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Feeding brown fat: dietary phytochemicals targeting non-shivering thermogenesis to control body weight

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Excessive adipose accumulation, which is the main driver for the development of secondary metabolic complications, has reached epidemic proportions and combined pharmaceutical, educational and nutritional approaches are required to reverse the current rise in global obesity prevalence rates. Brown adipose tissue (BAT) is a unique organ able to dissipate energy and thus a promising target to enhance BMR to counteract a positive energy balance. In addition, active BAT might support body weight maintenance after weight loss to prevent/reduce relapse. Natural products deliver valuable bioactive compounds that have historically helped to alleviate disease symptoms. Interest in recent years has focused on identifying nutritional constituents that are able to induce BAT activity and thereby enhance energy expenditure. This review provides a summary of selected dietary phytochemicals, including isoflavones, catechins, stilbenes, the flavonoids quercetin, luteolin and resveratrol as well as the alkaloids berberine and capsaicin. Most of the discussed phytochemicals act through distinct molecular pathways e.g. sympathetic nerve activation, AMP-kinase signalling, SIRT1 activity or stimulation of oestrogen receptors. Thus, it might be possible to utilise this multitude of pathways to co-activate BAT using a fine-tuned combination of foods or combined nutritional supplements.

Brown adipose tissue: Browning: Energy expenditure: Phytochemicals: Weight management

Obesity is a major public health threat of the 21st century, both from an individual as well as from an economic point of view. Epidemiological data from extreme cases such as the USA with more than 70% of adults being overweight or obese propose that obesity has sadly become the common or wild-type phenotype in certain nations^(1,2). Apart from the cosmetic imperfections arising from an obese phenotype, obesity-related metabolic complications and secondary diseases (different cancers, CVD, osteoarthritis, to name but a few) severely affect life quality and thus drive ongoing research efforts to identify weight lowering or stabilising pharmaceuticals. Possible approaches for the treatment of obesity were expanded by the discovery of physiologically relevant amounts of active brown adipose tissue (BAT) in adult human subjects⁽³⁻⁵⁾. Cold-induced BAT in human subjects negatively correlates with age and $BMI^{(3)}$, while RMR shows a positive correlation⁽⁴⁾. The principal cell within BAT is the highly specialised brown adipocyte, which defends body temperature in a cold environment through non-shivering thermogenesis. This is enabled by uncoupling protein 1 (UCP1),

Abbreviations: AR, adrenergic receptor; BAT, brown adipose tissue; BBR, berberine; BW, body weight; CAP, capsaicin; COMT, catechol-O-methyl-transferase; EE, energy expenditure; EGCG, (–)-epigallocatechin gallate; E2, 17β-oestradiol; ER, estrogen receptor; FGF21, fibroblast growth factor 21; HFD, high fat diet; LSI, lifestyle intervention; OPE, onion peel extract; PGC1a, PPAR γ co-activator 1a; PTS, pterostilbene; PKA, protein kinase A; RSV, resveratrol; SIRT1, sirtuin 1; SUV, standardised uptake value; TRPV1, transient receptor potential vanilloid 1; UCP1, uncoupling protein 1; WAT, white adipose tissue. *Corresponding author: C. Wolfrum, email christian-wolfrum@ethz.ch

localised to the inner mitochondrial membrane, which dissipates energy stored in the mitochondrial proton gradient generated by oxidative phosphorylation⁽⁶⁾. This catabolic process burns large amounts of fatty acids and glucose⁽⁷⁻⁹⁾ and therefore, active BAT likely serves as a nutrient sink to buffer excessive energetic intake or utilises energy reserves mobilised from white adipose tissue (WAT). In response to a cold stimulus, catecholamines are released from sympathetic nerve fibres that innervate BAT and function as inducers of the thermogenic programme. Other endogenous factors including fibroblast growth factor 21 (FGF21), natriuretic peptides, bone morphogenetic protein 8b, glucagon-like peptide 1, thyroid hormones or oestradiol are known modulators of BAT activity or recruitment⁽¹⁰⁻¹²⁾. In addition, the formation of brown-like adipocytes (beige/ brite) within the WAT can be induced by cold acclima-tisation or pharmacological means^(13,14). These beige/ brite adipocytes express UCP1 and thereby extend the pool of thermogenically active cells. In summary, different routes are conceivable of how active brown adipocytes can assist in regulating metabolism and ultimately body weight (BW): (1) activation of existing brown cells within BAT, (2) formation of new brown cells within BAT or (3) induction of WAT browning.

Throughout history, human subjects relied on natural products to counteract disease, due to their richness in bioactive substances. Furthermore, many leading drugs are derived or isolated from natural sources and indispensable for human pharmacotherapy (e.g. morphine, metformin, sodium-glucose co-transporter 2 inhibitors, artemisinin, glucagon-like peptide-1 receptor agonist exendin-4). In view of the extent of the overweight/obesity problem, dietary constituents (phytochemicals) capable of tuning BAT activity could be used to reinforce weight loss programmes or to stabilise BW. A prime example of such a phytochemical is ephedrine, the active principle found in plants of the *Ephedra* genus, whose leaves are traditionally consumed as a tea. Ephedrine is a natural sympathomimetic and an herbal preparation from Ma Huang (72 mg ephedrine daily) significantly reduced BW (-4 v. -0.8 kg) and fat mass (-2.1 v. 0.2 %) in overweight to obese (BMI >29 and $<35 \text{ kg/m}^2$) patients (male n 6, female n 29) within 8 weeks when compared to placebo (male *n* 5, female *n* 28)⁽¹⁵⁾.

Obesity is caused by gradual fat accumulation due to a chronic positive energy balance. An American adult gains about 0.5-1 kg BW annually. The critical gap between energy intake and expenditure that advances weight gain is thus estimated to be as little as 92-682 kJ daily⁽¹⁶⁾. In turn, the total average human BAT mass is approximately 168 g and can burn 301 kJ within 2 h of cold-mediated activation⁽¹⁷⁾. This is exemplified by the synthetic β3-adrenergic receptor (AR) agonist mirabegron, which acutely activates BAT in fluorodeoxygluemission tomography-computed cose positron tomography scans and enhances RMR by approximately 837 kJ daily in men (200 mg) and by 10.7 % (628 kJ daily) in females⁽¹⁸⁾. Chronic mirabegron intake (100 mg daily for 4 weeks) elevates resting energy expenditure by 5.8% (343 kJ daily) compared to baseline and increases BAT volume as well as BAT activity in women⁽¹⁹⁾. These treatment effects were notably more pronounced in women who possessed little BAT at baseline⁽¹⁹⁾. Thus, sustained β 3-AR stimulation is potentially advantageous in subjects with elevated BW associated with lower BAT activity/volume.

It is pertinent to mention that sex-specific differences in human BAT activity, BAT mass and BAT detectability have been delineated in diverse studies, which likely modulate outcomes of pharmacological interventions. Female subjects are more frequently identified as BAT-positive in positron emission tomography-computed tomography scans (female:male ratio 2:1) and display higher BAT mass as well as BAT activity compared to $men^{(3,20-22)}$. Various, yet not entirely resolved factors. such as the expression pattern of sex hormone receptors. levels of sex hormones, body size or the increased sensitivity of females to cold with distinct thermogenic responses might contribute to this sexual dimorphism^(23–25). Interestingly, BMI does not negatively correlate anymore with BAT activity or BAT mass in elderly men (43-82 years), while this correlation persists in female subjects (43-82 years)⁽²⁰⁾. This suggests that older overweight/obese women might benefit more from BAT activation to regulate adiposity than men. Contrarily, Fletcher et al.⁽²⁶⁾ could not confirm differences in cold-induced BAT activity or BAT distribution in healthy men v. women and the apparent higher BAT volume in men vanished after normalisation to body size⁽²⁶⁾.</sup>

This review discusses a selection of phytochemicals present in food or drinks based on scientific evidence for their efficacy in human and animal models to affect weight or energy expenditure (EE). In particular, we provide an overview of how these substances modulate BAT activity or WAT browning at the molecular level.

Stilbenes

Pterostilbene (PTS) is the dimethylated derivative of resveratrol (RSV) and its potential weight-lowering capacity has only recently gained attention. Interest in PTS rose when Rimando et al.⁽²⁷⁾ demonstrated that PTS strongly induced PPAR α activity in a rat hepatocyte cell line overexpressing a PPRE-luciferase reporter gene⁽²⁷⁾. PTS at $100 \,\mu\text{M}$ displayed 2-fold higher PPARa induction than the pharmacologic PPARa agonist ciprofibrate. The anti-hyperlipidaemic activity of PTS was verified in hamsters where PTS-fortification of high fat diet (HFD; 25 mg PTS/kg food) led to lower total and LDL-cholesterol levels⁽²⁷⁾. PPAR α is a nuclear transcription factor, which is highly enriched in the liver as well as BAT v. WAT where it controls a myriad of genes involved in fatty acid uptake as well as β - and per-oxisomal lipid oxidation^(28,29). The group of Maria Portillo examined the effect of PTS on adipose tissue in rat models of genetic and dietary-induced obesity^(30,31). In Zucker fatty rats, the daily oral application of 15 mg PTS/kg for 6 weeks resulted in significantly less total fat (10% v. 13.2%) compared to untreated animals⁽³⁰⁾.

During the dietary intervention, Wistar rats were fed an adipogenic high fat-high sucrose (20% each) diet enriched with PTS to ensure an estimated daily intake of either 15 mg/kg or 30 mg/kg BW. Although no effect on weight gain or BW was observed, total adipose tissue mass was dose-dependently reduced in comparison with control animals $(40.3 \text{ g}, 36.6 \text{ g}, 47.5 \text{ g})^{(31)}$. Moreover, enzymatic carnitine palmitoyltransferase 1a activity was higher in the liver while fatty acid synthase activity was blunted in $WAT^{(31)}$. The described phenotype might result from an interplay between increased fatty acid oxidation due to higher fatty acid import into mitochondria via carnitine palmitovltransferase 1a and reduced adipocyte lipogenesis due to low fatty acid synthase activity. Unfortunately, the only published human clinical trial with PTS as investigatory compound could not reproduce these findings from rats. Instead, 8 weeks of daily PTS ingestion (0, 100 or 250 mg daily) increased LDL-cholesterol and consequently total cholesterol levels in a cohort of male and female hypercholesterolemic Caucasian and African Americans⁽³²⁾. Overall, a mild but significant effect of PTS on BMI was observed when participants were stratified for cholesterol medication. BAT possesses a tremendous oxidative capacity and relies heavily on substrate e.g. fatty acid oxidation to fulfil its thermogenic function. PPARa agonists are known to induce UCP1 expression in murine BAT⁽³³⁾ and β -AR signalling increases PPAR α expression in brown adipocytes⁽³⁴⁾. To date, three publications have addressed the impact of PTS-fortified diet on WAT browning or BAT functionality in rodent models. Aguirre et al.⁽³⁵⁾ found that 15 or 30 mg PTS/kg BW upregulated various brown and mitochondrial marker genes in male Zucker fa/fa rats when administered daily for 6 weeks⁽³⁵⁾. These changes were accompanied by increased PPAR α and UCP1 protein abundance in the BAT. Additionally, higher carnitine palmitoyltransferase 1α activity was measured in the intrascapular BAT lysate of PTS-rats compared to controls⁽³⁵⁾. These molecular alterations imply that PTS enhances the oxidative capacity of BAT in genetically obese rats that translated to a beneficial phenotype as total adipose tissue weight and BW were reduced in PTS-rats⁽³⁵⁾. This conclusion is substantiated by data from Nagao and co-workers⁽³⁶⁾ who applied a 10× higher PTS dose (300 mg/kg BW daily) as a food supplement to male, genetically hyperphagic rats, which again caused lower body fat accumulation⁽³⁶⁾. Compared to control-fed animals, PTS stimulated oxygen consumption (3.91 v. 3.69 litre/ 100 g BW daily) and enhanced EE (80.4 v. 77.1 litre/ 100 g BW daily) after 4 weeks of feeding⁽³⁶⁾. The observed reduction in the respiratory quotient (0.82 v. 0.85) of PTS-treated rats was in agreement with higher lipid oxidation during the metabolically active dark phase⁽³⁶⁾. Interestingly, Nagao *et al.*⁽³⁶⁾ not only confirmed that PTS is a PPARa agonist, but also discovered that PTS at 50-250 µm increased sirtuin 1 (SIRT1) activity⁽³⁶⁾. SIRT1 is a NAD+-dependent protein deacetylase and modulates the functional activity of various enzymes and transcription factors. Deacetylation of PPAR γ by SIRT1 is crucial for browning of WAT as it

enables the recruitment of the PPARy coactivator PRDM16, which functions as a master regulator of the thermogenic gene programme in WAT^(37,38). In turn, food deprivation or pharmacological SIRT1 activation have been shown to blunt the PPARy-dependent expression of lipogenic genes in WAT through SIRT1-mediated blockade of the PPAR γ -coactivator NCoR⁽³⁹⁾. This repressive effect on PPARy promotes lipolysis and reduces fat storage in WAT⁽³⁹⁾. SIRT1 additionally affects mitochondrial biogenesis by two converging mechanisms. First, SIRT1 deacetylates and activates liver kinase B1, an upstream kinase and activator of AMPK, which in turn phosphorylates PPARy co-activator 1α (PGC1 α)⁽⁴⁰⁾. Secondly, SIRT1 directly deacety-lates PGC1 α ⁽⁴¹⁾. Consistent with the relevance of SIRT1 in WAT browning, chronic PTS administration to HFD-fed mice (90 mg/kg daily) caused lower weight gain and a trend towards browning of the inguinal WAT as measured by upregulation of browning-specific marker genes (CIDEA, PGC1a, EBF2, PPARy and TBX1)⁽⁴²⁾. The transcriptional and morphological changes during WAT remodelling in response to cold exposure are independent of PPARa as PPARa knockout mice display equal browning when compared with wild-type animals⁽⁴³⁾. Collectively, it seems that SIRT1 agonism by PTS is the dominant molecular trigger, which confers the physiological effects of long-term PTS consumption such as browning of WAT, enhanced oxidative capacity, lipid mobilisation and increased EE. The pharmacokinetic analysis of orally applied PTS (168 mg/kg BW) in rats revealed a high bioavailability of about 80% with a peak plasma PTS concentration of approximately $30 \,\mu M^{(44)}$. This dosage corresponds to an approximated human equivalent dose of 27 mg/ $kg^{(45)}$ and up to 250 mg/kg BW of PTS is evidentially safe for human $use^{(46)}$.

