

Bacteriological Spectrum and Antibiotic Susceptibility on Blood Culture in Newly Diagnosed Pediatric Patients With Acute Lymphoblastic Leukemia During the Induction Phase

Review began 05/16/2022

Review ended 05/26/2022

Published 05/30/2022

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Abstract

Background

Acute lymphoblastic leukemia (ALL) is the most common cancer diagnosed in children worldwide. This study was conducted to find out the trends in the bacteriological spectrum and antibiotic susceptibility on blood culture in newly diagnosed children with acute lymphoblastic leukemia during the induction phase at a pediatric oncology unit in South Punjab, Pakistan.

Methodology

This cross-sectional study was conducted from January 1, 2020, to June 30, 2021. A total of 263 newly diagnosed ALL cases of both genders aged up to 16 years were included. Adopting full aseptic measures, the blood samples of all children were sent for culture and sensitivity testing to the institutional laboratory immediately after collection on the eighth day of the induction phase in all children. Bacterial isolates and their sensitivity/resistance patterns were noted.

Results

Out of 263 children with ALL, 172 (65.4%) were males. Overall, the mean age was 7.4±3.4 years (ranging between 1 and 16 years). B-cell type was the commonest type noted in 204 (77.6%) children. Out of a total of 52 cases with positive blood culture findings for bacterial isolates, there were 28 (53.8%) cases with gram-negative bacterial isolates, while 24 (46.2%) were gram-positive bacterial isolates. *Escherichia coli* (*E. coli*) was the commonest type of gram-negative bacteria noted among 18/28 (64.3%) cases, while *Staphylococcus aureus* (*S. aureus*) was the most frequent gram-positive bacterial isolates in 13/24 (54.2%). We found meropenem, linezolid, clindamycin, piperacillin, tazobactam, and amikacin to have the highest antimicrobial sensitivities, while commonly adopted antibiotics such as ciprofloxacin, cefotaxime, cefoperazone, amoxiclav, and ampicillin were found to have high resistance rates.

Conclusion

Gram-negative bacterial isolates formed the majority of the positive blood culture cases. *Escherichia coli*, *Staphylococcus aureus*, and *Klebsiella pneumonia* (*K. pneumonia*) were the most common types of bacterial isolates. Routinely used antibiotics such as ciprofloxacin, cefotaxime, cefoperazone, and ampicillin were found to have high rates of resistance.

Categories: Pediatrics, Oncology

Keywords: staphylococcus aureus, e. coli, bacterial isolates, blood culture, acute lymphoblastic leukemia

Introduction

Acute lymphoblastic leukemia (ALL) is the most common cancer diagnosed in children worldwide. Although no population-level statistics are available from Pakistan, it is estimated that the annual incidence of ALL is 3.7-4.9 per 100,000 children in the United States [1]. Infection is considered to be the commonest cause of treatment-related morbidity and mortality in children with the diagnosis of cancer globally [2,3]. Among children with ALL, infections are noted to be one of the major reasons behind increased rates of mortality in developing countries. Compared to developed countries, infection-related mortality soars up to 10-fold among children with ALL in developing countries [4]. Timely identification and treatment of causative agents can significantly improve the survival rates among pediatric cancer patients [2]. Ward et al. who estimated global childhood cancer survival rates and priority settings revealed that supporting services and care including infection control have the highest survival gains [5].

How to cite this article

Fawad U (May 30, 2022) Bacteriological Spectrum and Antibiotic Susceptibility on Blood Culture in Newly Diagnosed Pediatric Patients With Acute Lymphoblastic Leukemia During the Induction Phase. Cureus 14(5): e25470. DOI 10.7759/cureus.25470

Geographical variations exist regarding infection-related mortality and outcomes even among different settings of the same geographical locations. These differences could be attributed to differences in the epidemiology of causative agents and variations in inaccessibility to resources thought to prevent, identify, and treat these infections [6]. Due to these differences, it is very difficult to implement a single approach to reduce morbidity and mortality among children with cancer. A study from India analyzing bacterial spectrum and sensitivity patterns of pathogens among patients of febrile neutropenic patients with hematological malignancies found *Escherichia coli* (*E. coli*) to be the commonest pathogen involved in 43% of culture-positive cases [7]. Another study from the United States evaluating children with newly diagnosed acute leukemia noted coagulase-negative staphylococci (CoNS) as the most commonly involved microorganism (12/29, 41.4% cases) [6].

