessed for the absorbed dose, and the majority were vertebral metastases. Real-life scenarios, such as impaired renal function or constipation of the patient, spillover from bone and paravertebral metastases to the bone marrow, changes in tumor PSMA expression, and total tumor volume over the therapeutic period, could create room for discrepancy from dosimetry estimates based on blood samples and images, especially for multiple cycles of therapy.

In conclusion, [^{177}Lu]Ludotadipep showed a dosimetry profile that is expected to be safe at a dose of 6 cycles of 3.7 GBq planned for a phase II clinical trial, with kidneys and bone marrow as the critical organs. At the projected maximum injected activity, the tumor-absorbed dose in human subjects was computed to be comparably high for [^{177}Lu]Ludotadipep as for various [^{177}Lu]Lu-labeled PSMA-targeting radiopharmaceuticals reported in the literature.

Supplement

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Availability of Data and Material

The datasets generated or analyzed during the study are available from the corresponding author on reasonable request.

Conflicts of Interest

Chansoo Park is an employee of FutureChem Co., Ltd., and Dae Yoon Chi is its chief executive officer. Seunggyun Ha has consultant agreement with FutureChem. Joo Hyun O has consultant agreements with FutureChem and Novartis, and as a member of the editorial board of the *Korean Journal of Radiology*, she was not involved in the editorial evaluation or decision to publish this article. All remaining authors have declared no conflicts of interest.

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REFERENCES

1. Silver DA, Pellicer I, Fair WR, Heston WD, Cordon-Cardo C. Prostate-specific membrane antigen expression in normal and malignant human tissues. *Clin Cancer Res* 1997;3:81-85
2. Hyvärkkä A, Virtanen V, Kempainen J, Grönroos TJ, Minn H, Sundvall M. More than meets the eye: scientific rationale behind molecular imaging and therapeutic targeting of prostate-specific membrane antigen (PSMA) in metastatic prostate cancer and beyond. *Cancers (Basel)* 2021;13:2244
3. Maurer T, Eiber M, Schwaiger M, Gschwend JE. Current use of PSMA-PET in prostate cancer management. *Nat Rev Urol* 2016;13:226-235
4. Kabasakal L, AbuQbeith M, Aygün A, Yeyin N, Ocak M, Demirci E, et al. Pre-therapeutic dosimetry of normal organs and tissues of (177)Lu-PSMA-617 prostate-specific membrane antigen (PSMA) inhibitor in patients with castration-resistant prostate cancer. *Eur J Nucl Med Mol Imaging* 2015;42:1976-1983
5. Violet J, Jackson P, Ferdinandus J, Sandhu S, Akhurst T, Iravani A, et al. Dosimetry of 177Lu-PSMA-617 in metastatic castration-resistant prostate cancer: correlations between pretherapeutic imaging and whole-body tumor dosimetry with treatment outcomes. *J Nucl Med* 2019;60:517-523
6. Sartor O, de Bono J, Chi KN, Fizazi K, Herrmann K, Rahbar K, et al. Lutetium-177-PSMA-617 for metastatic castration-resistant prostate cancer. *N Engl J Med* 2021;385:1091-1103
7. Hofman MS, Emmett L, Sandhu S, Iravani A, Joshua AM, Goh JC, et al. [177Lu]Lu-PSMA-617 versus cabazitaxel in patients with metastatic castration-resistant prostate cancer (TheraP): a randomised, open-label, phase 2 trial. *Lancet* 2021;397:797-804
8. Kramer V, Fernández R, Lehnert W, Jiménez-Franco LD, Soza-Ried C, Eppard E, et al. Biodistribution and dosimetry of a single dose of albumin-binding ligand [177Lu]Lu-PSMA-ALB-56 in patients with mCRPC. *Eur J Nucl Med Mol Imaging* 2021;48:893-903
9. Benešová M, Umbricht CA, Schibli R, Müller C. Albumin-binding PSMA ligands: optimization of the tissue distribution profile. *Mol Pharm* 2018;15:934-946
10. Zang J, Fan X, Wang H, Liu Q, Wang J, Li H, et al. First-in-human study of 177Lu-EB-PSMA-617 in patients with metastatic castration-resistant prostate cancer. *Eur J Nucl Med Mol Imaging* 2019;46:148-158
11. Kelly J, Amor-Coarasa A, Ponnala S, Nikolopoulou A, Williams C Jr, Schlyer D, et al. Trifunctional PSMA-targeting constructs for prostate cancer with unprecedented localization to LNCaP tumors. *Eur J Nucl Med Mol Imaging* 2018;45:1841-1851
12. Vaughn BA, Loveless CS, Cingoranelli SJ, Schlyer D, Lapi SE, Boros E. Evaluation of 177Lu and 47Sc picagala-linked, prostate-specific membrane antigen-targeting constructs for their radiotherapeutic efficacy and dosimetry. *Mol Pharm* 2021;18:4511-4519
13. Ling X, Latoche JD, Choy CJ, Kurland BF, Laymon CM, Wu Y, et al. Preclinical dosimetry, imaging, and targeted radionuclide therapy studies of Lu-177-labeled albumin-binding, PSMA-targeted CTT1403. *Mol Imaging Biol* 2020;22:274-284
14. Lee BS, Kim MH, Chu SY, Jung WJ, Jeong HJ, Lee K, et al. Improving theranostic gallium-68/lutetium-177-labeled PSMA