RSV attracted the interest of scientific community after Howitz et al.⁽⁴⁷⁾ identified RSV as a powerful small molecule activator of SIRT1⁽⁴⁷⁾. SIRT1 operates as one of the molecular regulators essential for the beneficial physiological effects of energetic restriction⁽⁴⁸⁾. This discovery introduced the idea that RSV might be a natural substance able to ameliorate obesity by mimicking a low energy state. Lagouge et al.⁽⁴⁹⁾ first demonstrated that a RSV-enriched (400 mg/kg BW daily) diet impairs weight gain in male HFD-fed mice⁽⁴⁹⁾. RSV mice displayed enhanced oxygen consumption, improved cold tolerance during an acute cold challenge as well as increased mitochondrial content in BAT. A geneenrichment analysis confirmed the induction of genes related to mitochondrial biogenesis and function after RSV therapy in muscle. The measured increase in EE was not attributable to spontaneous locomotor activity, suggesting that RSV modifies BAT functionality as a driver of adaptive thermogenesis. Another study explored the effects of lifelong RSV-containing HFD (0.04 % w/ w) on metabolic health in middle-aged, male mice⁽⁵⁰⁾. Here, RSV intake did not affect BW, possibly due to the lower concentrations used, but significantly attenuated the signs of ageing such as elevated fasting blood glucose levels, insulin resistance and pathological organ

male, non-human primates, 4 weeks daily RSV ingestion (200 mg/kg BW) suppressed BW gain and enhanced RMR by 29% compared to baseline⁽⁵¹⁾. In addition, the body temperature difference between the active phase and the hypothermic light phase was reduced, suggesting a thermogenic activity $^{(51)}$. To date, direct, robust evidence for a contribution of BAT to the systemic effects of RSV is scarce. Two months of daily RSV feeding of male mice (400 mg/kg BW) on a standard diet led to lower epididymal and retroperitoneal adipose tissue weight, higher oxygen consumption and about 2-fold upregulation of UCP1 and SIRT1 mRNA expression in BAT of RSV mice⁽⁵²⁾. The authors hypothesise that fat stores are mobilised to accommodate the increased fatty acid demand of more active BAT, which in turn lowers adiposity. However, no histological sections with UCP1-immunohistochemistry, UPC1 protein levels or mitochondrial parameters were presented to strengthen such a statement. The laboratory of Du⁽⁵³⁾ demonstrated the formation of brown-like adipocytes in the presence of 10 um RSV, when stromal vascular cells isolated from WAT were differentiated with a brown adipogenic cocktail⁽⁵³⁾. These brown-like adipocytes upregulated an array of essential brown marker genes. This finding was affirmed in female CD-1 mice, where RSV (0.1% in HFD) slowed weight gain and induced the same thermogenic gene set in the inguinal WAT when compared to controls⁽⁵⁴⁾. Consistent with the known morphological features of browning, the inguinal WAT of RSV-fed mice exhibited adipocytes with multilocular lipid droplets and a shift towards smaller cells. RSV-treated mice and cells further displayed increased metabolic rates and enhanced lipid oxidation. More intriguingly, all the specified molecular alterations and the consequent physiological outcomes were not observed when experiments were repeated in the absence of $AMPK^{(54)}$. Using the same experimental set-up, Wang et al.⁽⁵⁴⁾ counted more brown adipocytes in histological BAT sections, which implies that RSV might provoke the formation of brown adipocytes, in vivo. In parallel, RSV feeding resulted in higher UCP1 and PRDM16 protein content as well as the augmented levels of phosphoAMPK⁽⁵⁴⁾. Similar findings including reduced amounts of acetylated PGC1a were reported for the brown fat in male rats treated with $30 \text{ mg/kg BW}^{(55)}$.

changes⁽⁵⁰⁾. In a before-after study performed in six

Contrary to the overwhelming amount of literature addressing the benefits of RSV in animal models, no human clinical trials exist that specifically use EE, BAT activation or weight management as readouts. A plethora of studies (supplementary material) conducted in different target groups with daily RSV doses from 75 up to 2000 mg did not detect any effects of RSV on key metabolic parameters, anthropometric measures or molecular downstream markers of SIRT1 activation^(56–58). In strong contrast stands a cross-over trial completed in obese, healthy men, where daily RSV (150 mg) intake for 30 d modestly mimicked the physiological effects resembling energetic restriction⁽⁵⁹⁾. The participants had lower resting and sleeping metabolic rates (2–4 % lower EE), higher daily respiratory quotient as indicator for improved

metabolic flexibility, higher SIRT1 and phosphoAMPK protein abundance in muscle biopsies plus elevated carbonyl cyanide-4-(trifluoromethoxy)phenylhydrazoneuncoupled respiration. This striking discrepancy in study outcome might be due to such factors as the extent of obesity, degree of insulin resistance, age, sex, sample size, parallel *v*. crossover design (seasonal impact) and blood RSV concentrations^(56–59).

The structural similarity between PTS and RSV implies that they might share the same molecular mode of action. SIRT1 agonism. Nonetheless, the methoxy groups in PTS protect it from hepatic metabolisation and possibly cause the high oral bioavailability of PTS $(80\%)^{(44)}$. In contrast, the low oral bioavailability of RSV is one of its major drawbacks and might account for the poor reproducibility as well as comparability of human RSV trials. Although RSV is well absorbed in the intestine through passive diffusion (70%), the phenolic groups are rapidly glucuronidated or sulphonated in the liver and leave only 1.5% of RSV unmodified⁽⁶⁰⁾. Notably, inter-species differences exist in the formation of glucuronidated RSV (g-RSV) metabolites. In rats and mice, 90% of an applied RSV dose is glucuronidated compared to 65 % in human subjects and dogs. Lastly, enzymatic stereoselectivity leads to distinct ratios of g-RSV types: the dominant type (RSV-4'G) in human subjects and dogs is undetectable in mice and rats⁽⁶¹⁾. Consequently, findings from rodent studies are difficult to translate to human subjects and other models might be required to assess the physiological bioactivity/efficacy of RSV metabolites. The accepted difficulty in producing reproducible and meaningful effects with RSV in human subjects makes it currently unlikely to be upgraded from an over the counter nutritional supplement to a pharmacological remedy.

Flavonoids

Ouercetin is particularly abundant (16.5 mg/g) in the onion peel extract (OPE: 16.5 mg/g), which is frequently used as a quercetin matrix in animal experiments⁽⁶²⁾. OPE reduced</sup> diet-induced obesity by 6% and visceral fat in male rats when added to HFD (3.6 g/kg food) for 8 weeks⁽⁶³⁾. Ting et al.⁽⁶⁴⁾ described an even more pronounced weight phenotype (20 % difference in weight gain) for a quercetinrich supplement in male HFD-fed rats (185, 270 or 925 mg/kg BW daily), demonstrating that quercetin likely is the active ingredient in the $OPE^{(64)}$. In an extensive study with Zucker fatty rats, quercetin was provided daily via the food (10 mg/kg BW) and profoundly reduced weight gain when combined with either standard diet (12 v. 26%) or HFD (26 v. 33%) over a 10 week period⁽⁶⁵⁾. Quercetin-treated rats showed improved insulin sensitivity, reduced endothelial nitric oxide synthase protein and inflammatory TNF α release in the visceral WAT. Unfortunately, the aforementioned studies did not evaluate EE or physical activity as potential contributors.

In male mice, HFD enriched with 5 g OPE/kg for 8 weeks did not attenuate weight or fat mass compared to plain HFD but upregulated markers of WAT browning⁽⁶⁶⁾. One possibility for the lack of a weight phenotype

could be due to a stronger obese phenotype or different quercetin contents in the OPE (68 v. 276 mg/g dry weight). Browning of murine, male WAT following 12 weeks HFD feeding with pure quercetin (1 g/kg food) was confirmed by Kuipers *et al.*⁽⁶⁷⁾. Classical BAT was not affected by the treatment and browning was insufficient to modulate BW or EE, in accordance with the results from Stewart et al.⁽⁶⁸⁾ at a relatively high quercetin dose of 8 g/kg HFD⁽⁶⁸⁾. A recent report⁽⁶⁹⁾ describes browning of WAT alongside enhanced BAT-UCP1 content in male mice fed with HFD + 0.05% quercetin⁽⁶⁹⁾. Plasma norepinephrine concentrations were about three times higher in quercetin-exposed animals, which suggests an intensified sympathetic nervous activity as the underlying effector. Furthermore, protein kinase A (PKA) protein levels were elevated in subcutaneous WAT of quercetin mice. However, cAMP levels or phosphoPKA substrates, which are surrogate markers of adrenergic stimulation, were not determined. It remains to be shown whether these molecular adaptations translate into a distinct phenotype as body weight or EE were not measured⁽⁶⁹⁾. Higher postprandial norepinephrine levels were also found in male calves fed quercetin (50 mg/kg BW) supplemented milk replace-ment for 1 week⁽⁷⁰⁾. In human liver lysates, quercetin and its glycoside rutin showed catechol-O-methyltransferase (COMT) inhibitory activity at IC_{50} of 5.3 and 10.8 µM, respectively and could potentiate norepinephrine action⁽⁷¹⁾. Furthermore, quercetin dose dependently (1-250 µm) reinforced norepinephrine and isoproterenol-induced lipolysis in isolated male rat adipocytes by increasing cAMP concentrations⁽⁷²⁾. Even though the effects of quercetin on BW are inconclusive, quercetin might assist in improving metabolic homoeostasis. Notably, inter-species differences in quercetin efficacy are obvious as most studies in rats showed a weight-lowering effect after 8-12 weeks of intervention, which was mostly absent in mice. Surprisingly, an extended quercetin feeding diet regimen over 12 weeks (1 g/kg HFD) in male mice added up to reduced terminal BW because of a lower adipose tissue mass⁽⁷³⁾. In these mice, adipose tissue-resident macrophages and mast cells were significantly lowered, which caused attenuated visceral adipose tissue as well as systemic inflammation. Furthermore, UCP1 mRNA was upregulated in the BAT after quercetin treatment, suggesting a potential thermogenic effect⁽⁷³⁾. At a molecular level, quercetin led to higher SIRT1 protein amounts and phosphoAMPK levels in the visceral WAT⁽⁷³⁾. The antiinflammatory property of activated SIRT1 is well established^(74,75). Induction of the SIRT1-liver kinase B1-AMPK pathway as a mediator of quercetin's antiinflammatory activity was confirmed in bone-derived macrophages stimulated with lipopolysaccharides to mimic obesity-associated inflammation⁽⁷⁶⁾. Under this (20 µм) condition, quercetin increased SIRT1, phosphoAMPK and phospho liver kinase B1 protein levels in parallel to a heightened intracellular AMP: ATP ratio⁽⁷⁶⁾.

Although promising effects regarding BW and inflammatory states are evident in rodent studies, the current available human clinical trials do not reproduce this picture. An up to date meta-analysis covering doses from 100 up to 1000 mg daily and intervention durations from 2 to 12 weeks did not find any beneficial clinical effects of OPE or quercetin intake on human $BW^{(77)}$. The absence of any human physiologically relevant effect is underlined by the studies summarised in the supplementary material, mainly performed in Korean subjects^(78–80).

It is striking that circulating quercetin concentrations reported in rodent studies appear much higher than in human subjects. In male mice, 5.3 g/kg BW daily quercetin related to 90 μ M of fasting quercetin levels⁽⁸¹⁾ and a lower dose (100 mg/kg BW) to 10 µM after 12 weeks feed $ing^{(73)}$. In male and female human subjects, 150 mg daily over 5 weeks resulted in a mean fasting plasma quercetin concentration of $0.270 \,\mu\text{M}^{(82)}$, 500 mg daily over 12 weeks in about $1.3 \,\mu\text{m}^{(83)}$. Surprisingly, the mouse dose of 100 mg/kg BW translates to a human equivalent dose of 8 mg/kg BW and in human subjects, 500 mg daily reflects 6.5 mg/kg BW as calculated by the mean BW in this cohort. Consequently, although the doses lie in a comparable range, the circulating quercetin concentration in human subjects is roughly ten times lower. This comparison highlights potential inter-species differences in quercetin metabolism, absorption and excretion that affects final plasma concentrations and potentially the therapeutic outcome. Considering that human quercetin intake is safe up to 5 g daily⁽⁸⁴⁾, clinical trials of extended duration (17 weeks) and at higher doses are required to conclude on the efficacy of quercetin in human subjects.

Luteolin has been extensively studied due to its antiinflammatory activity and research focusing on weight loss following luteolin intake is in its early stages and limited to rodent studies. Distinct long-term dietary interventions with luteolin-enriched HFD (12 weeks, 0.01 % w/w or 16 weeks, 0.005% w/w) significantly reduced BW gain in male mice⁽⁸⁵⁻⁸⁷⁾. This phenotype was due to reduced visceral and subcutaneous WAT accumulation with a lower proinflammatory state of the WAT (e.g. macrophage infiltration, lower M1:M2 ratio)^(85–87). Kwon *et al.*⁽⁸⁶⁾ measured enhanced faecal lipid output in the luteolin group and higher rates of lipolysis in the WAT, which likely influenced adiposity⁽⁸⁶⁾. Luteolin was shown to elevate PPARy protein levels and tranadipocytes⁽⁸⁸⁾. 3T3-L1 scriptional activity in Modulation of PPARy might contribute to the healthier adipose tissue expansion but cannot explain the reduction in adipose tissue mass as established pharmacological PPARy agonists (thiazolidinediones) tend to increase fat $mass^{(89)}$. It is reasonable to assume that the effect of luteolin on BW is polymodal and due to many physiological changes such as attenuated inflammation, reduced intestinal lipid absorption and more lipolytic WAT that fuels activated BAT. The latter assumption comes from a study by Zhang *et al.*⁽⁹⁰⁾ who demonstrated higher EE in luteolin-HFD (0.01 % w/w) fed male mice compared to untreated mice⁽⁹⁰⁾. The histological examination of the BAT revealed an increased density of UCP1 + cells in the luteolin group combined with an induction of the thermogenic programme at the mRNA and protein levels. Moreover, browning of subcutaneous WAT was detected. Interestingly, this phenotype was preserved when HFD was replaced by standard diet and proves the inherent thermogenic activity of luteolin⁽⁹⁰⁾. At the molecular level, luteolin stimulated AMPK phosphorylation in vivo and the beneficial effects of luteolin on thermogenic genes was abolished in primary brown and white adipocytes when cells were pre-exposed to the AMPK inhibitor compound C. It appears that luteolin promotes an activated brown phenotype through the AMPK-SIRT1-PGC1a axis and human clinical studies are necessary for proof of concept/efficacy. Newest findings by Zhang *et al.*⁽⁹¹⁾ illustrate a significant role of mast cells in energy homoeostasis⁽⁹¹⁾. Genetic ablation or pharmacologic inactivation of mast cells in male mice amplified acute norepinephrine-triggered oxygen consumption and increased body temperature. In both conditions, massive browning occurred in the subcutaneous WAT together with higher UCP1 protein levels and the upregulation of thermogenic genes⁽⁹¹⁾. The authors claim that serotonin secretion from mast cells inhibits platelet derived growth factor receptor α + progenitor cell proliferation that can give rise to beige/brite adipocytes. In agreement with the anti-inflammatory features common to quercetin and luteolin as well as the attenuation of mast cells' infiltration into WAT following the chronic consumption of these compounds, one can speculate that quercetin and/or luteolin enhance thermogenesis and systemic EE by WAT browning, due to lower mast cell-derived serotonin release.