In Pakistan, not much work is seen regarding the pattern of bacterial isolates and their antibiotic susceptibility among children with ALL. It was imperative to design a study on the subject to fill up the much-needed gap and provide useful information about the most common causative agents and their sensitivity/resistance patterns to most commonly used antibiotics. The present study was conducted to find out trends in the bacteriological spectrum and antibiotic susceptibility on blood culture in newly diagnosed pediatric patients with ALL during the induction phase at a pediatric oncology unit in Southern Punjab, Pakistan.

Materials And Methods

This cross-sectional study was conducted at the Department of Pediatric Oncology, The Children's Hospital and Institute of Child Health, Multan Pakistan, from January 1, 2020, to June 30, 2021. Approval from the Institutional Ethical Committee was acquired (approval number: CHC/IEC/20-148). Informed and written consent was sought from parents or guardians or children.

A total of 263 newly diagnosed ALL children of both genders aged up to 16 years were included, which gives a reasonable power to the study considering the rare nature of this disease. The induction phase of chemotherapy lasts 28 days (day 8 to day 35), starting from day 8 of therapy following the prophase. Adopting full aseptic measures, the blood samples of all children were sent for culture and sensitivity testing to the institutional laboratory immediately after collection on the eighth day of the induction phase in all children. Laboratory tests were done at the institutional laboratory. The blood sample was taken by a nurse or a medical technician and immediately sent to the laboratory following the standard protocols of using the blood collection tube. Blood culture was performed using the traditional method for bacterial identification. The blood sample (5 mL) was mixed with *Trypanosoma* broth with a 1:9 ratio and incubated at 37°C. Signs of bacterial growth were checked routinely. Blood agar, chocolate agar, and MacConkey agar were used for subculture. In the case of positive blood culture, bacterial identification was made using colony characteristics, the gram reaction of the organism, and biochemical tests as per standard guidelines. The isolation of one or more known bacteria was observed from blood culture. The isolated bacteria were *Escherichia coli* (*E. coli*), *Staphylococcus aureus* (*S. aureus*), *Klebsiella pneumoniae* (*K. pneumoniae*), *Pseudomonas aeruginosa* (*P. aeruginosa*), *Salmonella typhi* (*S. Typhi*), coagulase-negative staphylococci (CoNS), and *Enterococcus faecalis* (*E. faecalis*). An antimicrobial susceptibility test was performed on all blood cultures isolated by disk diffusion. About 3-5 colonies of bacteria were transformed into a tube containing 5 mL sterile normal saline and gently mixed with a homogeneous suspension. The incubated tubes were left to dry, and a set of the antibiotic disc was then placed. The commonly used disc was meropenem, amikacin, piperacillin-tazobactam, gentamycin, clindamycin, vancomycin, ceftriaxone, ciprofloxacin, imipenem, linezolid, cefoperazone, amoxiclav, amoxicillin, clarithromycin, cefotaxime, and levofloxacin. A standard treatment schedule was adopted and adhered to during the induction phase. A special proforma was formed to record all study data.

For data analysis, the Statistical Package for Social Sciences (SPSS) version 26.0 (IBM Corp., Armonk, NY, USA) was utilized. Quantitative data such as age were represented as means and standard deviations (SD). Qualitative data such as gender, area of residence, types of ALL, bacterial isolates, and their sensitivity/resistance patterns were shown as frequencies and percentages.

Results

In a total of 263 newly diagnosed ALL cases, there were 172 (65.4%) males. Overall, the mean age was 7.4±3.4 years (ranging between 1 and 16 years), while 116 (44.1%) children were aged above 5-10 years. The majority of the children (147 (55.9%)) belonged to urban areas. B-cell type was the commonest type of ALL noted among 204 (77.6%) children. Blood culture studies reported positive findings for bacterial isolates in 52 (19.8%) children, while seven (2.7%) had an existence of fungus. Table 1 shows the characteristics of all children.

| Characteristics | | Number (%) |
|----------------------------|--------------------|-------------|
| Gender | Male | 172 (65.4%) |
| | Female | 91 (34.6%) |
| Age (Years) | <5 | 93 (35.4%) |
| | >5 to 10 | 116 (44.1%) |
| | >10 | 54 (20.5%) |
| Area of Residence | Urban | 147 (55.9%) |
| | Rural | 116 (44.1%) |
| ALL Types | B-Cell | 204 (77.6%) |
| | T-Cell | 52 (19.8%) |
| | Mixed Type | 7 (2.7%) |
| Major Symptoms | Fever | 72 (27.4%) |
| | Cough | 51 (19.4%) |
| | Dyspnea | 38 (14.4%) |
| | Diarrhea | 26 (9.9%) |
| Blood Culture for Isolates | Bacterial Isolates | 52 (19.8%) |
| | Fungus | 7 (2.7%) |
| | Negative | 204 (77.6%) |

