



Review

# Bioactive Carbohydrate Polymers—Between Myth and Reality

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Abstract: Polysaccharides are complex macromolecules long regarded as energetic storage resources or as components of plant and fungal cell walls. They have also been described as plant mucilages or microbial exopolysaccharides. The development of glycosciences has led to a partial and difficult deciphering of their other biological functions in living organisms. The objectives of glycobiochemistry and glycobiology are currently to correlate some structural features of polysaccharides with some biological responses in the producing organisms or in another one. In this context, the literature focusing on bioactive polysaccharides has increased exponentially during the last two decades, being sometimes very optimistic for some new applications of bioactive polysaccharides, notably in the medical field. Therefore, this review aims to examine bioactive polysaccharide, taking a critical look of the different biological activities reported by authors and the reality of the market. It focuses also on the chemical, biochemical, enzymatic, and physical modifications of these biopolymers to optimize their potential as bioactive agents.

Keywords: polysaccharide; oligosaccharide; bioactive agent; macromolecules



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# 1. Introduction—What Is a Bioactive Polysaccharide?

Polysaccharides are biopolymers of carbohydrates commonly found in all living organisms. Polysaccharides are probably among the more complex macromolecules existing in nature. As proteins or nucleic acids, they are biopolymers; however, contrary to them including, respectively, in their structures 4 nucleotides (nucleic acids) or 20 amino acids (proteins), up to 40–50 monosaccharides (mainly pentoses and hexoses) have been detected in polysaccharides. These pentoses and hexose have the ability to link each other by glycosidic bonds between the anomeric hydroxyl of one monosaccharide with any of the other ones from another unit. Therefore, with the sole example of the assembly of two monosaccharides such as glucose and galactose, 12 putative glycosidic bonds are potentially possible, leading to various disaccharides. Most polysaccharides found in nature occur as medium or high molecular weight biopolymers, including in their structure's numerous monosaccharides. Also called glycans, they differ by the length of their chains, the monosaccharides units composing them, and also the type of glycosiding linkages in

Molecules **2021**, 26, 7068 2 of 36

their structures and the degree of branching leading to linear or ramified polysaccharides. Homopolysaccharide contain only a single type of repeating unit, whereas heteropolysaccharides contain two or more different monosaccharide kinds. This high level of variability is reinforced by non-osidic chemical groups grafting the polysaccharidic backbone such as short organic acids (acetyl, pyruvyl, or succinyl groups) or sulfuric acid half ester (mainly in marine polysaccharides) [1–3]. The physiological functions of polysaccharides are diverse and are acquired after forming specific conformations. They are also strongly dependent on linkage modes functions and properties appearing accordingly. Some of them are carbon and energy storage macromolecules such as starch, glycogen, or laminaran. Others are structural elements, as in the case of the chitin of exoskeletons of some animals and fungi or cellulose in plants. More recently, some polysaccharides have been also identified as bioactive compounds and called bioactive polysaccharides. This definition refers to polysaccharides having biological effects on living organisms. Indeed, these biopolymers participate in many biological processes, notably cellular communication. Glycosciences have focused more and more on the identification of the native biological functions of some bioactive polysaccharides, as well as on the use and screening of others with no natural functions, with the objective of bringing them to market. This review detailed the natural or modified bioactive polysaccharides showing nutraceutic, therapeutic or toxic actions on microorganisms, animals, and plants. These biological activities are strongly affected by the chemical structures of polysaccharides, and the reviews are the state-of-the-art of recent progress in their functionalization of using physic, chemistry, or biochemistry, and aim to correlate some biological activities to structural features. Moreover, in front of the exponential increase in articles, for book chapters and articles dealing with new bioactive polysaccharide with promising industrial applications in various fields, notably in medicine, a special attention is given to the market reality. However, even if some polysaccharides have already found various applications as bioactive compounds in the therapeutic fields as an anticoagulant agent (heparin); for instance, hydrogels (hyaluronic acid), or in vaccines notably after conjugation with a protein carrier, have their development limited to niche markets such as those of nutraceutic (prebiotics and synbiotics) and cosmetics. However, exploration in other fields such those of plant elicitation and biostimulation, wound healing, drug delivery, or tissue engineering using notably 3D printing could exist in the future. The recent exploration of a new potential source of polysaccharides such as those produced by microalgae opens the way to a new structure of high potential.

# 2. Structure-Function Relationships

Polysaccharides perform essential biological functions in the human body, such as antioxidant, immunomodulatory, antitumor, etc., [2,4–12]. However, these properties are strongly correlated with the carbohydrate polymer structure: its molecular weight, monosaccharide composition, type of glycosidic bond, and the degree of chain branching [3,13–16].

# 2.1. Antioxidant Function of Polysaccharides

The radical cation 2,2'-Azino-bis (3-ethylbenzthiazoline-6-sulfonic acid) (ABTS) is often used to evaluate the general antioxidant activity of compounds [17]. Zhang et al. [18] showed that plant polysaccharides with higher mannose content and lower glucose content exhibit more pronounced scavenging activity against free radicals ABTS. Uronic acid with electrophilic groups (such as ketonic or aldehyde) in the acidic polysaccharide promotes the release of hydrogen from the OH bond [19]. Moreover, low molecular weight polysaccharides had more reductive hydroxyl ends (per unit weight) to accept and eliminate free radicals [18]. The activity of polysaccharides in scavenging ABTS radicals depends on the content of protein, uronic acid residues, and glucose [14,18,20,21].

There is a relationship between the average molecular weight of the polymer and the ability to scavenge 2,2-diphenyl-1-picrylhydrazyl (DPPH) radicals. The lower the molecular weight is, the higher the antioxidant activity relative to DPPH [14,18]. It can be assumed

Molecules **2021**, 26, 7068 3 of 36

that the value of the antioxidant activity is also influenced by the content of mannose, rhamnose, uronic acid, and protein components in the polymer composition [14,20,22,23]. Zhang et al. [18] demonstrated a significant correlation of activity within the content of arabinose and galactose. It is reported that the ability to scavenge DPPH radicals of neutral polysaccharides is greater than that of acid [24].

The scavenging activity of hydroxyl radicals increases with increasing concentration. Polysaccharides with a high content of uronic acid, mannose, and protein components have a higher antioxidant activity [14,25–27]. Zhang et al. [18] showed that a polysaccharide with a higher content of uronic acid and rhamnose and a lower glucose content exhibited the highest absorbing activity.

The activity of scavenging superoxide radicals depends on the content of uronic acid residues and protein components, as well as on the monosaccharide composition of the backbone [14,18,25]. High scavenging activity was exhibited by a heteropolysaccharide, which contained a high content of uronic acid and, consequently, a high content of carboxyl groups, which weakened the dissociation energy of the O–H bond, which led to higher scavenging activity [18]. A higher antioxidant activity against superoxide radicals was observed for samples with a large amount of protein components in the polysaccharide composition [14].

Zeng et al. [28] and Wang et al. [29] demonstrated that acidic polysaccharides, in which the monomeric units of the backbone are connected mainly by a  $\beta$ -glycosidic bond, had antioxidant activity. However, in Rozi et al. [30], the acidic polysaccharide, which had only  $\alpha$ -glycosidic bonds in its structure, had the highest antioxidant activity. This can be explained by the fact that these samples differed significantly in their weight-average molecular weight, which had a significant effect on the activity of the polymers.

With an increase in concentration to a certain value, the antioxidant activity increased, but the activity may decrease when this concentration value is exceeded [22,23,26,31]. The absorption capacity for metal ions increased with an increase in the polysaccharide concentration [20,22,23,29,32]. The galactose content also showed a significant correlation with the reducing ability of ferrous iron ions [33].

The xylose content had a significant effect on the chelating activity of metals [33]. It was shown in [29] that the neutral polysaccharide had a significantly higher chelating activity of  $Fe^{2+}$  than acidic polysaccharides.

An increase in the polysaccharide concentration also enhanced the effect of inhibiting the discoloration of  $\beta$ -carotene [22,25]. Polysaccharides, which mainly contained residues of mannose, ribose, glucose, galactose, xylose, and arabinose, had the potential for lipid peroxidation [22].

Thus, many polysaccharides are effective antioxidants. However, the mechanism is still not clear enough, and the relationship between the structure of the polysaccharide and its scavenging capacity has not been fully elucidated due to the wide variety and variability of the structural features of the molecules of these polymers.

## 2.2. Immunomodulatory Function of Polysaccharides

Plant polysaccharides are ideal candidates for immunomodulatory therapeutic agents due to their relatively low toxicity [18,34–39]. They can improve the viability of macrophage cells. Polysaccharides with a high proportion of galactose residues in their structure significantly promoted the proliferation of RAW264.7 cells [40–42]. However, a high concentration of carbohydrate polymers can also have the opposite effect [18,43].

Polysaccharides with higher molecular weights better activate NO production [18,35]. The activation of macrophages by plant polysaccharides is mediated by recognizing polysaccharide polymers by specific receptors [44]. High molecular weight polysaccharides can have even more repetitive receptor sites that can recognize receptors on macrophages [18]. Many polysaccharides exhibiting immunomodulatory activity contain significant amounts of mannose and galactose residues in their structure [18,29,35,45]. Sulfated polysaccharides have a high potential for immunological activity [32,46]. The NO production in

Molecules **2021**, 26, 7068 4 of 36

macrophages increased in a concentration-dependent manner upon treatment with various concentrations of the polysaccharide [25,36,43].

High molecular weight polysaccharides also activate cytokine production [34]. Pectin fractions with a large amount of galacturonic acid led to a higher production of IL-1 $\alpha$  and G-CSF cytokines by murine macrophages compared to other fractions [47]. The proliferation of splenocytes was influenced by polysaccharides with high molecular weights and containing residues of mannose or galactose [25,43]. Polysaccharides containing D-glucose and D-mannose residues in a certain concentration had a protective effect against the apoptosis of macrophages caused by  $H_2O_2$  [22,43].

The immunomodulatory activity of polysaccharides, determined in various studies, is summarized in Table 1.

**Table 1.** Immunomodulatory activity of polysaccharides with different structures.

Polysaccharide Components	Mw (kDa)	Immunomodulatory Activity	Sources
residues of arabinose, glucose, galactose, low content of protein components	401	did not have cytotoxicity, increased NO production, promoted the proliferation of spleen lymphocytes	[18]
residues of arabinose, glucose, galactose, low content of protein components	99	did not have cytotoxicity, increased NO production, promoted the proliferation of spleen lymphocytes	[18]
residues of uronic acid, arabinose, galactose, and glucose, low content of protein components	99	did not have cytotoxicity, increased NO production, promoted proliferation of spleen lymphocytes and proliferation of T-lymphocytes	[18]
residues of uronic acid, galactose, arabinose, and glucose, low content of protein components	43	did not have cytotoxicity, increased NO production, promoted proliferation of spleen lymphocytes and proliferation of B-lymphocytes	[18]
residues of glucose, mannose, ribose, galactose, xylose, and arabinose	-	increased survival of L02 cells caused by H <sub>2</sub> O <sub>2</sub>	[22]
residues of fucose, galactose, and 3-O-methylgalactose	120	increased NO production, stimulated splenocytes	[25]
type II arabinogalactan, galacturonic acid residues	-	did not have cytotoxicity, increased NO production by J774. A1 macrophage cells, and increased cytokine production	[34]
residues of D-mannose, D-glucose	394	had a stimulating effect on macrophage cells RAW 264.7, at high concentrations decreased cell viability, increased NO production, stimulated splenocytes and T-lymphocytes, had a protective effect against macrophage apoptosis caused by $\rm H_2O_2$	[43]
residues of D-mannose, D-glucose	362	had a stimulating effect on macrophage cells RAW 264.7, at high concentrations decreased cell viability, increased NO production, stimulated splenocytes and T-lymphocytes, had a protective effect against macrophage apoptosis caused by $H_2O_2$	[43]
galacturonic acid, arabinose, and galactose	-	increased the production of cytokines IL-1 $\alpha$ and G-CSF	[46]

# 2.3. Antitumor Function of Polysaccharides

Antitumor activity against cancer cells depends on the structure of the polysaccharide backbone [14]. Most polymers with such activity contain such monosaccharide residues as glucose and mannose [17,20,21,23,48]. However, the action of polysaccharides is specific for various cancer cells [17,20]. The concentration dependence of the antiproliferative activity

Molecules **2021**, 26, 7068 5 of 36

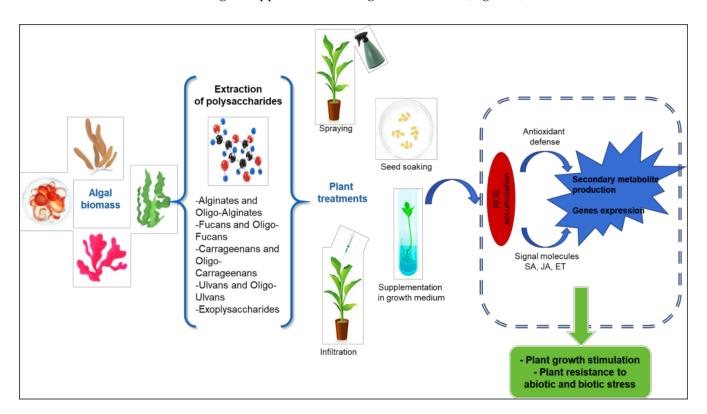
of polysaccharides was established [14,21,23,49,50]. A higher degree of branching led to stronger in vitro antitumor activity [51].

Thus, polysaccharides have different structural features, such as monosaccharide composition, type of glycosidic bonds, average molecular weight, degree of branching, presence or absence of protein components, or any functional groups. At the same time, the influence of various structural features on biological activity are interconnected with each other, therefore they must be considered in conjunction.

#### 3. Elicitation and Biostimulation

From an agronomic perspective, elicitor treatments with polysaccharides have gained prominence since their potential not only for modern agriculture practice but also as alternative tools to agrochemicals, allowing us to solve some environmental damage [52]. In the last decades, poly or oligosaccharides derived from algae have been more focus due to their ability to activate plant signals that enhance secondary metabolites' production and trigger plant defense responses [53,54]. Given their great diversity, these compounds have been also suggested to act as plant growth stimulators, which aim to intensify the absorption of substances or nutrients that will reduce chemical fertilizers [55].

Although it is largely established that polysaccharides act as effective biotechnological tools, their strategies' effects are not generalized. In fact, plant metabolism depends on the study model, characteristics of polysaccharides (structure, molecular mass, degree of polymerization and sulfate content) and types of treatment, e.g., foliar spraying, seed soaking or supplementation on growth medium (Figure 1).



**Figure 1.** Schematic representation of the general immune response of plants under algae polysaccharides treatments. SA: Salicylic acid; JA: Jasmonic acid; ET: Ethylene.

In recent years, tremendous advances have been made in understanding the general mode of action of algae polysaccharides as elicitors; however, their structure–activity relationships are still not well known.

Below, we briefly summarize the main results of the studied effects of poly- and oligosaccharides on plants' immune systems provided in the literature.

Molecules **2021**, 26, 7068 6 of 36

## 3.1. Polysaccharides as Inducers of Plant Defenses

The induction of resistance in plants by macroalgae polysaccharides is an effective method for disease control against environmental constraints which is linked to the activation or enhancement of several plant defense mechanisms such as the positive regulation of the expression of defense genes involved in the response of a wide range of abiotic and biotic stress, and the induction of the antioxidant system and the production of secondary metabolites [54,56–58]. Alginate or alginate oligosaccharides are the most algae carbohydrates described as potent elicitors in the literature (Table 2). For example, spray treatment with alginate in tomato induced the temporary production of reactive oxygen species, activated antioxidant defense, and increased the expression level of genes belonging to the SA-, JA-, and ET pathways conferring growth inhibition of Alternaria solani [59]. In addition, alginates of B. bifurcata and F. spiralis were shown to stimulate natural defenses of date palm by the activation of PAL activity and phenolic compound production [60]. Likewise, foliar application of κ-carrageenan in tomatoes reduced the severity of leaf spot caused by Septoria lycopersici via activation of antioxidant defense and modulation of chloroplast proteome [61]. Similarly, Tobacco infiltration with sulfated-carrageenan induced resistance to tobacco mosaic virus through the over-expression of defenses related genes encoding to PR proteins [62]. Sangha et al. [63] reported that only sulfated carrageenans have the ability to enhance resistance to Trichoplusia ni in Arabidopsis thaliana, indicating that the jasmonic acid and salicylic acid pathways are involved in this resistance. Fucan was also demonstrated to induce a multiplicity of plant defense events [54]. Klarzynski et al. [64] observed that fucan induced tobacco resistance to tobacco mosaic virus via the activation of some markers of systemic acquired resistance (SAR), such as the accumulation of salicylic acid and expression of the PR1 gene. Interestingly, and in the same way, microalgae polysaccharides were proved to trigger plant defense responses. Rachidi et al. [65] reported that polysaccharides extract from six different microalgae and cyanobacteria induced multiple signaling pathways in tomato, such as accumulation of ROS, pathogenesis-related proteins, and fatty acids, and improves the PAL and POX activities. Recently, Drira et al. [66] proved the potential of exopolysaccharides produced by *Porphyridium sordidum* to attenuate the severity of Fusarium oxysorum in Arabidpsis thaliana leaves. These authors provided evidence that EPS foliar applications activate the production of H<sub>2</sub>O<sub>2</sub> and enhance defense-related marker gene activities such as PAL, SOD, POD, CYP, and PR1.

Table 2. Alginate or oligoalginates generate resistance in different plants against diverse biotic and abiotic stresses.

Plant	Polysaccharide	Dose	Application Mode	Effect	Metabolism	References
Wheat	Alginate oligosaccharides	$1000~{ m mgL}^{-1}$	Supplementation in the growth medium	Tolerance to drought stress	-Enhancement of antioxidant system -Activation of related genes involved in ABA signal pathway (LEA1, SnRK2 and P5CS)	[67]
Kiwi fruit	Alginate oligosaccharides	50 mgL <sup>-1</sup>	Fruit soaking	Disease resistance to gray mold caused by <i>Botrytis</i> cinerea	-Enhancement of antioxidant system -Inducing of defense-related enzymes activities (PPO, PAL, and GLU)	[68]
Rice	Alginate oligosaccharides	$1~{ m mgmL}^{-1}$	Foliar spraying	Disease resistance to Magnaporthe grisea	-Activation of PAL, POD and CAT activities	[69]

Molecules **2021**, 26, 7068 7 of 36

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Plant	Polysaccharide	Dose	Application Mode	Effect	Metabolism	References
Safflowr	Alginate	0.075% and 0.15% (w/v)	Supplementation in growth medium	In vitro tolerance to salt stress	-Production of secondary metabolites (TPC, TFL, TFD, and Ant) -Enhancement in the antioxidant activity (CAT, TAC, and PAL)	[70]
Arabidopsis thaliana	Alginate Oligosaccharide	$^{25}$ mgL $^{-1}$	Foliar spraying	Resistance to Pst DC3000	-Production of early signal molecules (ROS, NO) - Activation of SA pathway	[71]
Cucumber	Alginate oligosaccharides	0.2% (w/v)	Foliar spraying	Water stress tolerance	-Decrease on MDA and (●OH) content -Activation of SOD and POD activities -Activation of genes involved in ABA signaling pathway	[72]

LEA1: Late embryogenesis abundant protein 1 gene, SnRK2: Sucrose nonfermenting 1-related protein kinase 2 gene, P5CS: Pyrroline-5-Carboxylate Synthetase gene, PPO: polyphenoloxidase, PAL: Phenylalanine ammonia lyase, GLU:  $\beta$ -1,3-glucanase, POD: Peroxidase, TPC: Total phenolic content, TFL: Total flavonoids (TFD), TFL: Total flavonoids, Ant: Anthocyanin, TAC: Total antioxidant capacity, ROS: Reactive oxygen species, NO: Nitric oxide, SA: Salicylic acid, ABA: Abscisic aid.

#### 3.2. Polysaccharides as Plant Growth Stimulator

In addition to their capacity to induce a defense response, algae polysaccharides can also stimulate plant growth. In fact, the growth promotion effect of polysaccharides extracted from microalgae [73–75] and macroalgae [55] was clearly demonstrated in terms of plant weight, plant size, leaves number, root length, chlorophyll content and photosynthetic activity. The biostimulant effect of purified poly or oligosaccharides has also been supported by several studies. For example, Yang et al. [76] have shown that the soaking of barley seeds in solutions of alginate-derived oligosaccharides enhanced seedling growth. This growth enhancement, which is linked to the molecular weights and Mannuronate/Guluronate ratio, was promoted by the stimulation on photosynthesis and amelioration of the adsorption activity. Similarly, Eucomis autumnalis bulbe coating by a low molecular mass oligoalginate increases plant height, florets number per inflorescence, and relative chlorophyll content [77]. In addition, foliar tobacco application by oligo carrageenans increases leaf biomass by stimulation photosynthesis efficiency, ribulose 1,5biphosphate carboxylase/oxygenase, and NAD(P)H-synthesizing enzymes activities [78]. Oligo carrageenans were also shown to enhance the level of growth-promoting hormones and C, N, and S assimilation in pine trees [79].

With increasing data from diverse research, algae polysaccharides appear to be an amazingly versatile elicitor agent. However, the potential interest of their commercializing is still in the early stages for agricultural use.

