Check for updates

OPEN ACCESS

EDITED BY Chrissa Kioussi, Oregon State University, United States

REVIEWED BY Sabrina Batonnet-Pichon, Université de Paris, France Clarissa Henry, University of Maine, United States

*CORRESPONDENCE Zhengxing Lian, lianzhx@cau.edu.cn Kun Yu, yukun@cau.edu.cn

⁺These authors have contributed equally to this work

SPECIALTY SECTION

This article was submitted to Molecular and Cellular Pathology, a section of the journal Frontiers in Cell and Developmental Biology

RECEIVED 08 June 2022 ACCEPTED 10 August 2022 PUBLISHED 30 August 2022

CITATION

Chen M, Wang Y, Deng S, Lian Z and Yu K (2022), Skeletal muscle oxidative stress and inflammation in aging: Focus on antioxidant and antiinflammatory therapy. *Front. Cell Dev. Biol.* 10:964130. doi: 10.3389/fcell.2022.964130

COPYRIGHT

© 2022 Chen, Wang, Deng, Lian and Yu. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or

reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Skeletal muscle oxidative stress and inflammation in aging: Focus on antioxidant and anti-inflammatory therapy

Mingming Chen^{1†}, Yiyi Wang^{2†}, Shoulong Deng³, Zhengxing Lian^{1*} and Kun Yu^{1*}

¹College of Animal Science and Technology, China Agricultural University, Beijing, China, ²Zhejiang A&F University, Zhejiang Provincial Key Laboratory of Characteristic Traditional Chinese Medicine Resources Protection and Innovative Utilization, Lin'an, China, ³NHC Key Laboratory of Human Disease Comparative Medicine, Institute of Laboratory Animal Sciences, Chinese Academy of Medical Sciences and Comparative Medicine Center, Peking Union Medical College, Beijing, China

With aging, the progressive loss of skeletal muscle will have negative effect on multiple physiological parameters, such as exercise, respiration, thermoregulation, and metabolic homeostasis. Accumulating evidence reveals that oxidative stress and inflammation are the main pathological characteristics of skeletal muscle during aging. Here, we focus on agingrelated sarcopenia, summarize the relationship between aging and sarcopenia, and elaborate on aging-mediated oxidative stress and oxidative damage in skeletal muscle and its critical role in the occurrence and development of sarcopenia. In addition, we discuss the production of excessive reactive oxygen species in aging skeletal muscle, which reduces the ability of skeletal muscle satellite cells to participate in muscle regeneration, and analyze the potential molecular mechanism of ROS-mediated mitochondrial dysfunction in aging skeletal muscle. Furthermore, we have also paid extensive attention to the possibility and potential regulatory pathways of skeletal muscle aging and oxidative stress mediate inflammation. Finally, in response to the abnormal activity of oxidative stress and inflammation during aging, we summarize several potential antioxidant and anti-inflammatory strategies for the treatment of sarcopenia, which may provide beneficial help for improving sarcopenia during aging.

KEYWORDS

skeletal muscle, aging, oxidative stress, inflammation, treatment strategy

Introduction

Skeletal muscle, as a powerful mediator of health and longevity, is the most massive plastic organ in the body, and plays a vital role in respiration, movement, metabolism, daily physical activity, protection, and maintenance of posture and body balance (Frontera and Ochala, 2015). The progressive loss of skeletal muscle with aging will have negative effect on multiple physiological parameters, such as exercise, respiration,

10.3389/fcell.2022.964130

thermoregulation, and metabolic homeostasis. This age-related muscle wasting, known as sarcopenia, is a long-term process caused by a number of complex factors in which the average muscle capacity decreased during aging, resulting in increased opportunities of instability, falls, and fractures (Tieland et al., 2018). In aging skeletal muscle, reduced biosynthesis, slower metabolism, and smaller mitochondrial size cause rapid loss of skeletal muscle mass and efficiency parameters. It is typically characterized by weakened and reduced muscle mass and fiber cross-sectional area, the changes in myosin isotype or fiber type, and the net loss of cellular components such as organelles, cytoplasm, and total proteins (Chemello et al., 2011; Yin et al., 2021). It is manifested as decreased muscle strength, easy fatigue, and decreased exercise capacity.

Although it has a profound impact, the molecular mechanisms causing skeletal muscle changes with age are not completely clear. Current evidence suggests that sarcopenia is regulated by a complex network. Mechanistically, sarcopenia involves several pathophysiological processes, including oxidative stress, inflammation, mitochondrial dysfunction, and subsequent activation or inhibition of signal transduction, such as the mTOR signal pathway, autophagy lysosome system, and ubiquitin-proteasome system. Specifically, the abnormal production of reactive oxygen species (ROS), chronic and low-grade inflammation, mitochondrial dysfunction, modified protein synthesis, synthetic metabolic passivation, denervation and reduced muscle regeneration are some of the key processes associated with sarcopenia (Cuthbertson et al., 2005; Breen and Phillips, 2013; Vasilaki and Jackson, 2013; Almada and Wagers, 2016; Pollock et al., 2017; Sakellariou et al., 2017; Scalabrin et al., 2019). During these processes, the excessive production of ROS, which controls redox signal pathway in muscle fibers, can cause oxidative damage and mitochondrial dysfunction, attenuate ATP production, increase protein degradation and reduce protein synthesis, further resulting in muscle loss and strength decline (Powers, 2014). Furthermore, the elevated inflammation causes oxidative stress and anabolic resistance, leading to muscle loss. For example, the chronic and low-grade inflammation affects the metabolism of skeletal muscle cells through interactions among various cytokines. IL-6 secreted by skeletal muscle activates the STAT3 protein through inhibiting JAK/STAT3, PI3K/Akt, and ERK signaling pathways, and plays a crucial role in protein degradation of skeletal muscle (Silva et al., 2015). In addition, accumulation of fat and fibrosis can lead to poor muscle mass, and the reduced capacity for self-renewal and differentiation of skeletal muscle satellite cells cause impaired muscle regeneration.

In short, oxidative stress and inflammation are important factors that are closely associated with aging. Here, we detailed elaborate on the relationship between aging and sarcopenia, as well as aging and oxidative stress, and clarify that aging-mediated skeletal muscle oxidative stress and oxidative damage are direct factors in the development of sarcopenia. Specifically, ROSmediated skeletal muscle mitochondrial dysfunction is a potential gatekeeper for skeletal muscle aging. Excessive ROS production in aging skeletal muscle causes mitochondrial dysfunction and further aggravates inflammation. Finally, we summarize several potential antioxidant and anti-inflammatory strategies that are expected to be used to treat sarcopenia, although to date, there is no effective or approved drug therapy.

Aging-mediated oxidative stress and oxidative damage in skeletal muscle

Aging and sarcopenia

Aging is a natural process related to the physical function deterioration. The progressive loss of muscle mass, strength and function is one of the most prominent signs of aging. This skeletal muscle degeneration associated with aging can lead to adverse consequences and seriously damage the patient's quality of life. The geriatricians defined sarcopenia as "the loss of skeletal muscle function and mass associated with age", and launched the newest diagnostic standards for sarcopenia, which including low muscle strength, diminished muscle quantity, reduced muscle quality or weak physical performance (Cruz-Jentoft et al., 2019). Sarcopenia is currently recognized as a serious geriatric problem and an important condition for predicting frailty in the elderly (Cruz-Jentoft et al., 2019). Muscle mass gradually decline from 30 to 40 years old as muscle funcition weakens (Denison et al., 2015). This loss of muscle mass is more severe in people with a sedentary or inactive lifestyle. From 50 to 60 years of age, the loss of muscle mass may reache 1%-2%year and 3-5% year for the elderly (Denison et al., 2015). Of these individuals, 30%-50% of their muscle mass may be completely disappear from the ages of 40-80 (Denison et al., 2015). As a multi-faceted elderly disease characterized by both progressive and systemic loss of muscle mass and function (Cruz-Jentoft and Sayer, 2019; Borsch et al., 2021), and the development and progression of sarcopenia is consistently accompanied by a variety negative consequences, including falls, broken bones, loss of movement and even death (Morley et al., 2011; Bischoff-Ferrari et al., 2015; De Buyser et al., 2016). Similar to any other complex syndrome, the pathogenesis and progression of sarcopenia are diverse. Although the potential mechanism of sarcopenia remains elusive, a number of age-related factors can lead to the deterioration of the structure and function of skeletal muscle, thereby resulting in sarcopenia. For example, skeletal muscle satellite cell dysfunction, imbalance in protein turnover, adipose tissue infiltration, oxidative stress, mitochondrial dysfunction, inflammation, insulin resistance, capillary, and neuromuscular injury (Meng and Yu, 2010; Fulop et al., 2017; Pagano et al., 2018). The detailed biological mechanism of aginginduced sarcopenia can be referred to other excellent review (Englund et al., 2021; Kim et al., 2021), which will not be repeated here.

Aging and oxidative stress

Of all the potential etiological bases of sarcopenia, ROS production, correlated oxidative damage, and shortcomings in redox signaling have been repeatedly proved to be closely related to various forms of muscle pathophysiology during aging (Jackson, 2016; Damiano et al., 2019). For example, muscle fibers isolated from older mice generate more ROS than young adult mice (Englund et al., 2021). During aging, ROS basal levels are increased in muscles and satellite cells (Minet and Gaster, 2012; Palomero et al., 2013), and high levels of ROS have been proved to enhance DNA damage, mitochondrial dysfunction and protein damage. Specifically, elevated ROS levels cause oxidation of DNA, proteins, and lipids; increased protein carbonylation; impaired myogenic proteins and autophagy process; and inhibited skeletal muscle cell differentiation (Scherz-Shouval et al., 2007; Sandiford et al., 2014; Sakellariou et al., 2016). However, with aging, more ROS is generate even the cells in quiescent state, which mainly from mitochondria and NADPH oxidase (NOX) (Jackson, 2016; Damiano et al., 2019). Accumulated ROS also trigger apoptotic signaling cascades with aging (Meng and Yu, 2010; Barbieri and Sestili, 2012). It is reported that activation of mitochondrial cystathione non-dependent apoptosis, cystathione 2 and JNK-mediated apoptosis leading to agerelated muscle loss (Braga et al., 2008; Marzetti et al., 2008). Interestingly, ROS is the cause of neuromuscular junction dysfunction in myopenia. The more ROS are produced in the neuromuscular junction region of older mice, which is related to the decrease of neurotransmitter release (Ivannikov and Van Remmen, 2015). This may lead to impaired action potential generation and result in reduced muscle strength associated with sarcopenia. In addition, ROS may boost anabolic resistance in sarcopenia at multiple levels. Increased levels of ROS, such as H₂O₂, can suppress the phosphorylation of mTOR, Akt, and downstream targets p70S6K and 4E-BP1 (Powers et al., 2016; Gomez-Cabrera et al., 2020). The common compensatory response to ROS accumulation is the enhanced antioxidant defenses of cells (Ji, 2015). Most studies showed increased activity of superoxide dismutase 1 (SOD1) (Sullivan-Gunn and Lewandowski, 2013), superoxide dismutase 2 (SOD2) (Gianni et al., 2004), catalase (CAT) (Gianni et al., 2004; Palomero et al., 2013; Sullivan-Gunn and Lewandowski, 2013), and glutathione peroxidase (GPx) (Palomero et al., 2013), and it has observed age-dependent alterations in antioxidant enzyme activity in muscle (Ji, 1993). Although antioxidant enzyme activity increases with aging, this compensatory adaptation mechanism does not fully offset the increase in oxidative stress. The excessive ROS generated in aging muscles might restrain the key components of the Akt/mTOR pathway, thus limiting their response to exercise stimulation (Powers et al., 2016; Gomez-Cabrera et al., 2020). In fact, decreasing oxidative stress during aging could enhance exercise adaptability. For instance, antioxidant supplements can restore the stimulation of protein synthesis by age-related deficient leucine in rats (Marzani et al., 2008). The above mentioned evidence suggests that excessive ROS-induced oxidative stress can inhibit protein synthesis during sarcopenia.

