

Contents lists available at ScienceDirect

# Journal of Clinical Tuberculosis and Other Mycobacterial Diseases



journal homepage: www.elsevier.com/locate/jctube

# In-silico design and ADMET predictions of some new imidazo[1,2-a] pyridine-3-carboxamides (IPAs) as anti-tubercular agents

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#### ARTICLE INFO

Keywords: In-silico design Tuberculosis Binding affinity Pharmacokinetics Molecular interactions Hydrogen bond

# ABSTRACT

Tuberculosis (TB) is one of the leading infectious diseases worldwide even with the ravaging COVID-19 pandemic in recent times. This mandated further search and exploration of more possible anti-TB drug candidates against *M. tuberculosis* strains. As an extension of our previous work on the homology modeled cytochrome *b* subunit of the bc1 complex (QcrB) of Mycobacterium tuberculosis, an in-silico design was carried out in order to further explore more newly potential anti-TB compounds. Ligand 26 was selected as the lead template (scaffold A) based on our previous docking results and its less bulky structure. Successively, eight (8) new ligands (A1–A8) were designed with better binding affinities in comparison to the scaffold template (–6.8 kcal/mol) and isoniazid standard drug (–6.00 kcal/mol) respectively. In addition, three (3) designed ligands namely, **A6**, **A2**, **and A7** with higher binding affinities were validated via ADME and toxicity prediction analysis, and the results showed zero violations of Lipinski rules with similar bioavailability, and high rate in gastrointestinal absorption, while toxicity parameters such as carcinogenicity and cytotoxicity were all predicted as non-toxic (inactiveness). The designed IPA compounds in the present study could serve as a promising gateway that could help the medicinal and synthetic chemist in the exploration of a new set of derivatives as anti-TB agents. Therefore, this research strongly recommends further experimental consideration of the newly designed IPA compounds through synthesis, in-vitro and in-vivo studies to validate the theoretical findings.

# 1. Introduction

*Mycobacterium* tuberculosis is the organism that causes one of the chronic infectious diseases popularly known as Tuberculosis (TB) responsible for the global high mortality rate [1]. The emergence of the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) as the cursor of the COVID-19 pandemic has continued to dominate the scientific research community and other media outlets in recent times [2,3]. Scientific evidence based on clinical perspective indicates that COVID-19 materializes regardless of TB manifestation, either after, during, or before an active diagnosis [2]. Therefore, TB should be given utmost attention even with its global declining rate of cases [1]. An imidazo [1, 2-a] pyridine-3-carboxamide (IPA) candidate (Q203) was reported to exhibit robust inhibitory activity against extensively drugresistant (XDR) and multidrug-resistant (MDR) strains and it is currently in clinical trials [4]. Researchers are currently developing a

keen interest in the synthesis of diverse series of compounds as anti-TB agents. Recently, benzo[d]imidazole-2-carboxamides and benzimidazoquinazoline derivatives as new anti-TB agents were designed, synthesized, and tested for biological responses respectively [5,6]. Hence, the rapid increase in the occurrences of TB drug resistance attracts the need to find new therapeutics as well to discover novel drug targets that could effectively kill M. tuberculosis when exploited. Some of the promiscuous targets inhibited by more than one compound include DprE1, MmpL3, QcrB, etc [7]. The novel derivatives of Q203 (IPAs) as anti-TB agents were also reported to have the ability to block the growth of MDR and XDR strains of M. tuberculosis by targeting the respiratory cytochrome bc1 complex (QcrB) [7]. The QcrB subunit is an important component of the electron transport chain necessary for the synthesis of ATP as it catalyzes the transfer of an electron from the ubiquinol to the cytochrome *c* [8]. However, the interaction of bonded ligand to the QcrB subunit receptor remains unclear and the crystal structure is not

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https://doi.org/10.1016/j.jctube.2021.100276

Available online 20 September 2021

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available in the Protein Data Bank (PDB) [9]. The search for more potent compounds is very tedious, costly, and time-consuming [10]. As such, the use of computational chemistry tools based on theoretical insights could come in handy with the aim to modify and design new compounds with better bioactivities. Some of the computational methods employed in computer-aided drug design include homology modeling, molecular docking simulation, pharmacokinetic predictions, and QSAR analysis amongst others. These computational approaches have been employed over the years to improve existing anti-tubercular agents through virtual screening for the identification and modification of potential hits [11,12]. Structure-based drug design (SBDD) solemnly depends on the knowledge and information of the 3D crystal structure of the targeted