#### 4. Medicine

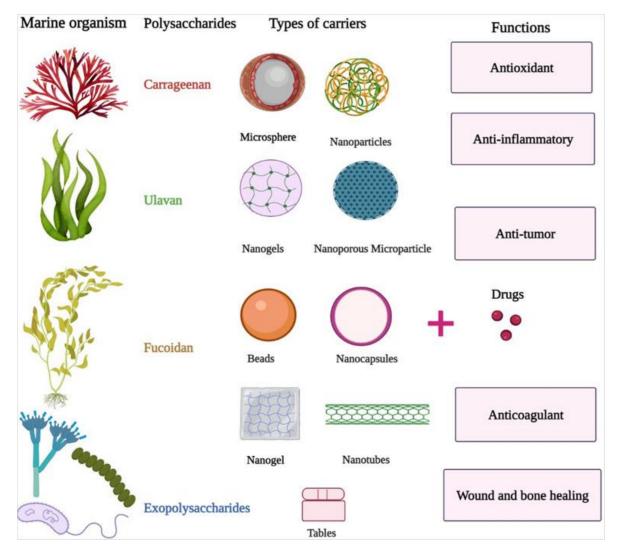
The potential of marine natural resources and recent advances in marine medicine biotechnology have led to the recent scale-up of the vast applications of marine resources in the medical area. Among a myriad of resources, recently, marine polysaccharides extracted from macroand microalgae and bacteria have attracted the attention researchers worldwide [80]. The biocompatibility, biodegradability, adhesivity, diversity of chemical structures, low toxicity, and the ability to form hydrogels in marine polysaccharides [4,81–83] resulted in their vast use

Molecules **2021**, 26, 7068 8 of 36

not only in food and cosmetic industries, but also led to their applications as materials for the incorporation of bioactive agents in drug delivery systems [84–92].

Marine sulfated polysaccharides including carrageenan, ulvan, and fucoidan extracted from macro algae or seaweeds [93–96], as well as microbial exopolysaccharides produced by extreme bacteria, fungi, and microalgae [97,98] are marine polysaccharides frequently reported during recent years because of their numerous biological properties, including antioxidant, anticoagulant, anticancer, antiviral, antiallergic, antiadhesive, antiangiogenic, and anti-inflammatory actions, as well as their high potential for deliver drug systems and tissue engineering [99–107]. New chemical modification methods have been developed during recent years to improve some of the biological activities of sulfated polysaccharides, to change their affinity to specific drugs, to increase their ability to incorporate drugs, and, finally, to increase the efficacy of their release [83,108].

Recently, in variety of carriers containing marine polysaccharides have been developed including nanoparticles, nanogels, nanotubes, nanocapsules, nanoporous microparticles, microspheres, hydrogels, beads, and tables, which effectively deliver drugs to target tissue in drug delivery systems [90,109–119] for medicine applications including gene therapy, cell therapy, pharmacokinetics, and tissue engineering (Figure 2). Here, we briefly review prominent developments of some dominant marine polysaccharides in deliver drug systems and tissue engineering.



**Figure 2.** The diversity of polysaccharides extracted and carriers developed from marine sources and potential pharmaceutical functions.

Molecules **2021**, 26, 7068 9 of 36

κ-carrageenan,  $\lambda$ -carrageenan, and  $\iota$ -carrageenan are three types of sulfated polysaccharides extracted from red algae [120] with the ability to form hydrogel for κ-carrageenan and  $\iota$ -carrageenan [121]. Microscale fibers, beads, nanotubes, spheres, nanoparticles containing carrageenan have been developed for diverse biomedical applications [88,122,123] (Figure 2). Some of the biological activities reported from carrageenan include antioxidant [124], antitumor, and immunomodulation activities [125], as well as reducing serum cholesterol and triglyceride levels [126], regulating growth factors [127,128], inhibiting syncytium formation [129], and promoting function HaLa cell and fibloblasts for healing wound [130].

In the field of tissue engineering, the carrageenans have been frequently used to deliver growth factors [118,119], to immobilize enzymes [131], for healing wounds [132], and to encapsulate several cell types, e.g., with human-adipose-derived stem cells, human nasal chondrocytes, or chondrocytic cells to deliver them in vivo for cell therapies and cartilage regeneration [119,120,133–135].  $\kappa$  and  $\iota$  –carrageenan are potential carriers for the controlled release of drugs [136–139]. Carrageenan-based hydrogels, beads and nanoparticles have been developed to deliver albumin [109,110,119], rosmarinic acid (RA) [140], acyclovir (antiviral drug) [5], ibuprofen [85], insulin aspart [87], as well as for their usage as a cell carrier [88] and echinochrome A [89].

Ulvan is sulfated polysaccharides extracted from the green algae with more diverse and complex structures, as well as biological activities less studied in compared to other sulfated polysaccharides [141–146]. Up to now, the diverse forms of carriers using ulvan including nanofibers, membranes, particles, hydrogels, and 3D porous structures have been developed (Figure 2) for delivery drugs, for peptide/protein [147–154], for wound dressing or bone tissue engineering [147–151,155–159], for inhibiting HeLa [160] and glioblastoma cells [161], for proliferation splenocyte [162], for enhancing fibroblasts growth and angiogenesis [163], for enhancing differentiation PC-12 cells [164], and for proliferation mesenchymal stem cells [165].

Fucoidan, constituting 5–10% of the dry biomass of brown seaweeds is sulfated polysaccharide with a high diversity of structure [166,167]. The ability of fucoidan to form gels and films is far less compared to carrageenan and ulvan [168], thus the mixing of fucoidan with other polymers can result in gels and films [169,170]. Among biological activities, the anticoagulant property of fucoidan has been well known [171–173]. Moreover, up to now, other activities such as antiviral [174], antimetastasis and antilymphangiogenesis [175,176], antitumor [177], anti-inflammatory [177–180], and anticancer effects [29,181–184] have been reported (Figure 2).

The diverse fucoidan-based carriers have been developed during recent years as nanoparticles for different purposes, such as for the releasing of the antitumor drug curcumin [90,91], the delivery of the anticancer drug doxorubicin [112], the encapsulation of the anticancer drug docetaxel (DTX) [113], for loading anticancer methotrexate (MTX) [114], for the delivery of antimicrobial and anti-inflammatory berberine [185–187], for the delivery of antitumor, anti-inflammatory, antioxidant, and hypoglycemic Oncocalyxone A [188], for releasing antibiotic gentamicin [189], for the carrying of basic fibroblast growth factor (bFGF) [190] and stromal cell-derived factor [191] as microparticles for the delivery lipoic acid, as hydrogels for releasing fibroblast growth factor-2 in vitro and in vivo [192], as nanoparticles functionalized with antibody-ErbB-2 for reducing tumor growth of lung [193], as 3D scaffolds for delivery vascular endothelial growth factor [115,116], and as polymeric micelles for loading the antitumor paclitaxel and curcumin [117] (Figure 2).

Exopolysaccharides (EPSs) are marine polysaccharides produced by marine bacteria [194–198], fungi [199,200], and microalgae [201–203] with new chemical and physical properties compared to polysaccharides produced by macro algae. The antioxidant function [204–209], preventing microbial infections [202,210–213], anticancer activity [86,92,201,214,215], healing bone [216,217], anticoagulant properties [218], antiviral activity [219–223], anti-inflammatory effects [224], potential for skin or cartilage grafting [222,225], inhibiting osteoclastogenesis [226], and promoting skin wound healing [227]

Molecules **2021**, 26, 7068 10 of 36

(Figure 2). An overview showed high diversity of biological properties, as well as the high potential of marine polysaccharide as multifunctional carriers in drug delivery systems and tissue engineering. It seems that the future exploration on new structures of marine polysaccharides, as well as their long-term toxicity assays, will prompt the developments of these biomaterials in drug delivery approaches and tissue engineering in near future.

#### 5. Food and Feed

## 5.1. Polysaccharides in Food Field

Food hydrocolloids are generally employed for their physical functions in stabilizing emulsions, viscous behavior, gelation, suspensions and foams, and control of crystal growth (Figure 3). The viscosity depends considerably on the preparation method. High temperature is particularly adverse, and the pH needs to be between 6 and 7 [228]. Seaweeds provide numerous several hydrocolloids to the food and feed industries [229], and the most important are agar, carrageenan (from red seaweeds), alginates, and sometimes sulfated fucoidans and laminarins (from brown seaweeds). Depending on their intrinsic structural characteristics and extrinsic environmental factors, polysaccharides frequently exhibit versatile rheological and physicochemical properties, which further affect their applications in food products. Typically, carbohydrates (especially polysaccharides) which present in liquid and/or solid food systems determine their structures and then their functions in industry.

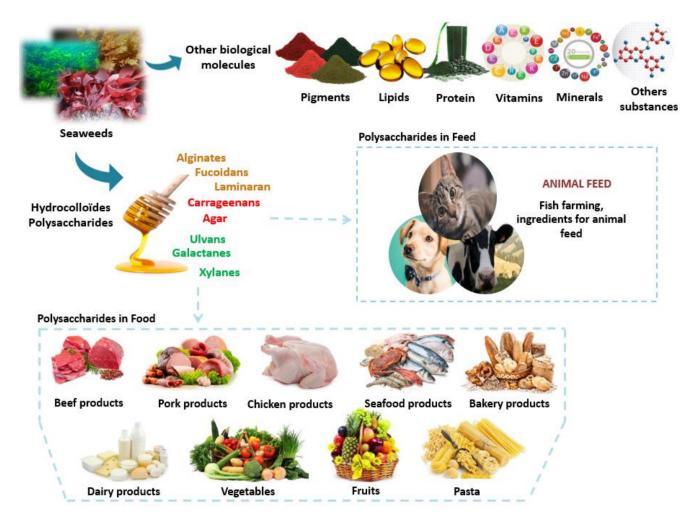


Figure 3. Algal polysaccharides in food and feed industry.

Molecules **2021**, 26, 7068 11 of 36

The influence of these biologically active macromolecules on smaller molecules, such as tastant compounds and aroma, has been investigated, with various studies concluding that apparent viscosity changes [230] and the physical entrapment of compounds [231] together explain perceptual differences [232]. These investigations tend to concentrate on the matrix structure and the release characteristics when envisaging changes in food perception.

Several studies have evaluated the effect of the incorporation of algal polysaccharides on nutritional, textural, and organoleptic properties of meat products (e.g., pork, beef, and fish products) (Figure 3). Recently, the antioxidative potential of laminarin (L), fucoidan (F), and an L/F extract from the brown seaweed *Laminaria digitata* was evaluated in pork homogenates and in horse heart oxymyoglobin. The results of this study demonstrated the feasibility of using these two polysaccharides (especially fucoidan) to increase the antioxidant activity of functional cooked meat products and improving the human antioxidant defense systems [233].

In their study, Jensen et al. [234] reported that alginate (used alone or in combination with other hydrocolloids) has an appetite regulator potential and thus could be used as a food supplement. However, according to [235], its incorporation into breakfast bars does not show significant differences as an appetite suppressant, compared to the control.

Albert et al. [236] demonstrated that alginate was also found to be effective as a coating film in microwave-cooked chicken nuggets, improving the heat distribution and thus shortening the cooking time. Sodium alginate was also used as a coating agent of bream, showing great results with additional antioxidant ability, which could delay the decay of the fish and enhance its shelf-life [237]. Coating foods with alginate improved the sensory quality and reduced the loss of water. It has also been reported that adding alginate to melon acts as a carrier for antimicrobials, which improved its shelf-life [238]. Alginates were also applied as a carrier for the anti-browning agents such as ascorbic acid and citric acid, which preserved the color of fresh cut Kent mangoes and improved the antioxidant potential. It has also been reported (i) that coating foods with alginate improved the sensory quality and reduced the loss of water and (ii) that adding alginate to melon acts as a carrier for antimicrobials, which improved its shelf-life [238].

Due to its thickening and gelling properties, its high melting temperatures, as well as its ability to hold into sugar to prevent crystallization, the agar extracted from rhodophyceae is highly sought in the food industry to prepare icings and bakery glazes [239]. The low gel strength matrix formed by agar is a property that makes its use possible in a wide range of food applications, including in liquid and spreadable foods (e.g., soft-texture confectionery) [240], as fat replacers, as cryoprotectants that minimize the damages occurring during the freezing/thawing process [241], and as edible films [232].

Carrageenan, isolated from red seaweeds, can function as a bulking agent, emulsifier, carrier, glazing agent, gelling agent, stabilizer, humectant, or thickener [232]. These sulfated polysaccharides are added to processed foods because it can bind water, promote gel formation, thicken, stabilize, and improve palatability and appearance through interaction with other substances in the food (e.g., carboxylmethyl-cellulose (CMC), galactomannan, starch, sodium, or calcium phosphates and proteins) [241]. Due to their important physicochemical and rheological properties, native carrageenans and semi-refined carrageenans are usually used in the food industry as an ingredient in dairy products such as ice cream, cheese, yoghurt, and milk-based products [242–245].

Other works have demonstrated the use of carrageenans in bakery products such as bread [246] and as a coating film to extend the shelf-life of fresh chicken breast [247].

Moreover, Piculell [241] showed that carrageenan can prevent separation and maintain texture in dairy products when added in small amounts of around 0.3% in milk gels (such as creamy fillings, flans, and custards), yoghourt, whipped cream, and milkshakes, and around 0.03% in liquid milk products and frozen desserts.

This sulfated polysaccharide can be used as a fat substitute in processed meats, as it restores tenderness and improves moisture retention in low-fat processed meats such as

Molecules **2021**, 26, 7068 12 of 36

hamburgers [248]. For example, in their research, Kumar and Sharma [249] showed that ground pork patties with less than 10.0% (w/w) total fat and carrageenan at important concentrations of 0.75% (w/w) actually had higher moisture retention after cooking and a similar texture compared to pork patties containing 20.0% (w/w) fat without carrageenan. It has been shown that carrageenan is successful in controlling discoloration, maintaining texture through shelf-life, and providing antibacterial protection when used as an edible fruit coating on sliced lychee bananas and mangoes [250].

## 5.2. Polysaccharides in Feed Field

Bioactive polysaccharides and oligosaccharides (BPO) are classified as a kind of indigestible but fermentable natural macromolecular carbohydrate. BPO are characterized by being biocompatible and biodegradable, along with their antibacterial, antioxidant, immunostimulating, and metabolic regulatory activities [1]. Because of these features, they can be used as effective alternatives to antibiotics in modulating gut microbiome. Several studies have investigated the prebiotic effects of oligosaccharides and polysaccharides derived from seaweeds in rats or mice being fed a seaweed-supplemented diet. Results conducted by Liu et al. [251] demonstrated a raise in the abundance of beneficial gut microbes such as *Bifidobacterium breve* and a diminution in pathogenic bacteria such as *Clostridium septicum* and *Streptococcus pneumonia* in rats supplemented with water-soluble polysaccharides from the red seaweed *Chondrus crispus*. Moreover, an increase in short chain fatty acids (SCFA) production and colonic growth was obtained, as well as an improvement of host immunity modulation through an elevation of the plasma immunoglobulin levels.

The supplementation of diets with extracts of the brown seaweeds  $\mathit{Undaria\ pinnati-fida}$  and  $\mathit{Laminaria\ japonica}$  has resulted in suppressed weight gain of rats, influenced by the composition of gut microbial communities associated with obesity by a reduction in the ratio of Firmicutes to Bacteroidetes and reduced populations of pathogenic bacteria, including  $\mathit{Clostridium}$ ,  $\mathit{Escherichia}$  and  $\mathit{Enterobacter\ genera\ [252]}$ . From Lean et al. [253], the oral administration of fucoidan from brown seaweeds has been shown to reduce the inflammatory pathology associated with dextran sulfate sodium (DSS)-induced colitis in mice, indicating its important potential for treating inflammatory bowel disease. Furthermore, Kuda et al. [254] showed that rats fed with a diet containing laminarin and low  $M_W$  alginate isolated from pheophyceae suppressed the production of indole, p-cresol, and sulfide, which are the putative risk markers for colon cancer. The neoagaro-oligosaccharides derived from the hydrolysis of agarose by  $\beta$ -agarase enzyme resulted in a rise in the numbers of  $\mathit{Lactobacillus}$  and  $\mathit{Bifidobacterium}$  in the feces or cecal content of mice, along with a decrease in putrefactive bacteria [255].

# 6. Antimicrobial/Antiviral Agents

The biodegradability, biocompatibility, and non-toxic nature of polysaccharides isolated from natural sources, make them valuable ingredients in different fields, such as pharmaceuticals, nutraceuticals, food, or cosmetic industries. They have been used in healthcare, namely in cancer diagnosis and treatment, in drug delivery, in tissue engineering, and as antimicrobial and antiviral agents [256].

In this section, a compilation of the literature of polysaccharides from natural sources with antimicrobial/antiviral potential were considered, as described in Table 3.

*Molecules* **2021**, *26*, 7068

 $\textbf{Table 3.} \ \ \textbf{Main polysaccharides from natural sources with antimicrobial and antiviral potential.}$ 

Polysaccharides	Source	Biological Activity	Species	References
Alginate/alginic acid	Seaweed	Antiviral	HIV-1, HPV, DENV, HSV	[257–259]
Almond gum	Plant	Antibacterial	Bacillus thuringiensis, Klebsiella pneumonia, Bacillus subtilis, Pseudomonas aeruginosa, Listeria monocytogenes	[260]
Carrageenan	Seaweed	Antibacterial	Chlamydia trachomatis, Saccharomyces cerevisiae, Staphylococcus aureus, Bacillus cereus, Escherichia coli, Pseudomonas aeruginosa	[257,258,261–265]
		Antiviral	HPV, HSV-1, HSV-2, HIV, HRV, Influenza A, DENV-2, DENV-3, VZV, RABV, EV71, SARS-CoV-2	
Ulvan	Seaweed	Antibacterial	Enterobacter cloace, Escherichia coli	[262,266]
Olvan	Seaweed	Antiviral	NDV, JEV	[202,200]
Rhodophyta Galactans	Seaweed	Antiviral	HSV-1 and HSV-2, DENV-2, HIV-1 and HIV-2, HAV, HPV, DENV	[257,259,267]
Calcium spirulan	Cyanobacteria	Antiviral	HSV-1, HCMV, Influenza A, Coxsackie virus, MV, HIV-1, PV, Mumps virus, HPV, DENV	[257,259,268]
Nostoflan	Cyanobacteria	Antiviral	HSV-1, HSV-2, HCMV, Influenza A	[257,259,269–271]
Chitin/chitosan	Animal	Antibacterial	Escherichia coli, Vibrio cholerae, Shigella dysenteriae, Bacteroides fragilis	[272,273]
Dextran	Bacteria	Antiviral	HPV	[258,266,274]
		Antifungal	Candida albicans	
Fucoidan	Seaweed	Antibacterial	Listeria monocytogenes, Micrococcus luteus, Staphylococcus aureus, Salmonella typhimurium, Streptococcus mutans, Streptococcus sanguinis, Streptococcus sobrinus, Strongyloides ratti, Streptococcus criceti, Streptococcus anginosus, Streptococcus gordonii, Aggregatibacter actinomycetemcomitans, Fusobacterium nucleatum, Prevotella intermedia, Porphyromonas gingivalis	[95,257,262,267, 271,275,276]
		Antiviral	HIV, HSV-1, HSV-2, DENV, HCMV, NDV, SARS-CoV-2	
		Antifungal	Aspergillus flavus, Aspergillus fumigatus, Mucor sp.	
Ginseng's polysaccha- ride	Plant	Antibacterial	Helicobacter pylori, Bacillus cereus, Staphylococcus aureus, Pseudomonas aeruginosa, Listeria monocytogenes, Salmonella enteritidis, Escherichia coli, Streptococcus pneumoniae	[277,278]
		Antifungal	Candida albicans	
		Antiviral	H1N1 Influenza virus, H5N1 Influenza vírus, HIV, HBV, RSV	
Heparin	Animal	Antiviral	HPV, SARS-CoV-2	[258,279–281]

Molecules **2021**, 26, 7068 14 of 36

Table 3. Cont.

Polysaccharides	Source	Biological Activity	Species	References
Laminarin	Seaweed	Antibacterial	Escherichia coli, Listeria monocytogenes, Staphylococcus aureus, Salmonella typhimurium	[257,259]
		Antiviral	HBV, HIV-1	
Lentinan	Fungi	Antiviral	SARS-CoV-2	[282]
Levan	Bacteria	Antiviral	(HPAI) A(H5N1), ad40	[283]
Pectin	Plant	Antibacterial	Citrobacter sp., Salmonella sp., Enterobacter sp., Shigella sp., Proteus sp., Klebsiella sp.	[256]
Polygonum multiflo- rum's polysaccha- ride	Plant	Antiviral	Coronavirus	[284]
Ganoderma polysaccha- rides	Fungi	Antibacterial	Erwinia carotovora, Bacillus cereus, Acinetobacter aerogenes, Acrobacter aerogenes, Arthrobacter citreus, Bacillus brevis, Bacillus subtilis, Corynebacterium insidiosum, Escherichia coli, Proteus vulgaris, Clostridium pasteurianum, Micrococcus roseus, Mycobacterium phlei, Staphylococcus aureus	[285]
		Antifungal	Penicillium digitatum, Aspergillus niger	
Xylan	Plant	Antibacterial	Klebsiella pneumoniae	[286]
Ay tari	I Idill	Antiviral	HSV	[200]

HIV—Human immunodeficiency virus, HPV—Human papillomavirus, DENV—Dengue virus, HSV—Herpes simplex virus, HRV—Human Rhinovirus, VZV—Varicella zoster vírus, RABV—Rabies virus, EV—Enterovirus, SARS-CoV-2—Severe Acute Respiratory Syndrome Coronavirus-2, NDV—Newcastle disease vírus, JEV—Japanese encephalitis virus, HAV—Hepatitis A, HCMV—Human cytomegalovirus, MV—measles virus, PV—Poliovirus, HBV—Hepatitis B virus, RSV—respiratory syncytial virus, (HPAI) A(H5N1)—Highly Pathogenic Asian Avian Influenza A(H5N1) Virus, ad40—Adenovirus type 40.

The great potential of polysaccharides from different sources as antiviral agents is undeniable, especially when the origin is seaweeds. Alginate, fucoidan, and laminarin, typically obtained from brown algae [95,256-259,262,271,275,287], carrageenan and galactans from red algae [257,258,261-265], and ulvan from green algae [262,266], often present antiviral activity against several viruses (Table 3). Indeed, this recognized biological activity immediately aroused the interest of the scientific community to search for solutions against the new SARS-CoV-2 virus responsible for causing COVID-19 [264,271,288]. Several polysaccharides have been explored with this propose, as is the case for carrageenan and fucoidan, from algae and heparin, with animal origin, that have already shown promising results against this virus [257,262,271,281]. Different antiviral mechanisms are associated to these compounds, such as the inhibition of enveloped and nonenveloped viruses through the inhibition of the binding or internalization of the virus into the host cells, the inhibition of virus replication through the suppression of the DNA polymerase activity, among others [257]. The role of heparin, however, should be highlighted. This polysaccharide, usually used as anticoagulant agent, is also known to contribute as an antiviral agent. For SARS-CoV-2, studies have shown that this bioactive compound strongly binds to the Spike protein, avoiding the entry in the host cells. Particularly for viruses causing respiratory problems, heparin also prevents pulmonary thrombosis, suggesting that heparin may act through multiple mechanisms [281].