- inhibitors with an albumin binder for prostate cancer. *Mol Cancer Ther* 2021;20:2410-2419
15. Shin D, Ha S, O JH, Rhew SA, Yoon CE, Kwon HJ, et al. A single dose of novel PSMA-targeting radiopharmaceutical agent [¹⁷⁷Lu]Ludotadipep for patients with metastatic castration-resistant prostate cancer: phase I clinical trial. *Cancers (Basel)* 2022;14:6225
 16. Lee I, Lim I, Byun BH, Kim BI, Choi CW, Woo SK, et al. A microdose clinical trial to evaluate [¹⁸F]florastamin as a positron emission tomography imaging agent in patients with prostate cancer. *Eur J Nucl Med Mol Imaging* 2021;48:95-102
 17. Rowe SP, Pienta KJ, Pomper MG, Gorin MA. PSMA-RADS version 1.0: a step towards standardizing the interpretation and reporting of PSMA-targeted PET imaging studies. *Eur Urol* 2018;73:485-487
 18. Nioche C, Orlhac F, Boughdad S, Reuzé S, Goya-Outi J, Robert C, et al. LIFEx: a freeware for radiomic feature calculation in multimodality imaging to accelerate advances in the characterization of tumor heterogeneity. *Cancer Res* 2018;78:4786-4789
 19. Siegel JA, Thomas SR, Stubbs JB, Stabin MG, Hays MT, Koral KF, et al. MIRD pamphlet no. 16: techniques for quantitative radiopharmaceutical biodistribution data acquisition and analysis for use in human radiation dose estimates. *J Nucl Med* 1999;40:375-615
 20. Ljungberg M, Celler A, Konijnenberg MW, Eckerman KF, Dewaraja YK, Sjögreen-Gleisner K, et al. MIRD pamphlet no. 26: joint EANM/MIRD guidelines for quantitative ¹⁷⁷Lu SPECT applied for dosimetry of radiopharmaceutical therapy. *J Nucl Med* 2016;57:151-162
 21. Forrer F, Krenning EP, Kooij PP, Bernard BF, Konijnenberg M, Bakker WH, et al. Bone marrow dosimetry in peptide receptor radionuclide therapy with [¹⁷⁷Lu-DOTA(0),Tyr(3)]octreotate. *Eur J Nucl Med Mol Imaging* 2009;36:1138-1146
 22. Sjögreen Gleisner K, Chouin N, Gabina PM, Ciccone F, Gnesin S, Stokke C, et al. EANM dosimetry committee recommendations for dosimetry of ¹⁷⁷Lu-labelled somatostatin-receptor- and PSMA-targeting ligands. *Eur J Nucl Med Mol Imaging* 2022;49:1778-1809
 23. Kabasakal L, Toklu T, Yeyin N, Demirci E, Abuqbeith M, Ocak M, et al. Lu-177-PSMA-617 prostate-specific membrane antigen inhibitor therapy in patients with castration-resistant prostate cancer: stability, bio-distribution and dosimetry. *Mol Imaging Radionucl Ther* 2017;26:62-68
 24. Gafita A, Wang H, Robertson A, Armstrong WR, Zaum R, Weber M, et al. Tumor sink effect in ⁶⁸Ga-PSMA-11 PET: myth or reality? *J Nucl Med* 2022;63:226-232
 25. Cremonesi M, Ferrari M, Bodei L, Tosi G, Paganelli G. Dosimetry in peptide radionuclide receptor therapy: a review. *J Nucl Med* 2006;47:1467-1475
 26. Bodei L, Cremonesi M, Ferrari M, Pacifici M, Grana CM, Bartolomei M, et al. Long-term evaluation of renal toxicity after peptide receptor radionuclide therapy with ⁹⁰Y-DOTATOC and ¹⁷⁷Lu-DOTATATE: the role of associated risk factors. *Eur J Nucl Med Mol Imaging* 2008;35:1847-1856