TABLE 1: Characteristics of Children With Acute Lymphoblastic Leukemia

Out of a total of 52 cases with positive blood culture findings for bacterial isolates, there were 28 (53.8%) gram-negative bacterial isolates, while 24 (46.2%) were gram-positive bacterial isolates. *Escherichia coli* was the commonest type of gram-negative bacteria noted among 18/28 (64.3%) cases, while *Staphylococcus aureus* was the most frequent gram-positive bacterial isolates found in 13/24 (54.2%). Table 2 shows the antibiotic sensitivity and resistance patterns concerning bacterial isolates among children with ALL.

| Bacterial Isolate | Most Sensitive Antibiotics | Sensitivity (%) | Most Resistant Antibiotics | Resistance (%) |
|--|----------------------------|-----------------|----------------------------|----------------|
| <i>Escherichia coli</i> (<i>E. coli</i>) (n=18) | Meropenem | 100 | Ampicillin | 89 |
| | Piperacillin Tazobactam | 94 | Cefoperazone | 83 |
| | Amikacin | 89 | Amoxiclav | 83 |
| <i>Staphylococcus aureus</i> (<i>S. aureus</i>) (n=13) | Clindamycin | 100 | Amoxicillin | 69 |
| | Gentamycin | 92 | Ciprofloxacin | 69 |
| | Vancomycin | 85 | Ampicillin | 62 |
| <i>Klebsiella pneumoniae</i> (<i>K. pneumoniae</i>) (n=9) | Meropenem | 89 | Clarithromycin | 67 |
| | Ceftriaxone | 89 | Amoxiclav | 67 |
| | Gentamycin | 78 | Cefotaxime | 67 |
| <i>Pseudomonas aeruginosa</i> (<i>P. aeruginosa</i>) (n=5) | Ceftriaxone | 80 | Gentamycin | 60 |
| | Ciprofloxacin | 80 | Ampicillin | 60 |
| | Amikacin | 60 | Piperacillin Tazobactam | 40 |
| <i>Salmonella typhi</i> (<i>S. typhi</i>) (n=3) | Meropenem | 100 | Ampicillin | 67 |
| | Imipenem | 100 | Levofloxacin | 67 |
| | Ceftriaxone | 67 | Ciprofloxacin | 67 |
| Coagulase-negative staphylococci (CoNS) (n=2) | Amikacin | 100 | Ceftriaxone | 50 |
| | Meropenem | 100 | Gentamycin | 50 |
| | Piperacillin Tazobactam | 50 | Ciprofloxacin | 50 |
| <i>Enterococcus faecalis</i> (<i>E. faecalis</i>) (n=2) | Vancomycin | 100 | Ciprofloxacin | 100 |
| | Linezolid | 100 | Gentamycin | 100 |
| | Clindamycin | 50 | Ampicillin | 50 |

TABLE 2: Antibiotic Sensitivity and Resistance Patterns With Respect to Bacterial Isolates Among Children With Acute Lymphoblastic Leukemia

Discussion

It is vital to periodically determine the most commonly involved organisms and their antimicrobial sensitivities in a particular population to enhance the clinical approach and treatment among children with ALL [8-10]. The present study is the first of its kind to analyze trends in the bacteriological spectrum and antibiotic susceptibility on blood culture in newly diagnosed pediatric patients with ALL during the induction phase at a pediatric oncology unit in South Punjab, Pakistan. We noted that the majority of the children with newly diagnosed ALL were male. A study by Burns et al. from the United States noted that 51.5% of children with newly diagnosed acute leukemia are male [6]. Local data from Lahore analyzing children with ALL observed that 79.2% of cases were male [11]. Another study from Karachi found that 66% of ALL cases were male [8]. In this study, we noted that B-cell was the most common type of ALL as noted among 77.6% of ALL cases. B-cell ALL is the most common type of ALL, accounting for more than 70% of ALL cases; therefore, our findings were consistent with what has been reported in the literature [11].