As one of the key events of aging, muscle loss affects the overall cellular homeostasis. Muscle plasticity allows muscles to tolerate any stress-related biological alteration. However, aging also affects muscle malleability in terms of energy intake and consumption (Distefano and Goodpaster, 2018). ROS-mediated oxidative stress is a common phenomenon under the conditions of energy intake and consumption, which may be a significant target of age-related complications. Skeletal muscle satellite cells can maintain the ability of muscle adaptation and regeneration, which is important to delay or prevent aging. The quiescent satellite cells rely on glycolysis to produce energy, which is conducive to maintaining low levels ROS in satellite cell pool and keeping the redox system in an active state (Radak et al., 2008; Thirupathi et al., 2020). However, the damage of the satellite cell cycle by ROS-induced oxidative stress may lead to the failure of quiescent satellite cells to activate in the process of aging, thereby effecting the ability of muscle regeneration and muscle contraction. In addition, studies have demonstrated that aged muscle have impaired regenerative capability and muscle functions (Joanisse et al., 2016; Pisot et al., 2016). After muscle atrophy, aged muscle also fail to regenerate, while young muscle tissue can recover (Verdijk et al., 2009). Although the mechanism of this regeneration process is still elusive, ROS-induced oxidative stress may be one of the reasons for this failure in muscle growth capacity. However, it should be noted that it remains unclear whether ROS can act as a direct signal transduction in the case of aging. In addition, only few genetic manipulations aimed at reducing ROS activities have led to increased lifespan in mammals, suggesting that oxidative stress may play a limited role in aging but that reducing oxidative stress delays lesions (Schriner et al., 2005; Yoshida et al., 2005; Salmon et al., 2010).

ROS may be a potential gatekeeper of mitochondrial dysfunction in aged skeletal muscle

Much of the interest in ROS and aging involves the potential role of mitochondria as a resource and target of ROS (Jang and Van Remmen, 2009; Salmon et al., 2010). Skeletal muscle fibers are enriched with mitochondria whose function is to synthesize ATP through the oxidative phosphorylation to supply energy for muscle contraction. Accumulating evidence have showed that mitochondrial dysfunctional may play a core role in the pathogenesis of sarcopenia, while maintaining the normal mitochondrial function is crucial for skeletal muscle development (Calvani et al., 2013; Bhatti et al., 2017).

The maintenance of mitochondrial function is a dynamic balance process of mitochondrial quality control, which is an elaborate and complex network in eukaryotic cells. Specifically, it maintain mitochondrial homeostasis through four key processes: mitochondrial protein homeostasis, biogenesis, kinetics and autophagy. The mitochondrial dysfunction magnified by defects in the quality control process is increasingly considered to be the main pathophysiological mechanism of sarcopenia. Skeletal muscle mitochondria are interconnected to form a dynamic network, and their composition and morphology can be changed by the dynamic regulation of mitochondrial fission and fusion (Ogata and Yamasaki, 1997; Picard et al., 2011; Hood et al., 2019). Among them, mitochondrial fusion is regulated by MFN1/2 and OPA1, while mitochondrial fission is regulated by DRP1 and FIS1. A recent study has demonstrated that the morphology and dynamics of mitochondria changed during aging (Leduc-Gaudet et al., 2015). Evidence has demonstrated that the level of FIS1 and the activity of DRP1 decrease in aging cells, which may affect satellite cell activation and skeletal muscle regeneration (Mai et al., 2010). In mice, muscle aging is associated with increased myofibrillar mitochondrial complexity and expansion of mitochondria under the sarcolemma (Leduc-Gaudet et al., 2015). The MFN2/ DRP1 ratio increased significantly in aged mice, suggesting an increase in mitochondrial fusion capacity during skeletal muscle aging (Leduc-Gaudet et al., 2015). In short, the control of mitochondrial dynamics may represent a mechanism of skeletal muscle aging, leading to the accumulation of mitochondrial dysfunction during aging, thereby leading to the development of sarcopenia. Furthermore, recent studies suggest that the alterations in mitophagy may be associated with the development of sarcopenia. It has been found that the Parkin-PINK1 mitophagy regulatory axis may be altered during muscle aging in humans, suggesting a phenomenon of mitophagy deficiency during muscle aging (Gouspillou et al., 2013; Drummond et al., 2014; Gouspillou et al., 2014). In addition, the voltage-dependent anion channels (VDAC) help to recruit Parkin to dysfunctional mitochondria, which is a necessary step to start mitochondrial autophagy (Faitg et al., 2017). However, the Parkin/VDAC ratio appears to be reduced in muscle of older men (Gouspillou et al., 2014). A recent study demonstrated that overexpression of Parkin in Drosophila can increase its lifespan, which is related to the increase of citrate synthase activity and the decrease of protein aggregates in aging muscle (Rana et al., 2013). Taken together, these results suggest that the mitophagy mechanisms are altered and autophagic potential is reduced during muscle aging. And such alterations would represent an attractive cellular mechanism to explain the accumulation of dysfunctional mitochondria during muscle aging.

In fact, among the various theories that have been proposed to explain sarcopenia, the mitochondrial aging theory assumes that the accumulation of mitochondrial dysfunction plays a causal role in muscle atrophy with aging. This theory is generally accepted that oxidative damage to mitochondrial proteins, lipids and DNA will accumulate with aging due to the generation of ROS inherent in the activity of the respiratory chain (Faitg et al., 2017). The ROS produced by mitochondria are also critical for maintaining muscle functions such as skeletal muscle development, muscle mass, mitochondrial biogenesis and injury repair. However, the excessive ROS will cause oxidative damage to many cell molecules and cell structures, and the accumulation of this damage is the root of cellular dysfunction and eventually leads to cellular aging (Harman, 1956). Moreover, this oxidative damage is considered to exacerbate the production of mitochondrial ROS, weaken the ability of mitochondria to adequately match cellular ATP demands and trigger mitochondria-mediated apoptosis (Faitg et al., 2017). Multiple studies agree that muscle aging is related to an increase of the oxidative stress markers (Chabi et al., 2008; Sinha-Hikim et al., 2013). In addition, several studies have also reported increased ROS production by muscle mitochondria during aging (Capel et al., 2005; Dirks et al., 2006; Chabi et al., 2008; Lanza et al., 2012). A study indicated that overexpressed CAT in mitochondria attenuated the adverse effects of aging on muscle strength in mice (Umanskaya et al., 2014). A recent study revealed that the mitochondrial calcium uptake family member 3 (MICU3) expression was down-regulated during aging in skeletal muscle, which is correlated with reduced myogenesis but elevated oxidative stress and apoptosis (Yang et al., 2021). Conversely, MICU3 reconstitution enhances antioxidants, prevents the accumulation of mitochondrial ROS, reduces apoptosis and increases myogenesis (Yang et al., 2021). However, it should be noted that although most studies agree that muscle aging is associated with the occurrence of oxidative stress, the involvement of mitochondria in this process is still unclear.

The differentiation of skeletal muscle satellite cells (SMSCs) into myotubes and further fusion into myofibers are decisive steps in muscle regeneration. Evidence suggests that increased ROS negatively affects the regenerative capacity of SMSCs during aging, which is partly mediated by increased ROS-induced oxidative stress. Specifically, the proliferation and differentiation of SMSCs decrease with aging (Dalle et al., 2017), and more SMSCs enter a permanent aging stage from reversible quiescent state in aging muscle (Sousa-Victor et al., 2014). At the same time, the antioxidant capacity of SMSCs also decreases with aging (Fulle et al., 2005), and what is worse, the production of ROS continuously increases during this process (Minet and Gaster, 2012). However, the exact mechanism of ROS-mediated reduction of SMSCs regeneration capacity during aging remains unclear. Current evidence indicates that multiple redox sensitive signaling in SMSCs are altered during aging, such as p38/MAPK, Wnt, Notch, and JAK/STAT3 (Szentesi et al., 2019; Foreman et al., 2021). It is unclear why these pathways are altered with aging, and why they no longer adapt to aging.

10.3389/fcell.2022.964130

Although aged SMSCs produce more ROS, it is conceivable that the inhibition of ROS could restore the proliferation and regeneration capabilities of aged SMSCs (Garcia-Prat et al., 2016).

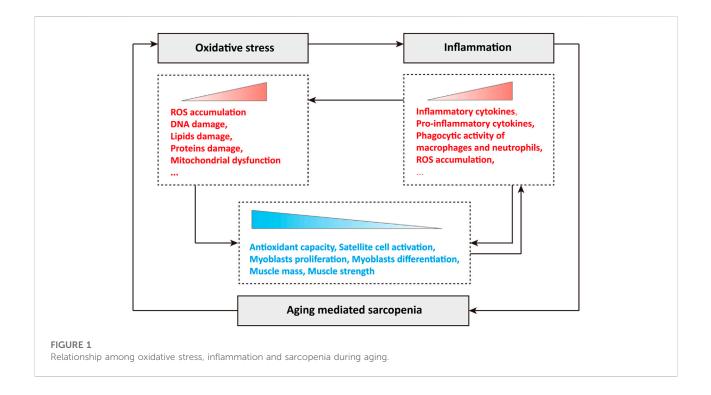
Skeletal muscle aging and oxidative stress mediated inflammatory response

A systemic, chronic, low-grade, sterile inflammatory state in aging was termed inflammatory aging (Cevenini et al., 2013). The inflammatory aging contributes to changes in skeletal muscle properties and sarcopenia, and mediates Alzheimer's disease and cognitive deficits in the elderly (Dalle et al., 2017; Zembron-Lacny et al., 2019). Inflammation is highly prevalent phenomenon in the elderly populations and is typified by increased levels of blood inflammatory markers in tissues and cells, leading to an increasing risk of chronic disease, weakness, disability and early death (Ferrucci and Fabbri, 2018). Inflammatory cells and muscle cells can generate inflammatory cytokines, contribute to the occurrence and development of sarcopenia, which is characterized by lower muscle mass, weakened muscle strength, and worse physical function, leading to an increased possibility of adverse events. Inflammatory cytokines, including pro-inflammatory and antiinflammatory cytokines interleukin 6 (IL-6) and tumor necrosis factor alpha (TNF- α), have been repeatedly proved to be closely associated with sarcopenia in humans and animals (Haddad et al., 2005; Dalle et al., 2017). Normally, the balance between skeletal muscle anabolism and catabolism is maintained by proinflammatory cytokines, but the expression of pro-inflammatory cytokines were increased during muscle atrophy, and leading to increased catabolism. Also, pro-inflammatory cytokines can inhibit skeletal muscle cells protein synthesis, impair muscle integrity and function, and thus lead to sarcopenia (Sharma and Dabur, 2020). The anti-inflammatory cytokines (such as IL-4, IL-10 and IL-15) can counteract the expression and activity of pro-inflammatory cytokines, especially IL-6 and TNF-a, to reduce muscle atrophy and delay sarcopenia (Marzetti et al., 2009; Hofmann et al., 2012). In addition, the anti-inflammatory cytokine, such as IL-4, can better glucose metabolism in muscle cells, and serve as a myogenic cells recruitment factor during muscle growth to promote muscle regeneration (Horsley et al., 2003; Heredia et al., 2013; Chang et al., 2019). The activation of the NF-KB pathway is a major trigger of inflammation in inactive muscle. The transgenic mice that overexpress IKK β , the main activator of NF-KB, have up-regulated expression of MuRF1, leading to the appearance of muscle atrophy (Cai et al., 2004). On the contrary, the absence of p105/p50 subunits of NF-κB has been proved to have an inhibitory effect on muscle atrophy (Hunter and Kandarian, 2004). As mentioned above, NF-KB activation is mainly related to the excessive production of proinflammatory cytokines. Also, NF- κ B can stimulate the expression of MuRF1, thereby degrading muscle contraction protein (Cai et al., 2004). In addition, TNF- α is the strongest activator of the NF- κ B pathway, which contributes to a positive feedback loop via activation of NF- κ B, thereby inducing TNF- α and driving NF- κ B-mediated muscle atrophy (Cai et al., 2004).