Catechins

Catechins are named after the catechu, an aqueous extract obtained from the wood of the acacia tree (Senegalia catechu) and rich in these polyphenolic substances. (-)-Epigallocatechin gallate (EGCG) and (-)-epigallocatechin are the most prevalent catechins in green tea infusions (92-94). The first notion that green tea/catechin consumption might affect BW traces back to a study investigating the effect of a standardised green tea extract on 24-h EE in healthy men⁽⁹⁵⁾. Surprisingly, three times daily intake of a combined caffeine (50 mg) plus EGCG (90 mg) preparation increased total 24-h EE by 3.5% whereas the caffeine (50 mg) only group did not show any effects⁽⁹⁵⁾. These findings were confirmed by Rudelle et al.⁽⁹⁶⁾, who measured a comparable 4.6% elevation (444 kJ daily) in the 24-h EE in lean men and women after a 3 d treatment diet regimen consisting of 282 mg EGCG and 300 mg caffeine daily⁽⁹⁶⁾. Bérubé-Parent et al.⁽⁹⁷⁾ next aimed to identify the necessary EGCG content in green tea capsules at a fixed caffeine dose (200 mg) to maximise the mixture's impact on 24-h EE in men⁽⁹⁷⁾. Consistent with previous reports, three times daily ingestion of the mixture enhanced 24-h EE at any EGCG concentration (minimum 270 mg daily up to maximum 1200 mg daily); however, no significant dose-response was observed. It seems plausible that the thermogenic activity of the ingested green tea mimetic is mediated either solely by EGCG and/or by a synergistic action of caffeine and EGCG,

whereas the effective amount of EGCG reaches a threshold beyond which no additional benefits on EE are to be expected. A 3 d trial focusing on the differential effects of EGCG or caffeine alone on EE in men (mean BMI 31) did not find any relevant impact at different EGCG concentrations (300 v. 600 mg) nor for caffeine (200 mg) or in combination⁽⁹⁸⁾. Nevertheless, EGCG enhanced fat oxidation as determined by the respiratory quotient 2 h after an overnight fast by 7% in the 300 mg EGCG group as well as in the postprandial phase, 2 h post-meal by 33%. The contradictory results concerning EE may be due to the mode of administration e.g. capsules v. beverage, the ethnicity of the participants⁽⁹⁹⁾, the degree of overweight/obese subjects and/or the utilised caffeine dose. Accordingly, EGCG-caffeine supplements seem to be more effective on weight loss or weight maintenance in subjects with low habitual caffeine intake^(99,100). A meta-analysis evaluating the influence of short-term EGCG consumption on EE and fat oxidation concluded that EGCG moderately boosts metabolic rate as indicated by significantly lower respiratory quotient and higher $EE^{(101)}$.

A large body of literature^(102–107) (supplementary material) has examined whether the possible thermogenic activity of catechins translates to a weight loss phenotype during extended supplement interventions. Overall, the effect of catechins on weight maintenance or loss as measured by BW, BMI and waist circumference is significant when combined with caffeine whereas only a mild clinical impact can be implied based on the described effect sizes^(99,108).

The exact molecular events as well as the physiological targets, which trigger fat loss and elevated EE in response to catechin ingestion are still under discussion. However, the general consent supported by solid experimental evidence points at the activation and/or recruitment of BAT as likely effectors. In male rats, the supplementation of HFD with green tea extract (20 mg/g) over 2 weeks lowered body fat accumulation and boosted EE⁽¹⁰⁹⁾. This phenotype was accompanied by higher BAT weight with increased total protein content, which is indicative for enhanced thermogenic BAT capacity. In turn, the positive impact of the green tea extract was abolished when the food was spiked with the β -AR inhibitor, propranolol⁽¹⁰⁹⁾, suggesting that noradrenergic stimulation is required for the observed phenotype. In line with this, EGCG or tea catechins do not directly affect UCP1 mRNA expression levels in BAT. Imaizumi and coworkers⁽¹¹⁰⁾ reported that in an 8 week dietary intervention a minor (70%) increase in UCP1 mRNA only when the tea catechins (40 % EGCG, 0.1 % caffeine) were administered in low fat diet to male rats⁽¹¹⁰⁾. Functionally relevant UCP1 protein abundance was not examined. In comparison, an EGCG (94%) enriched green tea extract did not alter BAT-UCP1 mRNA levels in male mice after 4 weeks feeding with HFD although total body fat content was reduced⁽¹¹¹⁾. An initial hypothesis proposed that the thermogenic action of EGCG originates from its ability to inhibit the COMT. resulting in higher norepinephrine levels in the postsynaptic cleft and therefore higher adrenergic input to

BAT. COMT methylates, inactivates and initiates the degradation of catecholamines and thereby controls the availability as well as activity of these neurotransmitters. Catechins were shown to exert COMT-inhibitory activity in human subjects, mouse and rat liver extracts, although the determined IC_{50} values range from 70 nM up to 1, 15 or 55 μ M depending on the substrate, catechin type or enzyme preparation used^(112–114). Nevertheless, the oral bioavailability of catechins is very low in men and women and plasma concentrations account for approximately 0.18% of ingested catechins (400 mg) from black tea $^{(115)}$. A pharmacokinetic analysis of daily green tea consumption (615 mg daily) detected peak plasma catechin levels 2 h post-prandially reaching roughly 85 ng/ml whereas EGCG levels were about 50 $nM^{(116)}$. Additionally, catechins are estimated to be cleared from the body within 10–12 h after intake⁽¹¹⁷⁾. Considering these pharmacokinetic parameters, circulating catechin concentrations are unlikely to reach the required IC₅₀ to exert a meaningful inactivation of COMT. Lorenz *et al.*⁽¹¹⁸⁾ proved that even a high dose of EGCG (750 mg) does not inhibit COMT activity in vivo when measured in human erythrocytes⁽¹¹⁸⁾.

An elegant, placebo-controlled crossover study performed in men⁽¹⁰²⁾ revealed the direct involvement of BAT in the acute thermogenic effect of a catechin-rich beverage (615 mg). The participants were allocated to either low- or high-BAT groups based on their BAT activity measured with fluorodeoxyglucose positron emission tomography-computed tomography after 2 h of cold acclimatisation. Only the high BAT group displayed a significant increase in EE after the consumption of the catechin drink while there was no difference between the other $groups^{(102)}$. Using the same technique, the authors found that prolonged daily catechin supply can increase cold-induced thermogenesis in the low-BAT group, which is paralleled by enhanced fat oxidation⁽¹⁰²⁾. Also, in healthy Japanese women a daily 540 mg dose of catechins increased BAT density in the supraclavicular region⁽¹⁰³⁾. These data indicate that catechins can recruit BAT to a physiologically relevant extent. Similar to capsinoids or capsaicinoids (described later), catechins might act through the transient receptor potential (TRP) vanilloid 1 (TRPV1) channels as first anticipated due to their astringent taste. TRPV1 are located on sensory neurons that signal to the central nervous system causing increased catecholamine release to stimulate BAT functionality. In vitro, EGCG has been shown to activate TRP channels in primary cultures of murine dorsal root ganglion neurons as well as in intestinal STC-1 cells^(119,120). The possible excitation of sensory neurons by catechins provides a plausible explanation for the observed threshold-dose of effective EGCG. Chronic exposure of TRP to high levels of capsinoids can induce receptor desensitisation and a comparable phenomenon might happen with high catechin $use^{(121)}$. In addition, intestinal catechin levels are not affected by the low systemic bioavailability and are likely sufficient to elicit TRP. Interestingly, there seems to be a significant interaction between EGCG (100 µM) and a priming dose of the β 3-receptor agonist ephedrine (0.1 μ M) on oxygen

consumption in male rats' BAT explants. The effect of EGCG on BAT respiratory rates was higher in the presence of ephedrine and was more profound with increasing ephedrine levels⁽¹²²⁾. TRP channels are multimodal sensors that integrate distinct signals to fine tune their activity and modulate cellular events. PKA is the downstream effector kinase of β -AR signalling and known to positively affect TRPV1 efficiency⁽¹²³⁾. Thus, EGCG signalling via TRPV1 might be effectively enforced by the β 3-AR–PKA axis. However, up to date corresponding human data are lacking to support the hypothesis concerning catechins and TRP.

Isoflavones: phytoestrogens

Sova represents the most dietary relevant isoflavone source with daidzein and genistein being the most prevalent forms and thus focus of this review. The positive nutritional profile of soya, specified by high protein, high isoflavone and low SFA contents, has long raised interest in weight management strategies, in particular due to favourable satiety and appetite $control^{(124)}$. Genistein/daidzein share structural similarity to the female sex hormone 17β-oestradiol (E2), which not only masters female reproduction but also profoundly integrates into energy homoeostasis by controlling appetite as well as BAT and WAT functionality through central and peripheral action. Centrally, E2 acts on the ventromedial hypothalamus where it inhibits AMPK-signalling via estrogen receptor (ER) a and enhances sympathetic tone towards BAT and upregulates BAT UCP1 and PGC1 $\alpha^{(125)}$. Similar to postmenopausal women, loss of E2 signalling in ovariectomised rats stimulates hyperphagia with resultant weight gain. E2 administration reverses this phenotype by increasing EE through the ERα-sympathetic nervous system-BAT axis and decreasing food intake. E2 is a membrane permeable, hydrophobic molecule and able to tune brown adipocyte function directly by signalling through ER α , ER β or g-protein coupled receptor 30^(126,127). In cultured brown adipocytes isolated from male mice, E2 increases β 3-AR mRNA expression and exerts the opposite effect on inhibitory α 2-levels, which could potentiate sympathetic inputs⁽¹²⁸⁾. E2 further promotes mitochondrial biogenesis by inhibiting phosphatase and tensin homolog, causing more active AKT-signalling and nuclear translocation of nuclear respiratory factor 1, the head of mitochondrial gene transcription^(129,130). Therefore, the question arises whether the oestrogenic potency of dietary isoflavones is effective at shaping energy homoeostasis to a physiologically relevant extent such as weight loss or stabilisation.

In male mice, HFD supplementation with the isoflavone-rich fraction (9 g/kg) of the Kudzu flower lowered weight gain as well as WAT and BAT weight after 7 weeks. Histological analysis of BAT from treated mice revealed more UPC1+ cells, suggesting that isoflavones activate BAT to burn more fatty acids⁽¹³¹⁾. Comparable findings were reported for male CD-1 mice fed with a soya-enriched (25% w/w), high phytoestrogenic diet pared to a soya-free, low phytoestrogenic diet⁽¹³²⁾. This dietary intervention resulted in a 7.6% lower weight gain in HP mice compared to low phytoestrogenic controls with significantly reduced adipose tissue mass. The BAT of HP mice was denser, concomitant with higher EE and apparent cold resistance during an acute cold challenge⁽¹³²⁾</sup>. As anticipated from the efficacy of exogenous oestradiol administration to ovariectomised rats, an isoflavone-rich diet (200 µg/g) prevented the severity of weight gain in these animals post-surgery⁽¹³³⁾. Similar findings, including the upregulation of UCP1 protein levels and higher plasma T3 concentrations, were reported by Lephart *et al.*⁽¹³⁴⁾ using a diet with an isoflavone content of 600 ppm in male rats⁽¹³⁴⁾</sup>. They showed that dietary isoflavone ingestion is effective in a model of low E2 and enhances metabolic rates at least partially due to brown fat activation. Several groups investigated the potency of the individual isoflavone types on weight development and EE in various models. In lean or obese rats, enrichment of high fat or standard diet with daidzein (50 mg/kg BW, 14 d) stabilised BW or induced weight loss compared to controls⁽¹³⁵⁾. Dose-dependent weight loss after daidzein intake (30 d) was also observed in obese male mice at 50 mg/kg BW (42.9 to 33.7 g) and at 100 mg/kg BW (42.5 to 32.4 g) but not at 25 mg/kg BW when compared to vehicle $controls^{(136)}$. Although EE was not measured, UCP1 immunofluorescence imaging showed higher UCP1 content in the BAT of the daidzein-exposed group⁽¹³⁵⁾. In vitro, daidzein stimulated hormone sensitive lipase mediated glycerol release from primary adipocytes from male rats indicating that elevated adipocyte lipolysis supports daidzein-induced weight loss^(136,137). Elevated fatty acid generation in brown adipocyte could enhance EE by directly activating UCP1. Additionally, daidzein was shown to inhibit phosphodiesterase activity at an IC₅₀ of 50 μ M⁽¹³⁸⁾, a critical enzyme which arrests β -AR signalling by inactivating cAMP.

(HP; 150 ppm daidzein, 190 ppm genistein) when com-

The literature is more extensive for the efficacy of genistein, probably due to the higher oestrogenicity^(139,140). In female mice, 8 weeks genistein treatment with HFD (0.25 % w/w) resulted in reduced weight gain and lower WAT expansion than in control HFD animals. Within the WAT, brite adipocyte marker genes were upregulated, which was probably centrally controlled⁽¹⁴¹⁾. Urocortin-3 (UCN3) was among the differentially regulated hypothalamic genes when HFD v. HFD + genistein and HFD v. control animals were compared. UCN3 is an anorexigenic peptide and a known player in energy homoeostasis. Male UCN3 transgenic mice are protected from HFD-induced weight gain⁽¹⁴²⁾ and intracerebroventricular UCN3 administration to male rats provoked thermogenesis⁽¹⁴³⁾ as well as higher BAT UCP1 expression levels⁽¹⁴⁴⁾. In another recent study, genistein-exposed (0.2% w/w) male mice displayed higher EE and the appearance of UCP1+ cells within the subcutaneous WAT compared to chow-fed controls, confirming the role of genistein in adipose browning $^{(145)}$. We confirmed a cell autonomous effect of both genistein and daidzein in mature immortalised brown adipocytes, which

enhanced UPC1 promoter activity in a UPC1-driven luciferase assay and led to higher UCP1 activity as measured by UCP1-immunofluorescence intensity⁽¹⁴⁶⁾. In 4-week old male and female mice, daily administration of genistein (200 mg/kg BW) for 15 d lowered body weight by 4 and 4.3%, respectively due to reduced food intake as well as reduced adipogenic gene expression mediated by $\text{ER\beta}^{(147)}$. In human fetal brown fat, ER α is the dominant oestrogen receptor type⁽¹⁴⁸⁾ but the ratio is unknown in adulthood. Ligand assays unveiled higher affinity of genistein and daidzein for ER β than ER α , demonstrating that isoflavones preferentially activate the oestrogen-response element via $ER\beta$. The EC₅₀s of ER α for genistein (15 µM) and daidzein $(>300 \ \mu\text{M})$ are supraphysiological and beyond circulating levels achievable⁽¹⁴⁹⁾ with dietary isoflavone supplementation (100 mg isoflavone capsules daily; thereof 14 mg daidzein and 3-4 mg genistein)⁽¹⁵⁰⁾. Given these concentrations, it is unlikely that isoflavones exert their physiological impact through $ER\alpha$, fitting to the overall impression that E2-treatment via sympathetic nervous system-ERa signalling elicits more pronounced alterations in energy homoeostasis. Nevertheless, an ER β -agonist (β -LGND2) conferred anti-obesity effects in male HFD-fed mice⁽¹⁵¹⁾. Pharmacological ER β activation (30 mg/kg BW daily, s.c.) elevated EE and acute cold tolerance by 14%. The thermogenic β-LGND2mediated response culminated in a fat mass loss of 50-60% compared to control animals. These beneficial effects were dramatically blunted, although not absent, in ER β -knockout mice. At the molecular level, β-LGND2 stimulated WAT browning and enhanced the mitochondrial respiratory capacity of white adipocytes.