Molecules **2021**, 26, 7068 15 of 36

Regarding the potential of polysaccharides against microbes other than viruses, the antibacterial activity stands out when compared with antifungal activity (Table 3). Several polysaccharides were found to be effective against a wide range of pathogenic bacteria. Between them, almond gum [260], carrageenan [263,265], laminarin [259], pectin [256], and polysaccharides obtained from species of *Ganoderma* sp. [285] demonstrated the ability to inhibit the growth of species as *Listeria monocytogenes*, *E. coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumonia*, *Chlamydia trachomatis*, *Staphylococcus aureus*, *Salmonella typhimurium*, *Helicobacter pylori*, *Prevotella*. *intermedia*, *Porphyromonas gingivalis*, among many others known to cause infections in humans.

A similar scenario is not observed for the antifungal activity of polysaccharides, possibly not due to the lack of activity, but due to the lower number of studies including this type of organisms. However, fucoidan and ginseng showed not only antiviral and antibacterial activity, but also antifungal activity as well [276–278]. Together with *Ganoderma* polysaccharides [285], pathogens extremely relevant not only for human health (as *Candida albicans*, *Aspergillus fumigatus*, and *A. flavus*), but also for agriculture (as *A. niger* and *Penicillium digitatum*), were identified, making them great candidates for the pharmaceutical and phytopharmaceutical industries. Taking this information in consideration and knowing that fungi are organisms capable of inflicting significant losses in several fields, more research should be considered in an attempt to find compounds able to control them.

In addition to the antimicrobial/antiviral potential of the polysaccharides presented in Table 3, other polysaccharides are known to indirectly contribute to those activities, as drug delivery carriers or even by stimulating defense mechanisms of hosts against pathogens or the growth of the regular microbiota; this fact is also extremely relevant for the pharmaceutical industry as well. The seaweed polysaccharide laminarin is known to stimulate the defense mechanisms of plants that are involved in the cascade of genes encoding proteins related with antimicrobial properties [289]. In the case of pullulan, a polysaccharide produced by the fungus *Aureobasidium pullulans*, its chemical structure is easily modified to deliver different drugs in the form of microparticles, nanoparticles, and hydrogels, among others [290]. Similarly, polysaccharides such as agar, hyaluronan, konjac glucomannan, schizophyllan, bacterial cellulose, xanthan gum, among others, have also been explored with this objective, presenting different therapeutic targets [279,291,292]. Another case is inulin, a polysaccharide produced by plants that is responsible for enhancing the proliferation of bacteria such as Bifidobacteria, known to be health-promoting, suppressing the growth of potential pathogens in the gut, and also acting as immunomodulators [293]. Although antibacterial activity is not described for levan, it has been proposed as a potential compound for the treatment of peptic ulcers typically associated to bacteria such as *Helicobacter pylori*. Due to the high adhesion ability and prebiotic activity, it has the capacity to protect and prevent the development of peptic ulcers [294].

Another feature to be explored in this field is the possibility to create synergisms between different compounds. A study conducted with fucoidan and antibiotics against oral pathogenic bacteria showed that the combination of this polysaccharide with antibiotics lead to an increase in the rate of the elimination of colony forming units per milliliter, when compared with the results obtained for the antibiotics or polysaccharides alone [275].

All these possibilities show the great potential of polysaccharides from different natural sources as antimicrobial and antiviral agents, not only in a direct way but also through different mechanisms that contribute to the same goal.

## 7. Chemical, Chemo-Enzymatic, and Enzymatic Functionalization of Polysaccharides

Polysaccharides can have various biological activities, for example, antiviral, antitumor, antioxidant, and immunomodulatory [4,9,13], which depends on their structure [4,36,295]. Polysaccharides have many highly reactive groups (acetamido, amino, carboxyl, hydroxyl groups) [296], which can be used in various functionalization ways. The biological activity of polysaccharides can be significantly increased with properly selected methods and conditions for carrying out structural modification [297,298].

Molecules **2021**, 26, 7068 16 of 36

#### 7.1. Chemical Functionalization

Chemical modification, in general, is the introduction of various functional groups into the polysaccharide structure [295–299]. This often leads to a decrease in the molecular weight of the polymer [300] and, consequently, an increase in its solubility in water [299]. The biological properties of modified polysaccharides depend not only on the method of chemical modification but also on the degree of substitution. Moderate substitution can enhance the biological activity of the polysaccharide [300]. In this case, a concentration-dependent effect of the activity of the carbohydrate polymer was observed [300,301].

One of the most common methods for modifying polysaccharides is sulfation. Polysaccharides functionalized in this way have higher immunoregulatory, antiviral, anticoagulant, antitumor, and antioxidant activities [298,302].

Another method of moderate substitution is the acetylation of polysaccharides. Acetyl groups can cause polysaccharide branches to stretch and change orientation, causing the formation of polysaccharide molecules with a transverse order [299]. As a result of a spatial arrangement change in the polysaccharide chains, the properties of macromolecules also change. It was found that the introduction of acetyl groups significantly increased the antioxidant [300,301], immunomodulatory [300,303,304], and anti-inflammatory [303] activities of the polysaccharides.

Phosphorylation and benzoylation can also increase the antioxidant activity of polysaccharides [305–307]. Xu et al. [15] demonstrated that acetylated and benzoylated polysaccharides were more effective in lowering blood glucose levels in mice when used as a cardioprotective agent compared to the native polymer.

Carboxymethylation can increase the antioxidant, antitumor, immunoregulatory, and antibacterial functions of polysaccharides [308]. It was shown in [309] that carboxymethylation converted polysaccharides into water-soluble products. Chen et al. [300] showed that carboxymethylation could slightly increase the ability to inhibit the discoloration of  $\beta$ -carotene. There was also a slight increase in the effect on the pinocytic activity of peritoneal macrophages in mice, which decreased the effect of the TNF- $\alpha$  protein on the secretion.

If necessary, it is possible to select the modification conditions with no decrease in the molecular weight of the polymer. The authors of [310] described a method for dextran phosphorylation, which did not lead to polysaccharide chain degradation. Silva et al. [311] determined the optimal conditions for carboxymethylation in which the least chain degradation was observed.

Chemical functionalization is the most common among the numerous methods for modifying polysaccharides due to the vast possibilities of introducing various functional groups into the structure of the polymer molecule. However, along with the advantages, this type of modification has some disadvantages. The major one is the frequent toxicity of the chemicals used.

#### 7.2. Chemo-Enzymatic Functionalization

Enzymes can be used as catalysts for the attachment of functional groups employing chemical reagents, and can themselves act as a functionalizing component, connecting with a polysaccharide molecule using a cross-linking agent [312]. Compared with the method of chemical binding, the method of modification catalyzed by enzymes is cheaper and safer [313].

The polysaccharides were modified with phenolic acids (gallic [314], caffeic [314,315], and ferulic [316]) in the presence of laccase as a catalyst. Chitosan molecules functionalized with phenolic acids showed much higher antioxidant activity in scavenging the radical cations 2,2'-Azino-bis-(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) [314] and 2,2-diphenyl-1-picrylhydrazyl (DPPH) [315 and, under certain modification conditions, a higher antimicrobial activity against *E. Coli*, as well as antifungal activity against *C. albicans* [314].

Molecules **2021**, 26, 7068 17 of 36

Lipase is often used to acylate the hydroxyl groups of polysaccharides [317–319]. Lipase, possessing regioselectivity, gives direction to the process of the structural modification of the polysaccharide [319].

Peroxidases are used to catalyze oxidative polymerization and graft the functional molecules to polymers in a two-step reaction. The first step involves the generation of radicals by peroxidase, then reactively oxidized target molecules crosslink the polymer [320]. Li et al. [321] reported that lytic polysaccharide monooxygenases could efficiently supply  $H_2O_2$  in situ to peroxidases using a gallic acid substrate for the functionalization of chitosan. In [322], a pectin polysaccharide was enzymatically modified by crosslinking ferulic acid groups using horseradish peroxidase. The resulting polysaccharides showed an improved ability to stabilize oil-in-water emulsions against coalescence and flocculation [322].

Tegl [315] demonstrated a chemo-enzymatic functionalization with glucose oxidase as a modifying reagent of particles of chitosan–zeolite and chitosan–zeolite modified with caffeic acid. Chitosan–zeolite particles modified with caffeic acid and then glucose oxidase demonstrated enhanced combined antioxidant and antimicrobial activity compared to the polysaccharide without modifications.

## 7.3. Enzymatic Functionalization

Enzymatic processing avoids the use of substances that are aggressive and harmful to human health. Enzymes are suitable tools for changing the structure of polysaccharides due to their specificity. Enzymatic technologies have great potential for modifying the properties of natural food ingredients such as polysaccharides to improve their functional characteristics [318]. However, the possibilities of such a modification are very limited and often come down to the cleavage of various constituents of the polysaccharide chain, the shortening of the chain, and, as a consequence, to a decrease in the values of molecular weight and intrinsic viscosity.

Oosterveld et al. [323] showed enzymatic deacetylation of pectin polysaccharide using pectin methylesterase, pectin acetylesterase, rhamnogalacturonan acetylesterase, arabinofuranosidase B, and rhamnogalacturonase in various combinations. The modification of rhamnogalacturonans and arabinans with enzyme mixtures such as endo-arabinase plus arabinofuranosidase, rhamnogalacturonase plus rhamnogalacturonan acetylesterase, and polygalacturonase plus pectin methyl esterase resulted in a decrease in molecular weights and intrinsic viscosity [323]. It was shown in [324] that the use of pronase E to modify exopolysaccharides isolated from lactic acid bacteria did not affect the molecular weight of the studied samples and, therefore, was suitable for protein removal.

Thus, numerous methods for modifying polysaccharides lead to a wide range of polymers with different biological properties. When modifying polysaccharides, it is possible to weaken their biological properties [300,314,316]. For this reason, the modification conditions and methods should be selected depending on the functionalization goals. It is essential to select the optimal conditions for the modification to obtain polymers with improved biological properties.

## 8. Current Markets

Long and laborious processes associated with both technology-transfer and regulatory constraints in taking a substance from research to the market is one of the main explanations for the fact that the list of carbohydrates with biotechnologically relevant properties currently on the market is shorter than that of all carbohydrates being researched for their biological activities. A compilation of currently marketed bioactive polysaccharides was performed and presented in Table 4.

Molecules **2021**, 26, 7068 18 of 36

**Table 4.** Currently marketed bioactive polysaccharides along with their biological origin and the main properties for which they are sold.

Polysaccharide	Mainly Sold As/For			
Animal Origin				
I Tana a mina	Anticoagulant (medical practice)			
Heparin	Scientific research (several novel medical uses)			
	Treatment of osteoarthritis in humans and other animals			
Chondroitin sulfate	Treatment of cataracts			
	Nutraceutical for cartilage, joint, and bone health <sup>1</sup>			
	Treatment of osteoarthritis and cataracts			
	Topical pharmaceutics (e.g., dry skin, dry eyes, wounds, or			
Hyaluronic acid <sup>2</sup>	oral inflammation)			
-	Plastic surgery filler			
	Cosmetic formulations (anti-aging properties)			
	Sizing and strengthening paper			
	Plant priming			
	Soil fertilizer/conditioner			
	Wine fining			
Chitin/chitosan <sup>3</sup>	Water treatment			
	Biomaterial (e.g., food packaging or wound dressings)			
	Nutraceutical for weight loss <sup>1</sup>			
	Nutraceutical with cholesterol lowering effect			
	Scientific research (several novel uses)			
	Plant Origin			
	Gelling agent			
	Stabilizer			
Pectin	Dietary fiber (supplement) with cholesterol lowering effec			
	Dietary fiber (supplement) that delays gastric emptying <sup>1</sup>			
	Dietary fiber (supplement) that ameliorates constipation <sup>1</sup>			
	Thickening agent			
	Emulsifier			
Konjac glucomannan	Dietary fiber (supplement) with cholesterol lowering effect			
	Dietary fiber (supplement) that ameliorates constipation			
	Nutraceutical for weight loss <sup>1</sup>			
	Nutraceutical for immune system stimulation			
Astragalus polysaccharide	Nutraceutical with diuretic effect			
Tistiuguius polysaccitatide	Nutraceutical with antiviral activity			
	Nutraceutical with several other medical applications <sup>1</sup>			
	Source for xylitol production			
	Plastic additive			
Xylan	Scientific research (novel packaging solutions and biomedic			
	applications)			
	Cosmetic ingredient			
	Nutraceutical with prebiotic and gut health promoting			
	activitiesNutraceutical with antidiabetic activity			
Inulin	Nutraceutical with weight loss activity			
mumi	Dietary fiber (supplement) that ameliorates constipation			
	Food ingredient to increase fiber content and substitute sug-			
	starch, and fats			
Dolugonum mailtiflamas	Nutraceutical for hair strengthening and color restoration			
Polygonum multiflorum	Nutraceutical for neuroprotection <sup>1</sup>			
polysaccharide	Nutraceutical with several other medical applications <sup>1</sup>			

Molecules **2021**, 26, 7068 19 of 36

 Table 4. Cont.

Polysaccharide	Mainly Sold As/For
	Nutraceutical to increase fiber intake (also, indirect method of weight loss)
	Nutraceutical with cholesterol lowering effect
	Food ingredient to thicken without gluten, gelatin, or eggs
	Dietary fiber (supplement) that ameliorates constipation or
Guar gum	diarrhea
	Multiple uses in paper and textile industries
	Thickener (multiple industries) Stabilizer (food industry)
	Binder (multiple industries)
	Laxative
	Seaweed Origin
	Superabsorbent
	Plant priming
	Drug-release carrier
	Inoculant carrier
	Thickening agent (multiple industries)
	Stabilizer (multiple industries)
	Gelling agent (multiple industries)
Alginate/Alginic acid	Emulsifier (multiple industries) Biomedical scaffolding
	Anti-poisoning <sup>1</sup>
	Nutraceutical with cholesterol lowering effect <sup>1</sup>
	Nutraceutical with anti-hypertensive effect <sup>1</sup>
	Nutraceutical to increase fiber intake (also, indirect method of
	weight loss) <sup>1</sup>
	Ingredient in peel-off skin masks
	Reflux treatment
	Gelling agent (multiple industries)
	Thickening agent (multiple industries)
	Stabilizer (multiple industries) Emulsifier (multiple industries)
Carrageenan	Lubricant
	Inactive excipient in pharmaceutical formulae
	Immobilization agent for cells/enzymes (biotechnology)
	Pro-inflammatory agent (scientific research)
	Gelling agent (multiple industries)
	Food ingredient to solidify liquids
Agar	Dietary fiber (supplement) that ameliorates constipation or diarrhea
	Scientific research (molecular biology and microbial/plant
	culture media)
	Nutraceutical with immunostimulant activity
	Nutraceutical with anticancer activity <sup>1</sup>
	Nutraceutical with anti-inflammatory activity <sup>1</sup>
Fucoidan	Nutraceutical with antihypertension and
	antihypercholesterolemia activity <sup>1</sup>
	Nutraceutical with anticoagulant and antithrombotic activities
	Nutraceutical with antioxidant activity Reagent for scientific research (bioactivities and enzyme
	activity)
Laminarin	Cosmeceutical with moisturizing capacity
	Dietary fiber (supplement)Nutraceutical with some
	health-related claims <sup>1</sup>
Caulerpa sulfated polysaccharide	Nutraceutical with immunostimulation activity
Same pa sallace polysaccialle	Nutraceutical with anticoagulant activity

*Molecules* **2021**, 26, 7068 20 of 36

 Table 4. Cont.

Polysaccharide Mainly Sold As/For					
Bacterial Origin					
_	Pharmaceutical for hypovolaemia treatment Antithrombotic and blood thinner				
Dextran	Medical lubricant				
	Pharmaceutical for parenteral nutrition/blood substituent				
	Research (several laboratory applications) Cosmeceutical ingredient with haircare and skin-whitening				
<b>T</b>	properties				
Levan	Food ingredient (dietary fiber supplement and sweetener) Scientific research (bioactivities, e.g., prebiotic,				
	anti-inflammatory, and antimicrobial)				
C 11	Gelling agent (multiple industries)				
Curdlan	Water-holding and stabilizing agent (food industry)				
	Scientific research (several therapeutic potentials)				
	Thickener (multiple industries) Stabilizer (multiple industries)				
Bacterial cellulose	Modern wound dressings				
	Several medical applications as structural material				
	Thickener (multiple industries)				
Xanthan Gum	Stabilizer (multiple industries)				
	Scientific research (biomedical scaffolding)				
	Thickener (multiple industries)				
C 11	Emulsifier and stabilizer				
Gellan gum	Drug-release carrier and cell encapsulation agent				
	Agar-substitute in culture media				
	Fungal Origin				
	Edible film-forming polysaccharide (oxygen-barrier)				
	Low-calories, tasteless food ingredient (bulking fiber,				
Pullulan	antifungal)				
	Adhesive and binder (multiple industries)				
	Drug-release agent				
	Thickening agent (food industry)				
	Suspension agent (food industry)				
	Water-holding agent (cosmeceutical industry)				
	Immunostimulant (especially against fungal infections)				
Scleroglucan	Edible films				
	Binder in tablets and drug-release carrier				
	Nutraceutical with hypolipidemic and hypoglycemic activities				
	Artificial tears and artificial saliva component				
	Anticancer pharmaceutical				
	Anti-HIV and anti-hepatitis pharmaceutical				
Lentinan	Malignant pleural effusion treatment				
	Nutraceutical with immunomodulation activity				
0.41	Nutraceutical with immunostimulation activity (indirect				
Grifolan	antitumor activity)				
	Nutraceutical with immunomodulation activity (indirect				
Schizophyllon	antitumor activity)				
Schizophyllan	Cosmetic ingredient with soothing/anti-inflammatory				
	properties				
Krestin	Nutraceutical with immunomodulation activity (indirect				
Ricotiii	antitumor activity)				
	Nutraceutical with immunomodulation activity (indirect				
Reishi polysaccharide	antitumor activity)				
	Several nutraceutical and pharmaceutical uses				

Molecules **2021**, 26, 7068 21 of 36

Table 4. Cont.

Polysaccharide Mainly Sold As/For				
	Lichen origin			
Lichenan Pustulan	Scientific research (bioactivity and enzyme activity) Scientific research (bioactivity and enzyme activity)			

<sup>&</sup>lt;sup>1</sup> Claim with insufficient scientific evidence; <sup>2</sup> Also extractable from bacteria; <sup>3</sup> Also extractable from fungi.

Noteworthy, most carbohydrate polymers in the market serve at least one of three main functions as a physico-chemical formulae modifier (e.g., thickener, stabilizer, binder, or emulsifier in food, feed, or cosmetic products), as a nutraceutical supplement with health claims, or as a pharmaceutical product. Importantly, more often in the case of nutraceuticals but also sometimes in the case of pharmaceuticals, some of the health claims are supported in preliminary scientific research (e.g., in vitro models) or even in the absence of scientific evidence (e.g., cultural or traditional medicine beliefs).

Many of the polymers detailed in Table 4 are sold in their purified form (e.g., heparin, pectin, carrageenan, among others), while some of the noted polysaccharides are not explicitly sold as such (e.g., *Astragalus* polysaccharide). In the case of the latter, it is often found that the whole source organism (e.g., in powder) or an extract of the source organism (standardized for a certain percentage of polysaccharides) is sold instead. In such cases, it becomes harder to quantify and characterize the market. An example of this difficulty is that of ginseng's polysaccharides. Despite being recognized in academia as one of the main bioactive components in ginseng products [325], ginseng itself as a whole product or in the form of extracts (which contain many more metabolites than the polysaccharides alone) are so popular that not one single polysaccharide-specific product could be found—it is for this reason that it is not included in Table 4.

Virtually all the listed polysaccharides in Table 4 present different variations in properties/claims and applications according to the specific source organism and method of extraction/refinement. Due to these two variables, polymers with different molecular weights and degrees of ramification are obtained, which in turn present different bioactivities. Due to an increased demand from consumers and rapid innovation in the fields of food, feed, cosmetics, and biopharmaceuticals technologies, as with most natural products, the markets for the listed polysaccharides are growing. This growth, however, has been modified (either positively or negatively) by the recent events of the COVID-19 pandemic. Low molecular weight heparin has been used prophylactically and therapeutically in COVID-19 patients [326], which has contributed to an increase in demand for this product (https://www.alliedmarketresearch.com/heparin-market-A06186 (accessed on 4 September 2021). On the other hand, non-essential polysaccharide products directly related to businesses that were shut down during the pandemic (e.g., xanthan gum in the beauty industry) observed a decrease in growth rate due to COVID-19 (https://www.theinsightpartners.com/reports/xanthan-gum-market (accessed on 4 September 2021).

