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ORIGINAL RESEARCH

IncRNA LOC100132354 promotes angiogenesis through VEGFA/VEGFR2 signaling pathway in lung adenocarcinoma

Yumin Wang^{1,*} Fan Zhang^{1,*} Junjun Wang¹ Lijuan Hu¹ Feng Jiang¹ Jian Chen¹ Jie Chen² Liangxing Wang³

¹Department of Laboratory Medicine, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou China; ²Intensive Center Unit, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou China; ³Department of Respiratory Medicine, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China

*These authors contributed equally to this work

Correspondence: Jie Chen Intensive Center Unit, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China Tel/fax +86 577 5557 9330 Email chenjie991300@163.com

Liangxing Wang Department of Respiratory, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou, China Tel/fax +86 577 5557 9272 Email wzyxywlx@163.com



Purpose: This study aimed to study the biological function and the molecular mechanisms associated with the promotion of angiogenesis by lncRNA LOC100132354 in lung adenocarcinoma (LAD).

Patients and methods: The mRNA expression levels of 100 pairs of LAD and normal tissue samples of LOC100132354, vascular endothelial growth factor A (VEGFA), VEGF receptor-2 (VEGFR2), basic fibroblast growth factor (bFGF), and thrombospondin-1 (TSP-1) were analyzed by qPCR. LOC100132354 was knockdown and overexpressed in SPCA-1 and A549 cell lines to analyze the protein and mRNA expression levels of VEGFA, VEGFR2, bFGF, TSP-1, and changes in protein expression levels of Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mekl/2, and P-Erk1/2. Tumor microvessel density (MVD) was analyzed in experimental nude mice.

Results: The qPCR results showed that the mRNA expression levels of LOC100132354, VEGFA, VEGFR2, and bFGF mRNA in LAD tissues were significantly increased, while TSP-1 mRNA was significantly decreased compared with the adjacent tissues. Survival analysis showed that VEGFA, VEGFR2, and bFGF were poor predictors, while TSP-1 was a good predictor in LAD. Knockdown or overexpression of LOC100132354 affected the expression levels of bFGF, VEGFA/VEGFR2 signaling pathway, and downstream target molecules, such as Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mekl/2, and P-Erk1/2, while decreased TSP-1. After knockdown or overexpression of VEGFA expression, no significant changes in the expression level of LOC100132354 were found. Tumorigenesis of nude mice confirmed that LOC100132354 can significantly increase the tumor MVD.

Conclusion: These findings suggest VEGFA was a downstream target gene of LOC100132354, promoting angiogenesis through VEGFA/VEGFR2 signaling pathway and downstream target molecules in LAD. So, LOC100132354 is considered as an antiangiogenic target in LAD. **Keywords:** lung cancer, lncRNA, angiogenesis, signaling pathway

Introduction

Lung cancer ranks first in terms of both morbidity and mortality of all cancers.¹ Of all lung cancers, non-small-cell lung cancer (NSCLC) accounts for approximately 85%. According to the histopathological classification, NSCLC is divided into lung adenocarcinoma (LAD) and squamous cell carcinoma. Despite several advancements in chemotherapy, radiation, and surgery, lung cancer still remains aggressive and resulted in mortality.² The average 5-year survival of lung cancer is below 15%.³⁻⁶ Recently, an increasing proportion of LAD has drawn a great attention on the socioeconomic environments.⁷ However, the mechanisms of LAD have not yet been uncovered.

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LncRNAs have been confirmed to play a very important role in the occurrence and progression of lung cancer. It has been found that some *lncRNAs*, such as HOTAIR,⁸ H19,⁹ MALAT1,10 SCAL1,11 AK126698,12 and GAS6-AS1,13 are closely related to lung cancer. Previously, we have found that lncRNA LOC100132354 was overexpressed in LAD and was localized at 6p21.1, with a total of 2282 bp RNA sequences encompassing the two exons by gene microarray. With these results, we have confirmed that LOC100132354 is an oncogenic lncRNA gene, and it promoted the progression and markedly enhanced the oncogenic ability in LAD.14 Therefore, we performed bioinformatics analysis of LOC100132354 and its adjacent coding gene (distance <300 kb) with transcriptome mRNA microarray results. These findings revealed that vascular endothelial growth factor A (VEGFA) may be an adjacent gene regulated by LOC100132354.