No human clinical trial so far included EE or BAT activation as outcome measures. Most reports⁽¹⁵²⁻¹⁵⁸⁾ (supplementary material) focus on anthropometric measures and body composition as surrogates for alterations in energy homoeostasis. The interpretation of human trials investigating the efficacy of isoflavone mixtures or individual compounds is complex as interactive factors such as sex, pre-/postmenopausal state, habitual isoflavone exposure/ethnicity and interindividual variation in metabolising enzymes (hepatic or intestinal, gut microflora) need to be taken into account. Within their food matrix, isoflavones exist usually as glycosides (genistin and daidzin)⁽¹⁵⁹⁾. However, they are rapidly hydrolysed to their aglycones after ingestion and undergo intestinal and hepatic metabolisation generating distinct metabolites, among others equol⁽¹⁵⁹⁾. Equol is built from daidzein by intestinal bacteria and is more bioactive than its precursor⁽¹⁶⁰⁾. Surprisingly, all experimental animals are able to generate equal but not every human subject is an equal producer⁽¹⁶¹⁾. Subject to differences in the gut microbiome, only 20-35% of Western individuals are equol-producers, whereas in the Asian region the rate is $50-60 \%^{(162-164)}$. There is also a sex and age bias towards postmenopausal women in the published trials. In this target group, dietary isoflavones might simply aid in minimising or compensating the effects of declining endogenous oestrogen levels and mitigate

weight postmenopausal gain. Presently, two meta-analyses dissect in depth the efficacy of isoflavones on body composition and weight development. These studies provide a clearer view on the effects in different subpopulations or influencing factors such as dosing, weight status or trial duration. The meta-analysis from Zhang et al.⁽¹⁶⁵⁾ concentrates specifically on postmenopausal non-Asian women and included nine trials for BW⁽¹⁶⁵⁾. Sova isoflavone supplementation was associated with a significant lower BW and subgroup analyses revealed that the difference is more pronounced in normal weight than obese women as well as with lower doses (<100 mg daily)⁽¹⁶⁵⁾. A second analysis performed by Akhlaghi *et al.*⁽¹⁶⁶⁾ includes both sexes, all ethnicities independent of age and comprised forty-seven trials with sova and seventeen trials with isoflavones as investigational subjects⁽¹⁶⁶⁾. BMI in response to isoflavone intake tended to be lower (P = 0.085), especially with lower doses and shorter trial length. The subgroup analysis suggests that postmenopausal and Caucasian women likely benefit more from isoflavone intake. Contrarily, soya intake does not affect BW, fat mass or waist circumference. The latter two are also not modified by isoflavone supplementation⁽¹⁶⁶⁾. Taken together, it seems that isoflavones have a mild beneficial effect on BW development and seem to be particularly beneficial in postmenopausal women to account for the reduced endogenous oestrogen levels. Although lower doses below 100 mg daily are more effective, they still exceed the amounts achievable through normal dietary sources, even in populations with high daily isoflavone intake $(25-50 \text{ mg})^{(167)}$. Thus, isoflavone fortification or supplements would be required to fulfil a physiological relevant intake.

Alkaloids

Alkaloids exert a plethora of pharmacological activities crucial for human pharmacotherapy (e.g. morphine, quinine and vinblastine)⁽¹⁶⁸⁾ and the alkaloids capsaicin (CAP) and berberine (BBR) have been shown to affect BAT activity or induce browning of WAT.

Capsaicinoids, especially CAP, confer the pungent sensation of red chilli from the *Capsicum* genus^(169,170). The thermogenic properties of CAP-rich food are reflected in a variety of physiological responses such as elevated body temperature, flushing, vasodilation and the concomitant onset of cooling mechanisms such as sweating. CAP stimulates afferent sensory neurons involved in thermo- and nociception by binding to TRPV1, which increases intracellular Ca^{2+} concentrations that trigger membrane depolarisation⁽¹⁷¹⁾. Many groups have examined the effect of acute or chronic capsaicinoids/capsinoids on EE, fat oxidation, weight management and the contribution of thermogenically active BAT, with diverging outcomes. Early studies investigating the influence of spicy foods using chilli sauce (3 g) or red pepper (30 mg CAP) on metabolic rates in men reported significant increases in postprandial EE by 25-32% compared to control meals^(172,173). This metabolic effect was abolished when the participants were pre-treated with the β 3-AR

antagonist propranolol⁽¹⁷³⁾, indicating the involvement of sympathetic nerve activity. Higher sympathetic:parasympathetic nervous system activity was confirmed in Caucasian men after the ingestion of a red pepper-spiked (6 g) appetiser compared to a control snack (1^{74}) . In contrast, Smeets *et al.* (1^{75}) did not find elevated diet-induced thermogenesis after a CAP-containing lunch (approximately 5.15 mg) nor altered substrate oxidation in men and women⁽¹⁷⁵⁾. However, the effect size might be too</sup> small, given a 6-fold lower CAP dose. Another placebocontrolled trial in men failed to identify a significant difference in the RMR of healthy subjects treated acutely with increasing amounts of dihydrocapsiate (0, 3 or 9 mg)⁽¹⁷⁶⁾. In vitro, CAP agonises TRPV1 at an effective dose (EC₅₀) of 50–100 nm. Contrarily, the EC₅₀ of capsiate and dihydrocapsiate are 580 and 670 nm, respectively $^{(177,178)}$. This discrepancy in potency between CAP and capsinoids combined with varying doses might resolve the absence of an effect in the dihydrocapsiate trial. A meta-analysis⁽¹⁷⁹⁾ covering the efficacy of capsiate and CAP intake on EE in human subjects reveals that capsiate enhances lipid utilisation and EE while CAP is only effective on fat oxidation. The effects of CAP on EE was only relevant in high dose trials (135-150 mg CAP), when CAP intake was stratified for $dosage^{(179)}$ For both substances, sympathetic nervous activity was stimulated where this outcome was recorded⁽¹⁷⁹⁾. Orally administrated CAP and capsiate (10 mg/kg BW) equally boosted oxygen consumption and serum catecholamine levels in mice (unknown sex) when compared to untreated animals⁽¹⁸⁰⁾. Kawabata *et al.*⁽¹⁸¹⁾ identified that the acute</sup>intragastric administration of capsinoids or CAP (10 mg/ kg BW, each) to male mice increased metabolic rate and stimulated fat oxidation in wild type, but not TRPV1 knockout mice, demonstrating the necessity of the vanilloid receptor as thermogenic mediator⁽¹⁸¹⁾. Adrenergic input is a potent activator of the thermogenic BAT programme as well as subcutaneous WAT browning and TRPV1 is expressed in both adipose types. Additionally, murine TRPV1 levels are upregulated in both BAT and WAT following a CAP-enriched diet^(182–184). Therefore, it was soon hypothesised that enforced BAT functionality or a bigger brown/brite adipocyte pool contributes to the physiologic effects of CAP. A direct effect of CAP on BAT activity was already implied in 1988, when Yoshida *et al.*⁽¹⁸⁵⁾ demonstrated that the intramuscular injection of 3 mg/kg CAP to rats elevated intrascapular BAT temperature locally and increased mitochondrial oxygen consumption⁽¹⁸⁵⁾. Similarly, 3 h intragastric CAP or capsinoid administration by Kawabata et al.⁽¹⁸¹⁾ (see earlier) heated up core and local BAT temperatures only in wild-type but not TRPV1 null mice⁽¹⁸¹⁾. A role of BAT in capsiate action is further supported by an acute capsiate exposure for 30 min that upregulated UCP1 mRNA levels in the BAT and evolved to higher UCP1 protein abundance in the BAT from male mice after 2 weeks of capsiate supplementation⁽¹⁸⁶⁾. A detailed study by Baskaran *et al.*⁽¹⁸²⁾ substantiates the interplay between \overrightarrow{CAP} , active BAT and TRPV1 in male mice⁽¹⁸²⁾. The addition of CAP to HFD (0.01 % w/w) over a 32 week intervention period prevented weight gain by almost 35%

compared to HFD controls and led to a final BW comparable to mice on standard diet. This impressive BW phenotype was not observed in TRPV1-knockout animals with the same diet regimen. Other rodent studies with males addressing obesity prevention by the supplementation of HFD $(0.014\%, rat)^{(187)}$ or standard diet $(10 \text{ mg/kg}, rat)^{(187)}$ (180) with CAP or capsiate up to 14 d equally reported lowered weight gain and stimulated fat loss from WAT compared to controls^(180,187). Regarding obesity treatment, CAP feeding attenuated additional weight gain in already obese male mice during a 10 week follow-up period (36.5 v. 41 g), resulting in reduced adiposity⁽¹⁸⁴⁾. Furthermore, CAP feeding lowered circulating TAG levels, which put forward the idea that CAP mobilises lipids from adipose depots and enhances lipid oxida-tion, likely in the BAT^(180,187). Apart from elevated $EE^{(180,182,183)}$, the RER of CAP-HFD male mice was significantly higher^(182,183) and points towards increased carbohydrate oxidation. Others found enhanced $fat^{(181,186)}$ and reduced carbohydrate⁽¹⁸¹⁾ oxidation after CAP-administration, which underlines a higher metabolic flexibility in these animals. Indications for metabolically more activated CAP-BAT were further evident in the heightened glycerol release from this tissue as surrogate for lipolysis in comparison with HFD-BAT⁽¹⁸²⁾. The lipolytic principle of CAP is very likely mediated by augmented catecholamine secretion and signalling through the β -AR-PKA-hormone-sensitive lipase pathway. Acute stimulation of brown adipocytes from male CAP-HFD mice with 1 µM CAP dramatically increased intracellular calcium influx, which was blunted in TRPV1-lacking adipocytes⁽¹⁸²⁾. A downstream analysis of the molecular events revealed that CAP activates TRPV1 leading to higher calcium levels and the activation of CaMKII. which in turn phosphorylates and activates AMPK. AMPK stimulates SIRT1 activity triggering the deacetylation of PGC1 α and PPAR γ as ultimate enhancers of BAT functionality. Baskaran and coworkers⁽¹⁸³⁾ demonstrated the browning of the inguinal WAT in response CAP diet, based on the same molecular to mechanism⁽¹⁸³⁾

Human clinical trials evaluating the long-term effect of CAP/capsiate on weight development are rare (supplementary material). A study by Janssens et al.⁽¹⁸⁸⁾ implies a role of CAP supplementation (2.56 mg/meal) in weight maintenance after weight loss in male and female subjects. Accordingly, the acute thermogenic effect of CAP could be sufficient to compensate for the reduction in EE that goes along with weight loss and energetic restriction⁽¹⁸⁸⁾. In a US placebo controlled-trial with obese male and female participants (mean BMI 30 kg/m^2) the daily ingestion of 6 mg capsinoids over a 12 weeks trended in a BW change (0.9 v. 0.5 kg) and significantly reduced visceral adiposity⁽¹⁸⁹⁾. However, more of these long-term studies with higher doses or in relation to weight stabilisation after weight loss are required to delineate the pharmacological potential of capsinoids. A recent crossover study directly captured BAT-activity during an acute capsinoid (12 mg) trigger in men and women by luorodeoxyglucose positron emission tomography-magnetic resonance in comparison with

cold-exposure using a cold vest (14.5°C). The mean standardised uptake value (SUV) for the capsinoid group did not reach the accepted threshold (SUV = 2) to classify a subject as BAT-positive, contrary to the cold-exposed group (mean SUV = 2.9). Only after an adjustment of the grey scale values, a mild uptake of glucose was notable in the capsinoid group. Nevertheless, the mean EE post-capsinoids administration was higher compared to baseline levels and this increase was significantly greater in subjects allocated as BAT-positive based on cold-exposed BAT detectability⁽¹⁹⁰⁾. Unfortunately, this study lacked a placebo control although the capsinoids were ingested as capsule containing additionally a mixture of rapeseed oil and medium chain TAG. A placebocontrolled, crossover trial with eighteen Japanese men verified an acute increase in EE after capsinoids intake (9 mg) in BAT-positive subjects when compared to placebo controls and BAT-negative individuals⁽¹⁹¹⁾.

In conclusion, it appears that the physiological outcomes associated with spicy food/CAP ingestion are the sum of four primary events: (1) enhanced sympathetic activity due to the stimulation of vagal afferent neurons and catecholamine levels, which (2) promote lipolysis in WAT and BAT and (3) activate non-thermogenesis in BAT; (4) the direct, cell autonomous trigger on adipocytes via TRPV1-AMPK-SIRT1 to enhance the thermogenic phenotype. The documented bioavailability (50-90% absorption) of capsaicinoids is high⁽¹⁸¹⁾. In mice, circulating plasma CAP concentration 1 h after an oral dose of 10 mg/kg BW reaches approximately $3.8 \,\mu M^{(192)}$. This value falls into a concentration range that induced browning in vitro in 3T3-L1 and indicates that circulating CAP levels are sufficient to elicit direct cellular effects⁽¹⁹³⁾.