In the present study, blood culture studies reported positive findings for bacterial isolates in 19.8% of children. The proportion of children with positive blood culture findings in the present study was less than what was reported in a recent study conducted by Rajeswari et al. who analyzed infections during the induction phase of children with ALL, where they found that 30.6% of the children have microbiologically confirmed infection [12]. Gram-negative bacterial isolates were noted in 53.8% of positive cases. Data from India in a recent study revealed that 80% of ALL cases with culture-positive findings for bacterial isolates have gram-negative organisms [12]. A study from China also revealed that 59.9% of cases with positive blood culture findings have a gram-negative bacterial presence [13].

We also noted that *E. coli* (gram-negative) was the most common bacterial isolate present in 34.6% of newly diagnosed ALL cases, followed by *Staphylococcus aureus* (gram-positive), which was found in 25% of cases. A study from Qatar by El-Mahallawy et al. observed that among 268 children with microbiologically confirmed bloodstream bacterial infections, coagulase-negative staphylococci were the commonest bacterial isolates noted in 19.8% of cases, followed by *Staphylococcus aureus* (16.4%) and *Acinetobacter* spp. (8.2%) [14]. Data from China found *Pseudomonas aeruginosa* to be the most common bacterial isolates noted in 11.6% of acute leukemia cases [13]. All these data further emphasize that significant differences exist in the distribution of the most commonly involved etiological agents among children with ALL, so frequent analysis of the most commonly involved pathogens must be conducted to obtain first-hand knowledge about the trends of the most prevalent microorganisms involved [15-18].

We found meropenem, linezolid, clindamycin, piperacillin, tazobactam, and amikacin to have the highest antimicrobial sensitivities, while commonly adopted antibiotics such as ciprofloxacin, cefotaxime, cefoperazone, amoxiclav, and ampicillin were found to have high resistance rates. Ampicillin, cefotaxime, and amoxiclav are commonly used antibiotics, but we noted high rates of resistance, which demands a reevaluation of these treatment options. A study conducted in Bangladesh observed that ciprofloxacin was effective against all bacteria, and it helped in reducing infections [17]. However, in the present study, we found that ciprofloxacin had the highest resistance to *S. aureus*, coagulase-negative staphylococci, and *E. faecalis* and maximum sensitivity to *P. aeruginosa*. In our study, ampicillin had resistance to nearly all bacteria excluding *K. pneumoniae*. In addition to ampicillin and ciprofloxacin, amoxicillin also had resistance to *S. aureus*. A study conducted in Pakistan by Irfan et al. observed that no resistance was found for imipenem and meropenem [19]. In our current study, we observed that vancomycin had the highest antimicrobial sensitivity against *E. faecalis*. Effective control of infections is directly linked to the survival of children with acute leukemia, so our findings reinforce that antibiotics should be used judiciously adopting the right dosage and duration while there is a need to revise local protocols for the treatment considering a change in the pattern of commonly present causative agents [20-23]. Some researchers have reported other factors such as prolonged antibiotic exposure, previous hospitalization, and prophylactic antibiotic usage for the development of antibiotic resistance [24-27].

Our study had some limitations as well. As this was a single-center study, our findings cannot be generalized. We were unable to identify risk factors associated with bacterial infections. Being a cross-sectional study, we could not document the outcome and follow-up data in the current set of patients, which would have given us valuable insight.

Conclusions

Gram-negative bacterial isolates formed the majority of the positive blood culture cases. *Escherichia coli*, *Staphylococcus aureus*, and *Klebsiella pneumonia* were the most common types of bacterial isolates. Routinely used antibiotics such as ciprofloxacin, cefotaxime, cefoperazone, and ampicillin were found to have high rates of resistance.

There is a need for continuous surveillance of the spectrum of locally prevalent pathogens and their susceptibility patterns that can surely help in formulating therapeutic options among newly diagnosed ALL patients.

Additional Information

Disclosures

Human subjects: Consent was obtained or waived by all participants in this study. The Ethical Committee of The Children's Hospital and Institute of Child Health, Multan, Pakistan, issued approval CHC/IEC/20-148.

Animal subjects: All authors have confirmed that this study did not involve animal subjects or tissue.

Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

Acknowledgements

We are thankful to M. Aamir (RESnTEC, Bahawalpur) for his assistance in this research.