As mentioned above, increased ROS is not a coincidence with age and sarcopenia. Oxidative stress, as an effective activator of macrophages and neutrophils, increases due to intense exercise, aging and ischemia (Bartnik et al., 2000). The phagocytic activity of macrophages and neutrophils leads to the generation and release of cytotoxic proteases, pro-inflammatory cytokines and free radicals, including reactive oxygen and nitrogen species (RONS) (Lu and Wahl, 2005). It is reported that the sensitivity of elderly rats to inflammatory injury increases, resulting in more pro-inflammatory cytokines, RONS and transcription factors (Chung et al., 2002). Among them, transcription factor NF-KB stimulates the expression of proinflammatory gene and regulates subsequent inflammation and muscle recovery from injury (Hnia et al., 2008). In aged rat muscle, age-related oxidative stress appears to be associated with NF-βB, macrophage infiltration and neutrophil activity (Ghaly and Marsh, 2010). In younger rodents, an appropriate inflammatory response is essential for muscle regeneration (Shen et al., 2008). It has been proved that this inflammatory response is regulated by the cyclooxygenase two and TGF-β pathways (Bondesen et al., 2004; Shen et al., 2008). Furthermore, the balance of notch-activation and TGF- β inhibition of satellite cells in aged muscle determines the differentiation of satellite cells into myogenic cells, which eventually dictates whether myogenesis and muscle regeneration in injured muscle can occur successfully (Carlson et al., 2008). Therefore, increased inflammatory response may lead to excessive TGF-B1 production, fibrosis scar formation and diminished muscle regeneration (Li et al., 2004).

In short, anti-inflammatory cytokines are a potential therapeutic target for sarcopenia. To maximize muscle regeneration efficiency, it is critical to minimize secondary inflammation of destructive leukocytes and maximize regeneration-promoting inflammatory cytokines and growth factors. However, studies on the interaction between sarcopenia and anti-inflammatory cytokines are very few, and need to be further investigated in depth.

In conclusion, oxidative stress and inflammation play a crucial role in the occurrence and development of sarcopenia during aging. Briefly, the production of ROS leads to oxidative stress in cells, and subsequent oxidative damage, such as DNA damage, lipid damage and protein damage, disrupts mitochondrial dynamics and quality control, resulting in mitochondrial dysfunction (Figure 1). Oxidative stress stimulates the abnormal accumulation of inflammatory cytokines, especially pro-inflammatory cytokines, activates the phagocytic activity of macrophages and neutrophils, and leads to



the production and release of cytotoxic protease, proinflammatory cytokines and ROS, and the excessive ROS further stimulates the occurrence of oxidative stress (Figure 1). During aging, the synergistic effects of oxidative stress and inflammation greatly reduce cellular antioxidant capacity, significantly inhibit the activation of skeletal muscle satellite cells, the proliferation and differentiation of myoblasts, and causing loss of muscle mass and muscle strength, which manifests as sarcopenia (Figure 1). In turn, sarcopenia during aging further exacerbates oxidative stress and inflammation, forming a poor vicious cycle (Figure 1).

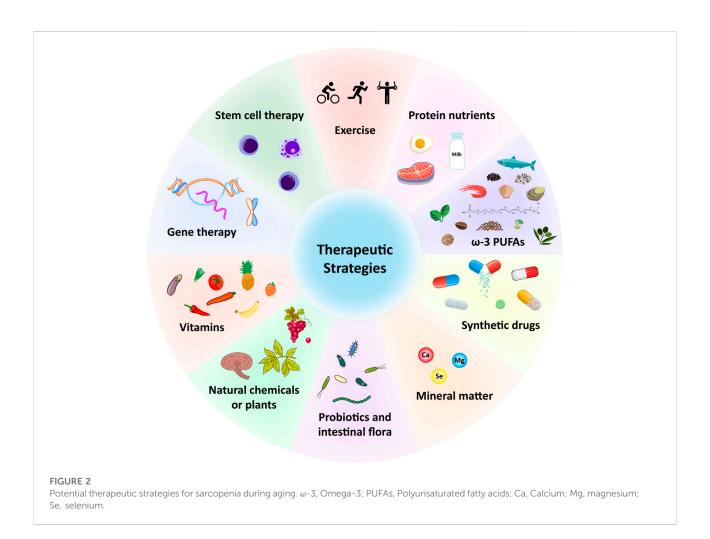
Potential therapeutic strategies for sarcopenia during aging

Multiple therapeutic strategies have been attempted and have proven effective in treating sarcopenia during aging. Since it is affected by many factors, the combination of exercise, potential antioxidant therapy and anti-inflammatory effect represents probably the most efficient strategy to prevent or treat skeletal muscle aging and sarcopenia. A large number of antioxidant and anti-aging therapies have been proposed (Figure 2; Table 1). For example, protein, amino acid and peptide supplements are thought to be highly effective in enhancing muscle metabolism and protecting muscle wasting during a prolonged immobilization (Dardevet et al., 2012) (Table 1). The supplementation of minerals such as calcium, magnesium, and selenium can effectively prolong or enhance age-related declines

in muscle function and strength (Veronese et al., 2014; Ter Borg et al., 2016; Fodor et al., 2020) (Table 1). Vitamins and fatty acids play a critical role in the prevention of sarcopenia through the inhibition of oxidative stress and relevant genes (Wang et al., 2021) (Table 1). In addition, a large number of phytochemicals such as polyphenols, polysaccharides, flavonoids, alkaloids, and triterpenoids may prevent and improve age-related sarcopenia through antioxidant and or anti-inflammatory (Li et al., 2004) (Table 1). Also, probiotics and intestinal flora have recently been considered to play a crucial role in the prevention of sarcopenia (An et al., 2019; Lahiri et al., 2019; Ni et al., 2019) (Table 1). For more detailed description, please refer to the excellent comments of Wang et al. and Nikawa et al. (Nikawa et al., 2021; Wang et al., 2021). Here, we only discuss some potential antioxidant substances or anti-inflammatory substances of interest for laboratory or preclinical studies to delay skeletal muscle aging and sarcopenia.

Carnosine

Carnosine is a dipeptide that is synthesized *in vivo* from β alanine and L-histidine in skeletal muscle (Jukic et al., 2021). Although its physiological role is not fully clear, carnosine is a nonenzymatic free radical scavenger and native antioxidant with neuroprotective and anti-inflammatory properties (Bellia et al., 2011). For example, carnosine can scavenge ROS and α - β unsaturated aldehydes produced by lipid peroxidation as a result of oxidative stress (Jukic et al., 2021). Carnosine



alleviates diabetic nephropathy and delay aging as it protects mesangial cells and podocytes (Jukic et al., 2021). Carnosine also resists saccharification by chelating divalent metal ions (Jukic et al., 2021). The amino acid β -alanine, as its main component, is often used as a nutritional supplement for athletes to improve exercise and muscle contraction (Jukic et al., 2021). In addition, its many beneficial effects are well confirmed, including pH buffering capacity, heavy metal chelating and antisaccharification activity (Hipkiss, 2009; Vistoli et al., 2012; Ghodsi and Kheirouri, 2018). It has been shown that longterm oral β -alanine can significantly increase the content of carnosine in skeletal muscle, which is the main site of carnosine generation and accumulation in the body (Derave et al., 2010). Similarly, another study indicated that β -alanine intake can effectively enhance muscle carnosine content, thereby improving exercise capacity in older subjects (del Favero et al., 2012). A large number of studies have provided evidence for the beneficial effects of carnosine supplements on cytokine release, and the excessive expression of the pro-inflammatory IL-6, IL-8 and TNF- α is involved in the inflammatory process in digestive system, especially in intestine, which can induce inflammatory

bowel disease (Katsanos and Papadakis, 2017). A few studies have demonstrated that carnosine inhibits the secretion and expression of IL-8 stimulated by hydrogen peroxide and TNF- α (Son et al., 2008). Altogether, these encouraging results demonstrate that supplementation with carnosine or its precursor substances may improve muscle strength, endurance, and function in the elderly. Therefore, considering the beneficial characteristics and its clinical application, carnosine is considered as a highly effective therapy without side effects.

Ganoderma lucidum

Ganoderma lucidum (*G. lucidum*, *Lingzhi*) is a large whiterot macrofungus of basidiomycete, which is widely used as "the mushroom of immortality" in China. In Chinese folklore, it is considered to be a panacea for various diseases. The spores, basidiocarp, and mycelia of *Ganoderma lucidum* are reported to contain about 400 different biologically active compounds, mainly including triterpenoids, polysaccharides, sterols, TABLE 1 The functions of potential therapeutic strategies for sarcopenia during aging.