BBR is arguably the most prominent known phytochemical capable of modulating BAT function and several preclinical studies have delivered interesting outcomes regarding the therapeutic impact of BBR on obesity-associated metabolic diseases. BBR has a long history of use in traditional medicines and interest in the metabolic side of BBR awoke only in 2004 when the blood cholesterol-lowering activity of BBR in hypercholesterolemic patients was published⁽¹⁹⁴⁾. Another study underpinned the therapeutic potential of BBR in genetically and fat-induced obese rodents⁽¹⁹⁵⁾. In male db/db mice a daily dose of BBR (5 mg/kg BW, i.p.) caused a 13% weight loss with a 10% reduction in fat mass within 26 d of treatment alongside with improved whole body metabolic homoeostasis⁽¹⁹⁵⁾. The same physiological effects were observed in male HFD-fed rats treated with either vehicle or BBR (380 mg/kg BW, orally)⁽¹⁹⁵⁾. In WAT, BBR downregulated lipogenic genes while oxidative and genes regulating mitochondria formation/function were enhanced in muscle and BAT (PPAR α and PGC1 α)⁽¹⁹⁵⁾. In 3T3-L1 cells, they subsequently identify that BBR acts as an AMPK activator, stimulating the phosphorylation of AMPK and acetyl-CoA carboxylase⁽¹⁹⁵⁾, possibly by inhibiting complex I causing a rise in the cellular AMP:ATP ratio⁽¹⁹⁶⁾. These findings motivated further research exploiting the effects of BBR on BAT activity as

AMPK plays an accepted role in BAT-mediated thermogenesis and EE⁽¹⁹⁷⁻¹⁹⁵⁾. Zhang et al.⁽²⁰⁰⁾ first reported a thermogenic effect by $BBR^{(200)}$. Apart from the estaband weight lished metabolic improvements, BBR-therapy to male db/db mice (5 mg/kg BW, i.p.) increased rectal temperature (about 1.5°C), consistently enhanced metabolic rate and defended body heat loss during cold exposure compared to vehicle treatment. Most strikingly, BBR dramatically enforced BAT thermogenesis directly as reflected by higher 18-FDG uptake monitored using micro-positron emission tomographymagnetic resonance. This activated BAT state was supported by reduced BAT mass, higher mitochondrial content and molecular evidence such as enhancement of the BAT-specific mRNA and protein signatures (PGC1a, UCP1. nuclear respiratory factor 1 and carnitine palmitoyltransferase) through activation of AMPK⁽²⁰⁰⁾. BBR likewise stimulated browning of the inguinal WAT. Interestingly, immunohistological analysis revealed higher tyrosine hydroxylase levels in the WAT and BAT, suggesting the contribution of augmented sympathetic output to the observed phenotype. When the animals were housed under thermoneutral conditions (30°C) to minimise BAT function, the beneficial effects of BBR administration were blunted, which strongly supports the necessity of active sympathetic activity as physiologic mediator of BBR activity⁽²⁰⁰⁾. WAT-browning and elevated EE were also noted in male mice on HFD treated for 5 weeks with BBR (5 mg/kg BW i.p.) or vehicle⁽²⁰¹⁾. Additionally, BBR upregulated hepatic FGF21 gene expression resulting in a concomitant elevation in plasma FGF21 levels, which was dependent on activated SIRT1, a known regulator of FGF21 transcription⁽²⁰²⁾. FGF21 was shown to promote WAT browning as well as the thermogenic BAT programme elsewhere and might depict an alternative mechanism of action of BBR^(203,204). Wu et al.⁽²⁰⁵⁾ clearly show the requirement of intact AMPK signalling in adipose tissues for an effective BBR treatment as the physiological responses to BBR administration were absent in adiponectin-Cre driven AMPK-floxed male $mice^{(205)}$.

In the same work $^{(205)}$, the effect of 1 month dietary BBR supplementation $(3 \times 500 \text{ mg daily})$ on human BAT activity was examined in overweight individuals. Using cold-stimulated fluorodeoxyglucose positron emission tomography-computed tomography imaging, they prove that human BAT is activated by BBR as mean SUV (2.6–3.3), BAT mass $(14.1-25.5 \text{ cm}^3)$ and activity $(103 \cdot 1 - 228 \cdot 2 \text{ ml} \times \text{SUV}_{ave} \times \text{g/ml})$ were all significantly increased in a paired before-after analysis. Patients (n 2) with no detectable BAT before treatment were not BAT-positive after 1 month of BBR intake. Thus, BBR is not sufficient to stimulate BAT formation or induce dormant brown adipocytes. The percentage change of BAT activity before-after BBR negatively correlated with the measured changes in BW and highlight the potency of BBR to combat overweight by means of BAT activation. A future pharmacological application of BBR to tackle obesity and associated metabolic complications is supported by additional human clinical trials. An open-label study in China investigated the

effect of a lifestyle intervention (LSI) with or without daily BBR $(3 \times 500 \text{ mg})$ or pioglitazone (15 mg) treatment on BW and metabolic health in individuals suffering from non-alcoholic fatty liver disease with diabetes or impaired glucose tolerance⁽²⁰⁶⁾. At the end of the intervention, the BBR group lost significantly more weight (-4.29 v. -1.44 kg) and waist circumference (-4.84 v. -2.14 cm) than LSI only, but also when compared to LSI plus pioglitazone. As there was no significant difference between LSI + pioglitazone and LSI + BBR on insulin sensitivity, glucose tolerance or glycated Hb levels, BBR improved glucose homoeostasis comparably to a known antidiabetic remedy with the additive effect of weight loss⁽²⁰⁶⁾. BBR was equally effective in placebocontrolled trials with Chinese type 2 diabetic and dyslipi-daemic patients (1 g daily)⁽²⁰⁷⁾ or poor glycaemic control $(1.5 \text{ g daily})^{(208)}$. The BBR group displayed significant improvements in BW (66.4 v. 70.5 kg), BMI (24.3 v. 25.4 kg/m²) and glycaemic control (fasting blood glucose 5.6 v. 6.4 mm/, HbA1c 6.6 v. 7.3 %) compared to controls at the end⁽²⁰⁷⁾. In a small pilot study (n 7) with obese Caucasian individuals over 13 weeks (1500 mg BBR daily) moderate changes in BW (2.3%), BMI (2.9) and fat content (3.6%) were determined compared to baselines values. Although these changes seem promising, they did not reach statistical significance likely due to the small sample size⁽²⁰⁹⁾. The available studies centring on BW control or weight loss are sparse and dominated bv Asian subjects (supplementary material). Nevertheless, the reported outcomes are overly encouraging and underline the need for further efforts aiming at different ethnicities and larger sample sizes. Furthermore, it would be interesting to study whether BBR enhances EE in human subjects. One of the current drawbacks is the extremely poor bioavailability of BBR in human and animal studies. In human subjects, oral BBR intake (300 mg daily) resulted in a mean plasma BBR concentration of 0.3 ng/ml and an acute BBR (400 mg) challenge led to a maximal concentration of $0.4 \text{ ng/ml}^{(210)}$. BBR combined with polyethylene glycol enhanced intestinal BBR absorption in rats by a factor of 3, indicating that the pharmacokinetic properties of BBR are adjustable with appropriate encapsulation strategies⁽²¹¹⁾. Improved BBR bioavailability is desirable as it affects the effective treatment dose and might reduce some of the mild side effects (e.g. constipation and intestinal discomfort) observed with the currently employed BBR dose of 1500 mg daily. Alternatively, the efficacy of the more absorbable analogue dihydroberberine with five times higher bioavailability than BBR in rats could be investigated for human $use^{(212)}$.

Conclusions

BAT is a remarkable organ with the capacity to ignite its thermogenic programme to 'waste' energies in response to environmental or endogenous cues. From the aforementioned reviewed phytochemicals we can draw several conclusions. First, preclinical evidence of efficacy from animal studies is limited in human trials and the reported NS Proceedings of the Nutrition Society

effect sizes in human subjects tend to be small. Differences in the gut microbiome and metabolising enzymes influence whether, how and to what extent a phytochemical is altered from its prodrug state to either a functionally more or less active metabolite that reaches the systemic circulation and target sites. Animal experiments are performed in inbred strains with near identical genetic background whereas in human subjects the interindividual variability is more pronounced. This is intensified by differential study designs (duration, dose, ethnicity, formulation, etc.) that might cause the large variability between human studies. Secondly, sex as biological variable adds extra complexity to the interpretation of results, which demands critical attention. Out of the thirty-nine human randomised controlled trials listed in the supplementary material, fifteen included male and female subjects, thirteen only males and nine only females. Females were notably overrepresented in trials with isoflavones due to their oestrogenic potential. No studies with 'females only' are listed for PTS, BBR and capsaicinoids and only one for catechins. The vast majority of the cited rodent studies were performed in male animals due to prevailing belief that females' hormonal cycle increase heterogeneity and results derived from male animals are translatable to females⁽²¹³⁾. In contrast, the female gonadal WAT is more innervated by sympathetic nerves and responds more effectively (browning) to a β 3-AR agonist compared to male gonadal WAT⁽²⁴⁾. Similarly, the BAT of female rats displays a more multilocular morphology, a denser mitochondrial network with higher UCP1 protein content and reduced inhibitory a2-AR expression compared to males⁽²¹⁴⁾. These sex-specific BAT differences convey an enhanced thermogenic potential and increased epinephrine-sensitivity to female animals. Consequently, it is crucial to include both sexes in studies aiming at modulators of non-shivering thermogenesis. In addition, various hormonal cues such as sex hormones (see phytoestrogens) or leptin affect BAT-mediated $EE^{(215,216)}$. Surprisingly, circulating leptin levels are higher in male v. female rats. while this ratio is reciprocal in human subjects^(217,218). These examples substantiate two statements. First, energy homoeostasis is inherently modulated by sexual dimorphism and secondly, the underlying regulatory system is inconsistent between males and females across species. Sex as biological variable further affects the pharmacokinetics and metabolism of phytochemicals, especially with regards to their metabolisation by the gut microflora or liver^(219,220). As such, a key hepatic cytochrome P540 enzyme (CYP3A4) is elevated in females over males⁽²²⁰⁾ and CYP3A4 is predominantly metabolising BBR⁽²²¹⁾, which could lead to sex-specific differences in the clearance of BBR. In the presented trials, the circulating metabolites are rarely determined and might not overlap between men and women causing differential study outcomes.

Finally, selected compounds clearly have a beneficial outcome in clinical trials and are accepted by meta-analyses. Regular catechin intake (300 mg) for example can result in about 418 kJ increase in EE. Spicy food hampers macronutrient intake by about 565

kJ and/or stabilises weight regain by compensating for the decrease in EE. This small contribution in itself might be sufficient to balance the energy gap to zero. Although these thermogenic effects are presently not sufficient to actively stimulate weight loss, they could be essential in preventing further fat accumulation and potentially stop the transition from overweight to obese states. As such, prevention of weight gain and weight maintenance are more easily achievable than a decrease in fat mass, as conserved mechanisms defend body weight rather than promote fat release⁽²²²⁾. In conclusion, dietary phytochemicals in their food matrix or as a supplement alone play an important role in obesity prevention by activating BAT functionality, targeting a broad population base.

Supplementary material

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References

- 1. Wang Y, Beydoun MA, Liang L et al. (2008) Will all Americans become overweight or obese? Estimating the progression and cost of the US obesity epidemic. *Obesity (Silver Spring, MD)* **16**, 2323–2330.
- NCD Risk Factor Collaboration (NCD-RisC) (2017) Worldwide trends in body-mass index, underweight, overweight, and obesity from 1975 to 2016: a pooled analysis of 2416 population-based measurement studies in 128-9 million children, adolescents, and adults. *Lancet (Lond, Engl)* 390, 2627–2642.
- 3. Cypess AM, Lehman S, Williams G et al. (2009) Identification and importance of brown adipose tissue in adult humans. N Engl J Med **360**, 1509–1517.
- van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM et al. (2009) Cold-activated brown adipose tissue in healthy men. N Engl J Med 360, 1500–1508.
- 5. Lean MEJ, James WPT, Jennings G *et al.* (1986) Brown adipose tissue uncoupling protein content in human infants, children and adults. *Clin Sci* **71**, 291–297.
- 6. Nedergaard J, Golozoubova V, Matthias A *et al.* (2001) UCP1: the only protein able to mediate adaptive nonshivering thermogenesis and metabolic inefficiency. *Biochim Biophys Acta* **1504**, 82–106.

- 7. Chondronikola M, Volpi E, Børsheim E *et al.* (2014) Brown adipose tissue improves whole-body glucose homeostasis and insulin sensitivity in humans. *Diabetes* **63**, 4089–4099.
- Bartelt A, Bruns OT, Reimer R *et al.* (2011) Brown adipose tissue activity controls triglyceride clearance. *Nat Med* 17, 200–205.
- Mercer SW & Trayhurn P (1983) Developmental changes in fatty acid synthesis in interscapular brown adipose tissue of lean and genetically obese (oblob) mice. *Biochem* J 212, 393–398.
- Whittle A, Relat-Pardo J & Vidal-Puig A (2013) Pharmacological strategies for targeting BAT thermogenesis. *Trends Pharmacol Sci* 34, 347–355.
- 11. Beiroa D, Imbernon M, Gallego R *et al.* (2014) GLP-1 agonism stimulates brown adipose tissue thermogenesis and browning through hypothalamic AMPK. *Diabetes* **63**, 3346–3358.
- Bordicchia M, Liu D, Amri E-Z et al. (2012) Cardiac natriuretic peptides act via p38 MAPK to induce the brown fat thermogenic program in mouse and human adipocytes. J Clin Invest 122, 1022–1036.
- Young P, Arch JR & Ashwell M (1984) Brown adipose tissue in the parametrial fat pad of the mouse. *FEBS Lett* 167, 10–14.
- Ohno H, Shinoda K, Spiegelman BM et al. (2012) PPARγ agonists induce a white-to-brown fat conversion through stabilization of PRDM16 protein. Cell Metab 15, 395–404.
- Boozer CN, Nasser JA, Heymsfield SB *et al.* (2001) An herbal supplement containing Ma Huang-Guarana for weight loss: a randomized, double-blind trial. *Int J Obes* 25, 316–324.
- Hill JO, Peters JC & Wyatt HR (2009) Using the energy gap to address obesity: a commentary. J Am Diet Assoc 109, 1848–1853.
- Carpentier AC, Blondin DP, Virtanen KA *et al.* (2018) Brown adipose tissue energy metabolism in humans. *Front Endocrinol* 9 [Epublication 7 August 2018].
- Cypess AM, Weiner LS, Roberts-Toler C *et al.* (2015) Activation of human brown adipose tissue by a β3-adrenergic receptor agonist. *Cell Metab* 21, 33–38.
- O'Mara AE, Johnson JW, Linderman JD *et al.* (2020) Chronic mirabegron treatment increases human brown fat, HDL cholesterol, and insulin sensitivity. *J Clin Invest* [Epublication ahead of print version].
- Pfannenberg C, Werner MK, Ripkens S et al. (2010) Impact of age on the relationships of brown adipose tissue with sex and adiposity in humans. *Diabetes* 59, 1789–1793.
- Lee P, Zhao JT, Swarbrick MM *et al.* (2011) High prevalence of brown adipose tissue in adult humans. *J Clin Endocrinol Metab* 96, 2450–2455.
- 22. Vijgen GHEJ, Bouvy ND, Teule GJJ et al. (2011) Brown adipose tissue in morbidly obese subjects. *PLoS ONE* 6, e17247.
- Valle A, García-Palmer FJ, Oliver J et al. (2007) Sex differences in brown adipose tissue thermogenic features during caloric restriction. *Cell Physiol Biochem* 19, 195–204.
- Kim S-N, Jung Y-S, Kwon H-J et al. (2016) Sex differences in sympathetic innervation and browning of white adipose tissue of mice. *Biol Sex Differ* 7 [Epublication 9 December 2016].
- Quarta C, Mazza R, Pasquali R *et al.* (2012) Role of sex hormones in modulation of brown adipose tissue activity. *J Mol Endocrinol* 49, R1–R7.
- 26. Fletcher LA, Kim K, Leitner BP et al. (2020) Sexual dimorphisms in adult human brown adipose tissue. Obesity (Silver Spring, MD) 28, 241–246.