The group of animal-based polysaccharides is, by far, the largest market for these polymers, with a combined market value superior to 20 billion USD in 2020 (https://www.alliedmarketresearch.com/heparin-market-A06186 (accessed on 4 September 2021), https://www.reportsanddata.com/report-detail/hyaluronic-acid-market (accessed on 4 September 2021), https://www.bccresearch.com/market-research/plastics/chitin-chitosan-derivatives-markets-report.html. (accessed on 4 September 2021), https://www.grandviewresearch.com/industry-analysis/chondroitin-sulfate-market (accessed on 4 September 2021). Of these, hyaluronic acid is the polymer with the largest market size (9.6 billion USD in 2020) (https://www.reportsanddata.com/report-detail/hyaluronic-acid-market (accessed on 4 September 2021), followed by heparin (6.5 billion USD in 2020) (https://www.alliedmarketresearch.com/heparin-market-A06186 (accessed on 4 September 2021), chitin/chitosan (projected 4.2 billion USD in 2021) (https://www.bccresearch.com/market-research/plastics/chitin-chitosan-derivatives-markets-report.html), and chondroitin sulfate (1.2 billion USD in 2020)

Molecules **2021**, 26, 7068 22 of 36

(https://www.grandviewresearch.com/industry-analysis/chondroitin-sulfate-market (accessed on 4 September 2021). The fact that animal-based polysaccharides are of such value is directly related to their value per kilogram, which is high due to their medical-grade processing requirements, along with the high-volume of sales of chitin/chitosan in the field of water treatment. Plant-based gums/fibers, as well as seaweed phycocolloids, have much higher volumes of sales, but lower prices per kilogram, given the less refined nature of these mostly food-grade polymers. An example of such a polymer is agar, which, despite its high volume of sales in kilograms, has a market size estimated of 239 million USD (in 2020) (https://www.industryarc.com/Research/Global-Agar-Market-Research-509553 (accessed on 4 September 2021). Nonetheless, the plant-based polysaccharide inulin, given its use as a nutraceutical prebiotic fiber, has a higher price range, rendering a market of 2.35 billion USD in 2020 (https://www.grandviewresearch.com/industry-analysis/inulin-market (accessed on 4 September 2021). The markets for three of the most popular food-grade thickening agents are each under the 1 billion USD threshold, namely pectin (888 million USD in 2020) (https: //www.mordorintelligence.com/industry-reports/pectin-market (accessed on 4 September 2021), alginate (728 million USD in 2020) (https://www.grandviewresearch.com/industryanalysis/alginate-market (accessed on 4 September 2021), and carrageenan (742 million USD in 2019) (https://www.grandviewresearch.com/industry-analysis/carrageenan-market (accessed on 4 September 2021). An alternative to these polymers is bacterial xanthan gum, which has an increasing market share, and is currently valued at 576 million USD (in 2020) (https://www.theinsightpartners.com/reports/xanthan-gum-market (accessed on 4 September 2021). Polysaccharides, discovered later and hence newer in the market, have smaller market sizes, such as the bacterial and fungal polysaccharides, e.g., dextran (191 million USD in 2019) (https://www.industryresearch.co/global-dextran-market-18478845 (accessed on 4 September 2021), bacterial cellulose (390 million USD in 2020) (https://www.industryresearch. co/global-microbial-and-bacterial-cellulose-market-18720450 (accessed on 4 September 2021), and pullulan (126 million USD in 2020) (https://www.industryresearch.co/global-pullulanmarket-18823725 (accessed on 4 September 2021). Despite presenting high growth rates, those with very low volumes of sales (strictly nutraceuticals, for instance) present much smaller markets, such as seaweed-based fucoidan (projected 30 million USD in 2020) (https: //www.360researchreports.com/global-fucoidan-market-14030270 (accessed on 4 September 2021) and laminarin (2 million USD in 2019) (https://www.360researchreports.com/globallaminarin-market-18049714 (accessed on 4 September 2021) or fungi-based lentinan (10 million USD in 2019) (https://www.360marketupdates.com/global-lentinan-market-14829124 (accessed on 4 September 2021) (Table 5).

Table 5. Polysacchrides used in markets and their potential values.

Polysaccharides	Potential Market Value in USD/Year	Year	Source
Hyaluronic acid	9.6 billions	2020	https://www.reportsanddata.com/ report-detail/hyaluronic-acid-market (accessed on 4 September 2021).
Heparin	6.5 billions	2020	https://www.alliedmarketresearch. com/heparin-market-A06186 (accessed on 4 September 2021).
Chitin/chitosan	4.2 billions	2021	https://www.bccresearch.com/market- research/plastics/chitin-chitosan- derivatives-markets-report.html (accessed on 4 September 2021).
Chondroitin sulfate	1.2 billions	2020	https://www.grandviewresearch.com/industry-analysis/chondroitin-sulfate-market (accessed on 4 September 2021).

Molecules **2021**, 26, 7068 23 of 36

Table 5. Cont.

Polysaccharides	Potential Market Value in USD/Year	Year	Source
			https:
Agar	239 millions	2020	//www.industryarc.com/Research/ Global-Agar-Market-Research-509553 (accessed on 4 September 2021).
			www.grandviewresearch.com/
Inulin	2.35 billions	2020	industry-analysis/inulin-market
			(accessed on 4 September 2021).
			https://www.mordorintelligence.com/
Pectin	888 millions	2020	industry-reports/pectin-market
			(accessed on 4 September 2021
	<b>20</b> 0 HH	2020	https://www.grandviewresearch.com/
Alginate	728 millions	2020	industry-analysis/alginate-market
			(accessed on 4 September 2021).
ā	E40 :11:	2010	https://www.grandviewresearch.com/
Carrageenan	742 millions	2019	industry-analysis/carrageenan-market
			(accessed on 4 September 2021).
vanthan	191 millions	2010	https://www.industryresearch.co/
xanthan	191 millions	2019	global-dextran-market-18478845
			(accessed on 4 September 2021). www.industryresearch.co/global-
			microbial-and-bacterial-cellulose-
Cellulose	390 millions	2020	market-18720450 (accessed on 4
			September 2021).
			https://www.industryresearch.co/
Pullulan	126 millions	2020	global-pullulan-market-18823725
Tununn	120 1111110113	2020	(accessed on 4 September 2021).
			https://www.360researchreports.com/
Fucoidan	30 millions	2020	global-fucoidan-market-14030270
			(accessed on 4 September 2021).
			https://www.360researchreports.com/
Laminarin	2 millions	2019	global-laminarin-market-18049714
			(accessed on 4 September 2021).
			www.360marketupdates.com/global-
Lentinan	10 millions	2019	lentinan-market-14829124 (accessed on
			4 September 2021).

# 9. Conclusions and Future Perspectives

Polysaccharides have received a great deal of attention during the two last decades notably with the development of glycosciences and glycobiology. To be realistic, despite the large number of bioactive polysaccharides identified and fully or partially characterized, only a small number of them have found significant commercial application. The reasons for this are numerous, but the main ones are probably their costs, their uncertain structures, their polydispersity, the maintenance of their quality, which may be difficult, and the presence of the market of competitive bioactive polysaccharides. For that, except bioactive polysaccharides having highly specific biological activities with no competitor on the market, it is very difficult for new ones to access to a viable business model.

A better understanding and resolving of their structures with modern analytical tools including NMR spectroscopy, mass spectrometry (ESI-IT MS, ESI-Q-TOF MS, MALDI-TOF MS, and others), HPAEC, GPC-MALLS, infrared spectroscopy, etc., opens the way for a better understanding of the relations between structures and biological functions. Accumulating data from these different structural analysis techniques sometimes with hyphenated approaches (LC-MS or LC-NMR) reinforces this opportunity. However, the main drawback that has up to now limited the development of bioactive polysaccharides in several fields of applications, notably the therapeutic one, is their polydispersity, and,

Molecules **2021**, 26, 7068 24 of 36

for the majority of them, the impossibility to propose a full and monodispersed structure. The obtaining of fully purified characterized oligosaccharides could be a good opportunity to solve this problem. Indeed, the correlation of a structure–activity relationship could help to produce biomimetic polysaccharides using non bioactive polysaccharides after their controlled modification using chemistry, biochemistry, or physics. In this field, enzymatic modifications are very promising as they target specific chemical groups to engineer new polysaccharide with a controlled structure from native polysaccharide extracted from plants, algae, and others. Polysaccharides have also received a great deal of attention as bioactive materials for high value applications benefiting from strong development of additive manufacturing. Another opportunity for bioactive polysaccharides is the recent development of technologies allowing the culturing of some microorganisms as new EPS producers. In this field, the photobioreactors technology and the creation of startups exploiting these microorganisms for the production of original polysaccharides is booming. However, their costs of production currently still limit their commercialization, albeit only in the field of cosmetic.

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#### References

1. Hentati, F.; Tounsi, L.; Djomdi, D.; Pierre, G.; Delattre, C.; Ursu, A.V.; Michaud, P. Bioactive polysaccharides from seaweeds. *Molecules* **2020**, 25, 3152. [CrossRef] [PubMed]

- 2. Delattre, C.; Pierre, G.; Laroche, C.; Michaud, P. Production, extraction and characterization of microalgal and cyanobacterial exopolysaccharides. *Biotechnol. Adv.* **2016**, *34*, 1159–1179. [CrossRef] [PubMed]
- 3. De Philippis, R.; Sili, C.; Paperi, R.; Vincenzini, M. Exopolysaccharide-producing cyanobacteria and their possible exploitation: A review. *J. Appl. Phycol.* **2001**, *13*, 293–299. [CrossRef]
- 4. Raposo, M.P.F.J.; Morais, R.M.S.C.; Morais, A.M.M.B. Bioactivity and applications of sulphated polysaccharides from marine microalgae. *Mar. Drugs* **2013**, *11*, 233–252. [CrossRef]
- 5. Li, Z.; He, X.; Liu, F.; Wang, J.; Feng, J. A review of polysaccharides from *Schisandra chinensis* and *Schisandra sphenanthera*: Properties, functions and applications. *Carbohydr. Polym.* **2018**, *184*, 178–190. [CrossRef]
- 6. Yen, H.W.; Hu, I.C.; Chen, C.H.; Ho, S.H.; Lee, D.J.; Chang, J.S. Microalgae-based biorefinery—From biofuels to natural products. *Bioresour. Technol.* **2013**, 135, 166–174. [CrossRef]
- 7. Pereira, S.; Zille, A.; Micheletti, E.; Moradas-Ferreira, P.; De Philippis, R.; Tamagnini, P. Complexity of cyanobacterial exopolysac-charides: Composition, structures, inducing factors and putative genes involved in their biosynthesis and assembly. *FEMS Microbiol. Rev.* 2009, 33, 917–941. [CrossRef]
- 8. Markou, G.; Nerantzis, E. Microalgae for high-value compounds and biofuels production: A review with focus on cultivation under stress conditions. *Biotechnol. Adv* **2013**, *31*, 1532–1542. [CrossRef] [PubMed]
- 9. Kraan, S. Algal polysaccharides, novel applications and outlook. In *Carbohydrates—Comprehensive Studies on GlycoBiology and Glycotechnology*; Chang, C.-F., Ed.; InTech: Rijeka, Croatia, 2012; pp. 489–532.
- 10. Arad, S.M.; Levy-Ontman, O. Red microalgae cell-wall polysaccharides: Biotechnological aspects. *Curr. Opin. Biotechnol.* **2010**, 21, 358–364. [CrossRef] [PubMed]
- 11. Tanna, B.; Mishra, A. Nutraceutical potential of seaweed polysaccharides: Structure, bioactivity, safety, and toxicity. *Compr. Rev. Food Sci. Food Saf.* **2019**, *18*, 817–831. [CrossRef] [PubMed]
- 12. Garcia-Vaquero, M.; Rajauria, G.; O'Doherty, J.V.; Sweeney, T. Polysaccharides from macroalgae: Recent advances, innovative technologies and challenges in extraction and purification. *Food Res. Int.* **2017**, *99*, 1011–1020. [CrossRef]
- 13. Misurcova, L.; Orsavova, J.; Ambrozova, J.V. Algal Polysaccharides and Health. Polysacch. Bioactivity Biotechnol. 2015, 110-144.

Molecules **2021**, 26, 7068 25 of 36

- 14. Chen, L.; Huang, G. Antitumor activity of polysaccharides: An overview. Curr. Drug Targets 2018, 19, 89–96. [CrossRef]
- 15. Xu, S.Y.; Huang, X.; Cheong, K.L. Recent advances in marine algae polysaccharides: Isolation, structure, and activities. *Mar. Drugs* **2017**, *15*, 388. [CrossRef]
- 16. Kakar, M.U.; Naveed, M.; Saeed, M.; Zhao, S.; Rasheed, M.; Firdos, S.; Dai, R. A review on structure, extraction, and Biol.ogical activities of polysaccharides isolated from *Cyclocarya paliurus* (Batalin) Iljinskaja. *Int. J. Biol. Macromol.* **2020**, *156*, 420–429. [CrossRef] [PubMed]
- 17. He, R.; Ye, J.; Zhao, Y.; Su, W. Partial characterization, antioxidant and antitumor activities of polysaccharides from *Philomycus bilineatus*. *Int. J. Biol. Macromol.* **2014**, *65*, 573–580. [CrossRef] [PubMed]
- 18. Zhang, Q.; Yu, J.; Zhang, L.; Hu, M.; Xu, Y.; Su, W. Extraction, characterization, and Biol.ogical activity of polysaccharides from *Sophora flavescens* Ait. *Int. J. Biol. Macromol.* **2016**, 93, 459–467. [CrossRef] [PubMed]
- 19. Wang, J.; Hu, S.; Nie, S.; Yu, Q.; Xie, M. Reviews on Mechanisms of in vitro Antioxidant Activity of Polysaccharides. *Oxid. Med. Cell. Longev.* **2016**, 2016, 5692852. [CrossRef] [PubMed]
- 20. Radzki, W.; Ziaja-Sołtys, M.; Nowak, J.; Rzymowska, J.; Dominik, J.; Sławińska, A.; Michalak-Majewska, M.; Zalewska-Korona, M.; Kuczumow, A. Effect of processing on the content and Biol.ogical activity of polysaccharides from *Pleurotus ostreatus* mushroom. *LWT Food Sci. Technol.* **2016**, *66*, 27–33. [CrossRef]
- 21. Han, L.; Suo, Y.; Yang, Y.; Meng, J.; Hu, N. Optimization, characterization, and Biol.ogical activity of polysaccharides from *Berberis dasystachya* Maxim. *Int. J. Biol. Macromol.* **2016**, *85*, 655–666. [CrossRef] [PubMed]
- 22. Tan, L.H.; Zhang, D.; Yu, B.; Zhao, S.-P.; Wang, J.-W.; Yao, L.; Cao, W.-G. Antioxidant activity and optimization of extraction of polysaccharide from the roots of *Dipsacus asperoides*. *Int. J. Biol. Macromol.* **2015**, *81*, 332–339. [CrossRef]
- 23. Yao, Y.L.; Shu, C.; Feng, G.; Wang, Q.; Yan, Y.-Y.; Yi, Y.; Wang, H.-X.; Zhang, X.-F.; Wang, L.-M. Polysaccharides from *Pyracantha fortuneana* and its Biol.ogical activity. *Int. J. Biol. Macromol.* **2020**, *150*, 1162–1174. [CrossRef]
- 24. Chen, F.; Huang, G. Antioxidant activity of polysaccharides from different sources of ginseng. *Int. J. Biol. Macromol.* **2019**, 125, 906–908. [CrossRef]
- 25. Jayaprakasha, G.K.; Singh, R.P.; Sakariah, K.K. Antioxidant activity of grape seed (*Vitis vinifera*) extracts on peroxidation models In Vitro. *Food Chem.* **2001**, *73*, 285–290. [CrossRef]
- 26. Meng, L.; Sun, S.; Li, R.; Shen, Z.; Wang, P.; Jiang, X. Antioxidant activity of polysaccharides produced by *Hirsutella* sp. and relation with their chemical characteristics. *Carbohydr. Polym.* **2015**, *117*, 452–457. [CrossRef] [PubMed]
- 27. Samanta, S.; Nandi, A.K.; Sen, I.K.; Maity, P.; Pattanayak, M.; Devi, K.S.P.; Khatua, S.; Maiti, T.K.; Acharya, K.; Islam, S.S. Studies on antioxidative and immunostimulating fucogalactan of the edible mushroom *Macrolepiota dolichaula*. *Carbohydr. Res.* **2015**, 413, 22–29. [CrossRef] [PubMed]
- 28. Zeng, H.; Zhang, Y.; Zhao, Y.; Tian, Y.; Miao, S.; Zheng, B. Extraction optimization, structure and antioxidant activities of *Fortunella margarita* Swingle polysaccharides. *Int. J. Biol. Macromol.* **2015**, 74, 232–242. [CrossRef] [PubMed]
- 29. Wang, Z.-J.; Xu, W.; Liang, J.-W.; Wang, C.-S.; Kang, Y. Effect of fucoidan on B16 murine melanoma cell melanin formation and apoptosis. *Afr. J. Tradit. Complement. Altern. Med.* **2017**, *14*, 149–155. [CrossRef]
- 30. Rozi, P.; Abuduwaili, A.; Mutailifu, P.; Gao, Y.; Rakhmanberdieva, R.; Aisa, H.A.; Yili, A. Sequential extraction, characterization, and antioxidant activity of polysaccharides from *Fritillaria pallidiflora* Schrenk. *Int. J Biol. Macromol.* **2019**, *131*, 97–106. [CrossRef] [PubMed]
- 31. Kang, Q.; Chen, S.; Li, S.; Wang, B.; Liu, X.; Hao, L.; Lu, J. Comparison on characterization and antioxidant activity of polysaccharides from *Ganoderma lucidum* by ultrasound and conventional extraction. *Int. J. Biol. Macromol.* **2019**, *124*, 1137–1144. [CrossRef]
- 32. Wang, Y.; Jia, J.; Ren, X.; Li, B.; Zhang, Q. Extraction, preliminary characterization and In Vitro antioxidant activity of polysaccharides from *Oudemansiella radicata* mushroom. *Int. J Biol. Macromol.* **2018**, 120, 1760–1769. [CrossRef] [PubMed]
- 33. Zhang, L.; Hu, Y.; Duan, X.; Tang, T.; Shen, Y.; Hu, B.; Liu, Y. Characterization and antioxidant activities of polysaccharides from thirteen boletus mushrooms. *Int. J. Biol. Macromol.* **2018**, *113*, 1–7. [CrossRef]
- 34. Schepetkin, I.A.; Faulkner, C.L.; Nelson-Overton, L.K.; Wiley, J.A.; Quinn, M.T. Macrophage immunomodulatory activity of polysaccharides isolated from *Juniperus scopolorum*. *Int. Immunopharmacol.* **2005**, *5*, 1783–1799. [CrossRef] [PubMed]
- 35. Nie, C.; Zhu, P.; Ma, S.; Wang, M.; Hu, Y. Purification, characterization and immunomodulatory activity of polysaccharides from stem lettuce. *Carbohydr. Polym.* **2018**, *188*, 236–242. [CrossRef]
- 36. Zeng, Y.J.; Yang, H.R.; Wu, X.L.; Peng, F.; Huang, Z.; Pu, L.; Lou, W.Y. Structure and immunomodulatory activity of polysaccharides from *Fusarium solani* DO7 by solid-state fermentation. *Int. J. Biol. Macromol.* **2019**, *137*, 568–575. [CrossRef]
- 37. Meng, M.; Cheng, D.; Han, L.; Chen, Y.; Wang, C. Isolation, purification, structural analysis and immunostimulatory activity of water-soluble polysaccharides from *Grifola Frondosa* fruiting body. *Carbohydr. Polym.* **2017**, 157, 1134–1143. [CrossRef]
- 38. Wang, Y.; Zhang, Y.; Shao, J.; Ren, X.; Jia, J.; Li, B. Study on the immunomodulatory activity of a novel polysaccharide from the lichen *Umbilicaria esculenta*. *Int. J Biol. Macromol.* **2019**, *121*, 846–851. [CrossRef]
- 39. Yin, M.; Zhang, Y.; Li, H. Advances in research on immunoregulation of macrophages by plant polysaccharides. *Front. Immunol.* **2019**, *10*, 145. [CrossRef] [PubMed]
- 40. Hu, Z.; Zhou, H.; Li, Y.; Wu, M.; Yu, M.; Sun, X. Optimized purification process of polysaccharides from Carex meyeriana Kunth by macroporous resin, its characterization and immunomodulatory activity. *Int. J. Biol. Macromol.* **2019**, *132*, 76–86. [CrossRef]

Molecules **2021**, 26, 7068 26 of 36

41. Ma, L.; Jiao, K.; Luo, L.; Xiang, J.; Fan, J.; Zhang, X.; Zhu, W. Characterization and macrophage immunomodulatory activity of two polysaccharides from the flowers of *Paeonia suffruticosa* Andr. *Int. J Biol. Macromol.* **2019**, 124, 955–962. [CrossRef] [PubMed]