Table 1

R<sub>1</sub>



protein to design the ligands that can serve as better inhibitors [13]. In the case where the 3D experimental structure of the targeted protein is not reported, the experimental amino acid sequence can be used to build a homology model [14]. The homology modeling technique predicts the 3D structure of the targeted protein sequence based on the alignment of an experimentally known homologous protein as a template [15]. In our previous report, homology modeling and molecular docking studies were carried out on some IPAs anti-TB agents targeting the OcrB subunit. The homology modeling of the receptor built and predicted a new 3D structure of QcrB target in M. tuberculosis using QcrB subunit of *M. smegmatis* as template [12,16]. Furthermore, the results of molecular docking in the study further revealed the binding profiling of the 35 IPA



ligands docked with the modeled protein. In the current study, the same 3D crystal structure of the QcrB modeled protein in *M. tuberculosis* was used to analyze the binding profiling and ADMET prediction of some newly designed compounds as potential hits of anti-TB candidates.

# 2. Methodology

# 2.1. Template selection and structural modifications

In our previous report, we have successfully carried out virtual screening of thirty-five (35) N-(2-phenoxy) ethyl imidazo[1,2-a] pyridine-3-carboxamides (IPAs) synthesized by Wang et al., (2019) with our homology modeled QcrB protein as the active target in the *Mycobacterium tuberculosis* [7,16]. As such, ligand 26 was selected as the template scaffold for further structural modification and rigorous molecular docking simulation. The structure of the newly designed ligands was drawn (Table 1) and optimized accurately at the density functional level of theory (B3LYP/6-31G\*\*) in a vacuum using Spartan 14 [17].

# 2.2. Molecular docking, ADME analysis, and toxicity prediction

Molecular docking is the most preferable technique in structurebased drug design to predict the binding free energy and the binding mode of the protein and ligand compound [18]. Therefore, molecular docking simulation was carried out to determine the binding affinities and the residual interactions when the ligand molecules bind with the active pockets of the protein as macromolecule using AutoDock 4.2 module implemented in PyRx 0.8. Blind docking was performed for all the designed ligand molecules to predict the active binding pockets of the modeled QcrB protein as the targeted macromolecule [19]. To ensure that all ligand molecules are properly docked, the 3D grid box dimensions were adjusted as X: 203.60, Y: 177.43, Z: 211.23 for grid center, and X: 88.26, Y: 86.09, Z: 82.38 for the number of points at the spacing of 1.875 Å on the whole protein structure to predict the best outcome of the docking task. Furthermore, the docking algorithm used was the Lamarckian Genetic Algorithm at default parametrized settings. After docking, protein and the ligands were obtained in PDBQT format, and complexes were formed using UCSF Chimera software while the visualization of residual interactions was done using Discovery Studio 2020 and UCSF Chimera software accordingly. The Swiss ADME online server (http://www.swissadme.ch/) was applied to predict absorption, distribution, metabolism, and excretion properties of the best ligands while ProTox-II online server (https://tox-new.charite.de/protox II/) was also used to determine their toxicity.

# 3. Results and discussions

# 3.1. Molecular docking analysis

The docking results of ligand molecules with the targeted protein showed the binding affinity ranging from (-8.5 kcal/mol to -11 kcal/mol). To compare the best binding affinity of the ligand molecules, we docked the standard drug with the modeled QcrB protein in *M. tuberculosis* and showed binding affinity as (-6.00 kcal/mol). All binding amino acid residues including non-bond interactions and binding affinities of the stable complexes formed were shown in Table 2.

A6 showed the best binding affinity (-11.0 kcal/mol) as a complex with the respected modeled QcrB protein and formed one conventional hydrogen bond with the amino acid residue of (GLY62 at a distance of 2.39142 Å) and Halogen (Fluorine), Amide-Pi Stacked, Alkyl, Pi-Alkyl bonds with the amino acid residues of (LEU58, LEU59, VAL63, ILE217, LEU65, LEU166, PRO167, PRO221, PHE69, TYR213) showed in Fig. 1. The complex of the A2 ligand molecule with the targeted modeled QcrB protein showed (-10.5 kcal/mol) binding affinity and formed one Conventional Hydrogen Bond with the amino acid residue (GLY62 at a distance of 2.08894 Å). Four different types of bonds such as Halogen

#### Table 2

Binding affinity (kcal/mol) and non-bonding interactions of the complexes.

