Targeting the FGFR signaling pathway in cholangiocarcinoma: promise or delusion?

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Abstract: Cholangiocarcinoma (CCA) is a refractory cancer with limited treatment options and poorly understood molecular mechanisms underlying tumor development. The most effective treatment is surgical resection; however, patients are highly prone to recurrence. Moreover, considering that most patients are diagnosed in advanced stages, treatment options are restricted to palliative care, which results in poor prognosis. Due to the limited effect of chemotherapy and radiotherapy, targeted therapy is becoming a hot topic in the field of biliary cancer treatment. The fibroblast growth factor/fibroblast growth factor receptor (FGF/FGFR) signaling pathway involves a variety of key biological processes for cell survival, differentiation, and metabolism. Next-generation sequencing data mining has shown that high levels of FGF/FGFR expression are associated with reduced overall survival (OS) in CAA, which indicates that the FGF/FGFR pathway may be an effective target for CAA treatment. This paper reviews the effect of FGF/FGFR signaling on CCA from onset to treatment and highlights the promise of FGF/FGFR signaling pathway inhibitors for targeting CCA.

Keywords: cholangiocarcinoma, fibroblast growth factor receptor, targeted therapy, tyrosine kinase inhibitors

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Highlights

- 1. Fibroblast growth factor/fibroblast growth factor receptor (FGF/FGFR) can regulate cell survival, proliferation, and mediate several vital physiological functions such as metabolic homeostasis, neuroendocrine balance, embryonic development, and tissue repair.
- 2. Dysregulation of the FGF/FGFR signaling pathway typically occurs through gene amplification, gain-of-function coding mutation, and gene fusion. This consequently affects a series of major biological processes and eventually causes malignancies, including cholangiocarcinoma (CCA).
- 3. CCA is a devastating cancer with a frightening 5-year survival rate of approximately 10% and few therapeutic options.
- 4. Mutations that alter *FGFRs 1–4* are frequently found in CCA, especially *FGFR2* fusion and *FGFR4* overexpression. Targeted therapies for FGFR signaling pathways in CCA, including small-molecule tyrosine

kinase inhibitors (TKIs), FGF ligand traps, and FGFR-targeted monoclonal antibodies, have proven effective and safe in a large number of preclinical and clinical trials.

5. Targeting FGF/FGFR signaling is a promising treatment approach for CCA. However, to better incorporate FGFR inhibitors into clinical practice, many variables need to be addressed, such as the mechanism underpinning FGFR-inhibitor resistance and possible solutions, the onset of chromosome aberration, and the key to establish effective targeted combinatorial therapies.

Background

Cholangiocarcinoma (CCA) is a highly malignant invasive carcinoma that originates from bile duct epithelial cells; however, the causes of CCA remain unclear. The established risk factors mainly include primary sclerosing cholangitis, bile duct abnormalities, biliary stones within the Ther Adv Med Oncol

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liver, infection with a liver fluke parasite (a common problem in Asia), exposure to certain chemicals and toxins, hepatitis B and hepatitis C virus infections, and so on.^{1,2} CCA is cancer with poorprognosis and low-incidence that is divided into intrahepatic cholangiocarcinoma (iCCA) and extrahepatic cholangiocarcinoma (eCCA) according to anatomical location.³ Although parts of the same bile duct, eCCA and iCCA were found to express different cell proteins and have different cell morphology, doubling time, chromosome karvotype, and chemotherapy sensitivity in vitro.4 Furthermore, in contrast with iCCA patients, patients with eCCA usually exhibit jaundice.5 Their clinical features and biological behaviors also have different characteristics, suggesting that CCAs are heterogeneous in their genotypes and must be studied separately depending on anatomic location.6

CCA is a devastating cancer with an alarmingly low 5-year survival rate of approximately 10% and few therapeutic options.⁷ Therefore, once CCA is suspected, comprehensive clinical examination should be performed to determine its clinical classification and staging. CCAs are relatively resistant to radiotherapy and chemotherapy; even their concurrent use (chemoradiotherapy) can barely improve the survival rate and prolong survival time in partial patients.⁸ Currently, the firstline chemotherapy for CCA is limited to 5-fluorouracil alone or its combined use with other chemotherapeutic drugs such as cisplatin. Second-line systemic treatment for advanced CCA typically shows dismal efficacy, with a median progression-free survival (PFS) value of approximately 2.7 months.9 Surgical resection has been considered the best treatment option for CCA. However, most patients are diagnosed with CCA at an advanced stage and have lost the chance to undergo radical resection (R0 resection). Therefore, it is urgent to find effective targeted therapies for CCA.

FGF/FGFR and its role in tumorigenesis

Greenman *et al.* reported that more than 1000 somatic mutations found in 274 Mb of DNA maintained consistency with the coding exons of 518 protein kinase genes in 210 diverse human malignancies.¹⁰