- 42. Zheng, T.; Gu, D.; Wang, X.; Shen, X.; Yan, L.; Zhang, W.; Fan, J. Purification, characterization and immunomodulatory activity of polysaccharides from *Leccinum crocipodium* (Letellier.) Watliag. *Int. J. Biol. Macromol.* **2020**, *148*, 647–656. [CrossRef]
- 43. Huang, K.; Li, Y.; Tao, S.; Wei, G.; Huang, Y.; Chen, D.; Wu, C. Purification, characterization and Biol.ogical activity of polysaccharides from *Dendrobium officinale*. *Molecules* **2016**, 21, 701. [CrossRef] [PubMed]
- 44. Schepetkin, I.A.; Quinn, M.T. Botanical polysaccharides: Macrophage immunomodulation and therapeutic potential. *Int. Immunopharmacol.* **2006**, *6*, 317–333. [CrossRef] [PubMed]
- 45. Wang, N.; Zhang, X.; Wang, S.; Guo, Q.; Li, Z.; Liu, H.; Wang, C. Structural characterisation and immunomodulatory activity of polysaccharides from white asparagus skin. *Carbohydr. Polym.* **2020**, 227, 115314. [CrossRef] [PubMed]
- 46. Huang, L.; Shen, M.; Morris, G.A.; Xie, J. Sulfated polysaccharides: Immunomodulation and signaling mechanisms. *Trends Food Sci. Technol.* **2019**, 92, 1–11. [CrossRef]
- 47. Li, J.; Wang, D.; Xing, X.; Cheng, T.J.R.; Liang, P.H.; Bulone, V.; Hsieh, Y.S. Structural analysis and Biol.ogical activity of cell wall polysaccharides extracted from Panax ginseng marc. *Int. J. Biol.* **2019**, *135*, 29–37.
- 48. Li, S.; Gao, A.; Dong, S.; Chen, Y.; Sun, S.; Lei, Z.; Zhang, Z. Purification, antitumor and immunomodulatory activity of polysaccharides from soybean residue fermented with *Morchella esculenta*. *Int. J. Biol. Macromol.* **2017**, *96*, 26–34. [CrossRef]
- 49. Wu, Z.; Li, H.; Wang, Y.; Yang, D.; Tan, H.; Zhan, Y.; Chen, G. Optimization extraction, structural features and antitumor activity of polysaccharides from *Z. jujuba cv.* Ruoqiangzao seeds. *Int. J. Biol. Macromol.* **2019**, *135*, 1151–1161. [CrossRef]
- 50. Yang, X.; Ji, H.; Feng, Y.; Yu, J.; Liu, A. Structural Characterization and Antitumor Activity of Polysaccharides from *Kaempferia galanga* L. *Oxidative Med. Cell. Longev.* **2018**, 2018, 1–10. [CrossRef]
- 51. Yu, J.; Ji, H.; Yang, Z.; Liu, A. Relationship between structural properties and antitumor activity of Astragalus polysaccharides extracted with different temperatures. *Int. J. Biol. Macromol.* **2019**, 124, 469–477. [CrossRef]
- 52. Zheng, F.; Chen, L.; Zhang, P.; Zhou, J.; Lu, X.; Tian, W. Carbohydrate Polymers Exhibit Great Potential as Effective Elicitors in Organic Agriculture: A Review. *Carbohydr. Polym.* **2020**, 230, 115637. [CrossRef]
- 53. Ahmad, B.; Khan, M.M.A.; Jahan, A.; Shabbir, A.; Jaleel, H. Increased Production of Valuable Secondary Products in Plants by Leaf Applied Radiation-Processed Polysaccharides. *Int. J. Biol. Macromol.* **2020**, *164*, 286–294. [CrossRef] [PubMed]
- 54. Vera, J.; Castro, J.; Gonzalez, A.; Moenne, A. Seaweed Polysaccharides and Derived Oligosaccharides Stimulate Defense Responses and Protection against Pathogens in Plants. *Mar. Drugs* **2011**, *9*, 2514–2525. [CrossRef]
- 55. González, A.; Castro, J.; Vera, J.; Moenne, A. Seaweed Oligosaccharides Stimulate Plant Growth by Enhancing Carbon and Nitrogen Assimilation, Basal Metabolism, and Cell Division. *J. Plant Growth Regul.* **2013**, 32, 443–448. [CrossRef]
- 56. Drira, M.; Ben Mohamed, J.; Ben Hlima, H.; Hentati, F.; Michaud, P.; Abdelkafi, S.; Fendri, I. Improvement of *Arabidopsis thaliana* salt tolerance using a polysaccharidic extract from the brown algae *Padina pavonica*. *Algal. Res.* **2021**, *56*, 102324. [CrossRef]
- 57. Andreeva, A.; Budenkova, E.; Babich, O.; Sukhikh, S.; Dolganyuk, V.; Michaud, P.; Ivanova, S. Influence of Carbohydrate Additives on the Growth Rate of Microalgae Biomass with an Increased Carbohydrate Content. *Mar. Drugs* **2021**, *19*, 381. [CrossRef]
- 58. Dolganyuk, V.; Andreeva, A.; Budenkova, E.; Sukhikh, S.; Babich, O.; Ivanova, S.; Prosekov, A.; Ulrikh, E. Study of Morphological Features and Determination of the Fatty Acid Composition of the Microalgae Lipid Complex. *Biomolecules* **2020**, *10*, 1571. [CrossRef]
- 59. Dey, P.; Ramanujam, R.; Venkatesan, G.; Nagarathnam, R. Sodium Alginate Potentiates Antioxidant Defense and PR Proteins against Early Blight Disease Caused by *Alternaria Solani* in *Solanum Lycopersicum* Linn. *PLoS ONE* **2019**, *14*, 1–26.
- 60. Bouissil, S.; El Alaoui-Talibi, Z.; Pierre, G.; Michaud, P.; El Modafar, C.; Delattre, C. Use of Alginate Extracted from Moroccan Brown Algae to Stimulate Natural Defense in Date Palm Roots. *Molecules* **2020**, 25, 720. [CrossRef]
- 61. Mani, S.D.; Govindan, M.; Muthamilarasan, M.; Nagarathnam, R. A Sulfated Polysaccharide κ-Carrageenan Induced Antioxidant Defense and Proteomic Changes in Chloroplast against Leaf Spot Disease of Tomato. *J. Appl. Phycol.* **2021**, 33, 2667–2681. [CrossRef]
- 62. Ghannam, A.; Abbas, A.; Alek, H.; Al-Waari, Z.; Al-Ktaifani, M. Enhancement of Local Plant Immunity against Tobacco Mosaic Virus Infection after Treatment with Sulphated-Carrageenan from Red Alga (*Hypnea musciformis*). *Physiol. Mol. Plant. Pathol.* **2013**, 84, 19–27. [CrossRef]
- Sangha, J.S.; Kandasamy, S.; Khan, W.; Bahia, N.S.; Singh, R.P.; Critchley, A.T.; Prithiviraj, B. λ-Carrageenan Suppresses Tomato Chlorotic Dwarf Viroid (TCDVd) Replication and Symptom Expression in Tomatoes. *Mar. Drugs* 2015, 13, 2875–2889. [CrossRef]
- 64. Klarzynski, O.; Descamps, V.; Plesse, B.; Yvin, J.C.; Kloareg, B.; Fritig, B. Sulfated Fucan Oligosaccharides Elicit Defense Responses in Tobacco and Local and Systemic Resistance against Tobacco Mosaic Virus. *Mol. Plant Microbe Interact.* **2003**, *16*, 115–122. [CrossRef]
- 65. Rachidi, F.; Benhima, R.; Kasmi, Y.; Sbabou, L.; Arroussi, H.El. Evaluation of Microalgae Polysaccharides as Biostimulants of Tomato Plant Defense Using Metabolomics and Biochemical Approaches. *Sci. Rep.* **2021**, *11*, 930. [CrossRef] [PubMed]
- 66. Drira, M.; Elleuch, J.; Hlima, H.B.; Hentati, F.; Gardarin, C.; Rihouey, C.; Cerf, D.L.; Michaud, P.; Abdelkafi, S.; Fendri, I. Optimization of Exopolysaccharides Production by *Porphyridium sordidum* and Their Potential to Induce Defense Responses in *Arabidopsis thaliana* against *Fusarium oxysporum*. *Biomolecules* **2021**, *11*, 282. [CrossRef] [PubMed]

Molecules **2021**, 26, 7068 27 of 36

67. Liu, H.; Zhang, Y.H.; Yin, H.; Wang, W.X.; Zhao, X.M.; Du, Y.G. Alginate Oligosaccharides Enhanced *Triticum Aestivum L.* Tolerance to Drought Stress. *Plant Physiol. Biochem.* **2013**, 62, 33–40. [CrossRef]

- 68. Zhuo, R.; Li, B.; Tian, S. Alginate Oligosaccharide Improves Resistance to Postharvest Decay and Quality in Kiwifruit (*Actinidia deliciosa* Cv. Bruno). *Hortic. Plant J.* **2021**. [CrossRef]
- 69. Zhang, S.; Tang, W.; Jiang, L.; Hou, Y.; Yang, F.; Chen, W.; Li, X. Elicitor Activity of Algino-Oligosaccharide and Its Potential Application in Protection of Rice Plant (*Oryza Saliva* L.) against *Magnaporthe grisea*. *Biotechnol*. *Biotechnol*. *Equip*. **2015**, 29, 646–652. [CrossRef]
- 70. Golkar, P.; Taghizadeh, M.; Noormohammadi, A. Effects of Sodium Alginate Elicitation on Secondary Metabolites and Antioxidant Activity of Safflower Genotypes under in Vitro Salinity Stress. *In Vitro Cell. Dev. Biol. Plant* **2019**, *55*, 527–538. [CrossRef]
- 71. Zhang, C.; Howlader, P.; Liu, T.; Sun, X.; Jia, X.; Zhao, X.; Shen, P.; Qin, Y.; Wang, W.; Yin, H. Alginate Oligosaccharide (AOS) Induced Resistance to Pst DC3000 via Salicylic Acid-Mediated Signaling Pathway in *Arabidopsis thaliana*. *Carbohydr. Polym.* **2019**, 225, 115221. [CrossRef]
- 72. Li, J.; Wang, X.; Lin, X.; Yan, G.; Liu, L.; Zheng, H.; Zhao, B.; Tang, J.; Guo, Y.D. Alginate-Derived Oligosaccharides Promote Water Stress Tolerance in Cucumber (*Cucumis Sativus* L.). *Plant Physiol. Biochem.* **2018**, 130, 80–88. [CrossRef] [PubMed]
- 73. Colla, G.; Rouphael, Y. Microalgae: New Source of Plant Biostimulants. Agronomy 2020, 10, 1240. [CrossRef]
- 74. Ronga, D.; Biazzi, E.; Parati, K.; Carminati, D.; Carminati, E.; Tava, A. Microalgal Biostimulants and Biofertilisers in Crop Productions. *Agronomy* **2019**, *9*, 192. [CrossRef]
- 75. El Arroussi, H.; Elmernissi, N.; Benhima, R.; El Kadmiri, I.M.; Bendaou, N.; Smouni, A.; Wahby, I. Microalgae Polysaccharides a Promising Plant Growth Biostimulant. *J. Algal Biomass. Utln.* **2016**, *7*, 55–63.
- Yang, J.; Shen, Z.; Sun, Z.; Wang, P.; Jiang, X. Growth Stimulation Activity of Alginate-Derived Oligosaccharides with Different Molecular Weights and Mannuronate/Guluronate Ratio on Hordeum vulgare L. J. Plant Growth Regul. 2021, 40, 91–100. [CrossRef]
- 77. Salachna, P.; Grzeszczuk, M.; Meller, E.; Soból, M. Oligo-Alginate with Low Molecular Mass Improves Growth and Physiological Activity of *Eucomis autumnalis* under Salinity Stress. *Molecules* **2018**, 23, 812. [CrossRef]
- 78. Castro, J.; Vera, J.; González, A.; Moenne, A. Oligo-Carrageenans Stimulate Growth by Enhancing Photosynthesis, Basal Metabolism, and Cell Cycle in Tobacco Plants (Var. Burley). *J. Plant Growth Regul.* **2012**, *31*, 173–185. [CrossRef]
- 79. Saucedo, S.; Contreras, R.A.; Moenne, A.; Oligo-Carrageenan Kappa Increases, C.N.S. Assimilation, Auxin and Gibberellin Contents, and Growth in Pinus Radiata Trees. *J. For. Res.* **2015**, *26*, 635–640. [CrossRef]
- 80. Albuquerque, I.R.; Cordeiro, S.L.; Gomes, D.L.; Dreyfuss, J.L.; Filgueira, L.G.A.; Leite, E.L.; Nader, H.B.; Rocha, H.A.O. Evaluation of Anti-Nociceptive and Anti-Inflammatory Activities of a Heterofucan from Dictyota menstrualis. *Mar. Drugs* **2013**, *11*, 2722–2740. [CrossRef] [PubMed]
- 81. Fedorov, S.N.; Ermakova, S.P.; Zvyagintseva, T.N.; Stonik, V.A. Anticancer and cancer preventive properties of marine polysac-charides: Some results and prospects. *Mar. Drugs* **2013**, *11*, 4876–4901. [CrossRef]
- 82. Usman, A.; Khalid, S.; Usman, A.; Hussain, Z.; Wang, Y. Chapter 5—Algal polysaccharides, novel application, and outlook. In *Algae Based Polymers, Blends, and Composites*; Zia, K.M., Zuber, M., Ali, M., Eds.; Elsevier: Amsterdam, The Netherlands, 2017; pp. 115–153.
- 83. D'Ayala, G.G.; Malinconico, M.; Laurienzo, P. Marine derived polysaccharides for biomedical applications: Chemical modification approaches. *Molecules* **2008**, *13*, 2069–2106. [CrossRef]
- 84. Li, P.; Wang, S.; Chen, H.; Zhang, S.; Yu, S.; Li, Y.; Cui, M.; Pan, W.; Yang, X. A novel ion-activated in situ gelling ophthalmic delivery system based on κ-carrageenan for acyclovir. *Drug Dev. Ind. Pharm.* **2018**, 44, 829–836. [CrossRef] [PubMed]
- 85. Obaidat, R.M.; Alnaief, M.; Mashaqbeh, H. Investigation of Carrageenan Aerogel Microparticles as a Potential Drug Carrier. *AAPS Pharm. Sci. Tech.* **2018**, *19*, 2226–2236. [CrossRef] [PubMed]
- 86. Farag, M.M.S.; Moghannem, S.A.M.; Shehabeldine, A.M.; Azab, M.S. Antitumor effect of exopolysaccharide produced by *Bacillus mycoides*. *Microb. Pathog.* **2020**, *140*, 103947. [CrossRef] [PubMed]
- 87. Lim, H.-P.; Ooi, C.-W.; Tey, B.-T.; Chan, E.-S. Controlled delivery of oral insulin aspart using pH-responsive alginate/κ-carrageenan composite hydrogel beads. *React. Funct. Polym.* **2017**, *120*, 20–29. [CrossRef]
- 88. Popa, E.G.; Gomes, M.E.; Reis, R.L. Cell delivery systems using alginate—carrageenan hydrogel beads and fibers for regenerative medicine applications. *Biomacromolecules* **2011**, *12*, 3952–3961. [CrossRef] [PubMed]
- 89. Yermak, I.M.; Gorbach, V.I.; Karnakov, I.A.; Davydova, V.N.; Pimenova, E.A.; Chistyulin, D.A.; Isakov, V.V.; Glazunov, V.P. Carrageenan gel beads for echinochrome inclusion: Influence of structural features of carrageenan. *Carbohydr. Polym.* **2021**, 272, 118479. [CrossRef]
- 90. Huang, Y.-C.; Lam, U.-I. Chitosan/Fucoidan pH Sensitive Nanoparticles for Oral Delivery System. *J. Chin. Chem. Soc.* **2011**, *58*, 779–785. [CrossRef]
- 91. Don, T.-M.; Chang, W.-J.; Jheng, P.-R.; Huang, Y.-C.; Chuang, E.-Y. Curcumin-laden dual-targeting fucoidan/chitosan nanocarriers for inhibiting brain inflammation via intranasal delivery. *Int. J. Biol. Macromol.* **2021**, *181*, 835–846. [CrossRef] [PubMed]
- 92. Yoshizawa, Y.; Enomoto, A.; Todoh, H.; Ametani, A.; Kaminogawa, S. Activation of Murine Macrophages by Polysaccharide Fractions from Marine Algae (*Porphyra yezoensis*). *Biosci. Biotechnol. Biochem.* **1993**, 57, 1862–1866. [CrossRef] [PubMed]
- 93. Olasehinde, T.A.; Mabinya, L.V.; Olaniran, A.O.; Okoh, A.I. Chemical characterization, antioxidant properties, cholinesterase inhibitory and anti-amyloidogenic activities of sulfated polysaccharides from some seaweeds. *Bioact. Carbohydr. Diet. Fibre* 2019, 18, 100182. [CrossRef]

Molecules **2021**, 26, 7068 28 of 36

94. Sanjeewa, K.K.A.; Kang, N.; Ahn, G.; Jee, Y.; Kim, Y.-T.; Jeon, Y.-J. Bioactive potentials of sulfated polysaccharides isolated from brown seaweed *Sargassum* spp in related to human health applications: A review. *Food Hydrocoll.* **2018**, *81*, 200–208. [CrossRef]

- 95. Wijesekara, I.; Pangestuti, R.; Kim, S.K. Biological activities and potential health benefits of sulfated polysaccharides derived from marine algae. *Carbohydr Polym* **2011**, *84*, 14–21. [CrossRef]
- 96. Zheng, L.-X.; Chen, X.-Q.; Cheong, K.-L. Current trends in marine algae polysaccharides: The digestive tract, microbial catabolism, and prebiotic potential. *Int. J. Biol. Macromol.* **2020**, *151*, 344–354. [CrossRef] [PubMed]
- 97. Caccamo, M.T.; Gugliandolo, C.; Zammuto, V.; Magazù, S. Thermal properties of an exopolysaccharide produced by a marine thermotolerant Bacillus licheniformis by ATR-FTIR spectroscopy. *Int. J. Biol. Macromol.* **2020**, *145*, 77–83. [CrossRef] [PubMed]
- 98. Casillo, A.; Lanzetta, R.; Parrilli, M.; Corsaro, M.M. Exopolysaccharides from Marine and Marine Extremophilic Bacteria: Structures, Properties, Ecological Roles and Applications. *Mar. Drugs* **2018**, *16*, 69. [CrossRef]
- 99. Alencar, P.O.C.; Lima, G.C.; Barros, F.C.N.; Costa, L.E.C.; Ribeiro, C.V.P.E.; Sousa, W.M.; Sombra, V.G.; Abreu, C.M.W.S.; Abreu, E.S.; Pontes, E.O.B.; et al. A novel antioxidant sulfated polysaccharide from the algae *Gracilaria caudata*: In vitro and in vivo activities. *Food Hydrocoll.* **2019**, *90*, 28–34. [CrossRef]
- 100. Barahona, T.; Chandía, N.P.; Encinas, M.V.; Matsuhiro, B.; Zúñiga, E.A. Antioxidant capacity of sulfated polysaccharides from seaweeds. A kinetic approach. *Food Hydrocoll.* **2011**, 25, 529–535. [CrossRef]
- 101. Cao, S.; He, X.; Qin, L.; He, M.; Yang, Y.; Liu, Z.; Mao, W. Anticoagulant and Antithrombotic Properties in Vitro and in Vivo of a Novel Sulfated Polysaccharide from Marine Green Alga *Monostroma nitidum*. *Mar. Drugs* **2019**, *17*, 247. [CrossRef]
- 102. Ciancia, M.; Quintana, I.; Cerezo, A.S. Overview of Anticoagulant Activity of Sulfated Polysaccharides from Seaweeds in Relation to their Structures, Focusing on those of Green Seaweeds. *Curr. Med. Chem.* **2010**, *17*, 2503–2529. [CrossRef] [PubMed]
- 103. Gheda, S.; El-Sheekh, M.; Abou-Zeid, A. In vitro anticancer activity of polysaccharide extracted from red alga *Jania rubens* against breast and colon cancer cell lines. *Asian Pac. J. Trop. Med.* **2018**, *11*, 583–589.
- 104. Krylova, N.V.; Ermakova, S.P.; Lavrov, V.F.; Leneva, I.A.; Kompanets, G.G.; Iunikhina, O.V.; Nosik, M.N.; Ebralidze, L.K.; Falynskova, I.N.; Silchenko, A.S.; et al. The Comparative Analysis of Antiviral Activity of Native and Modified Fucoidans from Brown Algae Fucus evanescens In Vitro and In Vivo. *Mar. Drugs* 2020, *18*, 224. [CrossRef] [PubMed]
- 105. Bouhlal, R.; Haslin, C.; Chermann, J.-C.; Colliec-Jouault, S.; Sinquin, C.; Simon, G.; Cerantola, S.; Riadi, H.; Bourgougnon, N. Antiviral activities of sulfated polysaccharides isolated from *Sphaerococcus coronopifolius* (Rhodophytha, Gigartinales) and Boergeseniella thuyoides (Rhodophyta, Ceramiales). *Mar. Drugs* **2011**, *9*, 1187–1209. [CrossRef]
- 106. Barboríková, J.; Šutovská, M.; Kazimierová, I.; Jošková, M.; Fraňová, S.; Kopecký, J.; Capek, P. Extracellular polysaccharide produced by *Chlorella vulgaris*—Chemical characterization and anti-asthmatic profile. *Int. J. Biol. Macromol.* **2019**, *135*, 1–11. [CrossRef] [PubMed]
- 107. Vo, T.-S.; Ngo, D.-H.; Kim, S.-K. Potential targets for anti-inflammatory and anti-allergic activities of marine algae: An overview. *Inflamm. Allergy Drug Targets* **2012**, *11*, 90–101. [CrossRef] [PubMed]
- 108. Dalavi, P.A.; Venkatesan, J.; Rani, V.; Anil, S. *Marine Polysaccharides Systems for Drug Delivery Applications BT—Advanced Biopolymeric Systems for Drug Delivery*; Nayak, A.K., Hasnain, M.S., Eds.; Springer: Cham, Switzerland, 2020; pp. 373–386.
- 109. Grenha, A.; Gomes, M.E.; Rodrigues, M.; Santo, V.E.; Mano, J.F.; Neves, N.M.; Reis, R.L. Development of new chitosan/carrageenan nanoparticles for drug delivery applications. *J. Biomed. Mater. Res.* **2010**, *92*, 1265–1272. [CrossRef] [PubMed]
- 110. Sarıyer, S.; Duranoğlu, D.; Doğan, Ö.; Küçük, İ. pH-responsive double network alginate/kappa-carrageenan hydrogel beads for controlled protein release: Effect of pH and crosslinking agent. *J. Drug Deliv. Sci. Technol.* **2020**, *56*, 101551. [CrossRef]
- 111. Fahmy, H.M.; Aly, A.A.; Sayed, S.M.; Abou-Okeil, A. K-carrageenan/Na-alginate wound dressing with sustainable drug delivery properties. *Polym. Adv. Technol.* **2021**, *32*, 1793–1801. [CrossRef]
- 112. Manivasagan, P.; Bharathiraja, S.; Bui, N.Q.; Jang, B.; Oh, Y.-O.; Lim, I.G.; Oh, J. Doxorubicin-loaded fucoidan capped gold nanoparticles for drug delivery and photoacoustic imaging. *Int. J. Biol. Macromol.* **2016**, *91*, 578–588. [CrossRef]
- 113. Lai, Y.-H.; Chiang, C.-S.; Hsu, C.-H.; Cheng, H.-W.; Chen, S.-Y. Development and Characterization of a Fucoidan-Based Drug Delivery System by Using Hydrophilic Anticancer Polysaccharides to Simultaneously Deliver Hydrophobic Anticancer Drugs. *Biomolecules* **2020**, *10*, 970. [CrossRef]
- 114. Wang, P.; Kankala, R.K.; Chen, B.; Long, R.; Cai, D.; Liu, Y.; Wang, S. Poly-allylamine hydrochloride and fucoidan-based self-assembled polyelectrolyte complex nanoparticles for cancer therapeutics. *J. Biomed. Mater. Res. Part. A* **2019**, 107, 339–347. [CrossRef]
- 115. Purnama, A.; Aid-Launais, R.; Haddad, O.; Maire, M.; Mantovani, D.; Letourneur, D.; Hlawaty, H.; Le Visage, C. Fucoidan in a 3D scaffold interacts with vascular endothelial growth factor and promotes neovascularization in mice. *Drug Deliv. Transl. Res.* **2015**, 5, 187–197. [CrossRef]
- 116. Vanavil, B.; Selvaraj, K.; Aanandhalakshmi, R.; Usha, S.K.; Arumugam, M. Bioactive and thermostable sulphated polysaccharide from *Sargassum swartzii* with drug delivery applications. *Int. J. Biol. Macromol.* **2020**, *153*, 190–200.
- 117. Phan, N.H.; Ly, T.T.; Pham, M.N.; Luu, T.D.; Vo, T.V.; Tran, P.H.; Tran, T.T. A Comparison of Fucoidan Conjugated to Paclitaxel and Curcumin for the Dual Delivery of Cancer Therapeutic Agents. *Anticancer. Agents Med. Chem.* **2018**, *18*, 1349–1355. [CrossRef] [PubMed]