Compounds	Binding affinity	Bonding types	Interacting amino acid	Distance (Å)
	(kcal/lilol)		residues	
Standard –6.00 drug		Conventional Hydrogen Bond	LEU58	2.09388
		Conventional Hydrogen Bond	LEU59	2.84072
		Pi-Anion	GLU159	3.32022
		Pi-Alkyl	LEU58	3.97204
	0.5	Pi-Alkyl	PRO221	5.18191
AI	-8.5	Univentional Hydrogen Bond	ALA385	2.52924
		Halogen (Eluorine)	LEU348	2.87618
		Pi-Sigma	PHF133	3 61502
		Pi-Sigma	ALA385	3.67506
		Pi-Sigma	ALA385	3.60692
		Pi-Pi T-shaped	PHE133	4.99664
		Amide-Pi Stacked	ALA385	4.12602
		Amide-Pi Stacked	ILE386	4.12602
		Alkyl	LEU129	5.40777
		Alkyl	ILE380	4.18/8/
		Alkyl	VAL345 ALA385	4.30783
		Alkyl	ALA385	4.32462
		Pi-Alkyl	ILE386	5.06303
		Pi-Alkyl	LEU129	5.19201
		Pi-Alkyl	PHE133	4.1159
		Pi-Alkyl	PHE134	4.35564
		Pi-Alkyl	PHE388	4.7971
4.2	10 5	Pi-Alkyl Comucentional	TYR389	4.44871
A2	-10.5	Hydrogen Bond	GL102	2.08894
		Halogen	GLU159	3.59989
		(Fluorine) Pi-Anion	GU1159	4 31 326
		Alkyl	LEU59	3.92938
		Alkyl	PRO221	4.39931
		Alkyl	LEU65	4.57881
		Alkyl	ARG111	4.54332
		Alkyl	PRO167	4.47863
		Alkyl	LEU65	4.48087
		Alkyl	LEU166 DBO167	5.41423
		Pi-Alkyl	ILF217	4 59328
		Pi-Alkyl	PRO221	4.71614
		Pi-Alkyl	PHE69	5.14437
		Pi-Alkyl	PHE69	4.72374
A3	-10.0	Halogen	HIS114	3.36308
		(Fluorine)		
		Pi-Anion	GLU159	3.94788
		Alkyl	LEU58	3.81904
		Alkyl	PRO221	4.09033
		Alkyl	LEU65	4.40346
		Alkyl	LEU166	4.97691
		Pi-Alkyl	LEU58	5.39169
		Pi-Alkyl	LEU59	5.27014
		Pi-Alkyl	PRO221	4.32695
		Pi-Alkyl	PHE69	4.72942
		Pi-Alkyl	HIS114	5.15802
A.4	0.1	PI-AIKYI Carbon Hydrogen	HI5216	5.28912
A4	-9.1	Bond	GL1105	5.51051
		Halogen	GLY163	3.31031
		Halogen	HIS114	3.68598
		(Fluorine) Halogen	HIS216	3.05615
		(Fluorine)	L D L C C	0 5055
		P1-Sigma	LEU65	3.7055
		Alkyl	ALA97 ILE100	3.09520 4 33311
		Alkyl	ARG111	4.58662
		Alkyl	PRO167	4.85181

(continued on next page)