Therapeutic strategies Exercise		Functions Improve muscle strength and increase muscle mass
	Carnosine (β-alanine)	ROS scavenging, antioxidant, anti-inflammatory, resist saccharification, neuroprotective properties (Bellia et al., 2011; Jukic et al., 2021)
Mineral matter (Calcium, magnesium, selenium)		Improve muscle performance and delay the decline of muscle strength (Veronese et al., 2014; Ter Borg et al., 2016; Fodor et al., 2020)
Vitamins (VC, VD, VE)		Eliminate both water-soluble and fat-soluble free radicals simultaneously (Geng et al., 2021), maintain muscle mass and strength; prevent or treat muscle atrophy (Kondo et al., 1992; Makanae et al., 2013; Nakamura et al., 2020)
ω-3 PUFAs (ALA, EPA, DPA, DHA)		Anti-inflammation, anti-aging, improving mitochondrial function, promoting myoblast differentiation and myotube fusion, enhancing muscle quality and strength (Smith et al., 2015; Chen et al., 2016; Custodero et al., 2018; Lovsletten et al., 2018; Tan et al., 2018; Xu et al., 2018)
Natural chemicals or plants	Polyphenols (Resveratrol, anthocyanins, phenolic Acids, Flavones)	Anti-inflammatory, antioxidant, improve mitochondrial quality, resist muscle atrophy (Salucci and Falcieri, 2020; Nikawa et al., 2021)
	Ganoderma lucidum	Inhibiting the production of ROS and lipid peroxidation, free radical scavenging, antioxidant, anti-aging, reduce muscle mass loss (Thyagarajan et al., 2006; Sanodiya et al., 2009; Zhonghui et al., 2014; Jatwani and Tulsawani, 2021)
Probiotics and intestinal flora		Enhance muscle mass and strength, anti-inflammatory (Ticinesi et al., 2019; Liu et al., 2021; van Krimpen et al., 2021)
Synthetic drugs	Anti-inflammatory Drugs (Thalidomide, tocilizumab, infliximab)	Preventing muscle atrophy via inhibition of inflammatory cytokines and their downstream effectors (Guasconi and Puri, 2008)
	Enobosarm	Induced muscle protein synthesis (Nikawa et al., 2021)
Stem cell therapy (Mesenchymal stem cell)		Increase skeletal muscle weight and muscle fiber cross-sectional area, activate resident skeletal muscle stem cells (Li et al., 2016; Wang et al., 2018)
Gene therapy (miR-1, miR-206, miR-133a, miR-133b, miR-208b)		Regulate inflammation and redox signals, regulate mitochondrial dysfunction and skeletal muscle regeneration (Barbiera et al., 2020)

Note: Whey protein, one of the primary proteins found in dairy products; VC, vitamin C; VD, vitamin D; VE, vitamin E; ALA, α-linolenic acid; EPA, eicosapentaenoic acid; DPA, docosapentaenoic acid; ω-3, Omega-3; PUFAs, Polyunsaturated fatty acids; ROS, reactive oxygen species.

steroids, nucleotides, fatty acids, peptides, proteins and trace elements, which makes Ganoderma lucidum have various pharmacological effects (Sanodiya et al., 2009). Ganoderma lucidum has been applied for thousands of years as an elixir, but its anti-aging effects still unrevealed, especially in skeletal muscle. Aging is associated with oxidation stress, free radical product and immunoregulation. It is currently believed that Ganoderma lucidum can extend lifespan through inhibiting the production of ROS, lipid peroxidation and advanced oxidation protein products (Sanodiya et al., 2009). In addition, it has antioxidant activity and immunomodulatory via enhancing free radical scavenging activity and diminishing iron antioxidant capacity (Sanodiya et al., 2009). The biologically active components of Ganoderma lucidum that have anti-aging related functions include triterpenoids, polysaccharides, and peptides, which all have antioxidant properties. For example, Ganoderma lucidum triterpenes can elevate the levels of SOD, GPx, CAT, and GSH in brain and liver tissues, protect the body from protein and lipid peroxidation that induced by oxidative stress (You and Lin, 2003). In addition, Ganoderma lucidum polysaccharides (GLPS-I, GLPS-II, GLPS-III, GLPS-IV) can increase hydroxyl and 2,2-diphenyl-1-picrylhydrazil radical scavenging activities and metal chelating activities (Shi et al., 2013; Wang et al., 2017). A study continuously administered GLPS (50, 100 and 200 mg/kg/day) to male mice for 28 days and found that GLPS could enhance the antioxidant enzymes activity in mouse skeletal muscle, including SOD, GPx, CAT, and reduce the level of malondialdehyde, indicating that the supplementation of GLPS could reduce exercise-induced oxidative stress in skeletal muscle (Zhonghui et al., 2014). Ganoderma lucidum peptide diminish lipid peroxidation primarily by scavenging hydroxyl radicals (*OH), which are the main activated ROS that participate in lipid oxygenation (Thyagarajan et al., 2006). Ganoderma lucidum peptide prevents lipid metabolism-related domino reaction through response with free [•]OH (Sun et al., 2004). In addition, Ganoderma lucidum peptide can eliminate excessive free radicals and chelate metals ions to minimize metal-induced oxidation (Sun et al., 2004). Ganoderma lucidum polysaccharide peptide, the combination of polysaccharide and peptide, can reduce the oxidase NADPH and the NADPH-dependent ROS production and diminish malonaldehyde levels in a kidney ischemia-reperfusion model, while enhancing the activities of antioxidant enzymes SOD, Mn-SOD, GSH, GSH-px and CAT (Sun et al., 2004; Wang et al., 2017). A recent study showed that supplementation with an aqueous extract of Ganoderma lucidum attenuated the loss of

muscle mass in rats through preventing oxidative stress and regulating myogenesis markers under low-pressure hypoxia (Jatwani and Tulsawani, 2021). Nevertheless, the anti-aging effect of *Ganoderma lucidum* is still a mystery, and the potential anti-aging mechanism of its clinical application remains to be revealed.

Omega-3 polyunsaturated fatty acid

Polyunsaturated fatty acids (PUFAs) are a class of fatty acids whose molecular structure is featured by two or more double bonds. PUFAs family mainly includes Omega-3 (ω -3) and Omega-6 (ω -3) family. ω -3 PUFAs are distinguished by the fact that the first double bond is located on the 3rd carbon. ω -3 PUFAs are mainly α -linolenic acid (ALA), eicosapentaenoic acid (EPA), docosapentaenoic acid (DPA) and docosahexaenoic acid (DHA). Among them, EPA, DPA and DHA are regarded as long-chain PUFAs, while ALA is recognized as a short-chain PUFAs (Vannice and Rasmussen, 2014). Accumulating evidence showed that ω -3 PUFAs have the functions of anti-inflammatory, anti-aging, anti-cancer, and have potential effects on regulating blood lipids and blood pressure, improving cardiovascular and cerebrovascular diseases, improving insulin resistance and diabetes treatment.

A recent meta-analysis confirmed a reduced c-reactive protein and IL-6 with ω -3 PUFAs supplementation in older adults (Custodero et al., 2018). Continuous supplementation of EPA and DHA in the elderly for 4 weeks can significantly reduce the level of IL-1 β , IL-6 and TNF- α , and this effect is more obvious after 8 weeks (Tan et al., 2018). As mentioned previously, chronic low-grade inflammation is considered to play a key role in the progression of sarcopenia. ω-3 PUFAs are potential therapeutics for the treatment of sarcopenia owing to their anti-inflammatory and anti-aging characteristics. Hence, the suppressing of this low-grade inflammation is generally recognized as a potential mechanism by which ω -3 PUFAs may antagonize sarcopenia. For example, ω -3 PUFAs supplementation for 6-months in healthy older adults dramatically enhanced muscle volume and strength compared with the placebo group (Smith et al., 2015). Additional evidence suggests that ω -3 PUFAs also might exert anabolism effects on muscle by activating mTOR signaling and reducing insulin resistance (Dupont et al., 2019). As mentioned above, the activation and functions of SMSCs are influenced by oxidative stress and inflammation during myogenesis, ω-3 PUFAs play a critical role in this process. For example, SMSCs were incubated with 50 mM DHA for 2 days found that DHA treatment resulted in longer multinucleated myotubes and higher myotube fusion rate compared to control (Xu et al., 2018). Treatment in human SCMSs with 100 µM EPA for 24 h increased cellular oxygen consumption rate, basal respiration, maximal respiration and proton leak, suggesting that EPA improves mitochondrial

function during myogenesis (Lovsletten et al., 2018). Additionally, co-culture of C2C12 myotubes with palmitic acid, EPA and DHA for 16 h can significantly inhibit the palmitic acid-induced expression of proinflammatory cytokine (Chen et al., 2016). In conclusion, ω -3 PUFAs are promising in the prevention or treatment of sarcopenia, either alone or in combination with classical treatment strategies. In addition, combined with exercise intervention, the supplementation of ω -3 PUFAs may enhance the gains in muscle mass and function obtained through exercise intervention.

Vitamins

Vitamins have been shown to play a critical role in preventing muscle atrophy by inhibiting oxidative stress and related genes. Vitamin C has the potential to treat or prevent muscle atrophy in mice, and its deficiency leads to muscle atrophy, while the muscle atrophy recovered after about 3 months of continuous vitamin C supplementation, which was attributed to the excessive production of ROS and the elevated expression of muscle atrophy marker genes MAFbx and MuRF1 (Takisawa et al., 2019). In addition, oral vitamin C diminished overload-induced skeletal muscle hypertrophy in male Wistar rats (Makanae et al., 2013). Dietary vitamin C supplementation helps reduce age-related muscle loss in the elderly (Lewis et al., 2020). The lack of Vitamin D might lead to muscle atrophy, while oral vitamin D prevents immobilization-induced muscle atrophy in mice through the vitamin D receptor (Nakamura et al., 2020). In addition, as an active form of Vitamin D, 1,25-dihydroxyvitamin D deficiency can lead to the development of age-related sarcopenia by inducing oxidative stress, muscle cell senescence and senescence-associated secretory phenotype, and inhibiting skeletal muscle regeneration (Yu et al., 2021). The possible mechanisms of vitamin D are associated with the elevated MAFbx expression and protein degradation (Wang et al., 2021). Vitamin E also prevented hindlimb muscle atrophy induced by immobilization in Wistar rats (Kondo et al., 1992). As one of the subgroups of vitamin E, tocopherols have beneficial effects in various pathophysiological models (i.e., hindlimb suspension, exercise, and ischemic perfusion injury) by reducing oxidative stress, inflammation, and atrophy while increasing regenerative capacity (Chung et al., 2018). Supplementation with α -tocopherol before and during muscle atrophy reduces hindlimb-induced slow twitch muscle atrophy, contributing to an increase in type I and type IIA muscle fiber size and a decrease in muscle proteolysis rate (Servais et al., 2007). Meanwhile, a study found that supplementation with α tocopherol at a dose of 800 IU for 48 days decreased the expression of oxidative stress markers in both young and older sedentary men (Margaritis et al., 1997). A study also showed that a-tocopherol supplementation 1 h prior to

10.3389/fcell.2022.964130

exercise under hypoxic conditions reduced cellular damage and inflammation in healthy young men (Santos et al., 2016). Notably, combinations of different vitamins may exert more beneficial effects. For example, parallelly use of vitamin C and E is effective in eliminating both water- and fat-soluble free radicals simultaneously, and protecting cell membranes from oxidative damage induced by free radicals (Geng et al., 2021). Oral vitamin D and E supplementation can maintain muscle mass and muscle strength, and further improve the life quality of patients with sarcopenia (Bo et al., 2019). Taken together, vitamins have therapeutic potential to induce muscle hypertrophy and inhibit muscle atrophy by mechanisms that inhibit ROSmediated oxidative stress, activate mitochondrial biogenesis, and suppress the expression of genes associated with muscle atrophy.