The growth and metastasis of lung cancer require the support of new blood vessels. Neovascularization plays a key role in the occurrence and development of lung cancer and is determined by the tumor microenvironment. Multiple signaling molecules were involved in its regulation.^{15–19} The proliferation of vascular endothelial cells is considered to be the core of neovascularization process. Vascular endothelial growth factor (VEGF) is the most important factor for inducing endothelial cell proliferation, and angiogenesis in tumor tissues is directly related to VEGF. VEGF consists of five family members, namely VEGFA, VEGFB, VEGFC, VEGFD, and VEGFE. VEGF binds to its receptors to induce the biological effects. The VEGF receptors (VEGFRs) mainly include Flt-1, KDR/Flk-1 (VEGFR2), Flt-4, and so on.^{20,21} VEGF, VEGFR, and their downstream signaling transduction pathways are used as specific tumor angiogenesis markers. Also VEGFA/VEGFR2 signaling transduction plays a leading role in the formation of vascular endothelial cells. VEGFA/VEGFR2 is a key factor for promoting angiogenesis in lung cancer,²² and it can promote tumor growth.²³ Therefore, inhibition of VEGFA/VEGFR2 signaling pathway can inhibit tumor cell growth by blocking angiogenesis.^{24,25} Basic fibroblast growth factor (bFGF) is also one of the angiogenic growth factors that can stimulate endothelial cells to synthesize collagenase and promote invasion of the surrounding stroma during angiogenesis. Moreover, VEGF can induce endothelial cell production and release bFGF, further promoting the role of angiogenesis.²⁶ While thrombospondin-1 (TSP-1) is a strong negative regulator of tumor angiogenesis.27

VEGFA/VEGFR2 can activate Ras by multiple pathways²⁸ and in turn activate Raf/Mek/Erk to exert the biological functions. Studies have shown that specific inhibitors of Mek1/2 can prevent the activation of Erk, thereby inhibiting the formation of blood vessels in vitro. Therefore, we speculate that LOC100132354 may regulate VEGFA/VEGFR2 signaling pathway to promote angiogenesis in LAD, but the specific regulatory mechanisms needs to be elucidated.

Patients and methods Patient samples

We collected 100 LAD tissue samples and corresponding normal tissue (NT) samples in the First Affiliated Hospital of Wenzhou Medical University, from January 2013 to August 2015. The TNM stage was determined according to the American Joint Committee on Cancer (AJCC) and the Union for International Cancer Control (UICC) in 2002. The diagnosis of LAD was confirmed using histopathology results. The clinical and characteristic data of these LAD patients are presented in Table 1. We followed-up the survival of these cases by telephone or other ways, and the range of follow-up time was 3–23 months. This study was approved by the Institutional Ethics Review Board of the First Affiliated Hospital of Wenzhou Medical University, and all patients provided written informed consent.

Quantitative PCR

Total RNA was obtained from fresh LAD tissues and cell lines with TRIzol Reagent kit (Invitrogen, Waltham, MA, USA) and was reverse-transcribed into cDNA with a reversetranscribed kit (TaKaRa, Shanghai, China) according to the manufacturer's instructions. LOC100132354, VEGFA, VEGFR2, bFGF, TSP-1, and β -actin mRNA expressions were detected by qPCR in ABI 7300 analyzer. The primers of six genes are shown in Table 2. Two micrograms of total RNA was reverse-transcribed into cDNA. qPCR was performed in a total reaction volume of 20 μ L, which consists of 2 μ L cDNA, 1 µL PCR forward primer (10 mM), 10 µL SYBR Premix (2×), 1 μ L PCR reverse primer (10 mM), and 6 μ L distilled water. The qPCR program was as follows: 10 minutes denaturation at 95°C, 40 cycles for 5 seconds at 95°C, 30 seconds at 60°C, and 5 minutes extension at 72°C. All the results were normalized to β -actin, and all the experiments were performed in triplicate. The median in each triplicate for calculating the relative lncRNA expressing concentrations (ΔCt =Ct median Ct median β -actin), and $2^{-\Delta\Delta Ct}$ formula for the relative expression.²⁹