# Table 2 (continued)

Compounds	Binding	Bonding types	Interacting	Distance		
	affinity (kcal/mol)	residues		(A)		
		Allari	II F217	4 56014		
		Alkyl	PRO221	5.48313		
		Pi-Alkyl	PRO167	5.10454		
		Pi-Alkyl	PHE69	5.29162		
		Pi-Alkyl	HIS114	4.68175		
		Pi-Alkyl	HIS216	5.24304		
A5	-10.3	Halogen	HIS114	3.50679		
		(Fluorine)	LEUEO	4.04964		
		Alkyl	PRO221	4.04304		
		Alkyl	LEU65	4.70392		
		Alkyl	ILE217	5.46661		
		Alkyl	LEU65	4.52788		
		Alkyl	LEU65	4.86007		
		Alkyl	LEU166	4.60995		
		Alkyl Di Allavl	PRO221	5.42632		
		Pi-Alkyl	PRO221	3.39003 4 44757		
		Pi-Alkyl	PHE69	4.87213		
		Pi-Alkyl	HIS114	5.14963		
		Pi-Alkyl	HIS114	5.12793		
		Pi-Alkyl	HIS216	5.28053		
A6	-11.0	Conventional	GLY62	2.39142		
		Hydrogen Bond	CLUIFO	2 66252		
		(Fluorine)	GLU159	3.00232		
		Amide-Pi Stacked	LEU58	4.97455		
		Amide-Pi Stacked	LEU59	4.97455		
		Alkyl	LEU58	4.97214		
		Alkyl	VAL63	4.49813		
		Alkyl	ILE217	4.54423		
		Alkyl	LEU65	4.95044		
		Alkyl	LEU100 LEU65	4 41666		
		Alkyl	PRO167	5.21434		
		Alkyl	PRO221	4.89313		
		Pi-Alkyl	ILE217	4.84499		
		Pi-Alkyl	PHE69	5.17173		
		Pi-Alkyl	PHE69	5.12895		
47	-10.5	PI-AIKYI Carbon Hydrogen	1 Y K213 HIS216	5.39932 3 78078		
117	-10.5	Bond	1113210	3.70770		
		Halogen	HIS114	3.60387		
		(Fluorine)				
		Alkyl	LEU58	4.03498		
		Alkyl	LEU59	3.97007		
		Alkyl	LEU65	5.01232		
		Alkyl	PRO167	5 11711		
		Alkyl	PRO221	5.46251		
		Pi-Alkyl	LEU59	5.39657		
		Pi-Alkyl	PRO221	4.5088		
		Pi-Alkyl	PHE69	5.20022		
		P1-AIKyl Di Allaul	HIS114	5.13828		
		Pi-Alkyl	HIS216	5 23027		
		Pi-Alkyl	HIS216	5.00678		
A8	-9.0	Conventional	ALA385	2.16555		
		Hydrogen Bond				
		Amide-Pi Stacked	ALA385	4.63904		
		Amide-Pi Stacked	ILE386	4.63904		
		Alkyl	LEU129 MET126	4.97501		
		Alkyl	VAL345	4 66888		
		Alkyl	VAL345	4.80002		
		Alkyl	LEU348	5.44256		
		Alkyl	ALA385	4.26799		
		Alkyl	ALA385	4.0649		
		Alkyl Di Allad	ALA385	4.78506		
		PI-AIKYI Pi-Alkyl	ALA385	5.14/48 4 62064		
		Pi-Alkyl	ILE386	4.76785		
		Pi-Alkyl	PHE133	4.51175		
		Pi-Alkyl	PHE388	4.91468		
		Pi-Alkyl	TYR389	3.85255		



**Fig. 1.** (a) Schematic representation of predicted **A6** ligand with protein complex interactions in the 2D diagram. Interactions are colored depending on their type. (b) The three-dimensional representation of the binding pose, interactions, H bond donor, and acceptor surface of predicted **A6** ligand with the protein complex. (c) Targeted protein is depicted in surface view and **A6** ligand compound as the stick in the binding pocket.

(Fluorine), Pi-Anion, Alkyl, Pi-Alkyl were visualized in the complex with the amino acid residues of (GLY62, GLU159, LEU59, PRO221, LEU65, ARG111, PRO167, LEU65, LEU166, ILE217, PHE69) showed in Fig. 2. A7 as a ligand compound expressed (-10.5 kcal/mol) binding affinity with the targeted modeled QcrB protein. Complex showed one Carbon Hydrogen Bond with the amino acid residue of (HIS216 at a distance of 3.78978 Å) and three different types of bonds such as Halogen

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**Fig. 2.** (a) Schematic representation of predicted **A2** ligand with protein complex interactions in the 2D diagram. Interactions are colored depending on their type. (b) The three-dimensional representation of the binding pose, interactions, H bond donor, and acceptor surface of predicted **A2** ligand with protein complex. (c) Targeted protein is depicted in surface view and **A2** ligand compound as a stick in the binding pocket.