Concluding remarks

Sarcopenia is one of the most severe geriatric syndromes. Under the trend of global aging, there is an urgent need to develop effective preventive and therapeutic measures. Although current research have clarified numerous pathophysiological factors in sarcopenia, such as the currently widely accepted mechanisms of oxidative stress and inflammation, and several therapeutic strategies have been proved to be effective in the treatment of sarcopenia. However, the majority of studies have been conducted in in vitro and in vivo models, while studies on humans are lacking. Overall, the molecular mechanisms that affect sarcopenia remain elusive. Focusing on the currently pathogenic mechanism of sarcopenia, it may be a long-term work to find, research and extensively apply antioxidants and anti-inflammatory substances, such as the synthesis of antioxidant and/or anti-inflammatory drugs. Currently, exercise is the only treatment that has been proven effective for delaying muscle loss. A combination of

References

Almada, A. E., and Wagers, A. J. (2016). Molecular circuitry of stem cell fate in skeletal muscle regeneration, ageing and disease. *Nat. Rev. Mol. Cell Biol.* 17 (5), 267–279. doi:10.1038/nrm.2016.7

An, J. M., Kang, E. A., Han, Y. M., Oh, J. Y., Lee, D. Y., Choi, S. H., et al. (2019). Dietary intake of probiotic kimchi ameliorated IL-6-driven cancer cachexia. *J. Clin. Biochem. Nutr.* 65 (2), 109–117. doi:10.3164/jcbn.19-10

Barbiera, A., Pelosi, L., Sica, G., and Scicchitano, B. M. (2020). Nutrition and microRNAs: Novel insights to fight sarcopenia. *Antioxidants (Basel)* 9 (10), E951. doi:10.3390/antiox9100951

Barbieri, E., and Sestili, P. (2012). Reactive oxygen species in skeletal muscle signaling. *J. Signal Transduct.* 2012, 982794. doi:10.1155/2012/982794

Bartnik, B. L., Juurlink, B. H., and Devon, R. M. (2000). Macrophages: Their myelinotrophic or neurotoxic actions depend upon tissue oxidative stress. *Mult. Scler.* 6 (1), 37-42. doi:10.1177/135245850000600108

exercise, potential antioxidant therapy, and antiinflammatory therapy probably is the most efficient strategy to prevent or treat sarcopenia during aging.

Author contributions

MC and YW conceptualized and wrote manuscript. SD assisted with the edited version. ZL and KY acquired the funding. All authors contributed to the article and approved the submitted version.

Funding

This work was supported by National Key Research and Development Project of China (2021YFF1000704), Natural Science Foundation of China (32072722), and National Transgenic Creature Breeding Grand Project (2016zx08008-003).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Bellia, F., Vecchio, G., Cuzzocrea, S., Calabrese, V., and Rizzarelli, E. (2011). Neuroprotective features of carnosine in oxidative driven diseases. *Mol. Asp. Med.* 32 (4-6), 258–266. doi:10.1016/j.mam.2011.10.009

Bhatti, J. S., Bhatti, G. K., and Reddy, P. H. (2017). Mitochondrial dysfunction and oxidative stress in metabolic disorders - a step towards mitochondria based therapeutic strategies. *Biochim. Biophys. Acta. Mol. Basis Dis.* 1863 (5), 1066–1077. doi:10.1016/j.bbadis.2016.11.010

Bischoff-Ferrari, H. A., Orav, J. E., Kanis, J. A., Rizzoli, R., SchloglM.Staehelin, H. B., et al. (2015). Comparative performance of current definitions of sarcopenia against the prospective incidence of falls among community-dwelling seniors age 65 and older. Osteoporos. Int. 26 (12), 2793–2802. doi:10.1007/s00198-015-3194-y

Bo, Y., Liu, C., Ji, Z., Yang, R., An, Q., Zhang, X., et al. (2019). A high whey protein, vitamin D and E supplement preserves muscle mass, strength, and quality of life in sarcopenic older adults: A double-blind randomized controlled trial. *Clin. Nutr.* 38 (1), 159–164. doi:10.1016/j.clnu.2017.12.020

Bondesen, B. A., Mills, S. T., Kegley, K. M., and Pavlath, G. K. (2004). The COX-2 pathway is essential during early stages of skeletal muscle regeneration. *Am. J. Physiol. Cell Physiol.* 287 (2), C475–C483. doi:10.1152/ajpcell.00088. 2004

Borsch, A., Ham, D. J., Mittal, N., Tintignac, L. A., Migliavacca, E., Feige, J. N., et al. (2021). Molecular and phenotypic analysis of rodent models reveals conserved and species-specific modulators of human sarcopenia. *Commun. Biol.* 4 (1), 194. doi:10.1038/s42003-021-01723-z

Braga, M., Sinha Hikim, A. P., Datta, S., Ferrini, M. G., Brown, D., Kovacheva, E. L., et al. (2008). Involvement of oxidative stress and caspase 2-mediated intrinsic pathway signaling in age-related increase in muscle cell apoptosis in mice. *Apoptosis* 13 (6), 822–832. doi:10.1007/s10495-008-0216-7

Breen, L., and Phillips, S. M. (2013). Interactions between exercise and nutrition to prevent muscle waste during ageing. *Br. J. Clin. Pharmacol.* 75 (3), 708–715. doi:10.1111/j.1365-2125.2012.04456.x

Cai, D., Frantz, J. D., Tawa, N. E., Melendez, P. A., Oh, B. C., Lidov, H. G. W., et al. (2004). IKKbeta/NF-kappaB activation causes severe muscle wasting in mice. *Cell* 119 (2), 285–298. doi:10.1016/j.cell.2004.09.027

Calvani, R., Joseph, A. M., Adhihetty, P. J., Miccheli, A., Bossola, M., Leeuwenburgh, C., et al. (2013). Mitochondrial pathways in sarcopenia of aging and disuse muscle atrophy. *Biol. Chem.* 394 (3), 393–414. doi:10.1515/hsz-2012-0247

Capel, F., Rimbert, V., Lioger, D., Diot, A., Rousset, P., Mirand, P. P., et al. (2005). Due to reverse electron transfer, mitochondrial H2O2 release increases with age in human vastus lateralis muscle although oxidative capacity is preserved. *Mech. Ageing Dev.* 126 (4), 505–511. doi:10.1016/j.mad.2004.11.001

Carlson, M. E., Hsu, M., and Conboy, I. M. (2008). Imbalance between pSmad3 and Notch induces CDK inhibitors in old muscle stem cells. *Nature* 454 (7203), 528–532. doi:10.1038/nature07034

Cevenini, E., Monti, D., and Franceschi, C. (2013). Inflamm-ageing. Curr. Opin. Clin. Nutr. Metab. Care 16 (1), 14–20. doi:10.1097/MCO.0b013e32835ada13

Chabi, B., Ljubicic, V., Menzies, K. J., Huang, J. H., Saleem, A., and Hood, D. A. (2008). Mitochondrial function and apoptotic susceptibility in aging skeletal muscle. *Aging Cell* 7 (1), 2–12. doi:10.1111/j.1474-9726.2007.00347.x

Chang, Y. H., Tsai, J-N., Chen, T-L., Ho, K-T., Cheng, H-Y., Hsiao, C-W., et al. (2019). Interleukin-4 promotes myogenesis and boosts myocyte insulin efficacy. *Mediat. Inflamm.* 2019, 4182015. doi:10.1155/2019/4182015

Chemello, F., Bean, C., Cancellara, P., Laveder, P., Reggiani, C., and Lanfranchi, G. (2011). Microgenomic analysis in skeletal muscle: Expression signatures of individual fast and slow myofibers. *PLoS One* 6 (2), e16807. doi:10.1371/journal. pone.0016807

Chen, S. C., Chen, P. Y., Wu, Y. L., Chen, C. W., Chen, H. W., Lii, C. K., et al. (2016). Long-chain polyunsaturated fatty acids amend palmitate-induced inflammation and insulin resistance in mouse C2C12 myotubes. *Food Funct.* 7 (1), 270–278. doi:10.1039/c5fo00704f

Chung, E., Mo, H., Wang, S., Zu, Y., Elfakhani, M., Rios, S. R., et al. (2018). Potential roles of vitamin E in age-related changes in skeletal muscle health. *Nutr. Res.* 49, 23–36. doi:10.1016/j.nutres.2017.09.005

Chung, H. Y., Kim, H. J., Kim, K. W., Choi, J. S., and Yu, B. P. (2002). Molecular inflammation hypothesis of aging based on the anti-aging mechanism of calorie restriction. *Microsc. Res. Tech.* 59 (4), 264–272. doi:10.1002/jemt.10203

Cruz-Jentoft, A. J., Bahat, G., Bauer, J., Boirie, Y., Bruyere, O., Cederholm, T., et al. (2019). Sarcopenia: Revised European consensus on definition and diagnosis. *Age Ageing* 48 (1), 601–631. doi:10.1093/ageing/afz046

Cruz-Jentoft, A. J., and Sayer, A. A. (2019). Sarcopenia. Lancet 393 (10191), 2636–2646. doi:10.1016/S0140-6736(19)31138-9

Custodero, C., Mankowski, R. T., Lee, S. A., Chen, Z., Wu, S., Manini, T. M., et al. (2018). Evidence-based nutritional and pharmacological interventions targeting chronic low-grade inflammation in middle-age and older adults: A systematic review and meta-analysis. *Ageing Res. Rev.* 46, 42–59. doi:10.1016/j. arr.2018.05.004

Cuthbertson, D., Smith, K., Babraj, J., Leese, G., Waddell, T., Atherton, P., et al. (2005). Anabolic signaling deficits underlie amino acid resistance of wasting, aging muscle. *FASEB J.* 19 (3), 422–424. doi:10.1096/fj.04-2640fje

Dalle, S., Rossmeislova, L., and Koppo, K. (2017). The role of inflammation in age-related sarcopenia. *Front. Physiol.* 8, 1045. doi:10.3389/fphys.2017.01045

Damiano, S., Muscariello, E., La Rosa, G., Di Maro, M., Mondola, P., and Santillo, M. (2019). Dual role of reactive oxygen species in muscle function: Can antioxidant dietary supplements counteract age-related sarcopenia? *Int. J. Mol. Sci.* 20 (15), E3815. doi:10.3390/ijms20153815

Dardevet, D., Rémond, D., Peyron, M-A., Papet, I., Savary-Auzeloux, I., and Moson, L. (2012). Muscle wasting and resistance of muscle anabolism: The

"anabolic threshold concept" for adapted nutritional strategies during sarcopenia. *ScientificWorldJournal* 2012, 269531. doi:10.1100/2012/269531

De Buyser, S. L., Petrovic, M., Taes, Y. E., Toye, K. R. C., Kaufman, J. M., Lapauw, B., et al. (2016). Validation of the FNIH sarcopenia criteria and SOF frailty index as predictors of long-term mortality in ambulatory older men. *Age Ageing* 45 (5), 602–608. doi:10.1093/ageing/afw071

del Favero, S., Roschel, H., Solis, M. Y., Hayashi, A. P., Artioli, G. G., Otaduy, M. C., et al. (2012). Beta-alanine (carnosyn) supplementation in elderly subjects (60-80 years): Effects on muscle carnosine content and physical capacity. *Amino Acids* 43 (1), 49–56. doi:10.1007/s00726-011-1190-x

Denison, H. J., Cooper, C., Sayer, A. A., and Robinson, S. M. (2015). Prevention and optimal management of sarcopenia: A review of combined exercise and nutrition interventions to improve muscle outcomes in older people. *Clin. Interv. Aging* 10, 859–869. doi:10.2147/CIA.S55842

Dirks, A. J., Hofer, T., Marzetti, E., Pahor, M., and Leeuwenburgh, C. (2006). Mitochondrial DNA mutations, energy metabolism and apoptosis in aging muscle. *Ageing Res. Rev.* 5 (2), 179–195. doi:10.1016/j.arr.2006.03.002

Distefano, G., and Goodpaster, B. H. (2018). Effects of exercise and aging on skeletal muscle. *Cold Spring Harb. Perspect. Med.* 8 (3), a029785. doi:10.1101/cshperspect.a029785

Drummond, M. J., Addison, O., Brunker, L., Hopkins, P. N., McClain, D. A., LaStayo, P. C., et al. (2014). Downregulation of E3 ubiquitin ligases and mitophagyrelated genes in skeletal muscle of physically inactive, frail older women: A crosssectional comparison. *J. Gerontol. A Biol. Sci. Med. Sci.* 69 (8), 1040–1048. doi:10. 1093/gerona/glu004