	Cases (n)	VEGFA ^a	P-value	VEGFR2 ^a	P-value	bFGF ^a	P-value	TSP-1ª	P-value
Sex			0.876		0.765		0.543		0.762
Male	40	8.12±0.57		6.23±0.32		4.01±0.22		0.62±0.02	
Female	60	7.99±0.65		6.16±0.27		3.98±0.20		0.61±0.01	
Clinical stage			0.001		0.008		0.019		0.021
la	22	4.01±0.12		2.34±0.07		1.23±0.13		0.91±0.12	
lb	31	4.19±0.14		2.46±0.12		1.65±0.12		0.86±0.11	
lla	15	5.23±0.13		3.25±0.13		2.02±0.14		0.71±0.09	
llb	16	5.89±0.21		4.42±0.18		2.32±0.14		0.35±0.05	
Illa	15	9.86±0.71		7.78±0.32		5.24±0.25		0.15±0.01	
Differentiation degree			0.008		0.002		0.021		0.032
Low	13	9.34±0.70		7.24±0.42		7.23±0.35		0.90±0.11	
Low-moderate	14	7.23±0.35		5.21±0.34		5.12±0.21		0.71±0.10	
Moderate	36	4.45±0.32		4.24±0.21		3.45±0.16		0.50±0.08	
High-moderate	19	3.83±0.25		2.34±0.18		1.45±0.12		0.31±0.05	
High	18	2.56±0.18		2.01±0.09		1.34±0.09		0.17±0.03	
LN metastasis			0.000		0.000		0.002		0.012
Yes	28	11.21±1.21		9.34±0.54		8.43±0.46		0.11±0.03	
No	72	3.21±0.23		2.24±0.12		1.87±0.16		0.91±0.15	
Smoking			0.854		0.734		0.521		0.773
Yes	37	8.23±0.56		6.34±0.33		4.11±0.23		0.65±0.02	
No	63	8.16±0.53		6.21±0.29		4.02±0.21		0.60±0.01	

Table I The clinical features of 100 patients with LAD

Notes: amRNA relative expression level.

Abbreviations: bFGF, basic fibroblast growth factor; LAD, lung adenocarcinoma; LN, lymph node; TSP-1, thrombospondin-1; VEGFA, vascular endothelial growth factor A; VEGFR2, vascular endothelial growth factor receptor-2.

Table 2 The primers of five genes

Gene ID	Sense primer	Antisense primer	Product size
LOC100132354	TTGGGCTGAGCACAGAGAAG	GTAGGAGTGTGTGTGTGG GG	198 bp
VEGFA	TCACCAAGGCCAGCACATAG	GAGGCTCCAGGGCATTAGAC	176 bp
VEGFR2	CGTCAACAAAGTCGGGAGA	CAGTGCACCACAAAGA CACG	153 bp
bFGF	CGGTCAACAAAGTCGGGAGA	CAGTGCACCACAAAGACACG	187 bp
TSP-1	TGCTTGCATCAGTGTGGACT	ATGGCCTCACAATAG CACCC	193 bp
β-actin	CCTGGCACCC AGCACAAT	GCTGATCCACATCTGCTGGAA	157 bp

Abbreviations: bFGF, basic fibroblast growth factor; TSP-1, thrombospondin-1; VEGFA, vascular endothelial growth factor A; VEGFR2, vascular endothelial growth factor receptor-2.

Cell culture

Two human LAD cell lines (SPCA-1, A549) were purchased from the Cell Bank of the Chinese Academy of Sciences and were cultured with complete medium (90% RPMI1640 supplemented with 10% FBS) at 37°C in 5% CO₂. The complete medium was replaced every 2–3 days.