Human fibroblast growth factor receptors (FGFRs) – a subfamily of receptor tyrosine kinases – comprise of four family members (FGFR1–4) that interact with 22 ligands (FGF1-14, FGF16-23).11,12 The oncogenic mechanisms of FGF/ FGFR signaling are very complicated and not fully understood; FGFs activate FGFRs through paracrine or autocrine mechanisms in cooperation with heparan sulfate proteoglycans.¹⁰ Dysregulation of the FGF/FGFR signaling pathway occurs typically through gene amplification, gain-of-function coding mutation, and gene fusion¹³; this is usually mediated by fibroblast growth factor receptor substrate 2 (FRS2), mitogen activated protein kinase (MAPK)/extracellular signal-regulated kinase 1/2 (ERK1/2), phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT) signaling pathways, Janus kinase-signal transducer and activator of transcription (JAK–STAT), phospholipase $C\gamma$ (PLC γ), ribosomal protein S6 kinase 2 (RSK2) 1,2, and so on.14,15 These processes then lead to intracellular phosphorylation of receptor kinase domains, cascading reactions to intracellular signals, and gene transcription.¹⁶ Many studies have confirmed that the carcinogenicity of FGF/FGFR is a result of acquiring potential mutations that lead to proteincoding and synthesis abnormalities in this pathway, which subsequently affects a series of major biological processes and eventually cause the tumors. However, under physiological conditions, FGF/FGFR can regulate cell survival and proliferation and mediate several vital physiological functions such as metabolic homeostasis, neuroendocrine balance, embryonic development, and tissue repair.¹⁷ In recent years, FGFRs have been also found to stimulate endothelial cell proliferation and promote cancer cell migration,¹⁸ regulate tumor cell proliferation,19 and activate antiapoptotic pathways, anti-tumor responses, and angiogenesis.20-22

The FGF/FGFR signaling pathway and CCA

In a study of 4853 tumors, FGFR aberrations were found in 7.1% of cancers, with 66% gene amplification, 26% mutations, and 8% rearrangements, by next-generation sequencing²³ ;notably, these aberrations were distributed as follows: 3.5% *FGFR1* (mostly amplification), 1.5%*FGFR2*, 2.0% *FGFR3*, and 0.5% *FGFR4*. Much research has found that FGFR (1–4) and FGF (1–10, 16–19, 22–23) contributes to many cancer-related cancers, such as bladder cancer, lung cancer, breast cancer, gastric cancer, endometrial carcinoma, urothelial carcinoma, bladder cancer, melanoma, hepatocellular carcinoma, multiple myeloma, renal cell carcinoma, prostate cancer, CCA, and so on.^{12,21,22,24} Previous genomic sequencing studies have revealed that 30–40% of iCCA have actionable mutations, including *IDH1/2* (5–20%), *FGFR2* fusions (4–16%), *ARID1A* alterations (7–16%), and *BAP1* mutations (1–38%).^{25,26} Many drugs are being actively developed for the clinical setting, especially those targeting the FGF/FGFR signaling pathway, which is involved in a variety of cancers, including CCA. *FGFR2* fusion events were identified in about 13% of iCCA,⁷ whereas *FGFR4* overexpression was noted in approximately 50% of all CCAs.²⁷ In addition, *FGFR1* and *FGFR3* mutations were also detected in CCA.²⁸

In a previous in vitro study on human CCA specimens, Raggi et al. demonstrated by immunohistochemistry that FGFR1 and FGFR2 were expressed in 30% and 65% of total samples, respectively.29 Evidently, FGFR1 expression is not consistent in CCA; thus, the of FGFR1 expression in the development of CCA and possible targeted treatment choices need further investigation. The most common FGFR chromosomal aberration in CCA is FGFR2-BICC1 fusion, which is constitutively active and plays a role in the activation of MAPK and PIK3CA/mammalian target of rapamycin (mTOR) pathways.³⁰ Moreover, a previous study found that 6.6% of iCCAs have the FGFR2 translocation and that FGFR2 amplification portended a better prognosis in 122 Chinese iCCA patients.³¹ Overexpression of FGFR2 fusion proteins, generated by genetic translocations, resulted in increased sensitivity to FGFR inhibitors both in vitro and in vivo.32 Besides, a number of selective FGFR inhibitors, including BGJ398, JNJ-42756493, and AZD4547 have shown meaningful clinical or preclinical activity and manageable toxicities in chemotherapyrefractory CCA.33-35

In addition, high expression of FGF15/19 and FGFR4 is closely associated with bile acid synthesis inhibition,³⁶ and is also able to slow the progression of inflammatory bile duct diseases.³⁷ Xu *et al.* investigated FGFR4 expression in 83 iCCAs and 116 eCCAs by immunohistochemistry, and found that FGFR4 was an independent prognostic factor in iCCAs and perihilar CCAs by multivariate analysis.³⁸ Moreover, FGFR4 can induce the proliferation, invasion, and epithelial-mesenchymal transition of FGF19⁺ cell lines *in vitro*; however, AP24354 – a GFR4 inhibitor – can suppress this phenomenon. Although FGFR4 was found to be associated with poor prognosis in

CCA *via* inducing proliferation, invasion, and suppressing apoptosis, Yoo *et al.* assessed the expression of 98 genes from 46 iCCAs and found that FGFR4-related genes (FGF19, FGF21, and FGFR4) were significantly associated with better disease-free survival (DFS) in iCCA; these authors even speculated they could be used as biomarkers to define the distinctive molecular phenotype of iCCA.³⁹

Therefore, targeting FGF/FGFR signaling could be a promising candidate for CCA therapy.

Therapies targeting FGF/FGFR signaling in CCA

Altered FGFR activation results from TKI inhibitor use and triggers intracellular signaling; FGF/ FGFR interactions at the extracellular level are associated with monoclonal antibodies and FGF ligand traps. Thus, FGFR inhibitors, which can be divided into FGFR-specific small-molecule TKIs, FGF ligand traps, and FGFR-targeting monoclonal antibodies, are currently being used in preclinical and clinical trials involving patients with advanced malignancies, including CCA.