Dupont, J., Dedeyne, L., Dalle, S., Koppo, K., and Gielen, E. (2019). The role of omega-3 in the prevention and treatment of sarcopenia. *Aging Clin. Exp. Res.* 31 (6), 825–836. doi:10.1007/s40520-019-01146-1

Englund, D. A., Zhang, X., Aversa, Z., and LeBrasseur, N. K. (2021). Skeletal muscle aging, cellular senescence, and senotherapeutics: Current knowledge and future directions. *Mech. Ageing Dev.* 200, 111595. doi:10.1016/j.mad.2021. 111595

Faitg, J., Reynaud, O., Leduc-Gaudet, J. P., and Gouspillou, G. (2017). Skeletal muscle aging and mitochondrial dysfunction: An update. *Med. Sci.* 33 (11), 955–962. doi:10.1051/medsci/20173311012

Ferrucci, L., and Fabbri, E. (2018). Inflammageing: Chronic inflammation in ageing, cardiovascular disease, and frailty. *Nat. Rev. Cardiol.* 15 (9), 505–522. doi:10. 1038/s41569-018-0064-2

Fodor, J., Al-Gaadi, D., Czirjak, T., Olah, T., Dienes, B., Csernoch, L., et al. (2020). Improved calcium homeostasis and force by selenium treatment and training in aged mouse skeletal muscle. *Sci. Rep.* 10 (1), 1707. doi:10.1038/s41598-020-58500-x

Foreman, N. A., Hesse, A. S., and Ji, L. L. (2021). Redox signaling and sarcopenia: Searching for the primary suspect. *Int. J. Mol. Sci.* 22 (16), 9045. doi:10.3390/ ijms22169045

Frontera, W. R., and Ochala, J. (2015). Skeletal muscle: A brief review of structure and function. *Calcif. Tissue Int.* 96 (3), 183–195. doi:10.1007/s00223-014-9915-y

Fulle, S., Di Donna, S., Puglielli, C., Pietrangelo, T., Beccafico, S., Bellomo, R., et al. (2005). Age-dependent imbalance of the antioxidative system in human satellite cells. *Exp. Gerontol.* 40 (3), 189–197. doi:10.1016/j.exger.2004.11.006

Fulop, T., Larbi, A., Dupuis, G., Le Page, A., Frost, E. H., Cohen, A. A., et al. (2017). Immunosenescence and inflamm-aging as two sides of the same coin: Friends or foes? *Front. Immunol.* 8, 1960. doi:10.3389/fimmu.2017.01960

Garcia-Prat, L., Martinez-Vicente, M., Perdiguero, E., Ortet, L., Rodriguez-Ubreva, J., Rebollo, E., et al. (2016). Autophagy maintains stemness by preventing senescence. *Nature* 529 (7584), 37–42. doi:10.1038/nature16187

Geng, R., Kang, S. G., Huang, K., and Tong, T. (2021). Boosting the photoaged skin: The potential role of dietary components. *Nutrients* 13 (5), 1691. doi:10.3390/nu13051691

Ghaly, A., and Marsh, D. R. (2010). Aging-associated oxidative stress modulates the acute inflammatory response in skeletal muscle after contusion injury. *Exp. Gerontol.* 45 (5), 381-388. doi:10.1016/j.exger.2010.03.004

Ghodsi, R., and Kheirouri, S. (2018). Carnosine and advanced glycation end products: A systematic review. *Amino Acids* 50 (9), 1177–1186. doi:10.1007/s00726-018-2592-9

Gianni, P., Jan, K. J., Douglas, M. J., Stuart, P. M., and Tarnopolsky, M. A. (2004). Oxidative stress and the mitochondrial theory of aging in human skeletal muscle. *Exp. Gerontol.* 39 (9), 1391–1400. doi:10.1016/j.exger.2004.06.002 Gomez-Cabrera, M. C., ArC-Chagnaud, C., SAlvAdor-PAscuAl, A., Brioche, T., ChopArd, A., Olaso-Gonzalez, G., et al. (2020). Redox modulation of muscle mass and function. *Redox Biol.* 35, 101531. doi:10.1016/j.redox.2020.101531

Gouspillou, G., Picard, M., Godin, R., Burelle, Y., and Hepple, R. T. (2013). Role of peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1 α) in denervation-induced atrophy in aged muscle: Facts and hypotheses. *Longev. Heal.* 2 (1), 13. doi:10.1186/2046-2395-2-13

Gouspillou, G., Sgarioto, N., Kapchinsky, S., Purves-Smith, F., Norris, B., Pion, C. H., et al. (2014). Increased sensitivity to mitochondrial permeability transition and myonuclear translocation of endonuclease G in atrophied muscle of physically active older humans. *FASEB J.* 28 (4), 1621–1633. doi:10.1096/fj.13-242750

Guasconi, V., and Puri, P. L. (2008). Epigenetic drugs in the treatment of skeletal muscle atrophy. *Curr. Opin. Clin. Nutr. Metab. Care* 11 (3), 233–241. doi:10.1097/MCO.0b013e3282fa1810

Haddad, F., ZaldivarF.Cooper, D. M., and Adams, G. R. (2005). IL-6-induced skeletal muscle atrophy. *J. Appl. Physiol.* 98 (3), 911–917. doi:10.1152/japplphysiol. 01026.2004

Harman, D. (1956). Aging: A theory based on free radical and radiation chemistry. J. Gerontol. 11 (3), 298-300. doi:10.1093/geronj/11.3.298

Heredia, J. E., Mukundan, L., Chen, F. M., Mueller, A. A., Deo, R. C., Locksley, R. M., et al. (2013). Type 2 innate signals stimulate fibro/adipogenic progenitors to facilitate muscle regeneration. *Cell* 153 (2), 376–388. doi:10.1016/j.cell.2013.02.053

Hipkiss, A. R. (2009). Carnosine and its possible roles in nutrition and health. Adv. Food Nutr. Res. 57, 87–154. doi:10.1016/S1043-4526(09)57003-9

Hnia, K., Gayraud, J., Hugon, G., Ramonatxo, M., De La Porte, S., Matecki, S., et al. (2008). L-arginine decreases inflammation and modulates the nuclear factorkappaB/matrix metalloproteinase cascade in mdx muscle fibers. *Am. J. Pathol.* 172 (6), 1509–1519. doi:10.2353/ajpath.2008.071009

Hofmann, S. R., Rosen-Wolff, A., Tsokos, G. C., and Hedrich, C. M. (2012). Biological properties and regulation of IL-10 related cytokines and their contribution to autoimmune disease and tissue injury. *Clin. Immunol.* 143 (2), 116–127. doi:10.1016/j.clim.2012.02.005

Hood, D. A., Memme, J. M., Oliveira, A. N., and Triolo, M. (2019). Maintenance of skeletal muscle mitochondria in health, exercise, and aging. *Annu. Rev. Physiol.* 81, 19–41. doi:10.1146/annurev-physiol-020518-114310

Horsley, V., Jansen, K. M., Mills, S. T., and Pavlath, G. K. (2003). IL-4 acts as a myoblast recruitment factor during mammalian muscle growth. *Cell* 113 (4), 483–494. doi:10.1016/s0092-8674(03)00319-2

Hunter, R. B., and Kandarian, S. C. (2004). Disruption of either the Nfkb1 or the Bcl3 gene inhibits skeletal muscle atrophy. *J. Clin. Invest.* 114 (10), 1504–1511. doi:10.1172/JCI21696

Ivannikov, M. V., and Van Remmen, H. (2015). Sod1 gene ablation in adult mice leads to physiological changes at the neuromuscular junction similar to changes that occur in old wild-type mice. *Free Radic. Biol. Med.* 84, 254–262. doi:10.1016/j. freeradbiomed.2015.03.021

Jackson, M. J. (2016). Reactive oxygen species in sarcopenia: Should we focus on excess oxidative damage or defective redox signalling? *Mol. Asp. Med.* 50, 33–40. doi:10.1016/j.mam.2016.05.002

Jang, Y. C., and Van Remmen, H. (2009). The mitochondrial theory of aging: Insight from transgenic and knockout mouse models. *Exp. Gerontol.* 44 (4), 256–260. doi:10.1016/j.exger.2008.12.006

Jatwani, A., and Tulsawani, R. (2021). Ganoderma lucidum induces myogenesis markers to avert damage to skeletal muscles in rats exposed to hypobaric hypoxia. *High. Alt. Med. Biol.* doi:10.1089/ham.2020.0172

Ji, L. (1993). Antioxidant enzyme response to exercise and aging. Med. Sci. Sports Exerc. 25 (2), 225–231. doi:10.1249/00005768-199302000-00011

Ji, L. L. (2015). Redox signaling in skeletal muscle: Role of aging and exercise. *Adv. Physiol. Educ.* 39 (4), 352–359. doi:10.1152/advan.00106.2014

Joanisse, S., Nederveen, J. P., Baker, J. M., Snijders, T., Iacono, C., and Parise, G. (2016). Exercise conditioning in old mice improves skeletal muscle regeneration. *FASEB J.* 30 (9), 3256–3268. doi:10.1096/fj.201600143RR

Jukic, I., Kolobaric, N., Stupin, A., Matic, A., Kozina, N., Mihaljevic, Z., et al. (2021). Carnosine, small but mighty-prospect of use as functional ingredient for functional food formulation. *Antioxidants (Basel)* 10 (7), 1037. doi:10.3390/antiox10071037

Katsanos, K. H., and Papadakis, K. A. (2017). Inflammatory bowel disease: Updates on molecular targets for biologics. *Gut Liver* 11 (4), 455-463. doi:10. 5009/gnl16308

Kim, J. W., Kim, R., Choi, H., Lee, S. J., and Bae, G. U. (2021). Understanding of sarcopenia: From definition to therapeutic strategies. *Arch. Pharm. Res.* 44 (9-10), 876–889. doi:10.1007/s12272-021-01349-z

Kondo, H., MiuraM..NakagakI, I., SaSaki, S., and Itokawa, Y. (1992). Trace element movement and oxidative stress in skeletal muscle atrophied by immobilization. *Am. J. Physiol.* 262 (5 1), E583–E590. doi:10.1152/ajpendo.1992. 262.5.E583

Lahiri, S., Kim, H., Garcia-Perez, I., Reza, M. M., Martin, K. A., Kundu, P., et al. (2019). The gut microbiota influences skeletal muscle mass and function in mice. *Sci. Transl. Med.* 11 (502), eaan5662. doi:10.1126/scitranslmed.aan5662

Lanza, I. R., Zabielski, P., Klaus, K. A., Morse, D. M., Heppelmann, C. J., Bergen, H. R., et al. (2012). Chronic caloric restriction preserves mitochondrial function in senescence without increasing mitochondrial biogenesis. *Cell Metab.* 16 (6), 777–788. doi:10.1016/j.cmet.2012.11.003

Leduc-Gaudet, J. P., Picard, M., St-Jean Pelletier, F., Sgarioto, N., Auger, M. J., Vallee, J., et al. (2015). Mitochondrial morphology is altered in atrophied skeletal muscle of aged mice. *Oncotarget* 6 (20), 17923–17937. doi:10.18632/oncotarget. 4235