Lentivirus-mediated siRNA and overexpression vector construction

We constructed siRNA GV248 vector and overexpression GV303 vector for LOC100132354 (GeneChem, Shanghai, China). SiRNA sequences of LOC100132354 were as follows: siRNA1: CCACCTGGATAGGGACAAA, siRNA2: GAAGAGAAATTGACAGGTT, siRNA3: CTGG

AGTAGAGAAGA GAAT. SiRNA sequences of VEGFA were as follows: siRNA1: AAGCCGTCCT GTGTGC-CGCTG, siRNA2: AACGATGAAGCCCTGGAGTGC, siRNA3: AAG TTAAAAGTGCCTGAACTG. Transfection was performed by adding 2×10^5 cells into 6-well plate. After 24 hours, the medium was changed, and the cells were incubated with the transfection complex according to the manufacturer's standard operating procedure. The multiplicity of infection values³⁰ was as follows: the MOI of A549 was 20 and that of SPCA-1 was 100. The A549 and SPCA-1 cells were infected with these lentiviruses for 72 hours, and the siRNA or overexpression efficiency was evaluated by qPCR. Puromycin screen experiment separated and obtained these cell lines by successfully transfecting with

lentivirus-mediated vectors. The study involved SPCA-1 LOC100132354, VEGFA siRNAs cell lines (siRNAs group, including siRNA1, siRNA2, and siRNA3). SPCA-1 was transfected with lentivirus-negative control LVCON077 vector (siRNA NC group), SPCA-1 (siRNA control group), and the A549 LOC100132354. VEGFA overexpressing cell line (OE group) and A549 were transfected with lentivirus negative control LVCON145 vector (OE NC group) and A549 (OE control group). We have previously constructed the best LOC100132354 interference sequence for SPCA-1 cell line and LOC100132354 overexpression level in A549 cells.¹⁴

Western blotting

The cell lysate was placed on ice for 30 minutes. The supernatant was electrophoresed on SDS-PAGE and then transferred onto a nitrocellulose membrane. Following the molecular cloning procedure, the primary antibody was blocked overnight and horseradish peroxidase secondary antibody was allowed to bind for 1 hour.³¹ The protein levels of VEGFA, VEGFR2, bFGF, TSP-1, Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mekl/2, and P-Erk1/2 in these groups were then analyzed.

Nude mice tumor formation and microvessel density (MVD) detection

The cells include SPCA-1 LOC100132354 siRNA2 cell line (siRNA group), siRNA NC group, OE group, and OE NC group. These cells at a density of 2×106 cells were inoculated into the axillary region of BALB/c nude mice subcutaneously. MVD was measured according to the method reported by Weidner et al and Maeda et al, and the sections were analyzed to determine the highest blood vessel density in the tumor, necrosis area with lesser blood vessels, and no blood vessels at the junction with NTs under low magnification microscope (100×). Any brown-yellow stained endothelial cells or cell plexuses that distinguish from the surrounding tumor tissues and connective tissue components were considered as blood vessels and confirmed after finding the highest density of blood vessels under the high magnification microscope (200×). If the structures are not connected, its branching structure also serves as a blood vessel.32,33 The number of microvessels of the five fields of vision was recorded for each sample, and the average number of groups was taken as the MVD value for the group. The investigation confirms with the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health (NIH Publication No. 85-23, revised 1996). The study protocol was approved by the Institutional Animal Experimental Ethics Review Board of Wenzhou Medical University.

Statistical methods

Differences in variables between the groups were assessed using Kruskal–Wallis test for non-normal distribution data or one-way ANOVA test for normal distribution data. Comparison between the two groups was evaluated by Student's *t*-test or Mann–Whitney *U* test or least significant difference test. Survival curve was analyzed with GraphPad Prism software 6.0, and chi-squared test analyzed the differences. P<0.05was considered to be statistically significant.

Results

The expression level and survival analysis of LOC100132354, VEGFA, VEGFR2, bFGF, and TSP-1 in LAD

The results of qPCR showed that the relative mRNA expression level of LOC100132354, VEGFA, VEGFR2, and bFGF was obviously higher than the NT tissues (Mann–Whitney U=103.00, 87.00, 79.00, 96.00 and P=0.0040, 0.0018, 0.0019, 0.0021), while TSP-1 mRNA was obviously lower than the NT tissues (Mann–Whitney U=165.00, P=0.0000) in LAD (Figure 1). This suggested significantly enhanced angiogenic capacity in the LAD tissues. One-way ANOVA test or *t*-test showed that the relative mRNA expression levels of VEGFA, VEGFR2, bFGF, and TSP-1 (P=0.001, 0.008, 0.019, 0.021. 0.008, 0.002, 0.021, 0.032. 0.000, 0.000, 0.002, 0.012) were obviously different in clinical stage, differentiation degree, and lymph node metastasis, while VEGFA, VEGFR2, bFGF, and TSP-1 (P=0.876, 0.765, 0.543, 0.762. 0.854, 0.734, 0.521, 0.773) showed no differences in sex and smoking (Table 1).