We used 'Cholangiocarcinoma/Bile duct cancer/ Biliary duct cancer' and 'FGFR' as key words to search for clinical trials on the clinicaltrials.gov site; we then collected detailed information on clinical trials related to FGFR pathway-targeting agents in CCA, as shown in Table 1.

Small-molecule TKIs of the FGFR

Small-molecule TKIs can either non-selectively inhibit FGFR signaling by competing for ATP binding domains (non-selective inhibitors) or selectively target the kinase domain of FGFRs (selective inhibitors). Considering that FGFRs are a superfamily of receptor tyrosine kinases (RTKs) with catalytic domain structural homology to a certain extent, the first-generation of TKIs was designed to express a multi-kinase activity.⁴⁰⁻⁴² Therefore, first-generation TKIs (such as nintedanib, brivanib, regorafenib, mastinib, E-3810, TSU-68, and BIBF1120) are nonselective inhibitors that can affect VEGFRs, PDGFRs, RET, KIT, and FGFRs. This suggests that, while they extensively target numerous tumorigenic growth factors, they might also indirectly drive tumor progression. Fortunately, however, the second-generation of TKIs (such as AZD4547, dovitinib, ponatinib, BGJ398, and

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Agent	Targets	Characteristics	Phase	ClinicalTrials. gov identifier:	Targeted tumors
BGJ398	FGFR1-3	FGFR-specific small- molecule TKIs	II	NCT02150967	CCA.
JNJ-42756493	FGFR1-4	FGFR-specific small- molecule TKIs	 	NCT01703481 NCT02699606	Advanced or refractory solid tumors or lymphoma. Advanced non-small-cell lung cancer, urothelial cancer, esophageal cancer or CCA.
Debio 1347-101	FGFR1-3	FGFR-specific small- molecule TKIs	Ι	NCT01948297	Advanced solid tumors.
TAS-120	FGFR1-4	FGFR-specific small- molecule TKIs	II	NCT02052778	Advanced solid tumors.
ARQ 087	FGFR1–3, RET, PDGF receptor, KIT, and SRC	FGFR-specific small- molecule TKIs	ll Ib/ll	NCT03230318 NCT0175290	Advanced intrahepatic CCA. Advanced solid tumors.
INCB054828	FGFR1-3	FGFR-specific small- molecule TKIs	Ι	NCT02393248	Advanced malignancies.
INCB062079	FGFR4	FGFR-specific small- molecule TKIs	I	NCT03144661	Advanced hepatocellular carcinoma and other malignancies.
H3B-6527	FGFR4	FGFR-specific small- molecule TKIs	Ι	NCT02834780	Advanced hepatocellular carcinoma.
Ponatinib	Abl, PDGFR α , VEGFR2, FGFR1 and Src	FGFR-specific small- molecule TKIs	II	NCT02265341 NCT02272998	Advanced biliary cancer.
Pazopanib	VEGFR1, VEGFR2, VEGFR3, PDGFRβ, c-Kit, FGFR1 and c-Fms	FGFR-specific small- molecule TKIs	II	NCT01855724	Biliary tree cancer.
FP-1039	FGFR1	FGF ligand traps	lb I	NCT01868022 NCT00687505	Solid malignancies. Advanced unresectable solid tumors.
Vofatamab (B-701)	FGFR3	FGFR-targeting monoclonal antibodies	I/II(b)	NCT02401542	Advanced urothelial cell carcinoma.
MFGR1887S	FGFR3	FGFR-targeting monoclonal antibodies	Ι	NCT01363024	Solid tumors.

Table 1. Clinical trials of FGF/FGFR signaling-targeted therapies for CCA.

CCA, cholangiocarcinoma; FGF, fibroblast growth factor; FGFR, fibroblast growth factor receptor; KIT, stem cell factor receptor; PDGF, platelet-derived growth factor; RET, rearranged during transfection; TKI, tyrosine kinase inhibitor; VEGF, vascular endothelial growth factor; VEGFR, vascular endothelial growth factor receptor.

LY2874455) serve as selective FGFR TKIs and are capable of playing targeted therapeutic roles with less toxic effects compared with those of non-selective inhibitors.

BGJ398 is a non-selective inhibitor with potential anti-angiogenic and anti-neoplastic activities; it also participates in the suppression of tumor cell differentiation and proliferation, tumor angiogenesis, and tumor cell survival. A phase II study of BGJ398 in CCAs, with FGFR2 fusions or other FGFR genetic alterations,³³ showed that the overall response rate (ORR) was 14.8%, the disease control rate was 75.4%, and the estimated median PFS was 5.8 months [95% confidence interval (CI) 4.3–7.6 months]. Furthermore, BGJ398 also exhibited manageable toxicities and a high disease control rate (75.4%).

JNJ-42756493 (Erdafitinib), also a non-selective inhibitor, has been granted Breakthrough Therapy Designation by the United States Food and Drug Administration (FDA) for the treatment of urothelial cancer and locally advanced or metastatic urothelial carcinoma in March 2018 and April 2019, respectively.⁴³ Moreover, a phase I, openlabel, multicenter, single-arm, dose-escalation study found that erdafitinib was well-tolerated in Japanese patients with advanced or refractory solid tumors including CCA; currently, a phase II study [ClinicalTrials.gov identifier: NCT02699606] to evaluate the clinical efficacy, safety, and pharmacokinetics of erdafitinib in Asian participants with CCA is ongoing.