Lewis, L. N., Hayhoe, R. P. G., Mulligan, A. A., Luben, R. N., Khaw, K. T., and Welch, A. A. (2020). Lower dietary and circulating vitamin C in middle- and olderaged men and women are associated with lower estimated skeletal muscle mass. *J. Nutr.* 150 (10), 2789–2798. doi:10.1093/jn/nxaa221

Li, T. S., Shi, H., Wang, L., and Yan, C. (2016). Effect of bone marrow mesenchymal stem cells on satellite cell proliferation and apoptosis in immobilization-induced muscle atrophy in rats. *Med. Sci. Monit.* 22, 4651–4660. doi:10.12659/msm.898137

Li, Y., Foster, W., Deasy, B. M., Chan, Y., Prisk, V., Tang, Y., et al. (2004). Transforming growth factor-beta1 induces the differentiation of myogenic cells into fibrotic cells in injured skeletal muscle: A key event in muscle fibrogenesis. *Am.* J. Pathol. 164 (3), 1007–1019. doi:10.1016/s0002-9440(10)63188-4

Liu, C., Cheung, W. H., Li, J., Chow, S. K. H., Yu, J., Wong, S. H., et al. (2021). Understanding the gut microbiota and sarcopenia: A systematic review. *J. Cachexia Sarcopenia Muscle* 12 (6), 1393–1407. doi:10.1002/jcsm.12784

Lovsletten, N. G., Bakke, S. S., Kase, E. T., Ouwens, D. M., Thoresen, G. H., and Rustan, A. C. (2018). Increased triacylglycerol - fatty acid substrate cycling in human skeletal muscle cells exposed to eicosapentaenoic acid. *PLoS One* 13 (11), e0208048. doi:10.1371/journal.pone.0208048

Lu, Y., and Wahl, L. M. (2005). Oxidative stress augments the production of matrix metalloproteinase-1, cyclooxygenase-2, and prostaglandin E2 through enhancement of NF-kappa B activity in lipopolysaccharide-activated human primary monocytes. *J. Immunol.* 175 (8), 5423–5429. doi:10.4049/jimmunol.175. 8.5423

Mai, S., Klinkenberg, M., Auburger, G., Bereiter-Hahn, J., and Jendrach, M. (2010). Decreased expression of Drp1 and Fis1 mediates mitochondrial elongation in senescent cells and enhances resistance to oxidative stress through PINK1. *J. Cell Sci.* 123 (6), 917–926. doi:10.1242/jcs.059246

Makanae, Y., Kawada, S., SasaKi, K., NaKazato, K., and IshiiN. (2013). Vitamin C administration attenuates overload-induced skeletal muscle hypertrophy in rats. *Acta Physiol.* 208 (1), 57–65. doi:10.1111/apha.12042

Margaritis, I., TessierF.Prou, E., Marconnet, P., and Marini, J. F. (1997). Effects of endurance training on skeletal muscle oxidative capacities with and without selenium supplementation. J. Trace Elem. Med. Biol. 11 (1), 37–43. doi:10.1016/ S0946-672X(97)80008-9

Marzani, B., Balage, M., Venien, A., Astruc, T., Papet, I., Dardevet, D., et al. (2008). Antioxidant supplementation restores defective leucine stimulation of protein synthesis in skeletal muscle from old rats. J. Nutr. 138 (11), 2205–2211. doi:10.3945/jn.108.094029

Marzetti, E., Carter, C. S., Wohlgemuth, S. E., Lees, H. A., Giovannini, S., Anderson, B., et al. (2009). Changes in IL-15 expression and death-receptor apoptotic signaling in rat gastrocnemius muscle with aging and life-long calorie restriction. *Mech. Ageing Dev.* 130 (4), 272–280. doi:10.1016/j.mad. 2008.12.008

Marzetti, E., Wohlgemuth, S. E., Lees, H. A., Chung, H. Y., Giovannini, S., and Leeuwenburgh, C. (2008). Age-related activation of mitochondrial caspaseindependent apoptotic signaling in rat gastrocnemius muscle. *Mech. Ageing Dev.* 129 (9), 542–549. doi:10.1016/j.mad.2008.05.005

Meng, S. J., and Yu, L. J. (2010). Oxidative stress, molecular inflammation and sarcopenia. Int. J. Mol. Sci. 11 (4), 1509–1526. doi:10.3390/ijms11041509

Minet, A. D., and Gaster, M. (2012). Cultured senescent myoblasts derived from human vastus lateralis exhibit normal mitochondrial ATP synthesis capacities with correlating concomitant ROS production while whole cell ATP production is decreased. *Biogerontology* 13 (3), 277–285. doi:10.1007/ s10522-012-9372-9

Morley, J. E., Abbatecola, A. M., Argiles, J. M., Baracos, V., Bauer, J., Bhasin, S., et al. (2011). Sarcopenia with limited mobility: An international consensus. J. Am. Med. Dir. Assoc. 12 (6), 403–409. doi:10.1016/j.jamda.2011.04.014

Nakamura, S., Sato, Y., Kobayashi, T., Kaneko, Y., Ito, E., Soma, T., et al. (2020). Vitamin D protects against immobilization-induced muscle atrophy via neural crest-derived cells in mice. *Sci. Rep.* 10 (1), 12242. doi:10.1038/s41598-020-69021-y

Ni, Y., Yang, X., Zheng, L., Wang, Z., Wu, L., Jiang, J., et al. (2019). Lactobacillus and bifdobacterium improves physiological function and cognitive ability in aged mice by the regulation of gut microbiota. *Mol. Nutr. Food Res.* 63 (22), e1900603. doi:10.1002/mnfr.201900603

Nikawa, T., Ulla, A., and Sakakibara, I. (2021). Polyphenols and their effects on muscle atrophy and muscle health. *Molecules* 26 (16), 4887. doi:10.3390/molecules26164887

Ogata, T., and Yamasaki, Y. (1997). Ultra-high-resolution scanning electron microscopy of mitochondria and sarcoplasmic reticulum arrangement in human red, white, and intermediate muscle fibers. *Anat. Rec.* 248 (2), 214–223. doi:10.1002/ (SICI)1097-0185(199706)248:2<214::AID-AR8>3.0.CO;2-S

Pagano, A. F., Brioche, T., Arc-Chagnaud, C., Demangel, R., Chopard, A., and Py, G. (2018). Short-term disuse promotes fatty acid infiltration into skeletal muscle. *J. Cachexia Sarcopenia Muscle* 9 (2), 335–347. doi:10.1002/jcsm.12259

Palomero, J., Vasilaki, A., Pye, D., McArdle, A., and Jackson, M. J. (2013). Aging increases the oxidation of dichlorohydrofluorescein in single isolated skeletal muscle fibers at rest, but not during contractions. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 305 (4), R351–R358. doi:10.1152/ajpregu.00530.2012

Picard, M., Taivassalo, T., Gouspillou, G., and Hepple, R. T. (2011). Mitochondria: Isolation, structure and function. *J. Physiol.* 589 (18), 4413–4421. doi:10.1113/jphysiol.2011.212712

Pisot, R., Marusic, U., Biolo, G., Mazzucco, S., Lazzer, S., Grassi, B., et al. (2016). Greater loss in muscle mass and function but smaller metabolic alterations in older compared with younger men following 2 wk of bed rest and recovery. *J. Appl. Physiol.* 120 (8), 922–929. doi:10.1152/japplphysiol.00858.2015

Pollock, N., Staunton, C. A., Vasilaki, A., McArdle, A., and Jackson, M. J. (2017). Denervated muscle fibers induce mitochondrial peroxide generation in neighboring innervated fibers: Role in muscle aging. *Free Radic. Biol. Med.* 112, 84–92. doi:10. 1016/j.freeradbiomed.2017.07.017

Powers, S. K. (2014). Can antioxidants protect against disuse muscle atrophy? Sports Med. 44 (2), S155–S165. doi:10.1007/s40279-014-0255-x

Powers, S. K., Morton, A. B., Ahn, B., and Smuder, A. J. (2016). Redox control of skeletal muscle atrophy. *Free Radic. Biol. Med.* 98, 208–217. doi:10.1016/j. freeradbiomed.2016.02.021

Radak, Z., Chung, H. Y., and Goto, S. (2008). Systemic adaptation to oxidative challenge induced by regular exercise. *Free Radic. Biol. Med.* 44 (2), 153–159. doi:10. 1016/j.freeradbiomed.2007.01.029

Rana, A., Rera, M., and Walker, D. W. (2013). Parkin overexpression during aging reduces proteotoxicity, alters mitochondrial dynamics, and extends lifespan. *Proc. Natl. Acad. Sci. U. S. A.* 110 (21), 8638–8643. doi:10.1073/pnas.1216197110

Sakellariou, G. K., Lightfoot, A. P., Earl, K. E., Stofanko, M., and McDonagh, B. (2017). Redox homeostasis and age-related deficits in neuromuscular integrity and function. *J. Cachexia Sarcopenia Muscle* 8 (6), 881–906. doi:10.1002/jcsm.12223

Sakellariou, G. K., Pearson, T., Lightfoot, A. P., Nye, G. A., Wells, N., Giakoumaki, I. I., et al. (2016). Mitochondrial ROS regulate oxidative damage and mitophagy but not age-related muscle fiber atrophy. *Sci. Rep.* 6, 33944. doi:10.1038/srep33944

Salmon, A. B., Richardson, A., and Perez, V. I. (2010). Update on the oxidative stress theory of aging: Does oxidative stress play a role in aging or healthy aging? *Free Radic. Biol. Med.* 48 (5), 642–655. doi:10.1016/j.freeradbiomed.2009.12.015

Salucci, S., and Falcieri, E. (2020). Polyphenols and their potential role in preventing skeletal muscle atrophy. *Nutr. Res.* 74, 10–22. doi:10.1016/j.nutres. 2019.11.004

Sandiford, S. D., Kennedy, K. A. M., Xie, X., Pickering, J. G., and Li, S. S. C. (2014). Dual oxidase maturation factor 1 (DUOXA1) overexpression increases reactive oxygen species production and inhibits murine muscle satellite cell differentiation. *Cell Commun. Signal.* 12, 5. doi:10.1186/1478-811X-12-5

Sanodiya, B. S., Thakur, G. S., Baghel, R. K., Prasad, G. B. K. S., and Bisen, P. S. (2009). Ganoderma lucidum: A potent pharmacological macrofungus. *Curr. Pharm. Biotechnol.* 10 (8), 717–742. doi:10.2174/138920109789978757

Santos, S. A., Silva, E. T., Caris, A. V., Lira, F. S., Tufik, S., and Dos Santos, R. V. T. (2016). Vitamin E supplementation inhibits muscle damage and inflammation after moderate exercise in hypoxia. *J. Hum. Nutr. Diet.* 29 (4), 516–522. doi:10.1111/jhn. 12361

Scalabrin, M., Pollock, N., Staunton, C. A., Brooks, S. V., McArdle, A., Jackson, M. J., et al. (2019). Redox responses in skeletal muscle following denervation. *Redox Biol.* 26, 101294. doi:10.1016/j.redox.2019.101294

Scherz-Shouval, R., Shvets, E., Fass, E., Shorer, H., Gil, L., and Elazar, Z. (2007). Reactive oxygen species are essential for autophagy and specifically regulate the activity of Atg4. *EMBO J.* 26 (7), 1749–1760. doi:10.1038/sj.emboj. 7601623