The survival analysis showed that VEGFA, VEGFR2, and bFGF (χ^2 =16.26, 10.78, 16.22 and *P*<0.0001, *P*=0.0010, *P*<0.0001) were considered as poor predictors, while TSP-1 (χ^2 =17.50, *P*<0.0001) was a good predictor in LAD (Figure 2). Studies have confirmed that VEGFA and VEGFR2 played a pivotal role in angiogenesis and bFGF was considered as an angiogenic factor, while TSP-1 was a negative regulator of vascular growth. So, our results confirmed that the growth of blood vessels was significantly increased in the LAD tissues.

LOC100132354 significantly enhanced angiogenic capacity in the LAD cell lines through VEGFA/VEGFR2 signaling pathway and downstream Ras/Raf/Mek/ Erk pathway

We further confirmed that the protein expression levels of VEGF, VEGFR2, bFGF, Ras, P-A-Raf, P-B-Raf, P-C-Raf,



Figure I Comparison of mRNA expression levels of LOC100132354, VEGFA, VEGFA, VEGFR2, bFGF, and TSP-1 in lung adenocarcinoma tissues and adjacent tumors. Notes: (A) Comparison of the relative expression levels of LOC100132354 in LAD and adjacent tissues showed that LOC100132354 expression level in LAD was significantly higher than the adjacent tissues. ***P<0.001. (B) Comparison of the relative expression levels of VEGFA in LAD and adjacent tissues showed that VEGFA expression level in LAD was distinctly higher than the adjacent tissues. ***P<0.001. (C) Comparison of the relative expression levels of VEGFA in LAD and adjacent tissues showed that VEGFA and Adjacent tissues. ***P<0.001. (C) Comparison of the relative expression levels of VEGFA in LAD and adjacent tissues showed that VEGFA and adjacent tissues. ***P<0.001. (D) Comparison of the relative expression levels of bFGF in LAD and adjacent tissues showed that VEGFA in LAD and adjacent tissues showed that VEGFA in LAD and adjacent tissues. ***P<0.001. (D) Comparison of the relative expression levels of bFGF in LAD and adjacent tissues showed that bFGF expression level in LAD was obviously higher than the adjacent tissues. ***P<0.001. (E) Comparison of the relative expression levels of TSP-1 in LAD and adjacent tissues showed that TSP-1 expression level in LAD was markedly lower than the adjacent tissues. ***P<0.001. Abbreviations: bFGF, basic fibroblast growth factor; LAD, lung adenocarcinoma; NT, normal tissue; TSP-1, thrombospondin-1; VEGFA, vascular endothelial growth factor receptor-2.

P-Mekl/2, and P-Erk1/2 (t=7.258, 8.850, 8.994, 4.920, 9.032, 7.734, 9.141, 7.143; *P*=0.018, 0.013, 0.012, 0.039, 0.012, 0.024, 0.012, 0.019) were significantly higher in the OE group than in the OE NC group, while TSP-1 protein expression levels were lower (t=-7.471, *P*=0.017). The protein expression levels of VEGF, VEGFR2, bFGF, Ras, P-A-Raf, P-B-Raf,

P-C-Raf, P-Mekl/2, and P-Erk1/2 (*t*=8.701, 7.574, 6.491, 5.421, 5.578, 5.443, 10.140, 8.424, 12.689; *P*=0.013, 0.021, 0.023, 0.032, 0.031, 0.032, 0.010, 0.014, 0.006) in the siRNA group were lower than those of the siRNA NC group, but the TSP-1 protein expression levels were higher than those of the siRNA NC group (*t*=-24.005, *P*=0.002) (Figure 3).