Molecules **2021**, 26, 7068 29 of 36

118. Santo, V.E.; Frias, A.M.; Carida, M.; Cancedda, R.; Gomes, M.E.; Mano, J.F.; Reis, R.L. Carrageenan-Based Hydrogels for the Controlled Delivery of PDGF-BB in Bone Tissue Engineering Applications. *Biomacromolecules* **2009**, *10*, 1392–1401. [CrossRef] [PubMed]

- 119. Rocha, P.M.; Santo, V.E.; Gomes, M.E.; Reis, R.L.; Mano, J.F. Encapsulation of adipose-derived stem cells and transforming growth factor-β1 in carrageenan-based hydrogels for cartilage tissue engineering. *J. Bioact. Compat. Polym.* **2011**, *26*, 493–507. [CrossRef]
- 120. Silva, T.H.; Alves, A.; Popa, E.; Reys, L.; Gomes, M.E.; Sousa, R.A.; Silva, S.S.; Mano, J.F.; Reis, R.L. Marine algae sulfated polysaccharides for tissue engineering and drug delivery approaches. *Biomatter* **2012**, *2*, 278–289. [CrossRef]
- 121. BeMiller, J.N. Carrageenans. In *Carbohydrate Chemistry for Food Scientists*, 3rd ed.; BeMiller, J.N., Ed.; AACC International Press: London, UK, 2019; pp. 279–291.
- 122. Mohamadnia, Z.; Zohuriaan-Mehr, M.J.; Kabiri, K.; Jamshidi, A.; Mobedi, H. Ionically cross-linked carrageenan-alginate hydrogel beads. *J. Biomater. Sci. Polym. Ed.* **2008**, *19*, 47–59. [CrossRef]
- 123. Hariyadi, D.M.; Purwanti, T.; Maulydia, D.; Estherline, C.A.; Hendradi, E.; Rahmadi, M. Performance and drug deposition of kappa-carrageenan microspheres encapsulating ciprofloxacin HCl: Effect of polymer concentration. *J. Adv. Pharm. Technol. Res.* **2021**, *12*, 242–249. [PubMed]
- 124. Yuan, H.; Song, J.; Zhang, W.; Li, X.; Li, N.; Gao, X. Antioxidant activity and cytoprotective effect of kappa-carrageenan oligosaccharides and their different derivatives. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 1329–1334. [CrossRef] [PubMed]
- 125. Zhou, G.; Sun, Y.; Xin, H.; Zhang, Y.; Li, Z.; Xu, Z. In vivo antitumor and immunomodulation activities of different molecular weight lambda-carrageenans from Chondrus ocellatus. *Pharmacol. Res.* **2004**, *50*, 47–53. [CrossRef]
- 126. Panlasigui, L.N.; Baello, O.Q.; Dimatangal, J.M.; Dumelod, B.D. Blood cholesterol and lipid-lowering effects of carrageenan on human volunteers. *Asia Pac. J. Clin. Nutr.* **2003**, *12*, 209–214.
- 127. Hoffman, R.C. arrageenans inhibit growth-factor binding. Biochem. J. 1993, 289, 331–334. [CrossRef] [PubMed]
- 128. Chen, H.-M.; Gao, Y.; Yan, X.-J. Carrageenan oligosaccharides inhibit growth-factor binding and heparanase activity. *Yao Xue Xue Bao* **2011**, *46*, 280–284.
- 129. Thompson, K.D.; Dragar, C. Antiviral activity of Undaria pinnatifida against herpes simplex virus. *Phytother. Res.* **2004**, *18*, 551–555. [CrossRef] [PubMed]
- 130. Tavakoli, S.; Kharaziha, M.; Kermanpur, A.; Mokhtari, H. Sprayable and injectable visible-light Kappa-carrageenan hydrogel for in-situ soft tissue engineering. *Int. J. Biol. Macromol.* **2019**, *138*, 590–601. [CrossRef] [PubMed]
- 131. Desai, P.D.; Dave, A.M.; Devi, S. Entrapment of lipase into K-carrageenan beads and its use in hydrolysis of olive oil in biphasic system. *J. Mol. Catal. B Enzym.* **2004**, *31*, 143–150. [CrossRef]
- 132. Ngwabebhoh, F.A.; Patwa, R.; Zandraa, O.; Saha, N.; Saha, P. Preparation and characterization of injectable self-antibacterial gelatin/carrageenan/bacterial cellulose hydrogel scaffolds for wound healing application. *J. Drug Deliv. Sci. Technol.* **2021**, 63, 102415. [CrossRef]
- 133. Popa, E.G.; Rodrigues, M.T.; Coutinho, D.F.; Oliveira, M.B.; Mano, J.F.; Reisab, R.L.; Gomes, M.E. Cryopreservation of cell laden natural origin hydrogels for cartilage regeneration strategies. *Soft Matter* **2013**, *9*, 875–885. [CrossRef]
- 134. Popa, E.G.; Carvalho, P.P.; Dias, A.F.; Santos, T.C.; Santo, V.E.; Marques, A.P.; Viegas, C.A.; Dias, I.R.; Gomes, M.E.; Reis, R.L. Evaluation of the in vitro and in vivo biocompatibility of carrageenan-based hydrogels. *J. Biomed. Mater. Res.* **2014**, 102, 4087–4097. [CrossRef]
- 135. Popa, E.; Reis, R.; Gomes, M. Chondrogenic phenotype of different cells encapsulated in κ-carrageenan hydrogels for cartilage regeneration strategies. *Biotechnol. Appl. Biochem.* **2012**, *59*, 132–141. [CrossRef]
- 136. Bornhöft, M.; Thommes, M.; Kleinebudde, P. Preliminary assessment of carrageenan as excipient for extrusion/spheronisation. *Eur. J. Pharm. Biopharm.* **2005**, *59*, 127–131. [CrossRef]
- 137. Ganta, S.; Devalapally, H.; Shahiwala, A.; Amiji, M. A review of stimuli-responsive nanocarriers for drug and gene delivery. *J. Control. Release* **2008**, 126, 187–204. [CrossRef]
- 138. Meyer, D.E.; Shin, B.C.; Kong, G.A.; Dewhirst, M.W.; Chilkoti, A. Drug targeting using thermally responsive polymers and local hyperthermia. *J. Control. Release* **2001**, 74, 213–224. [CrossRef]
- 139. Maciel, D.J.; de Mello Ferreira, I.L.; da Costa, G.M.; da Silva, M.R. Nanocomposite hydrogels based on iota-carrageenan and maghemite: Morphological, thermal and magnetic properties. *Eur. Polym. J.* **2016**, *76*, 147–155. [CrossRef]
- 140. Sahiner, N.; Sagbas, S.; Yılmaz, S. Microgels Derived from Different Forms of Carrageenans, Kappa, Iota, and Lambda for Biomedical Applications. *MRS Adv.* **2017**, *2*, 2521–2527. [CrossRef]
- 141. Percival, E. The polysaccharides of green, red and brown seaweeds: Their basic structure, biosynthesis and function. *Br. Phycol. J.* **1979**, *14*, 103–117. [CrossRef]
- 142. Pomin, V.H.; Mourão, P.A.S. Structure, Biology, evolution, and medical importance of sulfated fucans and galactans. *Glycobiology* **2008**, *18*, 1016–1027. [CrossRef]
- 143. Stengel, D.B.; Connan, S.; Popper, Z.A. Algal chemodiversity and bioactivity: Sources of natural variability and implications for commercial application. *Biotechnol. Adv.* **2011**, *29*, 483–501. [CrossRef]
- 144. Lewis, L.A.; McCourt, R.M. Green algae and the origin of land plants. Am. J. Bot. 2004, 91, 1535–1556. [CrossRef]
- 145. Domozych, D.S.; Stewart, K.D.; Mattox, K.R. The comparative aspects of cell wall chemistry in the green algae (Chlorophyta). *J. Mol. Evol.* **1980**, *15*, 1–12. [CrossRef] [PubMed]

Molecules **2021**, 26, 7068 30 of 36

146. Popper, Z.A.; Tuohy, M.G. Beyond the Green: Understanding the Evolutionary Puzzle of Plant and Algal Cell Walls. *Plant. Physiol.* **2010**, *153*, 373–383. [CrossRef]

- 147. Toskas, G.; Hund, R.-D.; Laourine, E.; Cherif, C.; Smyrniotopoulos, V.; Roussis, V. Nanofibers based on polysaccharides from the green seaweed Ulva Rigida. *Carbohydr. Polym.* **2011**, *84*, 1093–1102. [CrossRef]
- 148. Alves, A.; Pinho, E.D.; Neves, N.M.; Sousa, R.A.; Reis, R.L. Processing ulvan into 2D structures: Cross-linked ulvan membranes as new biomaterials for drug delivery applications. *Int. J. Pharm.* **2012**, 426, 76–81. [CrossRef] [PubMed]
- 149. Alves, A.; Duarte, A.R.C.; Mano, J.F.; Sousa, R.A.; Reis, R.L. PDLLA enriched with ulvan particles as a novel 3D porous scaffold targeted for bone engineering. *J. Supercrit. Fluids* **2012**, *65*, 32–38. [CrossRef]
- 150. Morelli, A.; Chiellini, F. Ulvan as a New Type of Biomaterial from Renewable Resources: Functionalization and Hydrogel Preparation. *Macromol. Chem. Phys.* **2010**, 211, 821–832. [CrossRef]
- 151. Alves, A.; Sousa, R.A.; Reis, R.L. Processing of degradable ulvan 3D porous structures for biomedical applications. *J. Biomed. Mater. Res.* **2013**, *101*, 998–1006. [CrossRef] [PubMed]
- 152. Massironi, A.; Morelli, A.; Grassi, L.; Puppi, D.; Braccini, S.; Maisetta, G.; Esin, S.; Batoni, G.; Pina, C.D.; Chiellini, F. Ulvan as novel reducing and stabilizing agent from renewable algal biomass: Application to green synthesis of silver nanoparticles. *Carbohydr. Polym.* **2019**, 203, 310–321. [CrossRef] [PubMed]
- 153. Tziveleka, L.-A.; Pippa, N.; Georgantea, P.; Ioannou, E.; Demetzos, C.; Roussis, V. Marine sulfated polysaccharides as versatile polyelectrolytes for the development of drug delivery nanoplatforms: Complexation of ulvan with lysozyme. *Int. J. Biol. Macromol.* **2018**, *118*, 69–75. [CrossRef]
- 154. Gajaria, T.K.; Bhatt, H.; Khandelwal, A.; Vasu, V.T.; Reddy, C.R.K.; Lakshmi, D.S. A facile chemical cross-linking approach toward the fabrication of a sustainable porous ulvan scaffold. *J. Bioact. Compat. Polym.* **2020**, *35*, 301–313. [CrossRef]
- 155. Dash, M.; Samal, S.K.; Morelli, A.; Bartoli, C.; Declercq, H.A.; Douglas, T.E.L.; Dubruel, P.; Chiellini, F. Ulvan-chitosan polyelectrolyte complexes as matrices for enzyme induced biomimetic mineralization. *Carbohydr. Polym.* **2018**, *182*, 254–264. [CrossRef]
- 156. Dash, M.; Samal, S.K.; Bartoli, C.; Morelli, A.; Smet, P.F.; Dubruel, P.; Chiellini, F. Biofunctionalization of Ulvan Scaffolds for Bone Tissue Engineering. *ACS Appl. Mater. Interfaces* **2014**, *6*, 3211–3218. [CrossRef] [PubMed]
- 157. Bang, T.H.; Van, T.T.; Hung, L.X.; Ly, B.M.; Nhut, N.D.; Thuy, T.T. Nanogels of acetylated ulvan enhance the solubility of hydrophobic drug curcumin. *Bull. Mater. Sci.* 2019, 42, 1. [CrossRef]
- 158. Mariia, K.; Arif, M.; Shi, J.; Song, F.; Chi, Z.; Liu, C. Novel chitosan-ulvan hydrogel reinforcement by cellulose nanocrystals with epidermal growth factor for enhanced wound healing: In vitro and in vivo analysis. *Int. J. Biol. Macromol.* **2021**, *183*, 435–446. [CrossRef]
- 159. Kikionis, S.; Ioannou, E.; Aggelidou, E.; Tziveleka, L.-A.; Demiri, E.; Bakopoulou, A.; Zinelis, S.; Kritis, A.; Roussis, V. The Marine Polysaccharide Ulvan Confers Potent Osteoinductive Capacity to PCL-Based Scaffolds for Bone Tissue Engineering Applications. *Int. J. Mol. Sci.* **2021**, 22, 3086. [CrossRef]
- 160. Costa, L.S.; Fidelis, G.P.; Cordeiro, S.L.; Oliveira, R.M.; Sabry, D.A.; Câmara, R.B.G.; Nobre, L.T.D.B.; Costa, M.S.S.P.; Almeida-Lima, J.; Farias, E.H.C.; et al. Biol.ogical activities of sulfated polysaccharides from tropical seaweeds. *Biomed. Pharmacother.* **2010**, 64, 21–28. [CrossRef]
- 161. Kesavan, S.; Smeen, K.; Sharmili, S.A.; Govindarajan, M.; Alharbi, N.S.; Kadaikunnan, S.; Khaled, J.M.; Alobaidi, A.S.; Alanzi, K.F.; Vaseeharan, B. Ulvan loaded graphene oxide nanoparticle fabricated with chitosan and D-mannose for targeted anticancer drug delivery. *J. Drug Deliv. Sci. Technol.* 2021, 65, 102760. [CrossRef]
- 162. Kim, K.; Cho, M.L.; Karnjanapratum, S.; Shin, I.-S.; You, S.G. In vitro and in vivo immunomodulatory activity of sulfated polysaccharides from Enteromorpha prolifera. *Int. J. Biol. Macromol.* **2011**, *49*, 1051–1058. [CrossRef]
- 163. Madub, K.; Goonoo, N.; Gimié, F.; Arsa, I.A.; Schönherr, H.; Bhaw-Luximon, A. Green seaweeds ulvan-cellulose scaffolds enhance in vitro cell growth and in vivo angiogenesis for skin tissue engineering. *Carbohydr. Polym.* **2021**, 251, 117025. [CrossRef]
- 164. Molino, P.J.; Garcia, L.; Stewart, E.M.; Lamaze, M.; Zhang, B.; Harris, A.R.; Winbergde, P.; Wallace, G.G. PEDOT doped with algal, mammalian and synthetic dopants: Polymer properties, protein and cell interactions, and influence of electrical stimulation on neuronal cell differentiation. *Biomater. Sci.* **2018**, *6*, 1250–1261. [CrossRef]
- 165. Tziveleka, L.-A.; Sapalidis, A.; Kikionis, S.; Aggelidou, E.; Demiri, E.; Kritis, A.; Ioannou, E.; Roussis, V. Hybrid Sponge-Like Scaffolds Based on Ulvan and Gelatin: Design, Characterization and Evaluation of Their Potential Use in Bone Tissue Engineering. *Materials* 2020, *13*, 1763. [CrossRef] [PubMed]
- 166. Usov, A.I.; Zelinsky, N.D. Chemical structures of algal polysaccharides. In *Functional Ingredients from Algae for Foods and Nutraceuticals*; (Woodhead Publishing Series in Food Science, Technology and Nutrition), Domínguez, F.N., Eds.; Woodhead Publishing: Sawston, UK, 2013; pp. 23–86.
- 167. Jiao, G.; Yu, G.; Zhang, J.; Ewart, H.S. Chemical structures and bioactivities of sulfated polysaccharides from marine algae. *Mar. Drugs* **2011**, *9*, 196–223. [CrossRef] [PubMed]
- 168. Cunha, L.; Grenha, A. Sulfated Seaweed Polysaccharides as Multifunctional Materials in Drug Delivery Applications. *Mar. Drugs* **2016**, *14*, 42. [CrossRef]
- 169. Sezer, A.D.; Cevher, E.; Hatipoğlu, F.; Oğurtan, Z.; Baş, A.L.; Akbuğa, J. Preparation of fucoidan-chitosan hydrogel and its application as burn healing accelerator on rabbits. *Biol. Pharm. Bull.* **2008**, *31*, 2326–2333. [CrossRef]
- 170. Lee, H.M.; Kim, J.-K.; Cho, T.-S. Applications of ophthalmic biomaterials embedded with fucoidan. *J. Ind. Eng. Chem.* **2012**, *18*, 1197–1201. [CrossRef]

Molecules **2021**, 26, 7068 31 of 36

171. Cumashi, A.; Ushakova, N.A.; Preobrazhenskaya, M.E.; D'Incecco, A.; Piccoli, A.; Totani, L.; Tinari, N.; Morozevich, G.E.; Berman, A.E.; Bilan, M.I.; et al. A comparative study of the anti-inflammatory, anticoagulant, antiangiogenic, and antiadhesive activities of nine different fucoidans from brown seaweeds. *Glycobiology* **2007**, *17*, 541–552. [CrossRef] [PubMed]