- 27. Rimando AM, Nagmani R, Feller DR *et al.* (2005) Pterostilbene, a new agonist for the peroxisome proliferator-activated receptor alpha-isoform, lowers plasma lipoproteins and cholesterol in hypercholesterolemic hamsters. *J Agric Food Chem* **53**, 3403–3407.
- 28. Kersten S (2014) Integrated physiology and systems biology of PPARα. *Mol Metab* **3**, 354–371.
- Rakhshandehroo M, Knoch B, Müller M et al. (2010) Peroxisome proliferator-activated receptor alpha target genes. PPAR Res [Epublication 26 September 2010].
- Etxeberria U, Hijona E, Aguirre L et al. (2017) Pterostilbene-induced changes in gut microbiota composition in relation to obesity. *Mol Nutr Food Res* 61 [Epublication 3 August 2016].
- Gómez-Zorita S, Fernández-Quintela A, Lasa A et al. (2014) Pterostilbene, a dimethyl ether derivative of resveratrol, reduces fat accumulation in rats fed an obesogenic diet. J Agric Food Chem 62, 8371–8378.
- 32. Riche DM, Riche KD, Blackshear CT et al. (2014) Pterostilbene on metabolic parameters: a randomized, double-blind, and placebo-controlled trial. Evidence-Based Complementary Altern Med **2014**, 459165.
- 33. Barbera MJ, Schluter A, Pedraza N et al. (2001) Peroxisome proliferator-activated receptor alpha activates transcription of the brown fat uncoupling protein-1 gene. A link between regulation of the thermogenic and lipid oxidation pathways in the brown fat cell. J Biol Chem 276, 1486–1493.
- 34. Mottillo EP, Bloch AE, Leff T *et al.* (2012) Lipolytic products activate peroxisome proliferator-activated receptor (PPAR) α and δ in brown adipocytes to match fatty acid oxidation with supply. *J Biol Chem* **287**, 25038–25048.
- 35. Aguirre L, Milton-Laskibar I, Hijona E *et al.* (2016) Effects of pterostilbene in brown adipose tissue from obese rats. *J Physiol Biochem* **73**, 457–464.
- Nagao K, Jinnouchi T, Kai S *et al.* (2017) Pterostilbene, a dimethylated analog of resveratrol, promotes energy metabolism in obese rats. *J Nutr Biochem* 43, 151–155.
- Seale P, Conroe HM, Estall J et al. (2011) Prdm16 determines the thermogenic program of subcutaneous white adipose tissue in mice. J Clin Invest 121, 96–105.
- Qiang L, Wang L, Kon N *et al.* (2012) Brown remodeling of white adipose tissue by SirT1-dependent deacetylation of PPARγ. *Cell* 150, 620–632.
- Picard F, Kurtev M, Chung N *et al.* (2004) Sirt1 promotes fat mobilization in white adipocytes by repressing PPAR-gamma. *Nature* 429, 771–776.
- Lan F, Cacicedo JM, Ruderman N et al. (2008) SIRT1 Modulation of the acetylation status, cytosolic localization, and activity of LKB1. Possible role in AMP-activated protein kinase activation. J Biol Chem 283, 27628–27635.
- Rodgers JT, Lerin C, Haas W et al. (2005) Nutrient control of glucose homeostasis through a complex of PGC-1alpha and SIRT1. *Nature* 434, 113–118.
- 42. La Spina M, Galletta E, Azzolini M et al. (2019) Browning effects of a chronic pterostilbene supplementation in mice fed a high-fat diet. Int J Mol Sci 20 [Epublication 29 October 2019].
- 43. Defour M, Dijk W, Ruppert P *et al.* (2018) The peroxisome proliferator-activated receptor α is dispensable for cold-induced adipose tissue browning in mice. *Mol Metab* **10**, 39–54.
- 44. Kapetanovic IM, Muzzio M, Huang Z et al. (2011) Pharmacokinetics, oral bioavailability, and metabolic profile of resveratrol and its dimethylether analog,

pterostilbene, in rats. *Cancer Chemother Pharmacol* 68, 593–601.

- Nair AB & Jacob S (2016) A simple practice guide for dose conversion between animals and human. J Basic Clin Pharm 7, 27–31.
- Riche DM, McEwen CL, Riche KD *et al.* (2013) Analysis of safety from a human clinical trial with pterostilbene. J *Toxicol* 2013, 463595.
- Howitz KT, Bitterman KJ, Cohen HY *et al.* (2003) Small molecule activators of sirtuins extend saccharomyces cerevisiae lifespan. *Nature* 425, 191–196.
- Bordone L & Guarente L (2005) Calorie restriction, SIRT1 and metabolism: understanding longevity. *Nat Rev Mol Cell Biol* 6, 298–305.
- 49. Lagouge M, Argmann C, Gerhart-Hines Z *et al.* (2006) Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1alpha. *Cell* **127**, 1109–1122.
- 50. Baur JA, Pearson KJ, Price NL *et al.* (2006) Resveratrol improves health and survival of mice on a high-calorie diet. *Nature* 444, 337–342.
- 51. Dal-Pan A, Blanc S & Aujard F (2010) Resveratrol suppresses body mass gain in a seasonal non-human primate model of obesity. *BMC Physiol* **10**, 11.
- 52. Andrade JMO, Frade ACM, Guimarães JB *et al.* (2014) Resveratrol increases brown adipose tissue thermogenesis markers by increasing SIRT1 and energy expenditure and decreasing fat accumulation in adipose tissue of mice fed a standard diet. *Eur J Nutr* **53**, 1503–1510.
- 53. Wang S, Liang X, Yang Q *et al.* (2015) Resveratrol induces brown-like adipocyte formation in white fat through activation of AMP-activated protein kinase (AMPK) α1. *Int J Obes* 2005 **39**, 967–976.
- 54. Wang S, Liang X, Yang Q *et al.* (2017) Resveratrol enhances brown adipocyte formation and function by activating AMP-activated protein kinase (AMPK) αl in mice fed high-fat diet. *Mol Nutr Food Res* **61** [Epublication].
- 55. Alberdi G, Rodríguez VM, Miranda J *et al.* (2013) Thermogenesis is involved in the body-fat lowering effects of resveratrol in rats. *Food Chem* **141**, 1530–1535.
- Poulsen MM, Vestergaard PF, Clasen BF et al. (2013) High-dose resveratrol supplementation in obese men. Diabetes 62, 1186–1195.
- Yoshino J, Conte C, Fontana L *et al.* (2012) Resveratrol supplementation does not improve metabolic function in nonobese women with normal glucose tolerance. *Cell Metab* 16, 658–664.
- Crandall JP, Oram V, Trandafirescu G et al. (2012) Pilot study of resveratrol in older adults with impaired glucose tolerance. J Gerontol A: Biol Sci Med Sci 67, 1307–1312.
- 59. Timmers S, Konings E, Bilet L *et al.* (2011) Calorie restriction-like effects of 30 days of resveratrol (resVida[™]) supplementation on energy metabolism and metabolic profile in obese humans. *Cell Metab* **14**, 612–622.
- 60. Springer M & Moco S (2019) Resveratrol and its human metabolites-effects on metabolic health and obesity. *Nutrients* **11** [Epublication].
- 61. Maier-Salamon A, Böhmdorfer M, Thalhammer T *et al.* (2011) Hepatic glucuronidation of resveratrol: interspecies comparison of enzyme kinetic profiles in human, mouse, rat, and dog. *Drug Metab Pharmacokinet* **26**, 364–373.
- 62. Kim SJ & Kim GH (2006) Quantification of quercetin in different parts of onion and its DPPH radical scavenging and antibacterial activity. *Food Sci Biotechnol* 15, 39–43.

- 63. Moon J, Do H-J, Kim OY *et al.* (2013) Antiobesity effects of quercetin-rich onion peel extract on the differentiation of 3T3-L1 preadipocytes and the adipogenesis in high fat-fed rats. *Food Chem Toxicol* **58**, 347–354.
- 64. Ting Y, Chang W-T, Shiau D-K et al. (2018) Antiobesity efficacy of quercetin-rich supplement on diet-induced obese rats: effects on body composition, serum lipid profile, and gene expression. J Agric Food Chem 66, 70–80.
- Rivera L, Morón R, Sánchez M *et al.* (2008) Quercetin ameliorates metabolic syndrome and improves the inflammatory status in obese Zucker rats. *Obesity (Silver Spring, MD)* 16, 2081–2087.
- Lee SG, Parks JS & Kang HW (2017) Quercetin, a functional compound of onion peel, remodels white adipocytes to brown-like adipocytes. *J Nutr Biochem* 42, 62–71.
- 67. Kuipers EN, van Dam AD, Held NM *et al.* (2018) Quercetin lowers plasma triglycerides accompanied by white adipose tissue browning in diet-induced obese mice. *Int J Mol Sci* **19** [Epublication 16 Jun 2018].
- Stewart LK, Soileau JL, Ribnicky D et al. (2008) Quercetin transiently increases energy expenditure but persistently decreases circulating markers of inflammation in C57BL/6J mice fed a high-fat diet. *Metabolism* 57, S39– S46.
- Choi H, Kim C-S & Yu R (2018) Quercetin upregulates uncoupling protein 1 in white/brown adipose tissues through sympathetic stimulation. J Obes Metab Syndr 27, 102–109.
- Gruse J, Görs S, Tuchscherer A *et al.* (2015) The effects of oral quercetin supplementation on splanchnic glucose metabolism in 1-week-old calves depend on diet after birth. *J Nutr* 145, 2486–2495.
- Gugler R & Dengler HJ (1973) Inhibition of human liver catechol-O-methyltransferase by flavonoids. Naunyn-Schmiedeberg's Arch Pharmacol 276, 223–233.
- Kuppusamy UR & Das NP (1994) Potentiation of beta-adrenoceptor agonist-mediated lipolysis by quercetin and fisetin in isolated rat adipocytes. *Biochem Pharmacol* 47, 521–529.
- Dong J, Zhang X, Zhang L *et al.* (2014) Quercetin reduces obesity-associated ATM infiltration and inflammation in mice: a mechanism including AMPKα1/SIRT1. *J Lipid Res* 55, 363–374.
- Cho KW & Lumeng CN (2011) Sirt1: a guardian at the gates of adipose tissue inflammation. *Diabetes* 60, 3100– 3102.
- 75. Yoshizaki T, Milne JC, Imamura T *et al.* (2009) SIRT1 exerts anti-inflammatory effects and improves insulin sensitivity in adipocytes. *Mol Cell Biol* **29**, 1363–1374.
- 76. Comalada M, Ballester I, Bailón E *et al.* (2006) Inhibition of pro-inflammatory markers in primary bone marrowderived mouse macrophages by naturally occurring flavonoids: analysis of the structure–activity relationship. *Biochem Pharmacol* 72, 1010–1021.
- 77. Huang H, Liao D, Dong Y et al. (2019) Clinical effectiveness of quercetin supplementation in the management of weight loss: a pooled analysis of randomized controlled trials. *Diabetes Metab Syndr Obes: Targets Ther* 12, 553–563.
- Kim J, Cha Y-J, Lee K-H *et al.* (2013) Effect of onion peel extract supplementation on the lipid profile and antioxidative status of healthy young women: a randomized, placebo-controlled, double-blind, crossover trial. *Nutr Res Pract* 7, 373–379.
- 79. Kim K-A & Yim J-E (2016) The effect of onion peel extract on inflammatory mediators in Korean overweight and obese women. *Clin Nutr Res* **5**, 261–269.

- Lee J-S, Cha Y-J, Lee K-H et al. (2016) Onion peel extract reduces the percentage of body fat in overweight and obese subjects: a 12-week, randomized, double-blind, placebo-controlled study. Nutr Res Pract 10, 175–181.
- Zhang L, Angst E, Park JL *et al.* (2010) Quercetin aglycone is bioavailable in murine pancreas and pancreatic xenografts. *J Agric Food Chem* 58, 7252–7257.
- 82. Egert S, Bosy-Westphal A, Seiberl J et al. (2009) Quercetin reduces systolic blood pressure and plasma oxidised low-density lipoprotein concentrations in overweight subjects with a high-cardiovascular disease risk phenotype: a double-blinded, placebo-controlled cross-over study. Br J Nutr 102, 1065–1074.
- Jin F, Nieman DC, Shanely RA *et al.* (2010) The variable plasma quercetin response to 12-week quercetin supplementation in humans. *Eur J Clin Nutr* 64, 692–697.
- 84. Lu NT, Crespi CM, Liu NM *et al.* (2016) A phase I dose escalation study demonstrates quercetin safety and explores potential for bioflavonoid antivirals in patients with chronic hepatitis C. *Phytother Res* **30**, 160–168.
- Xu N, Zhang L, Dong J et al. (2014) Low-dose diet supplement of a natural flavonoid, luteolin, ameliorates diet-induced obesity and insulin resistance in mice. *Mol Nutr Food Res* 58, 1258–1268.
- 86. Kwon E-Y, Jung UJ, Park T *et al.* (2015) Luteolin attenuates hepatic steatosis and insulin resistance through the interplay between the liver and adipose tissue in mice with diet-induced obesity. *Diabetes* **64**, 1658–1669.
- Zhang L, Han Y-J, Zhang X *et al.* (2016) Luteolin reduces obesity-associated insulin resistance in mice by activating AMPKα1 signalling in adipose tissue macrophages. *Diabetologia* 59, 2219–2228.
- Ding L, Jin D & Chen X (2010) Luteolin enhances insulin sensitivity via activation of PPARγ transcriptional activity in adipocytes. *J Nutr Biochem* 21, 941–947.
- de Souza CJ, Eckhardt M, Gagen K *et al.* (2001) Effects of pioglitazone on adipose tissue remodeling within the setting of obesity and insulin resistance. *Diabetes* 50, 1863–1871.
- 90. Zhang X, Zhang Q-X, Wang X et al. (2016) Dietary luteolin activates browning and thermogenesis in mice through an AMPK/PGC1α pathway-mediated mechanism. Int J Obes 2005 40, 1841–1849.
- Zhang X, Wang X, Yin H et al. (2019) Functional inactivation of mast cells enhances subcutaneous adipose tissue browning in mice. Cell Rep 28, 792–803. e4.
- 92. Graham HN (1992) Green tea composition, consumption, and polyphenol chemistry. *Prev Med* **21**, 334–350.
- Henning SM, Fajardo-Lira C, Lee HW et al. (2003) Catechin content of 18 teas and a green tea extract supplement correlates with the antioxidant capacity. Nutr Cancer 45, 226–235.
- 94. Arts ICW, van de Putte B & Hollman PCH (2000) Catechin contents of foods commonly consumed in The Netherlands. 2. Tea, wine, fruit juices, and chocolate milk. J Agric Food Chem 48, 1752–1757.
- 95. Dulloo AG, Duret C, Rohrer D et al. (1999) Efficacy of a green tea extract rich in catechin polyphenols and caffeine in increasing 24-h energy expenditure and fat oxidation in humans. Am J Clin Nutr 70, 1040–1045.
- 96. Rudelle S, Ferruzzi MG, Cristiani I *et al.* (2007) Effect of a thermogenic beverage on 24-hour energy metabolism in humans. *Obesity (Silver Spring, MD)* **15**, 349–355.
- Bérubé-Parent S, Pelletier C, Doré J et al. (2005) Effects of encapsulated green tea and Guarana extracts containing a mixture of epigallocatechin-3-gallate and caffeine

on 24 h energy expenditure and fat oxidation in men. Br J Nutr 94, 432–436.