Figure 2 The results of survival analysis in LAD.

Notes: (**A**) The survival analysis showing that LOC100132354 ($\chi^2=12.22$, P<0.001) as a poor predictor. (**B**) The survival analysis showing VEGFA ($\chi^2=16.26$, P<0.0001) as a poor predictor. (**C**) The survival analysis showing that VEGFR2 ($\chi^2=10.78$, P=0.0010) as a poor predictor. (**D**) The survival analysis showing that bFGF ($\chi^2=16.22$, P<0.0001) as a poor predictor. (**E**) Survival analysis showing TSP-1 as a good predictor in LAD ($\chi^2=17.50$, P<0.0001).

Abbreviations: bFGF, basic fibroblast growth factor; LAD, lung adenocarcinoma; TSP-1, thrombospondin-1; VEGFA, vascular endothelial growth factor A; VEGFR2, vascular endothelial growth factor receptor-2.

Compared with the OE NC group, the expression levels of VEGFA, VEGFR2, and bFGF mRNA (t=15.796, 16.041, 15.578; P=0.003, 0.002, 0.003) in the OE group were increased, while TSP-1 mRNA level was decreased (t=6.244; P=0.025). Compared with the siRNA NC group, the expression levels of VEGFA, VEGFR2, and bFGF mRNA (t=5.399, 5.903, 5.556; P=0.032, 0.023, 0.036) were decreased in the siRNA group, while TSP-1 mRNA level was increased (t=14.477; P=0.005) (Figure 4A).

These results confirmed that after inhibition or overexpression of LOC100132354, the key molecular proteins and mRNA levels in the VEGFA/VEGFR2 signaling pathway of the cell line were significantly changed.

VEGFA is a downstream target gene of LOC100132354

LOC100132354 RNA expression level showed no obvious changes after VEGFA knockdown (t=0.538; P=0.644) or



Figure 3 The proteins of VEGFA, VEGFR2, bFGF, TSP-1, Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mekl/2, and P-Erk1/2 from different cell lines by Western blotting. Notes: (A) The Western blotting figure of these proteins. (B) The results of VEGFA, VEGFR2, bFGF, TSP-1 in the four groups. (C) The results of Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mekl/2, and P-Erk1/2 in the four groups. *P<0.05, **P<0.01.

Abbreviations: bFGF, basic fibroblast growth factor; OE, overexpression; NC, negative control; TSP-1, thrombospondin-1; VEGFA, vascular endothelial growth factor A; VEGFR2, vascular endothelial growth factor receptor-2.



 $\label{eq:Figure 4} \textbf{Figure 4} \ \textbf{The expression levels of these genes in each treatment group.}$

Notes: (A) Compared with the OE NC group, the expression level of VEGFA, VEGFR2, and bFGF mRNA (t=15.796, 16.041, 15.578; P=0.003, 0.002, 0.003) are higher in the OE group, while TSP-1 mRNA level was decreased (t=6.244; P=0.025). Compared with the siRNA NC group, the expression level of VEGFA, VEGFR2, and bFGF mRNA (t=5.399, 5.903, 5.556; P=0.032, 0.023, 0.036) are lower in the siRNA group, while TSP-1 mRNA level was increased (t=14.477; P=0.005). (B) LOC100132354 RNA expression level did not change after VEGFA knockdown (t=0.538; P=0.644). (C) LOC100132354 RNA expression level did not change after VEGFA overexpression (t=0.144; P=0.898). *P<0.05, **P<0.01, "P<0.05, "#P<0.01.

Abbreviations: bFGF, basic fibroblast growth factor; OE, overexpression; NC, negative control; TSP-1, thrombospondin-1; VEGFA, vascular endothelial growth factor A; VEGFR2, vascular endothelial growth factor receptor-2.

overexpression (*t*=0.144; *P*=0.898; Figure 4B, C), suggesting that VEGFA was a downstream target gene of LOC100132354.