(Fluorine), Alkyl, Pi-Alkyl with the amino acid residues of (HIS114, LEU58, LEU59, LEU65, PRO167, PRO221, LEU59, PHE69, HIS114, HIS216) showed in Fig. 3. Furthermore, A3, A4, A5, A8 ligand molecules as complexes with the targeted modeled QcrB protein also revealed higher binding affinity than the template molecule and standard drug respectively. Based on the highest molecular docking scores as binding affinity, non-bond interactions and in comparison with the binding affinity of the standard drug, three ligand compounds (A6, A2, and A7) were considered for further analysis.

# 3.2. ADME and toxicity prediction

Molecular weight (acceptable range:  $\leq$ 500), number of hydrogen bond acceptors (acceptable range:  $\leq$ 10), lipophilicity (Log P)  $\leq$  5, and

**Fig. 3.** (a) Schematic representation of predicted **A7** ligand with protein complex interactions in the 2D diagram. Interactions are colored depending on their type. (b) The three-dimensional representation of the binding pose, interactions, H bond donor, and acceptor surface of predicted **A7** ligand with protein complex. (c) Targeted protein is depicted in surface view and **A7** ligand compound as the stick in the binding pocket.

molar refractivity (40–130) indicates the five rules of Lipinski, are crucial parameters for a successful drug candidate [20]. All the ADME parameters including drug-likeness, pharmacokinetic profile, and water solubility were analyzed for the selected ligand molecules showed in Table 3. All the ligand molecules as A6, A2, and A7 revealed 0 violations in Lipinski rules, similar bioavailability, and a high rate of gastrointestinal absorption. Only the A2 ligand molecule has glycoprotein permeability. Toxicity prediction was analyzed to determine the compounds were whether toxic or not. Predicted results were shown in Table 4. Determination of carcinogenicity and cytotoxicity of A6, A2, A7 were

#### Table 3

ADME and drug-likeness parameters of the selected IPAs.

ID	MW (g/mol)	nHBD	nHBA	Log S	GA	СРҮ	BBB	Pgp	BA	Log Po/w	SA	nLV
A6	373.51	1	3	-6.41	High	CPY2 D6 inhibitor	Yes	No	0.55	5.24	3.46	0
A2	343.25	1	3	-5.17	High	CYP2D6 inhibitor, CYP3A4 inhibitor	Yes	Yes	0.55	2.95	3.82	0
A7	373.51	1	3	-6.41	High	CYP2D6 inhibitor	Yes	No	0.55	5.24	3.46	0

Key: Molecular weight (MW), Number of hydrogen bond donor (nHBD), Water solubility (Log S), gastrointestinal absorption (GI), CYP isoform inhibitor (CPY), bloodbrain barrier permeant (BBB), P-glycoprotein substrate (Pgp), Bio-availability (B), consensus Log Po/w, Synthetic Accessibility (SA), Number of Lipinski violation (nLV).

#### Table 4

Toxicity prediction of the selected IPAs.

Compound	Carcinogenicity	Cytotoxicity
A6	Inactive	Inactive
A2	Inactive	Inactive
A7	Inactive	Inactive

predicted inactiveness (non-toxic).

#### 4. Conclusion

As an extension of our previous work, this research adopted the insilico approach in analyzing the binding profiling of some newly designed IPA compounds as potential hits of anti-TB candidates. The template scaffold (Ligand 26) was selected for the in-silico design strategy and ligand compounds (A1–A8) were designed which exhibited better binding affinities when compared with that of the scaffold template (6.8 kcal/mol) and isoniazid standard drug (6.00 kcal/mol). In addition, all docking results of designed ligands with the targeted protein showed binding affinities ranging from (-8.5 kcal/mol to -11 kcal/ mol). The drug-likeness and pharmacokinetic profile prediction results for the selected ligands with higher binding affinities (A6, A2, and A7) showed zero violations of Lipinski rules with similar bioavailability, and high rate in gastrointestinal absorption, while toxicity parameters such as carcinogenicity and cytotoxicity were all predicted as non-toxic (inactiveness).

#### Ethical statement

Not applicable

# CRediT authorship contribution statement

Mustapha Abdullahi: Conceptualization, Methodology, Data curation, Visualization, Investigation, Supervision, Writing - original draft. Niloy Das: Software, Visualization, Validation, Writing - review & editing. Shola Elijah Adeniji: Data curation, Formal analysis, Supervision. Alhassan Kabiru Usman: Investigation, Writing - review & editing. Ahmad Muhammad Sani: Writing - review & editing.

#### **Declaration of Competing Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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