Schriner, S. E., Linford, N. J., Martin, G. M., Treuting, P., Ogburn, C. E., Emond, M., et al. (2005). Extension of murine life span by overexpression of catalase targeted to mitochondria. *Science* 308 (5730), 1909–1911. doi:10.1126/science. 1106653

Servais, S., Letexier, D., Favier, R., Duchamp, C., and Desplanches, D. (2007). Prevention of unloading-induced atrophy by vitamin E supplementation: Links between oxidative stress and soleus muscle proteolysis? *Free Radic. Biol. Med.* 42 (5), 627–635. doi:10.1016/j.freeradbiomed.2006.12.001

Sharma, B., and Dabur, R. (2020). Role of pro-inflammatory cytokines in regulation of skeletal muscle metabolism: A systematic review. *Curr. Med. Chem.* 27 (13), 2161–2188. doi:10.2174/0929867326666181129095309

Shen, W., Li, Y., Zhu, J., Schwendener, R., and Huard, J. (2008). Interaction between macrophages, TGF-beta1, and the COX-2 pathway during the inflammatory phase of skeletal muscle healing after injury. *J. Cell. Physiol.* 214 (2), 405–412. doi:10.1002/jcp.21212

Shi, M., Zhang, Z., and Yang, Y. (2013). Antioxidant and immunoregulatory activity of Ganoderma lucidum polysaccharide (GLP). *Carbohydr. Polym.* 95 (1), 200–206. doi:10.1016/j.carbpol.2013.02.081

Silva, K. A., Dong, J., Dong, Y., Dong, Y., Schor, N., Tweardy, D. J., et al. (2015). Inhibition of Stat3 activation suppresses caspase-3 and the ubiquitin-proteasome system, leading to preservation of muscle mass in cancer cachexia. *J. Biol. Chem.* 290 (17), 11177–11187. doi:10.1074/jbc.M115.641514

Sinha-Hikim, I., Sinha-Hikim, A. P., Parveen, M., Shen, R., Goswami, R., Tran, P., et al. (2013). Long-term supplementation with a cystine-based antioxidant delays loss of muscle mass in aging. *J. Gerontol. A Biol. Sci. Med. Sci.* 68 (7), 749–759. doi:10.1093/gerona/gls334

Smith, G. I., Julliand, S., Reeds, D. N., Sinacore, D. R., Klein, S., and Mittendorfer, B. (2015). Fish oil-derived n-3 PUFA therapy increases muscle mass and function in healthy older adults. *Am. J. Clin. Nutr.* 102 (1), 115–122. doi:10.3945/ajcn.114.105833

Son, D. O., Satsu, H., Kiso, Y., Totsuka, M., and Shimizu, M. (2008). Inhibitory effect of carnosine on interleukin-8 production in intestinal epithelial cells through translational regulation. *Cytokine* 42 (2), 265–276. doi:10.1016/j.cyto. 2008.02.011

Sousa-Victor, P., Gutarra, S., Garcia-Prat, L., Rodriguez-Ubreva, J., Ortet, L., Ruiz-Bonilla, V., et al. (2014). Geriatric muscle stem cells switch reversible quiescence into senescence. *Nature* 506 (7488), 316–321. doi:10.1038/nature13013

Sullivan-Gunn, M. J., and Lewandowski, P. A. (2013). Elevated hydrogen peroxide and decreased catalase and glutathione peroxidase protection are associated with aging sarcopenia. *BMC Geriatr.* 13, 104. doi:10.1186/1471-2318-13-104

Sun, J., He, H., and Xie, B. J. (2004). Novel antioxidant peptides from fermented mushroom Ganoderma lucidum. *J. Agric. Food Chem.* 52 (21), 6646–6652. doi:10. 1021/jf0495136

Szentesi, P., Csernoch, L., Dux, L., and Keller-Pinter, A. (2019). Changes in redox signaling in the skeletal muscle with aging. *Oxid. Med. Cell. Longev.* 2019, 4617801. doi:10.1155/2019/4617801

Takisawa, S., Funakoshi, T., Yatsu, T., Nagata, K., Aigaki, T., Machida, S., et al. (2019). Vitamin C deficiency causes muscle atrophy and a deterioration in physical performance. *Sci. Rep.* 9 (1), 4702. doi:10.1038/s41598-019-41229-7

Tan, A., Sullenbarger, B., Prakash, R., and McDaniel, J. C. (2018). Supplementation with eicosapentaenoic acid and docosahexaenoic acid reduces high levels of circulating proinflammatory cytokines in aging adults: A randomized, controlled study. *Prostagl. Leukot. Essent. Fat. Acids* 132, 23–29. doi:10.1016/j.plefa.2018.03.010

Ter Borg, S., de Groot, L. C. P. G. M., Mijnarends, D. M., de Vries, J. H. M., Verlaan, S., Meijboom, S., et al. (2016). Differences in nutrient intake and biochemical nutrient status between sarcopenic and nonsarcopenic older adultsresults from the maastricht sarcopenia study. *J. Am. Med. Dir. Assoc.* 17 (5), 393–401. doi:10.1016/j.jamda.2015.12.015

Thirupathi, A., Pinho, R. A., and Chang, Y. Z. (2020). Physical exercise: An inducer of positive oxidative stress in skeletal muscle aging. *Life Sci.* 252, 117630. doi:10.1016/j.lfs.2020.117630

Thyagarajan, A., Jiang, J., Hopf, A., Adamec, J., and Sliva, D. (2006). Inhibition of oxidative stress-induced invasiveness of cancer cells by Ganoderma lucidum is mediated through the suppression of interleukin-8 secretion. *Int. J. Mol. Med.* 18 (4), 657–664. doi:10.3892/ijmm.18.4.657

Ticinesi, A., Nouvenne, A., Cerundolo, N., Catania, P., Prati, B., Tana, C., et al. (2019). Gut microbiota, muscle mass and function in aging: A focus on physical frailty and sarcopenia. *Nutrients* 11 (7), E1633. doi:10.3390/nu11071633

Tieland, M., Trouwborst, I., and Clark, B. C. (2018). Skeletal muscle performance and ageing. J. Cachexia Sarcopenia Muscle 9 (1), 3–19. doi:10.1002/jcsm.12238

Umanskaya, A., Santulli, G., Xie, W., Andersson, D. C., Reiken, S. R., and Marks, A. R. (2014). Genetically enhancing mitochondrial antioxidant activity improves muscle function in aging. *Proc. Natl. Acad. Sci. U. S. A.* 111 (42), 15250–15255. doi:10.1073/pnas.1412754111

van Krimpen, S. J., Jansen, F. A. C., Ottenheim, V. L., Belzer, C., van der Ende, M., and van Norren, K. (2021). The effects of pro-pre-and synbiotics on muscle wasting, a systematic review-gut permeability as potential treatment target. *Nutrients* 13 (4), 1115. doi:10.3390/nu13041115

Vannice, G., and Rasmussen, H. (2014). Position of the academy of nutrition and dietetics: Dietary fatty acids for healthy adults. *J. Acad. Nutr. Diet.* 114 (1), 136–153. doi:10.1016/j.jand.2013.11.001

Vasilaki, A., and Jackson, M. J. (2013). Role of reactive oxygen species in the defective regeneration seen in aging muscle. *Free Radic. Biol. Med.* 65, 317–323. doi:10.1016/j.freeradbiomed.2013.07.008

Verdijk, L. B., Gleeson, B. G., Jonkers, R. A. M., Meijer, K., Savelberg, H. H. C. M., Dendale, P., et al. (2009). Skeletal muscle hypertrophy following resistance training is accompanied by a fiber type-specific increase in satellite cell content in elderly men. *J. Gerontol. A Biol. Sci. Med. Sci.* 64 (3), 332–339. doi:10.1093/gerona/gln050

Veronese, N., Berton, L., Carraro, S., Bolzetta, F., De Rui, M., Perissinotto, E., et al. (2014). Effect of oral magnesium supplementation on physical performance in healthy elderly women involved in a weekly exercise program: A randomized controlled trial. *Am. J. Clin. Nutr.* 100 (3), 974–981. doi:10.3945/ajcn.113.080168

Vistoli, G., Straniero, V., Pedretti, A., Fumagalli, L., Bolchi, C., Pallavicini, M., et al. (2012). Predicting the physicochemical profile of diastereoisomeric histidinecontaining dipeptides by property space analysis. *Chirality* 24 (7), 566–576. doi:10. 1002/chir.22056

Wang, J., Cao, B., Zhao, H., and Feng, J. (2017). Emerging roles of Ganoderma lucidum in anti-aging. Aging Dis. 8 (6), 691–707. doi:10.14336/AD.2017.0410

Wang, Q. Q., Jing, X. M., Bi, Y. Z., Cao, X. F., Wang, Y. Z., Li, Y. X., et al. (2018). Human umbilical cord wharton's jelly derived mesenchymal stromal cells may attenuate sarcopenia in aged mice induced by hindlimb suspension. *Med. Sci. Monit.* 24, 9272–9281. doi:10.12659/MSM.913362

Wang, Y., Liu, Q., Quan, H., Kang, S. G., Huang, K., and Tong, T. (2021). Nutraceuticals in the prevention and treatment of the muscle atrophy. *Nutrients* 13 (6), 1914. doi:10.3390/nu13061914

Xu, J., Liu, D., Yin, H., Tong, H., Li, S., and Yan, Y. (2018). Fatty acids promote bovine skeletal muscle satellite cell differentiation by regulating ELOVL3 expression. *Cell Tissue Res.* 373 (2), 499–508. doi:10.1007/s00441-018-2812-3

Yang, Y. F., Yang, W., Liao, Z. Y., Wu, Y. X., Fan, Z., Guo, A., et al. (2021). MICU3 regulates mitochondrial Ca(2+)-dependent antioxidant response in skeletal muscle aging. *Cell Death Dis.* 12 (12), 1115. doi:10.1038/s41419-021-04400-5

Yin, L., Li, N., Jia, W., Wang, N., Liang, M., Yang, X., et al. (2021). Skeletal muscle atrophy: From mechanisms to treatments. *Pharmacol. Res.* 172, 105807. doi:10. 1016/j.phrs.2021.105807

Yoshida, T., Nakamura, H., Masutani, H., and Yodoi, J. (2005). The involvement of thioredoxin and thioredoxin binding protein-2 on cellular proliferation and aging process. *Ann. N. Y. Acad. Sci.* 1055, 1–12. doi:10. 1196/annals.1323.002

You, Y. H., and Lin, Z. B. (2003). Antioxidant effect of Ganoderma polysaccharide peptide. *Yao Xue Xue Bao* 38 (2), 85–88.

Yu, S., Ren, B., Chen, H., Goltzman, D., Yan, J., and Miao, D. (2021). 1, 25-Dihydroxyvitamin D deficiency induces sarcopenia by inducing skeletal muscle cell senescence. *Am. J. Transl. Res.* 13 (11), 12638–12649.

Zembron-Lacny, A., Dziubek, W., Wolny-Rokicka, E., Dabrowska, G., and Wozniewski, M. (2019). The relation of inflammaging with skeletal muscle properties in elderly men. *Am. J. Mens. Health* 13 (2), 1557988319841934. doi:10.1177/1557988319841934

Zhonghui, Z., Xiaowei, Z., and Fang, F. (2014). Ganoderma lucidum polysaccharides supplementation attenuates exercise-induced oxidative stress in skeletal muscle of mice. *Saudi J. Biol. Sci.* 21 (2), 119–123. doi:10.1016/j.sjbs.2013. 04.004