LOC100132354 significantly enhanced the MVD value of the nude mice

Our previous work showed that LOC100132354 significantly enhanced the tumorigenic ability in the nude mice experiment.¹⁴ Tumorigenesis of nude mice confirmed that the tumor MVD was decreased significantly (t=10.765; P=0.012) after inhibition of LOC100132354 and increased significantly (t=15.879; P=0.009) after overexpression of LOC100 132,354 (Figure 5 and Table 3).

Discussion

Tumor cells secrete VEGFA and bind to the VEGFR2 on the surface of endothelial cells to exert their biological effects.^{17,18,34} Studies have shown that the expression levels of VEGFA and VEGFR2 were significantly elevated in NSCLC tissues,^{35,36} indicating that angiogenesis was significantly enhanced in NSCLC tumors. Steady expression of VEGFA can lead to increase in tumor volume in lung cancer in mice,³⁶ while inhibition of VEGFA signaling pathways resulted in the inhibition of tumor growth and tumor angiogenesis.^{37,38}

In this study, the mRNA expression levels of LOC100132354, VEGFA, VEGFR2, and bFGF were significantly higher than the corresponding NT tissues, while TSP-1 mRNA are significantly lower than the corresponding NT tissues in LAD. The survival analysis showed that VEGFA, VEGFR2, and bFGF were poor predictors, while TSP-1 was a good predictor in LAD. These results revealed that the angiogenic capacity was enhanced significantly and could predict the prognosis of LAD.



Figure 5 Changes of MVD expression in tumors of nude mice. **Notes:** The tumorigenesis of nude mice confirmed that after inhibition of LOC100132354, the tumor MVD was decreased significantly (t=10.765; P=0.012) and after overexpression of LOC100 132354, the MVD was increased significantly (t=15.879; P=0.009). *P<0.05, ##P<0.01.

Abbreviations: MVD, microvessel density; OE, overexpression; NC, negative control.

Table 3 Comparison of MVD in nude mouse tumors ($x\pm$ SD) (n=25)

Group	MVD	<i>P</i> -value
siRNA NC	14.21±1.32	
siRNA	6.01±0.92	0.012
OE NC	10.01±1.01	
OE	18.12±1.21	0.009

Abbreviations: MVD, microvessel density; OE, overexpression; NC, negative control.

In our previous studies, we found that LOC100132354 increased the ability of colony formation, cell migration, adhesion, proliferation, invasion, S phase, and G2/M phase cells, while decreased G0/G1 phase cells and apoptosis.¹⁴ In this study, we further confirmed that the expression levels of VEGFA, VEGFR2, bFGF, Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mekl/2, and P-Erk1/2 were significantly higher from the OE group than those of the OE NC group, while TSP-1 expression levels were decreased. This showed that overexpression of LOC100132354 activated VEGFA/VEGFR2 pathway, further activating the downstream molecular Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mekl/2, and P-Erk1/2. Otherwise, VEGFA raised the levels of bFGF, while vascular growth negative regulator-TSP-1 protein expression levels were decreased. The expression levels of VEGFA, VEGFR2, bFGF, Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mekl/2, and P-Erk1/2 in the siRNA group were lower than those of the siRNA NC group, and only TSP-1 expression level was higher than that of the siRNA NC group. This suggested that the knockdown of LOC100132354 inactivated VEGFA/ VEGFR2 pathway, further inactivating the downstream molecular Ras, P-A-Raf, P-B-Raf, P-C-Raf, P-Mekl/2, and P-Erk1/2. Otherwise, VEGFA decreased the levels of bFGF, while TSP-1 expression levels were increased. So, the knockdown and overexpression experiments consistently confirmed that LOC100132354 regulated the angiogenic capacity by VEGFA/VEGFR2 pathways in LAD.

LOC100132354 RNA expression levels showed no changes after VEGFA knockdown or overexpression, otherwise LOC100132354 could significantly enhance the MVD value of the nude mice. This suggested that VEGFA was a downstream target gene of LOC100132354.

Conclusion

Our study ascertained that LOC100132354 increased the angiogenic capacity and the mechanism of regulation of angiogenic capacity in LAD. Our results confirmed that lncRNA LOC100132354 can act as an antiangiogenic target in LAD.

Acknowledgments

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Disclosure

The authors report no conflicts of interest in this work. There are not any of the family relationship between such authors.

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