TAS-120, which irreversibly inhibits all four FGFR subtypes, can selectively inhibit the growth of human cancer cell lines with *FGFR* gene abnormalities along with tumor growth in mouse xenograft models.⁴⁴ A clinical study reported that the irreversible FGFR inhibitor TAS-120 provides clinical benefit in patients with resistance to BGJ398 or Debio 1347 and overcomes several *FGFR2* mutations in iCCA models.⁴⁵ A phaseII study (ClinicalTrials.gov identifier: NCT02052778) to evaluate the ORR of TAS-120 is underway in approximately 100 iCCA patients with confirmed *FGFR2* gene fusions.

ARQ 087 (Derazantinib), a pan-FGFR inhibitor, is currently under clinical investigation [ClinicalTrials. gov identifier: NCT03230318] in patients with FGFR2 fusion-positive inoperable or advanced iCCAs; the results have not yet been announced. However, the latest clinical [ClinicalTrials.gov identifier: NCT01752920] shows that derazantinib has anti-tumor activity and a manageable safety profile in patients with advanced unresectable iCCA with FGFR2 fusion who progressed after chemotherapy; notably, its ORR was 20.7%, the disease control rate was 82.8%, and the estimated median PFS was 5.7 months, although, treatmentrelated adverse events (AEs) were observed in 93.1% of the patients, including asthenia/fatigue (69.0%), eye toxicity (41.4%), and hyperphosphatemia (75.9%); moreover, AEs with a grade ≥ 3 occurred in 27.6% patients.

INCB054828 (Pemigatinib) is a novel, selective, oral inhibitor of FGFR 1–3. The recent results of the single-arm, multicohort, phase II FIGHT-202 trial showed that 35.5% CCAs with FGFR2 fusions or rearrangements achieved ORR, included 2.8% complete responses, 33% partial

response.⁴⁶ But, for the other groups (CCAs with other *FGF/FGFR* alterations, CCAs with no *FGF/FGFR* alterations), no responses were observed. The median OS were 21.1 months, 6.7 months and 4.0 months, respectively, in the three groups. These data demonstrate the important role of Pemigatinib in the treatment of CCA with *FGFR2* fusions or rearrangements. Now, Pemigatinib has become the first and only FDA-approved targeted-drug for CCA. Results of studies with INCB062079, H3B-6527, and BAY1163877 regarding advanced malignancies, including CCA, are also expected to be released soon.

FGF ligand traps

This approach is based on the development of extracellular "FGF ligand traps" able to bind and isolate FGFs, thereby blocking their interaction with their receptors. For example, FP-1039 is a FGF ligand trap consisting of the extracellular domain of FGF receptor1 (FGFR1) fused with the Fc region of human immunoglobulin G1 (IgG1). It can selectively block mitogenic FGFs; however, it does not bind hormonal FGFs (FGF19, FGF21, and FGF23), which require co-receptors for binding and downstream signaling.47,48 The phaseI dose-finding study of FP-1039 [Clinical Trials.gov identifier: NCT00687505] and the phase Ib study of FP-1039 in FGFR1-amplified NSCLC [ClinicalTrials.gov identifier: NCT01868022] both showed that FP-1039 was well tolerated even in combination with chemotherapy.49,50 As a result, CCA patients are expected to have even good tolerance to FP-1039; thus, studies on the effects of FP-1039 in CCA treatment should be arranged.

The soluble pattern recognition receptor long pentraxin-3 (PTX3) acts as a multi-FGF inhibitor by binding various FGFs, such as FGF2, FGF6, FGF8b, FGF10, and FGF17.⁵¹ NSC12, a recently discovered small molecule PTX3-derived pan-FGF trap, is able to block FGF2/FGFR interaction and inhibit cell proliferation and tumor growth in human lung cancer cells both *in vitro* and in an *in vivo* murine model.⁵² This approach seems promising, and, thus, more thought needs to be put into its potential implementation for CCA treatment.

FGFR-targeted monoclonal antibodies

Monoclonal antibodies that specifically target FGFRs or FGFs help in reducing common adverse effects associated with inhibiting multiple

FGFR subtypes; for instance, FGFR2 and FGFR3 can be suppressed without compromising adult tissue homeostasis, whereas, since FGFR1 and FGFR4 are closely related, blocking them may lead to metabolic disturbances that have greater health risks.

Vofatamab (B-701) is a monoclonal antibody against mutated FGFR3. It was well-tolerated in a phase Ib/II trial [ClinicalTrials.gov identifier: NCT02401542], and was the first FGF/FGFR inhibitor to qualify for the FDA's fast-track designation in advanced or metastatic urothelial cell carcinoma.⁵³

In addition, studies have found that patients with activated/amplified FGFR2 signaling could benefit from GP369 (FGFR2-IIIb-specific monoclonal antibody) and R3Mab (FGFR3-specific monoclonal antibody).^{11,54}

MFGR1877S, a monoclonal antibody against FGFR3, was clinically evaluated [ClinicalTrials. gov identifier: NCT01363024] in patients with advanced solid tumors. The phaseI dose-escalation trial included 24 participants, but no results have been announced as of yet. More clinical trial data on FGFR-targeted monoclonal antibodies in CCA therapy are needed to accurately assess their effectiveness.

Disadvantages of current therapies

Resistance to FGFR inhibitions

Multiple nonspecific first-generation FGFR inhibitors are in full swing in CCA clinical trials and have demonstrated non-ideal clinical responses; the median duration of response was found to be only 5–6 months.⁵⁵ However, according to research data, second-generation FGFR inhibitors seemingly overcame FGFR-driven lack of specificity in CCA and have improved FGFR activity.⁴⁵ Nonetheless, detection of secondary FGFR2 mutations that may confer resistance is imperative in CCA cases.