- 172. Croci, D.O.; Cumashi, A.; Ushakova, N.A.; Preobrazhenskaya, M.E.; Piccoli, A.; Totani, L.; Ustyuzhanina, N.E.; Bilan, M.I.; Usov, A.I.; Grachev, A.A.; et al. Fucans, but not fucomannoglucuronans, determine the Biological activities of sulfated polysaccharides from Laminaria saccharina brown seaweed. *PLoS ONE* 2011, 6, e17283. [CrossRef] [PubMed]
- 173. Kuznetsova, T.A.; Besednova, N.N.; Mamaev, A.N.; Momot, A.P.; Shevchenko, N.M.; Zvyagintseva, T.N. Anticoagulant activity of fucoidan from brown algae *Fucus evanescens* of the Okhotsk Sea. *Bull. Exp. Biol. Med.* **2003**, *136*, 471–473. [CrossRef] [PubMed]
- 174. Lee, J.-B.; Hayashi, K.; Hashimoto, M.; Nakano, T.; Hayashi, T. Novel antiviral fucoidan from sporophyll of *Undaria pinnatifida* (Mekabu). *Chem. Pharm. Bull.* **2004**, 52, 1091–1094. [CrossRef] [PubMed]
- 175. Raghavendran, H.R.B.; Srinivasan, P.; Rekha, S. Immunomodulatory activity of fucoidan against aspirin-induced gastric mucosal damage in rats. *Int. Immunopharmacol.* **2011**, *11*, 157–163. [CrossRef] [PubMed]
- 176. Teng, H.; Yang, Y.; Wei, H.; Liu, Z.; Liu, Z.; Ma, Y.; Gao, Z.; Hou, L.; Zou, X. Fucoidan Suppresses Hypoxia-Induced Lymphangiogenesis and Lymphatic Metastasis in Mouse Hepatocarcinoma. *Mar. Drugs* **2015**, *13*, 3514–3530. [CrossRef] [PubMed]
- 177. Senthilkumar, K.; Manivasagan, P.; Venkatesan, J.; Kim, S.-K. Brown seaweed fucoidan: Biological activity and apoptosis, growth signaling mechanism in cancer. *Int. J. Biol. Macromol.* **2013**, *60*, 366–374. [CrossRef] [PubMed]
- 178. Kim, O.-K.; Lee, M.; Kwon, H.O.; Lee, D.; Park, J.; Kim, E.; You, Y.; Lim, Y.T.; Jun, W.; Lee, J. Costaria costata Extract Suppresses Development of Atopic Dermatitis in chloro-2,4-dinitrobenzene-treated NC/Nga Mice. Skin Pharmacol. Physiol. 2018, 31, 212–219. [CrossRef] [PubMed]
- 179. Sanjeewa, K.A.; Jayawardena, T.U.; Kim, S.Y.; Kim, H.S.; Ahn, G.; Kim, J.; Jeon, Y.J. Fucoidan isolated from invasive Sargassum horneri inhibit LPS-induced inflammation via blocking NF-κB and MAPK pathways. *Algal Res.* **2019**, *41*, 101561. [CrossRef]
- 180. Xu, Y.; Xu, J.; Ge, K.; Tian, Q.; Zhao, P.; Guo, Y. Anti-inflammatory effect of low molecular weight fucoidan from Saccharina japonica on atherosclerosis in apoE-knockout mice. *Int. J. Biol. Macromol.* **2018**, *118*, 365–374. [CrossRef]
- 181. Rui, X.; Pan, H.-F.; Shao, S.-L.; Xu, X.-M. Anti-tumor and anti-angiogenic effects of Fucoidan on prostate cancer: Possible JAK-STAT3 pathway. *BMC Complement. Altern. Med.* **2017**, 17, 378. [CrossRef] [PubMed]
- 182. Flórez-Fernández, N.; Torres, M.D.; González-Muñoz, M.J.; Domínguez, H. Potential of intensification techniques for the extraction and depolymerization of fucoidan. *Algal Res.* **2018**, *30*, 128–148. [CrossRef]
- 183. Kim, I.-H.; Kwon, M.-J.; Nam, T.-J. Differences in cell death and cell cycle following fucoidan treatment in high-density HT-29 colon cancer cells. *Mol. Med. Rep.* **2017**, *15*, 4116–4122. [CrossRef]
- 184. Park, H.Y.; Park, S.H.; Jeong, J.W.; Yoon, D.; Han, M.H.; Lee, D.S.; Choi, G.; Yim, M.J.; Lee, J.M.; Kim, D.H.; et al. Induction of p53-Independent Apoptosis and G1 Cell Cycle Arrest by Fucoidan in HCT116 Human Colorectal Carcinoma Cells. *Mar. Drugs* 2017, 15, 154. [CrossRef] [PubMed]
- 185. Zhao, D.; Xu, J.; Xu, X. Bioactivity of fucoidan extracted from *Laminaria japonica* using a novel procedure with high yield. *Food Chem.* **2018**, 245, 911–918. [CrossRef]
- 186. Yu, S.-H.; Wu, S.-J.; Wu, J.-Y.; Wen, D.-Y.; Mi, F.-L. Preparation of fucoidan-shelled and genipin-crosslinked chitosan beads for antibacterial application. *Carbohydr. Polym.* **2015**, *126*, 97–107. [CrossRef]
- 187. Wu, S.-J.; Don, T.-M.; Lin, C.-W.; Mi, F.-L. Delivery of berberine using chitosan/fucoidan-taurine conjugate nanoparticles for treatment of defective intestinal epithelial tight junction barrier. *Mar. Drugs* **2014**, 12, 5677–5697. [CrossRef] [PubMed]
- 188. Cavalcanti, I.D.; Ximenes, R.M.; Pessoa, O.D.; Magalhães, N.S.; de Britto Lira-Nogueira, M.C. Fucoidan-coated PIBCA nanoparticles containing oncocalyxone A: Activity against metastatic breast cancer cells. *J. Drug Deliv. Sci. Technol.* **2021**, *65*, 102698. [CrossRef]
- 189. Huang, Y.C.; Li, R.Y. Preparation and characterization of antioxidant nanoparticles composed of chitosan and fucoidan for antibiotics delivery. *Mar. Drugs.* **2014**, *12*, 4379–4398. [CrossRef] [PubMed]
- 190. Huang, Y.-C.; Yang, Y.-T. Effect of basic fibroblast growth factor released from chitosan-fucoidan nanoparticles on neurite extension. *J. Tissue Eng. Regen. Med.* **2016**, *10*, 418–427. [CrossRef] [PubMed]
- 191. Huang, Y.-C.; Liu, T.-J. Mobilization of mesenchymal stem cells by stromal cell-derived factor-1 released from chitosan/tripolyphosphate/fucoidan nanoparticles. *Acta Biomater.* **2012**, *8*, 1048–1056. [CrossRef]
- 192. Nakamura, S.; Nambu, M.; Ishizuka, T.; Hattori, H.; Kanatani, Y.; Takase, B.; Kishimoto, S.; Amano, Y.; Aoki, H.; Kiyosawa, T.; et al. Effect of controlled release of fibroblast growth factor-2 from chitosan/fucoidan micro complex-hydrogel on in vitro and in vivo vascularization. *J. Biomed. Mater. Res.* **2008**, *85*, 619–627. [CrossRef] [PubMed]
- 193. Oliveira, C.; Gonçalves, C.S.; Martins, E.P.; Neves, N.M.; Reis, R.L.; Costa, B.M.; Silva, T.H.; Martins, A. Fucoidan/chitosan nanoparticles functionalized with anti-ErbB-2 target breast cancer cells and impair tumor growth in vivo. *Int. J. Pharm.* **2021**, 600, 120548. [CrossRef]
- 194. Laurienzo, P. Marine polysaccharides in pharmaceutical applications: An overview. *Mar. Drugs* **2010**, *8*, 2435–2465. [CrossRef] [PubMed]
- 195. Nichols, C.A.M.; Guezennec, J.; Bowman, J.P. Bacterial exopolysaccharides from extreme marine environments with special consideration of the Southern Ocean, sea ice, and deep-sea hydrothermal vents: A review. *Mar. Biotechnol.* **2005**, *7*, 253–271. [CrossRef]

Molecules **2021**, 26, 7068 32 of 36

196. Wang, J.; Salem, D.R.; Sani, R.K. Two new exopolysaccharides from a thermophilic bacterium *Geobacillus* sp. WSUCF1: Characterization and bioactivities. *N. Biotechnol.* **2021**, *61*, 29–39. [CrossRef] [PubMed]

- 197. Charoenwongpaiboon, T.; Wangpaiboon, K.; Pichyangkura, R.; Nepogodiev, S.A.; Wonganan, P.; Mahalapbutr, P.; Field, R.A. Characterization of a nanoparticulate exopolysaccharide from *Leuconostoc holzapfelii* KM01 and its potential application in drug encapsulation. *Int. J. Biol. Macromol.* **2021**, *187*, 690–698. [CrossRef] [PubMed]
- 198. Komorowski, P.; Kołodziejczyk, A.; Makowski, K.; Kotarba, S.; Walkowiak, B. In situ-formed bacterial exopolysaccharide (EPS) as a potential carrier for anchorage-dependent cell cultures. *Eng. Biomater.* **2021**, *159*, 18–23.
- 199. Sun, C.; Wang, J.-W.; Fang, L.; Gao, X.-D.; Tan, R.-X. Free radical scavenging and antioxidant activities of EPS2, an exopolysaccharide produced by a marine filamentous fungus *Keissleriella* sp. YS 4108. *Life Sci.* **2004**, 75, 1063–1073. [CrossRef]
- 200. Sun, C.; Shan, C.Y.; Gao, X.D.; Tan, R.X. Protection of PC12 cells from hydrogen peroxide-induced injury by EPS2, an exopolysac-charide from a marine filamentous fungus *Keissleriella* sp. YS4108. *J. Biotechnol.* **2005**, *115*, 137–144. [CrossRef]
- 201. Gargouch, N.; Elleuch, F.; Karkouch, I.; Tabbene, O.; Pichon, C.; Gardarin, C.; Rihouey, C.; Picton, L.; Abdelkafi, S.; Fendri, I.; et al. Potential of Exopolysaccharide from *Porphyridium marinum* to Contend with Bacterial Proliferation, Biofilm Formation, and Breast Cancer. *Mar. Drugs* 2021, 19, 66. [CrossRef] [PubMed]
- Guzman-Murillo, M.A.; Ascencio, F. Anti-adhesive activity of sulphated exopolysaccharides of microalgae on attachment of red sore disease-associated bacteria and helicobacter pylori to tissue culture cells. Lett. Appl. Microbiol. 2000, 30, 473

  –478. [CrossRef]
- 203. Yang, Q.; Ge, Y.; Iqbal, N.M.; Yang, X.; Zhang, X. *Sulfitobacter alexandrii* sp. nov., a new microalgae growth-promoting bacterium with exopolysaccharides bioflocculanting potential isolated from marine phycosphere. *Antonie Van Leeuwenhoek* **2021**, 114, 1091–1106. [CrossRef]
- 204. Liu, F.; Ng, T.B. Antioxidative and free radical scavenging activities of selected medicinal herbs. *Life Sci.* **2000**, *66*, 725–735. [CrossRef]
- 205. Schinella, G.R.; Tournier, H.A.; Prieto, J.M.; Mordujovich de Buschiazzo, P.; Ríos, J.L. Antioxidant activity of anti-inflammatory plant extracts. *Life Sci.* **2002**, *70*, 1023–1033. [CrossRef]
- 206. Sun, H.-H.; Mao, W.-J.; Chen, Y.; Guo, S.-D.; Li, H.-Y.; Qi, X.-H.; Chen, Y.-L.; Xu, J. Isolation, chemical characteristics and antioxidant properties of the polysaccharides from marine fungus *Penicillium* sp. F23-2. *Carbohydr. Polym.* **2009**, 78, 117–124. [CrossRef]
- 207. Kim, S.-J.; Chung, B.H. Antioxidant activity of levan coated cerium oxide nanoparticles. *Carbohydr. Polym.* **2016**, *150*, 400–407. [CrossRef] [PubMed]
- 208. Yan, J.-K.; Qiu, W.-Y.; Wang, Y.-Y.; Wang, W.-H.; Yang, Y.; Zhang, H.-N. Fabrication and stabilization of biocompatible selenium nanoparticles by carboxylic curdlans with various molecular properties. *Carbohydr. Polym.* **2018**, *179*, 19–27. [CrossRef] [PubMed]
- 209. Barcelos, M.C.S.; Vespermann, K.A.C.; Pelissari, F.M.; Molina, G. Current status of Biotechnological production and applications of microbial exopolysaccharides. *Crit. Rev. Food Sci. Nutr.* **2020**, *60*, 1475–1495. [CrossRef] [PubMed]
- 210. Adebayo-Tayo, B.C.; Popoola, A.O. Biogenic synthesis and antimicrobial activity of silver nanoparticle using exopolysaccharides from Lactic Acid bacteria. *Int. J. Nano Dimens.* **2017**, *8*, 61–69.
- 211. Sun, M.L.; Zhao, F.; Chen, X.L.; Zhang, X.Y.; Zhang, Y.Z.; Song, X.Y.; Sun, C.Y.; Yang, J. Promotion of Wound Healing and Prevention of Frostbite Injury in Rat Skin by Exopolysaccharide from the Arctic Marine Bacterium *Polaribacter* sp. SM1127. *Mar. Drugs* 2020, *18*, 48. [CrossRef] [PubMed]
- 212. Tiwari, O.N.; Mondal, A.; Bhunia, B.; Kanti Bandyopadhyay, T.; Jaladi, P.; Oinam, G.; Indrama, T. Purification, characterization and Biotechnological potential of new exopolysaccharide polymers produced by cyanobacterium *Anabaena* sp. CCC 745. *Polymer* 2019, 178, 121695. [CrossRef]
- 213. Abinaya, M.; Vaseeharan, B.; Divya, M.; Sharmili, A.; Govindarajan, M.; Alharbi, N.S.; Kadaikunnan, S.; Khaled, J.M.; Benelli, G. Bacterial exopolysaccharide (EPS)-coated ZnO nanoparticles showed high antibiofilm activity and larvicidal toxicity against malaria and Zika virus vectors. *J. Trace Elem. Med. Biol.* **2018**, *45*, 93–103. [CrossRef] [PubMed]
- 214. Chihara, G.; Hamuro, J.; Maeda, Y.Y.; Arai, Y.; Fukuoka, F. Fractionation and Purification of the Polysaccharides with Marked Antitumor Activity, Especially Lentinan, from "Lentinus edodes" (Berk.) Sing. (an Edible Mushroom). Cancer Res. 1970, 30, 2776–2781.
- 215. Matsuda, M.; Yamori, T.; Naitoh, M.; Okutani, K. Structural Revision of Sulfated Polysaccharide B-1 Isolated from a Marine Pseudomonas Species and Its Cytotoxic Activity Against Human Cancer Cell Lines. *Mar. Biotechnol.* **2003**, *5*, 13–19. [CrossRef]
- 216. Colliec-Jouault, S.; Zanchetta, P.; Helley, D.; Ratiskol, J.; Sinquin, C.; Fischer, A.M.; Guezennec, J. Les polysaccharides microbiens d'origine marine et leur potentiel en thérapeutique humaine. *Pathol. Biol.* **2004**, *52*, 127–130. [CrossRef] [PubMed]
- 217. Sirin, S.; Aslim, B. Characterization of lactic acid bacteria derived exopolysaccharides for use as a defined neuroprotective agent against amyloid beta1–42-induced apoptosis in SH-SY5Y cells. *Sci. Rep.* **2020**, *10*, 8124. [CrossRef] [PubMed]
- 218. Guezennec, J.; Pignet, P.; Lijour, Y.; Gentric, E.; Ratiskol, J.; Colliec-Jouault, S. Sulfation and depolymerization of a bacterial exopolysaccharide of hydrothermal origin. *Carbohydr. Polym* **1998**, 37, 19–24. [CrossRef]
- 219. Hayashi, K.; Hayashi, T.; Kojima, I. A natural sulfated polysaccharide, calcium spirulan, isolated from Spirulina platensis: In vitro and ex vivo evaluation of anti-herpes simplex virus and anti-human immunodeficiency virus activities. *AIDS Res. Hum. Retrovir.* **1996**, 12, 1463–1471. [CrossRef]
- 220. Hayashi, T.; Hayashi, K.; Maeda, M.; Kojima, I. Calcium Spirulan, an Inhibitor of Enveloped Virus Replication, from a Blue-Green Alga Spirulina platensis. *J. Nat. Prod.* **1996**, *59*, 83–87. [CrossRef]

Molecules **2021**, 26, 7068 33 of 36

221. De Morais, M.G.; Stillings, C.; Dersch, R.; Rudisile, M.; Pranke, P.; Costa, J.A.V.; Wendorff, J. Preparation of nanofibers containing the microalga *Spirulina* (Arthrospira). *Bioresour. Technol.* **2010**, *101*, 2872–2876. [CrossRef]

- 222. Senni, K.; Pereira, J.; Gueniche, F.; Delbarre-Ladrat, C.; Sinquin, C.; Ratiskol, J.; Godeau, G.; Fischer, A.-M.; Helley, D.; Colliec-Jouault, S. Marine polysaccharides: A source of bioactive molecules for cell therapy and tissue engineering. *Mar. Drugs* **2011**, *9*, 1664–1681. [CrossRef]
- 223. Moussa, T.A.A.; Al-Qaysi, S.A.S.; Thabit, Z.A.; Kadhem, S.B. Microbial levan from *Brachybacterium phenoliresistens*: Characterization and enhancement of production. *Process. Biochem.* **2017**, *57*, 9–15. [CrossRef]
- 224. Matsui, M.S.; Muizzuddin, N.; Arad, S.; Marenus, K. Sulfated polysaccharides from red microalgae have antiinflammatory properties in vitro and in vivo. *Appl. Biochem. Biotechnol.* **2003**, *104*, 13–22. [CrossRef]
- 225. Rederstorff, E.; Weiss, P.; Sourice, S.; Pilet, P.; Xie, F.; Sinquin, C.; Colliec-Jouault, S.; Guicheux, J.; Laïb, S. An in vitro study of two GAG-like marine polysaccharides incorporated into injectable hydrogels for bone and cartilage tissue engineering. *Acta Biomater.* **2011**, *7*, 2119–2130. [CrossRef] [PubMed]
- 226. Velasco, C.R.; Baud'Huin, M.; Sinquin, C.; Maillasson, M.; Heymann, D.; Colliec-Jouault, S.; Padrines, M. Effects of a sulfated exopolysaccharide produced by *Altermonas infernus* on bone Biology. *Glycobiology* **2011**, 21, 781–795. [CrossRef]
- 227. Sun, M.-L.; Zhao, F.; Zhang, X.-K.; Zhang, X.-Y.; Zhang, Y.-Z.; Song, X.-Y.; Chen, X.-L. Improvement of the production of an Arctic bacterial exopolysaccharide with protective effect on human skin cells against UV-induced oxidative stress. *Appl. Microbiol. Biotechnol.* **2020**, *104*, 4863–4875. [CrossRef] [PubMed]
- 228. Chapman, V. Seaweeds and Their Uses; Springer Science & Business Media: New York, NY, USA, 2012.
- 229. Evans, F.D.; Critchley, A.T. Seaweeds for animal production use. J. Appl. Physiol 2014, 26, 891–899. [CrossRef]
- 230. Stevenson, R.J.; Mahmut, M.K. Experience dependent changes in odour–viscosity perception. *Acta Psychol.* **2011**, *136*, 60–66. [CrossRef]
- 231. Kuo, W.Y.; Lee, Y. Effect of food matrix on saltiness perception-implications for sodium reduction. *Compr. Rev. Food Sci. Food Saf.* **2014**, *13*, 906–923. [CrossRef]
- 232. Roohinejad, S.; Koubaa, M.; Barba, F.J.; Saljoughian, S.; Amid, M.; Greiner, R. Application of seaweeds to develop new food products with enhanced shelf-life, quality and health-related beneficial properties. *Food Res. Int.* **2017**, *99*, 1066–1083. [CrossRef] [PubMed]
- 233. Moroney, N.C.; O'Grady, M.N.; Lordan, S.; Stanton, C.; Kerry, J.P. Seaweed polysaccharides (laminarin and fucoidan) as functional ingredients in pork meat: An evaluation of anti-oxidative potential, thermal stability and bioaccessibility. *Mar. Drugs* **2015**, *13*, 2447–2464. [CrossRef] [PubMed]
- 234. Jensen, M.G.; Knudsen, J.C.; Viereck, N.; Kristensen, M.; Astrup, A. Functionality of alginate-based supplements for application in human appetite regulation. *Food Chem.* **2012**, *132*, 823–829. [CrossRef]
- 235. Mattes, R.D. Effects of a combination fiber system on appetite and energy intake in overweight humans. *Physiol. Behav.* **2007**, 90, 705–711. [CrossRef]
- 236. Albert, A.; Salvador, A.; Fiszman, S.M. A film of alginate plus salt as an edible susceptor in microwaveable food. *Food Hydrocoll.* **2012**, 27, 421–426. [CrossRef]
- 237. Song, Y.; Liu, L.; Shen, H.; You, J.; Luo, Y. Effect of sodium alginate-based edible coating containing different anti-oxidants on quality and shelf life of refrigerated bream (*Megalobrama amblycephala*). Food Control **2011**, 22, 608–615. [CrossRef]
- 238. Raybaudi-Massilia, R.M.; Mosqueda-Melgar, J.; Martín-Belloso, O. Edible alginate-based coating as carrier of antimicrobials to improve shelf-life and safety of fresh-cut melon. *Int. J. Food Microbiol.* **2008**, *121*, 313–327. [CrossRef] [PubMed]
- 239. Kohajdová, Z.; Karovičová, J. Application of hydrocolloids as baking improvers. Chem. Pap. 2009, 63, 26–38. [CrossRef]
- 240. Kowalski, S.; Lukasiewicz, M.; Juszczak, L.; Sikora, M. Sensory and textural profile of confectionery masses produced using natural honey and selected polysaccharide hydrocolloids as the basis. *Zywnosc Nauka Technologia Jakosc* **2011**, *3*, 40–52. [CrossRef]
- 241. Piculell, L. Gelling carrageenans. In *Food Polysaccharides and Their Applications*; Stephen, A.M., Phillips, G.O., Williams, P.A., Eds.; Taylor and Francis: London, UK, 2006; p. 239. Available online: <a href="http://www.taylorandfrancis.com">http://www.taylorandfrancis.com</a> (accessed on 4 September 2021).
- 242. Haghighimanesh, S.; Farahnaky, A. Ice cream powder production and investigation of its rheological and organoleptic properties. *Int. J. Food Eng.* **2011**, *7*. [CrossRef]
- 243. Černíková, M.; Buňka, F.; Pospiech, M.; Tremlová, B.; Hladká, K.; Pavlínek, V.; Březina, P. Replacement of traditional emulsifying salts by selected hydrocolloids in processed cheese production. *Int. Dairy J.* **2010**, *20*, 336–343. [CrossRef]
- 244. Ayub, M. Influence of different types of milk and stabilizers on sensory evaluation and whey separation of yoghurt. *Biol. Sci. PJSIR* **2004**, *47*, 398–402.
- 245. Bixler, H.J.; Johndro, K.; Falshaw, R. Kappa-2 carrageenan: Structure and performance of commercial extracts: II. Performance in two simulated dairy applications. *Food Hydrocoll.* **2001**, *15*, 619–630. [CrossRef]
- 246. Shon, J.; Yun, Y.; Shin, M.; Chin, K.B.; Eun, J.B. Effects of milk proteins and gums on quality of bread made from frozen dough. *J. Sci. Food Agric.* **2009**, *89*, 1407–1415. [CrossRef]
- 247. Seol, K.H.; Lim, D.G.; Jang, A.; Jo, C.; Lee, M. Antimicrobial effect of κ-carrageenan-based edible film containing ovotransferrin in fresh chicken breast stored at 5 C. *Meat Sci.* **2009**, *83*, 479–483. [CrossRef]
- 248. McHugh, D.J. *A Guide to the Seaweed Industry FAO Fisheries*; Technical Paper 441; Food and Agriculture Organization of the United Nations (FAO): Rome, Italy, 2003.