 27. Delker A, Fendler WP, Kratochwil C, Brunegraf A, Gosewisch A, Gildehaus FJ, et al. Dosimetry for ¹⁷⁷Lu-DKFZ-PSMA-617: a new radiopharmaceutical for the treatment of metastatic prostate cancer. *Eur J Nucl Med Mol Imaging* 2016;43:42-51
 28. Violet J, Sandhu S, Iravani A, Ferdinandus J, Thang SP, Kong G, et al. Long-term follow-up and outcomes of retreatment in an expanded 50-patient single-center phase II prospective trial of ¹⁷⁷Lu-PSMA-617 theranostics in metastatic castration-resistant prostate cancer. *J Nucl Med* 2020;61:857-865
 29. Vallabhajosula S, Kuji I, Hamacher KA, Konishi S, Kostakoglu L, Kothari PA, et al. Pharmacokinetics and biodistribution of ¹¹¹In- and ¹⁷⁷Lu-labeled J591 antibody specific for prostate-specific membrane antigen: prediction of ⁹⁰Y-J591 radiation dosimetry based on ¹¹¹In or ¹⁷⁷Lu? *J Nucl Med* 2005;46:634-641
 30. Uribe C, Peterson A, Van B, Fedrigo R, Carlson J, Sunderland J, et al. An international study of factors affecting variability of dosimetry calculations, part 1: design and early results of the SNMMI dosimetry challenge. *J Nucl Med* 2021;62(Suppl 3):36S-47S
 31. Mahmoudi E, Pirayesh E, Deevband MR, Amoui M, Rad MG, Ghorbani M. Patient-specific dosimetry in radioligand therapy (RLT) for metastatic prostate cancer using ¹⁷⁷Lu-DKFZ-PSMA-617. *Nucl Med Mol Imaging* 2021;55:237-244
 32. Bakht MK, Oh SW, Youn H, Cheon GJ, Kwak C, Kang KW. Influence of androgen deprivation therapy on the uptake of PSMA-targeted agents: emerging opportunities and challenges. *Nucl Med Mol Imaging* 2017;51:202-211
 33. Jacobson O, Kiesewetter DO, Chen X. Albumin-binding Evans blue derivatives for diagnostic imaging and production of long-acting therapeutics. *Bioconjug Chem* 2016;27:2239-2247
 34. Umbricht CA, Benešová M, Schibli R, Müller C. Preclinical development of novel PSMA-targeting radioligands: modulation of albumin-binding properties to improve prostate cancer therapy. *Mol Pharm* 2018;15:2297-2306
 35. Kuo HT, Lin KS, Zhang Z, Uribe CF, Merckens H, Zhang C, et al. ¹⁷⁷Lu-labeled albumin-binder-conjugated PSMA-targeting agents with extremely high tumor uptake and enhanced tumor-to-kidney absorbed dose ratio. *J Nucl Med* 2021;62:521-527
 36. Zang J, Liu Q, Sui H, Wang R, Jacobson O, Fan X, et al. ¹⁷⁷Lu-EB-PSMA radioligand therapy with escalating doses in patients with metastatic castration-resistant prostate cancer. *J Nucl Med* 2020;61:1772-1778
 37. Wahl RL, Sgouros G, Iravani A, Jacene H, Pryma D, Saboury B, et al. Normal-tissue tolerance to radiopharmaceutical therapies, the knowns and the unknowns. *J Nucl Med* 2021;62(Suppl 3):23S-35S
 38. Hindorf C, Lindén O, Tennvall J, Wingårdh K, Strand SE. Evaluation of methods for red marrow dosimetry based on patients undergoing radioimmunotherapy. *Acta Oncol* 2005;44:579-588