References

- Ribera JM, Oriol A: Acute lymphoblastic leukemia in adolescents and young adults. *Hematol Oncol Clin North Am.* 2009, 23:1033-42, vi. [10.1016/j.hoc.2009.07.002](https://doi.org/10.1016/j.hoc.2009.07.002)
- Inaba H, Pei D, Wolf J, et al.: Infection-related complications during treatment for childhood acute lymphoblastic leukemia. *Ann Oncol.* 2017, 28:386-92. [10.1093/annonc/mdw557](https://doi.org/10.1093/annonc/mdw557)
- Hwee J, Sutradhar R, Kwong JC, Sung L, Cheng S, Pole JD: Infections and the development of childhood

- acute lymphoblastic leukemia: a population-based study. *Eur J Cancer Prev.* 2020, 29:538-45. [10.1097/CEJ.0000000000000564](https://doi.org/10.1097/CEJ.0000000000000564)
4. Ceppi F, Antillon F, Pacheco C, Sullivan CE, Lam CG, Howard SC, Conter V: Supportive medical care for children with acute lymphoblastic leukemia in low- and middle-income countries. *Expert Rev Hematol.* 2015, 8:613-26. [10.1586/17474086.2015.1049594](https://doi.org/10.1586/17474086.2015.1049594)
 5. Ward ZJ, Yeh JM, Bhakta N, Frazier AL, Girardi F, Atun R: Global childhood cancer survival estimates and priority-setting: a simulation-based analysis. *Lancet Oncol.* 2019, 20:972-83. [10.1016/S1470-2045\(19\)30273-6](https://doi.org/10.1016/S1470-2045(19)30273-6)
 6. Burns JE, Reyes Pérez D, Li Y, et al.: Assessment of the impact of inpatient infectious events in pediatric patients with newly diagnosed acute leukemia at Dr. Robert Reid Cabral Children's Hospital, Dominican Republic. *PLoS One.* 2020, 15:e0243795. [10.1371/journal.pone.0243795](https://doi.org/10.1371/journal.pone.0243795)
 7. Burrows NR, Hora I, Geiss LS, Gregg EW, Albright A: US Department of Health and Human Services/Centers for Disease Control and Prevention: Incidence of end-stage renal disease attributed to diabetes among persons with diagnosed diabetes - United States and Puerto Rico, 2000-2014. *Morb Mortal Wkly Rep.* 2017, 66:1165-1205.
 8. Yasmeen N, Ashraf S: Childhood acute lymphoblastic leukaemia; epidemiology and clinicopathological features. *J Pak Med Assoc.* 2009, 59:150-5.
 9. Karanwal AB, Parikh BJ, Goswami P, Panchal HP, Parekh BB, Patel KB: Review of clinical profile and bacterial spectrum and sensitivity patterns of pathogens in febrile neutropenic patients in hematological malignancies: a retrospective analysis from a single center. *Indian J Med Paediatr Oncol.* 2013, 34:85-8. [10.4103/0971-5851.116184](https://doi.org/10.4103/0971-5851.116184)
 10. Castagnola E, Rossi MR, Cesaro S, et al.: Incidence of bacteremias and invasive mycoses in children with acute non-lymphoblastic leukemia: results from a multi-center Italian study. *Pediatr Blood Cancer.* 2010, 55:1103-7. [10.1002/psc.22750](https://doi.org/10.1002/psc.22750)
 11. Khan S, Anwar S, Latif MF, Farooq A, Faizan M: Induction-remission response in pediatric acute lymphoblastic leukaemia, Lahore protocol versus UKALL 2011 interim guidelines. *J Pak Med Assoc.* 2020, 70:591-6. [10.5455/JPMA.6586](https://doi.org/10.5455/JPMA.6586)
 12. Rajeswari B, Sukumaran Nair RK, Guruprasad CS, Nair M, Thankamony P, Parukutty K: Infections during induction chemotherapy in children with acute lymphoblastic leukemia - profile and outcomes: experience from a cancer center in South India. *Indian J Med Paediatr Oncol.* 2018, 39:188-92. [10.4103/ijmpo.ijmpo_95_17](https://doi.org/10.4103/ijmpo.ijmpo_95_17)
 13. Yao JF, Li N, Jiang J: Clinical characteristics of bloodstream infections in pediatric acute leukemia: a single-center experience with 231 patients. *Chin Med J (Engl).* 2017, 130:2076-81. [10.4103/0366-6999.213411](https://doi.org/10.4103/0366-6999.213411)
 14. El-Mahallawy H, Sidhom I, El-Din NH, Zamzam M, El-Lamie MM: Clinical and microbiologic determinants of serious bloodstream infections in Egyptian pediatric cancer patients: a one-year study. *Int J Infect Dis.* 2005, 9:43-51. [10.1016/j.ijid.2003.11.010](https://doi.org/10.1016/j.ijid.2003.11.010)