As we know, tumors are the result of a variety of genetic lesions; clinicians and researchers should not underestimate the ability of tumors to adapt to new stress conditions and resist anti-cancer drugs. Resistance mechanisms to overcome FGFR inhibition are driven by mutation activation or signaling pathway bypassing.

By analyzing pretreatment and post-progression cell-free circulating tumor DNA (cfDNA) in three advanced CCA patients after BGJ398 treatment, Goyal *et al.* found that FGFR2 point mutations (p.N549H, p.N549K, p.V564F, p.E565A, and p.K659M, p.N549H, p.E565A, p.L617V, and p.K641R) could lead to clinical resistance to FGF/FGFR inhibitions in CCA.⁵⁶ However, p.V564F was the only common mutation among the three patients, further highlighting its important role in FGFR2 progression.

Herrera-Abreu *et al.* used parallel RNA interference genetic screens to show that EGFR plays a role in reducing FGFR-inhibition sensitivity in *FGFR3*-mutant and *FGFR3*-translocated cell lines through a negative feedback loop.⁵⁷ Besides, activation of members of the EGFR family was also evident in triple-negative breast cancer as a mechanism to resist mutant-cancer gene inhibitors.⁵⁸ Moreover, much research has reported that FGFR3 fusions and c-MET may also be involved in resistance to FGFR inhibitors.^{47,59} Hence, the EGFR family, FGFR3, and c-MET need to be further investigated for possible roles in FGFR-inhibitor resistance in CCA.

Toxicity and side effects of FGFR inhibitors

Unlike VEGFR inhibitors, effective doses of FGFR inhibitors do not induce elevated blood pressure or proteinuria; their most common and severe toxicities include hyperphosphatemia, tissue calcification, and so on.^{33,60}

Increased FGF23 expression is known to be regulated by high serum phosphate and 1,25 (OH)₂D levels; FGF23 can down-regulate renal phosphate reabsorption and increase the effect of PTH to upregulate renal phosphate excretion by reducing the sodium phosphate co-transporters (NaPi-IIa and NaPi-IIc) in proximal tubules, whereas FGFR inhibitors induced the down-regulation of the FGF23-Klotho-FGFR1 complex, thereby leading to hypocalcemia and tissue calcification.⁶¹ Besides, hyperphosphatemia and tissue calcification are correlated with a higher cardiovascular mortality in patients with chronic renal failure, cardiomyocyte and vascular damage, and cardiovascular abnormalities such as impaired contractility, reduced cardiac output (CO), and arrhythmia.62,63

Bétrian *et al.* have reported two cases where patients developed severe straightening of scalp

Table 2. Common toxicities an	d side effects of FGFR inhibitors.
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Drugs	Tumors	Subject numbers	Phase	Adverse events	Severe adverse events
Rogaratinib ⁶⁵	Urothelial cancer, HNSCC, NSCLC, other tumor types	126	I	Hyperphosphatemia, diarrhea, decreased appetite, fatigue, and asymptomatic increased lipase	Decreased appetite and diarrhea, acute kidney injury, hypoglycemia, retinopathy, and vomiting
Debio 1347 ⁶⁶	Breast and biliary duct cancer	58	I	Dry mouth/eyes, hyperamylasemia, hypercalcemia, hyperbilirubinemia, hyperphosphatemia, and stomatitis	ΝΑ
BGJ398, Imatinib ⁶⁷	Gastrointestinal stromal tumor	16	1/11	CPK elevation, lipase elevation, hyperphosphatemia, anemia, and peripheral edema	No
Brivanib ⁶⁸	Advanced/metastatic solid tumors	68	I	Nausea, pyrexia, AST or ALT elevations, and thrombocytopenia	NA
ARQ 08769	Advanced solid tumors	80	I	Fatigue, nausea, AST increase, and diarrhea, hyperphosphatemia	ΝΑ
Erdafitinib ⁶⁰	Advanced or refractory solid tumors	6	I	Hyperphosphatemia, nausea, stomatitis, dysgeusia, and dry mouth	No
LY287445570	Gastric cancer and NSCLC	24	I	Hyperphosphatemia, diarrhea, and stomatitis	No

ALT, alanine aminotransferase; AST, aspartate aminotransferase; CPK, *creatine phosphokinase*; FGFR, fibroblast growth factor receptor; HNSCC, head and neck squamous cell carcinoma; NA, not available; NSCLC, non-small-cell lung carcinoma.

hair, eyelash trichomegaly, acral desquamation, xerostomia, and heel hyperkeratosis following the use of new selective pan-FGFR inhibitors.⁶⁴ We looked up clinical trials related to FGFR inhibitors to find out the corresponding treatmentrelated side effects. Table 2 shows some common FGFR-inhibitor related adverse events, such as hyperphosphatemia, diarrhea, decreased appetite, fatigue, and liver dysfunction. Severe adverse events were rare; however, it is likely that most of the results came from phase I trials and that the number of clinical trials was too small.

In view of the severity of these side effects, the application of FGFR inhibitors may have limits, especially in cancer patients with heart and kidney insufficiency; therefore, developing appropriate side-effect avoidance protocols and/or antagonistic side-effect drugs will be a key point in FGFRinhibitor promotion.