- Thielecke F, Rahn G, Böhnke J et al. (2010) Epigallocatechin-3-gallate and postprandial fat oxidation in overweight/obese male volunteers: a pilot study. Eur J Clin Nutr 64, 704–713.
- Hursel R, Viechtbauer W & Westerterp-Plantenga MS (2009) The effects of green tea on weight loss and weight maintenance: a meta-analysis. *Int J Obes* 2005 33, 956–961.
- 100. Westerterp-Plantenga MS, Lejeune MPGM & Kovacs EMR (2005) Body weight loss and weight maintenance in relation to habitual caffeine intake and green tea supplementation. *Obes Res* **13**, 1195–1204.
- 101. Kapoor MP, Sugita M, Fukuzawa Y et al. (2017) Physiological effects of epigallocatechin-3-gallate (EGCG) on energy expenditure for prospective fat oxidation in humans: a systematic review and meta-analysis.. *Nutr Biochem* 43, 1–10.
- 102. Yoneshiro T, Matsushita M, Hibi M *et al.* (2017) Tea catechin and caffeine activate brown adipose tissue and increase cold-induced thermogenic capacity in humans. *Am J Clin Nutr* **105**, 873–881.
- 103. Nirengi S, Amagasa S, Homma T *et al.* (2016) Daily ingestion of catechin-rich beverage increases brown adipose tissue density and decreases extramyocellular lipids in healthy young women. *SpringerPlus* **5**, 1363.
- 104. Matsuyama T, Tanaka Y, Kamimaki I et al. (2008) Catechin safely improved higher levels of fatness, blood pressure, and cholesterol in children. Obesity (Silver Spring, MD) 16, 1338–1348.
- 105. Nagao T, Komine Y, Soga S et al. (2005) Ingestion of a tea rich in catechins leads to a reduction in body fat and malondialdehyde-modified LDL in men. Am J Clin Nutr. 81, 122–129.
- 106. Auvichayapat P, Prapochanung M, Tunkamnerdthai O et al. (2008) Effectiveness of green tea on weight reduction in obese Thais: a randomized, controlled trial. *Physiol Behav* 93, 486–491.
- 107. Chantre P & Lairon D (2002) Recent findings of green tea extract AR25 (Exolise) and its activity for the treatment of obesity. *Phytomedicine* 9, 3–8.
- 108. Phung OJ, Baker WL, Matthews LJ et al. (2010) Effect of green tea catechins with or without caffeine on anthropometric measures: a systematic review and meta-analysis. *Am J Clin Nutr* 91, 73–81.
- 109. Choo JJ (2003) Green tea reduces body fat accretion caused by high-fat diet in rats through beta-adrenoceptor activation of thermogenesis in brown adipose tissue. J Nutr Biochem 14, 671–676.
- 110. Nomura S, Ichinose T, Jinde M *et al.* (2008) Tea catechins enhance the mRNA expression of uncoupling protein 1 in rat brown adipose tissue. *J Nutr Biochem* **19**, 840–847.
- 111. Klaus S, Pültz S, Thöne-Reineke C *et al.* (2005) Epigallocatechin gallate attenuates diet-induced obesity in mice by decreasing energy absorption and increasing fat oxidation. *Int J Obes* 2005 **29**, 615–623.
- 112. Nagai M, Conney AH & Zhu BT (2004) Strong inhibitory effects of common tea catechins and bioflavonoids on the *O*-methylation of catechol estrogens catalyzed by human liver cytosolic catechol-*O*-methyltransferase. *Drug Metab Dispos* 32, 497–504.
- 113. Shixian Q, VanCrey B, Shi J et al. (2006) Green tea extract thermogenesis-induced weight loss by epigallocatechin gallate inhibition of catechol-O-methyltransferase. J Med Food 9, 451–458.
- 114. Zhu BT, Shim J-Y, Nagai M et al. (2008) Molecular modelling study of the mechanism of high-potency inhibition of

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human catechol-O-methyltransferase by (-)-epigallocatechin-3-O-gallate. *Xenobiotica* **38**, 130–146.

- 115. Warden BA, Smith LS, Beecher GR *et al.* (2001) Catechins are bioavailable in men and women drinking black tea throughout the day. *J Nutr* **131**, 1731–1737.
- 116. Takahashi M, Miyashita M, Suzuki K *et al.* (2014) Acute ingestion of catechin-rich green tea improves postprandial glucose status and increases serum thioredoxin concentrations in postmenopausal women. *Br J Nutr* **112**, 1542–1550.
- 117. Yang CS, Chen L, Lee MJ et al. (1998) Blood and urine levels of tea catechins after ingestion of different amounts of green tea by human volunteers. *Cancer Epidemiol Biomarkers Prev* 7, 351–354.
- 118. Lorenz M, Paul F, Moobed M *et al.* (2014) The activity of catechol-*O*-methyltransferase (COMT) is not impaired by high doses of epigallocatechin-3-gallate (EGCG) *in vivo. Eur J Pharmacol* **740**, 645–651.
- 119. Kurogi M, Miyashita M, Emoto Y *et al.* (2012) Green tea polyphenol epigallocatechin gallate activates TRPA1 in an intestinal enteroendocrine cell line, STC-1. *Chem Senses* **37**, 167–177.
- 120. Kurogi M, Kawai Y, Nagatomo K et al. (2015) Auto-oxidation products of epigallocatechin gallate activate TRPA1 and TRPV1 in sensory neurons. Chem Senses 40, 27–46.
- 121. Sanz-Salvador L, Andrés-Borderia A, Ferrer-Montiel A et al. (2012) Agonist- and Ca²⁺-dependent desensitization of TRPV1 channel targets the receptor to lysosomes for degradation. J Biol Chem 287, 19462–19471.
- 122. Dulloo AG, Seydoux J, Girardier L *et al.* (2000) Green tea and thermogenesis: interactions between catechinpolyphenols, caffeine and sympathetic activity. *Int J Obes Relat Metab Disord* **24**, 252–258.
- 123. Ramsey IS, Delling M & Clapham DE (2006) An introduction to TRP channels. *Annu Rev Physiol* 68, 619–647.
- 124. Leidy HJ, Todd CB, Zino AZ *et al.* (2015) Consuming high-protein soy snacks affects appetite control, satiety, and diet quality in young people and influences select aspects of mood and cognition. *J Nutr* **145**, 1614–1622.
- 125. Martínez de Morentin PB, González-García I, Martins L et al. (2014) Estradiol regulates brown adipose tissue thermogenesis via hypothalamic AMPK. Cell Metab 20, 41–53.
- 126. Prossnitz ER & Barton M (2011) The G-protein-coupled estrogen receptor GPER in health and disease. *Nat Rev Endocrinol* **7**, 715–726.
- 127. Suetsugi M, Su L, Karlsberg K *et al.* (2003) Flavone and isoflavone phytoestrogens are agonists of estrogen-related receptors. *Mol Cancer Res* **1**, 981–991.
- 128. Monjo M, Rodríguez AM, Palou A *et al.* (2003) Direct effects of testosterone, 17 beta-estradiol, and progesterone on adrenergic regulation in cultured brown adipocytes: potential mechanism for gender-dependent thermogenesis. *Endocrinology* **144**, 4923–4930.
- 129. Mattingly KA, Ivanova MM, Riggs KA et al. (2008) Estradiol stimulates transcription of nuclear respiratory factor-1 and increases mitochondrial biogenesis. *Mol Endocrinol (Baltim, MD)* **22**, 609–622.
- Rodríguez-Cuenca S, Monjo M, Gianotti M et al. (2007) Expression of mitochondrial biogenesis-signaling factors in brown adipocytes is influenced specifically by 17beta-estradiol, testosterone, and progesterone. Am J Physiol Endocrinol Metab 292, E340–E346.
- 131. Kamiya T, Nagamine R, Sameshima-Kamiya M et al. (2012) The isoflavone-rich fraction of the crude extract

of the Puerariae flower increases oxygen consumption and BAT UCP1 expression in high-fat diet-fed mice. *Glob J Health Sci.* **4**, 147–155.

- Cederroth CR, Vinciguerra M, Kühne F *et al.* (2007) A phytoestrogen-rich diet increases energy expenditure and decreases adiposity in mice. *Environ Health Perspect* 115, 1467–1473.
- 133. Russell AL, Grimes JM, Cruthirds DF et al. (2017) Dietary isoflavone-dependent and estradiol replacement effects on body weight in the ovariectomized (OVX) rat. *Horm Metab Res/Horm Stoffwechselforsch/Horm Metab* 49, 457–465.
- 134. Lephart ED, Porter JP, Lund TD *et al.* (2004) Dietary isoflavones alter regulatory behaviors, metabolic hormones and neuroendocrine function in long-Evans male rats. *Nutr Metab* **1**, 16.
- 135. Crespillo A, Alonso M, Vida M et al. (2011) Reduction of body weight, liver steatosis and expression of stearoyl-CoA desaturase 1 by the isoflavone daidzein in diet-induced obesity. Br J Pharmacol 164, 1899–1915.
- 136. Guo Y, Wu G, Su X *et al.* (2009) Antiobesity action of a daidzein derivative on male obese mice induced by a high-fat diet. *Nutr Res (New York, NY)* **29**, 656–663.
- Szkudelska K, Szkudelski T & Nogowski L (2002) Daidzein, coumestrol and zearalenone affect lipogenesis and lipolysis in rat adipocytes. *Phytomedicine* 9, 338–345.
- 138. Ko W-C, Shih C-M, Lai Y-H *et al.* (2004) Inhibitory effects of flavonoids on phosphodiesterase isozymes from Guinea pig and their structure-activity relationships. *Biochem Pharmacol* **68**, 2087–2094.
- 139. Chrzan BG & Bradford PG (2007) Phytoestrogens activate estrogen receptor beta1 and estrogenic responses in human breast and bone cancer cell lines. *Mol Nutr Food Res* **51**, 171–177.
- 140. An J, Tzagarakis-Foster C, Scharschmidt TC et al. (2001) Estrogen receptor beta-selective transcriptional activity and recruitment of coregulators by phytoestrogens. J Biol Chem 276, 17808–17814.
- 141. Zhou L, Xiao X, Zhang Q *et al.* (2019) A possible mechanism: genistein improves metabolism and induces white fat browning through modulating hypothalamic expression of Ucn3, Depp, and Stc1. *Front Endocrinol* **10**, 478.
- 142. Jamieson PM, Cleasby ME, Kuperman Y *et al.* (2011) Urocortin 3 transgenic mice exhibit a metabolically favourable phenotype resisting obesity and hyperglycaemia on a high-fat diet. *Diabetologia* **54**. 2392–2403.
- 143. Telegdy G, Adamik A & Tóth G (2006) The action of urocortins on body temperature in rats. *Peptides* 27, 2289–2294.
- 144. Kotz CM, Wang C, Levine AS *et al.* (2002) Urocortin in the hypothalamic PVN increases leptin and affects uncoupling proteins-1 and -3 in rats. *Am J Physiol: Regul Integr Comp Physiol* **282**, R546–R551.
- 145. Palacios-González B, Vargas-Castillo A, Velázquez-Villegas LA *et al.* (2019) Genistein increases the thermogenic program of subcutaneous WAT and increases energy expenditure in mice. *J Nutr Biochem* **68**, 59–68.
- 146. Buhlmann E, Horváth C, Houriet J et al. (2019) Puerariae lobatae root extracts and the regulation of brown fat activity. *Phytomedicine* 64, 153075.
- 147. Penza M, Montani C, Romani A *et al.* (2006) Genistein affects adipose tissue deposition in a dose-dependent and gender-specific manner. *Endocrinology* **147**, 5740–5751.
- 148. Velickovic K, Cvoro A, Srdic B *et al.* (2014) Expression and subcellular localization of estrogen receptors α and β in human fetal brown adipose tissue. *J Clin Endocrinol Metab* **99**, 151–159.