Molecules **2021**, 26, 7068 34 of 36

249. Kumar, M.; Sharma, B.D. The storage stability and textural, physico-chemical and sensory quality of low-fat ground pork patties with Carrageenan as fat replacer. *Int. J. Food Sci. Tech.* **2004**, *39*, 31–42. [CrossRef]

- 250. Plotto, A.; Narciso, J.A.; Rattanapanone, N.; Baldwin, E.A. Surface treatments and coatings to maintain fresh-cut mango quality in storage. *J. Sci. Food Agric.* **2010**, *90*, 2333–2341. [CrossRef] [PubMed]
- 251. Liu, J.; Kandasamy, S.; Zhang, J.; Kirby, C.W.; Karakach, T.; Hafting, J.; Prithiviraj, B. Prebiotic effects of diet supplemented with the cultivated red seaweed *Chondrus crispus* or with fructo-oligo-saccharide on host immunity, colonic microbiota and gut microbial metabolites. *BMC Complement*. *Altern*. *Med*. **2015**, *15*, 279. [CrossRef] [PubMed]
- 252. Kim, J.Y.; Yu, D.Y.; Kim, J.A.; Choi, E.Y.; Lee, C.Y.; Hong, Y.H.; Cho, K. Effects of *Undaria pinnatifida* and *Laminaria japonica* on rat's intestinal microbiota and metabolite. *Int. J. Nutr. Food Sci.* **2016**, *6*, 502.
- 253. Lean, Q.Y.; Eri, R.D.; Fitton, J.H.; Patel, R.P.; Gueven, N. Fucoidan extracts ameliorate acute colitis. *PLoS ONE* **2015**, *10*, e0128453. [CrossRef]
- 254. Kuda, T.; Kosaka, M.; Hirano, S.; Kawahara, M.; Sato, M.; Kaneshima, T.; Kimura, B. Effect of sodium-alginate and laminaran on *Salmonella Typhimurium* infection in human enterocyte-like HT-29-Luc cells and BALB/c mice. *Carbohydr. Polym* 2015, 125, 113–119. [CrossRef]
- 255. Hu, B.; Gong, Q.; Wang, Y.; Ma, Y.; Li, J.; Yu, W. Prebiotic effects of neoagaro-oligosaccharides prepared by enzymatic hydrolysis of agarose. *Anaerobe* **2006**, 12, 260–266. [CrossRef]
- 256. Ullah, S.; Khalil, A.A.; Shaukat, F.; Song, Y. Sources, Extraction and Biomedical Properties of Polysaccharides. *Foods* **2019**, *8*, 304. [CrossRef]
- 257. Ahmadi, A.; Zorofchian Moghadamtousi, S.; Abubakar, S.; Zandi, K. Antiviral potential of algae polysaccharides isolated from marine sources: A review. *BioMed Res. Int.* **2015**, *2015*, 825203. [CrossRef]
- 258. Carse, S.; Bergant, M.; Schäfer, G. Advances in targeting hpv infection as potential alternative prophylactic means. *Int. J. Mol. Sci.* **2021**, 22, 2201. [CrossRef]
- 259. Kini, S.; Divyashree, M.; Mani, M.K.; Mamatha, B.S. Algae and cyanobacteria as a source of novel bioactive compounds for biomedical applications. In *Advances in Cyanobacterial Biology*; Academic Press: London, UK, 2020; pp. 173–194.
- 260. Bouaziz, F.; Koubaa, M.; Ellouz Ghorbel, R.; Ellouz Chaabouni, S. Biological properties of water-soluble polysaccharides and hemicelluloses from almond gum. *Int. J. Biol. Macromol.* **2017**, *95*, 667–674. [CrossRef] [PubMed]
- 261. Frediansyah, A. The antiviral activity of iota-, kappa-, and lambda-carrageenan against COVID-19: A critical review. *Clin. Epidemiol. Glob. Health* **2021**, *12*, 100826. [CrossRef]
- 262. Hans, N.; Malik, A.; Naik, S. Antiviral activity of sulfated polysaccharides from marine algae and its application in combating COVID-19: Mini review. *Bioresour. Technol. Rep.* **2021**, *13*, 100623. [CrossRef]
- 263. Lee, C. Carrageenans as Broad-Spectrum Microbicides: Current Status and Challenges. Mar. Drugs 2020, 18, 435. [CrossRef]
- 264. Moga, M.A.; Dima, L.; Balan, A.; Blidaru, A.; Dimienescu, O.G.; Podasca, C.; Toma, S. Are bioactive molecules from seaweeds a novel and challenging option for the prevention of HPV infection and cervical cancer therapy? A review. *Int. J. Mol. Sci.* **2021**, 22, 629. [CrossRef] [PubMed]
- 265. Pacheco-Quito, E.M.; Ruiz-Caro, R.; Veiga, M.D. Carrageenan: Drug Delivery Systems and Other Biomedical Applications. *Mar. Drugs* **2020**, *18*, 583. [CrossRef]
- 266. Tran, T.T.V.; Truong, H.B.; Tran, N.H.V.; Quach, T.M.T.; Nguyen, T.N.; Bui, M.L.; Thanh, T.T.T. Structure, conformation in aqueous solution and antimicrobial activity of ulvan extracted from green seaweed *Ulva reticulata*. *Nat. Prod. Res.* **2018**, 32, 2291–2296. [CrossRef]
- 267. Sen, I.K.; Chakraborty, I.; Mandal, A.K.; Bhanja, S.K.; Patra, S.; Maity, P. A review on antiviral and immunomodulatory polysaccharides from Indian medicinal plants, which may be beneficial to COVID-19 infected patients. *Int. J. Biol. Macromol.* **2021**, 181, 462–470. [CrossRef]
- 268. Hetta, M.; Mahmoud, R.; El-Senousy, W.; Ibrahim, M.; El-Taweel, G.; Ali, G. Antiviral and antimicrobial activities of *Spirulina platensis*. World J. Pharm. Sci. 2014, 3, 31–39.
- 269. Kanekiyo, K.; Hayashi, K.; Takenaka, H.; Lee, J.B.; Hayashi, T. Anti-herpes simplex virus target of an acidic polysaccharide, nostoflan, from the edible blue-green alga *Nostoc flagelliforme*. *Biol. Pharm. Bull.* **2007**, 30, 1573–1575. [CrossRef]
- 270. Liu, J.; Obaidi, I.; Nagar, S.; Scalabrino, G.; Sheridan, H. The antiviral potential of algal-derived Macromolecules. *Curr. Res. Biotechnol.* **2021**, *3*, 120–134. [CrossRef]
- 271. Satpati, G.G. Algal Sulfated Polysaccharides: Potent Immunomodulators against COVID-19 in Pandemic 2020. *Biosci. Biotechnol. Res. Asia.* **2020**, *17*, 601–605. [CrossRef]
- 272. Benhabiles, M.S.; Salah, R.; Lounici, H.; Drouiche, N.; Goosen, M.F.A.; Mameri, N. Antibacterial activity of chitin, chitosan and its oligomers prepared from shrimp shell waste. *Food Hydrocoll.* **2012**, *29*, 48–56. [CrossRef]
- 273. Khattak, S.; Wahid, F.; Liu, L.P.; Jia, S.R.; Chu, L.Q.; Xie, Y.Y.; Zhong, C. Applications of cellulose and chitin/chitosan derivatives and composites as antibacterial materials: Current state and perspectives. *Appl. Microbiol. Biotechnol.* **2019**, 103, 1989–2006. [CrossRef]
- 274. Rosca, I.; Petrovici, A.R.; Peptanariu, D.; Nicolescu, A.; Dodi, G.; Avadanei, M.; Ciolacu, D. Biosynthesis of dextran by *Weissella confusa* and its In vitro functional characteristics. *Int. J. Biol. Macromol.* **2018**, 107, 1765–1772. [CrossRef] [PubMed]
- 275. Lee, K.Y.; Jeong, M.R.; Choi, S.M.; Na, S.S.; Cha, J.D. Synergistic effect of fucoidan with antibiotics against oral pathogenic bacteria. *Arch. Oral Biol.* **2013**, *58*, 482–492. [CrossRef]

Molecules **2021**, 26, 7068 35 of 36

276. Phull, A.; Ali, A.; Ahmed, M.; Zia, M.; Haq, I.; Kim, S.J. In vitro antileishmanial, antibacterial, antifungal and anticancer activity of fucoidan from *Undaria pinnatifida*. *Int. J. Biosci.* **2017**, *11*, 219–227.

- 277. Ratan, Z.A.; Youn, S.H.; Kwak, Y.S.; Han, C.K.; Haidere, M.F.; Kim, J.K.; Cho, J. Y Adaptogenic effects of Panax ginseng on modulation of immune functions. *J. Ginseng Res.* **2021**, *45*, 32–40. [CrossRef] [PubMed]
- 278. Wang, L.; Huang, Y.; Yin, G.; Wang, J.; Wang, P.; Chen, Z.Y.; Ren, G. Antimicrobial activities of Asian ginseng, American ginseng, and notoginseng. *Phytother. Res.* 2020, *34*, 1226–1236. [CrossRef]
- 279. Akshay Kumar, K.P.; Zare, E.N.; Torres-Mendieta, R.; Wacławek, S.; Makvandi, P.; Černík, M.; Varma, R.S. Electrospun fibers based on botanical, seaweed, microbial, and animal sourced biomacromolecules and their multidimensional applications. *Int. J. Biol. Macromol.* 2021, 171, 130–149. [CrossRef] [PubMed]
- 280. Hippensteel, J.A.; LaRiviere, W.B.; Colbert, J.F.; Langout-Astri, C.J.; Schmidt, E.P. Heparin as a therapy for COVID-19: Current evidence and future possibilities. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2020**, *319*, L211–L217. [CrossRef] [PubMed]
- 281. Shi, C.; Tingting, W.; Li, J.P.; Sullivan, M.A.; Wang, C.; Wang, H.; Zhang, Y. Comprehensive landscape of heparin therapy for COVID-19. *Carbohydr. Polym.* 2021, 254, 117232. [CrossRef] [PubMed]
- 282. Shahzad, F.; Anderson, D.; Najafzadeh, M. The antiviral, anti-inflammatory effects of natural medicinal herbs and mushrooms and SARS-CoV-2 infection. *Nutrients* **2020**, *12*, 2573. [CrossRef]
- 283. Al-Hatamleh, M.A.; Hatmal, M.M.M.; Sattar, K.; Ahmad, S.; Mustafa, M.Z.; Bittencourt, M.D.C.; Mohamud, R. Antiviral and Immunomodulatory Effects of Phytochemicals from Honey against COVID-19: Potential Mechanisms of Action and Future Directions. *Molecules* 2020, 2, 5017. [CrossRef]
- 284. Adhikari, B.; Marasini, B.P.; Rayamajhee, B.; Bhattarai, B.R.; Lamichhane, G.; Khadayat, K.; Parajuli, N. Potential roles of medicinal plants for the treatment of viral diseases focusing on COVID-19: A review. *Phytother. Res.* **2021**, *35*, 1298–1312. [CrossRef] [PubMed]
- 285. Ferreira, I.C.F.R.; Heleno, S.A.; Reis, F.S.; Stojkovic, D.; Queiroz, M.J.R.P.; Vasconcelos, M.H.; Sokovic, M. Chemical features of Ganoderma polysaccharides with antioxidant, antitumor and antimicrobial activities. *Phytochemistry* **2015**, *114*, 38–55. [CrossRef] [PubMed]
- 286. Melo-Silveira, R.F.; Fidelis, G.P.; Pereira Costa, M.S.S.; Telles, C.B.S.; Dantas-Santos, N.; de Oliveira Elias, S.; Rocha, H.A.O. In vitro antioxidant, anticoagulant and antimicrobial activity and in inhibition of cancer cell proliferation by xylan extracted from corn cobs. *Int. J. Biol. Macromol.* **2012**, *13*, 409–426. [CrossRef] [PubMed]
- 287. Poveda-Castillo, G.D.C.; Rodrigo, D.; Martínez, A.; Pina-Pérez, M.C. Bioactivity of Fucoidan as an antimicrobial agent in a new functional beverage. *Beverages* **2018**, *4*, 64. [CrossRef]
- 288. Cardoso, I.; Cotas, J.; Rodrigues, A.; Ferreira, D.; Osório, N.; Pereira, L. Extraction and analysis of compounds with antibacterial potential from the red alga *Grateloupia turuturu*. *J. Mar. Sci. Eng.* **2019**, *7*, 220. [CrossRef]
- 289. Leandro, A.; Pereira, L.; Gonçalves, A.M.M. Diverse applications of marine macroalgae. Mar. Drugs 2020, 18, 17. [CrossRef]
- 290. Grigoras, A.G. Drug delivery systems using pullulan, a biocompatible polysaccharide produced by fungal fermentation of starch. *Environ. Chem. Lett.* **2019**, *17*, 1209–1223. [CrossRef]
- 291. Devaraj, R.D.; Reddy, C.K.; Xu, B. Health-promoting effects of konjac glucomannan and its practical applications: A critical review. *Int. J. Biol. Macromol.* **2019**, 126, 273–281. [CrossRef] [PubMed]
- 292. Du, B.; Meenu, M.; Liu, H.; Xu, B. A concise review on the molecular structure and function relationship of β-glucan. *Int. J. Mol. Sci.* **2019**, 20, 4032. [CrossRef] [PubMed]
- 293. Dobrange, E.; Peshev, D.; Loedolff, B.; Van Den Ende, W. Fructans as immunomodulatory and antiviral agents: The case of Echinacea. *Biomolecules* **2019**, *9*, 615. [CrossRef]
- 294. Ragab, T.I.M.; Shalaby, A.S.G.; Awdan, S.A.E.; El-Bassyouni, G.T.; Salama, B.M.; Helmy, W.A.; Esawy, M.A. Role of levan extracted from bacterial honey isolates in curing peptic ulcer: In vivo. *Int. J. Biol. Macromol.* **2020**, 142, 564–573. [CrossRef]
- 295. Chen, F.; Huang, G. Preparation and immunological activity of polysaccharides and their derivatives. *Int. J. Biol. Macromol.* **2018**, 112, 211–216. [CrossRef]
- 296. Garcia-Valdez, O.; Champagne, P.; Cunningham, M.F. Graft modification of natural polysaccharides via reversible deactivation radical polymerization. *Prog. Polym. Sci.* **2018**, *76*, 151–173. [CrossRef]
- 297. Huang, G.; Huang, H. The derivatization and antitumor mechanisms of polysaccharides. *Future Med. Chem.* **2017**, *9*, 1931–1938. [CrossRef] [PubMed]
- 298. Wang, Z.; Xie, J.; Shen, M.; Nie, S.; Xie, M. Sulfated modification of polysaccharides: Synthesis, characterization and bioactivities. *Trends Food Sci. Technol.* **2018**, *74*, 147–157. [CrossRef]
- 299. Li, S.; Xiong, Q.; Lai, X.; Li, X.; Wan, M.; Zhang, J.; Yan, Y.; Cao, M.; Lu, L.; Guan, J.; et al. Molecular Modification of Polysaccharides and Resulting Bioactivities. *Compr. Rev. Food Sci. Food Saf.* **2016**, *15*, 237–250. [CrossRef] [PubMed]
- 300. Chen, Y.; Zhang, H.; Wang, Y.; Nie, S.; Li, C.; Xie, M. Acetylation and carboxymethylation of the polysaccharide from *Ganoderma atrum* and their antioxidant and immunomodulating activities. *Food Chem.* **2014**, *156*, 279–288. [CrossRef] [PubMed]
- 301. Xu, P.; Li, J.; Hou, G.; Shi, F.; Ye, M. Cardioprotective effect of an exopolysaccharide from Lachnum YM130 and its derivatives on diabetic mice. *Proc. Biochem.* **2017**, *58*, 333–340. [CrossRef]
- 302. Li, J.; Chi, Z.; Yu, L.; Jiang, F.; Liu, C. Sulfated modification, characterization, and antioxidant and moisture absorption/retention activities of a soluble neutral polysaccharide from *Enteromorpha prolifera*. *Int. J. Biol. Macromol.* **2017**, *105*, 1544–1553. [CrossRef]

Molecules **2021**, 26, 7068 36 of 36

303. Yang, Y.; Chen, J.; Lei, L.; Li, F.; Tang, Y.; Yuan, Y.; Zhang, Y.; Wu, S.; Yin, R.; Ming, J. Acetylation of polysaccharide from *Morchella angusticeps* peck enhances its immune activation and anti-inflammatory activities in macrophage RAW264. 7 cells. *Food Chem. Toxicol.* **2019**, 125, 38–45. [CrossRef]

- 304. Liu, X.; Xie, J.; Jia, S.; Huang, L.; Wang, Z.; Li, C.; Xie, M. Immunomodulatory effects of an acetylated *Cyclocarya paliurus* polysaccharide on murine macrophages RAW264. 7. *Int. J. Biol. Macromol.* **2017**, *98*, 576–581. [CrossRef] [PubMed]
- 305. Liu, H.; Li, F.; Luo, P. Effect of Carboxymethylation and Phosphorylation on the Properties of Polysaccharides from Sepia esculenta Ink: Antioxidation and Anticoagulation In Vitro. *Mar. Drugs* **2019**, *17*, 626. [CrossRef]
- 306. Jiang, N.; Li, B.; Wang, X.; Xu, X.; Liu, X.; Li, W.; Chang, X.; Li, H.; Qi, H. The antioxidant and antihyperlipidemic activities of phosphorylated polysaccharide from *Ulva pertusa*. *Int. J. Biol. Macromol.* **2020**, *145*, 1059–1065. [CrossRef]
- 307. Huang, S.; Chen, F.; Cheng, H.; Huang, G. Modification and application of polysaccharide from traditional Chinese medicine such as Dendrobium officinale. *Int. J. Biol. Macromol.* **2020**, *157*, 385–393. [CrossRef]
- 308. Chakka, V.P.; Zhou, T. Carboxymethylation of polysaccharides: Synthesis and bioactivities. *Int. J. Biol. Macromol.* **2020**, 165, 2425–2431. [CrossRef]
- 309. Gabriel, L.; Tied, A.; Heinze, T. Carboxymethylation of polysaccharides—A comparative study. Cellul. Chem. Technol. 2020, 54, 82.
- 310. Suflet, D.M.; Chitanu, G.C.; Desbrières, J. Phosphorylated polysaccharides: 2: Synthesis and properties of phosphorylated dextran. *Carbohydr. Polym.* **2010**, *82*, 1271–1277. [CrossRef]
- 311. Silva, D.A.; de Paula, R.C.M.; Feitosa, J.; de Brito, A.C.F.; Maciel, J.S.; Paula, H.C.B. Carboxymethylation of cashew tree exudate polysaccharide. *Carbohydr. Polym.* **2004**, *58*, 163–171. [CrossRef]
- 312. Brumer, H. Enzymatic functionalization of cellulosic fibres for textile and other applications: Xyloglucan as a molecular anchor. In *Advances in Textile Biotechnology*; Woodhead Publishing: Cambridge, UK, 2010; pp. 266–287.
- 313. Liu, J.; Wang, X.; Yong, H.; Kan, J.; Jin, C. Recent advances in flavonoid-grafted polysaccharides: Synthesis, structural characterization, bioactivities and potential applications. *Int. J. Biol. Macromol.* **2018**, *116*, 1011–1025. [CrossRef]
- 314. Bozic, M.; Gorgieva, S.; Kokol, V. Laccase-mediated functionalization of chitosan by caffeic and gallic acids for modulating antioxidant and antimicrobial properties. *Carbohydr. Polym.* **2012**, *87*, 2388–2398. [CrossRef]
- 315. Tegl, G.; Stagl, V.; Mensah, A.; Huber, D.; Somitsch, W.; Grosse-Kracht, S.; Guebitz, G.M. The chemo enzymatic functionalization of chitosan zeolite particles provides antioxidant and antimicrobial properties. *Eng. Life Sci.* **2018**, *18*, 334–340. [CrossRef]
- 316. Vuillemin, M.E.; Muniglia, L.; Linder, M.; Bouguet-Bonnet, S.; Poinsignon, S.; dos Santos Morais, R.; Simard, B.; Paris, C.; Michaux, F.; Jasniewski, J. Polymer functionalization through an enzymatic process: Intermediate products characterization and their grafting onto gum Arabic. *Int. J. Biol. Macromol.* **2021**, *169*, 480–491. [CrossRef]
- 317. Yin, Y.; Lucia, L.A.; Pal, L.; Jiang, X.; Hubbe, M.A. Lipase-catalyzed laurate esterification of cellulose nanocrystals and their use as reinforcement in PLA composites. *Cellulose* **2020**, *27*, 6263–6273. [CrossRef]
- 318. Marjamaa, K.; Kruus, K. Enzyme Biotechnology in degradation and modification of plant cell wall polymers. *Physiol. Plant.* **2018**, 164, 106–118. [CrossRef]
- 319. Kaewprapan, K.; Wongkongkatep, J.; Panbangred, W.; Phinyocheep, P.; Marie, E.; Durand, A.; Inprakhon, P. Lipase-catalyzed synthesis of hydrophobically modified dextrans: Activity and regioselectivity of lipase from Candida rugosa. *J. Biosci. Bioeng.* **2011**, *112*, 124–129. [CrossRef] [PubMed]
- 320. Torres, E.; Ayala, M. Biocatalysis Based on Heme Peroxidases: Peroxidases as Potential Industrial Biocatalysts; Springer: Berlin/Heidelberg, Germany, 2010.
- 321. Li, X.; Li, S.; Liang, X.; McClements, D.J.; Liu, X.; Liu, F. Applications of oxidases in modification of food molecules and colloidal systems: Laccase, peroxidase and tyrosinase. *Trends Food Sci. Technol.* **2020**, *103*, 78–93. [CrossRef]
- 322. Zhang, L.; Shi, Z.; Shang, G.; Wen, Q.; Fang, Y.P.; Nishinari, K.; Phillips, G.O.; Jiang, F.T. Emulsification properties of sugar beet pectin after modification with horseradish peroxidase. *Food Hydrocoll.* **2015**, *43*, 107–113. [CrossRef]
- 323. Oosterveld, A.; Beldman, G.; Searle-Van Leeuwen, M.J.F.; Voragen, A.G.J. Effect of enzymatic deacetylation on gelation of sugar beet pectin in the presence of calcium. *Carbohydr. Polym.* **2000**, *43*, 249–256. [CrossRef]
- 324. Nachtigall, C.; Rohm, H.; Jaros, D. Degradation of Exopolysaccharides from Lactic Acid Bacteria by Thermal, Chemical, Enzymatic and Ultrasound Stresses. *Foods* **2021**, *10*, 396. [CrossRef] [PubMed]
- 325. Zhao, B.; Lv, C.; Lu, J. Natural occurring polysaccharides from Panax ginseng C. A. Meyer: A review of isolation, structures, and bioactivities. *Int. J. Biol. Macromol.* **2019**, 133, 324–336. [CrossRef] [PubMed]
- 326. Qin, W.; Dong, F.; Zhang, Z.; Hu, B.; Chen, S.; Zhu, Z.; Li, F.; Wang, X.; Zhang, Y.; Wang, Y.; et al. Low molecular weight heparin and 28-day mortality among patients with coronavirus disease 2019: A cohort study in the early epidemic era. *Thromb. Res.* **2021**, 198, 19–22. [CrossRef] [PubMed]