Conclusion and future perspective

In recent years, genome sequencing has provided a useful approach for cancer diagnosis; FGFR gene

mutations were found to be associated with the development of multiple tumors, included CCA. FGF/FGFR inhibitors have been studied extensively as a targeted therapy for CCA. Moreover, the current results have suggested that targeting the FGF/FGFR signaling pathway is promising in CCA.

However, a number of conceptually important questions still remain un-answered; several physiological and pathological processes contribute to the occurrence of CCAs, in which blood and lymphatic angiogenesis processes are also tightly regulated by several key angiogenic factors. The FGF, PDGF, VEGF, and angiopoietin families exert a synergistic effect on tumor blood and lymphatic vessels even though they have different effects on vascular maturation and function, respectively. However, when and how chromosome aberrations occur and the genetic point-of-no-return in CCA remain a mystery. In addition, knowing whether using FGF/VEGF/PDGF/IDH inhibitors in concert (combinatorial or sequential strategies) will provide added benefits for CCA patients, compared with those from their individual uses, is

essential information for clinical practice. We believe all of this will facilitate the development of FGFR in the field of CCA.

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Conflict of interest statement

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References

- Tyson GL and El-Serag HB. Risk factors for cholangiocarcinoma. *Hepatology* 2011; 54: 173–184.
- Sriamporn S, Pisani P, Pipitgool V, et al. Prevalence of *Opisthorchis viverrini* infection and incidence of cholangiocarcinoma in Khon Kaen, Northeast Thailand. *Trop Med Int Health* 2004; 9: 588–594.
- Marcano-Bonilla L, Mohamed EA, Mounajjed T, et al. Biliary tract cancers: epidemiology, molecular pathogenesis and genetic risk associations. *Chin Clin Oncol* 2016; 5: 61.
- Goeppert B. Biliary tract cancers: molecular characterization and identification of novel prognostic markers. *Pathologe* 2017; 38(Suppl. 2): 192–197.
- Nakeeb A, Pitt HA, Sohn TA, et al. Cholangiocarcinoma. A spectrum of intrahepatic, perihilar, and distal tumors. Ann Surg 1996; 224: 463–475.
- 6. Jang S, Chun SM, Hong SM, *et al.* High throughput molecular profiling reveals differential mutation patterns in intrahepatic cholangiocarcinomas arising in chronic advanced liver diseases. *Mod Pathol* 2014; 27: 731–739.

- Nakamura H, Arai Y, Totoki Y, *et al.* Genomic spectra of biliary tract cancer. *Nat Genet* 2015; 47: 1003–1010.
- Rizvi S, Khan SA, Hallemeier CL, et al. Cholangiocarcinoma—evolving concepts and therapeutic strategies. Nat Rev Clin Oncol 2018; 15: 95.
- Rogers JE, Law L, Nguyen VD, et al. Second-line systemic treatment for advanced cholangiocarcinoma. J Gastrointest Oncol 2014; 5: 408–413.
- Greenman C, Stephens P, Smith R, et al. Patterns of somatic mutation in human cancer genomes. *Nature* 2007; 446: 153–158.
- 11. Yin Y, Djakovic S, Marsters S, *et al.* Redesigning a monospecific anti-FGFR3 antibody to add selectivity for FGFR2 and expand antitumor activity. *Mol Cancer Ther* 2015; 14: 2270–2278.
- 12. Ornitz DM and Itoh N. The fibroblast growth factor signaling pathway. *Wiley Interdiscip Rev Dev Biol* 2015; 4: 215–266.
- 13. Katoh M. FGFR inhibitors: effects on cancer cells, tumor microenvironment and whole-body homeostasis. *Int J Mol Med* 2016; 38: 3–15.
- Zhou Y, Wu C, Lu G, *et al.* FGF/FGFR signaling pathway involved resistance in various cancer types. *J Cancer* 2020; 11: 2000–2007.
- Babina IS and Turner NC. Advances and challenges in targeting FGFR signalling in cancer. *Nat Rev Cancer* 2017; 17: 318–332.
- Jusakul A, Cutcutache I, Yong CH, et al. Whole-genome and epigenomic landscapes of etiologically distinct subtypes of cholangiocarcinoma. *Cancer Discov* 2017; 7: 1116–1135.
- Touat M, Ileana E, Postel-Vinay S, et al. Targeting FGFR signaling in cancer. Clin Cancer Res 2015; 21: 2684–2694.
- Narong S and Leelawat K. Basic fibroblast growth factor induces cholangiocarcinoma cell migration via activation of the MEK1/2 pathway. *Oncol Lett* 2011; 2: 821–825.
- Pardo OE, Latigo J, Jeffery RE, et al. The fibroblast growth factor receptor inhibitor PD173074 blocks small cell lung cancer growth in vitro and in vivo. *Cancer Res* 2009; 69: 8645–8651.
- Collin MP, Lobell M, Hübsch W, et al. Discovery of Rogaratinib (BAY 1163877): a pan-FGFR inhibitor. *ChemMedChem* 2018; 13: 437–445.
- 21. Yen TT, Thao DT and Thuoc TL. An overview on keratinocyte growth factor: from the molecular

properties to clinical applications. *Protein Pept Lett* 2014; 21: 306–317.