- 149. Kostelac D, Rechkemmer G & Briviba K (2003) Phytoestrogens modulate binding response of estrogen receptors alpha and beta to the estrogen response element. *J Agric Food Chem* **51**, 7632–7635.
- 150. van der Velpen V, Hollman PC, van Nielen M *et al.* (2014) Large inter-individual variation in isoflavone plasma concentration limits use of isoflavone intake data for risk assessment. *Eur J Clin Nutr* **68**, 1141–1147.
- 151. Ponnusamy S, Tran QT, Harvey I *et al.* (2017) Pharmacologic activation of estrogen receptor β increases mitochondrial function, energy expenditure, and brown adipose tissue. *FASEB* **31**, 266–281.
- 152. Liu Z-M, Ho SC, Chen Y-M *et al.* (2013) A six-month randomized controlled trial of whole soy and isoflavones daidzein on body composition in equol-producing post-menopausal women with prehypertension. *J Obes* **2013**, 359763.
- 153. Jones G, Dwyer T, Hynes K *et al.* (2003) A randomized controlled trial of phytoestrogen supplementation, growth and bone turnover in adolescent males. *Eur J Clin Nutr* **57**, 324–327.
- 154. Anderson JW, Fuller J, Patterson K *et al.* (2007) Soy compared to casein meal replacement shakes with energyrestricted diets for obese women: randomized controlled trial. *Metabolism* **56**, 280–288.
- 155. Ye Y-B, Chen A-L, Lu W *et al.* (2015) Daidzein and genistein fail to improve glycemic control and insulin sensitivity in Chinese women with impaired glucose regulation: a double-blind, randomized, placebo-controlled trial. *Mol Nutr Food Res* **59**, 240–249.
- 156. Amanat S, Eftekhari MH, Fararouei M *et al.* (2018) Genistein supplementation improves insulin resistance and inflammatory state in non-alcoholic fatty liver patients: a randomized, controlled trial. *Clin Nutr* **37**, 1210–1215.
- 157. Christie DR, Grant J, Darnell BE *et al.* (2010) Metabolic effects of soy supplementation in postmenopausal Caucasian and African American women: a randomized, placebo-controlled trial. *Am J Obstet Gynecol* **203**, 153.e1–153.e9.
- 158. Sites CK, Cooper BC, Toth MJ et al. (2007) Effect of a daily supplement of soy protein on body composition and insulin secretion in postmenopausal women. Fertil Steril 88, 1609–1617.
- 159. Barnes S, Prasain J, D'Alessandro T *et al.* (2011) The metabolism and analysis of isoflavones and other dietary polyphenols in foods and biological systems. *Food Funct* 2, 235–244.
- 160. Atkinson C, Frankenfeld CL & Lampe JW (2005) Gut bacterial metabolism of the soy isoflavone daidzein: exploring the relevance to human health. *Exp Biol Med* (*Maywood*, *NJ*) **230**, 155–170.
- 161. Muthyala RS, Ju YH, Sheng S *et al.* (2004) Equol, a natural estrogenic metabolite from soy isoflavones: convenient preparation and resolution of R- and S-equols and their differing binding and biological activity through estrogen receptors alpha and beta. *Bioorg Med Chem* **12**, 1559–1567.
- 162. Arai Y, Uehara M, Sato Y *et al.* (2000) Comparison of isoflavones among dietary intake, plasma concentration and urinary excretion for accurate estimation of phyto-estrogen intake. *J Epidemiol* **10**, 127–135.
- 163. Akaza H, Miyanaga N, Takashima N et al. (2004) Comparisons of percent equol producers between prostate cancer patients and controls: case-controlled studies of isoflavones in Japanese, Korean and American residents. *Jpn J Clin Oncol* 34, 86–89.

- 164. Liu B, Qin L, Liu A et al. (2010) Prevalence of the equolproducer phenotype and its relationship with dietary isoflavone and serum lipids in healthy Chinese adults. J Epidemiol 20, 377–384.
- 165. Zhang Y-B, Chen W-H, Guo J-J et al. (2013) Soy isoflavone supplementation could reduce body weight and improve glucose metabolism in non-Asian postmenopausal women – a meta-analysis. Nutrition 29, 8–14.
- 166. Akhlaghi M, Zare M & Nouripour F (2017) Effect of soy and soy isoflavones on obesity-related anthropometric measures: a systematic review and meta-analysis of randomized controlled clinical trials. *Adv Nutr* 8, 705– 717.
- Nagata C (2010) Factors to consider in the association between soy isoflavone intake and breast cancer risk. J Epidemiol 20, 83–89.
- 168. Matsuura HN & Fett-Neto AG (2017) Plant alkaloids: main features, toxicity, and mechanisms of action. In *Plant Toxins*, pp. 243–261 [P Gopalakrishnakone, C Carlini and R Ligabue-Braun, editors]. Dordrecht: Springer.
- Orellana-Escobedo L, Garcia-Amezquita LE, Olivas GI et al. (2013) Capsaicinoids content and proximate composition of Mexican chili peppers (*Capsicum* spp.) cultivated in the State of Chihuahua. CyTA – J Food 11, 179–184.
- 170. Scientific Committee on Food (2002) Opinion of the Scientific Committee on Food on Capsaicin. 12. Belgium: European Commission Health & Consumer protection directorate-general.
- 171. Bevan S & Szolcsányi J (1990) Sensory neuron-specific actions of capsaicin: mechanisms and applications. *Trends Pharmacol Sci* **11**, 330–333.
- 172. Henry CJ & Emery B (1986) Effect of spiced food on metabolic rate. *Hum Nutr Clin Nutr* **40**, 165–168.
- 173. Yoshioka M, Lim K, Kikuzato S et al. (1995) Effects of red-pepper diet on the energy metabolism in men. J Nutr Sci Vitaminol (Tokyo) 41, 647–656.
- 174. Yoshioka M, St-Pierre S, Drapeau V *et al.* (1999) Effects of red pepper on appetite and energy intake. *Br J Nutr* 82, 115–123.
- 175. Smeets AJ & Westerterp-Plantenga MS (2009) The acute effects of a lunch containing capsaicin on energy and substrate utilisation, hormones, and satiety. *Eur J Nutr* **48**, 229–234.
- Galgani JE & Ravussin E (2010) Effect of dihydrocapsiate on resting metabolic rate in humans. *Am J Clin Nutr* 92, 1089–1093.
- 177. Iida T, Moriyama T, Kobata K *et al.* (2003) TRPV1 activation and induction of nociceptive response by a non-pungent capsaicin-like compound, capsiate. *Neuropharmacology* 44, 958–967.
- 178. Sasahara I, Furuhata Y, Iwasaki Y et al. (2010) Assessment of the biological similarity of three capsaicin analogs (capsinoids) found in non-pungent chili pepper (CH-19 sweet) fruits. *Biosci Biotechnol Biochem* **74**, 274– 278.
- 179. Ludy M-J, Moore GE & Mattes RD (2012) The effects of capsaicin and capsiate on energy balance: critical review and meta-analyses of studies in humans. *Chem Senses* 37, 103–121.
- 180. Ohnuki K, Haramizu S, Oki K et al. (2001) Administration of capsiate, a non-pungent capsaicin analog, promotes energy metabolism and suppresses body fat accumulation in mice. *Biosci Biotechnol Biochem* 65, 2735–2740.

- 181. Kawabata F, Inoue N, Masamoto Y et al. (2009) Non-pungent capsaicin analogs (capsinoids) increase metabolic rate and enhance thermogenesis via gastrointestinal TRPV1 in mice. *Biosci Biotechnol Biochem* 73, 2690– 2697.
- 182. Baskaran P, Krishnan V, Fettel K *et al.* (2017) TRPV1 Activation counters diet-induced obesity through sirtuin-1 activation and PRDM-16 deacetylation in brown adipose tissue. *Int J Obes* 2005 **41**, 739–749.
- 183. Baskaran P, Krishnan V, Ren J et al. (2016) Capsaicin induces browning of white adipose tissue and counters obesity by activating TRPV1 channel-dependent mechanisms. Br J Pharmacol 173, 2369–2389.
- 184. Kang J-H, Goto T, Han I-S et al. (2010) Dietary capsaicin reduces obesity-induced insulin resistance and hepatic steatosis in obese mice fed a high-fat diet. *Obesity* (Silver Spring, MD) 18, 780–787.
- 185. Yoshida T, Yoshioka K, Wakabayashi Y et al. (1988) Effects of capsaicin and isothiocyanate on thermogenesis of interscapular brown adipose tissue in rats. J Nutr Sci Vitaminol (Tokyo) 34, 587–594.
- 186. Masuda Y, Haramizu S, Oki K *et al.* (2003) Upregulation of uncoupling proteins by oral administration of capsiate, a nonpungent capsaicin analog. *J Appl Physiol (Bethesda, MD)* 1985 **95**, 2408–2415.
- 187. Kawada T, Hagihara K & Iwai K (1986) Effects of capsaicin on lipid metabolism in rats fed a high fat diet. J Nutr 116, 1272–1278.
- 188. Janssens PLHR, Hursel R, Martens EAP *et al.* (2013) Acute effects of capsaicin on energy expenditure and fat oxidation in negative energy balance. *PLoS ONE* **8**, e67786.
- Snitker S, Fujishima Y, Shen H et al. (2009) Effects of novel capsinoid treatment on fatness and energy metabolism in humans: possible pharmacogenetic implications. Am J Clin Nutr 89, 45–50.
- 190. Sun L, Camps SG, Goh HJ et al. (2018) Capsinoids activate brown adipose tissue (BAT) with increased energy expenditure associated with subthreshold 18-fluorine fluorodeoxyglucose uptake in BAT-positive humans confirmed by positron emission tomography scan. Am J Clin Nutr 107, 62–70.
- 191. Yoneshiro T, Aita S, Kawai Y *et al.* (2012) Nonpungent capsaicin analogs (capsinoids) increase energy expenditure through the activation of brown adipose tissue in humans. *Am J Clin Nutr* **95**, 845–850.
- 192. Rollyson WD, Stover CA, Brown KC *et al.* (2014) Bioavailability of capsaicin and its implications for drug delivery. *J Controlled Release* **196**, 96–105.
- 193. Baboota RK, Singh DP, Sarma SM *et al.* (2014) Capsaicin induces 'brite' phenotype in differentiating 3T3-L1 preadipocytes. *PLoS ONE* **9**, e103093.
- 194. Kong W, Wei J, Abidi P *et al.* (2004) Berberine is a novel cholesterol-lowering drug working through a unique mechanism distinct from statins. *Nat Med* **10**, 1344–1351.
- 195. Lee YS, Kim WS, Kim KH *et al.* (2006) Berberine, a natural plant product, activates AMP-activated protein kinase with beneficial metabolic effects in diabetic and insulin-resistant states. *Diabetes* **55**, 2256–2264.
- 196. Turner N, Li J-Y, Gosby A *et al.* (2008) Berberine and its more biologically available derivative, dihydroberberine, inhibit mitochondrial respiratory complex I: a mechanism for the action of berberine to activate AMP-activated protein kinase and improve insulin action. *Diabetes* 57, 1414–1418.
- 197. Perdikari A, Kulenkampff E, Rudigier C et al. (2017) A high-throughput, image-based screen to identify kinases

involved in brown adipocyte development. *Sci Signal* **10** [Epublication].

- 198. Ahmadian M, Abbott MJ, Tang T *et al.* (2011) Desnutrin/ ATGL is regulated by AMPK and is required for a brown adipose phenotype. *Cell Metab* **13**, 739–748.
- 199. Wu L, Zhang L, Li B *et al.* (2018) AMP-Activated protein kinase (AMPK) regulates energy metabolism through modulating thermogenesis in adipose tissue. *Front Physiol* **9**, 122.
- 200. Zhang Z, Zhang H, Li B *et al.* (2014) Berberine activates thermogenesis in white and brown adipose tissue. *Nat Commun* **5**, 5493.
- 201. Sun Y, Xia M, Yan H *et al.* (2018) Berberine attenuates hepatic steatosis and enhances energy expenditure in mice by inducing autophagy and fibroblast growth factor 21. Br J Pharmacol 175, 374–387.
- 202. Li Y, Wong K, Giles A *et al.* (2014) Hepatic SIRT1 attenuates hepatic steatosis and controls energy balance in mice by inducing fibroblast growth factor 21. *Gastroenterology* **146**, 539–549.
- 203. Hondares E, Iglesias R, Giralt A et al. (2011) Thermogenic activation induces FGF21 expression and release in brown adipose tissue. J Biol Chem 286, 12983–12990.
- 204. Fisher FM, Kleiner S, Douris N *et al.* (2012) FGF21 Regulates PGC-1α and browning of white adipose tissues in adaptive thermogenesis. *Genes Dev* **26**, 271–281.
- 205. Wu L, Xia M, Duan Y *et al.* (2019) Berberine promotes the recruitment and activation of brown adipose tissue in mice and humans. *Cell Death Dis* **10**, 468.
- 206. Yan H-M, Xia M-F, Wang Y et al. (2015) Efficacy of berberine in patients with non-alcoholic fatty liver disease. *PLoS ONE* 10, e0134172.
- 207. Zhang Y, Li X, Zou D *et al.* (2008) Treatment of type 2 diabetes and dyslipidemia with the natural plant alkaloid berberine. *J Clin Endocrinol Metab* **93**, 2559–2565.
- Yin J, Xing H & Ye J (2008) Efficacy of berberine in patients with type 2 diabetes mellitus. *Metabolism* 57, 712–717.
- 209. Hu Y, Ehli EA, Kittelsrud J *et al.* (2012) Lipid-lowering effect of berberine in human subjects and rats. *Phytomedicine* **19**, 861–867.
- Hua W, Ding L, Chen Y *et al.* (2007) Determination of berberine in human plasma by liquid chromatographyelectrospray ionization-mass spectrometry. *J Pharm Biomed Anal* 44, 931–937.
- 211. Chen W, Miao Y-Q, Fan D-J *et al.* (2011) Bioavailability study of berberine and the enhancing effects of TPGS on intestinal absorption in rats. *AAPS PharmSciTech* **12**, 705–711.
- 212. Feng R, Shou J-W, Zhao Z-X *et al.* (2015) Transforming berberine into its intestine-absorbable form by the gut microbiota. *Sci Rep* **5**, 1–15.
- 213. Beery AK & Zucker I (2011) Sex bias in neuroscience and biomedical research. *Neurosci Biobehav Rev* **35**, 565–572.
- 214. Rodriguez-Cuenca S, Pujol E, Justo R *et al.* (2002) Sex-dependent thermogenesis, differences in mitochondrial morphology and function, and adrenergic response in brown adipose tissue. *J Biol Chem* **277**, 42958–42963.
- 215. Enriori PJ, Sinnayah P, Simonds SE *et al.* (2011) Leptin action in the dorsomedial hypothalamus increases sympathetic tone to brown adipose tissue in spite of systemic leptin resistance. *J Neurosci* **31**, 12189–12197.
- Siegrist-Kaiser CA, Pauli V, Juge-Aubry CE *et al.* (1997) Direct effects of leptin on brown and white adipose tissue. *J Clin Invest* 100, 2858–2864.

- 217. Hickey MS, Israel RG, Gardiner SN *et al.* (1996) Gender differences in serum leptin levels in humans. *Biochem Mol Med* **59**, 1–6.
- 218. Landt M, Gingerich RL, Havel PJ *et al.* (1998) Radioimmunoassay of rat leptin: sexual dimorphism reversed from humans. *Clin Chem* **44**, 565–570.
- 219. Org E, Mehrabian M, Parks BW *et al.* (2016) Sex differences and hormonal effects on gut microbiota composition in mice. *Gut Microbes* **7**, 313–322.
- 220. Waxman DJ & Holloway MG (2009) Sex differences in the expression of hepatic drug metabolizing enzymes. *Mol Pharmacol* **76**, 215–228.
- 221. Li G, Zhao M, Qiu F *et al.* (2018) Pharmacokinetic interactions and tolerability of berberine chloride with simvastatin and fenofibrate: an open-label, randomized, parallel study in healthy Chinese subjects. *Drug Des Devel Ther* **13**, 129–139.
- 222. Hill JO, Wyatt HR & Peters JC (2012) Energy balance and obesity. *Circulation* **126**, 126–132.