 15. Vaithiyam VS, Rastogi N, Ranjan P, et al.: Antimicrobial resistance patterns in clinically significant isolates from medical wards of a tertiary care hospital in North India. *J Lab Physicians.* 2020, 12:196-202. [10.1055/s-0040-1721161](https://doi.org/10.1055/s-0040-1721161)
 16. Ma J, Li N, Liu Y, et al.: Antimicrobial resistance patterns, clinical features, and risk factors for septic shock and death of nosocomial *E coli* bacteremia in adult patients with hematological disease: a monocenter retrospective study in China. *Medicine (Baltimore).* 2017, 96:e6959. [10.1097/MD.0000000000006959](https://doi.org/10.1097/MD.0000000000006959)
 17. Rahman MM, Khan MA: Levofloxacin prophylaxis to prevent bacterial infection in chemotherapy-induced neutropenia in acute leukemia. *Bangladesh Med Res Counc Bull.* 2009, 35:91-4. [10.3329/bmrcb.v35i3.4130](https://doi.org/10.3329/bmrcb.v35i3.4130)
 18. Abdollahi A, Hakimi F, Doomanlou M, Azadegan A: Microbial and antibiotic susceptibility profile among clinical samples of patients with acute leukemia. *Int J Hematol Oncol Stem Cell Res.* 2016, 10:61-9.
 19. Irfan S, Idrees F, Mehraj V, Habib F, Adil S, Hasan R: Emergence of Carbapenem resistant Gram negative and vancomycin resistant Gram positive organisms in bacteremic isolates of febrile neutropenic patients: a descriptive study. *BMC Infect Dis.* 2008, 8:80. [10.1186/1471-2334-8-80](https://doi.org/10.1186/1471-2334-8-80)
 20. Bo SN, Bo J, Ning YZ, et al.: Relationship between time to positivity of blood culture with clinical characteristics and hospital mortality in patients with *Escherichia coli* bacteremia. *Chin Med J (Engl).* 2011, 124:350-4. [10.3760/cma.j.issn.0366-6999.2011.03.002](https://doi.org/10.3760/cma.j.issn.0366-6999.2011.03.002)
 21. Aberuyi N, Rahgozar S, Ghodousi ES, Ghaedi K: Drug resistance biomarkers and their clinical applications in childhood acute lymphoblastic leukemia. *Front Oncol.* 2019, 9:1496. [10.3389/fonc.2019.01496](https://doi.org/10.3389/fonc.2019.01496)
 22. Patini R, Mangino G, Martellacci L, Quaranta G, Masucci L, Gallenzi P: The effect of different antibiotic regimens on bacterial resistance: a systematic review. *Antibiotics (Basel).* 2020, 9:10.3390/antibiotics9010022
 23. Thamlikitkul V, Rattanaumpawan P, Boonyasiri A, et al.: Thailand antimicrobial resistance containment and prevention program. *J Glob Antimicrob Resist.* 2015, 3:290-4. [10.1016/j.jgar.2015.09.003](https://doi.org/10.1016/j.jgar.2015.09.003)
 24. Tedim AP, Ruíz-Garbajosa P, Rodríguez MC, et al.: Long-term clonal dynamics of *Enterococcus faecium* strains causing bloodstream infections (1995-2015) in Spain. *J Antimicrob Chemother.* 2017, 72:48-55. [10.1093/jac/dkw366](https://doi.org/10.1093/jac/dkw366)
 25. Sánchez-Díaz AM, Cuartero C, Rodríguez JD, et al.: The rise of ampicillin-resistant *Enterococcus faecium* high-risk clones as a frequent intestinal colonizer in oncohaematological neutropenic patients on levofloxacin prophylaxis: a risk for bacteraemia?. *Clin Microbiol Infect.* 2016, 22:59.e1-8. [10.1016/j.cmi.2015.08.008](https://doi.org/10.1016/j.cmi.2015.08.008)
 26. Alatorre-Fernández P, Mayoral-Terán C, Velázquez-Acosta C, et al.: A polyclonal outbreak of bloodstream infections by *Enterococcus faecium* in patients with hematologic malignancies. *Am J Infect Control.* 2017, 45:260-6. [10.1016/j.ajic.2016.10.002](https://doi.org/10.1016/j.ajic.2016.10.002)
 27. Nanayakkara AK, Boucher HW, Fowler VG Jr, Jezek A, Outtersson K, Greenberg DE: Antibiotic resistance in the patient with cancer: escalating challenges and paths forward. *CA Cancer J Clin.* 2021, 71:488-504. [10.3322/caac.21697](https://doi.org/10.3322/caac.21697)