- 22. Fischbach A, Rogler A, Erber R, *et al.* Fibroblast growth factor receptor (FGFR) gene amplifications are rare events in bladder cancer. *Histopathology* 2015; 66: 639–649.
- 23. Helsten T, Elkin S, Arthur E, *et al.* The FGFR landscape in cancer: analysis of 4,853 tumors by next-generation sequencing. *Clin Cancer Res* 2016; 22: 259–267.
- 24. Xue WJ, Li MT, Chen L, *et al.* Recent developments and advances of FGFR as a potential target in cancer. *Future Med Chem* 2018; 10: 2109–2126.
- Jain A, Kwong LN and Javle M. Genomic profiling of biliary tract cancers and implications for clinical practice. *Curr Treat Options Oncol* 2016; 17: 58.
- 26. Chun YS and Javle M. Systemic and adjuvant therapies for intrahepatic cholangiocarcinoma. *Cancer Control* 2017; 24: 1073274817729241.
- 27. Ross JS, Wang K, Gay L, *et al.* New routes to targeted therapy of intrahepatic cholangiocarcinomas revealed by next-generation sequencing. *Oncologist* 2014; 19: 235–242.
- Lee H, Wang K, Johnson A, *et al.* Comprehensive genomic profiling of extrahepatic cholangiocarcinoma reveals a long tail of therapeutic targets. *J Clin Pathol* 2016; 69: 403–408.
- Raggi C, Fiaccadori K, Pastore M, *et al.* Antitumor activity of a novel fibroblast growth factor receptor inhibitor for intrahepatic cholangiocarcinoma. *Am J Pathol* 2019; 189: 2090–2101.
- Lombardi P, Marino D, Fenocchio E, et al. Emerging molecular target antagonists for the treatment of biliary tract cancer. Expert Opin Emerg Drugs 2018; 23: 63–75.
- Pu XH, Ye Q, Yang J, *et al.* Low-level clonal FGFR2 amplification defines a unique molecular subtype of intrahepatic cholangiocarcinoma in a Chinese population. *Hum Pathol* 2018; 76: 100–109.
- 32. Wu YM, Su F, Kalyana-Sundaram S, *et al.* Identification of targetable FGFR gene fusions in diverse cancers. *Cancer Discov* 2013; 3: 636–647.
- Javle M, Lowery M, Shroff RT, et al. Phase II study of BGJ398 in patients with FGFR-altered advanced cholangiocarcinoma. J Clin Oncol 2018; 36: 276–282.
- 34. Tabernero J, Bahleda R, Dienstmann R, et al. Phase I dose-escalation study of JNJ-42756493, an oral pan-fibroblast growth factor receptor inhibitor, in patients with advanced solid tumors. J Clin Oncol 2015; 33: 3401–3408.

- 35. Hall TG, Yu Y, Eathiraj S, *et al.* Preclinical activity of ARQ 087, a novel inhibitor targeting FGFR dysregulation. *PLoS One* 2016; 11: e0162594.
- Poupon R. Ursodeoxycholic acid and bile-acid mimetics as therapeutic agents for cholestatic liver diseases: an overview of their mechanisms of action. *Clin Res Hepatol Gastroenterol* 2012; 36(Suppl. 1): S3–S12.
- Sinha J, Chen F, Miloh T, et al. β-Klotho and FGF-15/19 inhibit the apical sodium-dependent bile acid transporter in enterocytes and cholangiocytes. Am J Physiol Gastrointest Liver Physiol 2008; 295: G996–G1003.
- Xu YF, Yang XQ, Lu XF, et al. Fibroblast growth factor receptor 4 promotes progression and correlates to poor prognosis in cholangiocarcinoma. Biochem Biophys Res Commun 2014; 446: 54–60.
- 39. Yoo C, Kang J, Kim D, *et al.* Multiplexed gene expression profiling identifies the FGFR4 pathway as a novel biomarker in intrahepatic cholangiocarcinoma. *Oncotarget* 2017; 8: 38592–38601.
- 40. De Luca A, Frezzetti D, Gallo M, *et al.* FGFRtargeted therapeutics for the treatment of breast cancer. *Expert Opin Invest Drugs* 2017; 26: 303–311.
- 41. Dienstmann R, Rodon J, Prat A, *et al.* Genomic aberrations in the FGFR pathway: opportunities for targeted therapies in solid tumors. *Ann Oncol* 2014; 25: 552–563.
- 42. Abdel-Rahman O. Targeting FGF receptors in colorectal cancer: from bench side to bed side. *Future Oncol* 2015; 11: 1373–1379.
- 43. Naing A. A new screening tool for FGFR inhibitor treatment? *Lancet Oncol* 2019; 20: 1340–1342.
- 44. Ochiiwa H, Fujita H, Itoh K, *et al.* TAS-120, a highly potent and selective irreversible FGFR inhibitor, is effective in tumors harboring various FGFR gene abnormalities. *Mol Cancer Ther* 2013; 12S.
- 45. Goyal L, Shi L, Liu LY, *et al.* TAS-120 overcomes resistance to ATP-competitive FGFR inhibitors in patients with FGFR2 fusion-positive intrahepatic cholangiocarcinoma. *Cancer Discov* 2019; 9: 1064–1079.
- 46. Abou-Alfa GK, Sahai V, Hollebecque A, et al. Pemigatinib for previously treated, locally advanced or metastatic cholangiocarcinoma: a multicentre, open-label, phase 2 study. Lancet Oncol 2020; 21: 671–684.
- 47. Hallinan N, Finn S, Cuffe S, *et al.* Targeting the fibroblast growth factor receptor family in cancer. *Cancer Treat Rev* 2016; 46: 51–62.

- Harding TC, Long L, Palencia S, *et al.* Blockade of nonhormonal fibroblast growth factors by FP-1039 inhibits growth of multiple types of cancer. *Sci Transl Med* 2013; 5: 178ra39.
- Tolcher AW, Papadopoulos KP, Patnaik A, et al. A phase I, first in human study of FP-1039 (GSK3052230), a novel FGF ligand trap, in patients with advanced solid tumors. *Ann Oncol* 2016; 27: 526–532.
- Morgensztern D, Karaseva N, Felip E, et al. An open-label phase IB study to evaluate GSK3052230 in combination with paclitaxel and carboplatin, or docetaxel, in FGFR1-amplified non-small cell lung cancer. Lung Cancer 2019; 136: 74–79.
- Ronca R, Giacomini A, Di Salle E, et al. Longpentraxin 3 derivative as a small-molecule FGF trap for cancer therapy. *Cancer Cell* 2015; 28: 225–239.
- 52. Castelli R, Giacomini A, Anselmi M, et al. Synthesis, structural elucidation, and biological evaluation of NSC12, an orally available fibroblast growth factor (FGF) ligand trap for the treatment of FGF-dependent lung tumors. J Med Chem 2016; 59: 4651–4663.
- Casadei C, Dizman N, Schepisi G, et al. Targeted therapies for advanced bladder cancer: new strategies with FGFR inhibitors. Ther Adv Med Oncol 2019; 11: 1758835919890285.
- Bai A, Meetze K, Vo NY, et al. GP369, an FGFR2-IIIb-specific antibody, exhibits potent antitumor activity against human cancers driven by activated FGFR2 signaling. *Cancer Res* 2010; 70: 7630–7639.
- 55. Mahipal A, Tella SH, Kommalapati A, et al. FGFR2 genomic aberrations: Achilles heel in the management of advanced cholangiocarcinoma. *Cancer Treat Rev* 2019; 78: 1–7.
- 56. Goyal L, Saha SK, Liu LY, et al. Polyclonal secondary FGFR2 mutations drive acquired resistance to FGFR inhibition in patients with FGFR2 fusion-positive cholangiocarcinoma. *Cancer Discov* 2017; 7: 252–263.
- Herrera-Abreu MT, Pearson A, Campbell J, et al. Parallel RNA interference screens identify EGFR activation as an escape mechanism in FGFR3-mutant cancer. *Cancer Discov* 2013; 3: 1058–1071.

58. Issa A, Gill JW, Heideman MR, et al. Combinatorial targeting of FGF and ErbB receptors blocks growth and metastatic spread of breast cancer models. *Breast Cancer Res* 2013; 15: R8.

- Stransky N, Cerami E, Schalm S, *et al.* The landscape of kinase fusions in cancer. *Nat Commun* 2014; 5: 4846.
- 60. Nishina T, Takahashi S, Iwasawa R, *et al.* Safety, pharmacokinetic, and pharmacodynamics of erdafitinib, a pan-fibroblast growth factor receptor (FGFR) tyrosine kinase inhibitor, in patients with advanced or refractory solid tumors. *Invest New Drugs* 2018; 36: 424–434.
- 61. Bergwitz C and Juppner H. Regulation of phosphate homeostasis by PTH, vitamin D, and FGF23. *Annu Rev Med* 2010; 61: 91–104.
- 62. Jono S, McKee MD, Murry CE, *et al.* Phosphate regulation of vascular smooth muscle cell calcification. *Circ Res* 2000; 87: E10–E17.
- 63. Yanochko GM, Vitsky A, Heyen JR, *et al.* Pan-FGFR inhibition leads to blockade of FGF23 signaling, soft tissue mineralization, and cardiovascular dysfunction. *Toxicol Sci* 2013; 135: 451–464.
- 64. Betrian S, Gomez-Roca C, Vigarios E, *et al.* Severe onycholysis and eyelash trichomegaly following use of new selective pan-FGFR inhibitors. *JAMA Dermatol* 2017; 153: 723–725.
- Schuler M, Cho BC, Sayehli CM, et al. Rogaratinib in patients with advanced cancers selected by FGFR mRNA expression: a phase 1 dose-escalation and dose-expansion study. *Lancet* Oncol 2019; 20: 1454–1466.
- 66. Voss MH, Hierro C, Heist RS, et al. A phase I, open-label, multicenter, dose-escalation study of the oral selective FGFR inhibitor Debio 1347 in patients with advanced solid tumors harboring FGFR gene alterations. *Clin Cancer Res* 2019; 25: 2699–2707.
- 67. Kelly CM, Shoushtari AN, Qin LX, *et al.* A phase Ib study of BGJ398, a pan-FGFR kinase inhibitor in combination with imatinib in patients with advanced gastrointestinal stromal tumor. *Invest New Drugs* 2019; 37: 282–290.
- 68. Jonker DJ, Rosen LS, Sawyer MB, et al. A phase I study to determine the safety, pharmacokinetics and pharmacodynamics of a dual VEGFR and FGFR inhibitor, brivanib, in patients with advanced or metastatic solid tumors. *Ann Oncol* 2011; 22: 1413–1419.
- 69. Papadopoulos KP, El-Rayes BF, Tolcher AW, et al. A phase 1 study of ARQ 087, an oral pan-FGFR inhibitor in patients with advanced solid tumours. Br J Cancer 2017; 117: 1592–1599.
- 70. Michael M, Bang YJ, Park YS, et al. A phase 1 study of LY2874455, an oral selective pan-FGFR inhibitor, in patients with advanced cancer. *Target Oncol* 2017; 12: 463–474.

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